

# **Unnatural amino acids increase sensitivity and provide for the design of highly selective caspase substrates**

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## **Supplemental section**









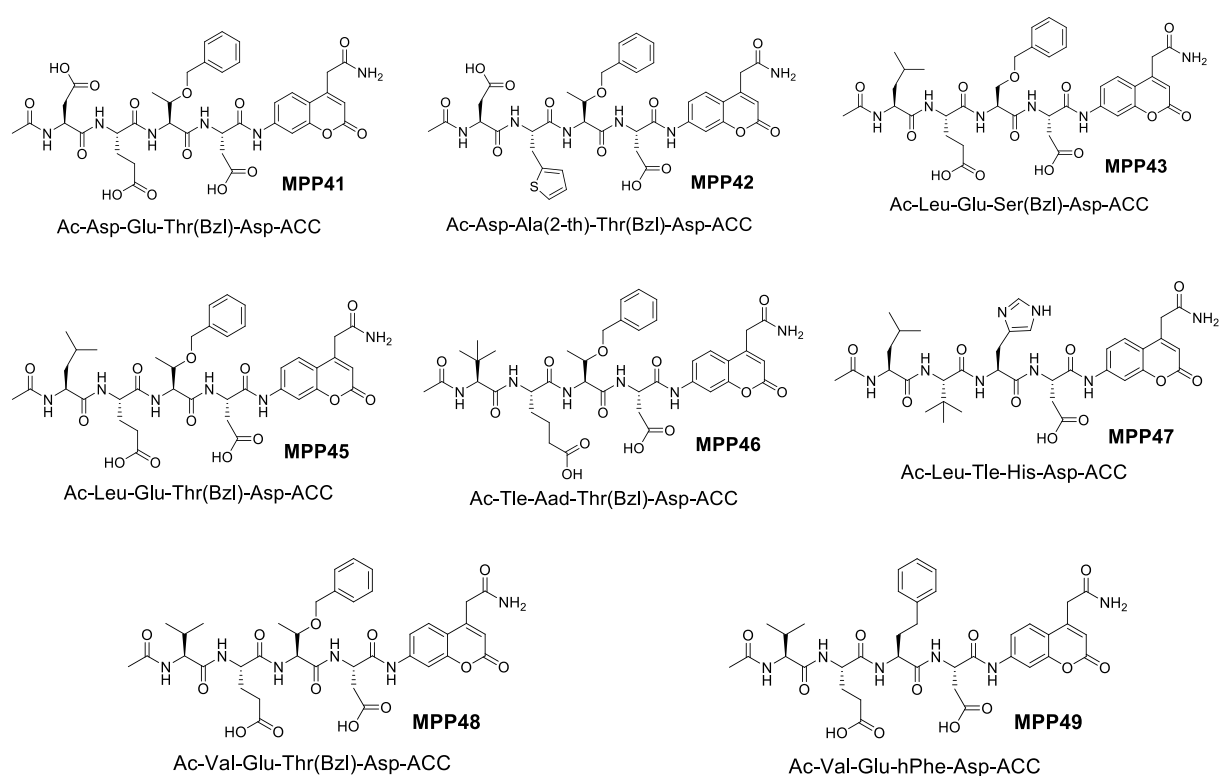






substrate	enzyme	$k_{\text{cat}}, \text{s}^{-1}$	$K_{\text{M}}, \mu\text{M}$	$k_{\text{cat}}/K_{\text{M}}, \text{M}^{-1}\text{s}^{-1}$
MPP41	Casp-3	30.1	28.2	1,050,000
MPP42	Casp-3	27.8	33.5	825,000
MPP48	Casp-6	4.08	23.0	186,300
MPP49	Casp-6	2.94	12.6	229,000
MPP41	Casp-7	6.88	41.6	167,000
MPP42	Casp-7	9.66	61.5	159,000
MPP45	Casp-8	0.361	1.04	349,000
MPP46	Casp-8	0.717	1.31	525,300
MPP47	Casp-9	1.18	32.1	37,000
MPP43	Casp-10	1.59	9.11	176,200

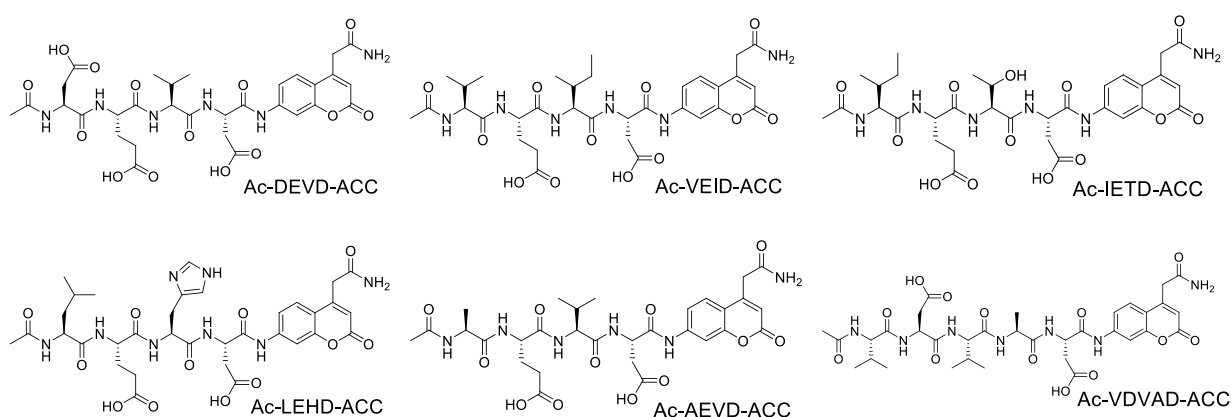
**Supplementary Table 1. Kinetic analysis of the best caspases substrates containing unnatural amino acids.**



**Supplementary Figure 7. The structures of the best short peptide caspases substrates**

sequence	enzyme		Casp-3	Casp-6	Cas-7	Casp-8	Casp-9	Casp-10
<b>DEVD</b>	Casp-3/7	$K_M, \mu\text{M}$	20.7	473	49.6	6.94	104	24.3
		$k_{\text{cat}}, \text{s}^{-1}$	10.0	0.642	3.92	0.496	0.033	0.867
		$k_{\text{cat}}/K_M, \text{M}^{-1}\text{s}^{-1}$	474,400	1,360	82,000	71,600	310	34,600
<b>VEID</b>	Casp-6	$K_M, \mu\text{M}$	218	45.6	285	2.12	25.6	20.9
		$k_{\text{cat}}, \text{s}^{-1}$	3.26	2.02	0.553	0.300	0.078	1.25
		$k_{\text{cat}}/K_M, \text{M}^{-1}\text{s}^{-1}$	14,700	44,800	1,870	14,100	3,300	58,400
<b>IETD</b>	Casp-8	$K_M, \mu\text{M}$	156	141	401	6.43	59.0	20.2
		$k_{\text{cat}}, \text{s}^{-1}$	0.962	1.72	0.121	0.607	0.144	1.76
		$k_{\text{cat}}/K_M, \text{M}^{-1}\text{s}^{-1}$	6,120	12,200	300	94,900	2,450	88,200
<b>LEHD</b>	Casp-9	$K_M, \mu\text{M}$	163	209	419	13.7	102	26.3
		$k_{\text{cat}}, \text{s}^{-1}$	1.61	2.83	0.305	2.54	1.35	3.87
		$k_{\text{cat}}/K_M, \text{M}^{-1}\text{s}^{-1}$	9,670	13,600	730	186,000	12,700	143,700
<b>AEVD</b>	Casp-10	$K_M, \mu\text{M}$	126	n.d.	143	8.59	57.8	33.1
		$k_{\text{cat}}, \text{s}^{-1}$	4.04	n.d.	2.64	0.686	0.136	0.510
		$k_{\text{cat}}/K_M, \text{M}^{-1}\text{s}^{-1}$	32,900	1,100	18,500	80,200	2,300	15,700
<b>VDVAD</b>	Casp-2	$K_M, \mu\text{M}$	30.2	n.d.	98.5	48.5	110	27.9
		$k_{\text{cat}}, \text{s}^{-1}$	3.46	n.d.	1.47	0.868	0.00941	0.710
		$k_{\text{cat}}/K_M, \text{M}^{-1}\text{s}^{-1}$	115,000	100	14,900	17,900	80	25,600

**Supplementary Table 2. Kinetic analysis of caspases fluorogenic substrates (Ac-P4-P3-P2-Asp-ACC) with reference tetrapeptide sequences and a caspase 2 reference substrate (Ac-VDVAD-ACC).**  $K_M$  values for Ac-AEVD-ACC and Ac-VDVAD-ACC substrates for caspase 6 were above 500  $\mu\text{M}$ , so we calculated only  $k_{\text{cat}}/K_M$  parameter (see Equation 2 in Supplementary Figure 12), n.d. not determined.



**Supplementary Figure 8. Reference structures of generally used peptidyl caspase-targeting sequences.**

substrate	enzyme	Casp-3	Casp-6	Casp-7	Casp-8	Casp-9	Casp-10
MPP38	Casp-3/7	<b>318,200</b>	96	<b>71,600</b>	2,654	27	2,020
MPP39		<b>181,000</b>	36	<b>40,500</b>	1,500	16	300
MPP40		<b>238,000</b>	39	<b>38,000</b>	1,430	7	1,560
MPP36	Casp-6	1,343	<b>26,600</b>	560	590	0	5,800
MPP28	Casp-8	520	110	200	<b>53,400</b>	600	7,800
MPP30		2,100	140	920	<b>137,500</b>	1,880	2,600
MPP8	Casp-9	0	30	0	2,600	<b>32,200</b>	960
MPP10		360	350	45	1,400	<b>27,500</b>	3,200
MPP12		0	0	0	1,580	<b>31,500</b>	1,120
MPP17	Casp-10	3,650	4,300	950	5,400	400	<b>48,600</b>
MPP21		40	300	74	380	490	<b>12,600</b>
MPP50		580	1,600	34	11,100	1,090	<b>59,800</b>
MPP51		120	25	0	10,800	590	<b>62,400</b>
MPP52		70	20	36	130	320	<b>10,300</b>
MPP53		50	86	51	20	160	<b>5,800</b>

**Supplementary Table 3. Kinetic analysis of the most specific substrates tested toward six human recombinant caspases.** Values are  $k_{cat}/K_M$  with units of  $M^{-1}s^{-1}$ .

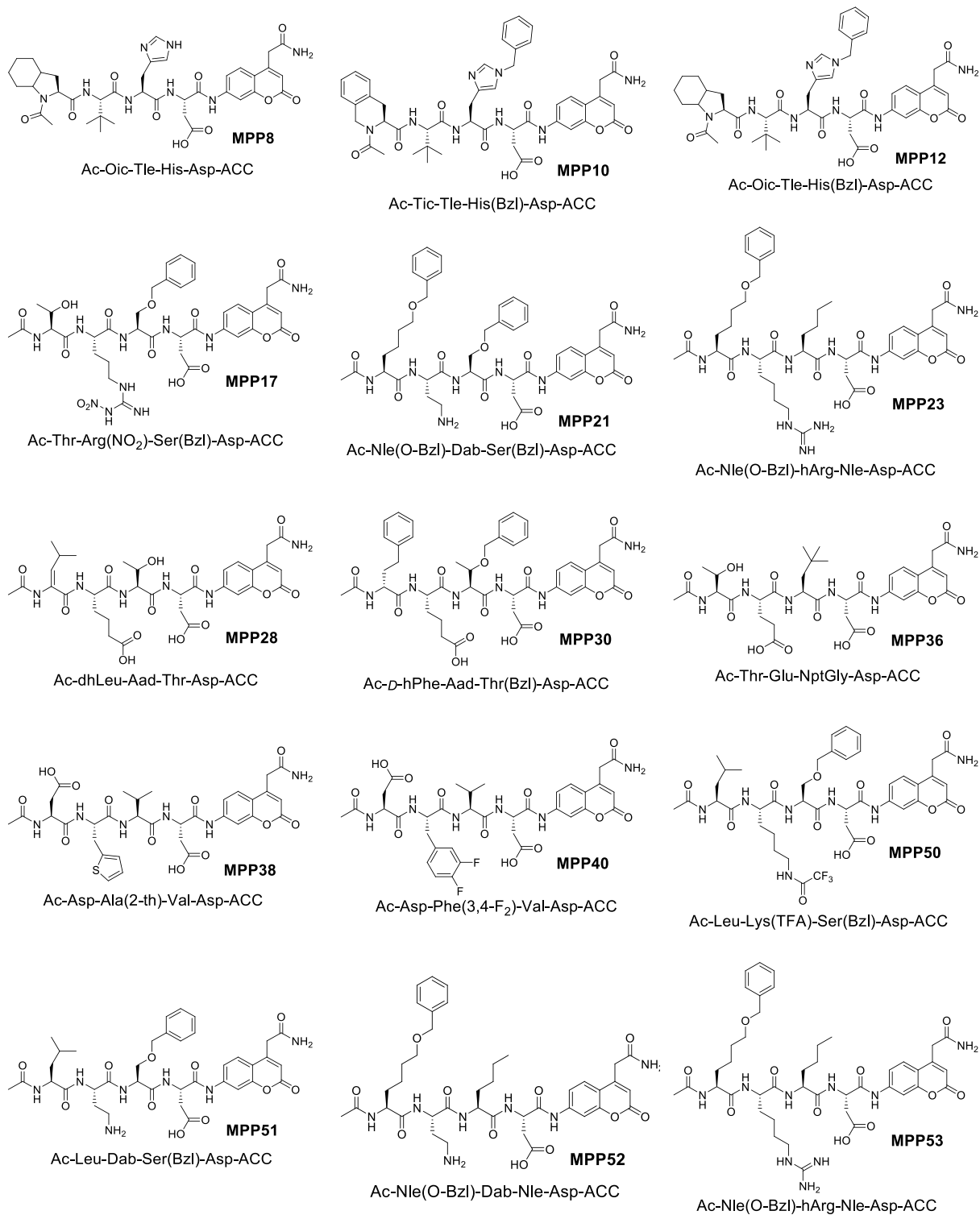
substrate	enzyme	$k_{\text{cat}}, \text{s}^{-1}$	$K_{\text{M}}, \mu\text{M}$	$k_{\text{cat}}/K_{\text{M}}, \text{M}^{-1}\text{s}^{-1}$
MPP38	Caspase-3	8.34	26.2	318,000
MPP39		6.44	35.6	181,000
MPP40		8.03	33.8	238,000
MPP36	Caspase-6	1.63	59.6	26,600
MPP38	Caspase-7	2.11	29.4	71,600
MPP39		2.01	51.8	40,500
MPP40		2.37	64.3	38,000
MPP28	Caspase-8	0.536	10.1	53,400
MPP30		0.222	1.59	137,000
MPP8	Caspase-9	2.07	68.6	32,200
MPP10		0.348	13.1	27,500
MPP12		0.642	21.1	31,500
MPP17	Caspase-10	1.07	22.6	48,600
MPP21		0.102	7.64	12,600
MPP50		0.204	3.55	59,800
MPP51		0.863	13.9	62,400
MPP52		0.121	12.1	10,300
MPP53		0.072	12.7	5,800

**Supplementary Table 4. Detailed kinetic analysis of the most selective caspases substrates.**

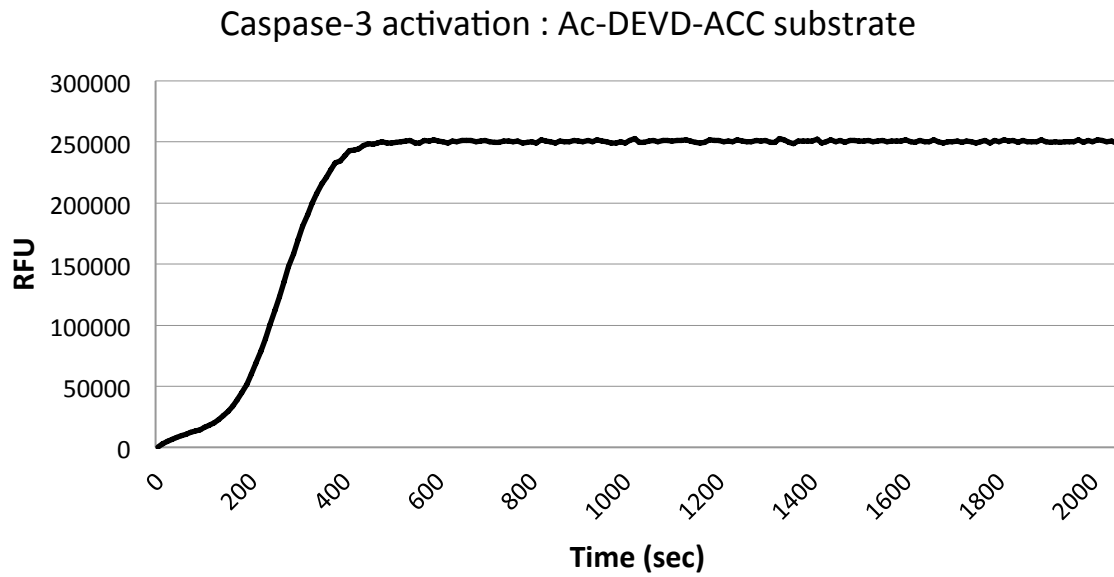
substrate/enzyme	Casp-3	Casp-6	Casp-7	Casp-8	Casp-9	Casp-10
Commercial substrates						
DEVD	<b>1415</b>	79	<b>470</b>	62	2	120
VEID	172	<b>371</b>	16	36	12	188
IETD	60	101	3	<b>78</b>	19	326
LEHD	92	307	6	614	<b>103</b>	618
AEVD	394	56	134	68	17	<b>83</b>
VDVAD	542	0	90	71	1	125
Substrates containing unnatural amino acids						
MPP39	<b>872</b>	0	<b>305</b>	11	0	3
MPP36	14	<b>345</b>	3	5	0	36
MPP28	3	0	0	<b>94</b>	5	120
MPP30	34	1	7	<b>72</b>	0	3
MPP8	0	0	0	23	<b>256</b>	20
MPP10	0	0	0	22	<b>145</b>	25
MPP52	0	0	0	0	0	<b>47</b>
MPP23	0	0	0	3	1	<b>26</b>
MPP50	6	7	0	21	2	<b>154</b>

**Supplementary Table 5. Selectivity of individual substrates.** In this experiment we compared substrates with previously reported sequences with our most selective substrates toward six recombinant caspases. Each substrate concentration was held at 100  $\mu$ M, and all caspases were used in the final concentration of 50 nM. Results are presented as the initial rate of ACC liberation versus time (RFU/s). Each experiment was performed in standard caspase buffer (buffers for caspases 8, 9, and 10 were supplemented with 1.0 M of sodium citrate).

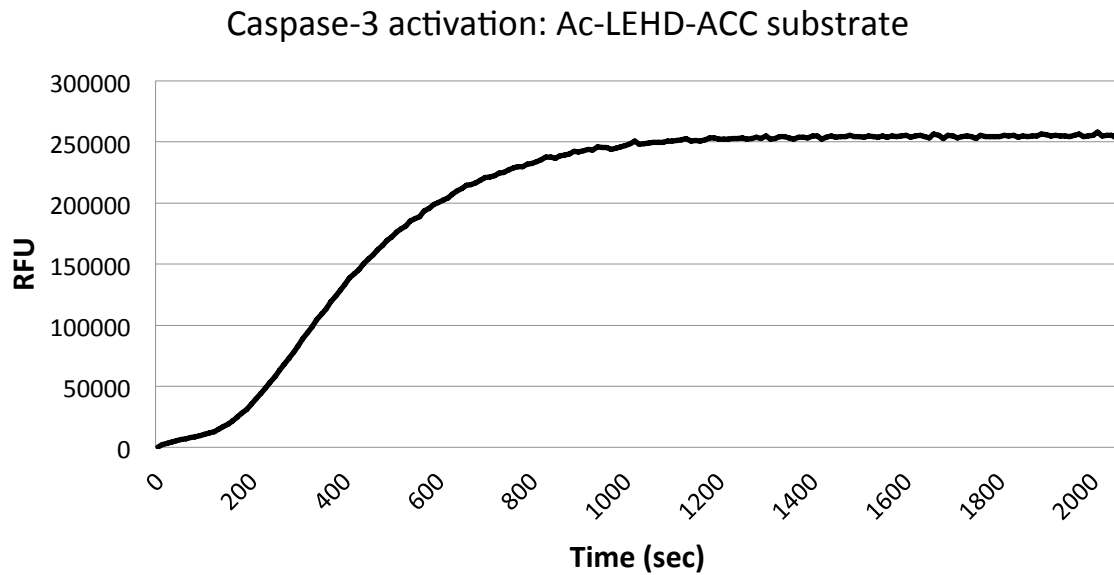




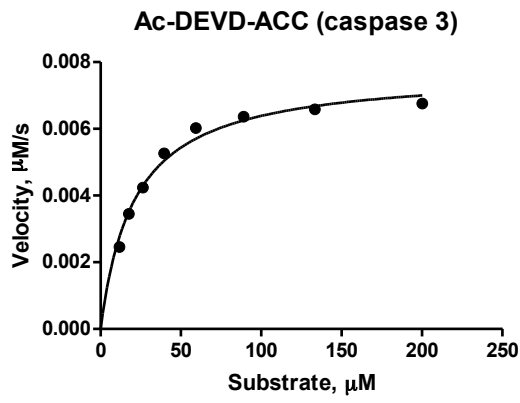
**Supplementary Figure 9. Structures of caspases-selective fluorogenic substrates.**



**Supplementary Figure 10. Caspase 3 activation measured with Ac-DEVD-ACC.** Substrate concentration 250  $\mu$ M. Cytosolic extract was treated with cytochrome C and dATP (see Materials and Methods) and used without pre-incubation. The assay was performed at 37°C.



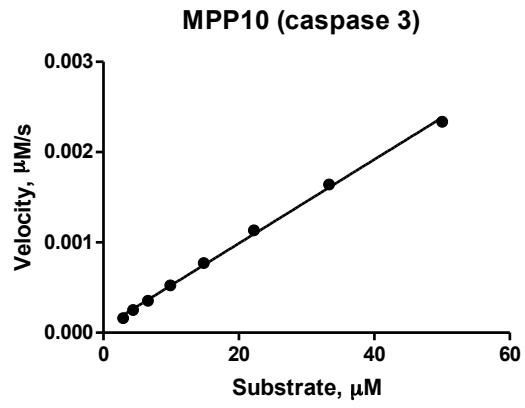
**Supplementary Figure 11. The Ac-LEHD-ACC hydrolysis time course.** Detailed kinetic analysis with the use of XIAP-BIR2 and XIAP-BIR3 clearly demonstrated that Ac-LEHD-ACC is hydrolyzed mainly by caspase 3, making this substrate useless as caspase 9 probe. Substrate concentration 250  $\mu$ M. Cytosolic extract was stimulated with cytochrome C and dATP (see Materials and Methods) and used without incubation. Assay was performed at 37°C.



Equation 1:

$$V_{max} = k_{cat} * [E] \rightarrow k_{cat} = \frac{V_{max}}{[E]}$$

$$\frac{k_{cat}}{K_m} = \frac{V_{max}}{[E] * K_m}$$

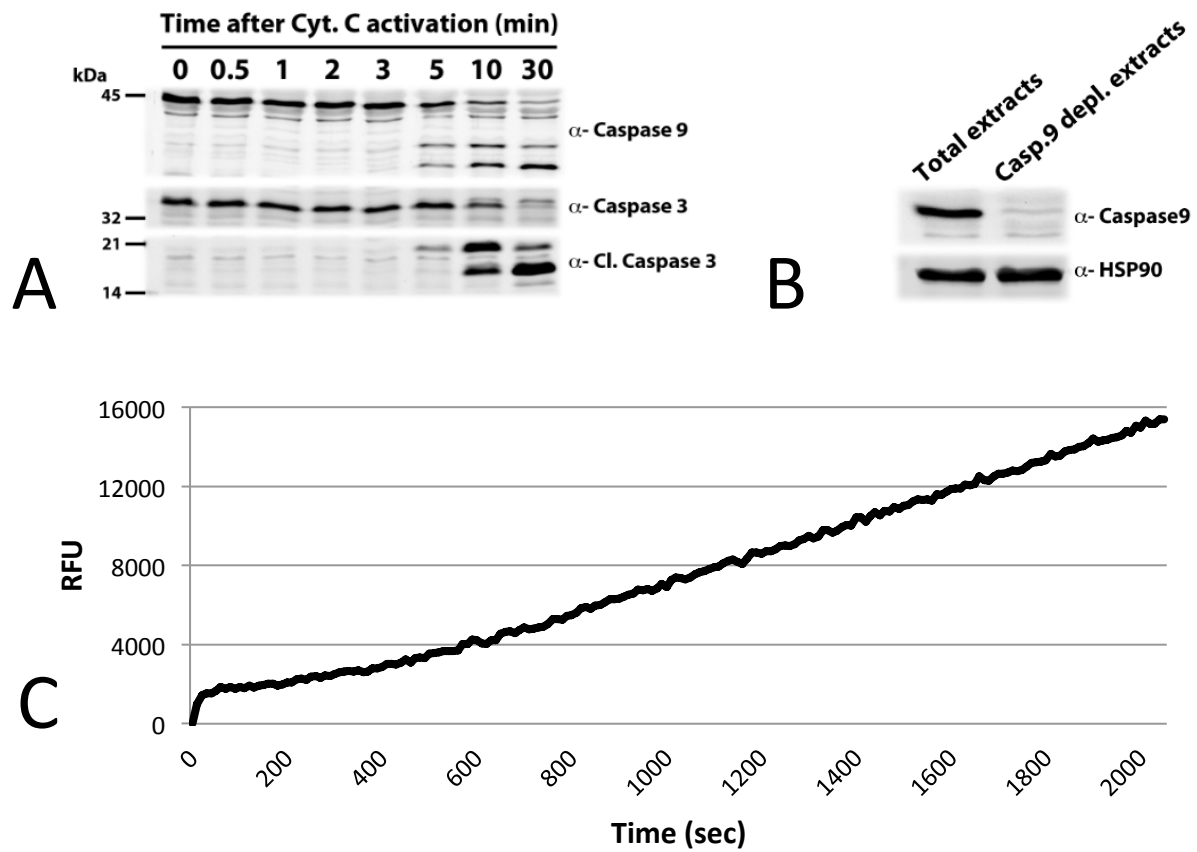


Equation 2:

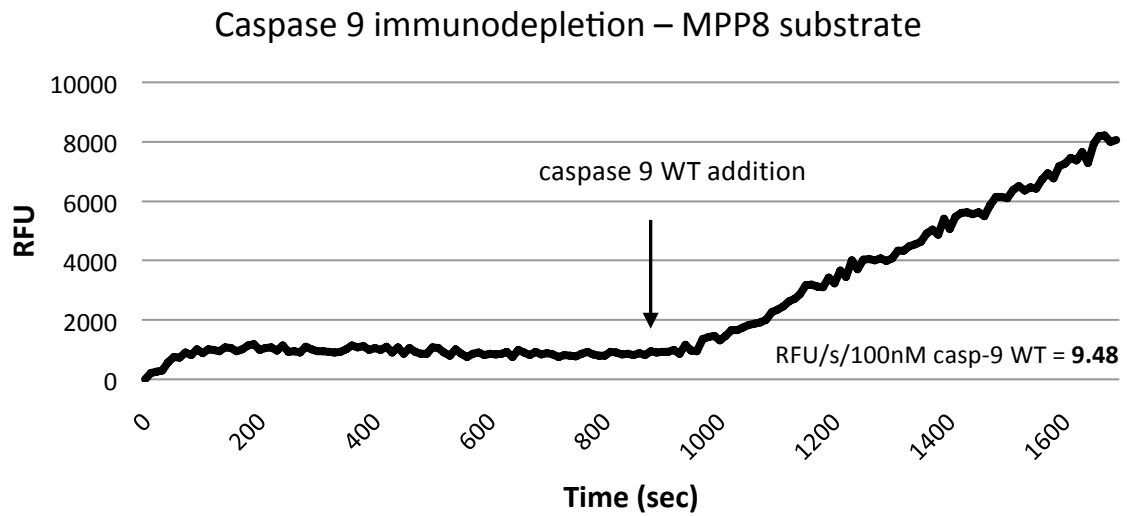
$$slope = \frac{\Delta V}{\Delta [S]}$$

$$\frac{k_{cat}}{K_m} = \frac{slope}{[E]} = \frac{\Delta V}{\Delta [S] * [E]}$$

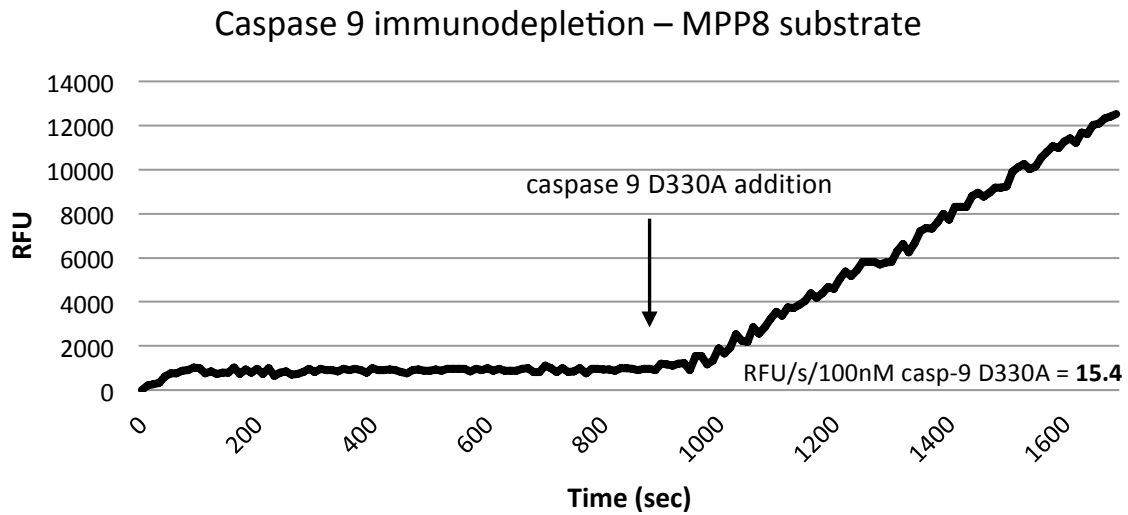
**Supplementary Figure 12. Calculation of  $k_{cat}/K_M$  parameters – for good substrates (left) and for poor substrates (right).**



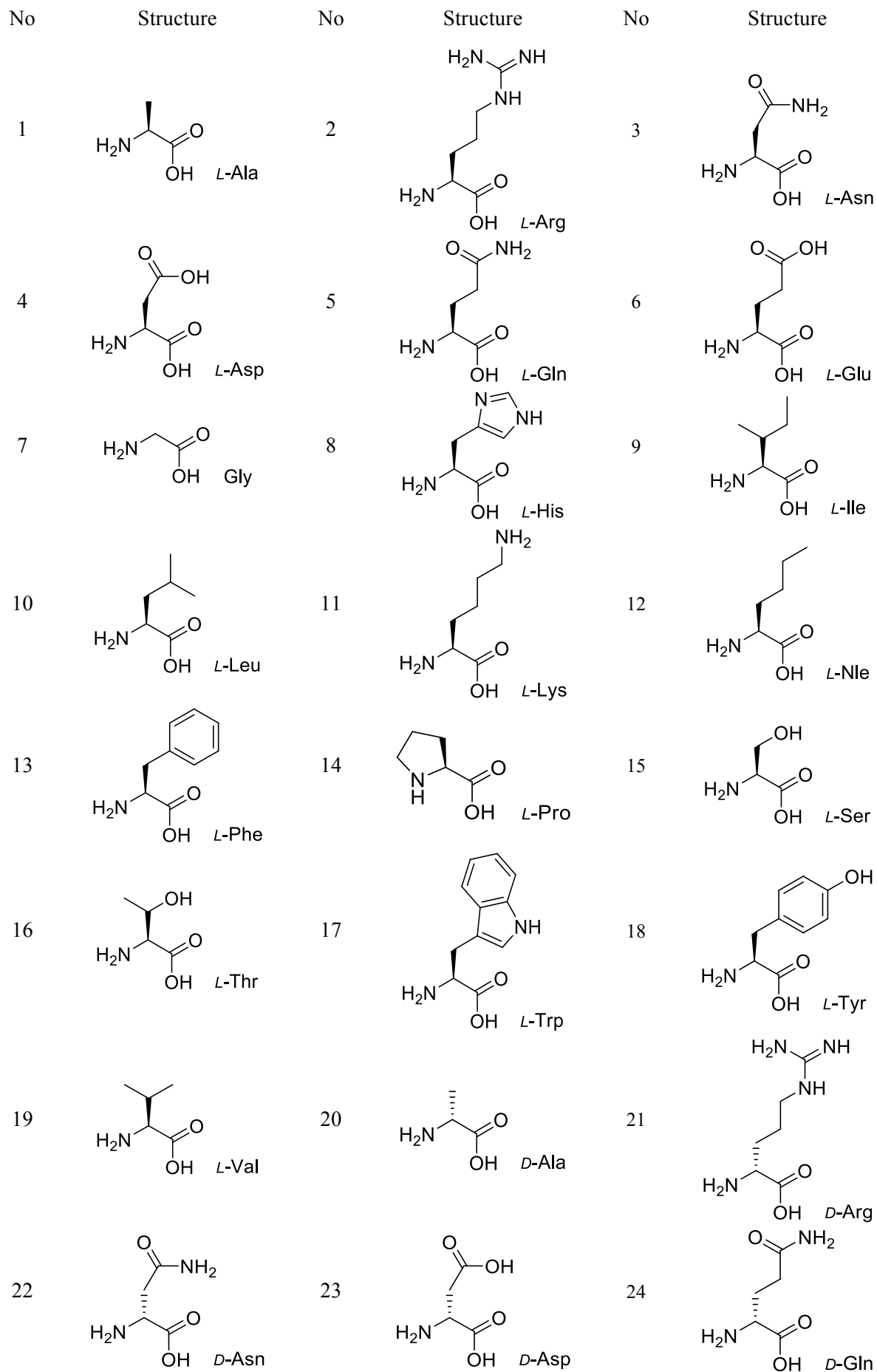
**Supplementary Figure 13. Time course of caspase 9 activation.** (A) Cytosolic extracts were stimulated with cytochrome C and caspase 9 and caspase 3 cleavage was monitored over the time course. (B) Caspase 9 immunodepletion from cytosolic extracts. Western Blot showing the absence of Caspase 9 in the immunodepleted sample. (C) Caspase 9 activation in cellular extract measured with the use of MPP8, a selective short peptide-based caspase 9 substrate. The “burst” in first 40-60 seconds is caused by cytochrome C and it appears also in the absence of cytosolic extracts (data not shown). The activity of caspase 9 can be measured after 60 seconds after cytochrome stimulation.



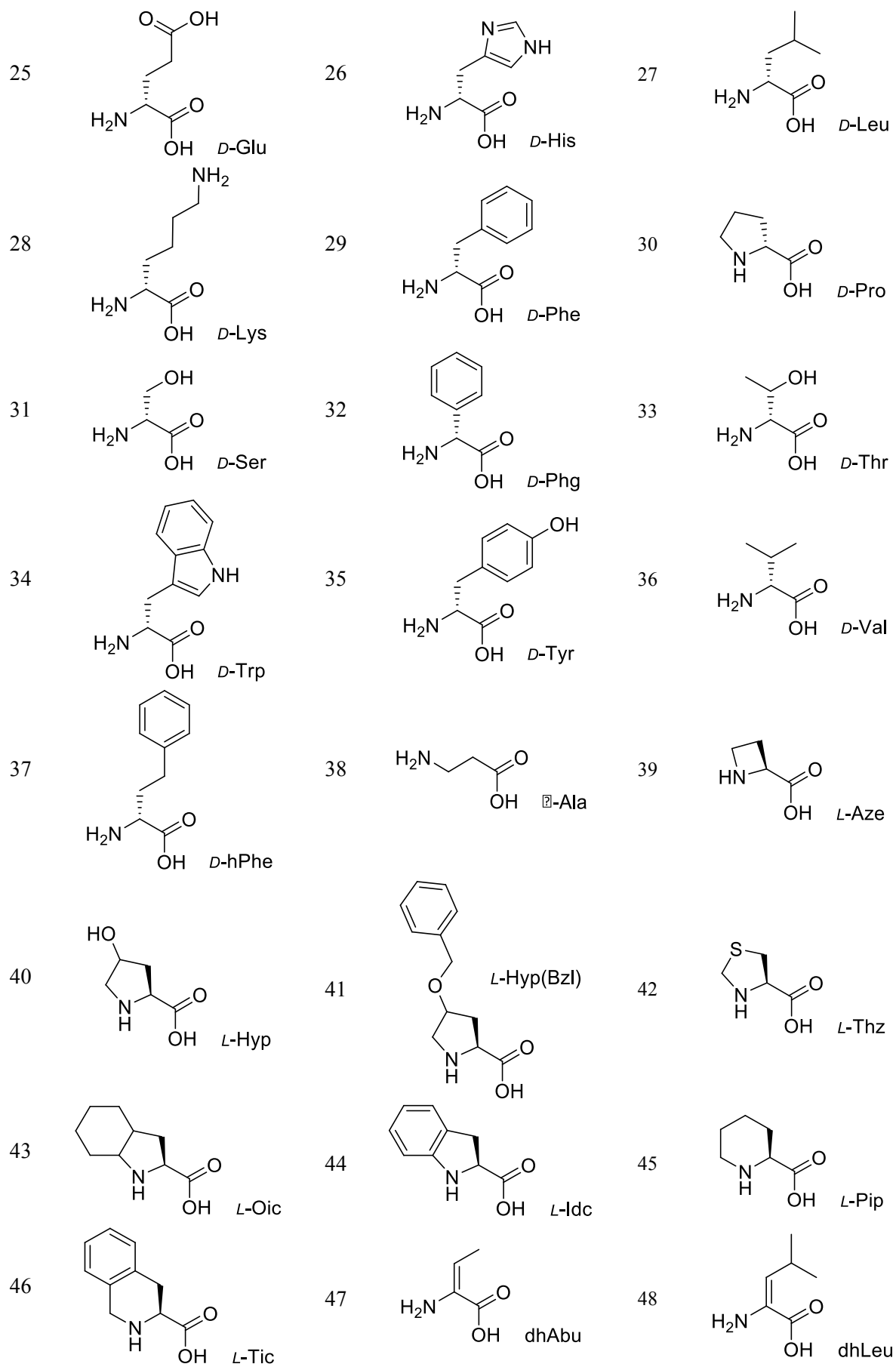
**Supplementary Figure 13. Monitoring of the MPP8 hydrolysis by the addition of caspase 9 WT to the caspase 9 immunodepleted cytosolic extract.** The proteolytic activity of caspase 9 immunodepleted cytosolic extract has been monitored with the use of MPP8 substrate (200  $\mu$ M). After 15 minutes a recombinant wild type caspase 9 was added (100 nM) and the measurement was continued.

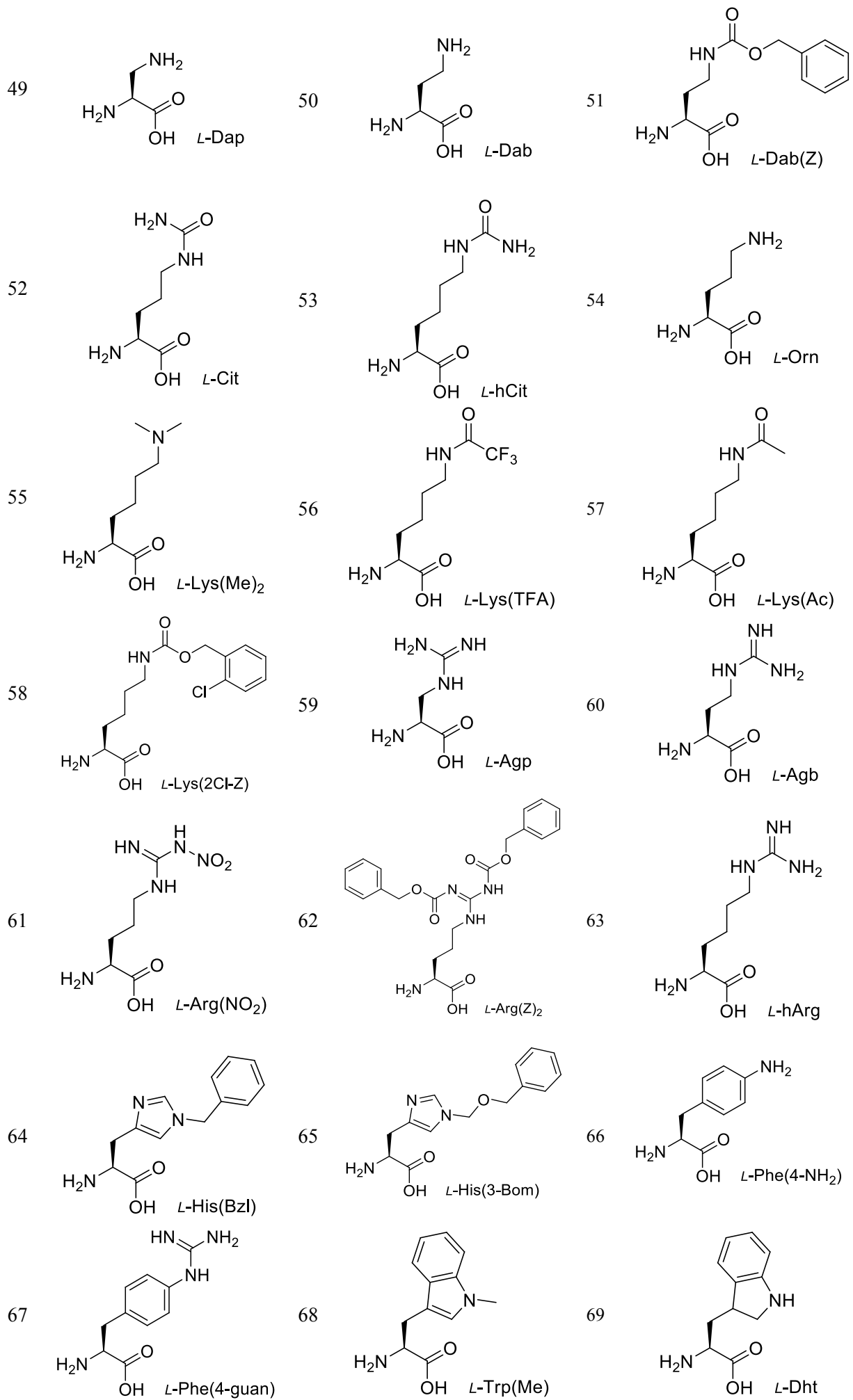


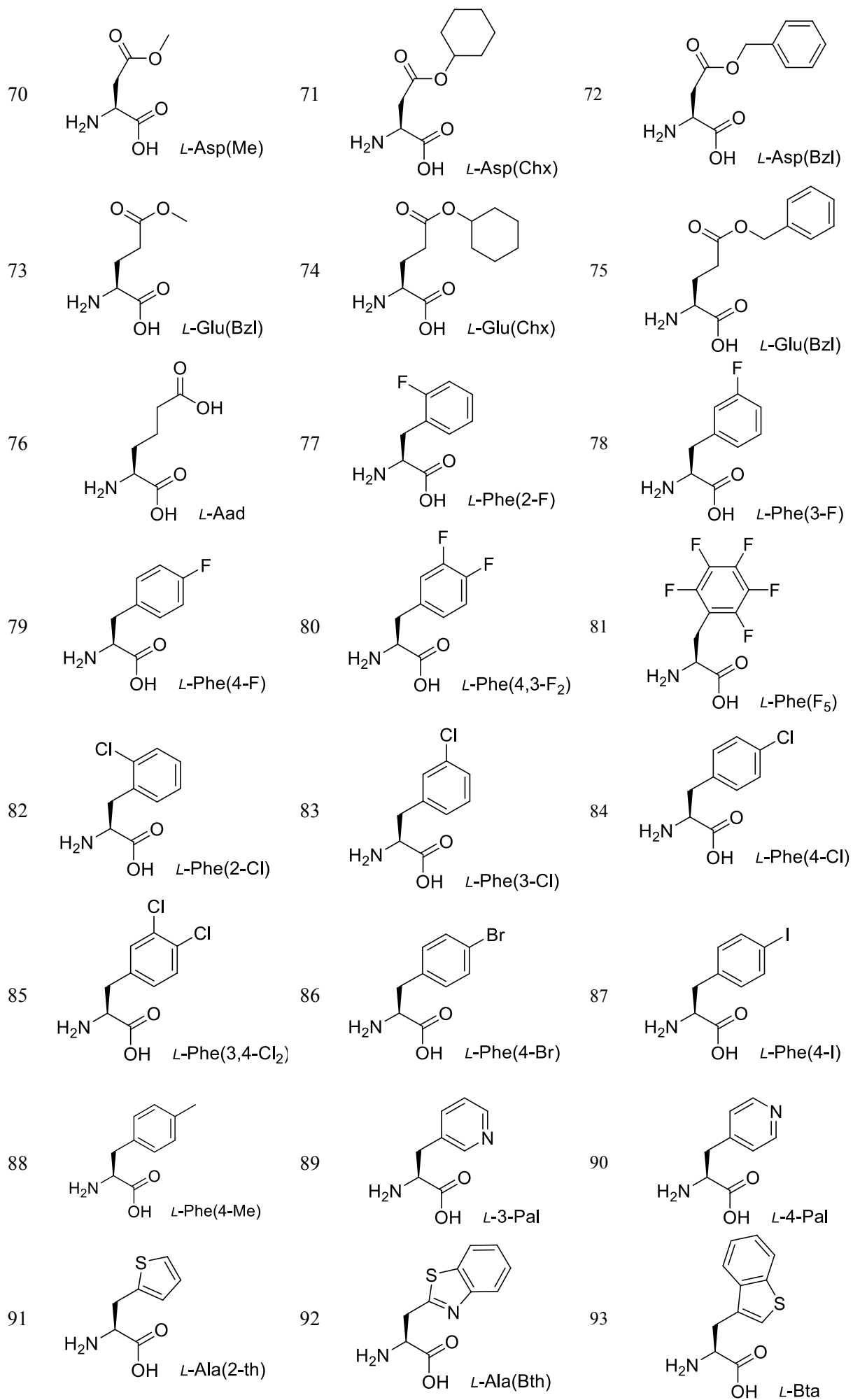
**Supplementary Figure 15. Monitoring of the MPP8 hydrolysis by the addition of caspase 9 D330A mutant to the caspase 9 immunodepleted cytosolic extract.** The proteolytic activity of caspase 9 immunodepleted cytosolic extract has been monitored with the use of MPP8 substrate (200  $\mu$ M). After 15 minutes a recombinant caspase 9 D330A mutant was added (100 nM) and the measurement was continued.

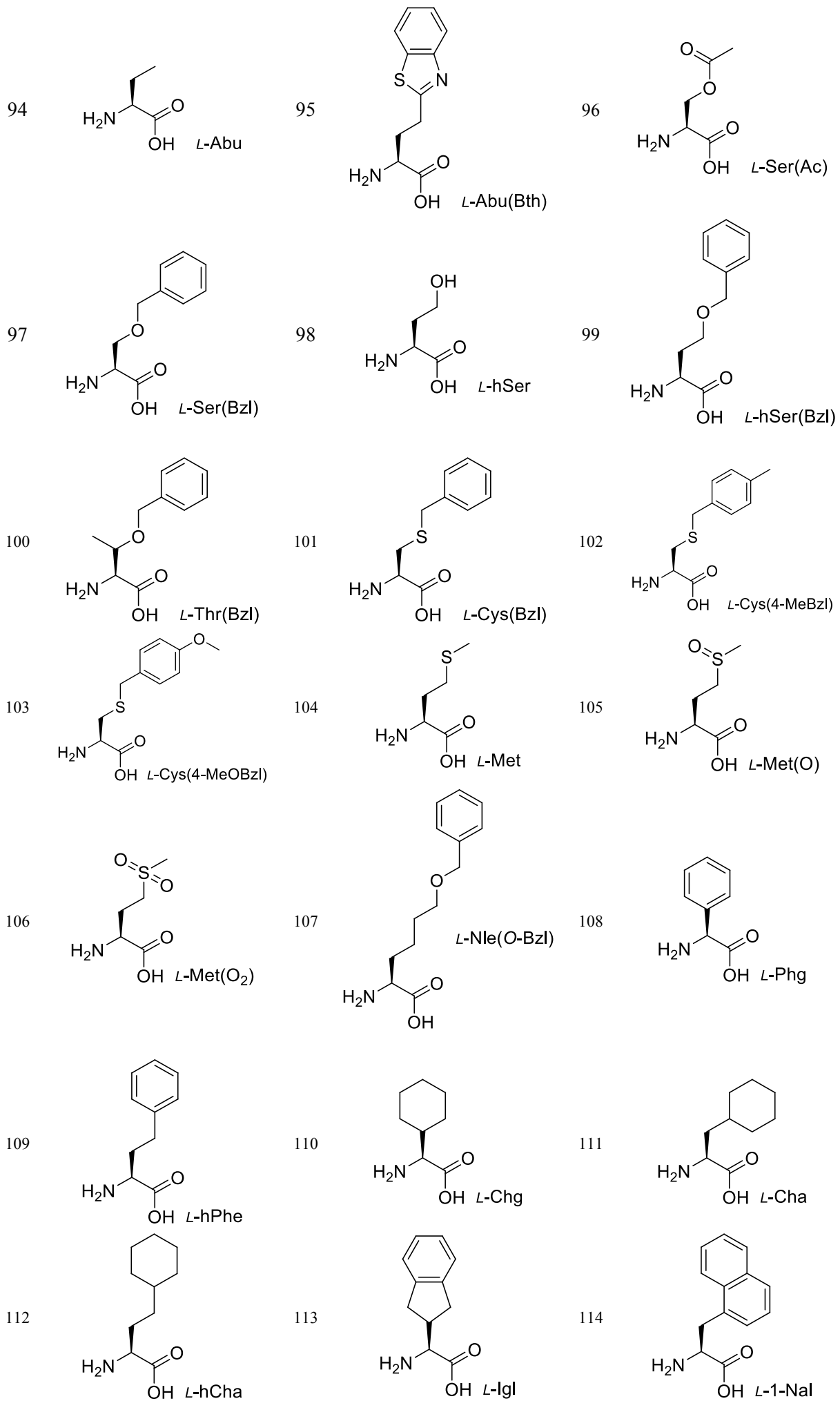


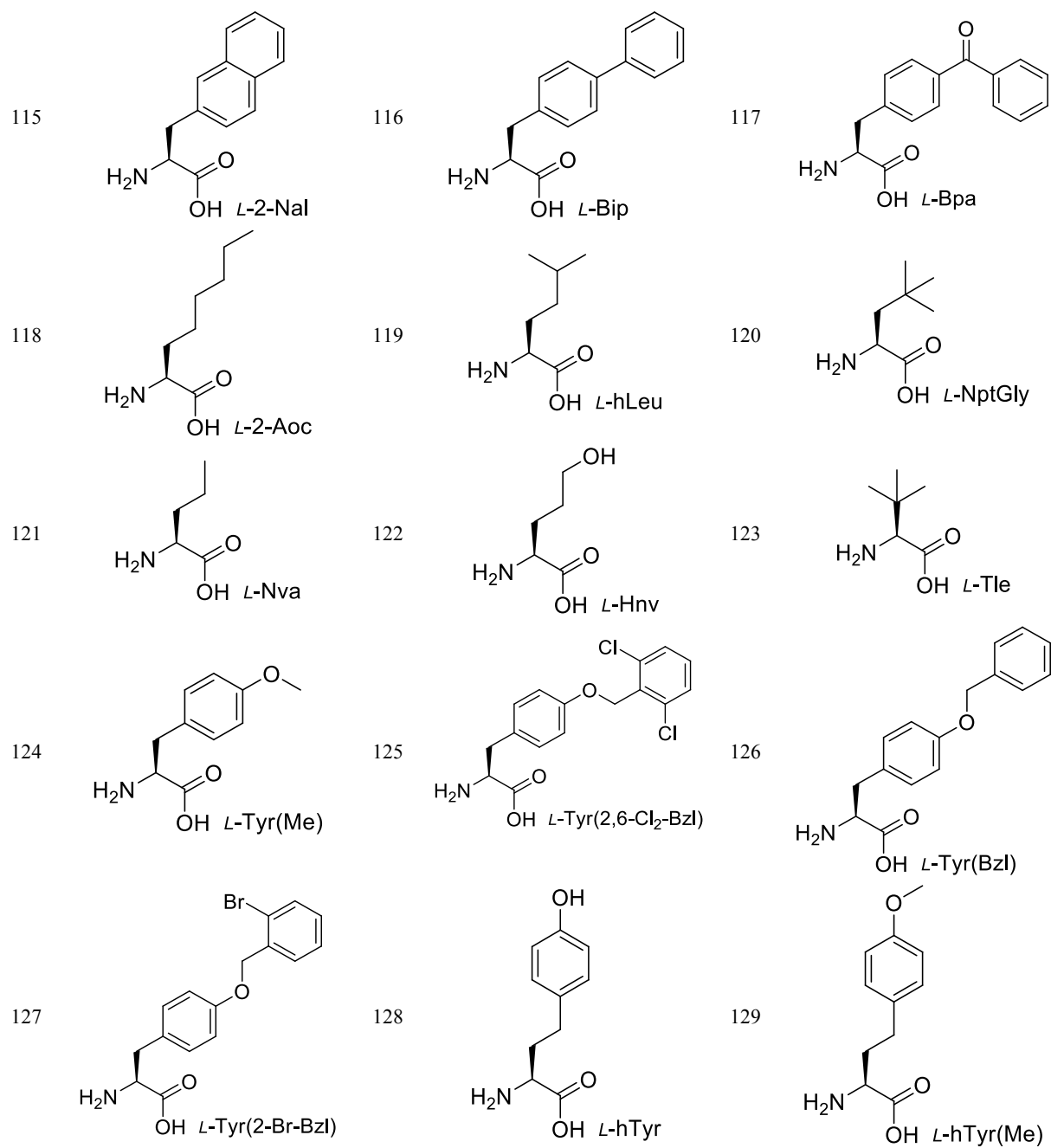












**Supplementary Table 5. Structure of amino acids used in HyCoSuL synthesis.**

## Compounds analysis

All individual compounds used for kinetic and biological studies were purified by HPLC on a Waters M600 solvent delivery module with a Waters M2489 detector system using Waters Spherisorb S50DS2 column and characterized by high-resolution mass spectrometry (HRMS) using on High Resolution Mass Spectrometer WATERS LCT Premier XE with Electrospray ionization (ESI). Overall yields for the complete synthesis and HRMS data are listed below.

**Ac-DEVD-ACC**, 32.1% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>31</sub>H<sub>38</sub>N<sub>6</sub>O<sub>14</sub>, 719.6652 found, 719.2532

**Ac-LEHD-ACC**, 29.3% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>34</sub>H<sub>42</sub>N<sub>8</sub>O<sub>12</sub>, 755.2922 found, 755.2966

**Ac-IETD-ACC**, 25.8% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>32</sub>H<sub>42</sub>N<sub>6</sub>O<sub>13</sub>, 719.2810 found, 719.2859

**Ac-AEVD-ACC**, 24.6% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>30</sub>H<sub>38</sub>N<sub>6</sub>NaO<sub>12</sub>, 697.2440 found, 697.2444

**Ac-VEID-ACC**, 33.3% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>33</sub>H<sub>44</sub>N<sub>6</sub>NaO<sub>12</sub>, 739.2909 found, 739.2908

**Ac-VDVAD-ACC**, 34.5% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>34</sub>H<sub>45</sub>N<sub>7</sub>NaO<sub>13</sub>, 782.2967 found, 782.2966

**MPP8**, 28.9% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>38</sub>H<sub>48</sub>N<sub>8</sub>O<sub>10</sub>, 777.3493 found, 777.3580

**MPP10**, 39.0% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>46</sub>H<sub>50</sub>N<sub>8</sub>O<sub>10</sub>, 875.3650 found, 875.3729

**MPP12**, 35.7% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>45</sub>H<sub>54</sub>N<sub>8</sub>O<sub>10</sub>, 867.3963 found, 867.3593

**MPP17**, 35.45% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>37</sub>H<sub>46</sub>N<sub>10</sub>O<sub>14</sub>, 855.3195 found, 855.3266

**MPP21**, 32.2% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>44</sub>H<sub>53</sub>N<sub>7</sub>O<sub>12</sub>, 872.3752 found, 872.3831

**MPP23**, 27.5% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>43</sub>H<sub>59</sub>N<sub>9</sub>O<sub>11</sub>, 878.4334 found, 878.4421

**MPP28**, 31.2% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>33</sub>H<sub>42</sub>N<sub>6</sub>O<sub>13</sub>, 731.2810 found, 731.2879

**MPP30**, 32.2% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>44</sub>H<sub>50</sub>N<sub>6</sub>NaO<sub>13</sub>, 893.3328 found, 893.3328

**MPP36**, 17.9% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>33</sub>H<sub>44</sub>N<sub>6</sub>O<sub>13</sub>, 733.2966 found, 733.3005

**MPP38**, 27.5% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>33</sub>H<sub>38</sub>N<sub>6</sub>NaO<sub>12</sub>S, 765.2160 found, 765.2180

**MPP39**, 31.1% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>35</sub>H<sub>35</sub>F<sub>5</sub>N<sub>6</sub>NaO<sub>12</sub>, 849.2125 found, 849.2155

**MPP40**, 32.8% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>35</sub>H<sub>38</sub>F<sub>2</sub>N<sub>6</sub>NaO<sub>12</sub>, 795.2408 found, 795.2421

**MPP41**, 28.6% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>37</sub>H<sub>42</sub>N<sub>6</sub>O<sub>15</sub>, 811.2708 found, 811.2769

**MPP42**, 26.5% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>39</sub>H<sub>42</sub>N<sub>6</sub>O<sub>13</sub>S, 835.2531 found, 835.2628

**MPP43**, 27.1% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>38</sub>H<sub>46</sub>N<sub>6</sub>NaO<sub>13</sub>, 817.3015 found, 817.2991

**MPP45**, 30.1% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>39</sub>H<sub>48</sub>N<sub>6</sub>NaO<sub>13</sub>, 831.3162 found, 831.3210

**MPP46**, 34.2% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>40</sub>H<sub>50</sub>N<sub>6</sub>NaO<sub>13</sub>, 845.3328 found, 845.3335

**MPP47**, 21.4% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>35</sub>H<sub>46</sub>N<sub>8</sub>O<sub>10</sub>, 739.3337 found, 739.3400

**MPP48**, 29.8% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>38</sub>H<sub>46</sub>N<sub>6</sub>NaO<sub>13</sub>, 817.3015 found, 817.3004

**MPP49**, 24.3% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>37</sub>H<sub>44</sub>N<sub>6</sub>NaO<sub>12</sub>, 787,2909 found, 787,2919

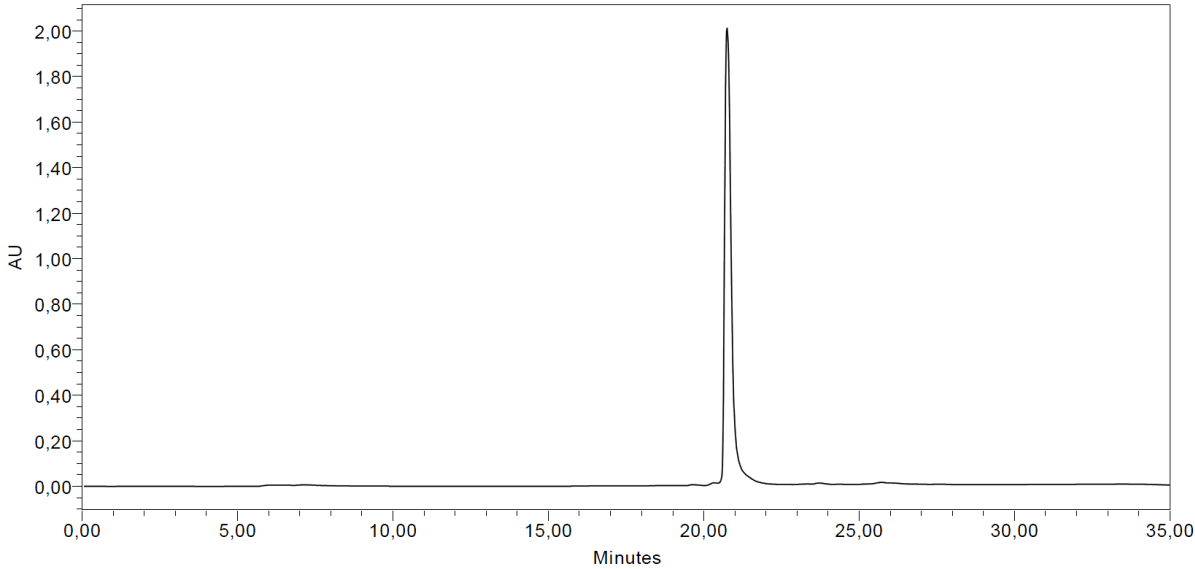
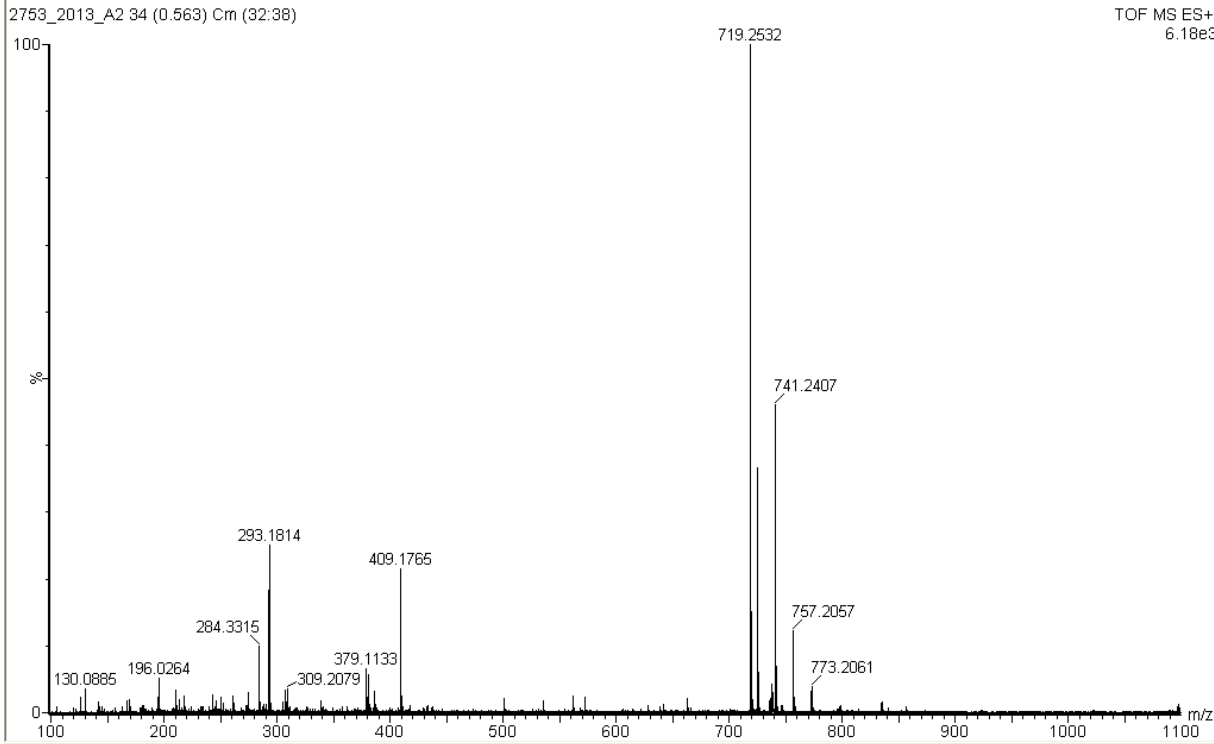
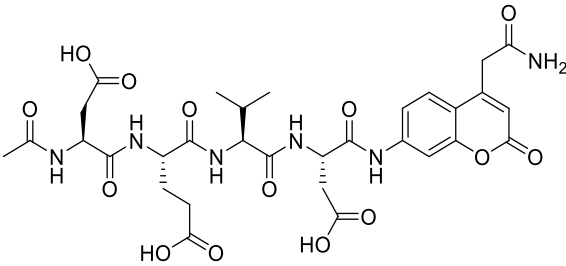
**MPP50**, 33.3% yield. HRMS (m/z): [MNa]<sup>+</sup> calcd for C<sub>42</sub>H<sub>50</sub>F<sub>3</sub>N<sub>7</sub>NaO<sub>12</sub>, 912.3362 found, 912.3362

**MPP51**, 26.4% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>37</sub>H<sub>47</sub>N<sub>7</sub>O<sub>11</sub>, 766.3334 found, 766.3416

**MPP52**, 29.6% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>40</sub>H<sub>53</sub>N<sub>7</sub>O<sub>11</sub>, 808.3803 found, 808.3878

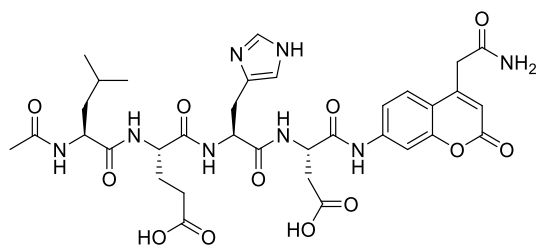
**MPP53**, 22.5% yield. HRMS (m/z): [M]<sup>+</sup> calcd for C<sub>43</sub>H<sub>59</sub>N<sub>9</sub>O<sub>11</sub>, 878.4334 found, 878.4424

Ac-DEVD-ACC



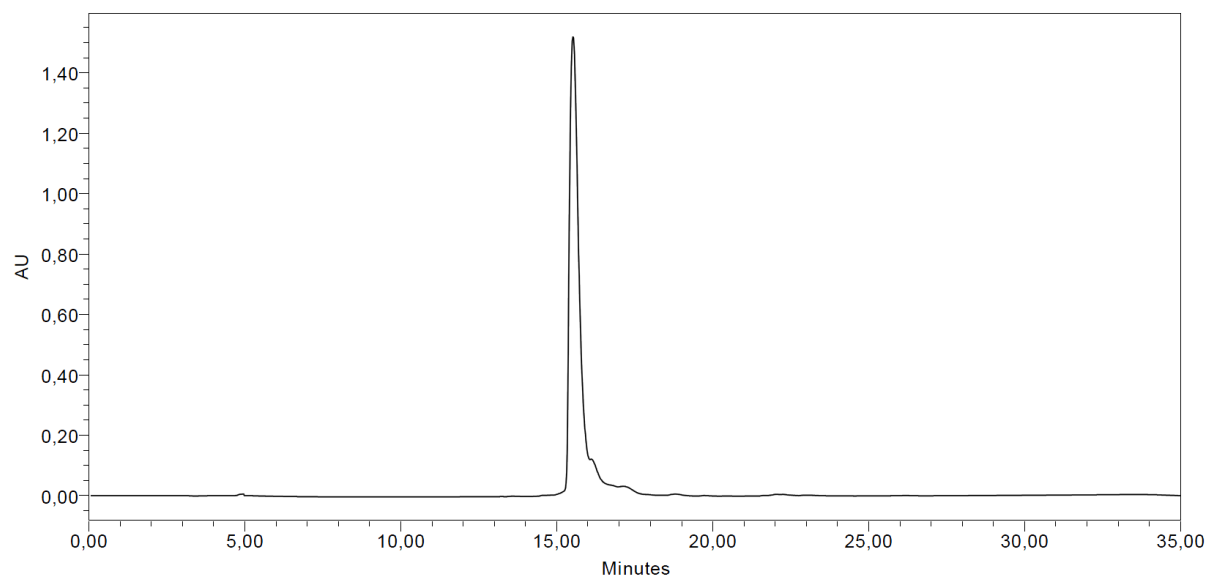
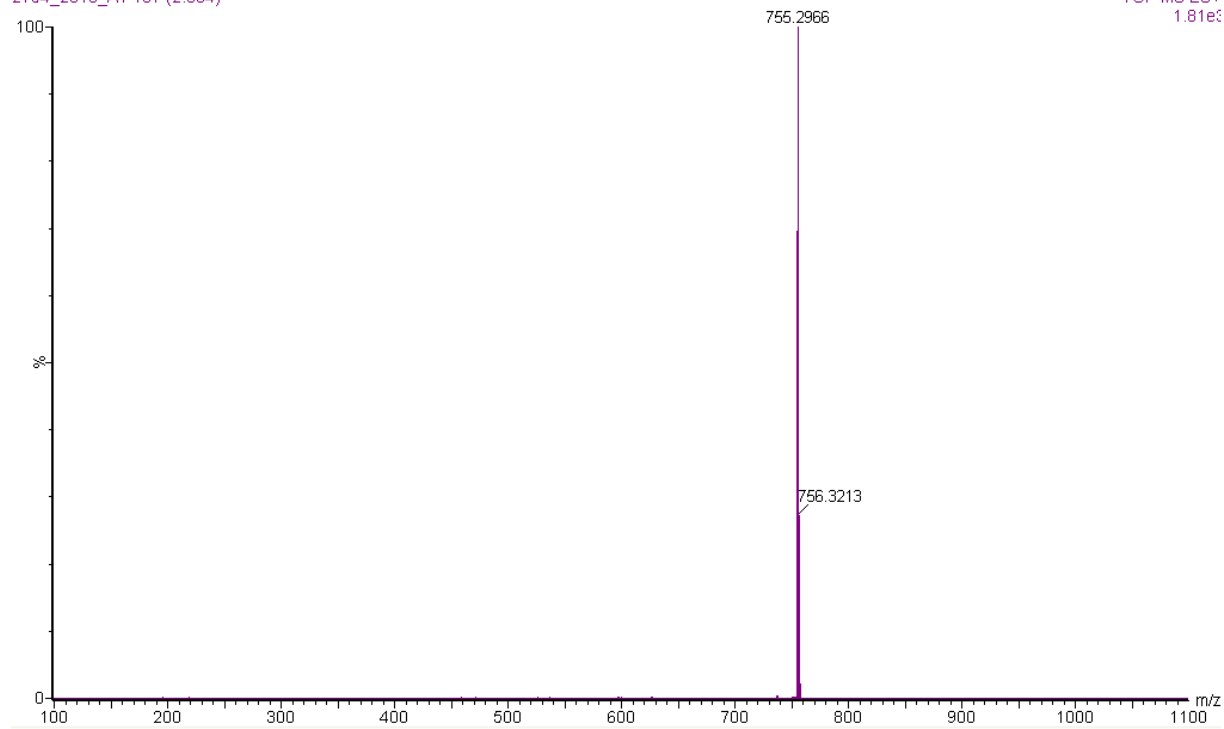


# Ac-LEHD-ACC

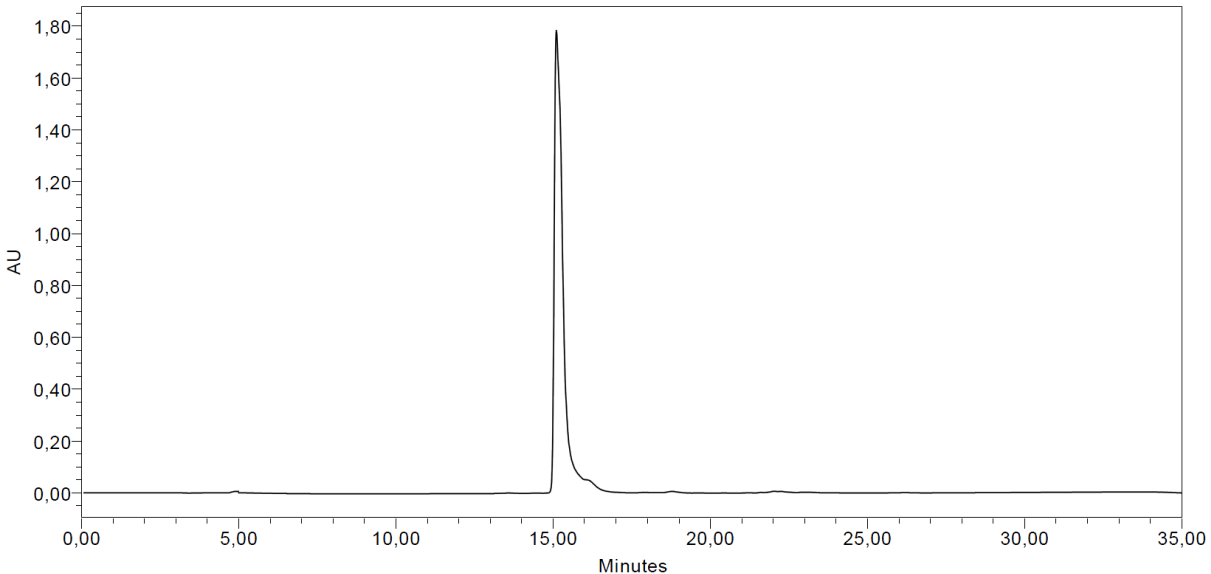
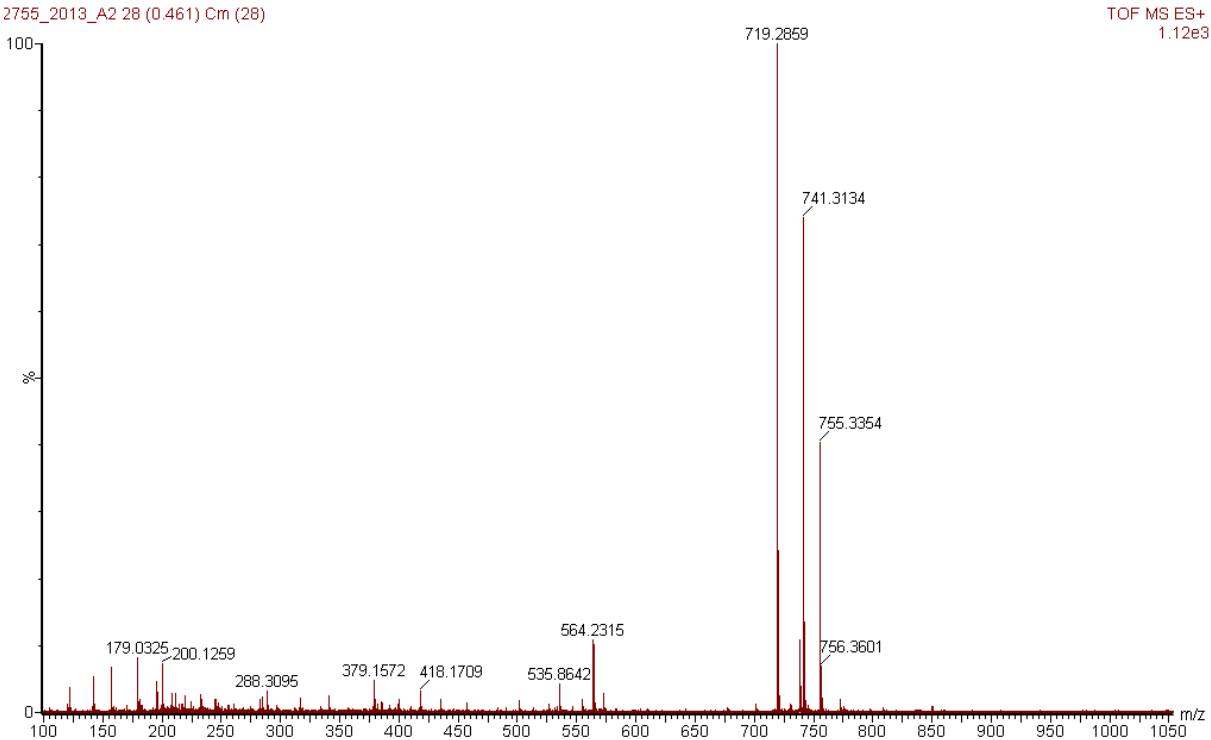
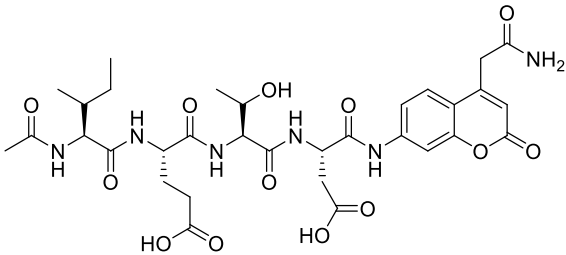


2754\_2013\_A1 167 (2.834)

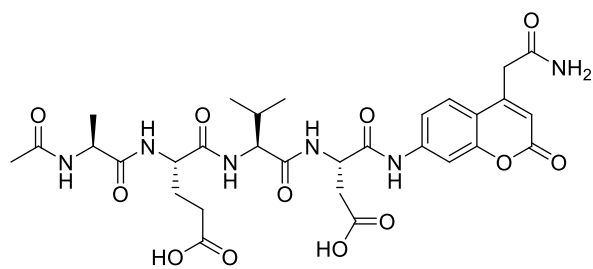
TOF MS ES+  
1.81e3



Ac-IETD-ACC

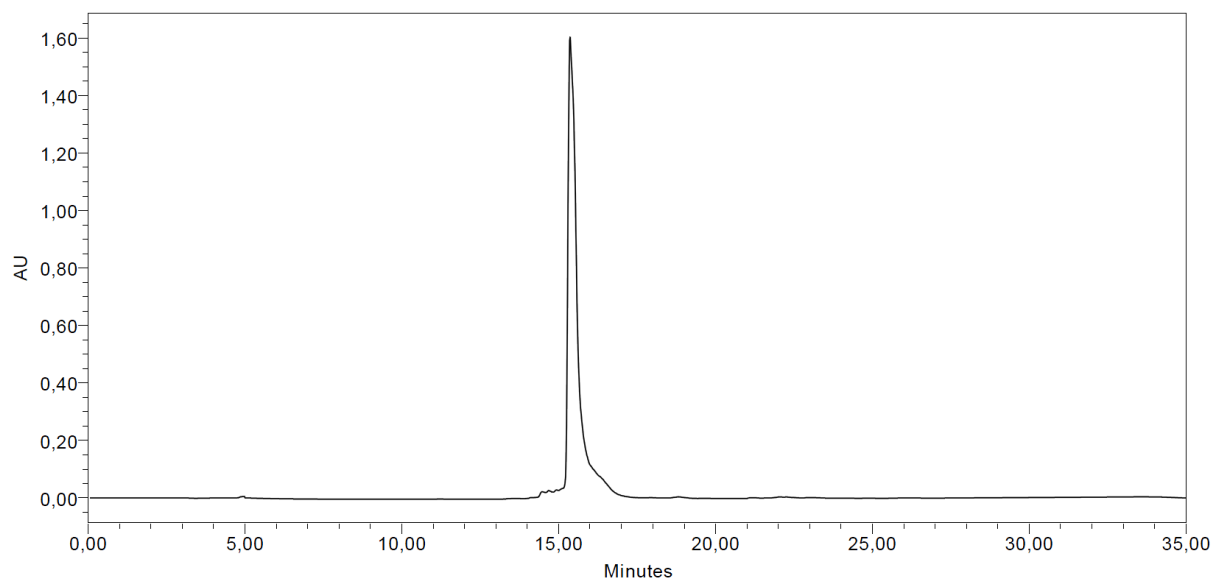
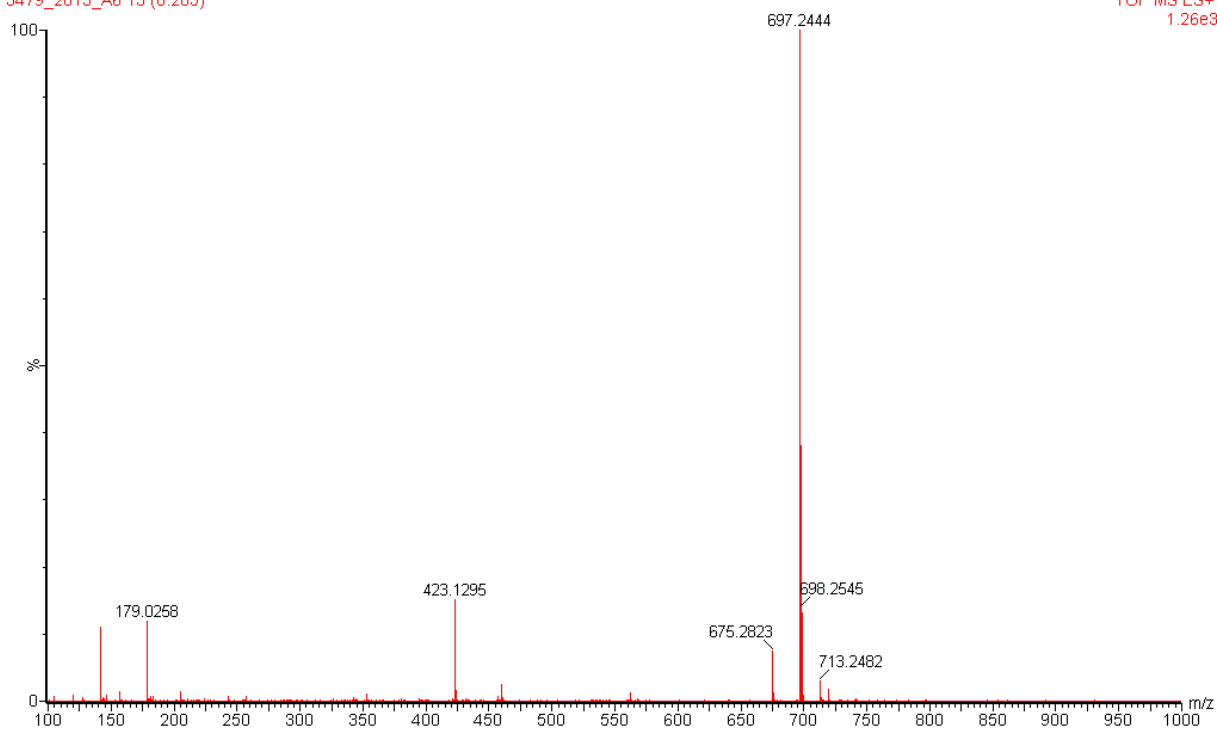


# Ac-AEVD-ACC



3479\_2013\_A6 13 (0.205)

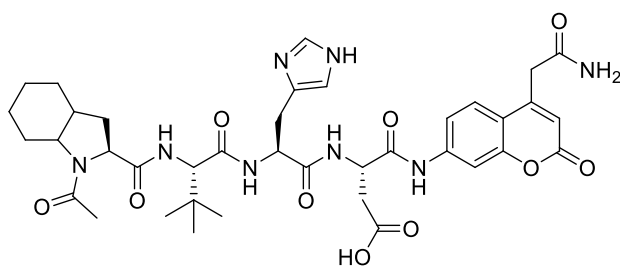
TOF MS ES+  
1.26e3



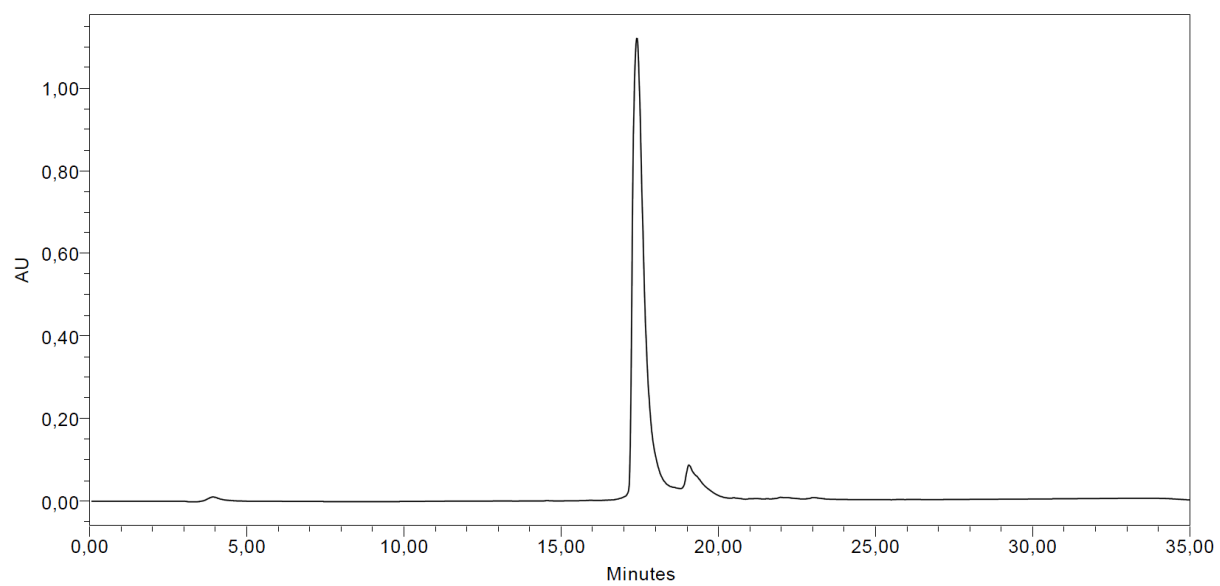
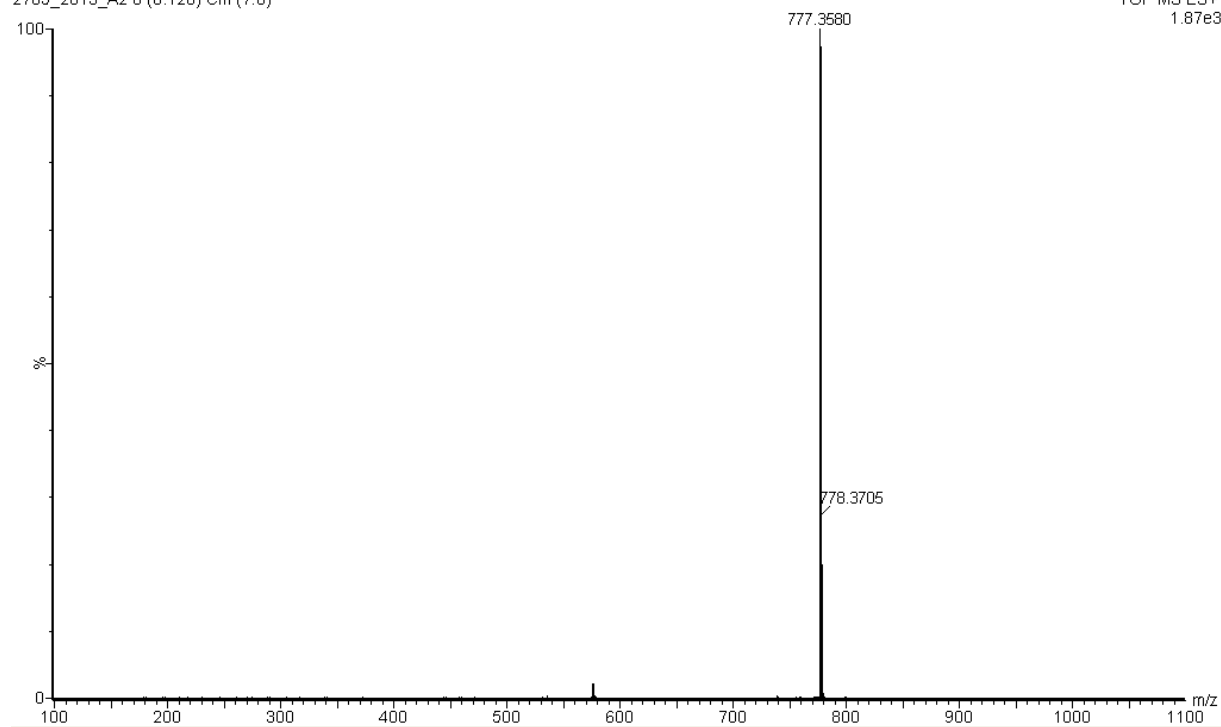




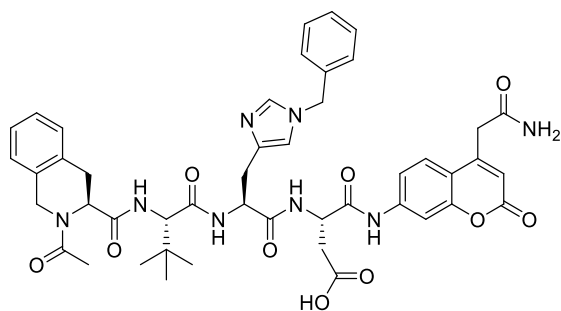
# MPP8



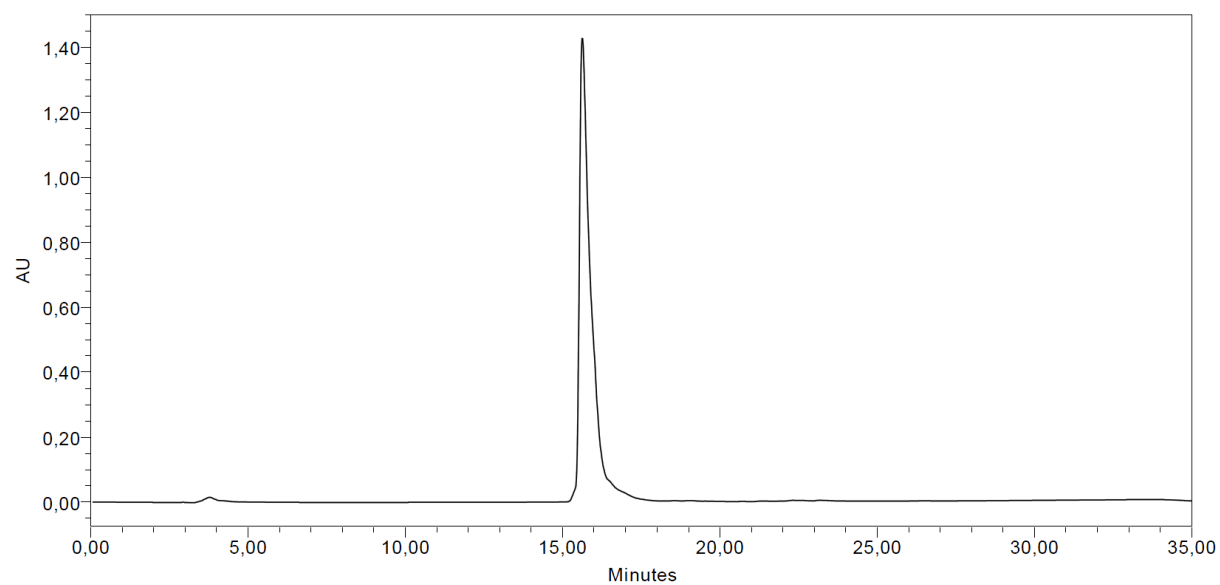
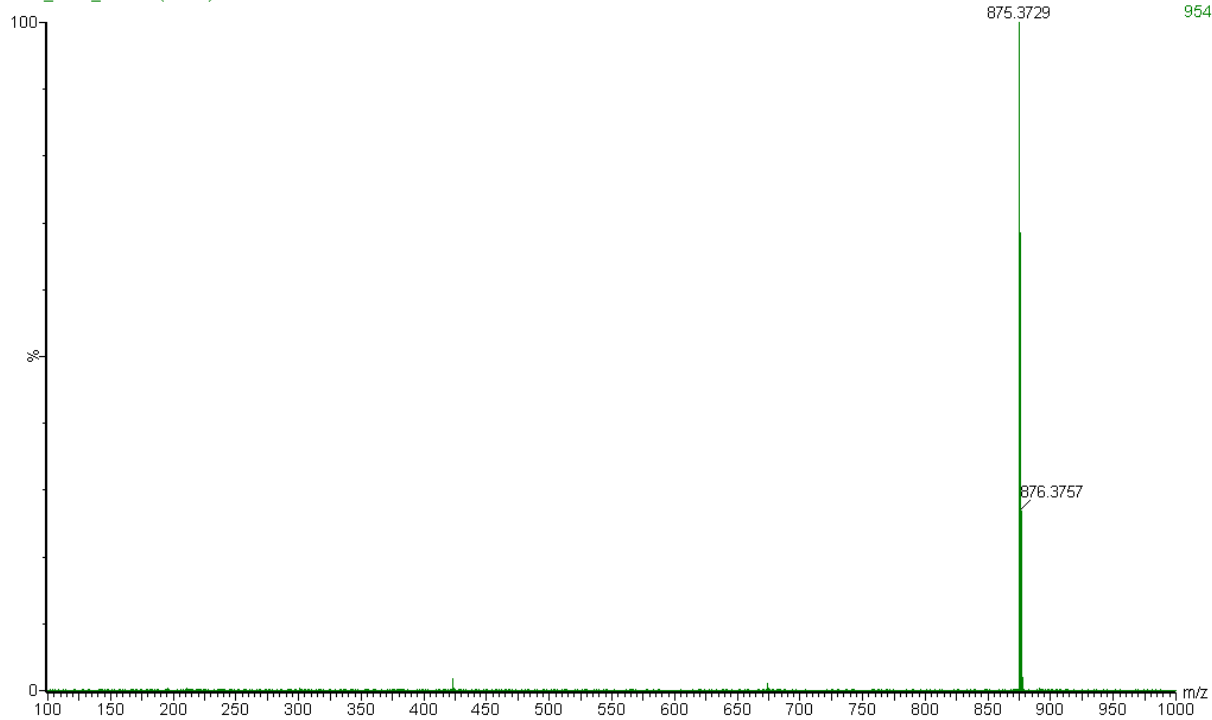
2765\_2013\_A2 8 (0.120) Cm (7:8)



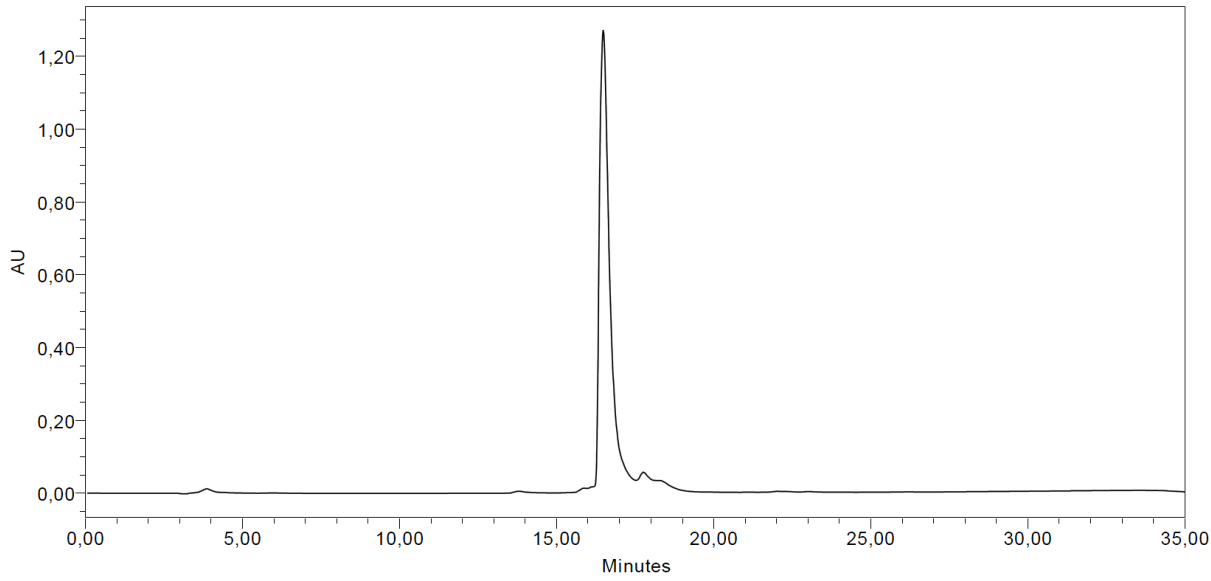
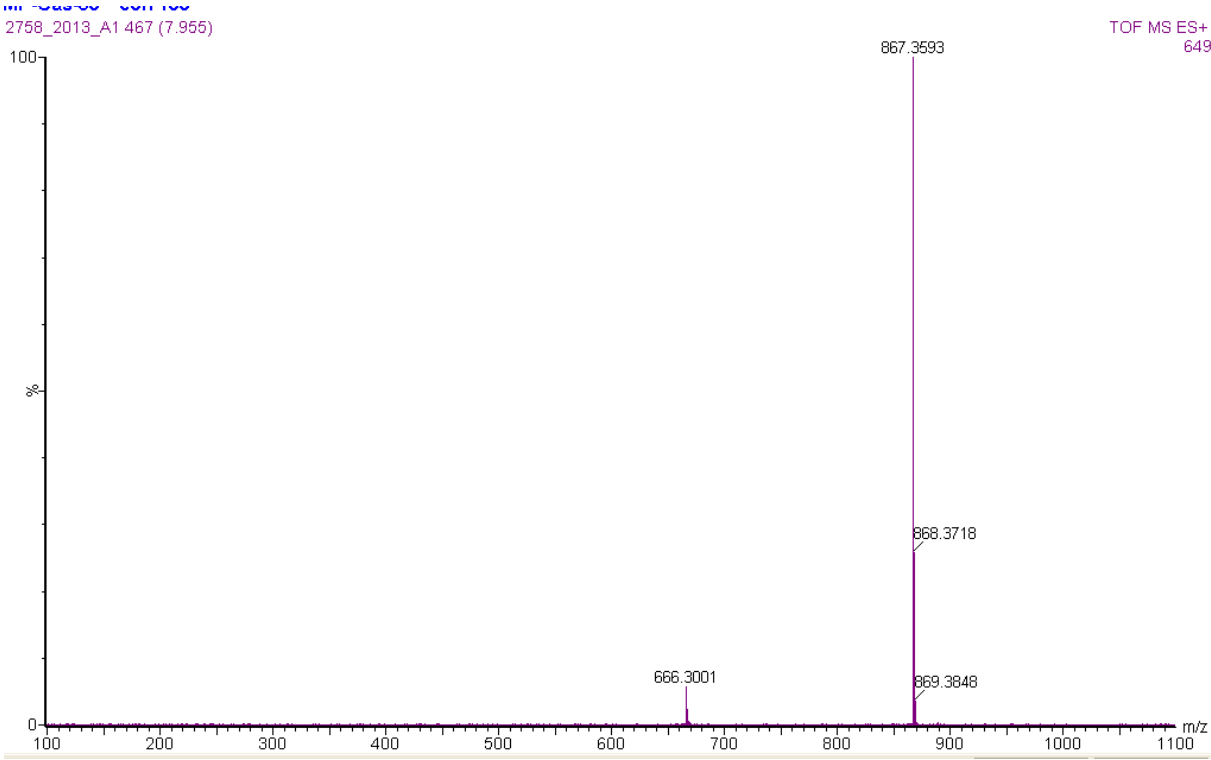
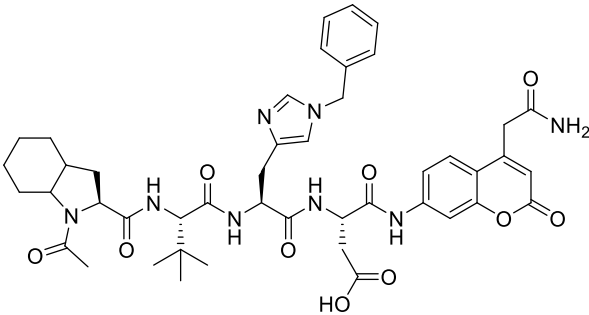
# MPP10



3482\_2013\_A1 101 (1.706)

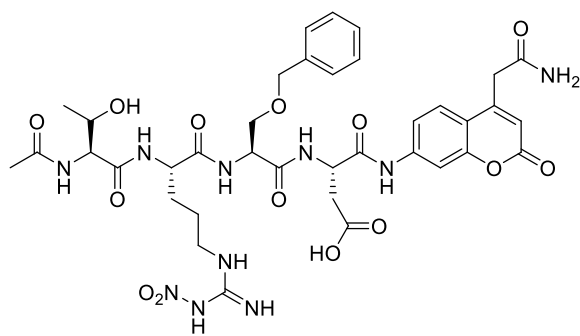


MPP12

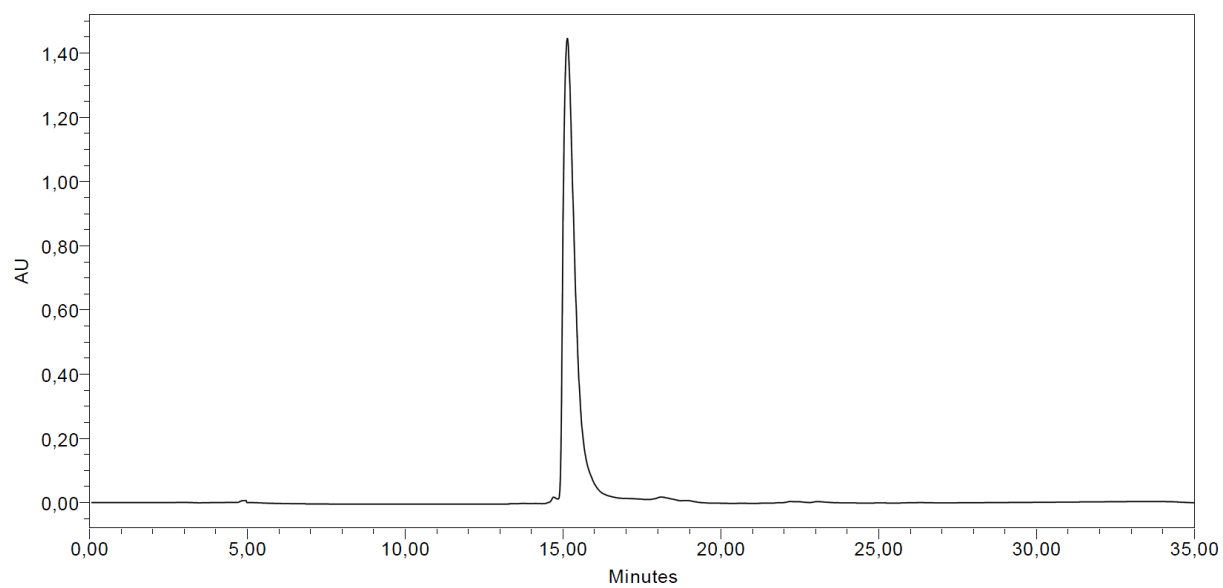
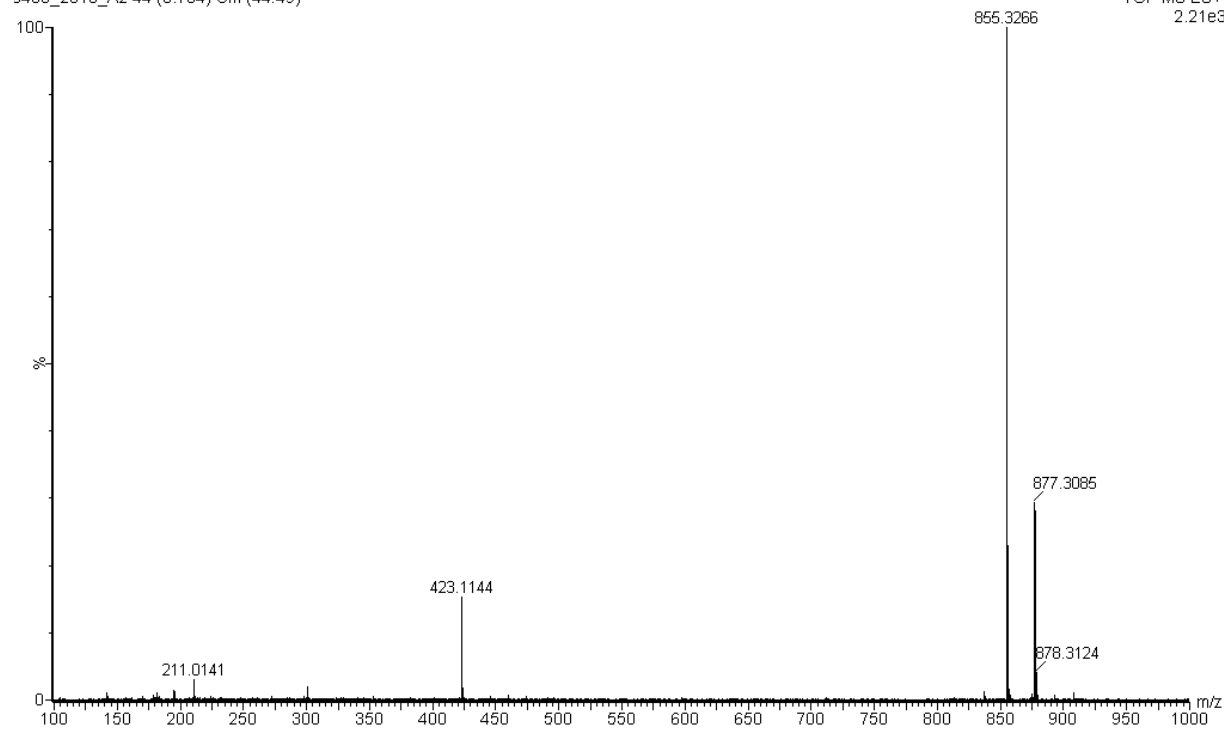




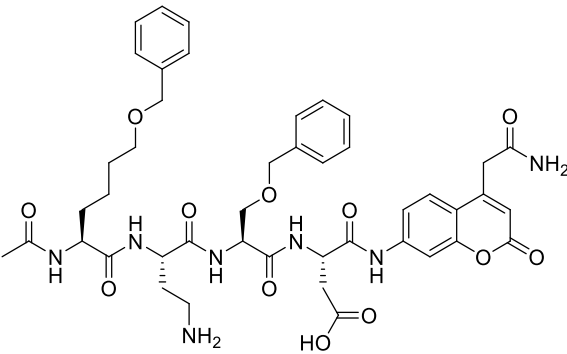
# MPP17



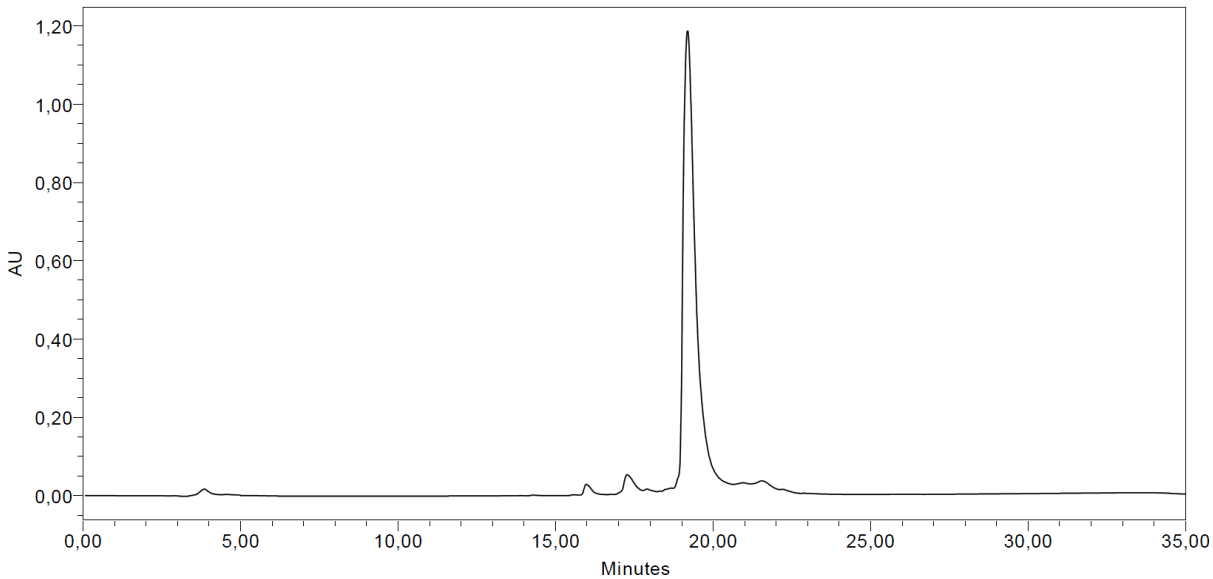
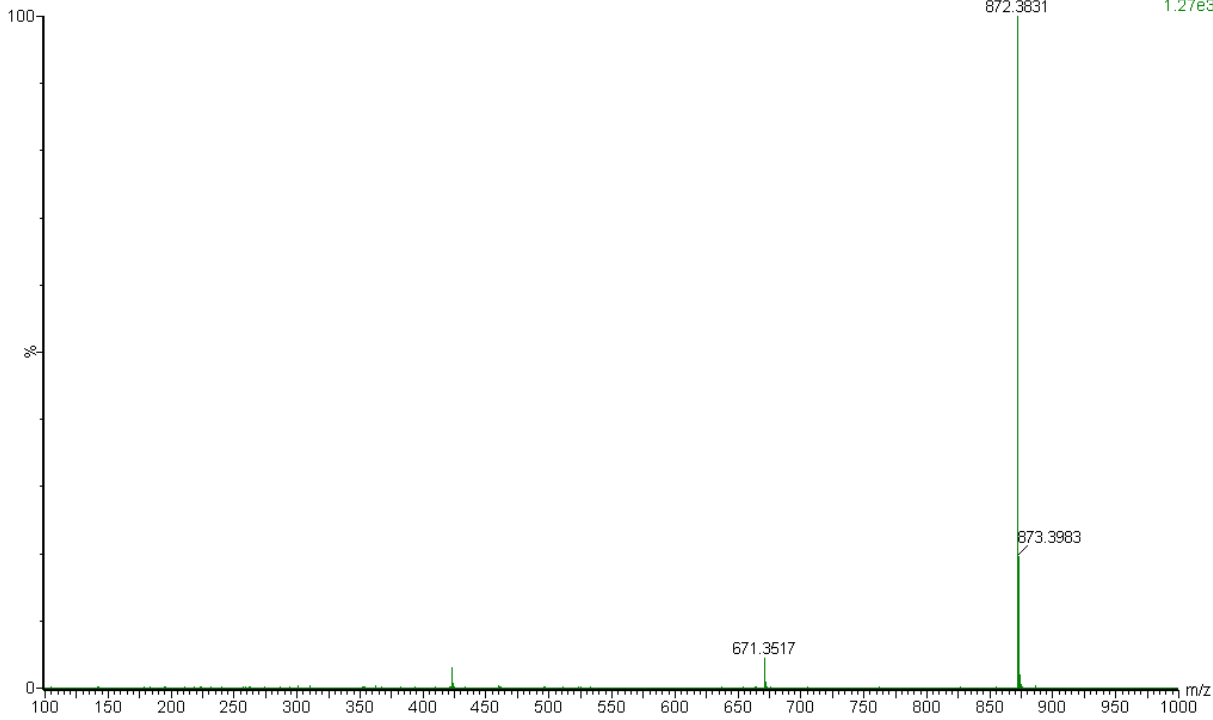
3483\_2013\_A2 44 (0.734) Cm (44:49)



MPP21

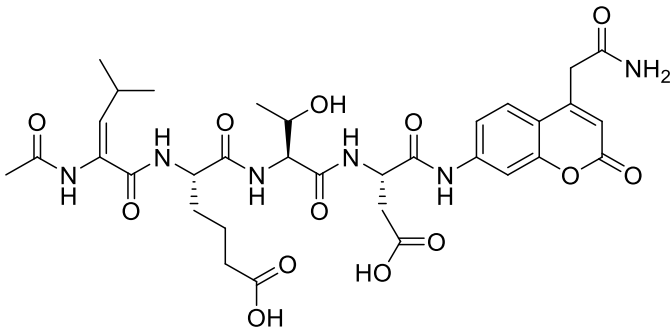


3484\_2013\_A1 64 (1.075) Cm (64:68)



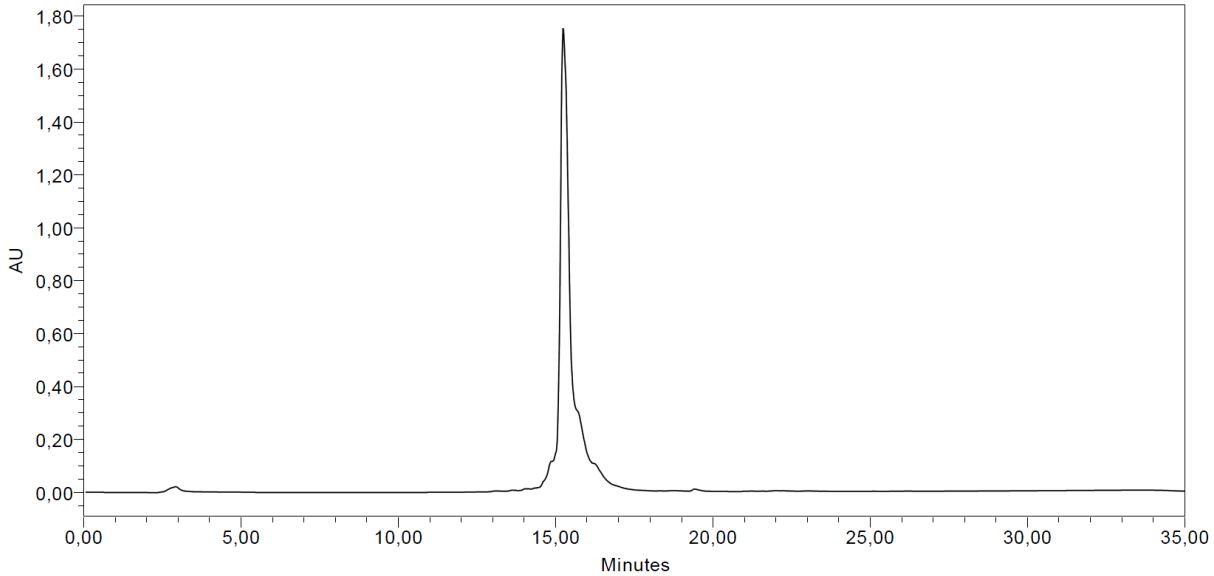
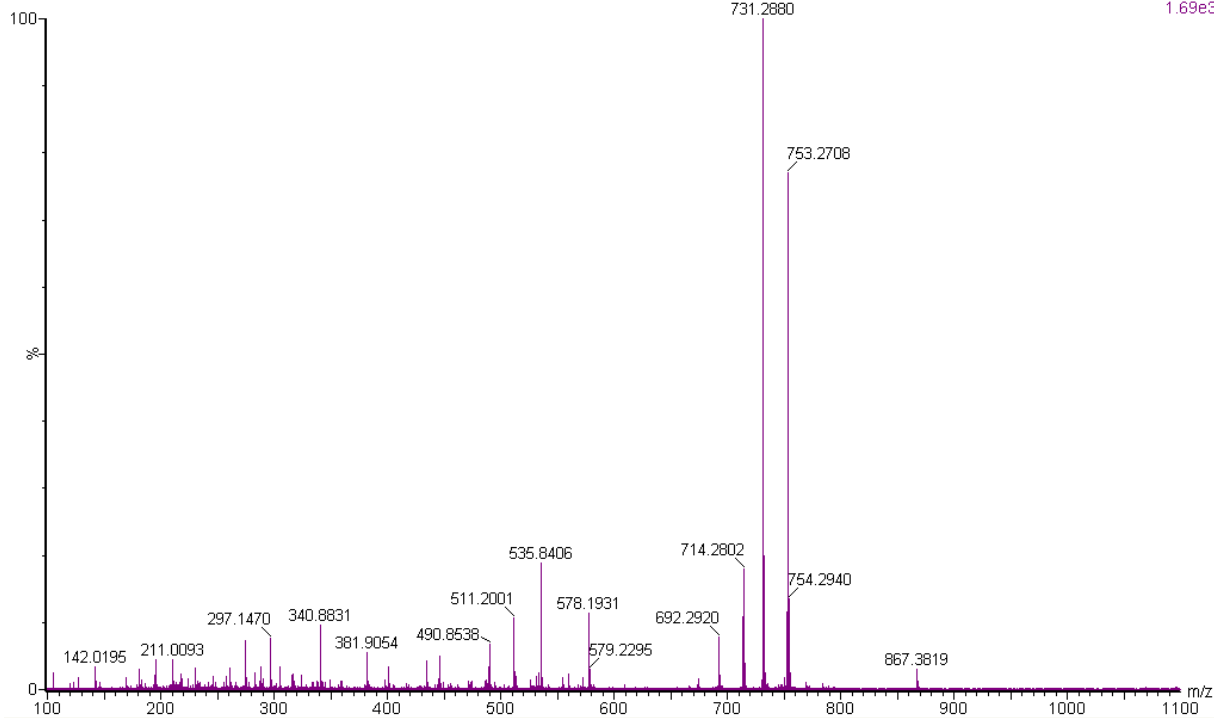


MPP28

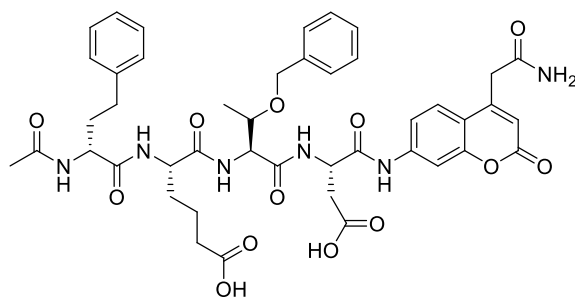


2759\_2013\_A2 15 (0.239) Cm (10:15)

TOF MS ES+  
1.69e3

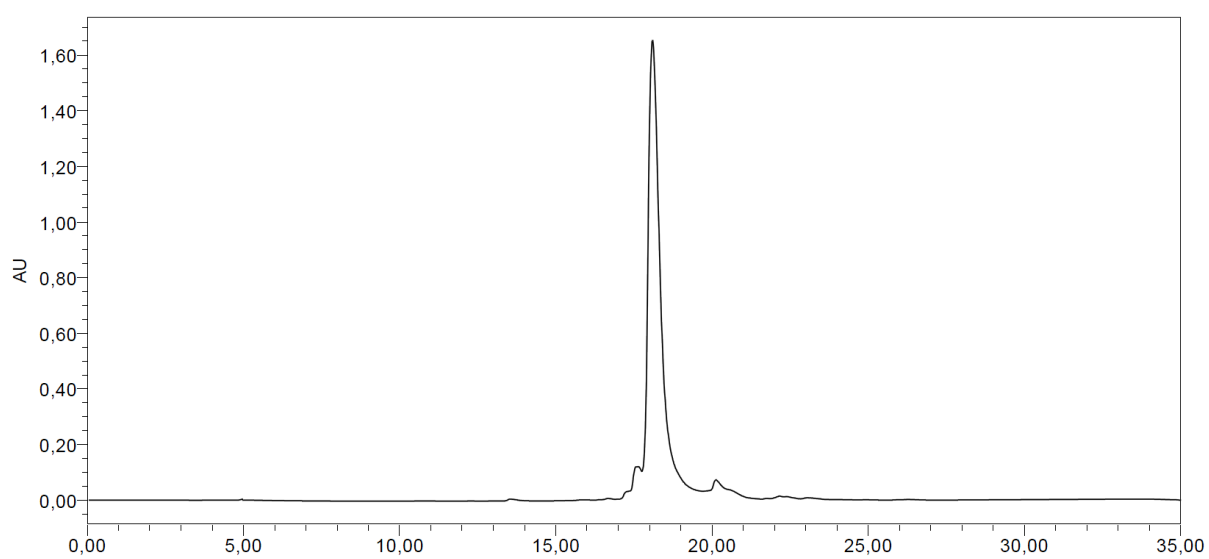
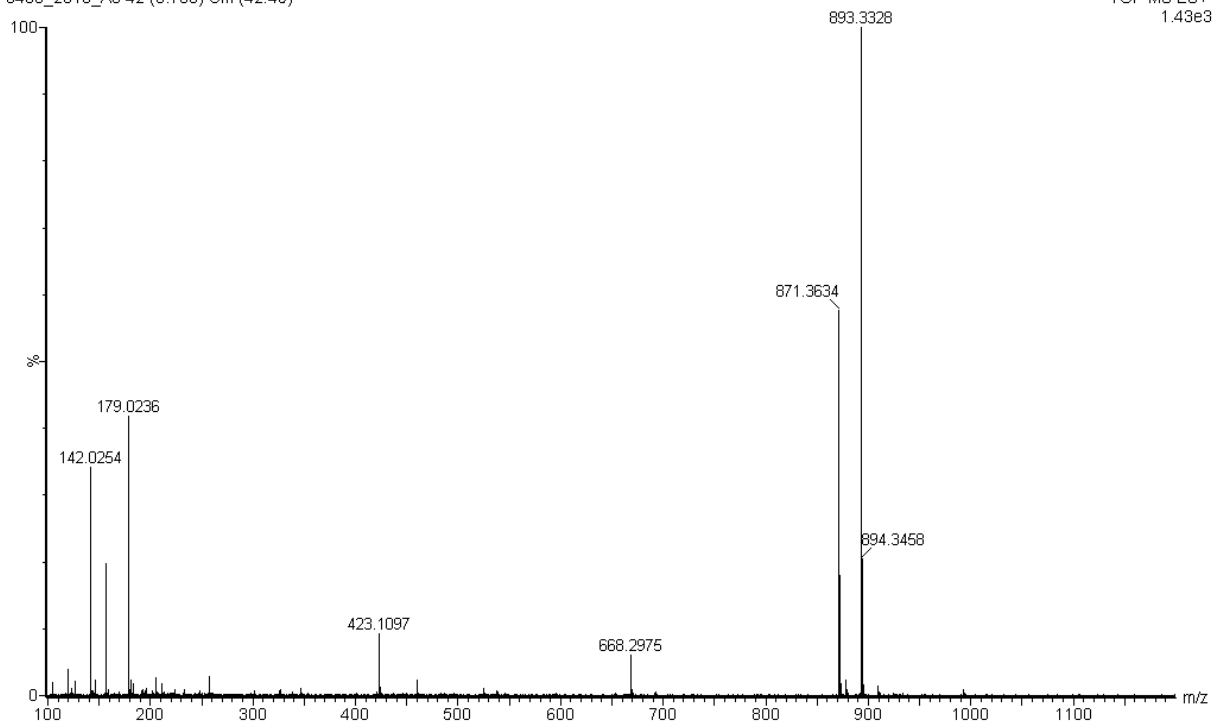


# MPP30

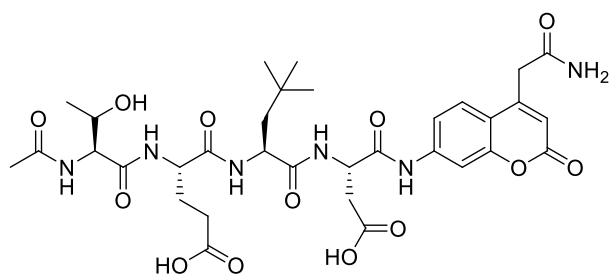


3486\_2013\_A3 42 (0.700) Cm (42:45)

TOF MS ES+  
1.43e3

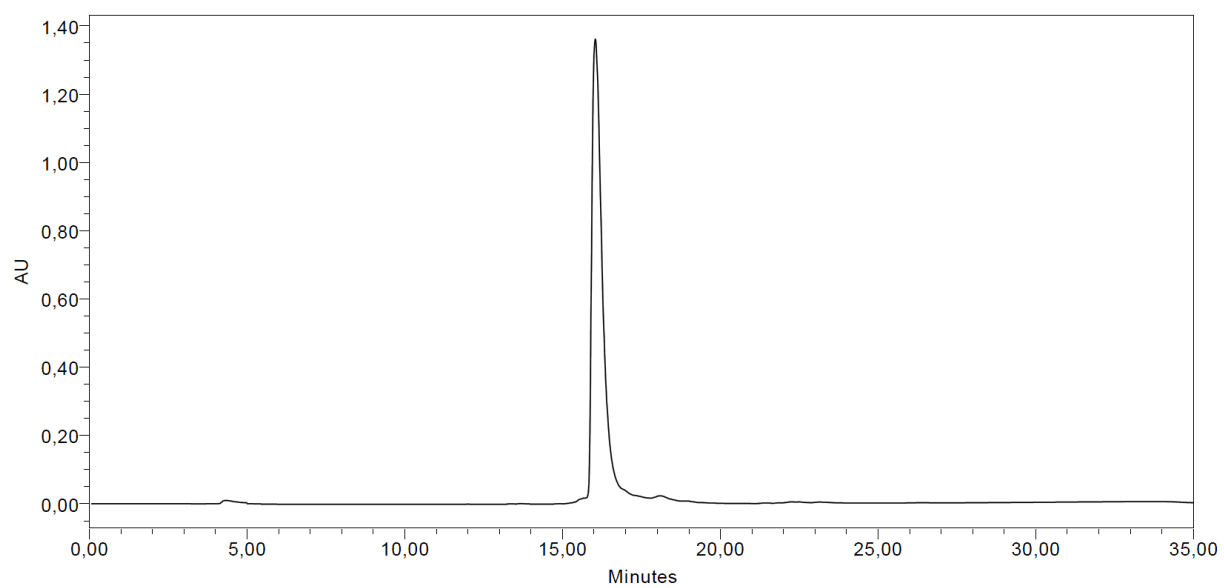
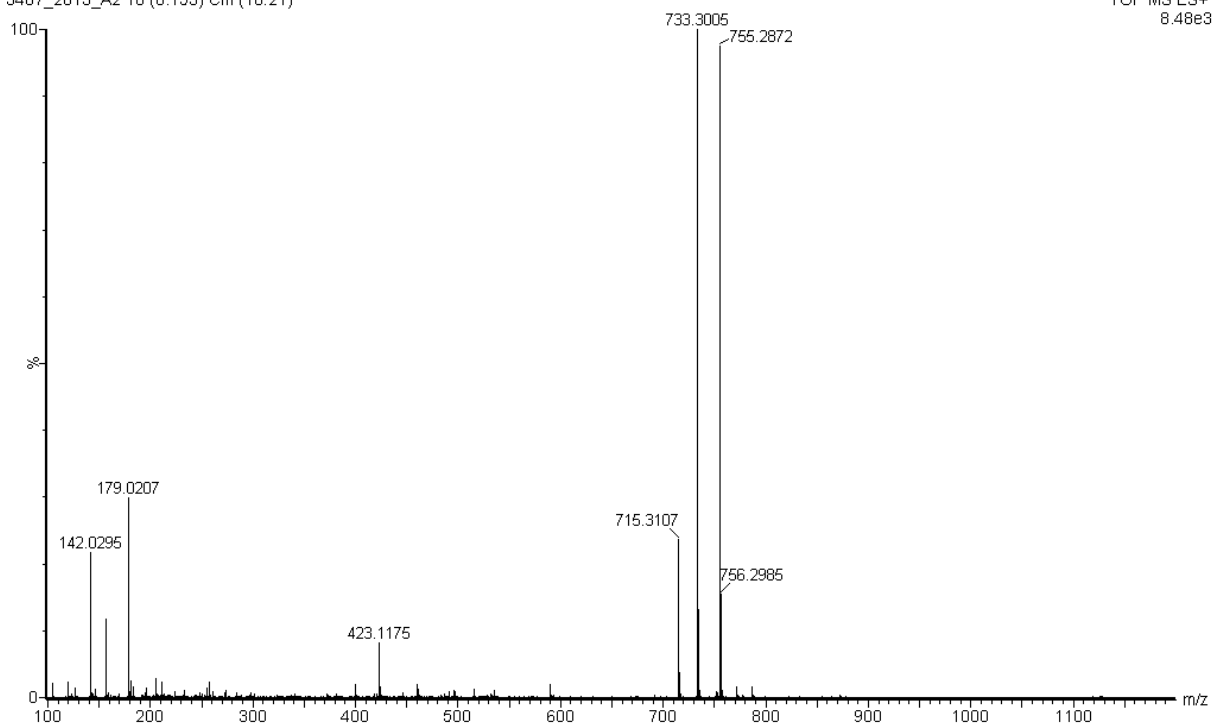


# MPP36

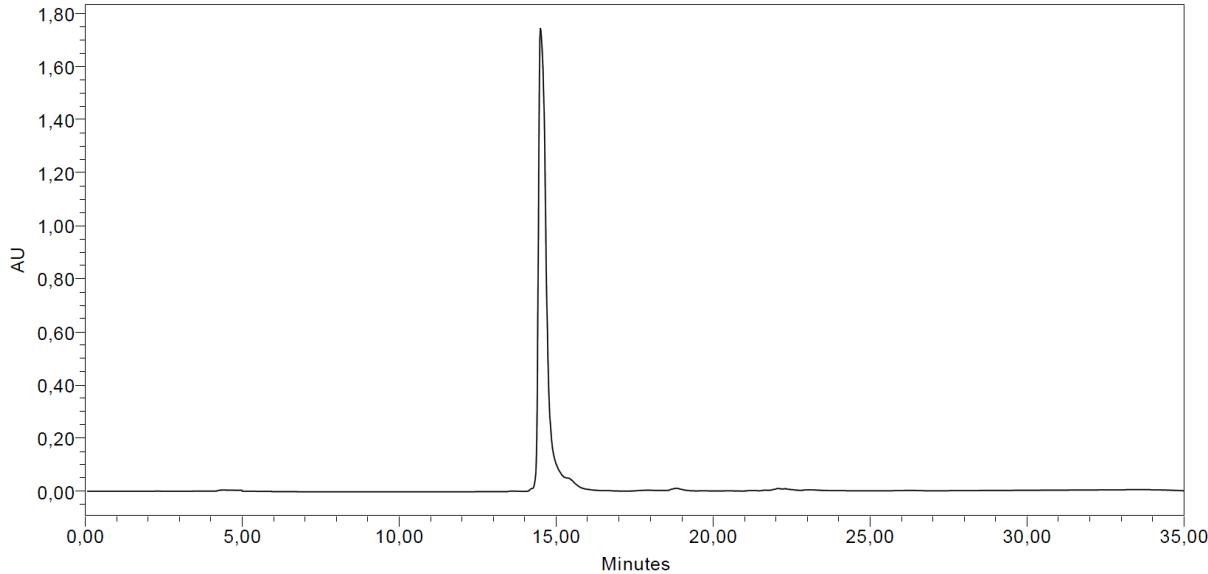
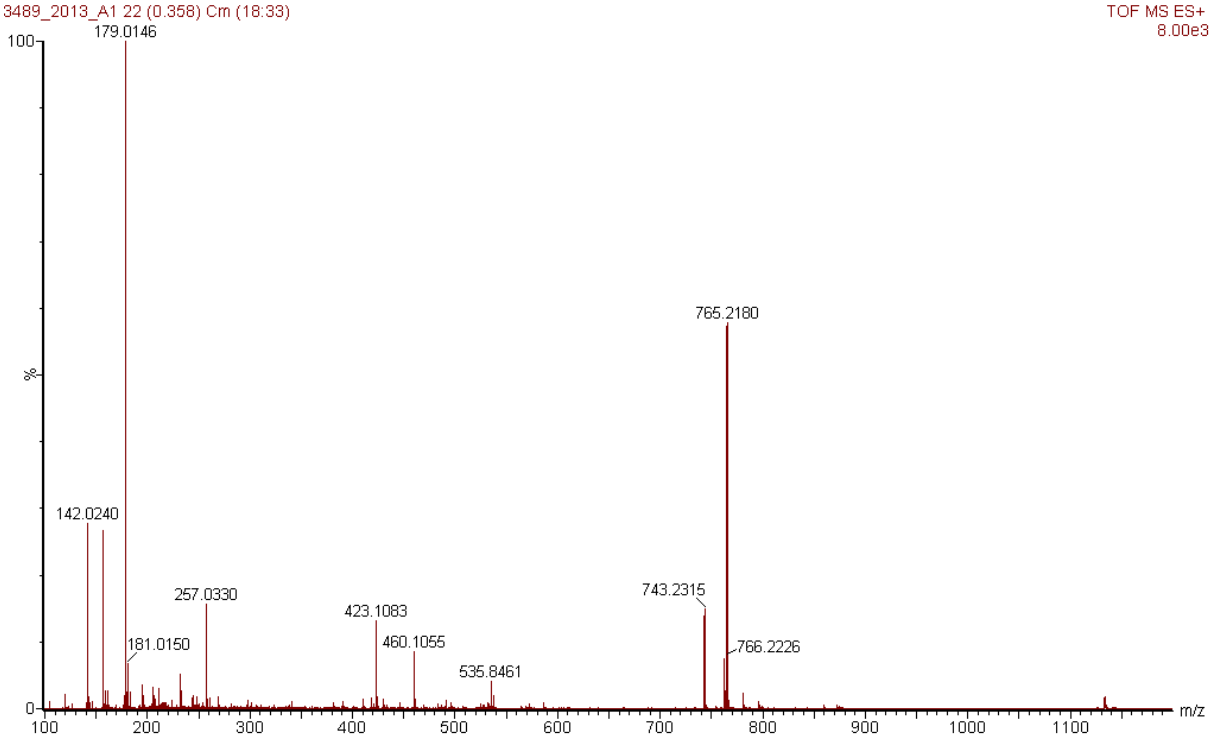
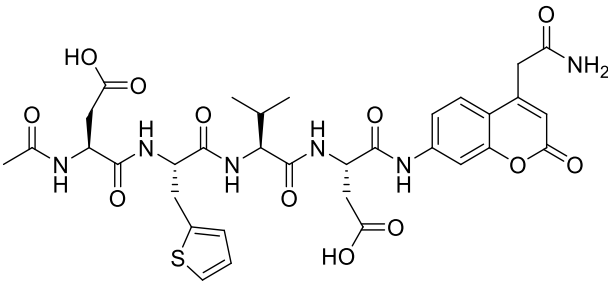


3487\_2013\_A2 10 (0.153) Cm (10:21)

TOF MS ES+  
8.48e3



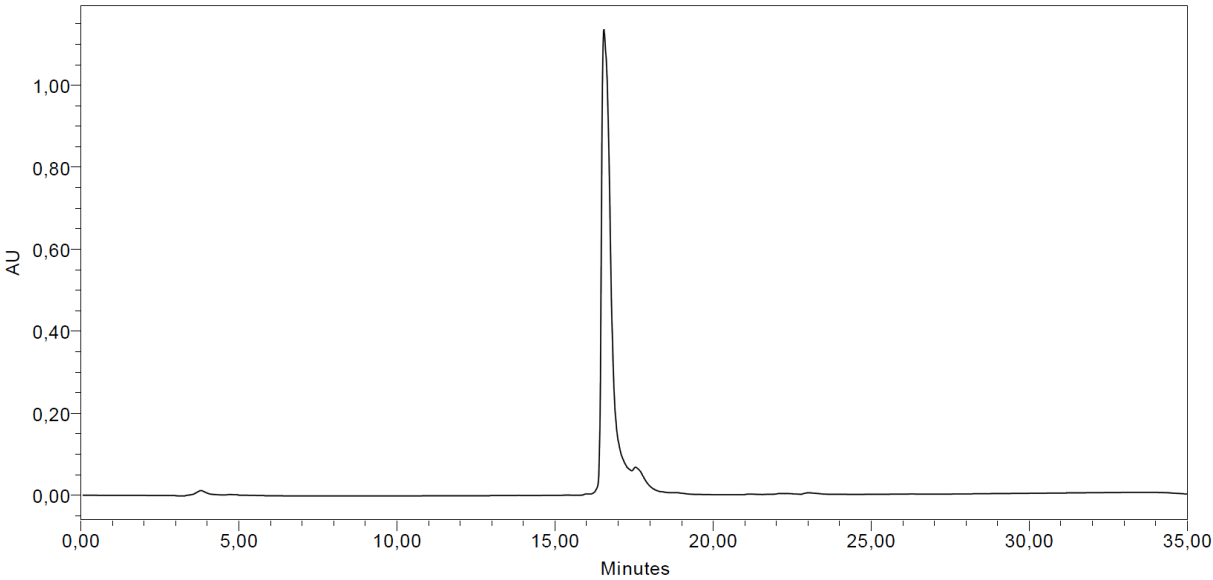
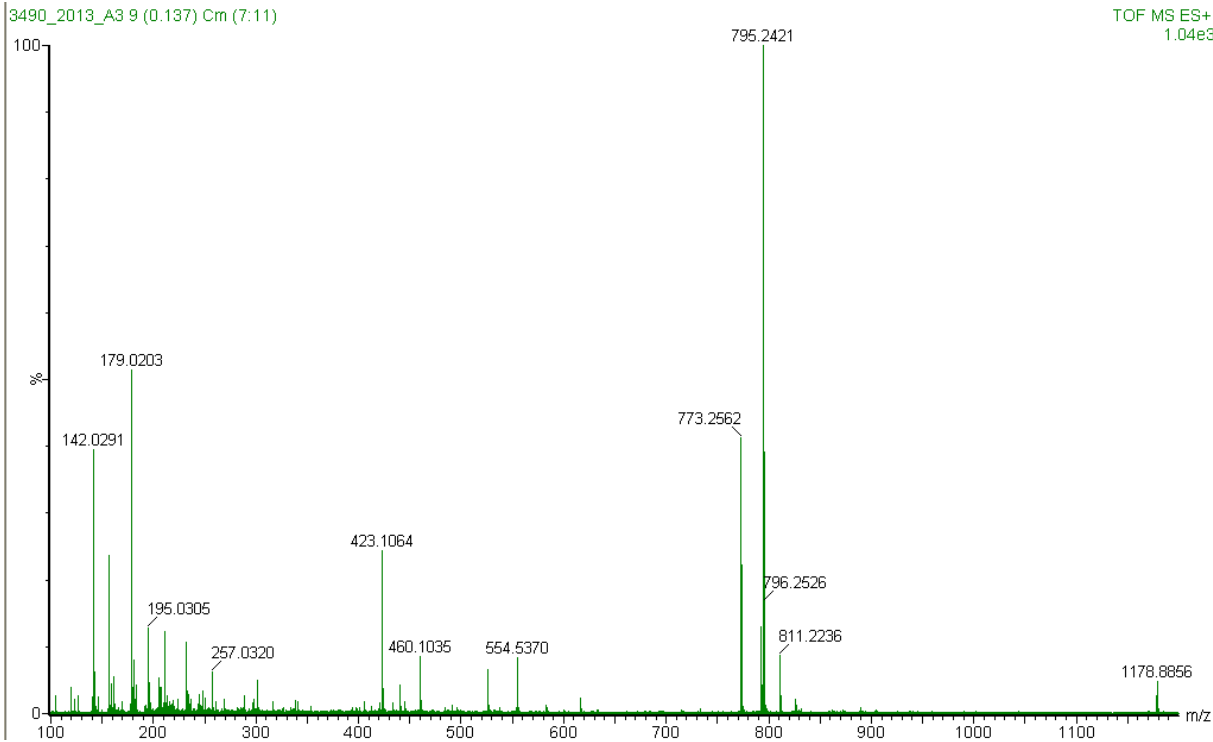
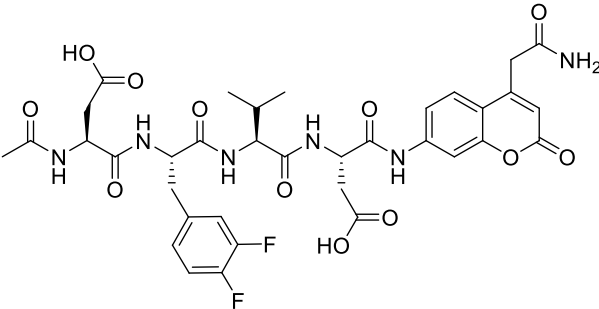
MPP38



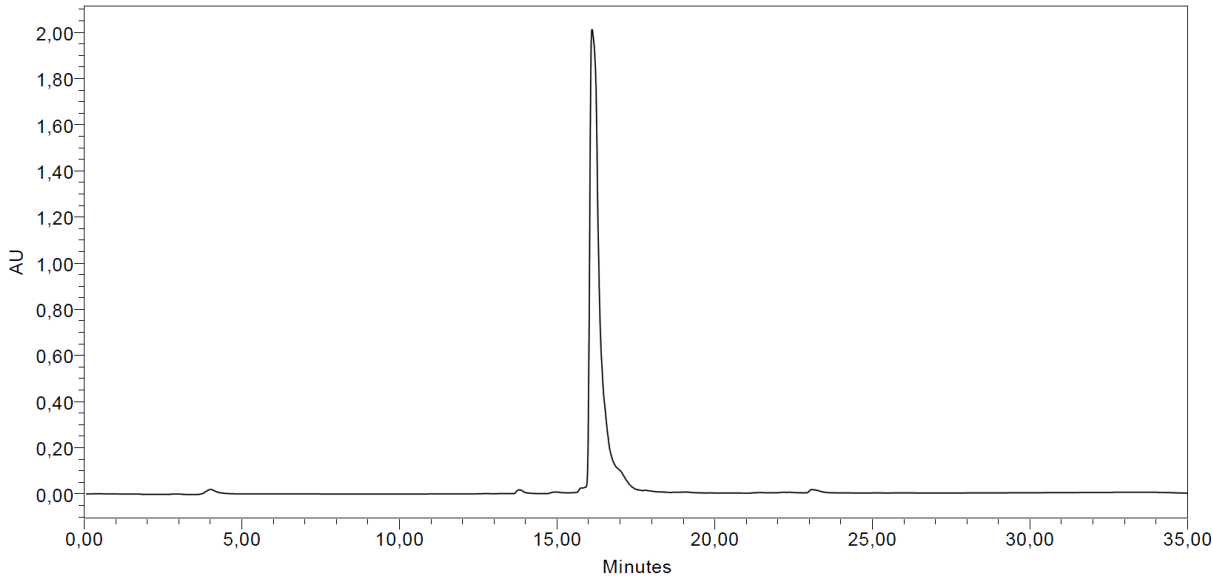
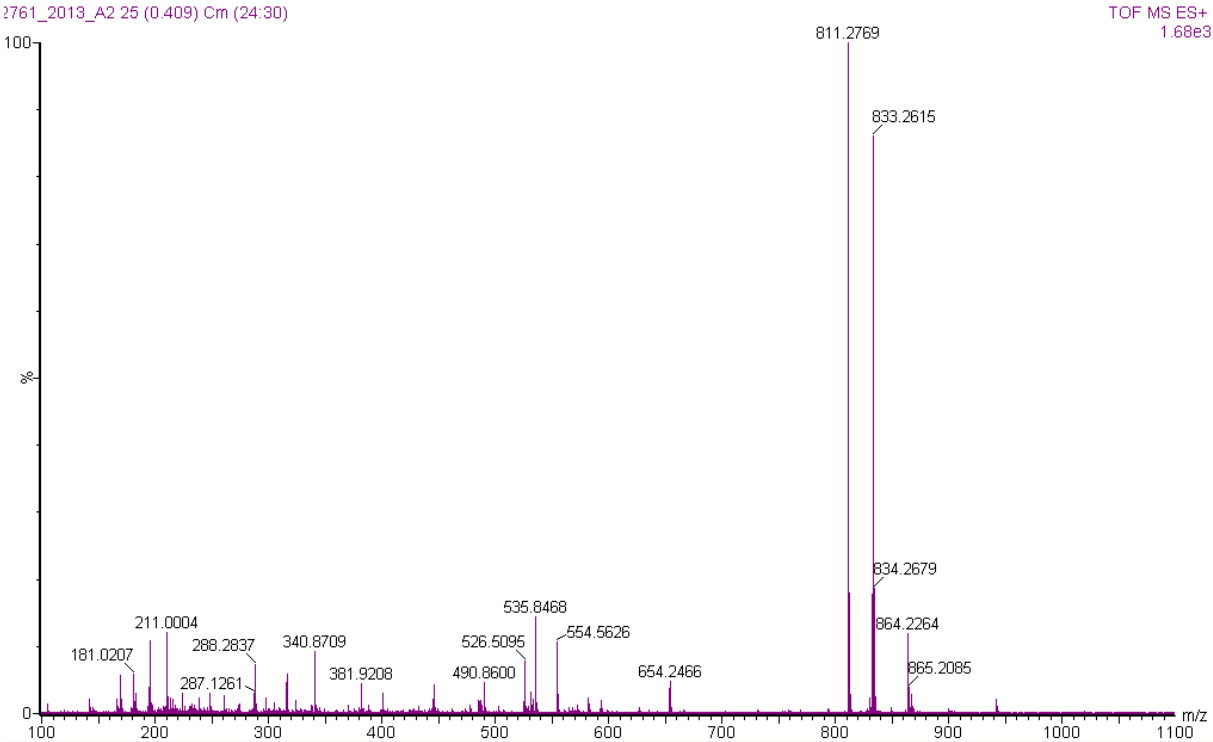
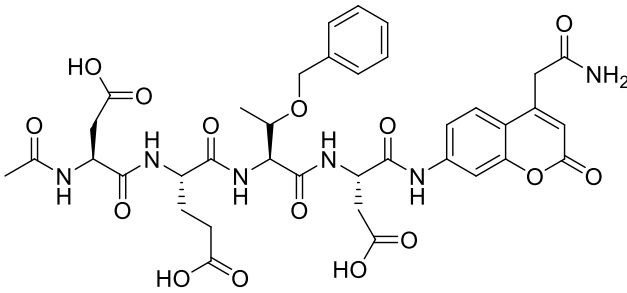




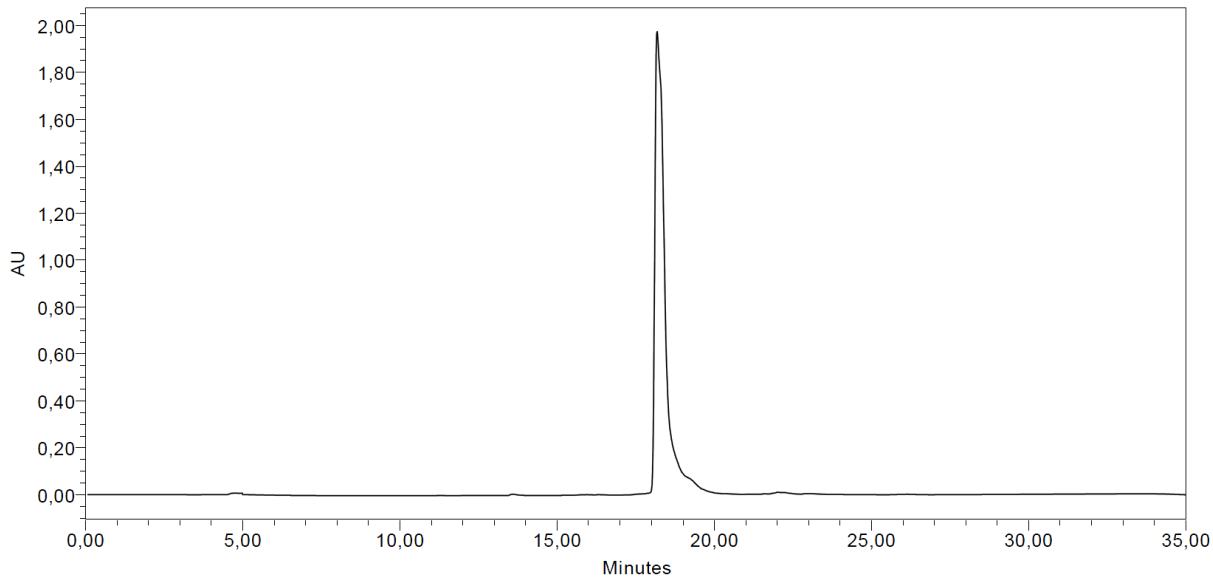
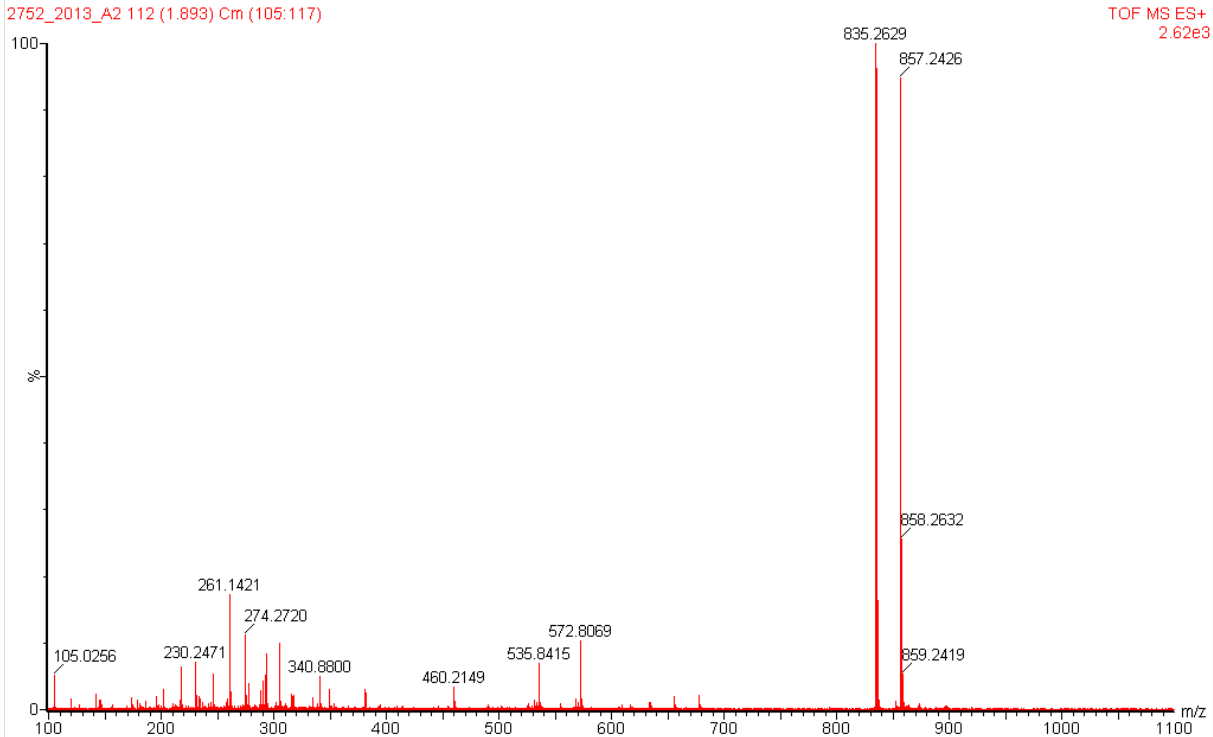
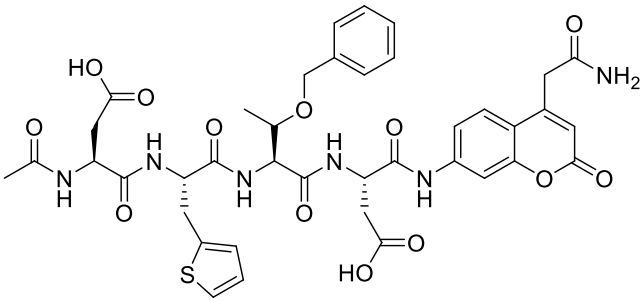
MPP40



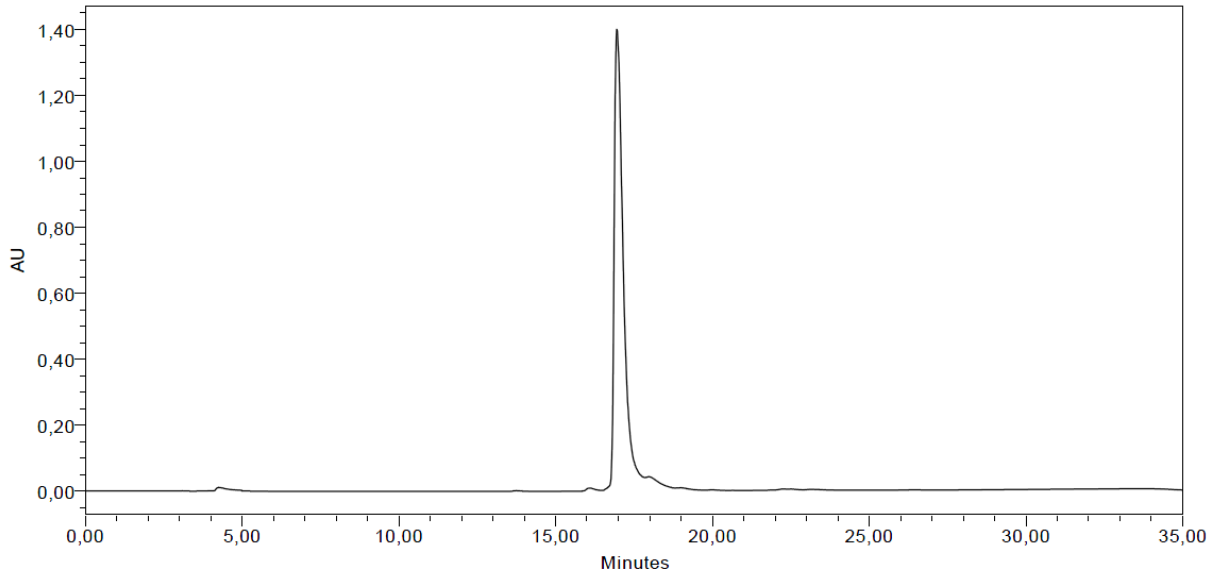
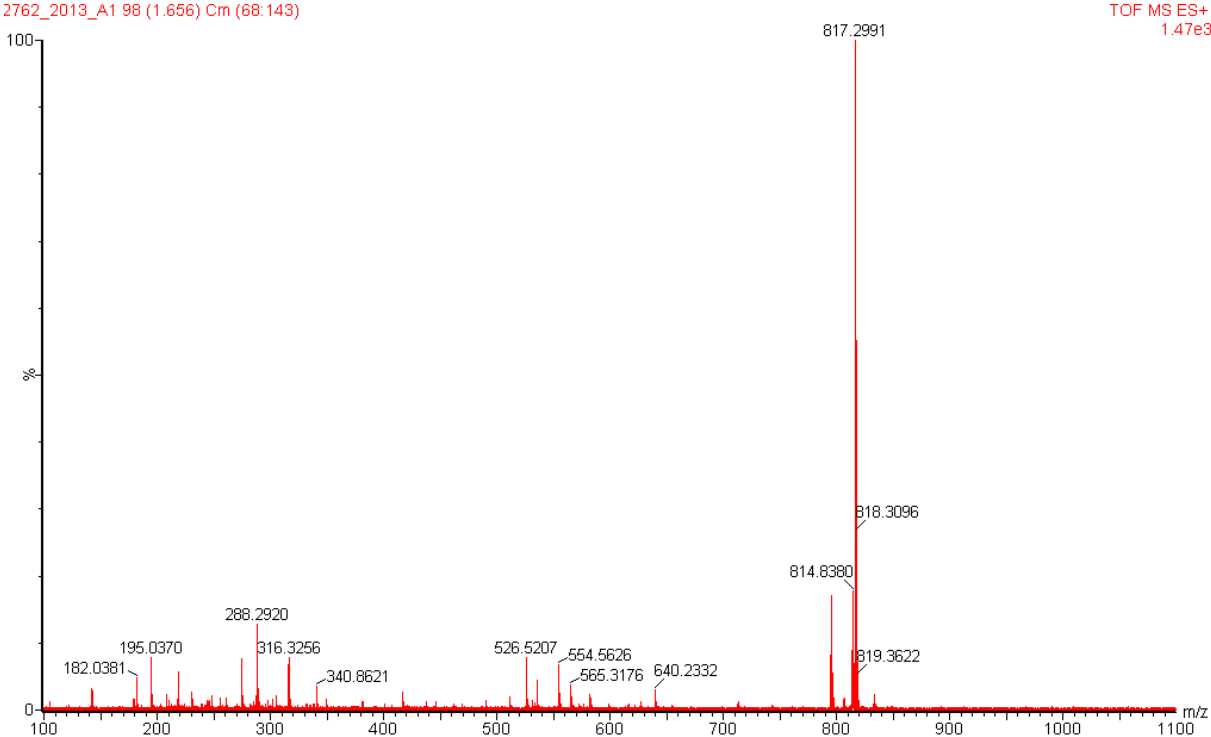
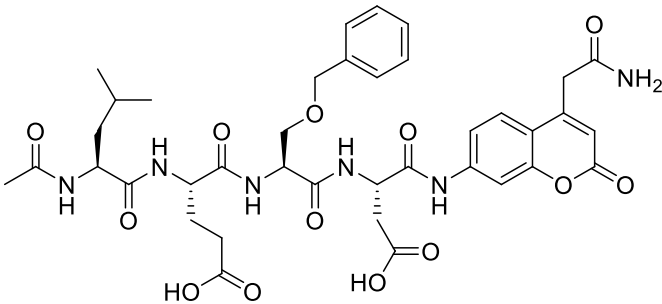
MPP41



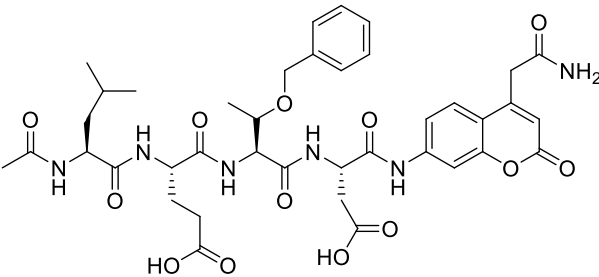
MPP42



MPP43

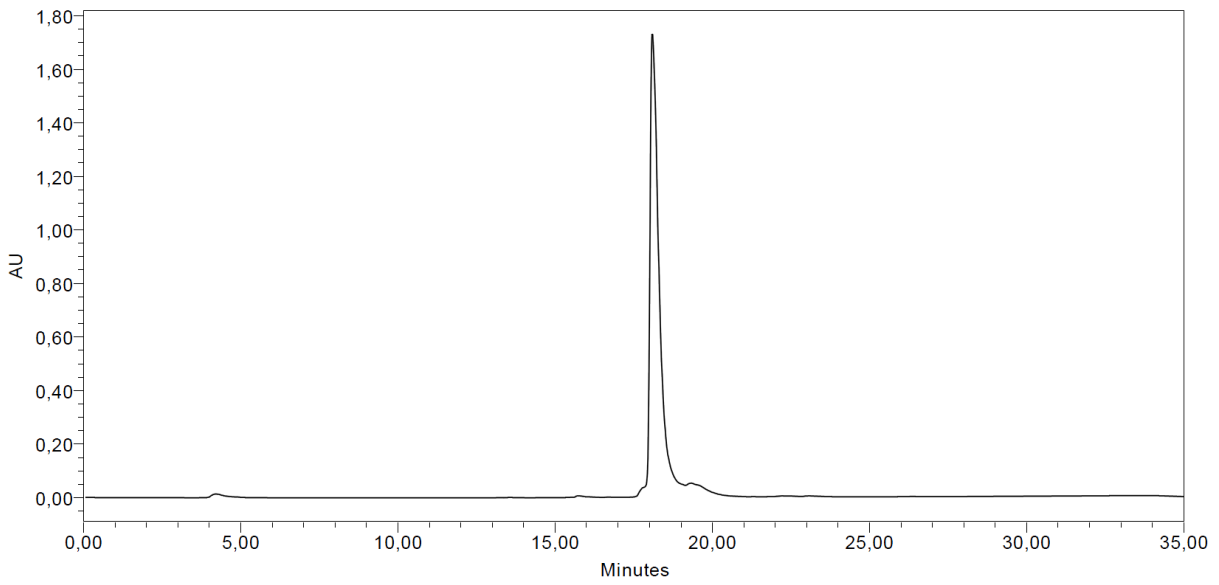
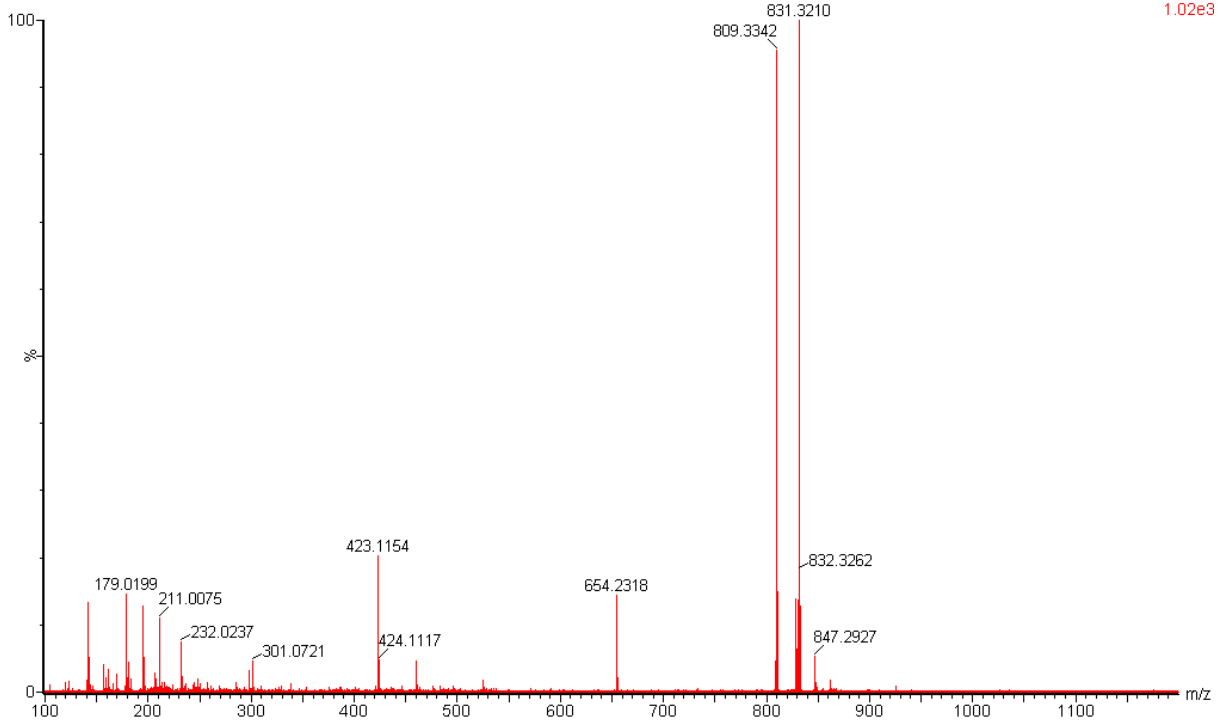


MPP45

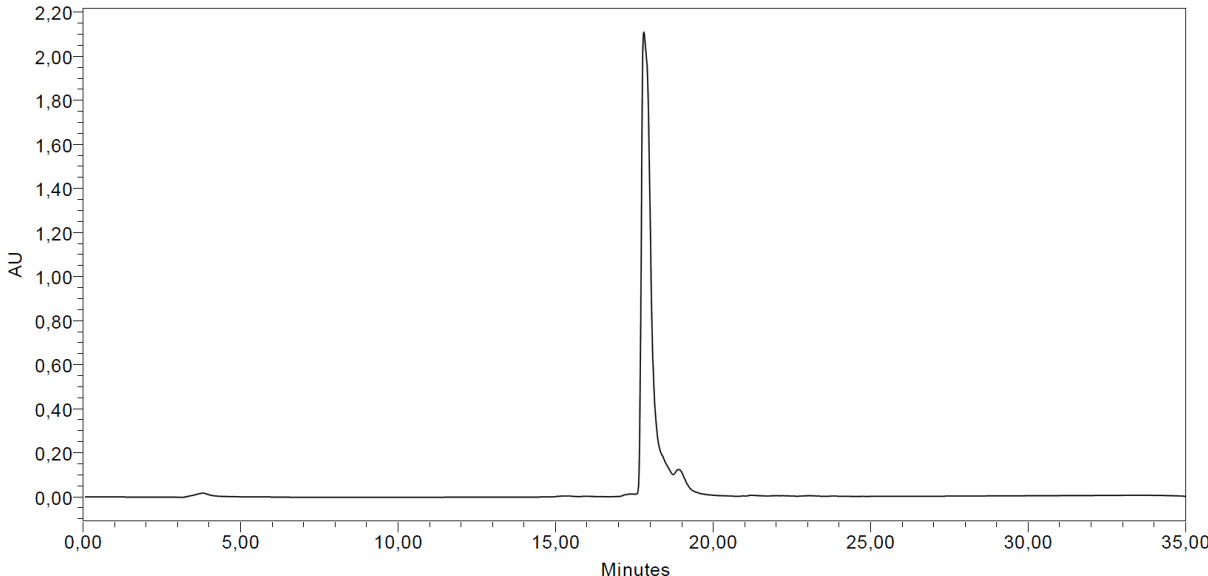
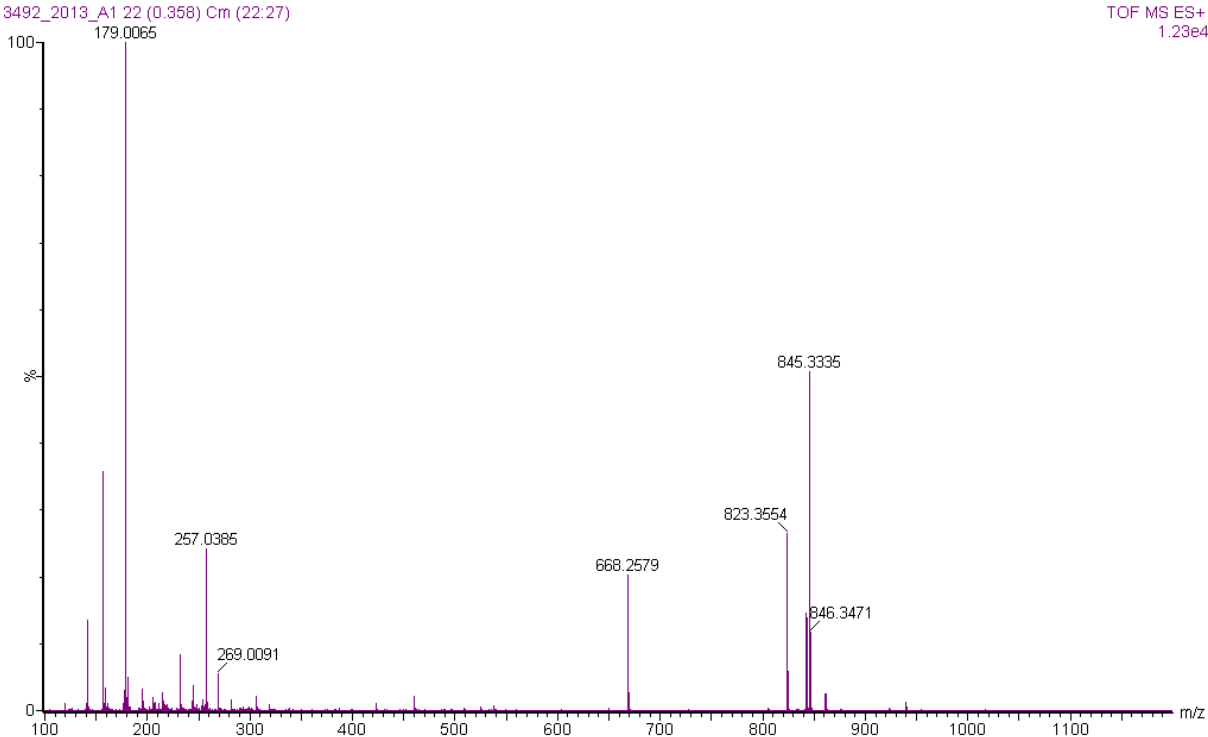
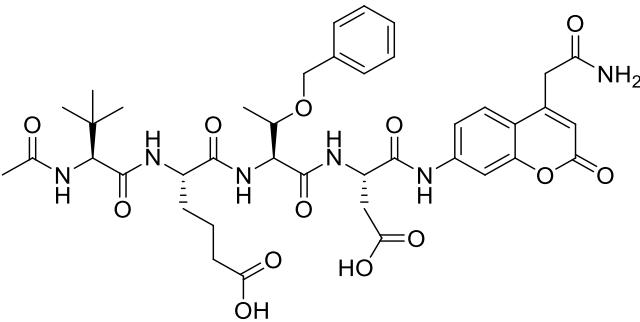


3491\_2013\_A1 71 (1.194) Cm (69.71)

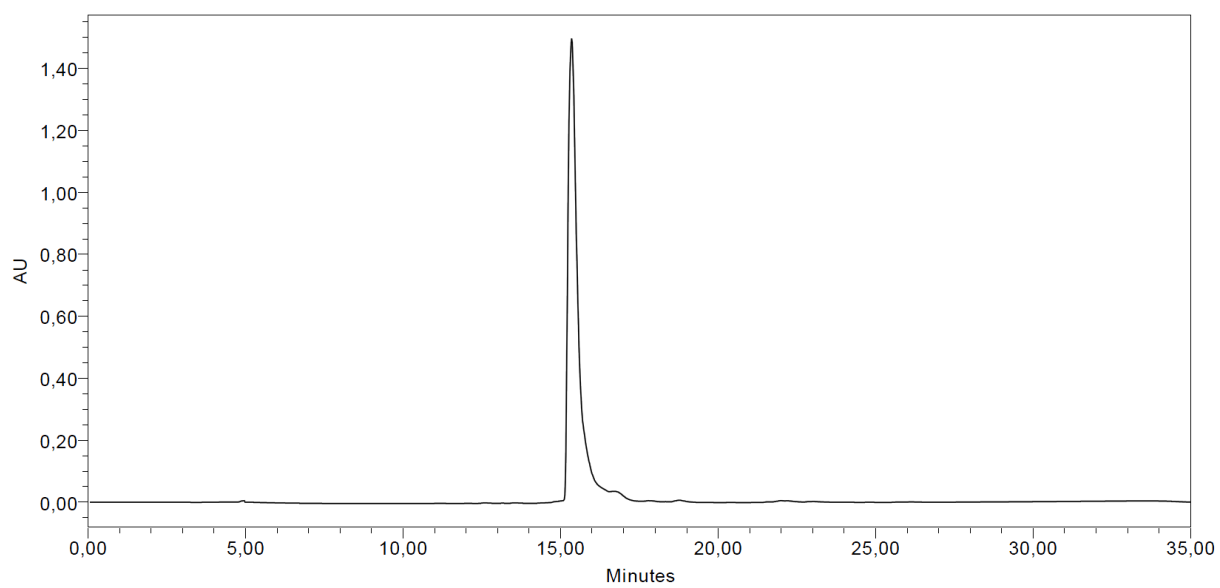
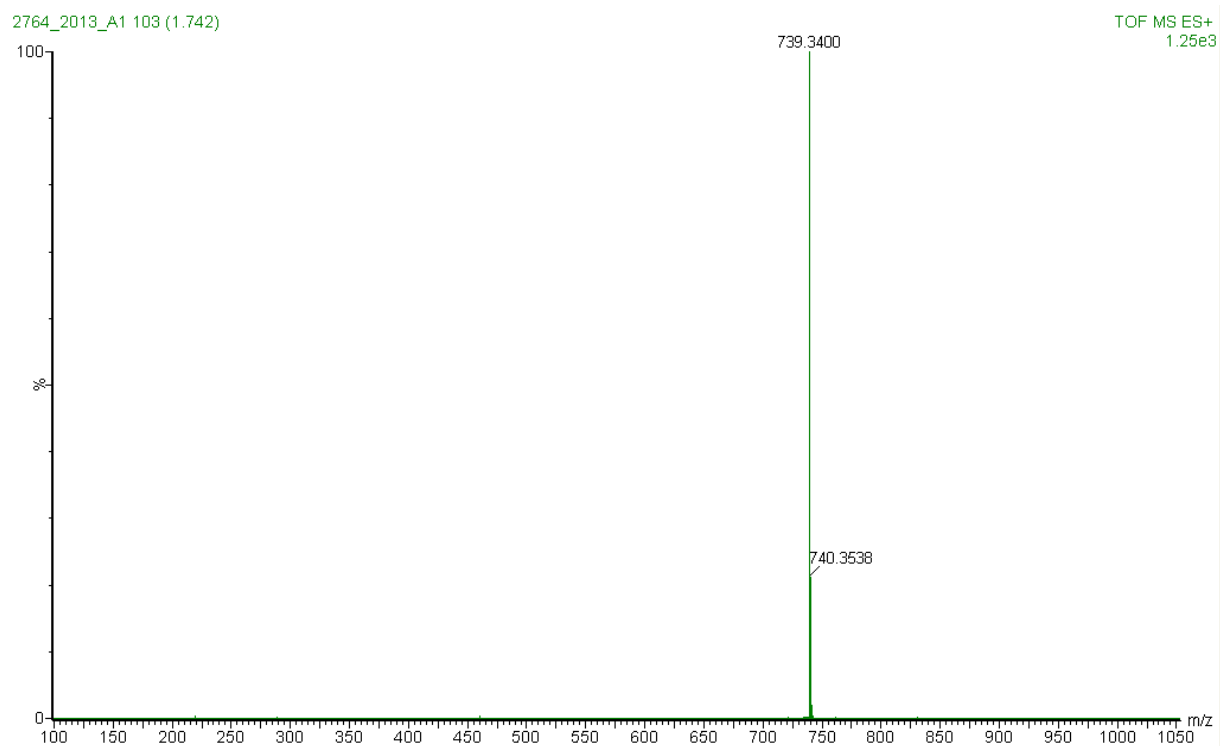
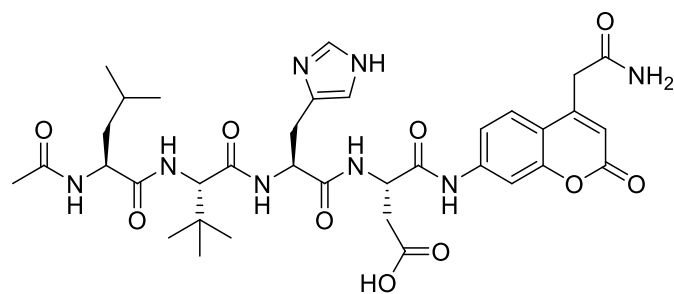
TOF MS ES+  
1.02e3



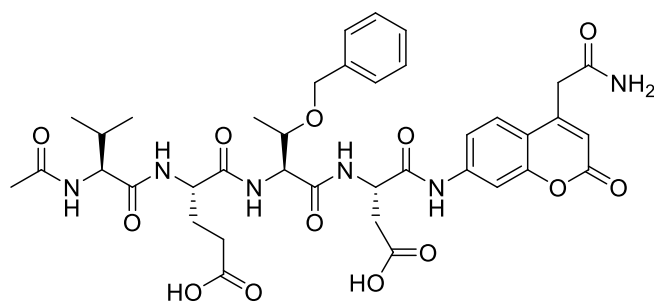
MPP46



# MPP47

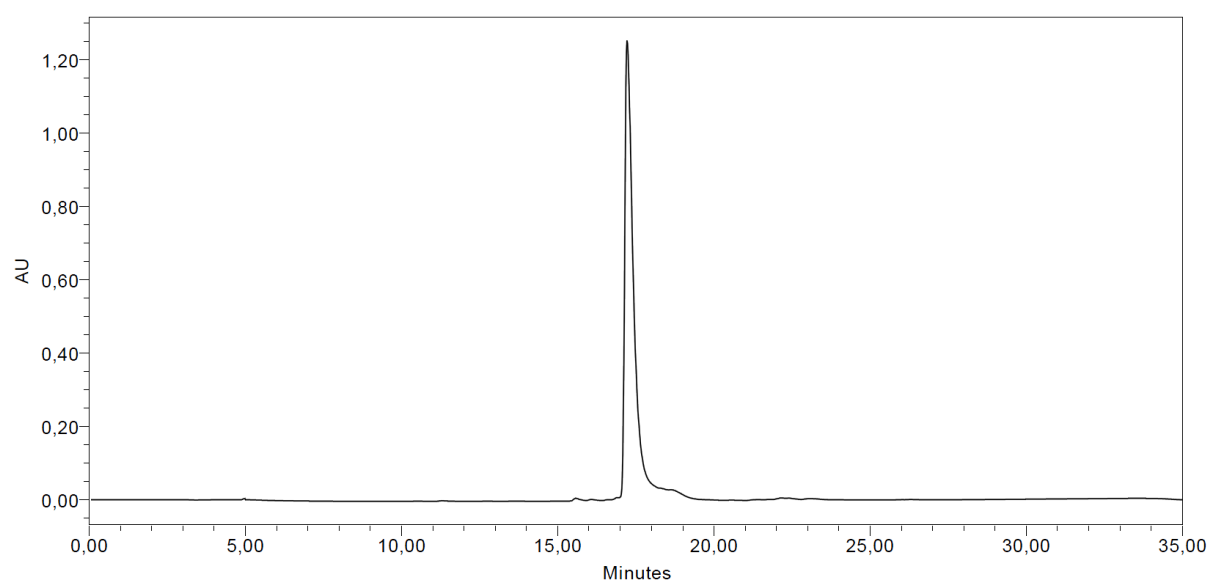
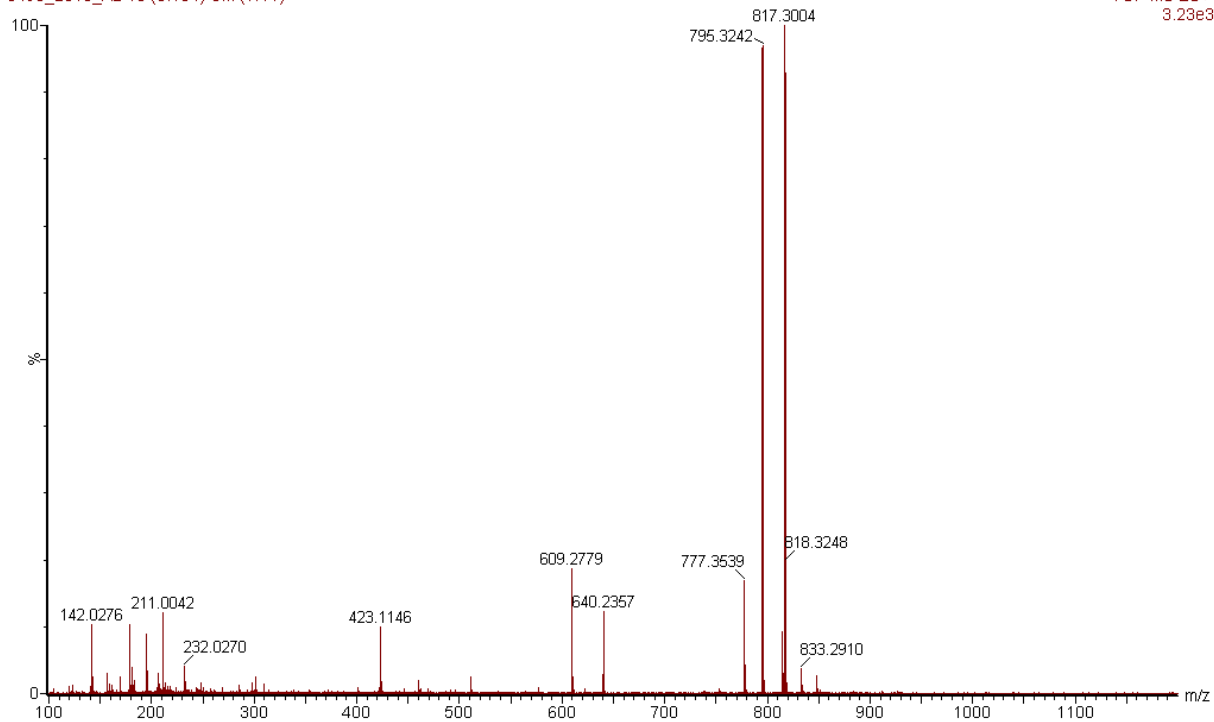


# MPP48



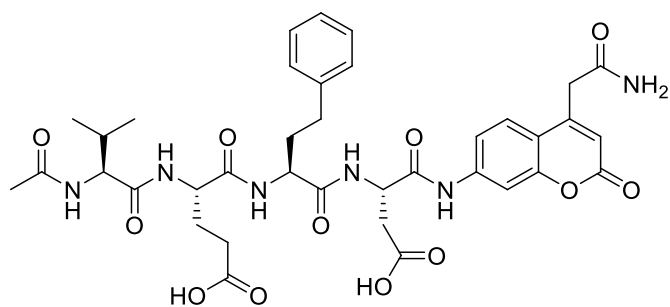
3493\_2013\_A2 10 (0.154) Cm (7:11)

TOF MS ES+  
3.23e3



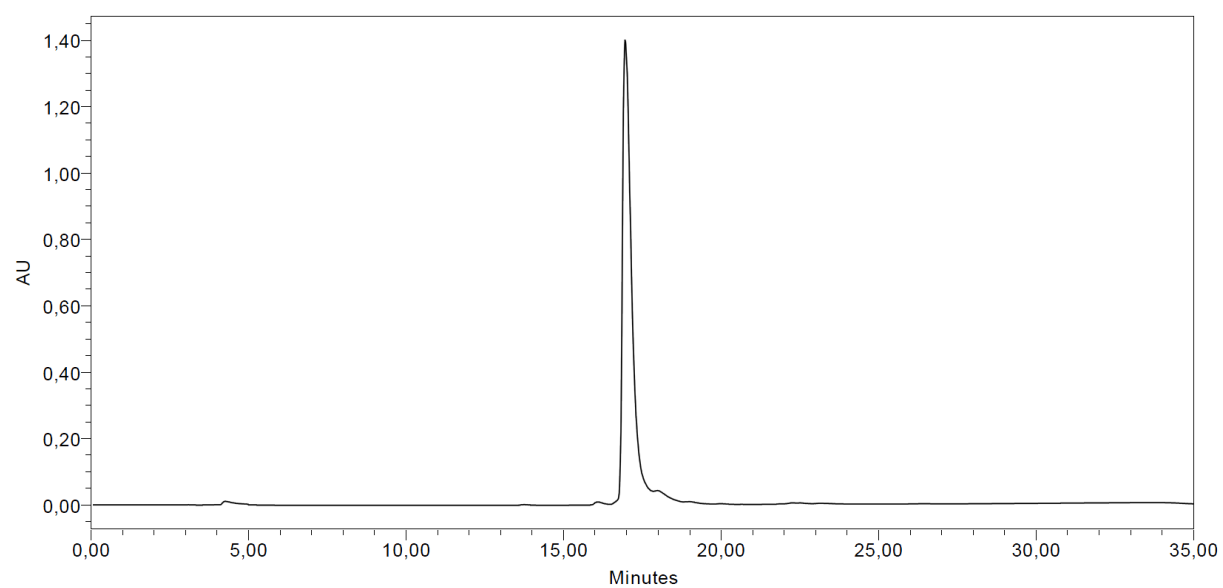
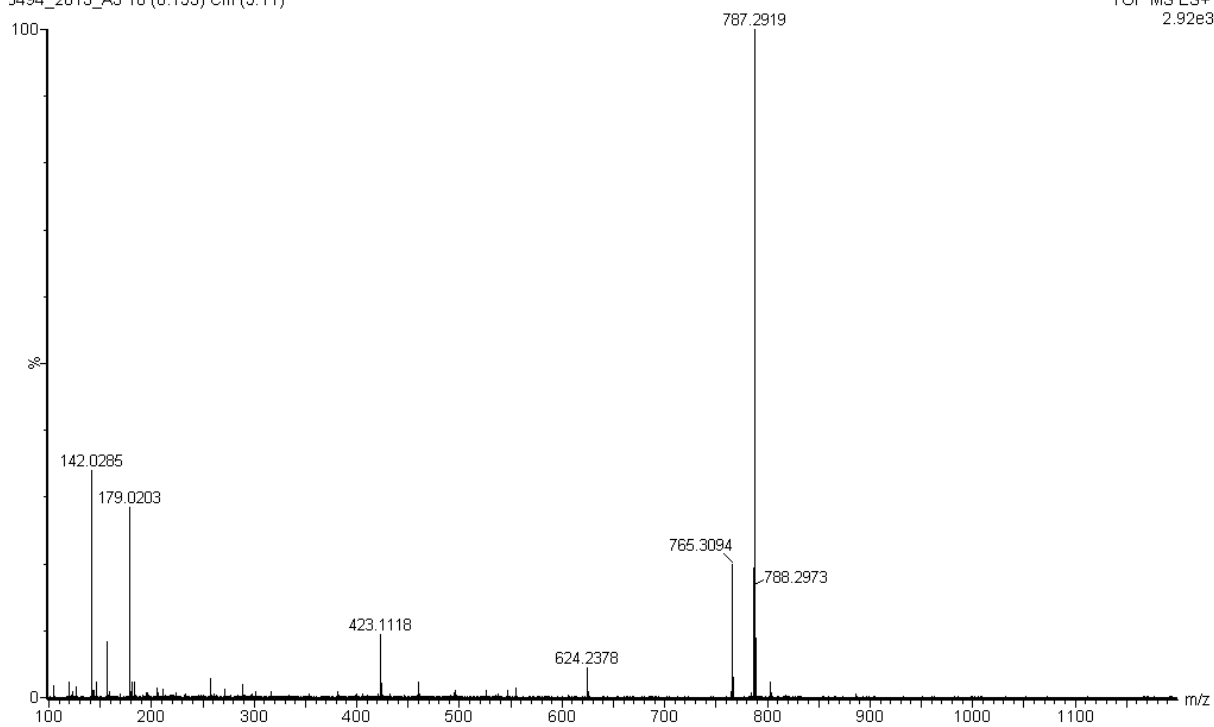


# MPP49

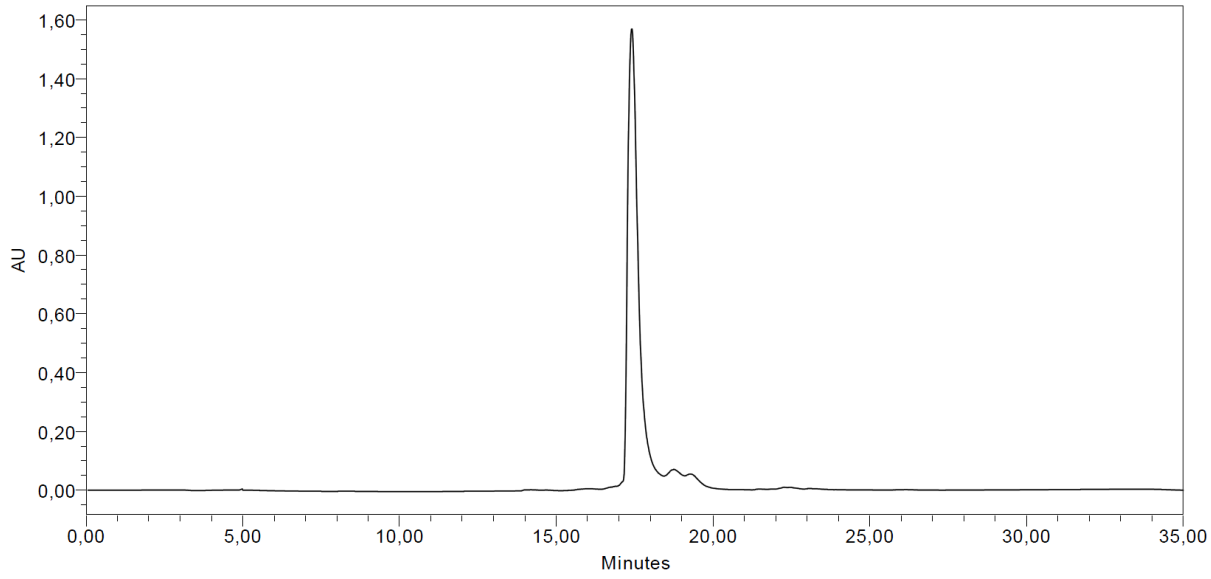
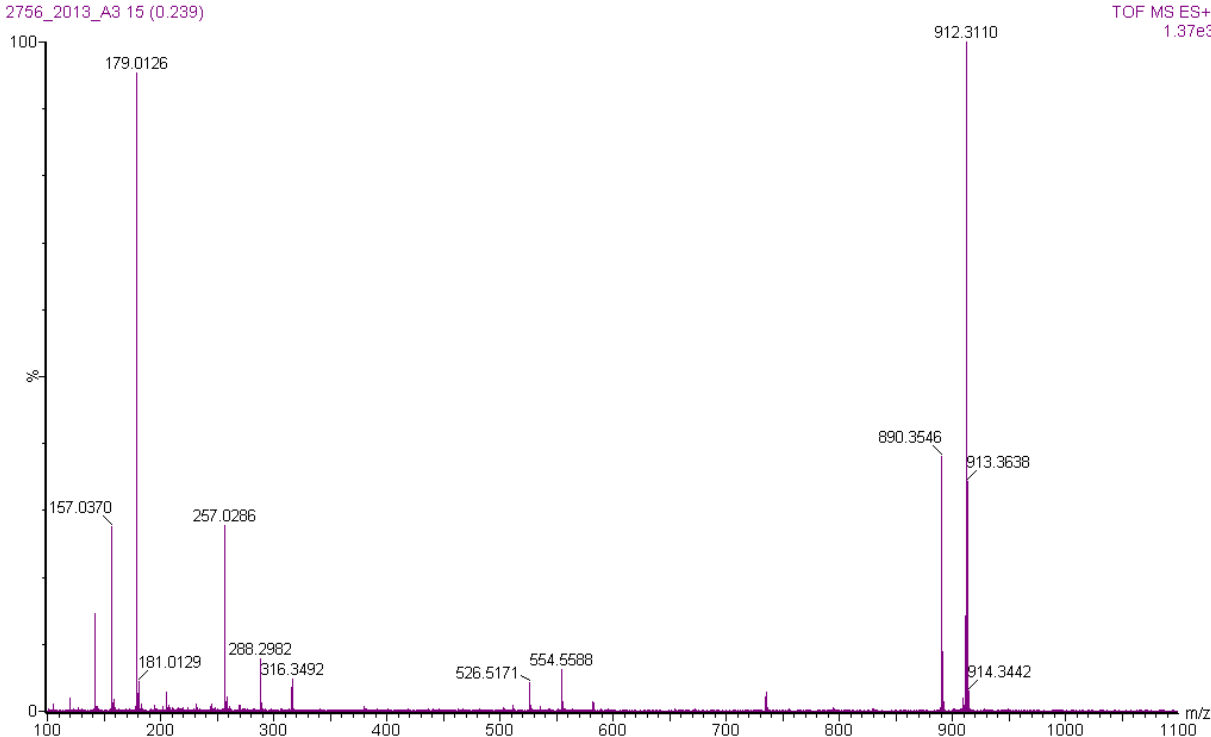
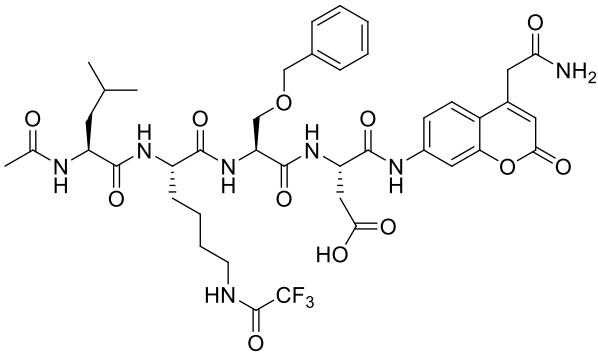


3494\_2013\_A3 10 (0.153) Cm (5:11)

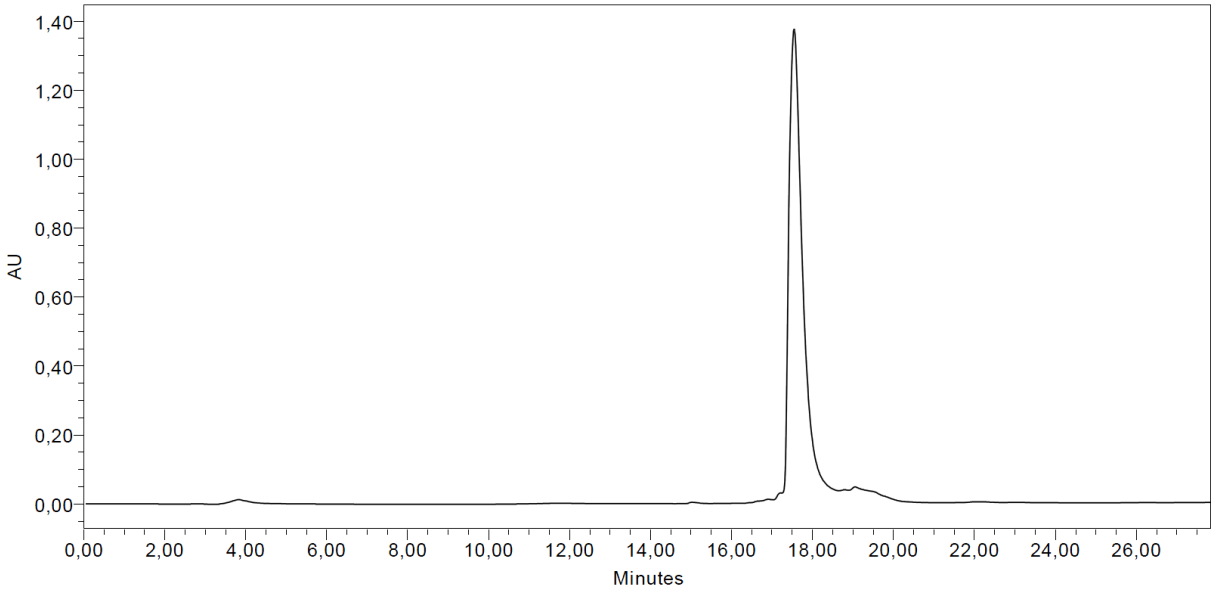
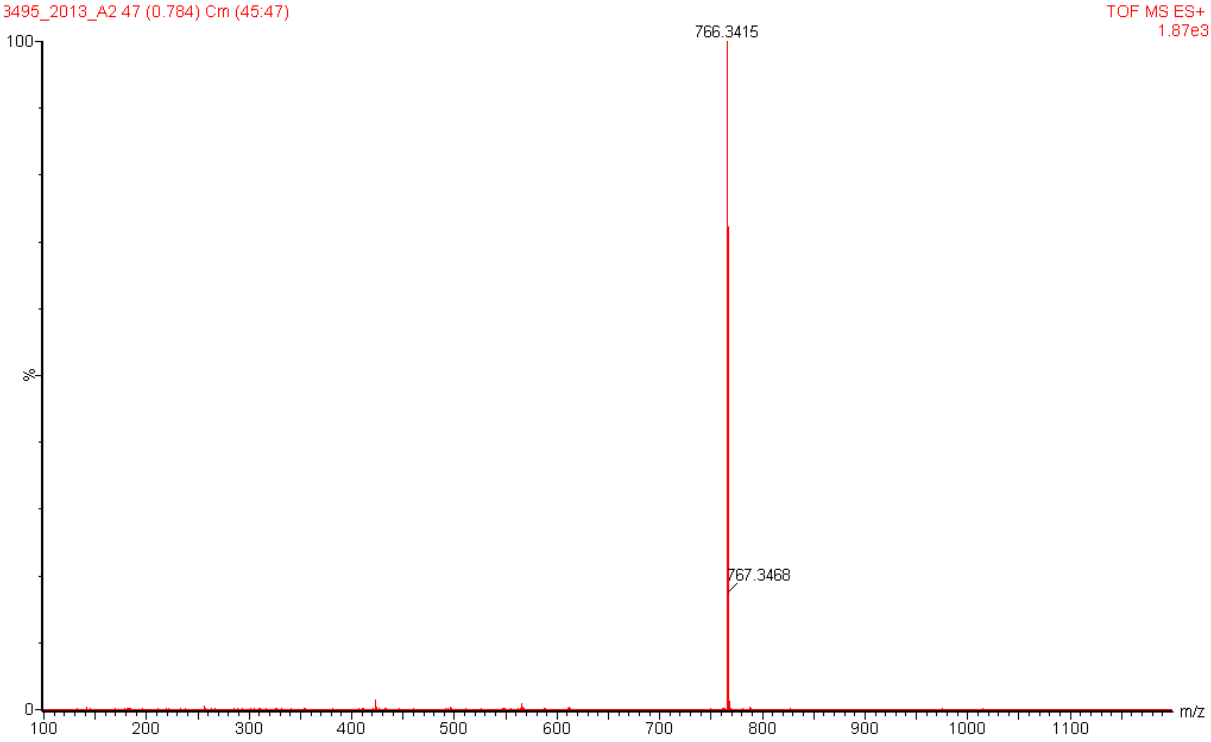
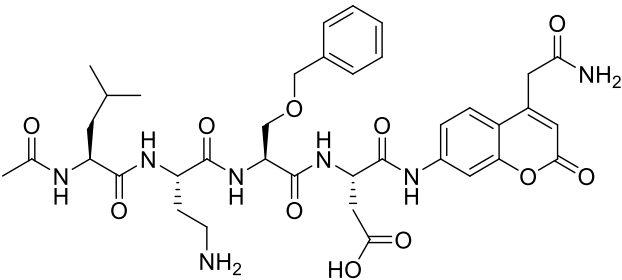
TOF MS ES+  
2.92e3



MPP50



MPP51



MPP52

