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GGTTTTTCTTTTGACTATTCCCATCGCAGAAAACGACGCATCA
TCTTTAACGATGCGCGAATATTTAACTTGAACAAGCGGAAAT
AAATAGAGCAGCTATTAGATTATTCTTATGTTGGGTCTATTAA
GGTTATGTTAATTGTTAGTTTGCTATGCTAGTAGTAGATTGTA
TAAATGTTTATGGTCACAAATG→
    pagP
  
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SlyA binding site consensus: (t/g)T(g/a)GCAA(g/t)C(a/t)AA

Supplementary Figure S6. Complementation of Δhns and $\Delta slyA$ mutants. (A) *E. coli* BW25113 pBAD33, BW25113 Δhns *pagP-lacZ* pBAD33, and BW25113 Δhns *pagP-lacZ* pBAD33-hns were grown to log-phase in M63B1 0.4% glucose, then induced in M63B1 0.4% glycerol 0.2% arabinose. After 2 h of induction, expression of *pagP* was assessed by measuring β-galactosidase activity (miller units). (B) Promoter region of the *pagP* gene, showing putative SlyA binding sites (boxes) and SlyA binding site consensus. (C) BW25113 pCA24N, BW25113 $\Delta slyA$ *pagP-lacZ* pCA24N and BW25113 $\Delta slyA$ *pagP-lacZ* pCA24N-slyA were grown in planktonic cultures overnight in M63B1 0.4% glucose (Pk) or in biofilm for 96 h (Bf) in M63B1 0.4% glucose 0.1 mM isopropyl-β-d-thiogalactopyranoside (IPTG). The average values ± SD of three independent experiments are shown. Statistical significance was assessed using a two-tailed unpaired *t*-test (* p < 0.05).