A new and more powerfully activating diamine for practical and scalable enantioselective aldehyde crotylsilylation reactions

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Supporting Information

General Information. All reactions were carried out under an atmosphere of nitrogen in flamedried glassware with magnetic stirring unless otherwise indicated. Degassed solvents were purified by passage through an activated alumina column. Thin-layer chromatography (TLC) was carried out on glass backed silica gel TLC plates (250 mm) from Silicycle; visualization by UV light and/or phosphomolybdic acid (PMA). HPLC analysis was carried out on an Agilent 1200 Series using either a Chiralpak AD-H $(250 \times 4.5 \text{ mm ID})$ column or Chiralcel OD-H $(250 \times 4.5 \text{ mm ID})$ column. ¹H NMR spectra were recorded on a Bruker AVIII 300 (300 MHz), AVIII 400 (400 MHz), AVIII 500 (500 MHz) or AVIII 500 Ascend (500 MHz) spectrometer and are reported in ppm, relative to residual protonated solvent peak (CDCl3, 7.26 ppm; C_6D_6 , 7.16 ppm). Data are reported as follows: (bs= broad singlet, s = singlet, d = doublet, t = triplet, $m =$ multiplet, $dd =$ doublet of doublets, $dd =$ doublet of doublet of doublets, $dd =$ doublet of doublet of triplets, td = triplet of doublets; coupling constant(s) in Hz; integration). Proton decoupled ¹³C NMR spectra were recorded on a Bruker AVIII 400 (100 MHz), AVIII 500 (126 MHz) or AVIII 500 Ascend (126 MHz) spectrometer and are reported in ppm from CDCl₃ internal standard (77.23 ppm). ²⁹Si NMR spectra were recorded on a Bruker AVIII 400 (79 MHz) or AVIII 500 (100 MHz) and are reported in ppm relative to TMS (0.00 ppm) internal standard. ¹⁹F NMR spectra were recorded on a Bruker AVIII 400 (376 MHz) and are reported in ppm relative to α, α, α -trifluorotoluene (-63.72 ppm) internal standard. Infrared spectra were recorded on a Nicolet Avatar 370DTGS FT-IR. Optical rotations were recorded on a Jasco DIP-1000 digital polarimeter. (APCI)-MS was conducted on a JMS-LCmate LCMS (JEOL). Melting points were determined using a Stanford Research Systems DigiMelt apparatus.

Preparation of diaminophenol 6:

Compound 13 was prepared using a modified literature procedure.¹ To a cooled (0 $^{\circ}$ C) solution of conc. HCl (32.4 mL, 394 mmol) in MeOH (126 mL) was added (*R,R*)-diaminocyclohexane (45.0 g, 394 mmol). The ice water bath was removed, and after 15 min, water (42.0 mL) was added. After 30 min, a solution of Boc2O (90.6 mL, 394 mmol) in MeOH (42 mL) was added slowly and the resulting mixture was stirred for 12 h. The mixture was concentrated and the residue was resuspended in $Et₂O$ (100 mL), and collected by filtration, rinsing with $Et₂O$ to remove any unprotected diaminocyclohexane. The resulting residue was treated with 3 N NaOH (285 mL) and extracted with CH_2Cl_2 (3 x 125 mL). The combined organic layers were dried over MgSO4, filtered and concentrated to give **13** as a beige solid (55.2 g, 257 mmol, 65%) that was used without further purification. ¹H NMR (400 MHz, CDCl₃) δ 4.43 (bs, 1H), 3.23-2.95 (m, 1H), 2.31 (td, *J* = 10.4, 4.0 Hz, 1H), 2.06-1.88 (m, 2H), 1.78-1.63 (m, 2H), 1.45 (s, 9H), $1.38-0.99$ (m, $6H$). The $1H$ NMR spectroscopic data is in agreement with data reported in literature. $1,2$

Aldehyde **14** was prepared using a modified literature procedure.³ To a solution of 2-*tert*-butyl phenol (30.6 mL, 200 mmol) in acetonitrile (400 mL) was added paraformaldehyde (40.4 g, 1.35 mol), $MgCl₂$ (28.6 g, 300 mmol) and Et₃N (104.4 mL, 748 mmol). The resulting mixture was heated to reflux and stirred for 5 h. The mixture was cooled to room temperature and 5% HCl (200 mL) was added. The mixture was extracted with CH₂Cl₂ (3 x 125 mL). The combined organic layers were concentrated and the residue was partitioned between Et₂O (250 mL) and H₂O (250 mL). The layers were separated and the Et₂O layer was washed with brine $(1 \times 100 \text{ mL})$, dried over Na₂SO₄, filtered and concentrated. The resulting oil was vacuum distilled (bp ~73 °C @ ~5 mm Hg) to give aldehyde **14** as a pale yellow oil (20.7 g, 116 mmol, 58% yield). ¹**H NMR** (300 MHz, CDCl₃) δ 11.78 (s, 1H), 9.88 (s, 1H), 7.53 (dd, *J* = 7.7, 1.5 Hz, 1H), 7.40 (dd, $J = 7.7$, 1.7 Hz, 1H), 6.95 (t, $J = 7.7$ Hz, 1H), 1.43 (s, 9H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.³

⁽¹⁾ D. W. Lee, H. Ha and W. K. Lee, *Synth. Commun.*, 2007, **37**, 737-742.

⁽²⁾ X. Zhang, T. J. Emge and K. C. Hultzsch, *Angew. Chem. Int. Ed.*, 2012, **51**, 394-398.

⁽3) N. Gisch, J. Balzarini and C. Meier, *J. Med. Chem.*, 2007, **50**, 1658-1667.

To a solution of compound **13** (24.9 g, 116 mmol) in EtOH (1.1 L), was added aldehyde **14** (20.7 g, 116 mmol). The mixture was heated to reflux and stirred for 3 h. The mixture was cooled to room temperature and concentrated. The residue was recrystallized from minimal boiling EtOH to give imine **15** as long yellow crystals (39.2 g, 105 mmol, 90% yield). **m.p.** 153-155 °C; **1 H NMR** (400 MHz, CDCl3) δ 13.73 (bs, 1H), 8.35 (s, 1H), 7.32 (dd, *J* = 7.8, 1.7 Hz, 1H), 7.11 (dd, *J* = 7.6, 1.7 Hz, 1H), 6.81 (t, *J* = 7.6 Hz, 1H), 4.37 (bs, 1H), 3.73-3.40 (m, 1H), 3.04 (bs, 1H), 2.18-2.02 (m, 1H), 1.98-1.87 (m, 1H), 1.87- 1.64 (m, 3H), 1.46-1.30 (m, 3H) 1.45 (s, 9H), 1.31 (s, 9H); **13C** NMR (101 MHz, CDCl3) δ 165.02, 160.64, 155.39, 137.55, 129.85, 129.44, 118.87, 117.82, 79.49, 77.43, 72.87, 54.55, 35.02, 33.64, 32.00, 29.57, 28.37, 25.06, 24.31; **IR** (thin film, cm-1) 3437, 3338, 2936, 2859, 2243, 1688, 1630, 1505, 1436, 1390, 1365, 1267, 733; **HRMS (FAB+)**: calcd for C₂₂H₃₅O₃N₂ [M+H]⁺: 375.2648, found 375.2654.

A 2-L roundbottom flask equipped with an addition funnel was charged with LiAlH4 (13.0 g, 343 mmol) and THF (600 mL) and the resulting mixture was cooled to 0 °C. A solution of **15** (42.8 g, 114 mmol) in THF (300 mL) was added slowly *via* the addition funnel, with a THF rinse (100 mL). The mixture was warmed to room temperature and stirred for 2 h. The addition funnel was replaced with a reflux condenser and the mixture was heated to reflux for 12 h. The mixture was cooled to 0 °C, and the reaction was quenched by the *CAREFUL* and *SLOW* addition of water (100 mL). The mixture was extracted with Et₂O (5 x 100 mL). The combined organic layers were dried over MgSO₄, filtered and concentrated. The resulting beige solid was purified by recrystallization from minimal boiling hexanes to give diaminophenol (*R*,*R*)-**6** as white crystals (30.2 g, 104 mmol, 91% yield). **m.p.** 114-116 °C; **¹ H NMR** (400 MHz, CDCl3) δ 7.17 (dd, *J* = 7.9, 1.6 Hz, 1H), 6.92-6.80 (m, 1H), 6.70 (t, *J* = 7.6 Hz, 1H), 4.01 (d, *J* = 13.4 Hz, 1H), 3.83 (d, *J* = 13.5 Hz, 1H), 2.39 (s, 3H), 2.24-2.08 (m, 4H), 1.82-1.64 (m, 2H), 1.42 (s, 9H), 1.31-1.11 (m, 3H), 1.03-0.85 (m, 1H); ¹³C **NMR** (126 MHz, CDCl₃) δ 157.49, 136.88, 126.27, 125.77, 124.47, 118.16, 62.46, 62.22, 51.02, 34.85, 33.59, 31.24, 31.16, 29.74, 25.33, 24.83; **IR** (thin

film, cm-1) 3299, 3217, 2927, 2854, 2797, 2641, 1590, 1430, 1352, 1240, 1084, 747; **HRMS (FAB+)**: calcd for $C_{18}H_{31}ON_2$ [M+H]⁺: 291.2436, found 291.2443.

Preparation of *cis***-crotyltrichlorosilane 10 and** *trans***-crotyltrichlorosilane 11:**

$$
\begin{array}{cccc}\n & & 1 \text{ mol } \% & & \text{Me} \\
\leftarrow & + & \text{HSiCl}_3 & \xrightarrow{\text{THF}, -78 \text{ to } 23 \text{ } ^\circ \text{C}} & & & \text{Me} \\
\end{array}
$$

cis-Crotyltrichlorosilane 10 was prepared using a modified literature procedure.^{4,5} To a cooled (-78 °C) solution of Pd(PPh₃)₄ (0.88 g, 0.76 mmol) in THF (400 mL) was added 1,3-butadiene (50 mL, 573 mmol, condensed into a graduated cylinder cooled to -78 °C) followed by trichlorosilane (38.6 mL, 382 mmol). After 15 min, the cooling bath was removed and the mixture was allowed to warm to room temperature. After 17 h, an aliquot was removed and ¹H NMR analysis showed complete consumption of the trichlorosilane. The reaction flask was fitted with a short-path distillation head and the THF was removed by distillation. The residue was transferred to a distillation-head equipped, 250-mL roundbottom flask with a THF rinse. The THF was removed by distillation and the residue was distilled under reduced pressure (bp ~60 °C ω ~30 mm Hg) to give 10 as a clear and colorless liquid (56.2 g, 297 mmol, 78%) yield). ¹ H NMR spectroscopic analysis revealed that the *cis* to *trans* ratio was ≥99:1. **¹ H NMR** (300 MHz, CDCl₃) δ 5.84-5.62 (m, 1H), 5.54-5.31 (m, 1H), 2.50-2.24 (m, 2H), 1.76-1.58 (m, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.⁵

$$
\begin{array}{ccccccc}\n\text{H}_0 & & & & \text{CI}_3 \text{CCOCCI}_3 & & & & \text{B mol % CUCI} \\
\text{H}_0 & & & & & \text{HSiCl}_3, \text{EI}_3 \text{N} & & & \\
\text{H}_1 & & & & \text{CII} & & \\
\end{array}
$$

trans-Crotyltrichlorosilane 11 was prepared according to a modified literature procedure.^{5,6} *trans*-Crotylalcohol was purchased from Sigma Aldrich as a 19:1 (E:Z) mixture, which was confirmed by ${}^{1}H$ NMR spectroscopic analysis. To a cooled (0 °C) solution of *trans*-crotylalcohol (121 mL, 1.41 mol) in hexachloroacetone (470 mL) was added PPh₃ (386 g, 1.47 mol) portion-wise over 3 h. The ice/water bath was allowed to melt and warm to room temperature slowly, and after 12 h the reaction flask was fitted with a short-path distillation head. Distillation (bp ~85°C @ 760 mm Hg) gave *trans*-crotylchloride as a clear and colorless liquid (83.2 g, 919 mmol, 65% yield). **¹ H NMR** (500 MHz, CDCl3) δ 5.88-5.73 (m, 1H), 5.69-5.52 (m, 1H), 4.06-3.98 (m, 2H), 1.75-1.68 (m, 3H).

⁽4) J. Tsuji, M. Hara and K. Ohno, *Tetrahedron*, 1974, **30**, 2143-2146.

⁽5) K. Iseki, Y. Kuroki, M. Takahashi, S. Kishimoto and Y. Kobayashi, *Tetrahedron*, 1997, **53**, 3513-3526.

⁽6) M. Kira, T. Hino and H. Sakurai, *Tetrahedron Lett.*, 1989, **30**, 1099-1102.

A 2 L round bottom flask equipped with an addition funnel was charged with $Et₃N$ (104 mL, 1.03 mol) and Et₂O (450 mL). The solution was cooled to 0 °C and copper(I) chloride (4.2 g, 43 mmol) was added. A solution of trichlorosilane (104 mL, 1.03 mol) and *trans*-crotylchloride (77.4 g, 855 mmol) in Et₂O (150 mL) was prepared and transferred to the addition funnel. This solution was then added *very slowly* (over the course of 1 h) to the reaction mixture, so as to maintain a reaction mixture temperature near 0 °C. After the addition was complete, the mixture was allowed to warm to room temperature and after 2 h, analysis of an aliquot by ¹H NMR spectroscopy showed complete consumption of the *trans*crotylchloride. The solution was transferred to a 2 L round bottom flask by cannulation through a glass microfiber filter (Grade GF/D) equipped teflon tube $(3/16 \text{ i.d.})$ to filter the Et₃N•HCl salts. The flask was fitted with a distillation head and the Et₂O and Et₃N were removed by distillation. The residue was transferred to a 250 mL pear-shaped flask by cannula with an $Et₂O$ rinse. The flask was fitted with a short-path distillation head and the excess Et₂O was removed by distillation and the residue was distilled (bp ~140-145 °C @ 760 mm Hg) to give *trans*-crotyltrichlorosilane **11** as a clear and colorless liquid (74.4 g, 392 mmol, 46% yield). ¹ H NMR spectroscopic analysis revealed that the *trans* to *cis* ratio was 95:5. ¹**H NMR** (400 MHz, CDCl₃) δ 5.67-5.53 (m, 1H), 5.48-5.32 (m, 1H), 2.31-2.20 (m, 2H), 1.76-1.69 $(m, 3H)$. The ${}^{1}H$ NMR spectroscopic data is in agreement with data reported in literature.⁵

Preparation and characterization of silanes (*S***,***S***)-3 and (***S***,***S***)-8:**

To a cooled (0 °C) solution of (S,S) -1 (1.00 g, 1.8 mmol) in CH₂Cl₂ (7.0 mL) was added DBU (270 µL, 1.8 mmol) followed by freshly distilled phenol (170 mg, 1.8 mmol). The ice/water bath was removed and after 1.5 h, the flask was fitted with a distillation head and the volatiles were removed by distillation. The residue was treated with pentane (10 mL) and the mixture was stirred vigorously for 10 min to ensure complete precipitation of the DBU•HCl salts. The mixture was then filtered through an airfree filter frit, and the filtrate was concentrated to give (*S*,*S*)-**3** as a thick oil, which was analyzed without further purification. ¹**H NMR** (400 MHz, CDCl₃) δ 7.40-7.35 (m, 2H), 7.31-7.27 (m, 2H), 7.25-7.19 (m, 4H), 7.08-6.97 (m, 3H), 6.90-6.84 (m, 2H), 5.04-4.84 (m, 2H), 4.09 (dd, *J* = 15.6, 3.6 Hz, 2H), 3.80 (dd, *J* = 18.8, 15.6 Hz, 2H), 2.87-2.64 (m, 2H), 1.76-1.48 (m, 6H), 1.22-1.03 (m, 2H), 1.02-0.82 (m, 2H); **13C NMR** (101 MHz, CDCl3) δ 154.62, 141.40, 141.29, 132.80, 131.27, 131.23, 129.62, 129.35, 122.00,

120.46, 120.28, 120.15, 115.37, 66.68, 65.99, 48.12, 48.05, 31.19, 30.94, 24.91, 20.27; **29Si NMR** (79 MHz, CDCl₃) δ -25.21.

To a cooled (0 °C) solution of (S,S) -6 (1.00 g, 3.4 mmol) in CH₂Cl₂ (11 mL) was added DBU (1.54 mL, 10.3 mmol). Allyltrichlorosilane (0.55 mL, 3.8 mmol) was then added slowly. The reaction mixture was allowed to warm to room temperature and after 1 h the flask was fitted with a distillation head and the volatiles were removed by distillation. The residue was treated with $Et₂O$ (15 mL), and the resulting mixture was stirred vigorously for 10 min to ensure complete precipitation of the DBU•HCl salts. The mixture was then filtered through an air-free filter frit, and the filtrate was concentrated to give (*S*,*S*)-8 as a thick oil, which was analyzed without further purification. ¹H NMR (500 MHz, CDCl₃) δ 7.23-7.18 (m, 1H), 6.90-6.84 (m, 1H), 6.82 (t, *J* = 7.5 Hz, 1H), 5.95 (ddt, *J* = 16.9, 10.1, 7.8 Hz, 1H), 5.16-5.07 (m, 1H), 5.04-4.95 (m, 1H), 4.23 (d, *J* = 15.6 Hz, 1H), 3.69 (d, *J* = 15.7 Hz, 1H), 2.54-2.42 (m, 1H), 2.47 (s, 3H), 2.13 (ddd, *J* = 10.5, 9.0, 3.3 Hz, 1H), 2.06-1.90 (m, 4H), 1.74-1.63 (m, 2H), 1.39 (s, 9H), 1.26-1.15 (m, 1H), 1.14-1.03 (m, 2H), 0.86-0.73 (m, 1H); **13C NMR** (126 MHz, CDCl3) δ 152.93, 139.64, 135.86, 132.00, 130.16, 129.31, 127.17, 125.92, 121.56, 120.54, 115.69, 64.94, 62.57, 54.65, 48.97, 47.52, 38.13, 34.85, 32.33, 30.26, 30.13, 29.98, 29.94, 29.86, 29.60, 29.31, 24.96, 24.75, 24.29, 19.81, 18.88; ²⁹Si NMR (99 MHz, CDCl₃) δ -15.38; **HRMS (FAB**+): calcd for C₂₁H₃₃ON₂Si [M+H]⁺: 357.2362, found 357.2351.

The stereostructure of **8** was proved by NMR spectroscopic analysis. COSY, HSQC, and HMBC experiments allowed the unambiguous assignment of all of the relevant protons in the ¹H NMR spectrum, and a NOESY experiment revealed the illustrated interactions which are consistent only with the silicon stereochemistry shown in structure (*S*,*S*)-**8** (Supp. Fig. 1). Copies of the COSY, HSQC, HMBC, and NOESY spectra are provided below.

Supplementary Figure 1. NOESY data confims the stereochemical assignment at silicon in (*S*,*S*)-**8**.

General Procedure for the one-pot allylation or crotylation of aldehydes with (*R***,***R***)-6:**

To a cooled (0 °C) solution of (R,R) -6 (1.45 g, 5.0 mmol, 1.1 equiv.) in CH₂Cl₂ (16.5 mL) is added DBU (2.24 mL, 15.0 mmol, 3.3 equiv). The allyltrichlorosilane **9** or *cis*-crotyltrichlorosilane **10** or *trans*crotyltrichlorosilane **11** (**9**: 0.80 mL, **10** or **11**: 0.84 mL, 5.5 mmol, 1.2 equiv) is then added slowly. The ice/water bath is removed and after 1 h the mixture is recooled to 0 °C. The aldehyde (4.5 mmol, 1.0 equiv) is added and the solution is maintained at 0 °C for 1 h. The progress of the reaction may be monitored by TLC, 1 h is generally sufficient for full conversion.

(A) General procedure for the acidic workup with recovery of 6: The mixture is concentrated and the residue is suspended in $Et₂O$ (27.0 mL). The mixture is stirred vigorously for 20 min to ensure complete precipitation of the DBU•HCl salts. The mixture is then filtered through a frit, and the filtrate is treated with *n*-Bu4NF (5.0 mL, 1 M in THF, 5.0 mmol, 1.1 equiv). After 2 h, 1 M aqueous HCl (25.0 mL, 25.0 mmol, 5.5 equiv) is added and the mixture is transferred to a separatory funnel. The layers are separated and the aqueous layer is extracted with $Et₂O$ (3 x 50.0 mL). The combined organic layers are washed with H₂O (2 x 25.0 mL) and saturated aqueous NaHCO₃ (1 x 25.0 mL), dried over MgSO₄, filtered and concentrated. The residue is purified by chromatography on silica gel to provide the allylation or crotylation product, and we have found that a simple filtration through a pad of silica is generally sufficient.

Recovery of 6: The combined aqueous layers from above are treated with 1 M aqueous NaOH (50) mL, 50 mmol) and extracted with CH₂Cl₂ (5 x 50.0 mL). The combined organic layers are washed with water (2 x 25.0 mL), dried over MgSO₄, filtered and concentrated. The residue is dissolved in minimal hot 9:1 MeOH:H₂O (during the dissolution process the temperature should not be allowed to exceed 80 $^{\circ}$ C, as the ligand may start to undergo decomposition at higher temperatures). The hot saturated solution is allowed to cool to room temperature, and distilled water (10.0 mL) was added to ensure complete crystallization of **6**. The white solid is collected by filtration through a frit with a cold 1:1 MeOH:H₂O rinse and then dried *in vacuo* (overnight, with gentle warming with an oil bath set to 40 °C) to give recovered diaminophenol **6**.

(B) General procedure for the simplified workup for acid-sensitive substrates without recovery of 6: The reaction mixture is treated with *n*-Bu₄NF (4.5 mL, 1 M in THF, 4.5 mmol, 1.0 equiv) and the ice/water bath is removed. After 30 min the mixture is concentrated and the residue is purified by chromatography on silica gel to provide the allylation or crotylation product.

Aldehydes employed in this study:

The six aldehydes employed in Table 1 are all known compounds and were prepared according to the literature procedures.^{7,8,9,10,11,12}

Data for the products in Table 1:

 \overline{a}

Workup procedure **A** was used. Product **16** was isolated as a colorless oil (83% yield, >99% ee). ¹**H NMR** (400 MHz, CDCl₃) δ 7.42-7.27 (m, 5H), 5.83 (ddt, *J* = 17.2, 10.2, 7.1 Hz, 1H), 5.20-5.04 (m, 2H), 4.56 (s, 2H), 3.92-3.85 (m, *J* = 10.0, 6.7, 3.4 Hz, 1H), 3.52 (dd, *J* = 9.5, 3.4 Hz, 1H), 3.38 (dd, *J* = 9.5, 7.4 Hz, 1H), 2.31 (d, $J = 3.5$ Hz, 1H), 2.29-2.25 (m, 2H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.¹³ The enantiomeric excess was determined by chiral HPLC analysis: OD-H column, 98.5:1.5 hexanes:*i*PrOH, 1 mL/min, 254 nm. The absolute configuration was determined by optical rotation and comparison to the literature value: $[\alpha]_D^{2^2}$ +2.3° (CHCl₃, *c* 2.0); lit: $[\alpha]_D^{23}$ +2.0° (CHCl₃, *c* 2.3) for the (*S*) enantiomer with 94% ee.¹³

⁽⁷⁾ S. Sano, Y. Kobayashi, T. Kondo, M. Takebayashi, S. Maruyama, T. Fujita and Y. Nagao, *Tetrahedron Lett.*, 1995, **36**, 2097-2100.

⁽⁸⁾ S. F. Vanier, G. Larouche, R. P. Wurz and A. B. Charette, *Org. Lett.*, 2010, **12**, 672-675.

⁽⁹⁾ C. L. Flowers and P. Vogel, *Chemistry (Weinheim an der Bergstrasse, Germany)*, 2010, **16**, 14074-14082.

⁽¹⁰⁾ A. B. Smith, T. J. Beauchamp, M. J. LaMarche, M. D. Kaufman, Y. Qiu, H. Arimoto, D. R. Jones and K. Kobayashi, *J. Am. Chem. Soc.*, 2000, **122**, 8654-8664.

⁽¹¹⁾ Kaugars, S. J. Nelson, F. E. Dutton and S. E. Martin, *Synth. Commun.*, 1993, **23**, 797-809.

⁽¹²⁾ A. B. Smith, S. S.-Y. Chen, F. C. Nelson, J. M. Reichert and B. A. Salvatore, *J. Am. Chem. Soc.*, 1995, **117**, 12013-12014.

⁽13) G. E. Keck and D. Krishnamurthy, *Organic Syntheses*, 1998, **75**, 12-18.

Workup procedure **B** was used and product **17** was isolated as a colorless oil (81% yield, 98% ee). **1 H NMR** (500 MHz, CDCl3) δ 5.84 (ddt, *J* = 17.2, 10.2, 7.1 Hz, 1H), 5.17-5.03 (m, 2H), 3.74-3.67 (m, 1H), 3.63 (dd, *J* = 9.9, 3.7 Hz, 1H), 3.46 (dd, *J* = 9.9, 6.9 Hz, 1H), 2.39 (d, *J* = 3.9 Hz, 1H), 2.30-2.19 (m, 2H), 0.91 (s, 9H), 0.08 (s, 6H). The 1 H NMR spectroscopic data is in agreement with data reported in literature.¹⁴ The enantiomeric excess was determined by ¹⁹F NMR (376 MHz, CDCl₃) analysis of the Mosher ester.¹⁵ The absolute configuration was determined by optical rotation and comparison to the literature value: $[\alpha]_D^{2^2}$ +2.4° (CHCl₃, *c* 2.0); lit: $[\alpha]_D^{2^0}$ +1.7° (CHCl₃, *c* 0.24) for the (*S*) enantiomer with 59% ee.¹⁴

⁽¹⁴⁾ W. R. Roush, L. K. Hoong, M. A. J. Palmer, J. A. Straub and A. D. Palkowitz, *J. Org. Chem.* 1990, **55**, 4117-4126. (15) (a) J. A. Dale, D. L. Dull and H. S. Mosher, *J. Org. Chem.*, 1969, **34**, 2543-2549; (b) T. R. Hoye, C. S. Jeffrey and F. Shao, *Nature protocols*, 2007, **2**, 2451-2458.

Workup procedure **B** was used to prepare product **12** (for convenience on small scale, not because it is acid sensitive), which was isolated as a colorless oil (94% yield, 96% ee, 98:2 dr). ¹H NMR (500 MHz, CDCl3) δ 7.43-7.27 (m, 5H), 5.74 (ddd, *J* = 17.2, 10.3, 7.9 Hz, 1H), 5.14-4.98 (m, 2H), 4.55 (s, 2H), 3.65-3.62 (m, 1H), 3.56 (dd, *J* = 9.5, 3.1 Hz, 1H), 3.40 (dd, *J* = 9.5, 7.7 Hz, 1H), 2.40-2.27 (m, 2H), 1.08 (d, $J = 6.8$ Hz, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.¹⁶ The enantiomeric excess and diastereomer ratio were determined by chiral HPLC analysis: OD-H column, 98:2 hexanes:*i*PrOH, 1 mL/min, 254 nm. The assay was developed using a 3:1 *anti*:*syn* mixture of racemic diastereomers. The absolute configuration was determined by optical rotation and comparison to the literature value: $[\alpha]_D^{22} +23.8^\circ$ (CH₂Cl₂, *c* 2.0); lit: $[\alpha]_D^{20} +24.6^\circ$ (CH₂Cl₂, *c* 1.8) for the (S,R) enantiomer with 96% ee.¹⁶

Workup procedure **A** was used to prepare product **18**, which was isolated as a colorless oil (88% yield, 97% ee, 94:6 dr). **¹ H NMR** (400 MHz, CDCl3) δ 7.44-7.27 (m, 5H), 5.90-5.74 (m, 1H), 5.13-5.03 (m, 2H), 4.59-4.50 (s, 2H), 3.71-3.63 (m, 1H), 3.55 (dd, *J* = 9.6, 3.2 Hz, 1H), 3.43 (dd, *J* = 9.6, 7.6 Hz, 1H), 2.43-2.29 (m, 1H), 2.25 (d, $J = 3.3$ Hz, 1H), 1.04 (d, $J = 6.9$ Hz, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.¹⁷ The enantiomeric excess and diastereomer ratio were determined by chiral HPLC analysis: OD-H column, 98:2 hexanes:*i*PrOH, 1 mL/min, 254 nm. The assay was developed using a ~3:1 *anti*:*syn* mixture of racemic diastereomers. The absolute configuration was determined by optical rotation and comparison to the literature value: $[\alpha]_D^2$ -7.1° (CHCl₃, *c* 2.0); lit: $[\alpha]_D^{20}$ -5.6° (CHCl₃, *c* 1.2) for the (*S,S*) enantiomer with 99% ee.¹⁶

⁽¹⁶⁾ B. M. Hackman, P. J. Lombardi and J. L. Leighton, *Org. Lett.*, 2004, **6**, 4375-4377.

⁽17) D. K. Mohapatra, P. P. Das, M. R. Pattanayak and J. S. Yadav, *Chemistry (Weinheim an der Bergstrasse, Germany)*, 2010, **16**, 2072-2078.

Workup procedure **A** was used and product **19** was isolated as a pale yellow oil (92% yield, 93% ee, >99:1 dr). **¹ H NMR** (500 MHz, CDCl3) δ 7.26-7.14 (m, 2H), 6.94-6.78 (m, 2H), 5.78 (ddd, *J* = 17.2, 10.4, 7.6 Hz, 1H), 5.13-4.98 (m, 2H), 4.45 (s, 2H), 3.80 (s, 3H), 3.70 (ddd, *J* = 9.2, 5.7, 4.6 Hz, 1H), 3.66 (m, 1H), 3.61 (ddd, *J* = 9.3, 8.2, 4.4 Hz, 1H), 2.87 (d, *J* = 3.0 Hz, 1H), 2.30-2.21 (m, 1H), 1.81-1.64 (m, 2H), 1.05 (d, $J = 6.8$ Hz, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.¹⁶ The enantiomeric excess and diastereomeric ratio were determined by chiral HPLC analysis of the 3,5-dinitrobenzoate ester derivative: AD-H column, 98:1:1 hexanes:EtOH:MeOH, 1 mL/min, 254 nm. The assay was developed using a ~3:1 *anti*:*syn* mixture of racemic diastereomers. We previously determined that the (R, R) -diastereomer elutes third.¹⁶

Workup procedure **A** was used and product **20** was isolated as a pale yellow oil (82% yield, 98:2 dr). **¹ H NMR** (300 MHz, CDCl3) δ 7.26-7.18 (m, 2H), 7.00-6.79 (m, 2H), 6.03-5.79 (m, 1H), 5.23-5.01 (m, 2H), 4.45 (s, 2H), 3.80 (s, 3H), 3.65-3.52 (m, 2H), 3.46 (dd, *J* = 9.2, 7.1 Hz, 1H), 3.24 (bs, 1H), 2.43- 2.27 (m, 1H), 2.26-2.10 (m, 1H), 1.96-1.79 (m, 1H), 0.91 (d, J = 7.0 Hz, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.¹⁸ The diastereomer ratio was determined by ¹H NMR spectroscopy $(C_6D_6, 400 \text{ MHz})$.

⁽18) T. M. Trygstad, Y. Pang and C. J. Forsyth, *J. Org. Chem.*, 2009, **74**, 910-913.

Workup procedure **A** was used and product **21** was isolated as a pale yellow oil (82% yield, 92:8 dr. **¹ H NMR** (500 MHz, CDCl3) δ 7.26-7.20 (m, 2H), 6.95-6.79 (m, 2H), 5.85 (ddd, *J* = 17.5, 10.5, 7.3 Hz, 1H), 5.10-4.97 (m, 2H), 4.44 (s, 2H), 3.80 (s, 3H), 3.63 (dd, *J* = 9.1, 4.1 Hz, 1H), 3.46 (dd, *J* = 9.1, 6.4 Hz, 1H), 3.39 (t, *J* = 5.7 Hz, 1H), 3.22 (bs, 1H), 2.39-2.24 (m, 1H), 1.99-1.85 (m, 1H), 1.04 (d, *J* = 6.7 Hz, 3H), 0.95 (d, $J = 7.0$ Hz, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.¹⁹ The diastereomer ratio was determined by ¹H NMR spectroscopy (C_6D_6 , 400 MHz).

⁽19) K. Tanaka, Y. Fujimori, Y. Saikawa and M. Nakata, *J. Org. Chem.*, 2008, **73**, 6292-6298.

Workup procedure **A** was used and product **22** was isolated as a pale yellow oil (80% yield, 93:7 dr). **¹ H NMR** (500 MHz, CDCl3) δ 7.26-7.22 (m, 2H), 6.90-6.85 (m, 2H), 5.90 (ddd, *J* = 16.6, 11.0, 8.4 Hz, 1H), 5.10-5.00 (m, 2H), 4.44 (s, 2H), 3.80 (s, 3H), 3.55 (dd, *J* = 9.2, 4.3 Hz, 1H), 3.48 (dd, *J* = 9.2, 7.6 Hz, 1H), 3.39 (bs, 1H), 3.35 (dd, *J* = 8.0, 3.7 Hz, 1H), 2.41-2.29 (m, 1H), 1.97-1.84 (m, 1H), 1.10 (d, *J* $= 6.9$ Hz, 3H), 0.87 (d, $J = 6.9$ Hz, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.²⁰ The diastereomer ratio was determined by ¹H NMR spectroscopy (C₆D₆, 400 MHz).

⁽20) S. R. Chemler and W. R. Roush, *J. Org. Chem.*, 2003, **68**, 1319-1333.

Workup procedure **A** was used and product **23** was isolated as a pale yellow oil (80% yield, 98:2 dr). **¹ H NMR** (400 MHz, CDCl3) δ 7.26-7.22 (m, 2H), 6.90-6.85 (m, 2H), 5.84 (ddt, *J* = 17.2, 10.1, 7.0 Hz, 1H), 5.16-5.02 (m, 2H), 4.44 (s, 2H), 3.84-3.79 (m, 1H), 3.81 (s, 3H), 3.49 (d, *J* = 5.4 Hz, 2H), 2.52 (d, $J = 3.8$ Hz, 1H), 2.29-2.15 (m, 2H), 1.94-1.81 (m, 1H), 0.95 (d, $J = 7.1$ Hz, 3H). The ¹H NMR spectroscopic data is in agreement with data reported in literature.²¹ The diastereomer ratio was determined by ¹H NMR spectroscopy (C_6D_6 , 400 MHz).

⁽21) K. C. Nicolaou, A. P. Patron, K. Ajito, P. K. Richter, H. Khatuya, P. Bertinato, R. A. Miller and M. J. Tomaszewski, *Chem. Eur. J.*, 1996, **2**, 847-868.

Workup procedure **A** was used and product **24** was isolated as a pale yellow oil (80% yield, 94:6 dr). **¹ H NMR** (500 MHz, CDCl3) δ 7.26-7.20 (m, 2H), 6.90-6.85 (m, 2H), 5.63 (ddd, *J* = 17.2, 10.3, 8.7 Hz, 1H), 5.10-4.92 (m, 2H), 4.49-4.39 (m, 2H), 3.81 (s, 3H), 3.57-3.46 (m, 3H), 2.62 (bs, 1H), 2.29 (ddt, *J* = 15.5, 8.8, 6.6 Hz, 1H), 1.92 (ddt, *J* = 7.0, 4.8, 2.4 Hz, 1H), 1.09 (d, *J* = 6.6 Hz, 3H), 0.95 (d, *J* = 7.0 Hz, 3H). The 1 H NMR spectroscopic data is in agreement with data reported in literature.²² The diastereomer ratio was determined by ¹H NMR spectroscopy (C_6D_6 , 400 MHz).

⁽22) E. de Lemos, F.-H. Porée, A. Bourin, J. Barbion, E. Agouridas, M.-I. Lannou, A. Commerçon, J.-F. Betzer, A. Pancrazi and J. Ardisson, *Chemistry (Weinheim an der Bergstrasse, Germany)*, 2008, **14**, 11092-11112.

Workup procedure **A** was used and product **25** was isolated as a pale yellow oil (80% yield, 95:5 dr). **¹ H NMR** (500 MHz, CDCl3) δ 7.26-7.23 (m, 2H), 6.91-6.84 (m, 2H), 5.79 (ddd, *J* = 17.3, 10.3, 8.4 Hz, 1H), 5.14-5.05 (m, 2H), 4.45 (s, 2H), 3.80 (s, 3H), 3.59-3.42 (m, 3H), 2.33-2.19 (m, 2H), 2.01-1.88 $(m, 1H)$, 1.00-0.93 $(m, 6H)$. The $1H$ NMR spectroscopic data is in agreement with data reported in literature.²³ The diastereomer ratio was determined by ¹H NMR spectroscopy (C_6D_6 , 400 MHz).

⁽23) W.-H. Jung, C. Harrison, Y. Shin, J.-H. Fournier, R. Balachandran, B. S. Raccor, R. P. Sikorski, A. Vogt, D. P. Curran and B. W. Day, *J. Med. Chem.*, 2007, **50**, 2951-2966.

Workup procedure **B** was used and product **26** was isolated as a colorless oil (87% yield, 96:4 dr). **1 H NMR** (500 MHz, CDCl3) δ 5.91-5.75 (m, 1H), 5.20-5.10 (m, 2H), 4.37-4.28 (m, 1H), 4.09 (dd, *J* = 7.9, 6.0 Hz, 1H), 3.94-3.86 (m, 1H), 3.53 (t, *J* = 8.1 Hz, 1H), 2.36-2.20 (m, 3H), 1.78 (ddd, *J* = 14.2, 7.4, 3.0 Hz, 1H), 1.72-1.67 (m, 1H), 1.67-1.59 (m, 4H), 0.90 (td, $J = 7.5$, 3.0 Hz, 6H). The ¹H NMR spectroscopic data is in agreement with data reported in literature and comparison of the optical rotation to the literature value of the enantiomer verifies the absolute configuration of 26: $[\alpha]_D^2$ ¹ +8.0° (CHCl₃, *c* 2.0); lit: $[\alpha]_D^{20}$ -6.8° (CHCl₃, *c* 1.5).¹² The diastereomer ratio was determined by ¹H NMR spectroscopy $(C_6D_6, 400 MHz).$

Summary of Computational Method

All computations were performed using Jaguar, version 7.8, Schrodinger, LLC, New York, NY, 2010.

The geometries of the two silanes were optimized using the B3LYP functional and the 6-31G^{**} basis set. Effective potentials (LACVP) were used for the Br atoms. At the optimized geometries the wavefunctions were recalculated using the cc-pVTZ basis. We include the Cartesian coordinates and final total energies for both optimized geometries below.

B3LYP/6-31G**/LACVP final total energy: -1625.001295 hartrees
B3LYP/cc-pVTZ final total energy: -6747.602235 hartrees final total energy: -6747.602235 hartrees

final geometry:

		angstroms	
atom	x	у	\mathbf{z}
C1	-0.4982009010	-0.5640678668	0.8474852299
C ₂	-0.1576222468	-0.1958625197	2.1534727283
C ₃	1.0727613256	0.4195061570	2.4103339770
C4	1.9480308209	0.6792080770	1.3545975611
C ₅	1.6100992271	0.3226800174	0.0485496634
C ₆	0.3863010488	-0.3031291056	-0.1978759771
07	-1.0334436822	-0.4956775657	3.1641396989
Si8	-1.7700455138	0.4549984989	4.3390466916
C ₉	-2.0418846886	2.1770231235	3.6001871698
C10	-2.9453582217	2.2023827073	2.3981006494
C11	-4.1739416179	2.7254940397	2.3657413176
N12	-0.9690280043	0.5010854285	5.8820652026
C13	-0.0182402632	1.5211069219	6.2959736263

Summary of Results for (*S***,***S***)-8:**

B3LYP/6-31G** final total energy: -1293.855729 hartrees B3LYP/cc-pVTZ final total energy: -1294.193939 hartrees

¹H NMR

²⁹Si NMR

-25.21

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