SUPPLEMENTAL FIGURE LEGENDS

Figure S1. Cytochrome c released by HCCS variants

Recombinant GST-HCCS variants with substitutions in A) Domain I, B) Domain II, C) Domain III, and D) Domain IV were co-expressed with cytochrome c in Δ ccm *E.coli* in 1 L cultures. Cells were lysed by sonication and fractionated by ultracentrifugation. UV/vis absorption spectra were recorded for the resulting soluble fractions using 1 mg of total protein, quantified by Nanodrop. The alpha peak region of the recorded spectra is depicted and each colored spectrum represents the indicated mutant in the corresponding color.

Figure S2. N128A and M130A HCCS mutants exhibit distinguishable heme characteristics

Recombinant GST-HCCS: cytochrome c co-complexes were purified from Δ ccm *E. coli* and prepared for UV/vis absorption spectroscopy. Shown are spectra for A) N128A HCCS/ cyt c and B) M130A HCCS cyt c following purification (black line) and chemical reduction with sodium dithionite (red). Arrows indicate the wavelength (nm) of peak absorption maxima. All spectra were performed with equal amounts (100 µg) of total purified protein. For all proteins, Bradford quantitation was confirmed by Coomassie staining, which also indicated that GST-HCCS proteins were obtained at >90% purity.

Figure S3. ALA treatment enhances the synthetase activity of several HCCS variants

Recombinant GST-HCCS variants were co-expressed with cytochrome c in Δ ccm *E. coli* either in the presence (+) or absence of ALA (-). Cells were lysed with BPER reagent and 100 µg of

protein extracts were resolved by SDS-PAGE and transferred to nitrocellulose. Released cytochrome c was detected by heme stain. Data shown are representative of 3 replicate experiments.



Figure S1



Figure S2



Figure S3

	Oligo ID	Sequence (5'-3')	Constructed plasmid	
1	W118A-F	AGC AGA TTC AGA GAA AAA GGC GGT TTA CCC TTC TGA GCA G	HCCS_W118A	
	V120A/D121A E	CIUCIC AGA AGO GIA AAC COC CIT III CIC IGA AIC IGC I	HCCS V120A/	
2	V120A/P121A-P		P121A	
	V120A F		1121A	
3	V120A-F		HCCS_Y120A	
	D120A-K	GAG AAA AAG TGG GTT TAC GCT TCT GAG CAG ATG TTC T		
4	P121A-P	AGA ACA TCT GCT CAG AAG CGT AAA CCC ACT TTT TCT C	HCCS_P121A	
	N128A/M130A-F	CCC TTC TGA GCA GAT GTT CTG GGC TGC AGC GTT AAA GAA AGG GTG GAA GTG G	HCCS N128A/	
5	N128A/M130A-R	CCA CTI CCA CCC TTI CTI TAA CGC TGC AGC CCA GAA CAT CTG CTC AGA AGG G	M130A	
	N128A-F	CCT TCT GAG CAG ATG TTC TGG GCT GCA ATG TTA AAG AAA GGG TG		
6	N128A-R	CAC CCT TTC TTT AAC ATT GCA GCC CAG AAC ATC TGC TCA GAA GG	HCCS_N128A	
	M130A-F	GAG CAG ATG TTC TGG AAT GCA GCG TTA AAG AAA GGG TGG AAG TG		
7	M130A-R	CAC TTC CAC CCT TTC TTT AAC GCT GCA TTC CAG AAC ATC TGC TC	HCCS_M130A	
	N155A-F	GGA TAT GTA TAA TAT CAT TAG AAT TCA CGC TCA GAA TAA CGA GCA GGC TTG GAA G		
8	N155A-R	CTT CCA AGC CTG CTC GTT ATT CTG AGC GTG AAT TCT AAT GAT ATT ATA CAT ATC C	HCCS_N155A	
	E159K-F	ATT AGA ATT CAC AAT CAG AAT AAC AAG CAG GCT TGG AAG GAG		
9	E159K-R	CTC CTT CCA AGC CTG CTT GTT ATT CTG ATT GTG AAT TCT AAT	HCCS_E159K	
1.0	E159A-F	TCA CAA TCA GAA TAA CGC GCA GGC TTG GAA GGA GA	MOGO ELGON	
10	E159A-R	TCT CCT TCC AAG CCT GCG CGT TAT TCT GAT TGT GA	HCCS_E159A	
	E159D-F	AAT TCA CAA TCA GAA TAA CGA TCA GGC TTG GAA GGA GAT TTT G		
11	E159D-R	CAA AAT CTC CTT CCA AGC CTG ATC GTT ATT CTG ATT GTG AAT T	HCCS_E159D	
10	W162A-F	TCA GAA TAA CGA GCA GGC TGC GAA GGA GAT TTT GAA GTG G		
12	W162A-R	CCA CTT CAA AAT CTC CTT CGC AGC CTG CTC GTT ATT CTG A	HCCS_W162A	
12	W168A/E169A-F	GCT TGG AAG GAG ATT TTG AAG GCG GCA GCC CTT CAT GCT GCA	HCCS W168A/	
13	W168A/E169A-R	TGC AGC ATG AAG GGC TGC CGC CTT CAA AAT CTC CTT CCA AGC	E169A	
1.4	W168A-F	TGG AAG GAG ATT TTG AAG GCG GAA GCC CTT CAT GCT GC		
14	W168A-R	GCA GCA TGA AGG GCT TCC GCC TTC AAA ATC TCC TTC CA	HCCS_W168A	
15	E169A-F	GGA GAT TTT GAA GTG GGC AGC CCT TCA TGC TGC AG	LICCS E160A	
15	E169A-R	CTG CAG CAT GAA GGG CTG CCC ACT TCA AAA TCT CC	HCCS_E109A	
16	E169K-F	AGG AGA TTT TGA AGT GGA AAG CCC TTC ATG CTG CA	HCCS E160V	
10	E169K-R	TGC AGC ATG AAG GGC TTT CCA CTT CAA AAT CTC CT	HCCS_E109K	
17	W213A-F	GCC TTT TGA TAG GCA CGA TGC GAT CAT AAA CCG TTG CGG G	HCCS W213A	
17	W213A-R	CCC GCA ACG GTT TAT GAT CGC ATC GTG CCT ATC AAA AGG C	IICC5_W2I5A	
18	R217C-F	AGG CAC GAT TGG ATC ATA AAC TGT TGC GGG ACA GAA	HCCS R217C	
10	R217C-R	TTC TGT CCC GCA ACA GTT TAT GAT CCA ATC GTG CCT	nees_k217e	
19	R217A-F	AGG CAC GAT TGG ATC ATA AAC GCT TGC GGG ACA GAA G	HCCS R217A	
17	R217A-R	CTT CTG TCC CGC AAG CGT TTA TGA TCC AAT CGT GCC T	nees_k21/A	
20	R217K-F	CTT TTG ATA GGC ACG ATT GGA TCA TAA ACA AGT GCG GGA CAG AAG TTA GA	HCCS R217K	
20	R217K-R	TCT AAC TTC TGT CCC GCA CTT GTT TAT GAT CCA ATC GTG CCT ATC AAA AG	nees_iezi/k	
21	R217D-F	ATA GGC ACG ATT GGA TCA TAA ACG ATT GCG GGA CAG AAG T	HCCS R217D	
21	R217D-R	ACT TCT GTC CCG CAA TCG TTT ATG ATC CAA TCG TGC CTA T	11000_1021710	
22	D227A-F	GAC AGA AGT TAG ATA TGT GAT TGC TTA TTA TGA TGG TGG TGA AGT CA	HCCS D227A	
22	D227A-R	TGA CTT CAC CAC CAT CAT AAT AAG CAA TCA CAT ATC TAA CTT CTG TC	nees_bzz/m	
23	Y229A-F	CGG GAC AGA AGT TAG ATA TGT GAT TGA TTA TGC TGA TGG TGG TGA AGT C	HCCS Y229A	
	Y229A-R	GAC TTC ACC ACC ATC AGC ATA ATC AAT CAC ATA TCT AAC TTC TGT CCC G		
24	R246A-F	CAC CAT CCT GGA CGT CGC TCC TGC CTT AGA TTC A	HCCS R246A	
L	R246A-R	TGA ATC TAA GGC AGG AGC GAC GTC CAG GAT GGT G	11000_10/11	
25	D257A/R258A-F	TCA CTT TCG GCA GTA TGG GCC GCA ATG AAA GTC GCT TGG TGG	HCCS_D257A/	
	D257A/R258A-R	CCA CCA AGC GAC TTT CAT TGC GGC CCA TAC TGC CGA AAG TGA	R258A	
26	D257A-F	ACT TIC GGC AGT ATG GGC CAG AAT GAA AGT CGC TT	HCCS D257A	
	D257A-R	AAG CGA CTT TCA TTC TGG CCC ATA CTG CCG AAA GT		
27	D257K-F	TTC ACT TTC GGC AGT ATG GAA GAG AAT GAA AGT CGC TTG GT	HCCS D257K	
- '	D257K-R	ACC AAG CGA CITI TCA TICI TCT TCC ATA CTG CCG AAA GTG AA		
28	R258A-F	CTT TCG GCA GTA TGG GAC GCA ATG AAA GTC GCT TGG TG	HCCS R258A	
	R258A-R	CAC CAA GCG ACT TTC ATT GCG TCC CAT ACT GCC GAA AG		
29	R258E-F	CAC TIT CGG CAG TAT GGG ACG AGA TGA AAG TCG CTT GGT GGC G	HCCS R258E	
	R258E-R	CGC CAC CAA GCG ACT TTC ATC TCG TCC CAT ACT GCC GAA AGT G		

Table S1. Oligonucleotide primers and plasmids

Putative Domain	HCCS variant	BPER ^a	UV/vis ^b
	WT	1	1
	W118A	1.7±0.25	1.6±0.26
	Y120A/P121A	1.1±0.13	0.5±0.09
	Y120A	0.7±0.28	1.2±0.03
Domain I	P121A	0.9±0.91	1.9±0.22
	N128A/M130A	1.3±0.38	1.9±0.29
	N128A	1.5±0.15	1.9±0.27
	M130A	1.2±0.41	1.3±0.17
	N155A	2.0±0.33	1.9±0.12
	E159K	0.1±0.01	0.7±0.05
	E159A	1.9±0.76	1.8±0.11
	E159D	1.3±0.75	$0.9{\pm}0.08$
Domain II	W162A	0.3±0.10	0.3±0.04
	W168A/E169A	< 0.1	nd
	W168A	< 0.1	nd
	E169A	< 0.1	nd
	E169K	< 0.1	nd
Domain III	W213A	1.4±0.20	1.1 ± 0.06
	R217C	< 0.1	nd
	R217K	< 0.1	nd
	R217D	< 0.1	nd
	R217A	< 0.1	nd
	D227A	< 0.1	nd
Domain IV	Y229A	< 0.1	nd
Domain I v	R246A	< 0.1	nd
	D257A/R258A	< 0.1	nd
	D257A	1.3±0.48	0.9±0.31
	D257K	0.2±0.19	0.4 ± 0.16
	R258A	< 0.1	0.6±0.13
	R258E	< 0.1	nd

Table S2. Quantitation of released cytochrome c from two methods

Each value is relative to the amount of cytochrome c released by WT HCCS, which has been set to 1. Data shown represent the average of at least two separate experiments \pm SEM.

^{*a*}BPERS were performed as described in Materials and Methods. Calculated values were based on the densitometric analysis of chemiluminescent signal from heme stains.

^bCalculated values were based on the reduced alpha peak height in the UV/vis absorption spectrum of 1 mg total protein from the soluble fractions of 1 L cultures co-expressing each respective variant with cytochrome c.

		Protein Levels	
	HCCS variant		Co-expressed
Putative		Expressed	with
Domain		alone	cytochrome c
			acceptor
	WT	1	1
	W118A	0.55	0.91
	Y120A/P121A	0.50	0.53
	Y120A	0.82	0.40
Domain I	P121A	0.95	0.28
	N128A/M130A	0.95	0.65
	N128A	0.76	0.61
	M130A	0.75	0.61
	N155A	0.48	0.54
	E159K	0.51	0.39
	E159A	0.65	0.55
	E159D	0.76	0.23
Domain II	W162A	0.20	0.15
	W168A/E169A	0.14	0.22
	W168A	0.29	< 0.10
	E169A	0.30	< 0.10
	E169K	0.20	< 0.10
Domain III	W213A	0.45	0.59
	R217C	0.14	0.21
	R217K	0.18	0.12
	R217D	0.23	< 0.10
	R217A	0.23	< 0.10
	D227A	0.52	0.15
Domain IV	Y229A	0.80	0.11
Domain I v	R246A	0.23	< 0.10
	D257A/R258A	0.33	0.13
	D257A	0.59	0.33
	D257K	0.20	0.15
	R258A	0.33	0.45
	R258E	0.13	0.11

Table S3. Quantitation of purified protein

Each value is based on the Bradford measurement of total milligrams of GST-HCCS protein purified from 1 L of culture relative to the amount of total protein purified from 1 L of WT HCCS, which has been set to 1, under each respective condition. Bradford quantitation was confirmed by Coomassie staining, which also indicated that GST-HCCS proteins were obtained at >90% purity. Data shown represent the average yield from at least two separate experiments.