

Online-Figure 1. A. Using TEVC whole-cell Kir4.1-Kir5.1 currents were recorded from an oocyte 3 days post-injection of the Kir4.1-Kir5.1 dimer cDNA along with the mu-opioid receptor (MOR) cDNA. With 90 mM K⁺ in the extracellular solution inward rectifying currents were recorded at baseline. Membrane potential (V_m) was held at 0 mV. A series of command pulse potentials from -160 mV to 140 mV with a 20-mV increment was applied to the cell. Exposure to 1 μ M DAMGO failed to affect the channel currents, suggesting that the Kir4.1-Kir5.1 channel is modulated by G _{α q} receptors and not G_s receptors. **B.** The time profile shows that the current amplitude did not change when DAMGO was present in the bath solution.

Online-Figure 2. A. Whole-cell currents were recorded from an oocyte 3 days post-injection of the Kir4.1-Kir5.1 tandem-dimer cDNA along with the 5-HT_{2A} receptor cDNA using TEVC. Currents were recorded in the presence of high K⁺ (90 mM). Exposure to 40 μM 5-HT (serotonin) inhibited the currents by 35%. **B.** When baseline and affected currents were scaled to the same magnitude at -160 mV, the I/V relationship of the currents recorded in these two conditions were superimposed, suggesting that the effects are voltage-independent. **C.** TEVC recording from an oocyte using a ramp protocol, clearly shows inhibition of the inward currents by 5-HT.

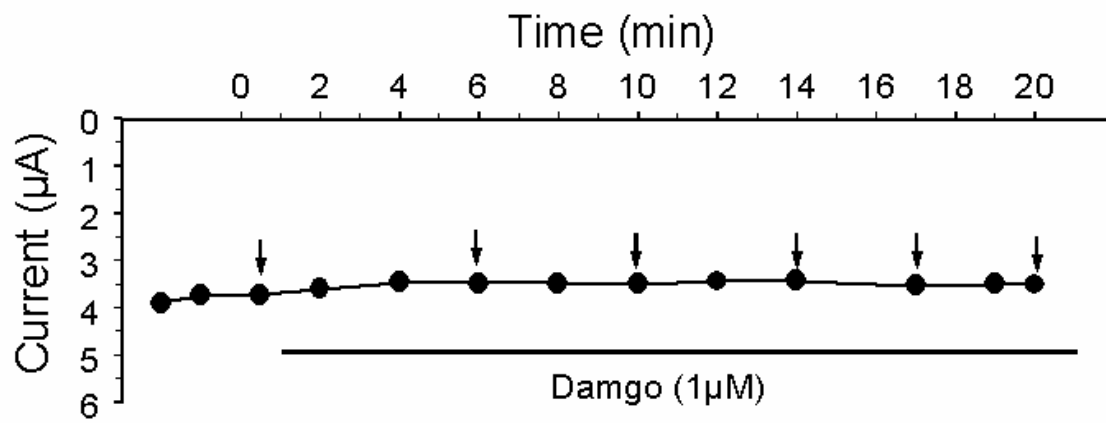
Online-Figure 3. A, B. Currents were recorded from an oocyte in the same condition as in Figure 4A. Following exposure to 15 nM PMA the cells were subsequently treated with 1 μM SP or 40 μM DOI. Prior exposure to PMA resulted in a much lower channel response to the neurotransmitters. Instead of 36% current inhibition for SP and 43% current inhibition for DOI, the Kir4.1-Kir5.1 currents were barely inhibited (>10%).

A

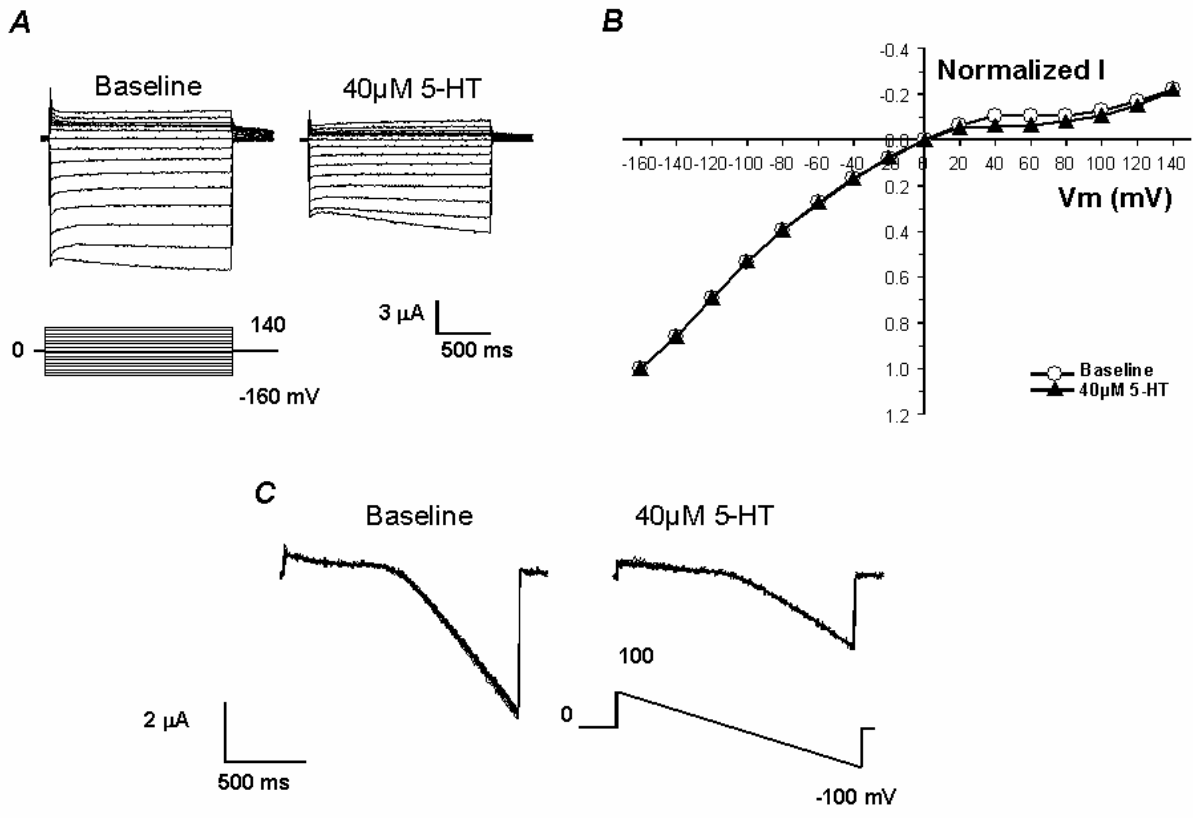
Kir4.1-Kir5.1+MOR



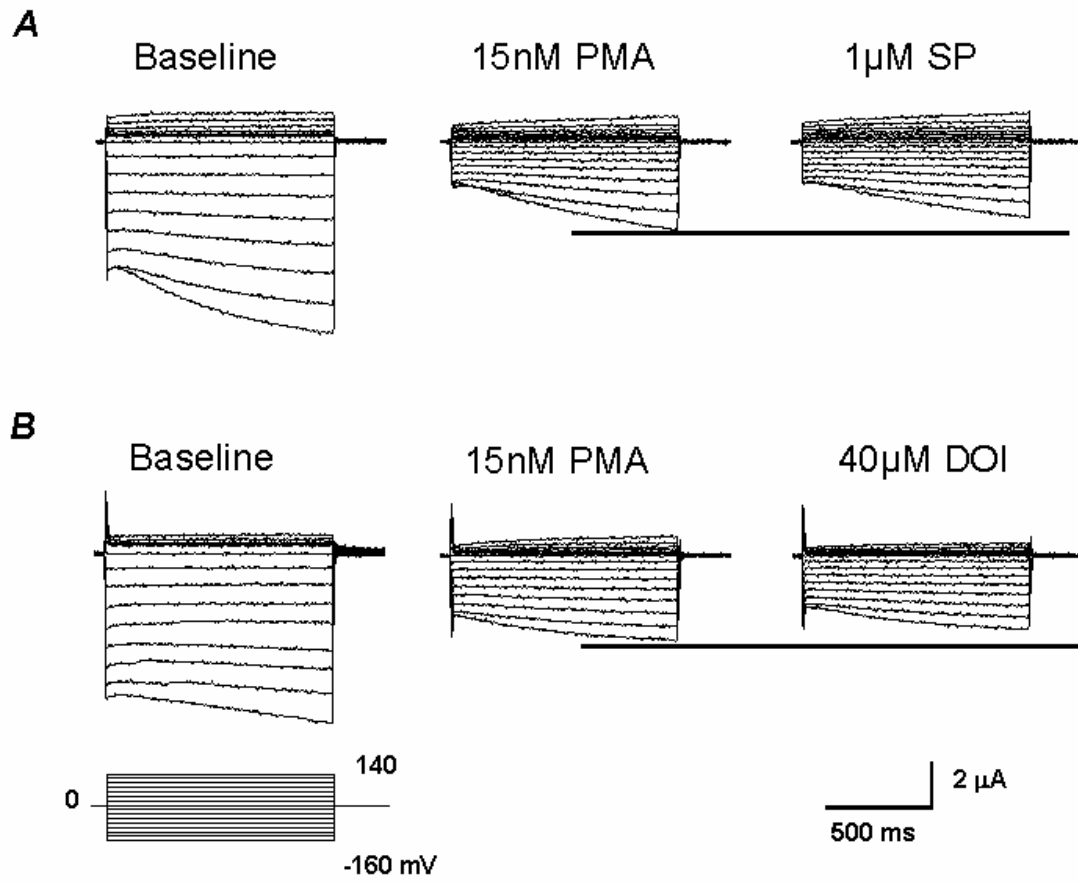
B



Online Fig. 1



Online Fig. 2



Online Fig. 3