

SUPPLEMENTARY MATERIAL

Fig. S1. The expression of Notch targets is suppressed in endocycling follicle cells where E(y)1 function is compromised. (A,A') Induction of an independent *e(y)I*-RNAi transgene suppressed *hnt* expression in the *e(y)I*-RNAi follicle cells of a stage-7 egg chamber, marked by the expression of RFP (red, A). (B,B') Expression of *brE-lacZ* was abolished in mid-stage follicle cells with *e(y)I* depleted [expressing GFP (red, B)].

Fig. S2. The M/E switch is disrupted in the *e(y)I*-RNAi follicle cells. (A,A') The nuclei of *e(y)I*-depleted follicle cells (expressing RFP) in a stage-8 egg chamber are much smaller and more crowded than those of their neighboring wild-type cells (lacking RFP). (B,B') In an egg chamber at stage 8, PH3 was detected in the *e(y)I*-knockdown follicle cells (expressing RFP), but not in wild-type cells (lacking RFP). (C,C') After stage 7, CycB, which is normally suppressed in wild-type endocycling follicle cells (lacking RFP), was randomly expressed in the *e(y)I*-RNAi follicle cell clones (expressing RFP). Cell nuclei were co-stained with DAPI (green in A' and blue in B,C).

Fig. S3. Poor viability of *e(y)I* mutant cells in the wing disc and Lateral inhibition defects in *e(y)I*-depleted tissues. (A-A'') Wild-type *FRT19A* mock clones in a wing disc. (B-B'') The sizes of *e(y)I* mutant clones generated in a *Minute*^{+/-} [*M(1)*^{osp/+}] background, marked by the absence of GFP, are extremely small, compared with those of control clones in (A-A''). Mosaic clones were induced by heat-shocking early second instar larvae at 37°C for 1 hour, and wing discs were dissected after 72 hours at 25°C. Cell nuclei were labeled with DAPI in red (A'', B''). (C) An adult scutellum bearing *FRT19A* mock clones had four well-organized bristles (white arrows). (D) A fly with *e(y)I* mutant clones carried an extra bristle in scutellum (yellow arrow, in 8 out of 50 adult flies observed). (E) Portion of an adult wing with *FRT19A* mock clones. (F) Extra vein-like structures (black arrows) observed in an adult wing carrying *e(y)I* mutant clones. (G) Portion of a *C96-Gal4* control wing. (H) Dense wing margin bristles (black arrows) in *C96-Gal4*>*e(y)I*-RNAi flies .

Fig. S4. Notch gain-of-function phenotypes in Su(H)-VP16-expressing tissues. (A-B') *en-Gal4*, *UAS-RFP/+*; *tub-Gal80ts/+* control discs showed normal expression patterns of Notch targets, Cut (A') and Wg (B'). (C-D') Ectopic expression of *Su(H)-VP16* by *en-Gal4* caused wing disc overgrowth and Notch signaling activation, as monitored by expression of Cut (C') and Wg (D') in the posterior compartment of wing discs. (E-H) Although the *C96-Gal4*>*Su(H)-VP16* adult wing did not show obvious overgrowth (F), when compared with the *C96-Gal4* control (E), it displayed partial loss of wing margin bristles (H, red arrows), a typical Notch gain-of-function phenotype.

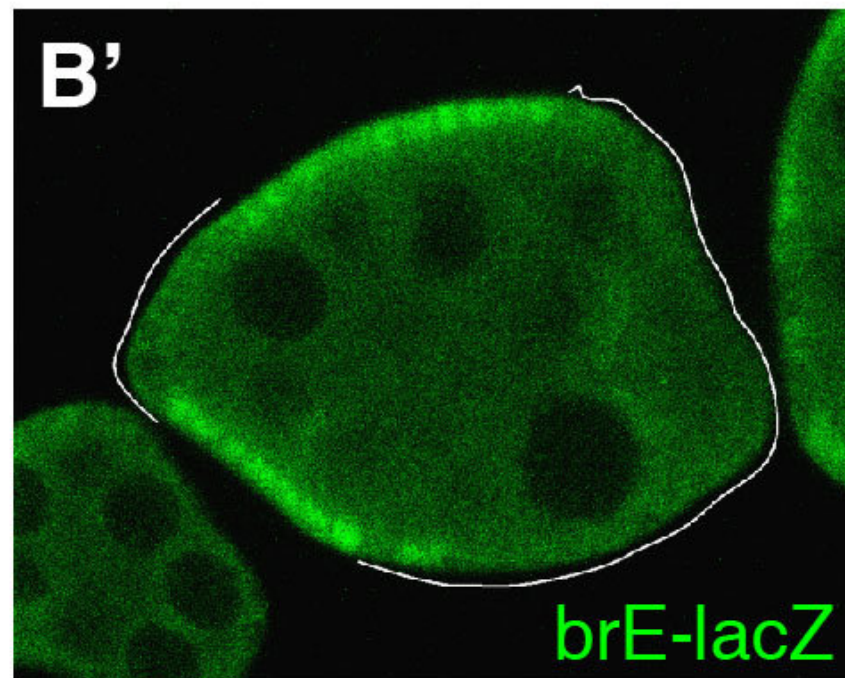
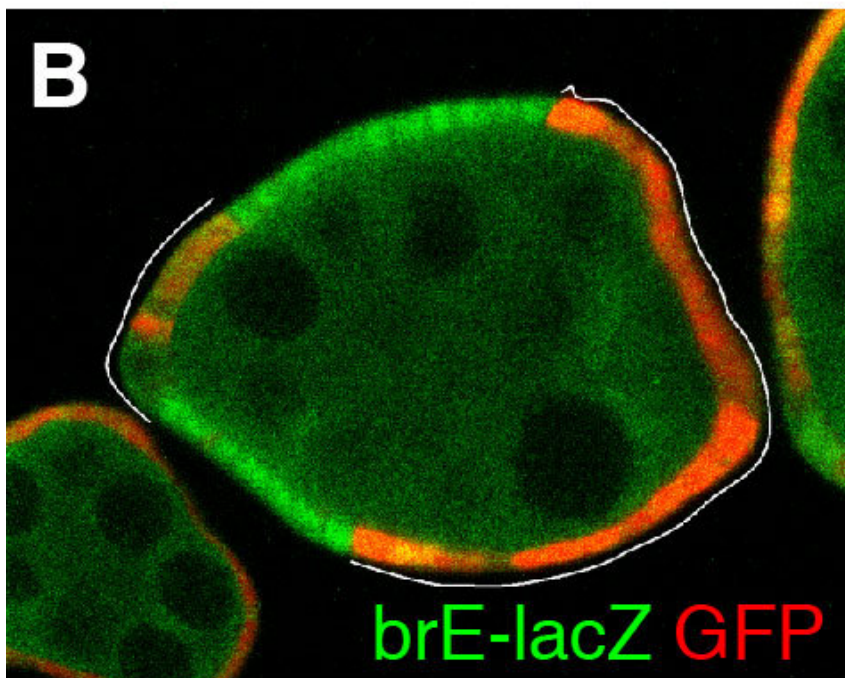
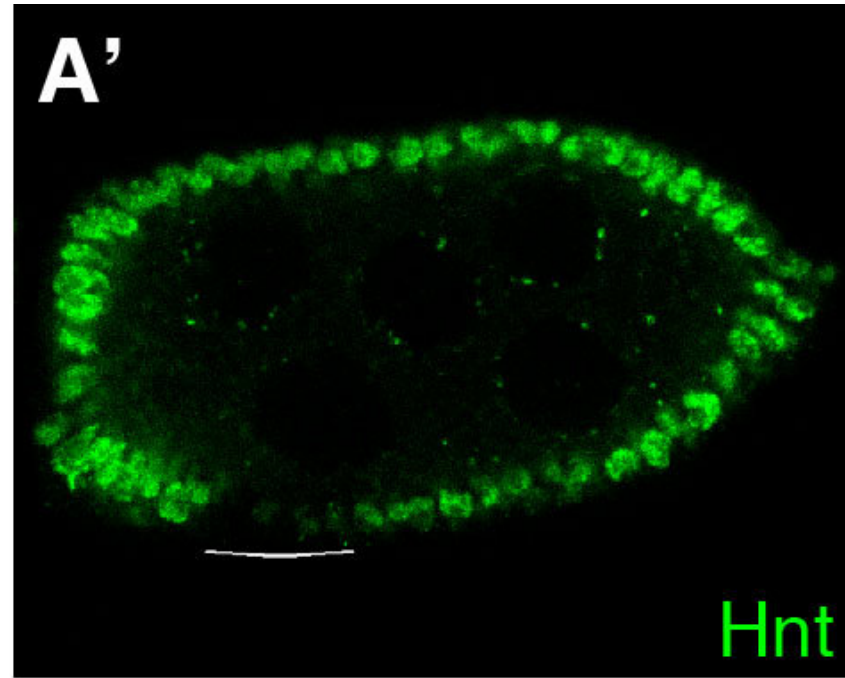
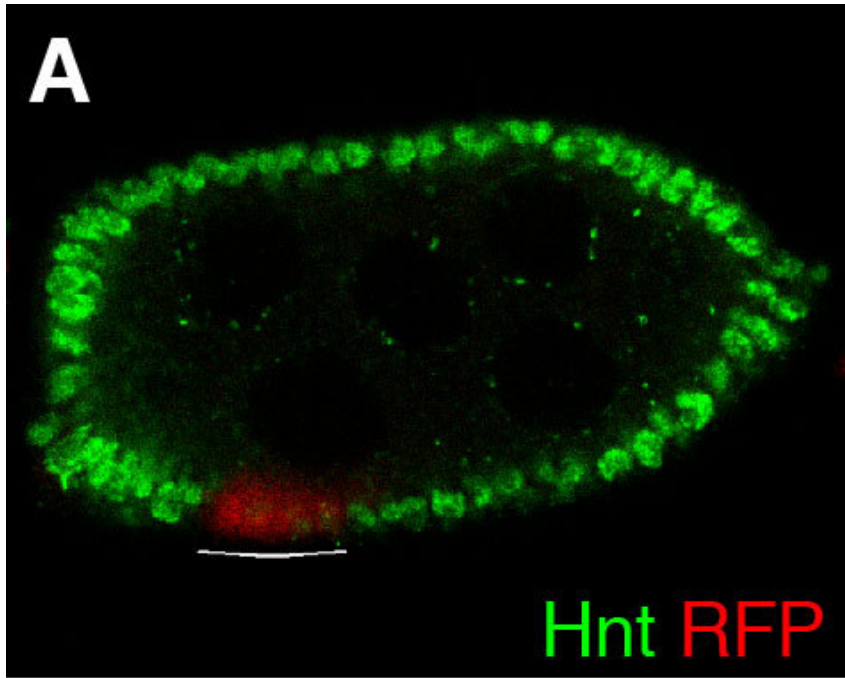


Fig. S1

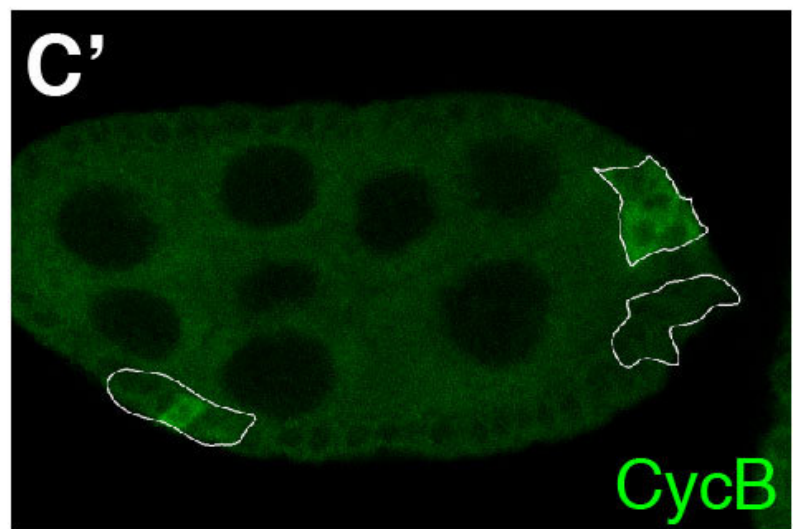
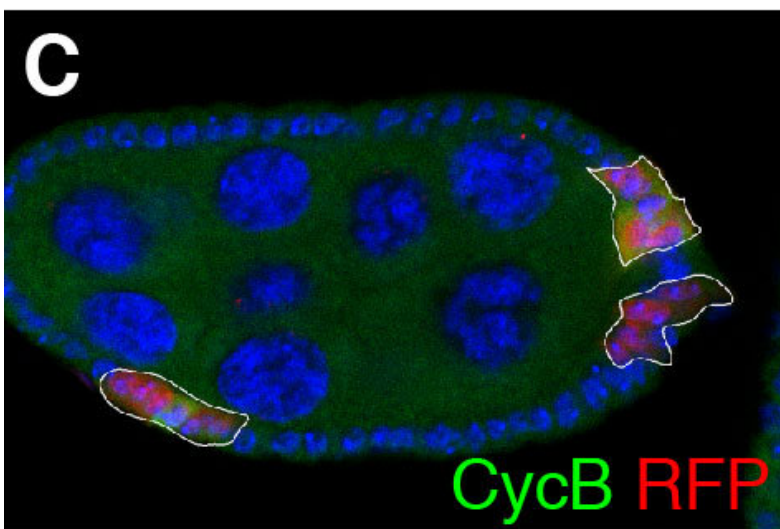
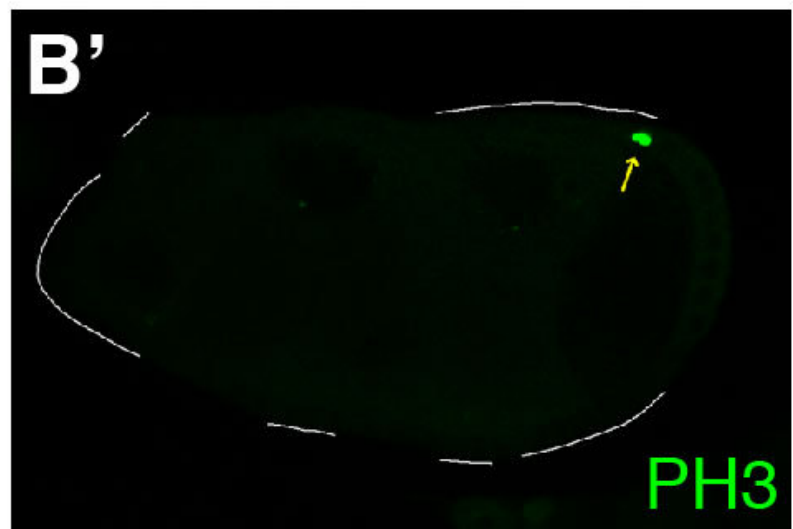
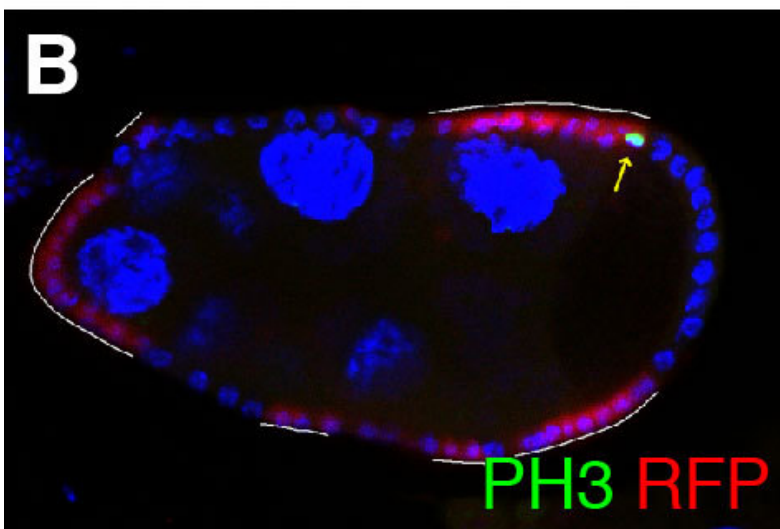
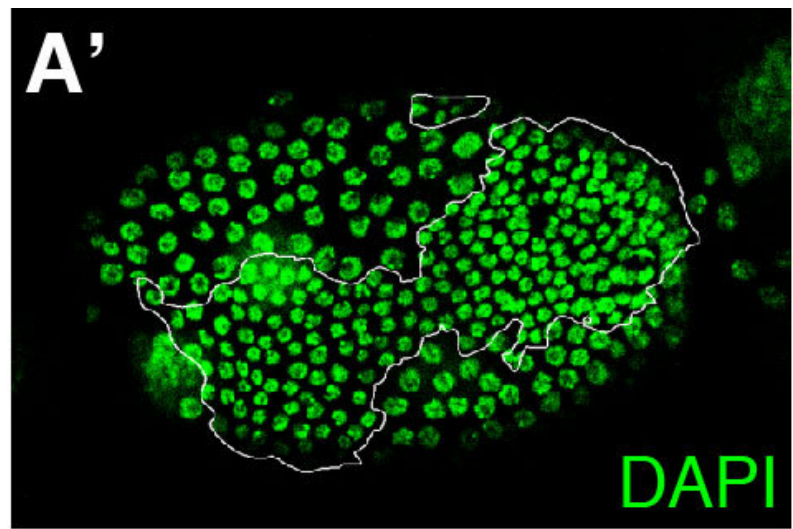
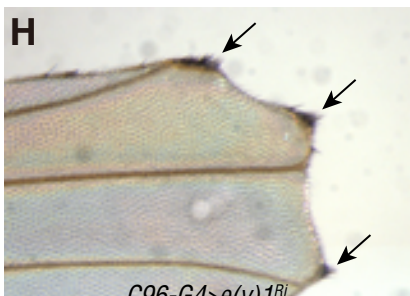
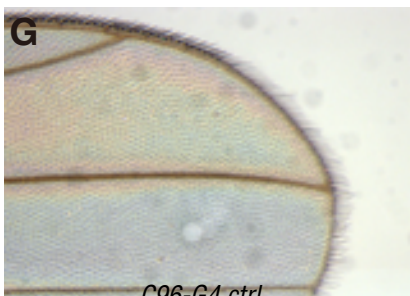
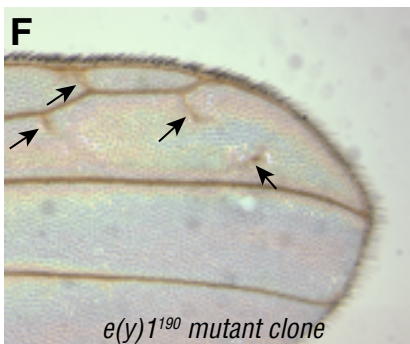
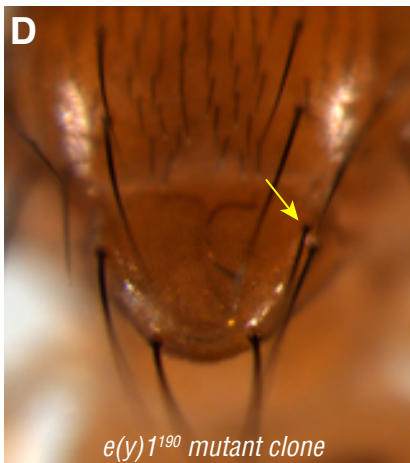
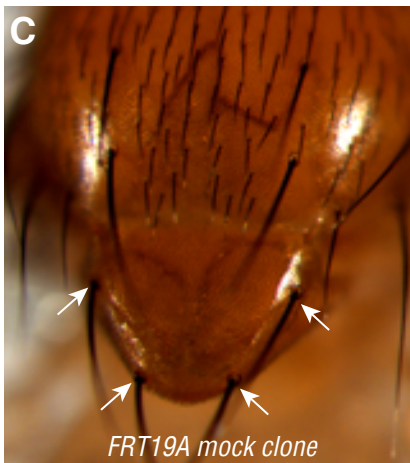
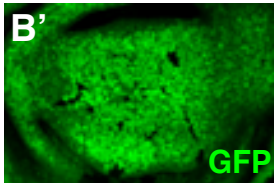
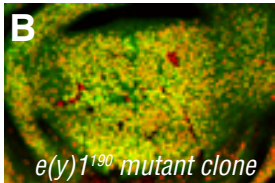
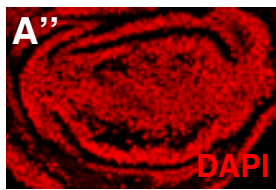
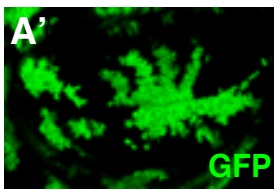
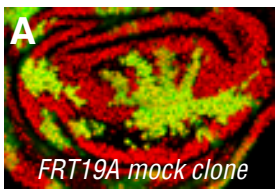
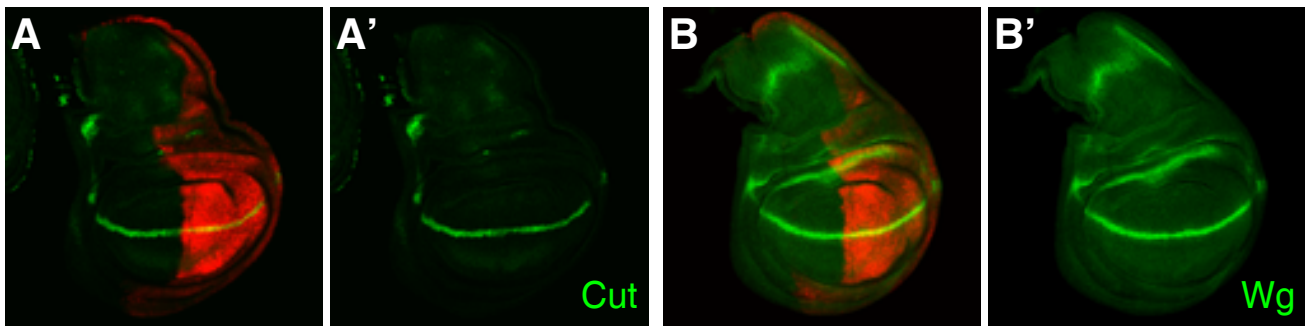


Fig. S2



en-G4, UAS-RFP/+; tub-G80ts/+



en-G4, UAS-RFP/UAS-Su(H)-VP16; tub-G80ts/+

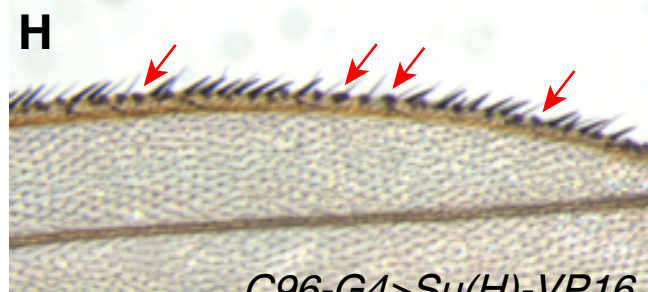
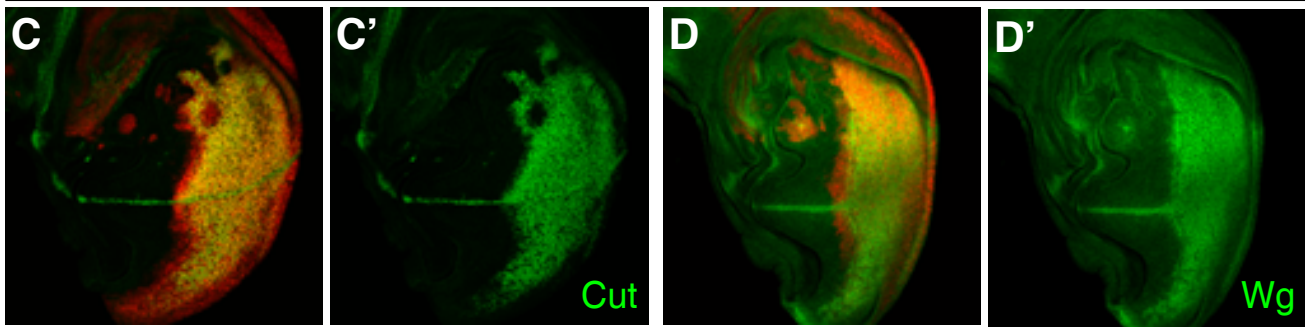


Table S1: Genotypes related to figures

| Figure name | Genotype |
|-------------|---------------------------------------------------------------------------------------------------------------------------|
| Fig. 1A-B' | <i>hsFLP/+;; act>CD2>Gal4, UAS-RFP/UAS-e(y)I^{RNAi-BL32345}</i> (> = FRT in this and all following figures) |
| Fig. 1C,C' | <i>hsFLP/+;E(spl)mβ-CD2/+; act>y⁺>Gal4, UAS-lacZ/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 1D,D' | <i>hsFLP/+; act>y⁺>Gal4, UAS-GFP/+; E(spl)m7-lacZ/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 2B-C'' | <i>ubi-mRFPnls, w, hsFLP¹²² FRT19A/w, hsFLP¹²² e(y)I¹⁹⁰ FRT19A</i> |
| Fig. 2D | <i>w, ubi-GFP,M(1)^{osp} FRT19A/w, hsFLP¹²² e(y)I¹⁹⁰ FRT19A;; +/TM6B</i> |
| Fig. 2E | <i>w, ubi-GFP,M(1)^{osp} FRT19A/w, hsFLP¹²² e(y)I¹⁹⁰ FRT19A;; Dp(1;3)DC335/+</i> |
| Fig. 3A | <i>w^{J118}</i> |
| Fig. 3B | <i>C96-Gal4/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 3C | <i>N²⁶⁴⁻³⁹/+</i> |
| Fig. 3D | <i>N²⁶⁴⁻³⁹/+;; C96-Gal4/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 3E | <i>N²⁶⁴⁻³⁹/P{GT1}e(y)I^{BG00948}</i> |
| Fig. 3F | <i>N²⁶⁴⁻³⁹/e(y)I¹⁹⁰</i> |
| Fig. 3G | <i>N¹/+</i> |
| Fig. 3H | <i>N¹/e(y)I¹⁹⁰</i> |
| Fig. 3I,J | <i>en-Gal4, UAS-GFP/+; tub-Gal80ts/+</i> |
| Fig. 3K,L | <i>en-Gal4, UAS-GFP/+; tub-Gal80ts/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 4A,A' | <i>hsFLP/+;; act>CD2>Gal4, UAS-RFP/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 4B,B' | <i>hsFLP/+; UAS-N^{EXT}/+; act>CD2>Gal4, UAS-RFP/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 4C,C' | <i>hsFLP/+; UAS-N^{CD}/+; act>CD2>Gal4, UAS-RFP/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 4D,D' | <i>hsFLP/+; UAS-Su(H)-VP16/+; act>CD2>Gal4, UAS-RFP/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 4E | <i>C96-Gal4/+</i> |
| Fig. 4F | <i>C96-Gal4/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 4G | <i>UAS-N^{CD}/+; C96-Gal4/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 4H | <i>UAS-Su(H)-VP16/+; C96-Gal4/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. 6A | <i>en-Gal4, UAS-GFP/+; tub-Gal80ts/UAS-TAF12^{RNAi-BL34852}</i> |
| Fig. 6B | <i>en-Gal4, UAS-GFP/+; tub-Gal80ts/UAS-TAF1^{RNAi-BL32421}</i> |
| Fig. 6C | <i>en-Gal4, UAS-GFP/UAS-e(y)2^{RNAi-BL42524}; tub-Gal80ts/+</i> |
| Fig. 6D | <i>C96-Gal4/UAS-TAF12^{RNAi-BL34852}</i> |
| Fig. 6E | <i>C96-Gal4/UAS- UAS-TAF1^{RNAi-BL32421}</i> |
| Fig. 6F | <i>UAS-e(y)2^{RNAi-BL42524}/+; C96-Gal4/+</i> |
| Fig. S1A,A' | <i>hsFLP/+; UAS-e(y)I^{NIG-6474R-1}/+; act>CD2>Gal4, UAS-RFP/UAS-dcr2</i> |

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|---------------|-----------------------------------------------------------------------------------------------------|
| Fig. S1B,B' | <i>hsFLP/+; act>y⁺>Gal4, UAS-GFP/+; br-lacZ/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. S2A-C' | <i>hsFLP/+;; act>CD2>Gal4, UAS-RFP/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. S3A-A'' | <i>w, ubi-GFP,M(1)^{osp} FRT19A/w, hsFLP¹²² FRT19A</i> |
| Fig. S3 B-B'' | <i>w, ubi-GFP,M(1)^{osp} FRT19A/w, hsFLP¹²² e(y)I¹⁹⁰ FRT19A</i> |
| Fig. S3C | <i>ubi-mRFPnls, w, hsFLP¹²² FRT19A/w, hsFLP¹²² FRT19A</i> |
| Fig. S3D | <i>ubi-mRFPnls, w, hsFLP¹²² FRT19A/w, hsFLP¹²² e(y)I¹⁹⁰ FRT19A</i> |
| Fig. S3E | <i>w, ubi-GFP,M(1)^{osp} FRT19A/w, hsFLP¹²² FRT19A</i> |
| Fig. S3F | <i>w, ubi-GFP,M(1)^{osp} FRT19A/w, hsFLP¹²² e(y)I¹⁹⁰ FRT19A</i> |
| Fig. S3G | <i>C96-Gal4/+</i> |
| Fig. S3H | <i>C96-Gal4/UAS-e(y)I^{RNAi-BL32345}</i> |
| Fig. S4A-B' | <i>en-Gal4, UAS-RFP/+; tub-Gal80ts/+</i> |
| Fig. S4C-D' | <i>en-Gal4, UAS-RFP/UAS-Su(H)-VP16; tub-Gal80ts/+</i> |
| Fig. S4E,G | <i>C96-Gal4/+</i> |
| Fig. S4F,H | <i>UAS-Su(H)-VP16/+; C96-Gal4/+</i> |