

Supporting Information

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Development of Cyclobutene- and Cyclobutane-Functionalized Fatty Acids with Inhibitory Activity against *Mycobacterium tuberculosis*

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cmdc_201402067_sm_miscellaneous_information.pdf

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Benzyl 9-decenoate was prepared by a known procedure.¹ Thionyl chloride (36.9 mL, 51 mmol) was added to a stirred solution of 9-decenoic acid DA (8.67 g, 51 mmol) and catalytic dimethylformamide (DMF, 0.12 mL, 1.5 mmol) in toluene (400 mL). The reaction was stirred at rt until the starting material was consumed (TLC, ~ 24 h). The reaction was concentrated and the residue redissolved in toluene (400 mL) and triethylamine (8.5 mL). The solution was stirred for 24 h and washed with water. The aqueous layer was then extracted with ether. The combined organic solution were combined, dried with Na₂SO₄, and concentrated under vacuum. The residue was purified by flash column chromatography on silica gel (5-10% EtOAc in Hex) to give benzyl 9-decenoate (13.1 g, 99%) as a colorless oil: $R_f = 0.40$ (10% EtOAc/ Hex); IR (ZnSe, cm⁻¹): 3072, 2927, 2855, 1734, 1161, 908, 730 cm⁻¹; ¹H NMR: δ 7.38-7.32 (m, 5H), 5.83 (ddt, J = 17, 10.2, 6.6 Hz, 1H), 5.14 (s, 2H), 5.02 (ddd, J = 7.6 Hz, 2H), 5.00 (dq, J = 17.0, 1.7 Hz, 1H), 4.96 (d of quintet, J = 10.2, 1.2 Hz, 1H), 2.37 (t, J = 7.3 Hz, 2H), 2.10-2.02 (br q, J = 7.3Hz, 2H), 1.70-1.63 (br m, 2H), 1.41-1.48 (m, 10H); ¹³C NMR: δ 173.5, 138.9, 136.1, 128.4, 128.1, 128.0, 114.1, 65.9, 34.2, 33.7, 28.99, 28.97, 28.8, 28.7, 24.8, 22.6, 14.0; HRESI-MS: calcd. for C₁₇H₂₄O₂Na: [M+Na]⁺ 283.1674; Found: 283.1671.

(Z)-9-Octadecenoate, benzyl ester (benzyl oleate) was prepared by a variation of a known procedure.² Benzyl alcohol (3.83 g, 35.4 mmol) was added to a stirred solution of oleic acid (5.136 g, 18.1 mmol) and DMAP (43.7 mg, 3.57 mmol) in CH₂Cl₂ (40 mL). The solution was cooled to 0 °C and DCC (4.211 g, 20.4 mmol) was added. The mixture was allowed to warm to rt and stirred for 2.5 h. The resulting suspension was filtered through a cotton plug and the precipitate was washed with CH₂Cl₂. The CH₂Cl₂ solution was washed with water and dried over Na₂SO₄. Evaporation of the organic solvent and flash chromatography of the residue over silica gel, using 5% EtOAc/Hex, gave benzyl oleate (5.73 g, 85 %) as a colorless oil: $R_f = 0.83$ (10%

EtOAc/Hex); IR (ZnSe, cm⁻¹) 2924, 2853, 1738, 1456, 1162, 696 cm⁻¹; ¹H NMR: δ 7.38-7.31 (5H), 5.41-5.31 (2H), 5.13 (s, 2H), 2.37 (t, *J* = 7.6 Hz, 2H), 2.05-2.00 (4H), 1.70-1.62 (m, 2H), 1.40-1.29 (20H), 0.90 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 173.6, 136.12, 129.9, 129.7, 128.5 (two overlapping signals), 128.11, 128.09, 66.0, 34.3, 31.9, 29.7, 29.6, 29.50, 29.3, 29.1, 29.1, 27.2, 27.1, 24.9, 22.7, 14.1.

(*E*)-9-octadecenoic acid, benzyl ester (benzyl elaidate)³ was prepared from the reaction of elaidic acid (0.9836, 3.5 mmol), DCC (0.79 g, 3.8 mmol), DMAP (0.087 g, 0.7 mmol), and benzyl alcohol (0.73 mL, 7 mmol) in CH₂Cl₂ (10 mL) by a similar procedure as employed for benzyl oleate. The product was purified by flash column chromatography on silica gel (2.5% / 5% EtOAc/Hex) to afford 1.2280 g (95%) of benzyl elaidate as a colorless oil: Rf = 0.57 (10% EtOAc/Hex). IR (ZnSe, cm⁻¹) 2922, 2852, 2737, 1160, 966, 696 cm⁻¹; ¹H NMR: δ 7.38-7.34 (m, 5H), 5.41-5.38 (m, 2H), 5.13 (s, 2H), 2.37 (t, *J* = 7.2 Hz, 2H), 1.98 (br s, 4H), 1.68-1.63 (m, 2H), 1.38-1.18 (br m, 20H), 0.90 (t, *J* = 7.2 Hz, 3H); ¹³C NMR δ 173.6, 136.1, 130.4, 130.2, 128.5, 128.1, 66.0, 34.3, 32.57, 32.52, 31.9, 29.6, 29.52, 29.46, 29.3, 29.2, 29.1, 29.0, 28.9, 24.9, 22.7, 14.1.

Data from cytotoxicity studies



Figure S1. Analogs investigated

Compound	Average % viability at indicated concentration (μM)					
	0	25	50	100	250	Control ^a
1 (DA-CB)	100	67.8	58.5	48.1	0.9	0.2
2 (DA-satCB)	100	72.9	49.2	7.5	0.6	0.2
4 (DA-alcCB)	100	86.5	78.7	77.5	58.8	0.2
5 (OA-CB)	100	93.8	91.8	89.3	73.4	0.2
6 (OA-satCB)	100	87.8	84.6	82.5	78.9	0.2
8 (OA-alcCB)	100	5.5	4.4	4.4	3.7	0.2
DA	100	83.5	77.3	73	38.3	0.2
D-cycloserine	100	81.8	77.5	72.2	69.4	0.2
Isoniazid	100	76.5	75.1	71.7	64	0.2

Table S1. Toxicity of Fatty acid analogs in RAW 2647 macrophages

a. Sodium dodecyl sulfate (SDS)

Thermal Stability (Figure S1-S11)

Differential scanning calorimetry (DSC) and thermal gravimetric analysis (TGA) were acquired simultaneously on 4-10 mg samples of the indicated molecules, using a Perkin Elmer STA 6000 analyzer operated at a temperature ramp of 2 °C/minute except for compound **5** at a 5 °C/minute



Figure S1. Thermal stability of 1. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S2. Thermal stability of 2. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S3. Thermal stability of 3. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S4. Thermal stability of 4. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S5. Thermal stability of 5. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S6. Thermal stability of 6. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S7. Thermal stability of 7. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S8. Thermal stability of 8. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S9. Thermal stability of **9.** TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S10. Thermal stability of 10. TGA-measured mass (red) and DSC-measured heat flow (blue).



Figure S11. Thermal stability of 11. TGA-measured mass (red) and DSC-measured heat flow (blue).



Decenoic acid: aggregation assay in phosphate buffer



Analog **2** (C_{10} cyclobutane); aggregation assay in phosphate buffer



Analog 4 (C_{10} cyclobutanol); aggregation assay in phosphate buffer



Oleic acid: aggregation assay in phosphate buffer







Analog 7 (C_{18} cis-cyclobutanone); aggregation assay in phosphate buffer



Analog 8 (C_{18} cyclobutanol); aggregation assay in phosphate buffer







Analog **10** (C_{18} trans-cyclobutene); aggregation assay in phosphate buffer



Analog **11** (C_{18} trans-chloroketone); aggregation assay in phosphate buffer

References

¹ J, J. Li, C. Limberakis, D. A. Pflum, *Modern Organic Synthesis in the Laboratory: A Collection of Standard Experimental Procedures*, Oxford University Press, New York, **2007**, p 45.

² B. Neises, W. Steglich, Angew. Chem. Int. Ed. **1978**, 17, 522–524. Previous report: E. G. Maleeva, *Zhurnal Obshchei Khimii* **1953**, 23, 1662–1664; K. Murai, G. J. Akazome, Jpn Oil Chem. Soc. **1955**, 4, 125–127. (no detail for preparation)

³ E. G. Maleeva, *Zhurnal Obshchei Khimii*. **1953**, *23*, 1662–1664 [*Chem. Abstr.* **1954**, *48*, 77506].



Figure S1.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum.



Figure S2.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 15.



Figure S3.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum.





Figure S4.2. 13 C-NMR (100 MHz, chloroform-*d*) spectrum of 16.



80

70

60

50

40

30



180 170 160 150 140 130 120 110 100 90

P1 PL1 SF01

CPDPRG2

NUC2 NUC2 PCPD2 PL2 PL12 PL13 SFO2

SI VDW SВ

GB -PC

ppm

CHANNEL f2 ===

waltz16

1H 70.00 usec -4.00 dB 12.90 dB 12.90 dB 400.1316005 MHz

32768 100.6127704 MHz EM

0 1.00 Hz 0 1.40



Figure S6.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of 2.



Figure S6.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 2.



Figure S7.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 17.

90

80

70

60

50

40

30

160 150 140 130 120 110 100

170

===

CPDPRG2

PCPD2 PCPD2 PL2 PL12 PL13 SFO2 SI SF WDW

SSB

LB GB TPC

ppm

CHANNEL f2 ===

waltz16

Waltz16 1H 70.00 usec -4.00 dB 12.90 dB 12.90 dB 400.1316005 MHz 32768

32768 100.6127901 MHz EM

0 1.00 Hz

0



Figure S8.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of 3.



Figure S8.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of **3**.



Figure S9.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of 18.



Figure S9.2. 13 C-NMR (100 MHz, chloroform-*d*) spectrum of 18.



Figure S10.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of 4.



Figure S10.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 4.



Figure S11.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of 19.



Figure S11.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 19.



Figure S12.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of the crude.



Figure S12.2. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of first eluting *cis* isomer.



Figure S12.3. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of first eluting *cis* isomer.



Figure S12.4. ¹H-NMR (400 MHz, chloroform-d) spectrum of first eluting *trans* isomer.



Figure S12.5. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of first eluting *tran* isomer.



Figure S12.6. ¹H-NMR(400 MHz, chloroform-*d*) spectrum of second eluting *cis* isomer.



Figure S12.7. ¹³C-NMR(100 MHz, chloroform-*d*)spectrum of second eluting *cis* isomer.



Figure S12.8. ¹H-NMR(400MHz, chloroform-*d*)spectrum of second eluting *trans* isomer.



Figure S12.9. ¹³C-NMR(100MHz,chloroform-d) spectrum of second eluting *trans* isomer.



Figure S13.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of crude 20.



Figure S13.2. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of first eluting *trans* isomer of **20**.



Figure S13.3. ¹³C-NMR(100 MHz, chloroform-*d*) spectrum of first eluting *trans* isomer of 20.



FigureS13.4. ¹H-NMR(400MHz,chloroform-*d*)spectrum of second eluting *trans* isomer of 20.



FigureS13.5.¹³C-NMR(100MHz,chloroform-*d*)spectrum of second eluting *trans* isomer of 20.



Figure S13.6. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of first eluting *cis* isomer of **20**.



Figure S13.7. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of first eluting *cis* isomer of 20.



Figure S13.8. ¹H-NMR(400 MHz, chloroform-*d*) spectrum of second eluting *cis* isomer of **20**.



Figure S13.9. ¹³C-NMR(100 MHz, chloroform-*d*)spectrum of second eluting *cis* isomer of 20.



Figure S20.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of compound 5.



Figure S20.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of compound 5.



Figure S21.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of compound 6.



Figure S21.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of compound 6.



Figure S22.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of compound 21.



Figure S23.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of compound **7**.



Figure S23.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of compound 7.



Figure S24.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum.



Figure S24.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum.



Figure S25.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of compound 8.



Figure S25.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of compound **8**.



Figure S26.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of compound 22.



Figure S27.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of compound 9.



Figure S27.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of compound 9.



Figure S28.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum.



Figure S29.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 23.



Figure S30.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum.



Figure S30.3. ¹³C-NMR(100 MHz, chloroform-*d*) spectrum of first eluting *trans* isomer.



Figure S30.5. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of first eluting *cis* isomer.



Figure S30.7.¹³C-NMR (100MHz, chloroform-*d*) spectrum of second eluting *trans* isomer.



Figure S30.9. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of second eluting *cis* isomer.



Figure S31.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 24.



Figure S32.1. ¹H-NMR (400 MHz, chloroform-*d*) spectrum of 10.



Figure S32.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 10.



Figure S33.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum.



Figure S34.2. ¹³C-NMR (100 MHz, chloroform-*d*) spectrum of 11.