

1 **Supplemental material**

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4 **Supplemental MATERIALS AND METHODS**

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6 **Southern blot analysis.** Southern blot analysis was performed using a probe
7 amplified with the primers (attB4)Aolah-up-F and (attB1)Aolah-up-R for verification of
8 *Aolah* gene disruption. For confirmation of *Aowsc* gene disruption, Southern blot
9 analysis was performed using a probe amplified with the primers (attB4)Aowsc-up-F
10 and (attB1)Aowsc-up-R. After electrophoresis, genomic DNAs digested with restriction
11 enzymes *EcoT22I* or *ApaI* were transferred onto Hybond N+ membrane (GE Healthcare,
12 Buckinghamshire, UK). The ECL (enhanced chemiluminescence) Direct Nucleic Acid
13 Labeling and Detection system (GE Healthcare) and a LAS-1000plus luminescent
14 image analyzer (Fuji Photo Film, Tokyo, Japan) were used for detection.

15

16 **Protein extraction and Western blot analysis** The *A. oryzae* strains expressing
17 full-length AoLAH (AoLAH-3×HA) or middle-region deleted AoLAH
18 (AoLAH[(1-2039)+(4710-5727)]-3×HA) were grown in DPY liquid nutrient medium as
19 shaking cultures for 24 h at 30°C. Cell extracts were prepared by homogenizing the
20 mycelia using liquid nitrogen in an elution buffer (50 mM Tris/HCl, pH 7.5, 1 mM
21 PMSF, and 1:100 protease inhibitor cocktail [Sigma-Aldrich, St. Louis, MO]). Total cell
22 lysates were centrifuged at 500×g for 3 min to remove cell debris, and the obtained
23 supernatants were further centrifuged (10,000×g, 10 min, 4°C). The resulting pellet
24 fraction was used for detection of middle-region deleted AoLAH. To prepare a fraction

25 highly enriched with Woronin body proteins, the 10,000×g pellet fraction was
26 re-suspended in the elution buffer supplemented with 0.5% Triton X-100, and then
27 centrifuged at 20,000×g, 4°C for 10 min, of which the pellet fraction was used for
28 detection of full-length AoLAH. The pellet fractions were dissolved in sampling buffer
29 and analyzed by Western blotting. NuPAGE® Novex 3%-8% Tris-Acetate Gel,
30 NuPAGE®LDS Sample Buffer (4X), and HiMark™ Pre-Stained Protein marker
31 (Invitrogen Life Technologies, Carlsbad, CA) were used for Western blotting analysis.
32 The primary antibodies used was mouse anti-HA monoclonal antibody (12CA5; Roche,
33 Mannheim, Germany), and the secondary was peroxidase-conjugated anti-mouse
34 antibody (Vector Laboratories, Burlingame, CA). Protein bands were detected and
35 analyzed by the ECL detection reagents (Pierce, Rockford, IL) and a LAS-1000plus
36 luminescent image analyzer.

37 **Table S1** Primers used in this study

38	Name	Sequence (5'-3')
39	(attB4)Aolah-up-F	GGGGACAAC TTTGTATAGAAAAGTTGGGAGGATTGCCTCCGCATACAATAC
40	(attB1)Aolah-up-R	GGGGACTGCTTTTTTGTACAAACTTGGCCTTGATCGCTCTCTGCCCACT
41	(attB2)Aolah-down-F_2	GGGGACAGCTTTCTTGTACAAAGTGGGATGACTATGATGACTGCCAACATCTCC
42	(attB3)Aolah-down-R_2	GGGGACAAC TTTGTATAATAAAAGTTGCGATATGGAACCGATTCAAAGTCAACTCC
43	(attB1) Aolah_1-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTGATGTTTAAGGCCTTATTGGCCGGGGCCGT
44	(attB2) Aolah_6243-R	GGGGACCACTTTGTACAAGAAAGCTGGGTCTCCAGCCC GCCATCGTCTTTTACACCGA
45	Aolah-Cter-F (14254)	GGGGACAAGTTTGTACAAAAAAGCAGGCTATGGAATGGCCTTTGATAGATTGGGAGAAGGAA
46	Aolah-Cter-R (17400)	GGGGACCACTTTGTACAAGAAAGCTGGGTCAATCACATTGCTCATGTCCATGGTTCGACGA
47	Aolah_N-R-fusion	TATCAAAGGCCATTCCTCCAGCCC GCCATCGTCTTTTACACCGAC
48	Aolah_C-F-fusion	GATGGCGGGCTGGAGGAATGGCCTTTGATAGATTGGGAGAAGGAA
49	(attB4) Aowsc-up-F	GGGGACAAC TTTGTATAGAAAAGTTGAGATGAGAGCATAGCGCGGTACC
50	(attB1) Aowsc-up-R	GGGGACTGCTTTTTTGTACAAACTTGGATGGCGGTTGATGCGGTTGCGT
51	(attB2) Aowsc-down-F	GGGGACAGCTTTCTTGTACAAAGTGGATAGCGTTACGACCAACGTCGCG
52	(attB3) Aowsc-down-R	GGGGACAAC TTTGTATAATAAAAGTTGTGCCTCCAAGAGGCGAAAGTCAGT
53	(attB1)-DsRed-M-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTCTATGGACAACACCGAGGACGTCATC
54	(attB2)-PTS1-R	GGGGACCACTTTGTACAAGAAAGCTGGGTT CATAATTTGGACTGGGAGCCGGAGTGGCG

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58 **Supplemental figure legends**

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60 **FIG. S1** Amino acid sequence alignment of N-terminal regions from *Aspergillus* LAH
61 proteins. Based on comparison with other LAH proteins, the *Aolah* gene was
62 re-predicted to contain additional 1,198 amino acids at N-terminus (indicated by an
63 arrow) of the original ORF AO090011000895 from the *A. oryzae* genome database
64 (DOGAN: Database of the Genomes Analyzed at NITE [National Institute of
65 Technology and Evaluation, Japan]; <http://www.bio.nite.go.jp/dogan/Top>). Amino acid
66 sequence data of LAH proteins from *A. fumigatus* (AfLAH) and *A. nidulans* (AnLAH)
67 were taken from the *Aspergillus* Genome Database (AspGD; <http://www.aspgd.org/>).

68

69 **FIG. S2** Amino acid sequences of poly-lysine regions in the middle region of LAH
70 proteins.

71

72 **FIG. S3** Southern blot analysis of the *Aolah* disruptant. Genomic DNAs of the parent
73 strain (NSRku70-1-1; P) and *Aolah* disruptant (NSK- Δ lah2; Δ) were digested with
74 *Eco*T22I, and then subjected to Southern blot analysis. Filled bars indicate the used
75 probe.

76

77 **FIG. S4** Domain prediction of WSC proteins. Amino acid sequence data of WSC
78 proteins from *A. fumigatus* and *A. nidulans* were taken from the *Aspergillus* Genome
79 Database (AspGD; <http://www.aspgd.org/>). SMART (<http://smart.embl-heidelberg.de/>)
80 was used for prediction of domains. ClustalW program
81 (<http://www.genome.jp/tools/clustalw/>) was used for the sequence identity analysis.

82

83 **FIG. S5** Southern blot analysis of the *Aowsc* disruptant. Genomic DNAs of the parent
84 strain (NSRku70-1-1; P) and *Aowsc* disruptant (NSK- Δ wsc1; Δ) were digested with
85 *ApaI*, and then subjected to Southern blot analysis (lower). Filled bars indicate the used
86 probe.

87

88 **FIG. S6** Western blot analysis of full length (A) and middle-region deleted (B) AoLAHs.
89 Arrowheads indicate the bands corresponding to each protein. Lanes: C, wild-type
90 control strain (NSRku70-1-1A); FL, AoLAH full-length expressing strain; Δ M: strain
91 expressing middle-region deleted AoLAH.

92

93 **FIG. S7** Disorder prediction analysis for AoLAH. The disorder probability of AoLAH
94 was predicted by PrDOS (Protein DisOrder prediction System;
95 <http://prdos.hgc.jp/cgi-bin/top.cgi>). The plot of disorder probability of each residue
96 along the sequence is shown. Residues beyond the black threshold line in this plot are
97 predicted to be disordered. Note that most of the AoLAH N-terminal and middle regions
98 is predicted to be disordered, and that part of the AoLAH C-terminal region is predicted
99 to be ordered.

100

101 **FIG. S8** Amino acid content of the middle region of LAH protein. The amino acid
102 content of the LAH proteins was calculated and the three most abundant amino acids are
103 listed. Important amino acids are highlighted by different colors; E and K are
104 highlighted by green and red, respectively.

105

106 **Supplemental Video S1** Time-lapse movie of Woronin body localization in the
107 wild-type strain. Woronin bodies were visualized by expressing AoLAH[1-2039]-EGFP
108 and asterisks indicate the septum.

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110 **Supplemental Video S2** Time-lapse movie of Woronin body localization in the *Aolah*
111 disruptant. Woronin bodies were visualized by expressing AoLAH[1-2039]-EGFP and
112 asterisks indicate the septum.

113

114 **Supplemental Video S3** Time-lapse movie of Woronin bodies tethered to the septum in
115 strain expressing full-length AoLAH. Woronin bodies were visualized by expressing
116 AoLAH[1-2039]-EGFP.

117

118 **Supplemental Video S4** Time-lapse movie of Woronin bodies tethered to the septum in
119 strain expressing middle-region deleted AoLAH. Woronin bodies were visualized by
120 expressing AoLAH[1-2039]-EGFP.

<i>A. oryzae</i>			<i>A. fumigatus</i>			<i>A. nidulans</i>		
AoLAH			LAH			LAH		
2316	KKKKKDKKKK	2325	2345	KKKKKKNKKK	2354	2670	SKKAKKKKKK	2679
2945	KKGKKKKKNR	2954	2473	KKSJKKKKKK	2482	2823	KASKKKKKNK	2832
3067	KKNKKKKKKK	3076	2738	KKNKKKNKRK	2747	2988	KKSJKKNKKK	2997
3155	KKKTKEQKK	3164	2874	KKKAKKKKNR	2883	3272	KNAKKKKKKK	3281
3491	KKDKKKKKKQ	3500	3003	KKSJKNNKKK	3012	3445	KKKNKKKKKK	3454
3593	KKKAKKDKKK	3602	3089	KKTKEKKKK	3098	3575	KKKVKKDKKK	3584
3819	KDKKKKKKRK	3828	3330	KKKDKKKKKK	3339	3734	KKRAKKEKKR	3743
3942	KKAKKERKR	3951	3438	KKKAKDKKK	3447	4273	KKKNKKAKKQ	4282
4443	KKQKKAKKQ	4452	3642	KSJKNNKKKK	3651	4443	SKKEKKLKK	4452
			3718	KKKAKKDKK	3727			
			4246	SKKSKKAKK	4255			

FIG. S2 Amino acid sequences of poly-lysine regions in the middle region of LAH proteins.

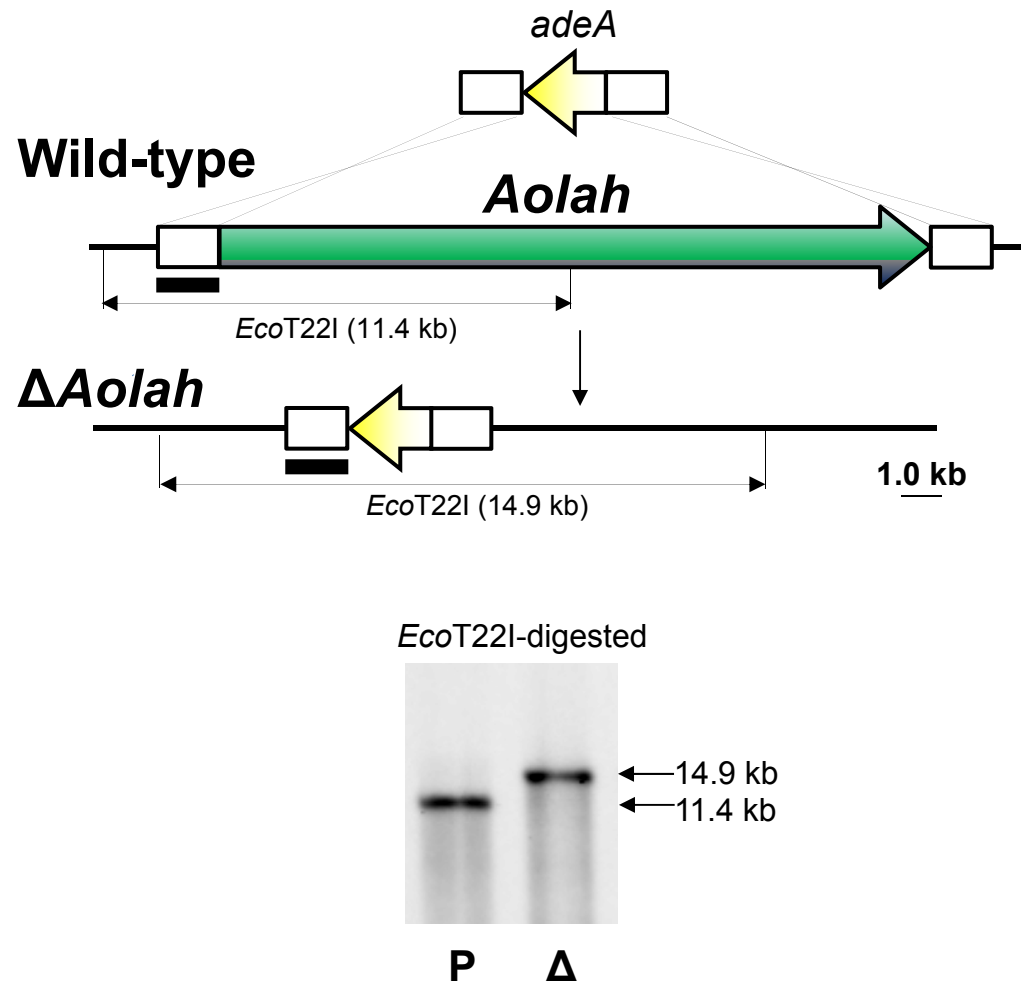


FIG. S3 Southern blot analysis of the *Aolah* disruptant. Genomic DNAs of the parent strain (NSRku70-1-1; P) and *Aolah* disruptant (NSK- $\Delta lah2$; Δ) were digested with *EcoT22I*, and then subjected to Southern blot analysis. Filled bars indicate the used probe.

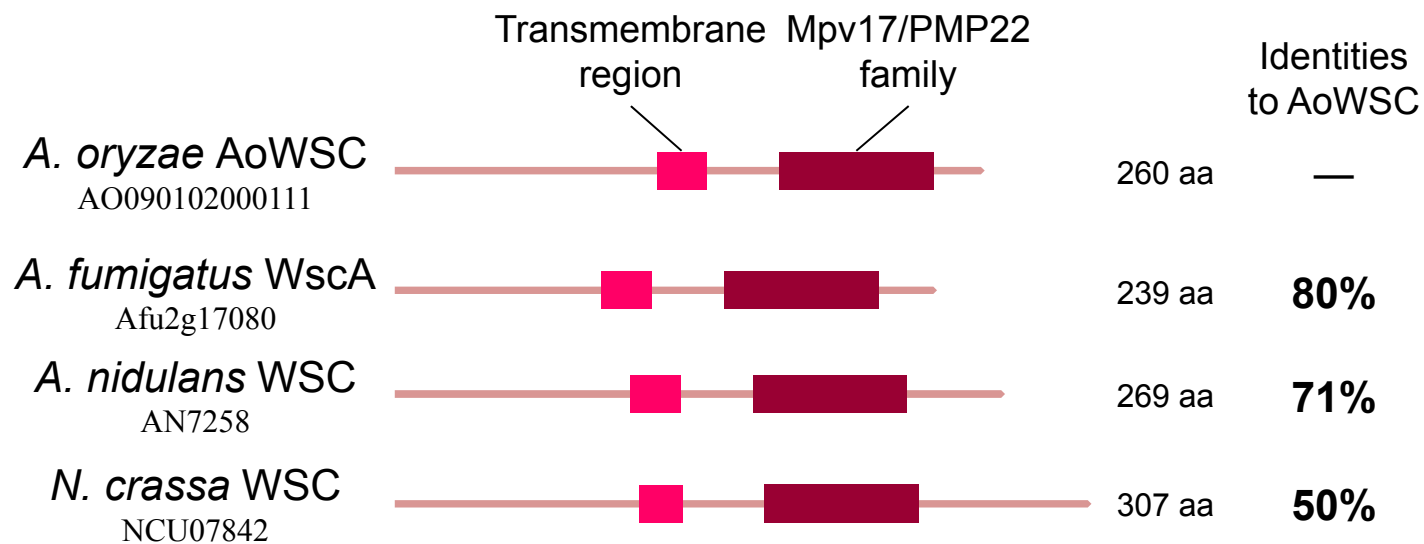


FIG. S4 Domain prediction of WSC proteins. Amino acid sequence data of WSC proteins from *A. fumigatus* and *A. nidulans* were taken from the *Aspergillus* Genome Database (AspGD; <http://www.aspgd.org/>). SMART (<http://smart.embl-heidelberg.de/>) was used for prediction of domains. ClustalW program (<http://www.genome.jp/tools/clustalw/>) was used for the sequence identity analysis.

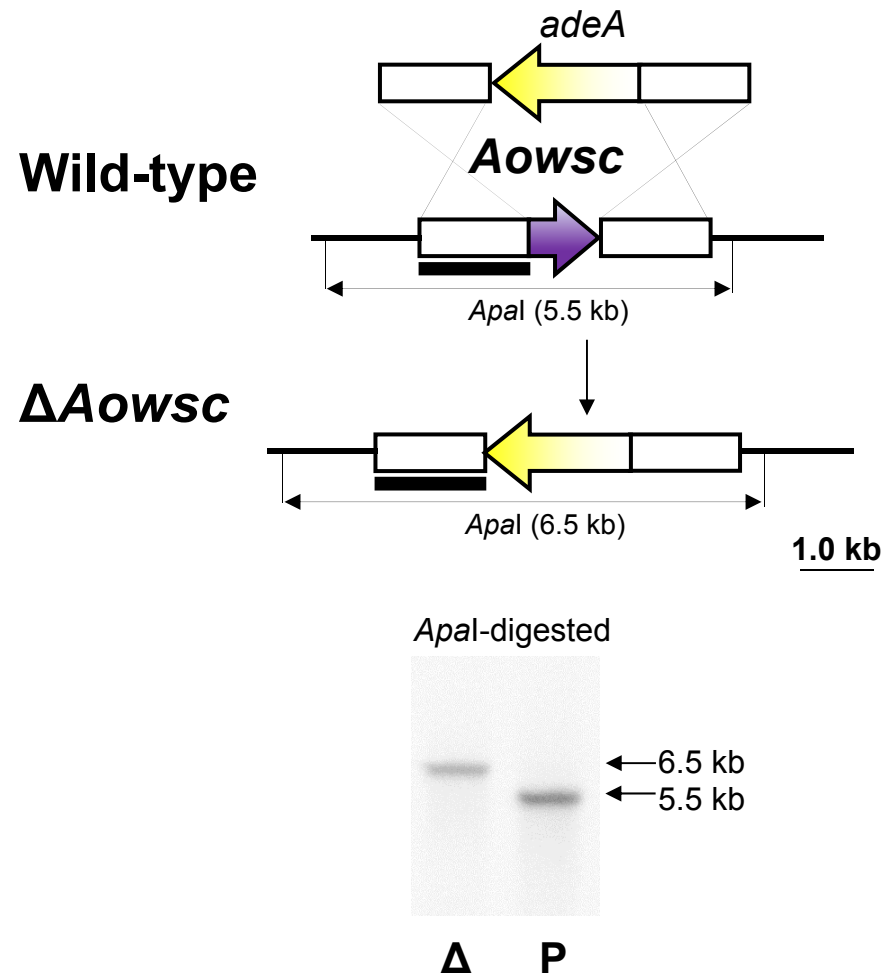


FIG. S5 Southern blot analysis of the *Aowsc* disruptant. Genomic DNAs of the parent strain (NSRku70-1-1; P) and *Aowsc* disruptant (NSK-Δwsc1; Δ) were digested with *ApaI*, and then subjected to Southern blot analysis (lower). Filled bars indicate the used probe.

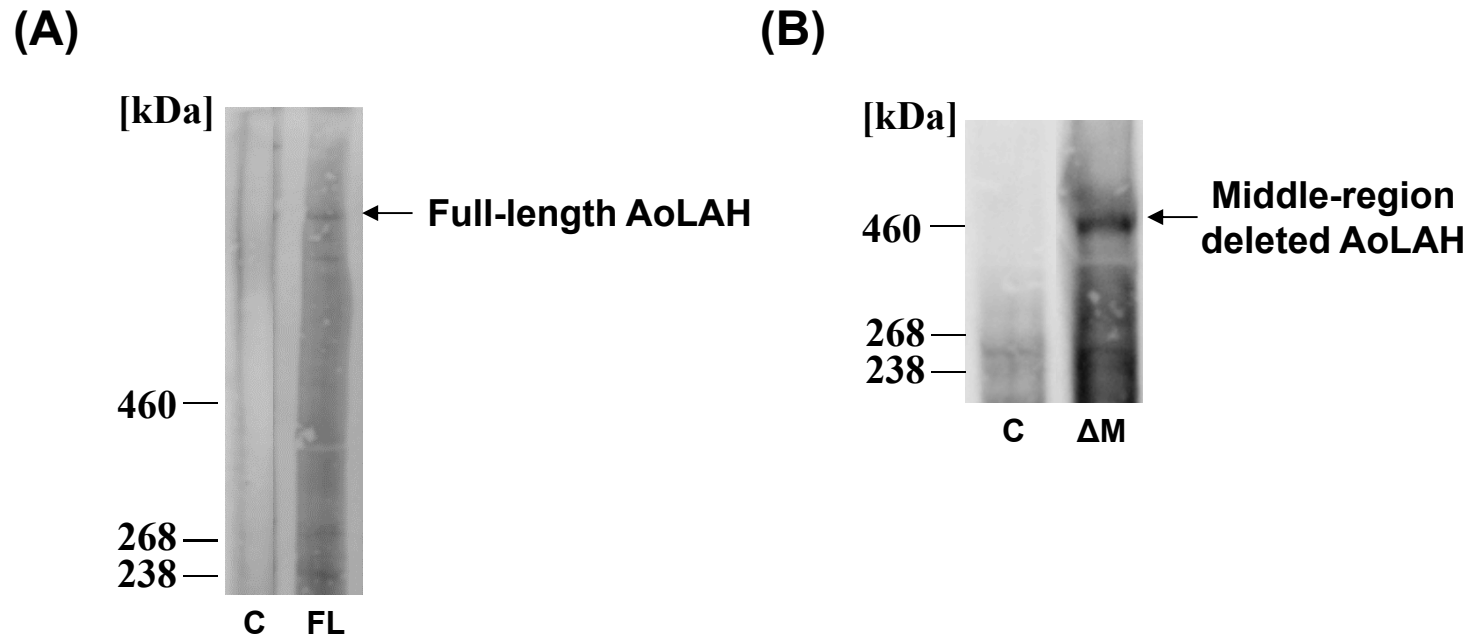


FIG. S6 Western blot analysis of full length (A) and middle-region deleted (B) AoLAHs. Arrowheads indicate the bands corresponding to each protein. Lanes: C, wild-type control strain (NSRku70-1-1A); FL, AoLAH full-length expressing strain; Δ M: strain expressing middle-region deleted AoLAH.

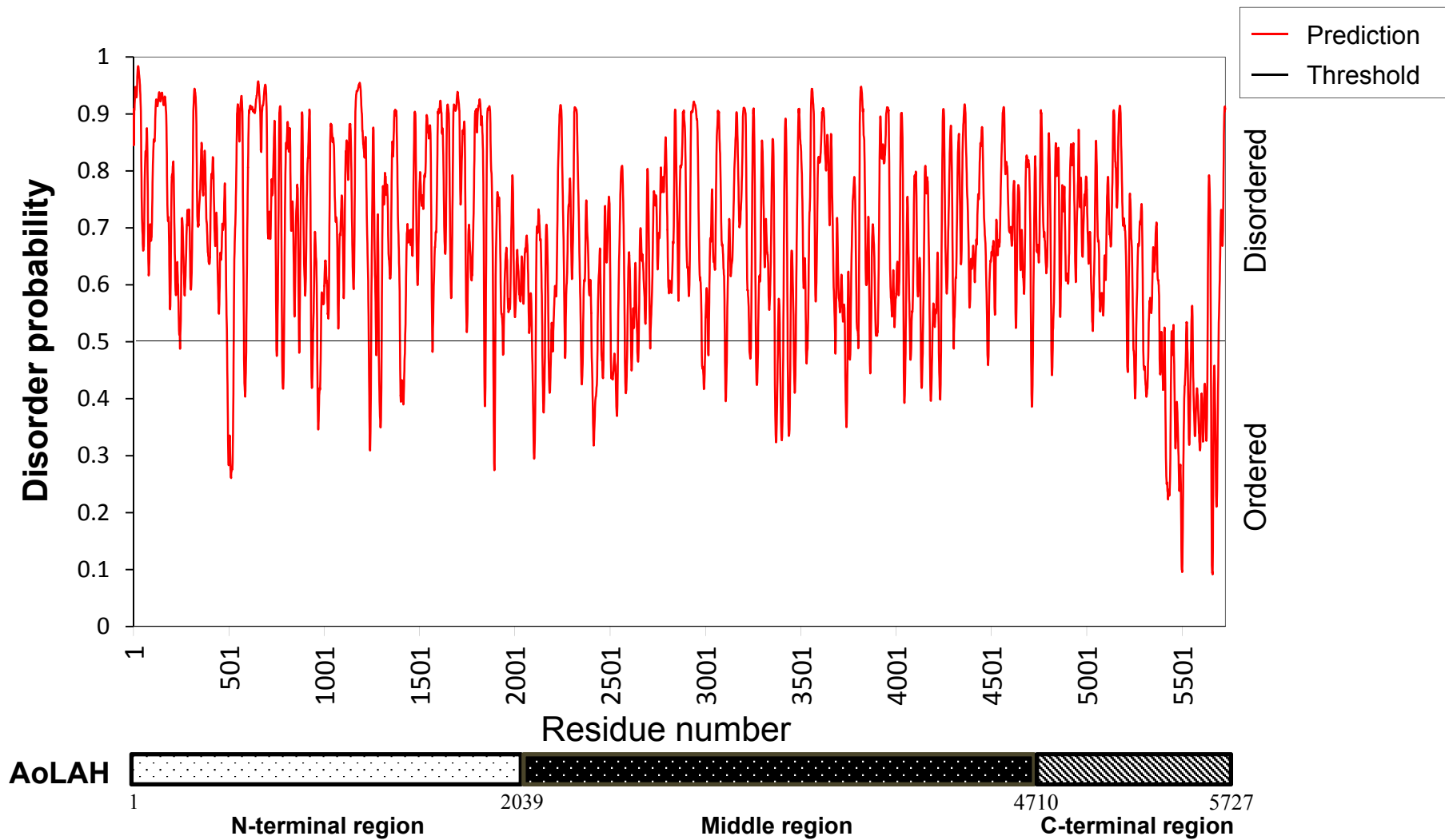


FIG. S7 Disorder prediction analysis for AoLAH. The disorder probability of AoLAH was predicted by PrDOS (Protein DisOrder prediction System; <http://prdos.hgc.jp/cgi-bin/top.cgi>). The plot of disorder probability of each residue along the sequence is shown. Residues beyond the black threshold line in this plot are predicted to be disordered. Note that most of the AoLAH N-terminal and middle regions is predicted to be disordered, and that part of the AoLAH C-terminal region is predicted to be ordered.

