



Fig. S1 (related to Fig. 2): Extended phenotypic analysis of cultured IL-1R1^{hi} ILC3 cells. (A) IL-1R1^{hi} ILC3s were FACS purified from SLT, and cultured in indicated conditions (IL-15 plus DMSO (□), CH-223191 (■), or FICZ (dashed line)). On d 14, flow cytometry was used to assess expression of the indicated markers. Histograms depict staining in a representative donor (n ≥ 2 for each marker). Antibodies used to detect CD11b, CD27, NKG2C, NKG2D, and granzyme B were purchased from BD Biosciences; CD16, CD69, CD127, CD161, NKp44, NKp46 were purchased from Beckman Coulter. “Pan-KIR” staining was detected using a mix comprised of the following antibodies: NKAT2 and NKB1 from BD; KIR2DL3 and KIR3DL1 from R&D Systems; and CD158a, CD158b, and KIRp70 from Beckman Coulter. (B) Single IL-1R1^{hi} ILC3s were cultured on OP9-GFP⁺ stroma in α-MEM medium containing IL-15 plus either DMSO carrier or AHR agonist FICZ. Half the medium was removed every 2-3 d and replaced with fresh media containing 2x cytokines. After 14 d, each of 60 replicate wells was individually assessed for CD94 surface expression via flow cytometry. OP9 cells were excluded from analysis by gating on GFP⁻ lymphocytes. Percentages of cells in each well expressing CD94 represent the proportion of cells which were CD94⁺ among total live GFP⁻ cells detected in that well. One hundred percent of the wells scored positive for at least some CD94 expression. Data represent mean ± SEM from 2 independent experiments with similar results.