

Stereoconvergent Arylations and Alkenylations of Unactivated Alkyl Electrophiles: The Catalytic Enantioselective Synthesis of Secondary Sulfonamides and Sulfones

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Supporting Information

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I. General Information

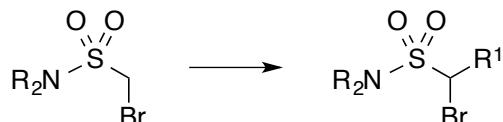
The following reagents were purchased and used as received: $\text{NiCl}_2 \cdot \text{glyme}$ (Strem), ZnI_2 (Strem), and Cp_2ZrHCl (Strem). Ligands **L1** (available from Aldrich) and **L6** were prepared according to a literature procedure.¹ Grignard reagents were prepared from aryl bromides and magnesium turnings (Strem) or from aryl iodides and *i*-PrMgCl (Aldrich; 2.0 M in THF); on occasion, we have found purchased Grignard reagents to be less suitable. THF was deoxygenated and dried by sparging with argon followed by passage through an activated alumina column (S. G. Water) prior to use. All reactions were carried out in oven-dried glassware under an inert atmosphere.

^1H NMR data and ^{13}C NMR data were collected on a VARIAN 500 MHz spectrometer at ambient temperature. HPLC analyses were carried out on an Agilent 1100 series system with Daicel CHIRALPAK® columns or Daicel CHIRALCEL® columns (internal diameter 4.6 mm, column length 250 mm, particle size 5 μm or 3 μm). GC analyses were carried out on an Agilent 6890 series system with an HP-5 column (length 30 m, I.D. 0.25 mm).

(1) Choi, J.; Fu, G. C. *J. Am. Chem. Soc.* **2012**, *134*, 9102–9105.

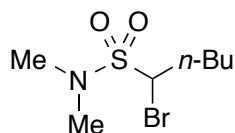
II. Preparation of Electrophiles

These procedures have not been optimized.



Representative experimental procedure for the preparation of α -bromosulfonamides.

LDA was prepared by the dropwise addition of *n*-BuLi (1.6 M in hexanes; 13.8 mL, 22 mmol) to a solution of *i*-Pr₂NH (3.36 mL, 24.0 mmol) in THF (71 mL) in a 500-mL round-bottom flask at -78 °C. The reaction mixture was stirred at 0 °C for 15 min, and then it was cooled to -78 °C. A solution of the 1-bromomethanesulfonamide (20.0 mmol; prepared according to a literature procedure from bromomethanesulfonyl chloride² and a secondary amine³) in THF (40.0 mL) was added over 15 min to the LDA solution at -78 °C. The mixture was stirred for 30 min, and then a solution of the alkyl bromide (26.0 mmol) in THF (43.3 mL) was added over 15 min. The solution was stirred at -78 °C for 2 h, and then it was allowed to slowly warm to r.t. The reaction mixture was stirred at r.t. for 12 h, and then the reaction was quenched by the addition of saturated aqueous NH₄Cl (100 mL). The mixture was extracted with Et₂O (3 × 50 mL), and the combined organic layers were rinsed with brine (50 mL), dried over MgSO₄, and concentrated.



1-Bromo-*N,N*-dimethylpentane-1-sulfonamide. The title compound was prepared from 1-bromo-*N,N*-dimethylmethanesulfonamide (5.00 g, 24.7 mmol) and 1-bromobutane (3.45 mL, 32.2 mmol). The product was purified by column chromatography (3% → 20% ethyl acetate/hexanes): 3.00 g (47%). Colorless oil.

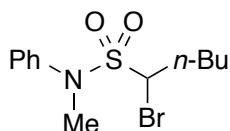
¹H NMR (500 MHz, CDCl₃) δ 4.81 (dd, 1H, *J* = 10.7, 3.1 Hz), 3.03 (s, 6H), 2.34 (dddd, 1H, *J* = 14.4, 10.0, 5.5, 3.1 Hz), 2.10–2.02 (m, 1H), 1.70–1.61 (m, 1H), 1.47–1.30 (m, 3H), 0.94 (t, 3H, *J* = 7.2 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 63.3, 38.8, 32.9, 29.2, 21.9, 13.9.

FT-IR (neat) 2958, 2873, 2814, 1483, 1458, 1435, 1414, 1380, 1342, 1287, 1237, 1203, 1171, 1145, 1106, 1064, 973, 930, 782, 750, 734 cm⁻¹.

MS (EI) *m/z* (M⁺) calcd for C₇H₁₆BrNO₂S: 257, found: 257.

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- (2) Gao, F.; Yan, X.; Zahr, O.; Larsen, A.; Vong, K.; Auclair, K. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5518–5522.
- (3) Brienne, M.-J.; Varech, D.; Leclercq, M.; Jacques, J.; Radembino, N.; Dessalles, M.-C.; Mahuzier, G.; Gueyouche, C.; Bories, C. Loiseau, P.; Gayral, P. *J. Med. Chem.* **1987**, *30*, 2232–2239.



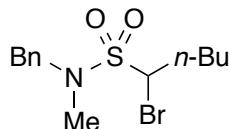
1-Bromo-N-methyl-N-phenylpentane-1-sulfonamide. The title compound was prepared from 1-bromo-N-methyl-N-phenylmethanesulfonamide (3.82 g, 14.5 mmol) and 1-bromobutane (2.02 mL, 18.8 mmol). The product was purified by column chromatography on silica gel (2%→15% ethyl acetate/hexanes) and then on C-18 silica gel (10%→100% acetonitrile/water): 3.60 g (78%). Colorless oil.

^1H NMR (500 MHz, CDCl_3) δ 7.50–7.48 (m, 2H), 7.43–7.39 (m, 2H), 7.35–7.31 (m, 1H), 4.74 (dd, 1H, J = 10.5, 3.1 Hz), 3.52 (s, 3H), 2.27 (dddd, 1H, J = 14.5, 10.2, 5.3, 3.1 Hz), 2.11–2.03 (m, 1H), 1.66–1.58 (m, 1H), 1.40–1.24 (m, 3H), 0.89 (t, 3H, J = 7.2 Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 140.8, 129.7, 128.1, 127.3, 63.3, 42.0, 32.7, 29.1, 21.9, 13.9.

FT-IR (neat) 3062, 3039, 2957, 2931, 2872, 1595, 1493, 1466, 1453, 1436, 1351, 1270, 1237, 1183, 1143, 1106, 1068, 1026, 917, 886, 767, 725 cm^{-1} .

MS (ESI) m/z (M^++H) calcd for $\text{C}_{12}\text{H}_{19}\text{BrNO}_2\text{S}$: 320, found: 320.



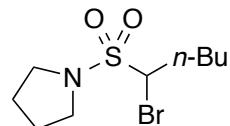
N-Benzyl-1-bromo-N-methylpentane-1-sulfonamide. The title compound was prepared from *N*-benzyl-1-bromo-*N*-methylmethanesulfonamide (3.75 g, 13.5 mmol) and 1-bromobutane (1.88 mL, 17.5 mmol). The product was purified by column chromatography (2%→15% ethyl acetate/hexanes): 2.04 g (45%). Light-yellow oil.

^1H NMR (500 MHz, CDCl_3) δ 7.39–7.35 (m, 4H), 7.34–7.30 (m, 1H), 4.84 (dd, 1H, J = 10.7, 3.1 Hz), 4.61 (d, 1H, J = 14.8 Hz), 4.36 (d, 1H, J = 14.8 Hz), 2.88 (s, 3H), 2.40 (dddd, 1H, J = 14.4, 10.0, 5.6, 3.1 Hz), 2.15–2.07 (m, 1H), 1.72–1.64 (m, 1H), 1.50–1.31 (m, 3H), 0.95 (t, 3H, J = 7.2 Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 135.8, 128.9, 128.3, 128.2, 64.1, 55.5, 35.5, 32.9, 29.3, 21.9, 13.9.

FT-IR (neat) 3088, 3064, 3031, 2958, 2931, 2872, 1605, 1587, 1496, 1467, 1455, 1338, 1278, 1212, 1196, 1151, 1106, 1077, 1029, 994, 944, 910, 858, 787, 733 cm^{-1} .

MS (ESI) m/z (M^++H) calcd for $\text{C}_{13}\text{H}_{21}\text{BrNO}_2\text{S}$: 334, found: 334.



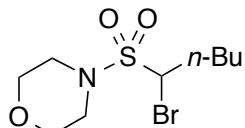
1-((1-Bromopentyl)sulfonyl)pyrrolidine. The title compound was prepared from 1-((bromomethyl)sulfonyl)pyrrolidine (3.02 g, 13.2 mmol) and 1-bromobutane (1.85 mL, 17.2 mmol). The product was purified by column chromatography (2%→15% ethyl acetate/hexanes): 1.96 g (52%). Light-yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 4.84 (dd, 1H, *J* = 10.7, 3.1 Hz), 3.62–3.56 (m, 2H), 3.49–3.43 (m, 2H), 2.36 (dddd, 1H, *J* = 14.4, 10.0, 5.6, 3.1 Hz), 2.11–2.03 (m, 1H), 1.99–1.94 (m, 4H), 1.70–1.61 (m, 1H), 1.48–1.30 (m, 3H), 0.94 (t, 3H, *J* = 7.2 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 63.7, 49.4, 32.7, 29.3, 26.1, 21.9, 13.9.

FT-IR (neat) 2957, 2872, 1461, 1334, 1238, 1200, 1148, 1076, 1014, 929, 781 cm⁻¹.

MS (EI) *m/z* (M⁺) calcd for C₉H₁₈BrNO₂S: 283, found: 283.



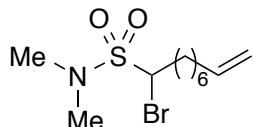
4-((1-Bromopentyl)sulfonyl)morpholine. The title compound was prepared from 4-((bromomethyl)sulfonyl)morpholine (3.01 g, 12.3 mmol) and 1-bromobutane (1.72 mL, 16.0 mmol). The product was purified by column chromatography (2%→20% ethyl acetate/hexanes): 1.28 g (35%). White solid.

¹H NMR (500 MHz, CDCl₃) δ 4.72 (dd, 1H, *J* = 10.7, 3.1 Hz), 3.75–3.68 (m, 4H), 3.50–3.42 (m, 4H), 2.32 (dddd, 1H, *J* = 14.3, 9.9, 5.5, 3.0 Hz), 2.05–1.97 (m, 1H), 1.67–1.58 (m, 1H), 1.45–1.27 (m, 3H), 0.92 (t, 3H, *J* = 7.2 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 67.0, 63.8, 47.3, 32.8, 29.1, 21.8, 13.8.

FT-IR (neat) 2959, 2925, 2860, 1467, 1460, 1450, 1434, 1347, 1328, 1299, 1261, 1237, 1204, 1153, 1114, 1074, 1014, 958, 846, 778, 732 cm⁻¹.

MS (EI) *m/z* (M⁺) calcd for C₉H₁₈BrNO₃S: 299, found: 299.



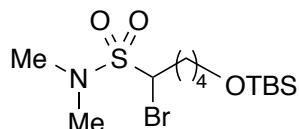
1-Bromo-*N,N*-dimethylnon-8-ene-1-sulfonamide. The title compound was prepared from 1-bromo-*N,N*-dimethylmethanesulfonamide (1.76 g, 8.71 mmol) and 8-bromo-1-octene (1.90 mL, 11.3 mmol). The product was purified by column chromatography (2%→20% ethyl acetate/hexanes): 1.30 g (48%). Colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 5.80 (ddt, 1H, *J* = 16.9, 10.2, 6.7 Hz), 5.00 (ddt, 1H, *J* = 17.1, 2.2, 1.6 Hz), 4.94 (ddt, 1H, *J* = 10.2, 2.2, 1.2 Hz), 4.81 (dd, 1H, *J* = 10.6, 3.1 Hz), 3.03 (s, 6H), 2.33 (dddd, 1H, *J* = 14.3, 10.0, 5.8, 3.1 Hz), 2.10–2.02 (m, 3H), 1.71–1.63 (m, 1H), 1.48–1.25 (m, 7H).

¹³C NMR (126 MHz, CDCl₃) δ 139.0, 114.5, 63.3, 38.8, 33.8, 33.1, 28.8, 28.6, 27.1.

FT-IR (neat) 3075, 2923, 2852, 1640, 1479, 1454, 1414, 1340, 1285, 1204, 1143, 1063, 971, 907, 783 cm⁻¹.

MS (ESI) *m/z* (M⁺+H) calcd for C₁₁H₂₃BrNO₂S: 312, found: 312.



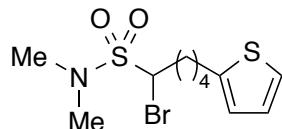
1-Bromo-5-((*tert*-butyldimethylsilyloxy)-*N,N*-dimethylpentane-1-sulfonamide. A 250-mL round-bottom flask was charged with 1-bromo-*N,N*-dimethylmethanesulfonamide (0.808 g, 4.00 mmol) and toluene (24 mL). *tert*-Butyl(4-iodobutoxy)dimethylsilane (5.03 g, 16.0 mmol), aqueous NaOH (50% w/v; 24 mL), and benzyltriethylammonium chloride (0.911 g, 4.00 mmol) were added to the solution at r.t. The resulting mixture was stirred at r.t. for 24 h, and then water (50 mL) was added. The organic phase was separated, and the aqueous solution was extracted with ethyl acetate (2 × 25 mL). The combined organic layers were dried over MgSO₄ and concentrated. The product was purified by column chromatography (hexanes → 30% ethyl acetate/hexanes): 1.21 g (78%). Colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 4.82 (dd, 1H, *J* = 10.7, 3.1 Hz), 3.62 (t, 2H, *J* = 6.1 Hz), 3.02 (s, 6H), 2.38–2.32 (m, 1H), 2.13–2.04 (m, 1H), 1.79–1.70 (m, 1H), 1.67–1.45 (m, 3H), 0.89 (s, 9H), 0.05 (s, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 63.2, 62.6, 38.8, 33.0, 31.8, 26.1, 23.7, 18.5, -5.2.

FT-IR (neat) 2952, 2929, 2885, 2856, 1471, 1462, 1389, 1343, 1287, 1256, 1205, 1146, 1127, 1106, 1006, 973, 939, 836, 812, 776, 740 cm⁻¹.

MS (ESI) *m/z* (M⁺+H) calcd for C₁₃H₃₁BrNO₃SSi: 388, found: 388.



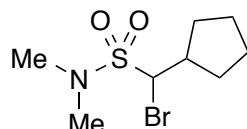
1-Bromo-*N,N*-dimethyl-5-(thiophen-2-yl)pentane-1-sulfonamide. The title compound was prepared from 1-bromo-*N,N*-dimethylmethanesulfonamide (3.00 g, 14.8 mmol) and 2-(4-bromobutyl)thiophene (4.23 g, 19.3 mmol). The product was purified by column chromatography (3% → 20% ethyl acetate/hexanes): 1.44 g (29%). Light-yellow solid.

¹H NMR (500 MHz, CDCl₃) δ 7.12 (dd, 1H, *J* = 5.1, 1.2 Hz), 6.92 (dd, 1H, *J* = 5.1, 3.4 Hz), 6.79 (dddd, 1H, *J* = 3.3, 1.0, 1.0, 1.0 Hz), 4.80 (dd, 1H, *J* = 10.5, 3.2 Hz), 3.02 (s, 6H), 2.92–2.82 (m, 2H), 2.40–2.34 (m, 1H), 2.14–2.07 (m, 1H), 1.80–1.69 (m, 3H), 1.56–1.47 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 144.8, 126.9, 124.4, 123.2, 63.0, 38.8, 33.0, 30.9, 29.7, 26.6.

FT-IR (neat) 2935, 2857, 1480, 1454, 1414, 1340, 1286, 1203, 1180, 1145, 1063, 972, 850, 784 cm⁻¹.

MS (ESI) *m/z* (M⁺+H) calcd for C₁₁H₁₉BrNO₂S₂: 340, found: 340.



1-Bromo-1-cyclopentyl-*N,N*-dimethylmethanesulfonamide. The title compound was prepared from 1-bromo-*N,N*-dimethylmethanesulfonamide (3.03 g, 15.0 mmol) and cyclopentyl

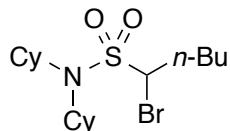
4-methylbenzenesulfonate (4.69 g, 19.5 mmol). The product was purified by column chromatography (10% ethyl acetate/hexanes): 668 mg (16%). Colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 4.99 (d, 1H, *J* = 4.8 Hz), 3.00 (s, 6H), 2.66–2.59 (m, 1H), 1.99–1.88 (m, 2H), 1.75–1.55 (m, 5H), 1.54–1.45 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 68.5, 41.8, 38.7, 31.8, 30.0, 25.6, 25.5.

FT-IR (neat) 2947, 2869, 2812, 1481, 1452, 1413, 1333, 1284, 1205, 1180, 1142, 1063, 969, 898, 862, 786 cm⁻¹.

MS (EI) *m/z* (M⁺–Br) calcd for C₈H₁₆NO₂S: 190, found: 190.



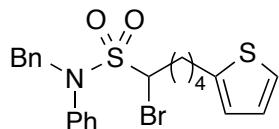
1-Bromo-*N,N*-dicyclohexylpentane-1-sulfonamide. The title compound was prepared from 1-bromo-*N,N*-dicyclohexylmethanesulfonamide (2.10 g, 6.21 mmol) and 1-bromobutane (0.867 mL, 8.07 mmol). The product was purified by column chromatography (1%→8% ethyl acetate/hexanes): 2.06 g (84%). Colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 4.57 (dd, 1H, *J* = 10.6, 2.9 Hz), 3.38–3.33 (br m, 2H), 2.35 (dddd, 1H, *J* = 14.3, 10.1, 5.3, 2.9 Hz), 2.07–2.00 (m, 1H), 1.95–1.91 (m, 2H), 1.86–1.59 (m, 13H), 1.45–1.23 (m, 7H), 1.09 (qt, 2H, *J* = 13.1, 3.5 Hz), 0.92 (t, 3H, *J* = 7.2 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 66.6, 59.3, 33.9, 33.3, 32.4, 29.5, 26.6, 25.4, 22.0, 13.9.

FT-IR (neat) 2931, 2855, 1467, 1454, 1401, 1381, 1329, 1275, 1256, 1235, 1188, 1166, 1142, 1101, 1074, 1048, 1027, 997, 982, 929, 917, 895, 856, 847, 824, 801, 774, 760, 749, 733 cm⁻¹.

MS (EI) *m/z* (M⁺) calcd for C₁₇H₃₂BrNO₂S: 393, found: 393.



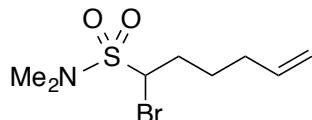
N-Benzyl-1-bromo-*N*-phenyl-5-(thiophen-2-yl)pentane-1-sulfonamide. The title compound was prepared from *N*-benzyl-1-bromo-*N*-phenylmethanesulfonamide (2.70 g, 7.94 mmol) and 2-(4-bromobutyl)thiophene (2.26 g, 10.3 mmol). The product was purified by column chromatography on silica gel (2%→12% ethyl acetate/hexanes) and then preparative HPLC on C-18 silica gel (80%→100% acetonitrile/water; water was doped with 0.1% AcOH): 0.881 g (23%). White solid.

¹H NMR (500 MHz, CDCl₃) δ 7.34–7.27 (m, 5H), 7.26–7.20 (m, 5H), 7.11 (dd, 1H, *J* = 5.1, 1.2 Hz), 6.90 (dd, 1H, *J* = 5.1, 3.4 Hz), 6.76 (dddd, 1H, *J* = 3.2, 1.0, 1.0, 1.0 Hz), 5.34 (d, 1H, *J* = 14.8 Hz), 4.75 (dd, 1H, *J* = 10.5, 3.1 Hz), 4.69 (d, 1H, *J* = 14.9 Hz), 2.88–2.78 (m, 2H), 2.38–2.31 (m, 1H), 2.21–2.13 (m, 1H), 1.79–1.65 (m, 3H), 1.51–1.43 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 144.8, 138.1, 136.3, 129.6, 129.4, 128.7, 128.6, 128.5, 127.9, 126.9, 124.4, 123.2, 63.3, 58.9, 32.7, 30.9, 29.6, 26.6.

FT-IR (neat) 3064, 3031, 2932, 2858, 1594, 1492, 1454, 1439, 1348, 1214, 1178, 1150, 1093, 1066, 1028, 917, 868, 822, 781 cm^{-1} .

MS (ESI) m/z (M^++H) calcd for $\text{C}_{22}\text{H}_{25}\text{BrNO}_2\text{S}_2$: 478, found: 478.



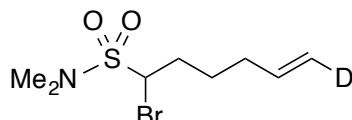
1-Bromo-*N,N*-dimethylhex-5-ene-1-sulfonamide. The title compound was prepared from 1-bromo-*N,N*-dimethylmethanesulfonamide (4.00 g, 19.8 mmol) and 5-bromo-1-pentene (3.05 mL, 25.7 mmol). The product was purified by column chromatography (3%→15% ethyl acetate/hexanes): 2.29 g (43%). Colorless oil.

^1H NMR (500 MHz, CDCl_3) δ 5.79 (ddt, 1H, $J = 16.9, 10.2, 6.7$ Hz), 5.05 (dq, 1H, $J = 17.1, 1.7$ Hz), 5.01 (ddt, 1H, $J = 10.2, 1.9, 1.2$ Hz), 4.82 (dd, 1H, $J = 10.5, 3.2$ Hz), 3.02 (s, 6H), 2.35 (dddd, 1H, $J = 14.5, 10.2, 6.0, 3.2$ Hz), 2.19–2.04 (m, 3H), 1.84–1.75 (m, 1H), 1.60–1.51 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 137.5, 115.8, 63.0, 38.8, 32.8, 32.7, 26.4.

FT-IR (neat) 3076, 2918, 1640, 1482, 1454, 1415, 1341, 1285, 1204, 1143, 1063, 970, 912, 856, 786, 738 cm^{-1} .

MS (ESI) m/z (M^++H) calcd for $\text{C}_8\text{H}_{17}\text{BrNO}_2\text{S}$: 270, found: 270.



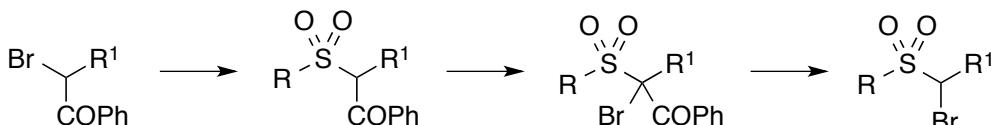
(E)-1-Bromo-*N,N*-dimethylhex-5-ene-1-sulfonamide-6-d. The title compound was prepared from 1-bromo-*N,N*-dimethylmethanesulfonamide (762 mg, 3.77 mmol) and (*E*)-pent-4-en-1-yl-5-*d* 4-methylbenzenesulfonate (1.18 g, 4.90 mmol). The product was purified by column chromatography (2%→20% ethyl acetate/hexanes): 408 mg (40%). Colorless oil.

^1H NMR (500 MHz, CDCl_3) δ 5.78 (dt, 1H, $J = 16.9, 6.5$ Hz), 5.05–5.00 (m, 1H), 4.82 (dd, 1H, $J = 10.5, 3.2$ Hz), 3.01 (s, 6H), 2.37–2.30 (m, 1H), 2.18–2.03 (m, 3H), 1.83–1.74 (m, 1H), 1.59–1.50 (m, 1H).

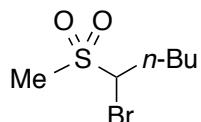
^{13}C NMR (126 MHz, CDCl_3) δ 137.4, 115.5 (t, $J = 24$ Hz), 63.1, 38.8, 32.8, 32.7, 26.4.

FT-IR (neat) 3028, 2949, 2862, 2264, 1621, 1483, 1455, 1435, 1414, 1342, 1287, 1204, 1183, 1144, 1064, 972, 868, 785, 744 cm^{-1} .

MS (ESI) m/z (M^++H) calcd for $\text{C}_8\text{H}_{16}\text{DBrNO}_2\text{S}$: 271, found: 271.



Representative experimental procedure for the preparation of α -bromosulfones. The target molecules were prepared according to literature procedures from α -bromoketones.^{4,5} A 100-mL round-bottom flask was charged with the α -bromo- β -keto-sulfone (10.0 mmol) and aqueous KOH (30% w/v; 50 mL), and the mixture was stirred at r.t. for 48 h. When the reaction was complete (monitored by TLC), the reaction mixture was extracted with dichloromethane (3 \times 30 mL). The combined organic layers were dried over MgSO_4 and concentrated.



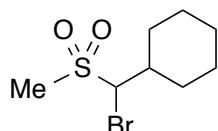
1-Bromo-1-(methylsulfonyl)pentane. The title compound was prepared from 2-bromo-2-(methylsulfonyl)-1-phenylhexan-1-one (12.0 g, 36.0 mmol). The product was purified by column chromatography (hexanes \rightarrow 20% ethyl acetate/hexanes): 8.08 g (98%). White solid.

^1H NMR (500 MHz, CDCl_3) δ 4.61 (dd, 1H, J = 11.0, 3.0 Hz), 3.09 (s, 3H), 2.43 (dd, 1H, J = 14.4, 9.9, 5.7, 3.0 Hz), 1.96 (dd, 1H, J = 14.2, 11.1, 9.5, 4.4 Hz), 1.72–1.64 (m, 1H), 1.50–1.31 (m, 3H), 0.94 (t, 3H, J = 7.2 Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 64.3, 37.6, 30.1, 29.2, 21.8, 13.8.

FT-IR (neat) 3010, 2958, 2932, 2873, 1467, 1454, 1434, 1413, 1381, 1311, 1237, 1208, 1140, 1121, 1106, 956, 928, 815, 771, 748, 735 cm^{-1} .

MS (ESI) m/z (M $^+$ +H) calcd for $\text{C}_6\text{H}_{14}\text{BrO}_2\text{S}$: 229, found: 229.



(Bromo(methylsulfonyl)methyl)cyclohexane. The bromination of 2-cyclohexyl-2-(methylsulfonyl)-1-phenylethan-1-one was conducted at 60 °C, and extra KBr and H_2O_2 were added until the reaction was complete. The title compound was prepared from 2-bromo-2-cyclohexyl-2-(methylsulfonyl)-1-phenylethan-1-one (8.13 g, 22.6 mmol). The reaction was run at 40 °C for 96 h. The product was purified by column chromatography on silica gel (5% \rightarrow 30% ethyl acetate/hexanes) and then on C-18 silica gel (10% \rightarrow 100% acetonitrile/water): 1.69 g (29%). White solid.

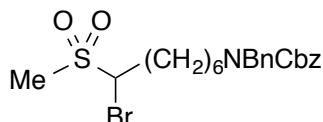
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- (4) Suryakiran, N.; Reddy, T. S.; Ashalatha, K.; Lakshman, M.; Venkateswarlu, Y. *Tetrahedron Lett.* **2006**, *47*, 3853–3856.
- (5) Suryakiran, N.; Prabhakar, P.; Reddy, T. S.; Mahesh, K. C.; Rajesh, K.; Venkateswarlu, Y. *Tetrahedron Lett.* **2007**, *48*, 877–881.

¹H NMR (500 MHz, CDCl₃) δ 4.60 (d, 1H, *J* = 2.7 Hz), 3.10 (s, 3H), 2.41–2.35 (m, 1H), 2.08–2.04 (m, 1H), 1.84–1.76 (m, 2H), 1.72–1.67 (m, 1H), 1.64–1.61 (m, 1H), 1.48–1.30 (m, 4H), 1.21–1.12 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 71.0, 39.9, 37.4, 31.8, 28.5, 26.0, 25.6, 25.3.

FT-IR (neat) 3011, 2930, 2855, 1452, 1411, 1370, 1310, 1240, 1171, 1138, 1090, 1080, 1060, 1032, 968, 922, 896, 885, 848, 792, 774, 728 cm⁻¹.

MS (ESI) *m/z* (M⁺+H) calcd for C₈H₁₆BrO₂S: 255, found: 255.



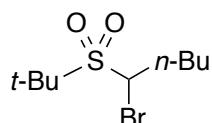
Benzyl benzyl(7-bromo-7-(methylsulfonyl)heptyl)carbamate. The title compound was prepared from benzyl benzyl(7-bromo-7-(methylsulfonyl)-8-oxo-8-phenyloctyl)carbamate (3.08 g, 5.13 mmol). The product was purified by column chromatography on silica gel (10%→50% ethyl acetate/hexanes) and then on C-18 silica gel (10%→100% acetonitrile/water): 1.57 g (62%). Viscous colorless oil.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.39–7.20 (br m, 10H), 5.18–5.14 (m, 2H), 4.68–4.60 (m, 1H), 4.50 (s, 2H), 3.28–3.20 (m, 2H), 3.06 (s, 3H), 2.40–2.30 (br m, 1H), 1.97–1.86 (br m, 1H), 1.70–1.25 (br m, 8H).

¹³C NMR (126 MHz, CD₂Cl₂) δ 156.9, 156.4, 138.6, 137.6, 128.82, 128.78, 128.2, 128.05, 127.98, 127.5, 67.3, 64.7, 50.8, 50.5, 47.4, 46.7, 37.8, 30.7, 28.6, 28.3, 27.9, 27.2, 26.7.

FT-IR (neat) 3087, 3062, 3030, 2930, 2858, 1692, 1605, 1585, 1496, 1467, 1467, 1453, 1421, 1365, 1315, 1230, 1140, 1119, 1072, 1028, 955, 915, 819, 768, 733 cm⁻¹.

MS (ESI) *m/z* (M⁺+H) calcd for C₂₃H₃₁BrNO₄S: 496, found: 496.



1-Bromo-1-(*tert*-butylsulfonyl)pentane. A mixture of 2-bromo-1-phenylhexan-1-one (5.10 g, 20.0 mmol), 2-methyl-2-propanethiol (1.80 g, 20.0 mmol), benzyltriethylammonium bromide (0.272 g, 1.00 mmol), and NaOH (3.00 g, 75.0 mmol) in dichloromethane (40 mL) and water (40 mL) in a 250-mL round-bottom flask was stirred at r.t. for 8 h. Then, water (100 mL) was added, and the mixture was extracted with dichloromethane (3 × 50 mL). The combined organic layers were dried over MgSO₄ and concentrated. The residue was dissolved in MeOH (50 mL) and water (50 mL), and then oxone® (30.7 g, 100 mmol) was added. The reaction mixture was stirred at r.t. overnight, and most of the MeOH was removed under reduced pressure. The resulting aqueous mixture was extracted with dichloromethane (3 × 30 mL). The combined organic layers were dried over MgSO₄ and concentrated. 2-(*tert*-Butylsulfonyl)-1-phenylhexan-1-one was purified by column chromatography (5%→60% ethyl acetate/hexanes): 5.34 g (90%). White solid.

2-Bromo-2-(*tert*-butylsulfonyl)-1-phenylhexan-1-one was prepared from 2-(*tert*-butylsulfonyl)-1-phenylhexan-1-one following the described procedure.

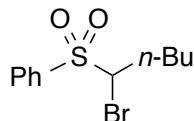
The title compound was prepared from 2-bromo-2-(*tert*-butylsulfonyl)-1-phenylhexan-1-one (2.30 g, 6.13 mmol). The reaction was conducted at 40 °C. The product was purified by column chromatography (hexanes→20% ethyl acetate/hexanes): 1.41 g (85%). White solid.

¹H NMR (500 MHz, CDCl₃) δ 4.84 (dd, 1H, *J* = 10.5, 3.0 Hz), 2.43 (dddd, 1H, *J* = 14.5, 10.2, 5.3, 2.9 Hz), 2.11–2.03 (m, 1H), 1.75–1.66 (m, 1H), 1.55 (s, 9H), 1.48–1.30 (m, 3H), 0.94 (t, 3H, *J* = 7.2 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 63.3, 59.2, 30.8, 28.8, 25.2, 22.0, 13.9.

FT-IR (neat) 2959, 2933, 2873, 1479, 1467, 1399, 1366, 1305, 1192, 1167, 1118, 1104, 1020, 986, 964, 929, 801, 733 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₉H₁₉BrNaO₂S: 293, found: 293.



((1-Bromopentyl)sulfonyl)benzene. The title compound was prepared from 2-bromo-1-phenyl-2-(phenylsulfonyl)hexan-1-one (12.0 g, 30.4 mmol). The reaction was conducted at 60 °C. The product was purified by column chromatography (hexanes→20% ethyl acetate/hexanes): 8.50 g (96%). White solid.

¹H NMR (500 MHz, CDCl₃) δ 7.98–7.95 (m, 2H), 7.72–7.68 (m, 1H), 7.61–7.57 (m, 2H), 4.70 (dd, 1H, *J* = 11.1, 2.9 Hz), 2.41 (dddd, 1H, *J* = 14.3, 9.9, 5.8, 2.9 Hz), 1.89 (dddd, 1H, *J* = 14.1, 11.1, 9.4, 4.4 Hz), 1.67–1.58 (m, 1H), 1.45–1.26 (m, 3H), 0.91 (t, 3H, *J* = 7.2 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 135.5, 134.6, 130.2, 129.2, 66.0, 31.0, 29.2, 21.8, 13.8.

FT-IR (neat) 3065, 2958, 2932, 2872, 1584, 1478, 1466, 1447, 1381, 1324, 1309, 1236, 1203, 1149, 1133, 1083, 1024, 999, 929, 792, 778, 746 cm⁻¹.

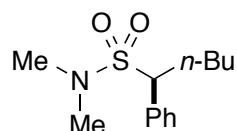
MS (ESI) *m/z* (M⁺+H) calcd for C₁₁H₁₆BrO₂S: 291, found: 291.

III. Enantioselective Arylations

General Procedure. An oven-dried 8-mL vial equipped with a magnetic stir bar was capped with a PTFE-lined septum cap, cooled under vacuum, and then filled with nitrogen. ZnI₂ (290 mg, 0.910 mmol) was added to the vial, and the vial was then immediately placed under vacuum and refilled with nitrogen (three cycles). Next, THF (2.73 mL) was added to the vial, followed by a solution of ArMgBr (prepared according to a literature procedure;¹ 1.00 M in THF; 0.910 mL, 0.910 mmol). The mixture was stirred at r.t. for 30 min. An oven-dried 20-mL vial equipped with a magnetic stir bar was charged with NiCl₂·glyme (15.4 mg, 0.070 mmol), (*R,R*)-L1 (30.4 mg, 0.091 mmol), and the electrophile (0.70 mmol). The vial was sealed with a PTFE-lined septum cap, placed under vacuum, and then filled with nitrogen; this cycle was repeated three times. THF (4.14 mL) was added, and the mixture was stirred at r.t. for 20 min, at which

time it had become homogenous. Both vials were wrapped with electrical tape, attached with nitrogen-filled balloons, and cooled to $-20\text{ }^{\circ}\text{C}$ for 15 min. The heterogeneous mixture of the nucleophile was then transferred by syringe over 2 min to the vial that contained the electrophile. The nitrogen-filled balloon was removed, and the septum cap was covered with grease. The reaction mixture was stirred at $-20\text{ }^{\circ}\text{C}$ for 24 h, and then the reaction was quenched by the addition of ethanol (0.70 mL). The solution was allowed to warm to r.t., and then it was filtered through a pad of silica (eluted with Et_2O). The filtrate was concentrated, and the residue was purified by column chromatography.

A second run was conducted with (*S,S*)-**L1**.



(*S*)-*N,N*-Dimethyl-1-phenylpentane-1-sulfonamide (Table 2, Entry 1). 1-Bromo-*N,N*-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (20% \rightarrow 25% Et_2O /hexanes). Light-yellow solid. First run: 159 mg (89%, 96% ee). Second run: 162 mg (91%, 96% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (1% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 10.9$ min (major), 13.4 min (minor).

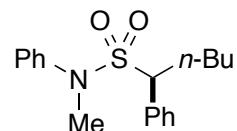
^1H NMR (500 MHz, CDCl_3) δ 7.42–7.34 (m, 5H), 4.08 (dd, 1H, $J = 11.3, 3.8$ Hz), 2.53 (s, 6H), 2.34 (dddd, 1H, $J = 13.7, 10.2, 6.5, 3.8$ Hz), 2.15 (dddd, 1H, $J = 13.6, 11.4, 10.0, 5.1$ Hz), 1.38–1.23 (m, 2H), 1.22–1.09 (m, 2H), 0.84 (t, 3H, $J = 7.3$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 133.9, 129.6, 129.0, 128.9, 67.7, 37.8, 29.6, 28.9, 22.4, 13.9.

FT-IR (neat) 3017, 2952, 2930, 2872, 1497, 1455, 1326, 1305, 1288, 1204, 1137, 1109, 1064, 973, 820, 808 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{13}\text{H}_{21}\text{NNaO}_2\text{S}$: 278, found: 278.

$[\alpha]^{25}_D = -30^\circ$ ($c = 1.02$, CHCl_3).



(*S*)-*N*-Methyl-*N,1*-diphenylpentane-1-sulfonamide (Table 2, Entry 2). 1-Bromo-*N*-methyl-*N*-phenylpentane-1-sulfonamide (224 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (10% Et_2O /hexanes). Light-yellow solid. First run: 211 mg (95%, 93% ee). Second run: 211 mg (95%, 95% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (2% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 10.5$ min (major), 11.7 min (minor).

^1H NMR (500 MHz, CDCl_3) δ 7.41–7.35 (m, 5H), 7.31–7.27 (m, 2H), 7.21–7.18 (m, 1H), 7.17–7.14 (m, 2H), 4.11 (dd, 1H, $J = 11.4, 3.7$ Hz), 2.88 (s, 3H), 2.32 (dddd, 1H, $J = 13.6, 10.1, 6.5, 3.7$

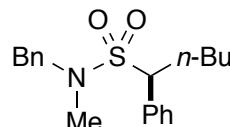
Hz), 2.14 (dddd, 1H, J = 13.4, 11.4, 9.9, 5.2 Hz), 1.34–1.18 (m, 2H), 1.17–1.03 (m, 2H), 0.80 (t, 3H, J = 7.3 Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 141.7, 133.7, 129.9, 129.1, 129.0, 128.8, 126.5, 125.8, 68.2, 39.2, 30.0, 28.9, 22.4, 13.9.

FT-IR (neat) 3063, 3030, 2957, 2932, 2872, 1596, 1493, 1455, 1423, 1380, 1342, 1266, 1179, 1143, 1108, 1067, 1028, 1003, 969, 917, 880, 801, 765 cm^{-1} .

MS (ESI) m/z (M^++Na) calcd for $\text{C}_{18}\text{H}_{23}\text{NNaO}_2\text{S}$: 340, found: 340.

$[\alpha]^{25}_{\text{D}} = -105^\circ$ ($c = 1.01$, CHCl_3).



(S)-N-Benzyl-N-methyl-1-phenylpentane-1-sulfonamide (Table 2, Entry 3). N -Benzyl-1-bromo- N -methylpentane-1-sulfonamide (234 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (7% ethyl acetate/hexanes). Light-yellow solid. First run: 219 mg (94%, 94% ee). Second run: 221 mg (95%, 93% ee).

The ee was determined by HPLC on a CHIRALPAK AD-H column (2% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 25.8$ min (major), 28.9 min (minor).

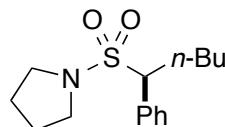
^1H NMR (500 MHz, CDCl_3) δ 7.42–7.36 (m, 5H), 7.31–7.23 (m, 3H), 7.22–7.18 (m, 2H), 4.12 (dd, 1H, J = 11.3, 3.8 Hz), 4.01 (d, 1H, J = 14.7 Hz), 3.68 (br d, 1H, J = 11.0 Hz), 2.42 (s, 3H), 2.38 (dddd, 1H, J = 13.7, 10.1, 6.2, 3.8 Hz), 2.21 (dddd, 1H, J = 13.5, 11.3, 9.8, 5.2 Hz), 1.42–1.26 (m, 2H), 1.26–1.12 (m, 2H), 0.85 (t, 3H, J = 7.3 Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 136.3, 133.9, 129.7, 129.0, 128.9, 128.6, 128.3, 127.9, 68.5, 54.2, 34.6, 29.6, 29.0, 22.4, 13.9.

FT-IR (neat) 3063, 3030, 2954, 2930, 2870, 1495, 1454, 1363, 1327, 1214, 1149, 1133, 1075, 1003, 944, 890, 807, 760 cm^{-1} .

MS (ESI) m/z (M^++Na) calcd for $\text{C}_{19}\text{H}_{25}\text{NNaO}_2\text{S}$: 354, found: 354.

$[\alpha]^{25}_{\text{D}} = -54^\circ$ ($c = 1.03$, CHCl_3).



(S)-1-((1-Phenylpentyl)sulfonyl)pyrrolidine (Table 2, Entry 4). 1-((1-Bromopentyl)sulfonyl)pyrrolidine (199 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (10% ethyl acetate/hexanes). White solid. First run: 166 mg (84%, 96% ee). Second run: 170 mg (86%, 96% ee).

The ee was determined by HPLC on a CHIRALPAK AD-H column (2% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 13.8$ min (minor), 20.2 min (major).

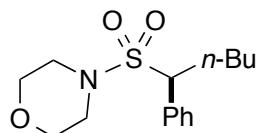
¹H NMR (500 MHz, CDCl₃) δ 7.43–7.39 (m, 2H), 7.39–7.33 (m, 3H), 4.11 (dd, 1H, *J* = 11.3, 3.8 Hz), 3.21–3.13 (m, 2H), 2.84–2.77 (m, 2H), 2.33 (dddd, 1H, *J* = 13.8, 10.1, 6.4, 3.8 Hz), 2.17 (dddd, 1H, *J* = 13.5, 11.3, 9.6, 5.2 Hz), 1.74–1.67 (m, 2H), 1.67–1.58 (m, 2H), 1.39–1.24 (m, 2H), 1.24–1.10 (m, 2H), 0.84 (t, 3H, *J* = 7.3 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 134.3, 129.7, 128.8, 128.7, 67.7, 48.2, 29.2, 29.0, 25.9, 22.4, 13.9.

FT-IR (neat) 3436, 2957, 2887, 2872, 2857, 1498, 1467, 1456, 1325, 1294, 1240, 1198, 1143, 1128, 1084, 1015, 829, 806, 728 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₅H₂₃NNaO₂S: 304, found: 304.

[α]_D²⁵ = -51° (c = 0.97, CHCl₃).



(S)-4-((1-Phenylpentyl)sulfonyl)morpholine (Table 2, Entry 5). 4-((1-Bromopentyl)sulfonyl)morpholine (210 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (20% ethyl acetate/hexanes). White solid. First run: 197 mg (95%, 98% ee). Second run: 186 mg (89%, 95% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (3% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 13.9 min (major), 16.7 min (minor).

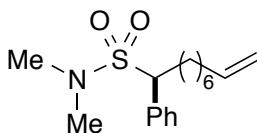
¹H NMR (500 MHz, CDCl₃) δ 7.44–7.35 (m, 5H), 4.02 (dd, 1H, *J* = 11.3, 3.8 Hz), 3.56–3.52 (m, 2H), 3.48–3.43 (m, 2H), 3.06–3.02 (m, 2H), 2.75 (br s, 2H), 2.34 (dddd, 1H, *J* = 13.8, 10.1, 6.3, 3.8 Hz), 2.13 (dddd, 1H, *J* = 13.4, 11.3, 9.8, 5.1 Hz), 1.39–1.23 (m, 2H), 1.23–1.08 (m, 2H), 0.84 (t, 3H, *J* = 7.3 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 133.5, 129.7, 129.2, 129.0, 68.5, 67.0, 46.3, 29.7, 28.9, 22.4, 13.9.

FT-IR (neat) 2955, 2923, 2859, 1496, 1455, 1336, 1323, 1257, 1214, 1152, 1128, 1110, 1076, 955, 924, 848, 803 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₅H₂₃NNaO₃S: 320, found: 320.

[α]_D²⁵ = -34° (c = 1.02, CHCl₃).



(S)-N,N-Dimethyl-1-phenylnon-8-ene-1-sulfonamide (Table 2, Entry 6). 1-Bromo-N,N-dimethylnon-8-ene-1-sulfonamide (219 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (5% → 10% ethyl acetate/hexanes). Light-yellow solid. First run: 192 mg (89%, 95% ee). Second run: 189 mg (87%, 95% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (1% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 12.8 min (major), 20.6 min (minor).

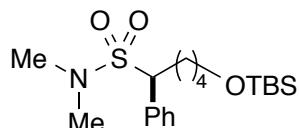
¹H NMR (500 MHz, CDCl₃) δ 7.41–7.34 (m, 5H), 5.76 (dd, 1H, J = 16.9, 10.2, 6.7, 6.7 Hz), 4.96 (ddd, 1H, J = 17.1, 2.2, 1.6, 1.6 Hz), 4.91 (ddd, 1H, J = 10.2, 2.3, 1.2, 1.2 Hz), 4.08 (dd, 1H, J = 11.3, 3.9 Hz), 2.53 (s, 6H), 2.30 (ddd, 1H, J = 13.7, 10.2, 6.5, 3.9 Hz), 2.19–2.11 (m, 1H), 2.01–1.96 (m, 2H), 1.35–1.11 (m, 8H).

¹³C NMR (126 MHz, CDCl₃) δ 139.1, 133.9, 129.6, 129.0, 128.9, 114.4, 67.7, 37.8, 33.8, 29.8, 29.1, 28.8, 26.7.

FT-IR (neat) 3062, 2924, 2853, 1640, 1497, 1468, 1456, 1414, 1327, 1208, 1137, 1066, 977, 912, 824 cm⁻¹.

MS (ESI) m/z (M⁺+Na) calcd for C₁₇H₂₇NNaO₂S: 332, found: 332.

[α]²⁵_D = -19.2° (c = 0.98, CHCl₃).



(S)-5-((tert-Butyldimethylsilyl)oxy)-N,N-dimethyl-1-phenylpentane-1-sulfonamide (Table 2, Entry 7). 1-Bromo-5-((tert-butylidimethylsilyl)oxy)-N,N-dimethylpentane-1-sulfonamide (272 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (2% → 20% ethyl acetate/hexanes). White solid. First run: 248 mg (92%, >99% ee). Second run: 250 mg (93%, 98% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (2% i-PrOH/hexanes, 1.0 mL/min) with t_r = 8.2 min (major), 11.0 min (minor).

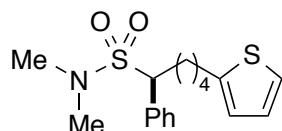
¹H NMR (500 MHz, CDCl₃) δ 7.41–7.33 (m, 5H), 4.09 (dd, 1H, J = 11.3, 3.9 Hz), 3.56–3.49 (m, 2H), 2.53 (s, 6H), 2.35–2.28 (m, 1H), 2.21–2.13 (m, 1H), 1.57–1.42 (m, 2H), 1.27–1.18 (m, 2H), 0.82 (s, 9H), -0.02 (s, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 133.7, 129.6, 129.0, 128.9, 67.6, 62.7, 37.8, 32.4, 29.7, 26.0, 23.1, 18.4, -5.2.

FT-IR (neat) 3065, 2931, 2897, 2860, 1458, 1385, 1359, 1329, 1280, 1257, 1200, 1143, 1132, 1110, 1092, 966, 900, 872, 833, 808, 779, 736 cm⁻¹.

MS (ESI) m/z (M⁺+Na) calcd for C₁₉H₃₅NNaO₃SSi: 408, found: 408.

[α]²⁵_D = -15.0° (c = 0.98, CHCl₃).



(S)-N,N-Dimethyl-1-phenyl-5-(thiophen-2-yl)pentane-1-sulfonamide (Table 2, Entry 8). 1-Bromo-N,N-dimethyl-5-(thiophen-2-yl)pentane-1-sulfonamide (238 mg, 0.700 mmol) and phenylzinc iodide (1.05 mmol) were used. The product was purified by column chromatography (10% → 15% ethyl acetate/hexanes). Yellow solid. First run: 128 mg (54%, 90% ee). Second run: 131 mg (55%, 91% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (5% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 18.3$ min (major), 22.8 min (minor).

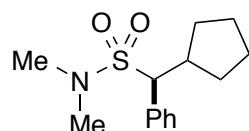
^1H NMR (500 MHz, CDCl_3) δ 7.41–7.34 (m, 5H), 7.08 (dd, 1H, $J = 5.1, 1.2$ Hz), 6.88 (dd, 1H, $J = 5.1, 3.4$ Hz), 6.71 (dddd, 1H, $J = 3.3, 1.0, 1.0, 1.0$ Hz), 4.08 (dd, 1H, $J = 11.2, 3.9$ Hz), 2.82–2.70 (m, 2H), 2.52 (s, 6H), 2.39–2.32 (m, 1H), 2.23–2.15 (m, 1H), 1.74–1.61 (m, 2H), 1.35–1.21 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 145.1, 133.8, 129.6, 129.0, 128.9, 126.8, 124.2, 123.0, 67.6, 37.8, 31.4, 29.65, 29.59, 26.2.

FT-IR (neat) 3064, 2932, 2856, 1495, 1480, 1454, 1331, 1282, 1200, 1140, 1062, 1030, 967, 849, 820 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{17}\text{H}_{23}\text{NNaO}_2\text{S}_2$: 360, found: 360.

$[\alpha]^{25}_D = -9.4^\circ$ ($c = 0.99$, CHCl_3).



(S)-1-Cyclopentyl-N,N-dimethyl-1-phenylmethanesulfonamide (Table 2, Entry 9). 1-Bromo-1-cyclopentyl-N,N-dimethylmethanesulfonamide (189 mg, 0.700 mmol) and phenylzinc iodide (1.05 mmol) were used. The product was purified by column chromatography (first purification: 10% ethyl acetate/hexanes; second purification: 12%→100% dichloromethane/hexanes). White solid. First run: 86 mg (46%, >99% ee). Second run: 79 mg (42%, >99% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (1% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 12.7$ min (major), 14.6 min (minor).

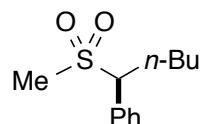
^1H NMR (500 MHz, CDCl_3) δ 7.41–7.38 (m, 2H), 7.37–7.31 (m, 3H), 3.92 (d, 1H, $J = 10.3$ Hz), 2.78–2.69 (m, 1H), 2.43 (s, 6H), 2.29–2.22 (m, 1H), 1.75–1.67 (m, 1H), 1.66–1.60 (m, 1H), 1.59–1.41 (m, 4H), 1.03–0.95 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 135.2, 129.7, 128.69, 128.67, 73.2, 41.8, 37.6, 32.3, 32.1, 25.5, 24.1.

FT-IR (neat) 3090, 3064, 3025, 2960, 2871, 2812, 1496, 1479, 1452, 1323, 1293, 1206, 1188, 1131, 1081, 1063, 1030, 1003, 969, 911, 872, 848, 807, 732 cm^{-1} .

MS (EI) m/z ($\text{M}^+ - \text{SO}_2\text{NMe}_2$) calcd for $\text{C}_{12}\text{H}_{15}$: 159, found: 159.

$[\alpha]^{25}_D = -43^\circ$ ($c = 1.04$, CHCl_3).



(S)-(1-(Methylsulfonyl)pentyl)benzene (Table 3, Entry 1). 1-Bromo-1-(methylsulfonyl)pentane (160 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (20%→30% ethyl acetate/hexanes). White solid. First run: 150 mg (95%, 94% ee). Second run: 153 mg (97%, 94% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (5% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 17.6$ min (major), 20.8 min (minor).

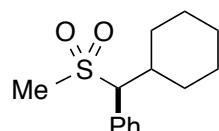
^1H NMR (500 MHz, CDCl_3) δ 7.42–7.37 (m, 5H), 3.99 (dd, 1H, $J = 11.5, 3.7$ Hz), 2.59 (s, 3H), 2.45–2.37 (m, 1H), 2.12 (dddd, 1H, $J = 13.6, 11.5, 9.6, 5.3$ Hz), 1.41–1.26 (m, 2H), 1.25–1.14 (m, 2H), 0.85 (t, 3H, $J = 7.1$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 133.4, 129.5, 129.28, 129.27, 70.4, 38.7, 28.9, 26.7, 22.4, 13.9.

FT-IR (neat) 3088, 3065, 3051, 3011, 2931, 2869, 1496, 1468, 1456, 1417, 1379, 1292, 1277, 1263, 1211, 1158, 1130, 1107, 1072, 1036, 966, 936, 904, 805, 722 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{12}\text{H}_{18}\text{NaO}_2\text{S}$: 249, found: 249.

$[\alpha]^{25}_D = -6.2^\circ$ ($c = 1.00$, CHCl_3).



(S)-(Cyclohexyl(methylsulfonyl)methyl)benzene (Table 3, Entry 2).

(Bromo(methylsulfonyl)methyl)cyclohexane (179 mg, 0.700 mmol) and phenylzinc iodide (1.05 mmol) were used. The product was purified by column chromatography (10% \rightarrow 15% ethyl acetate/hexanes). White solid. First run: 145 mg (82%, 99% ee). Second run: 148 mg (84%, 99% ee).

The ee was determined by HPLC on a CHIRALPAK AD-H column (4% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 16.7$ min (minor), 25.8 min (major).

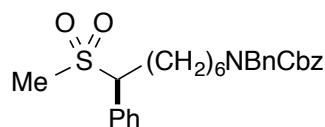
^1H NMR (500 MHz, CDCl_3) δ 7.43–7.36 (m, 5H), 3.87 (d, 1H, $J = 7.9$ Hz), 2.53–2.45 (m, 1H), 2.46 (s, 3H), 2.29–2.24 (m, 1H), 1.80–1.74 (m, 1H), 1.67–1.56 (m, 3H), 1.42–1.33 (m, 1H), 1.28–1.18 (m, 2H), 1.14–1.05 (m, 1H), 0.93–0.85 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 133.9, 129.8, 129.2, 129.1, 75.9, 41.4, 38.1, 32.4, 30.6, 26.11, 26.06, 26.0.

FT-IR (neat) 3004, 2930, 2853, 1496, 1454, 1413, 1378, 1348, 1319, 1302, 1292, 1244, 1221, 1170, 1127, 1076, 1036, 970, 896, 854, 804, 742 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{14}\text{H}_{20}\text{NaO}_2\text{S}$: 275, found: 275.

$[\alpha]^{25}_D = -40^\circ$ ($c = 1.06$, CHCl_3).



Benzyl (S)-benzyl(7-(methylsulfonyl)-7-phenylheptyl)carbamate (Table 3, Entry 3). Benzyl benzyl(7-bromo-7-(methylsulfonyl)heptyl)carbamate (199 mg, 0.400 mmol) and phenylzinc iodide (0.520 mmol) were used. The product was purified by column chromatography on silica gel (25% ethyl acetate/hexanes) and then preparative HPLC on C-18 silica gel (80% \rightarrow 100% acetonitrile/water; water was doped with 0.1% AcOH). Viscous colorless oil. First run: 149 mg (75%, 89% ee). Second run: 145 mg (73%, 91% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (20% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 31.5 min (major), 40.0 min (minor).

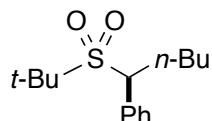
^1H NMR (500 MHz, CD_2Cl_2) δ 7.44–7.17 (m, 15H), 5.14–5.12 (m, 2H), 4.45 (s, 2H), 4.01–3.95 (m, 1H), 3.22–3.15 (m, 2H), 2.59 (s, 3H), 2.35–2.23 (br m, 1H), 2.12–1.99 (br m, 1H), 1.48–1.40 (br m, 2H), 1.35–1.09 (br m, 6H).

^{13}C NMR (126 MHz, CD_2Cl_2) δ 156.9, 156.3, 138.7, 137.6, 133.6, 129.9, 129.40, 129.36, 128.8, 128.7, 128.2, 128.01, 127.95, 127.5, 70.3, 67.3, 50.8, 50.4, 47.4, 46.7, 38.9, 29.2, 28.4, 27.9, 27.4, 26.9, 26.8.

FT-IR (neat) 3088, 3063, 3031, 3007, 2931, 2858, 1697, 1605, 1586, 1496, 1468, 1454, 1422, 1366, 1305, 1232, 1137, 1086, 1071, 1029, 1002, 954, 916, 801 cm^{-1} .

MS (ESI) m/z (M^++H) calcd for $\text{C}_{29}\text{H}_{36}\text{NO}_4\text{S}$: 494, found: 494.

$[\alpha]^{25}_{\text{D}} = -0.037^\circ$ ($c = 4.1$, CHCl_3).



(S)-(1-(*tert*-Butylsulfonyl)pentyl)benzene (Table 3, Entry 4). 1-Bromo-1-(*tert*-butylsulfonyl)pentane (190 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (15% ethyl acetate/hexanes). White solid. First run: 179 mg (95%, 99% ee). Second run: 182 mg (97%, 98% ee).

The ee was determined by HPLC on a CHIRALPAK IB-3 column (1% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 8.4 min (major), 9.9 min (minor).

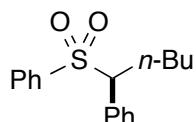
^1H NMR (500 MHz, CDCl_3) δ 7.47–7.45 (m, 2H), 7.39–7.32 (m, 3H), 4.14 (dd, 1H, $J = 11.6, 3.3$ Hz), 2.47 (dddd, 1H, $J = 13.6, 10.6, 6.2, 3.3$ Hz), 2.06 (dddd, 1H, $J = 13.4, 11.6, 10.2, 4.9$ Hz), 1.40–1.30 (m, 1H), 1.29–1.21 (m, 1H), 1.16 (s, 9H), 1.15–1.01 (m, 2H), 0.82 (t, 3H, $J = 7.3$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 134.9, 129.6, 129.0, 128.9, 65.3, 62.1, 28.9, 28.7, 24.4, 22.4, 13.9.

FT-IR (neat) 3032, 2986, 2954, 2872, 1497, 1466, 1455, 1366, 1279, 1190, 1115, 1100, 782 cm^{-1} .

MS (ESI) m/z (M^++Na) calcd for $\text{C}_{15}\text{H}_{24}\text{NaO}_2\text{S}$: 291, found: 291.

$[\alpha]^{25}_{\text{D}} = -20.3^\circ$ ($c = 1.01$, CHCl_3).



(S)-((1-Phenylpentyl)sulfonyl)benzene (Table 3, Entry 5). ((1-Bromopentyl)sulfonyl)benzene (204 mg, 0.700 mmol) and phenylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (10%→20% Et_2O /hexanes). White solid. First run: 195 mg (97%, 86% ee). Second run: 193 mg (96%, 83% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (1% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 13.6 min (major), 18.7 min (minor).

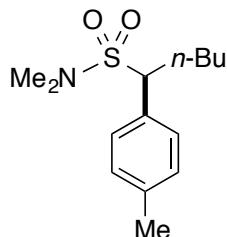
¹H NMR (500 MHz, CDCl₃) δ 7.54–7.49 (m, 3H), 7.38–7.34 (m, 2H), 7.29–7.26 (m, 1H), 7.24–7.20 (m, 2H), 7.10–7.07 (m, 2H), 4.01 (dd, 1H, *J* = 11.6, 3.6 Hz), 2.46–2.39 (m, 1H), 2.20–2.10 (m, 1H), 1.38–1.23 (m, 2H), 1.22–1.13 (m, 2H), 0.83 (t, 3H, *J* = 7.3 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 137.6, 133.5, 132.6, 130.0, 129.2, 128.8, 128.7, 128.6, 71.8, 29.0, 27.1, 22.4, 13.9.

FT-IR (neat) 2952, 2926, 2857, 1584, 1496, 1467, 1455, 1447, 1379, 1316, 1304, 1294, 1214, 1147, 1084, 1070, 1037, 1024, 998, 968, 800, 758, 713 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₇H₂₀NaO₂S: 311, found: 311.

[α]_D²⁵ = -78° (c = 1.08, CHCl₃).



(S)-N,N-Dimethyl-1-(*p*-tolyl)pentane-1-sulfonamide (Table 4, Entry 1). 1-Bromo-N,N-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and *p*-tolylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (20% Et₂O/hexanes). Light-yellow oil. First run: 172 mg (91%, 96% ee). Second run: 165 mg (87%, 95% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (2% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 8.1 min (major), 10.1 min (minor).

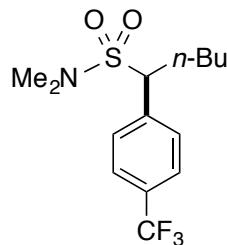
¹H NMR (500 MHz, CDCl₃) δ 7.30–7.27 (m, 2H), 7.19–7.17 (m, 2H), 4.05 (dd, 1H, *J* = 11.3, 3.8 Hz), 2.54 (s, 6H), 2.36 (s, 3H), 2.29 (dddd, 1H, *J* = 13.7, 10.1, 6.4, 3.8 Hz), 2.12 (dddd, 1H, *J* = 13.5, 11.4, 9.7, 5.3 Hz), 1.38–1.22 (m, 2H), 1.22–1.09 (m, 2H), 0.83 (t, 3H, *J* = 7.3 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 138.8, 130.8, 129.6, 129.5, 67.4, 37.8, 29.6, 28.9, 22.4, 21.3, 13.9.

FT-IR (neat) 3025, 2956, 2932, 2872, 2811, 1515, 1479, 1457, 1413, 1380, 1331, 1283, 1204, 1141, 1107, 1062, 1022, 968, 843, 832, 716 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₄H₂₃NNaO₂S: 292, found: 292.

[α]_D²⁵ = -30° (c = 0.99, CHCl₃).



(S)-N,N-Dimethyl-1-(4-(trifluoromethyl)phenyl)pentane-1-sulfonamide (Table 4, Entry 2). An oven-dried 8-mL vial equipped with a magnetic stir bar was capped with a PTFE-lined septum cap, cooled under vacuum, and filled with nitrogen. 4-Iodobenzotrifluoride (248 mg,

0.910 mmol) and THF (1.35 mL) were added to the vial, followed by the dropwise addition over 1 min of *i*-PrMgCl (1.92 M in THF; 0.474 mL, 0.910 mmol), and the resulting mixture was stirred at r.t. for 1 h. An oven-dried 4-mL vial equipped with a magnetic stir bar was capped with a PTFE-lined septum cap, cooled under vacuum, and filled with nitrogen. ZnI₂ (290 mg, 0.910 mmol) was added into the vial. The vial was immediately evacuated and refilled with nitrogen (three cycles), and then THF (1.82 mL) was added to the vial. The solution of ZnI₂ was transferred by syringe to the Grignard reagent, and then the reaction mixture was stirred at r.t. for 30 min.

1-Bromo-*N,N*-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and (4-(trifluoromethyl)phenyl)zinc iodide (0.910 mmol) were used. The product was purified by column chromatography (20% Et₂O/hexanes). Light-yellow solid. First run: 209 mg (92%, 98% ee). Second run: 216 mg (95%, 98% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (3% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 8.8 min (major), 12.0 min (minor).

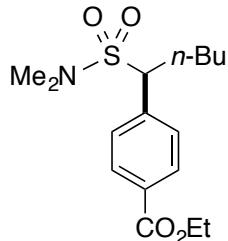
¹H NMR (500 MHz, CDCl₃) δ 7.65 (d, 2H, *J* = 8.2 Hz), 7.55 (d, 2H, *J* = 8.2 Hz), 4.14 (dd, 1H, *J* = 11.4, 3.8 Hz), 2.58 (s, 6H), 2.32 (dddd, 1H, *J* = 13.9, 10.3, 6.3, 3.9 Hz), 2.14 (dddd, 1H, *J* = 13.7, 11.4, 10.1, 4.9 Hz), 1.38–1.24 (m, 2H), 1.22–1.06 (m, 2H), 0.84 (t, 3H, *J* = 7.3 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 138.2 (d, *J*_{CF} = 1.3 Hz), 131.1 (q, *J*_{CF} = 32.7 Hz), 130.0, 125.8 (q, *J*_{CF} = 3.7 Hz), 123.6 (q, *J*_{CF} = 272.2 Hz), 67.3, 37.8, 29.6, 28.8, 22.3, 13.8.

FT-IR (neat) 2958, 2875, 1325, 1167, 1122, 1069, 1019, 968, 856, 727 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₄H₂₀F₃NNaO₂S: 346, found: 346.

[α]²⁵_D = -23.3° (c = 1.02, CHCl₃).



Ethyl (S)-4-(1-(*N,N*-dimethylsulfamoyl)pentyl)benzoate (Table 4, Entry 3). An oven-dried 8-mL vial equipped with a magnetic stir bar was capped with a PTFE-lined septum cap, cooled under vacuum, and then filled with nitrogen. Ethyl 4-iodobenzoate (251 mg, 0.910 mmol) was added to the vial, and then the vial was evacuated and refilled with nitrogen (three cycles). Next, THF (1.31 mL) was added to the vial, and the vial was wrapped with electrical tape and fitted with a nitrogen-filled balloon. Then, the reaction mixture was cooled to -20 °C. *i*-PrMgCl (1.78 M in THF; 0.511 mL, 0.910 mmol) was added over 1 min, and the mixture was stirred at -20 °C for 2 h. An oven-dried 4-mL vial equipped with a magnetic stir bar was capped with a PTFE-lined septum cap, cooled under vacuum, and filled with nitrogen. ZnI₂ (291 mg, 0.910 mmol) was added to the vial. The vial was immediately placed under vacuum and then filled with nitrogen. This evacuation-refill cycle was repeated three times, and then THF (1.82 mL) was added to the vial. The solution of ZnI₂ was transferred by syringe to the Grignard reagent,

and then the reaction mixture was stirred at -20°C for 30 min. The reaction mixture was allowed to warm to r.t. and stirred for an additional 30 min.

1-Bromo-*N,N*-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and (4-(ethoxycarbonyl)phenyl)zinc iodide (0.910 mmol) were used. The product was purified by column chromatography (20% ethyl acetate/hexanes). Colorless oil. First run: 229 mg (>99%, 97% ee). Second run: 229 mg (>99%, 97% ee).

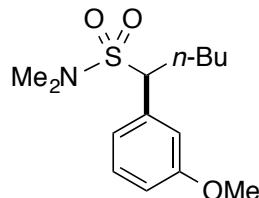
The ee was determined by HPLC on a CHIRALCEL OD-H column (5% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 8.7$ min (major), 11.4 min (minor).

^1H NMR (500 MHz, CDCl_3) δ 8.07–8.05 (m, 2H), 7.50–7.48 (m, 2H), 4.38 (q, 2H, $J = 7.1$ Hz), 4.14 (dd, 1H, $J = 11.3, 3.8$ Hz), 2.55 (s, 6H), 2.32 (dddd, 1H, $J = 14.0, 10.2, 6.2, 3.8$ Hz), 2.16 (dddd, 1H, $J = 13.6, 11.3, 10.0, 4.8$ Hz), 1.40 (t, 3H, $J = 7.1$ Hz), 1.37–1.21 (m, 2H), 1.21–1.05 (m, 2H), 0.82 (t, 3H, $J = 7.3$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 166.2, 139.0, 131.1, 130.0, 129.6, 67.5, 61.4, 37.9, 29.6, 28.9, 22.4, 14.5, 13.9.

FT-IR (neat) 2956, 2934, 2872, 2813, 1718, 1611, 1576, 1507, 1477, 1457, 1417, 1367, 1334, 1278, 1182, 1143, 1110, 1063, 1021, 968, 867, 799, 776, 753, 712 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ - \text{SO}_2\text{NMe}_2$) calcd for $\text{C}_{14}\text{H}_{19}\text{O}_2$: 219, found: 219.
 $[\alpha]^{25}_{\text{D}} = -36^\circ$ ($c = 1.00$, CHCl_3).



(S)-1-(3-Methoxyphenyl)-*N,N*-dimethylpentane-1-sulfonamide (Table 4, Entry 4). 1-Bromo-*N,N*-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and (3-methoxyphenyl)zinc iodide (0.910 mmol) were used. The product was purified by column chromatography (15% ethyl acetate/hexanes). White solid. First run: 175 mg (88%, 95% ee). Second run: 174 mg (87%, 96% ee).

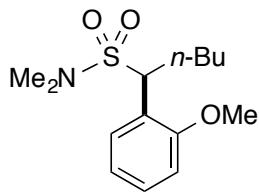
The ee was determined by HPLC on a CHIRALCEL OD-H column (3% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 10.5$ min (major), 12.9 min (minor).

^1H NMR (500 MHz, CDCl_3) δ 7.30–7.27 (m, 1H), 6.99–6.97 (m, 2H), 6.90–6.88 (m, 1H), 4.05 (dd, 1H, $J = 11.3, 3.8$ Hz), 3.82 (s, 3H), 2.56 (s, 6H), 2.29 (dddd, 1H, $J = 13.6, 10.2, 6.6, 3.8$ Hz), 2.12 (dddd, 1H, $J = 13.5, 11.3, 9.8, 5.2$ Hz), 1.38–1.23 (m, 2H), 1.23–1.10 (m, 2H), 0.84 (t, 3H, $J = 7.3$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 159.9, 135.4, 129.8, 122.0, 115.2, 114.2, 67.6, 55.5, 37.8, 29.7, 28.9, 22.4, 13.9.

FT-IR (neat) 3002, 2956, 2873, 2839, 1601, 1585, 1489, 1456, 1438, 1380, 1330, 1262, 1204, 1142, 1111, 1049, 968, 888, 806 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{14}\text{H}_{23}\text{NNaO}_3\text{S}$: 308, found: 308.
 $[\alpha]^{25}_{\text{D}} = -28^\circ$ ($c = 1.00$, CHCl_3).



(S)-1-(2-Methoxyphenyl)-N,N-dimethylpentane-1-sulfonamide (Table 4, Entry 5). 1-Bromo-N,N-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and (2-methoxyphenyl)zinc iodide (1.05 mmol) were used. The product was purified by column chromatography on silica gel (10%→15% ethyl acetate/hexanes) and then on C-18 silica gel (10%→100% acetonitrile/water). Light-yellow oil. First run: 125 mg (63%, 96% ee). Second run: 128 mg (64%, 96% ee).

The ee was determined by HPLC on a CHIRALPAK AS-H column (5% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 16.6$ min (major), 19.3 min (minor).

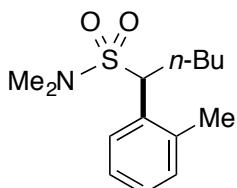
^1H NMR (500 MHz, CDCl_3) δ 7.59 (dd, 1H, $J = 7.8, 1.7$ Hz), 7.30 (ddd, 1H, $J = 8.2, 7.4, 1.7$ Hz), 7.00 (ddd, 1H, $J = 7.6, 7.6, 1.1$ Hz), 6.91 (dd, 1H, $J = 8.3, 1.1$ Hz), 4.87 (dd, 1H, $J = 11.4, 3.9$ Hz), 3.87 (s, 3H), 2.51 (s, 6H), 2.32 (dddd, 1H, $J = 13.7, 10.2, 6.4, 3.9$ Hz), 2.13–2.05 (m, 1H), 1.36–1.22 (m, 2H), 1.22–1.07 (m, 2H), 0.83 (t, 3H, $J = 7.3$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 157.7, 129.7, 129.6, 122.2, 121.1, 110.5, 57.3, 55.8, 37.6, 29.7, 28.6, 22.4, 13.9.

FT-IR (neat) 3070, 3005, 2957, 2873, 1601, 1587, 1494, 1463, 1442, 1380, 1330, 1290, 1247, 1202, 1142, 1124, 1090, 1052, 1026, 967, 796, 756, 726 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{14}\text{H}_{23}\text{NNaO}_3\text{S}$: 308, found: 308.

$[\alpha]^{25}_D = +40^\circ$ ($c = 1.03$, CHCl_3).



(S)-N,N-Dimethyl-1-(*o*-tolyl)pentane-1-sulfonamide (Table 4, Entry 6). 1-Bromo-N,N-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and *o*-tolylzinc iodide (1.05 mmol) were used. The product was purified by column chromatography (5%→10% ethyl acetate/hexanes). Light-yellow oil. First run: 148 mg (78%, 97% ee). Second run: 149 mg (79%, 97% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (2% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 9.8$ min (major), 11.9 min (minor).

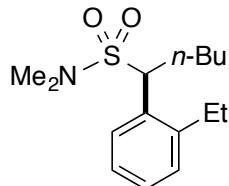
^1H NMR (500 MHz, CDCl_3) δ 7.62–7.58 (m, 1H), 7.25–7.19 (m, 3H), 4.43 (dd, 1H, $J = 11.3, 3.8$ Hz), 2.60 (s, 6H), 2.39 (s, 3H), 2.33 (dddd, 1H, $J = 13.6, 10.1, 6.1, 3.8$ Hz), 2.16–2.08 (m, 1H), 1.37–1.22 (m, 2H), 1.21–1.06 (m, 2H), 0.83 (t, 3H, $J = 7.3$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 137.4, 132.3, 130.7, 128.4, 128.3, 126.6, 63.2, 38.0, 30.7, 28.7, 22.6, 20.1, 13.9.

FT-IR (neat) 3064, 3023, 2957, 2872, 2813, 1604, 1493, 1461, 1380, 1329, 1283, 1203, 1178, 1141, 1119, 1063, 967, 834, 802 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₄H₂₃NNaO₂S: 292, found: 292.

[α]²⁵_D = +7.9° (c = 1.05, CHCl₃).



(S)-1-(2-Ethylphenyl)-N,N-dimethylpentane-1-sulfonamide (Table 4, Entry 7). 1-Bromo-N,N-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol), (2-ethylphenyl)zinc iodide (1.40 mmol), NiCl₂•glyme (30.8 mg, 0.140 mmol), and (*R,R*)-L1 (60.9 mg, 0.182 mmol) were used. The product was purified by column chromatography (first purification: 10% ethyl acetate/hexanes; second purification: 15%→90% dichloromethane/hexanes). Light-yellow oil. First run: 178 mg (90%, 97% ee). Second run: 165 mg (83%, 97% ee).

The ee was determined by HPLC on a CHIRALPAK IC column (15% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 16.9 min (minor), 22.7 min (major).

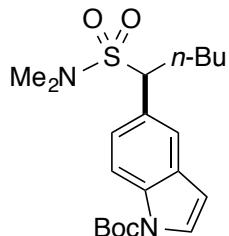
¹H NMR (500 MHz, CD₂Cl₂) δ 7.57–7.54 (m, 1H), 7.30–7.22 (m, 3H), 4.46 (dd, 1H, *J* = 11.1, 4.0 Hz), 2.79 (dq, 1H, *J* = 14.9, 7.5 Hz), 2.68 (dq, 1H, *J* = 15.2, 7.6 Hz), 2.62 (s, 6H), 2.29–2.22 (m, 1H), 2.15–2.07 (m, 1H), 1.39–1.24 (m, 2H), 1.22 (t, 3H, *J* = 7.6 Hz), 1.24–1.15 (m, 1H), 1.13–1.04 (m, 1H), 0.84 (t, 3H, *J* = 7.3 Hz).

¹³C NMR (126 MHz, CD₂Cl₂) δ 144.1, 131.9, 129.2, 128.7, 128.5, 126.5, 62.8, 38.0, 30.8, 29.3, 26.0, 23.0, 15.7, 13.9.

FT-IR (neat) 3063, 3021, 2959, 2933, 2873, 2813, 1490, 1455, 1378, 1330, 1282, 1201, 1177, 1141, 1120, 1062, 968, 803, 760 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₅H₂₅NNaO₂S: 306, found: 306.

[α]²⁵_D = +8.9° (c = 1.03, CHCl₃).



tert-Butyl (S)-5-(1-(N,N-dimethylsulfamoyl)pentyl)-1H-indole-1-carboxylate (Table 4, Entry 8). An oven-dried 8-mL vial equipped with a magnetic stir bar was capped with a PTFE-lined septum cap, cooled under vacuum, and filled with nitrogen. *tert*-Butyl 5-iodo-1*H*-indole-1-carboxylate (360 mg, 1.05 mmol) was added to the vial, and then the vial was evacuated and refilled with nitrogen (three cycles). THF (1.56 mL) was added to the vial, and the vial was

wrapped with electrical tape and fitted with a nitrogen-filled balloon. Then, the reaction mixture was cooled to -20°C . *i*-PrMgCl (1.93 M in THF; 0.544 mL, 1.05 mmol) was added over 1 min, and the mixture was stirred at -20°C for 2 h. An oven-dried 4-mL vial equipped with a magnetic stir bar was capped with a PTFE-lined septum cap, cooled under vacuum, and filled with nitrogen. ZnI₂ (338 mg, 1.06 mmol) was added to the vial. The vial was immediately placed under vacuum and then filled with nitrogen. This evacuation-refill cycle was repeated three times, and then THF (2.10 mL) was added to the vial. The solution of ZnI₂ was transferred by syringe to the Grignard reagent, and then the reaction mixture was stirred at -20°C for 30 min. The reaction mixture was allowed to warm to r.t. and stirred for an additional 30 min.

1-Bromo-*N,N*-dimethylpentane-1-sulfonamide (181 mg, 0.700 mmol) and (1-(*tert*-butoxycarbonyl)-1*H*-indol-5-yl)zinc iodide (1.05 mmol) were used. The product was purified by column chromatography on silica gel (10%→15% ethyl acetate/hexanes) and then on C-18 silica gel (10%→100% acetonitrile/water). Yellow solid. First run: 180 mg (65%, 88% ee). Second run: 200 mg (72%, 90% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (2% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 11.7$ min (major), 15.4 min (minor).

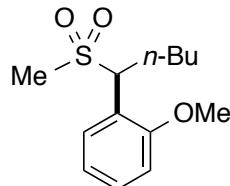
¹H NMR (500 MHz, CDCl₃) δ 8.14 (d, 1H, *J* = 8.5 Hz), 7.63–7.62 (m, 2H), 7.33 (dd, 1H, *J* = 8.6, 1.8 Hz), 6.58 (dd, 1H, *J* = 3.7, 0.8 Hz), 4.18 (dd, 1H, *J* = 11.4, 3.8 Hz), 2.51 (s, 6H), 2.36 (dddd, 1H, *J* = 13.7, 10.2, 6.4, 3.8 Hz), 2.20 (dddd, 1H, *J* = 13.6, 11.4, 10.0, 5.0 Hz), 1.68 (s, 9H), 1.38–1.23 (m, 2H), 1.22–1.08 (m, 2H), 0.82 (t, 3H, *J* = 7.3 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 149.7, 135.4, 130.9, 128.0, 126.9, 125.7, 121.9, 115.4, 107.4, 84.2, 67.6, 37.9, 29.9, 28.9, 28.3, 22.4, 13.9.

FT-IR (neat) 3152, 3120, 2956, 2934, 2873, 1736, 1536, 1470, 1445, 1374, 1351, 1329, 1256, 1218, 1193, 1164, 1138, 1107, 1084, 1042, 1024, 968, 841, 768, 729 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₂₀H₃₀N₂NaO₄S: 417, found: 417.

[α]_D²⁵ = -23.7° (c = 1.04, CHCl₃).



(S)-1-Methoxy-2-(1-(methylsulfonyl)pentyl)benzene (Table 4, Entry 9). 1-Bromo-1-(methylsulfonyl)pentane (160 mg, 0.700 mmol) and (2-methoxyphenyl)zinc iodide (0.910 mmol) were used. The product was purified by column chromatography (20%→25% ethyl acetate/hexanes). Colorless oil. First run: 148 mg (82%, 96% ee). Second run: 154 mg (86%, 96% ee).

The ee was determined by HPLC on a CHIRALCEL OD-H column (5% *i*-PrOH/hexanes, 1.0 mL/min) with $t_r = 17.6$ min (minor), 18.9 min (major).

¹H NMR (500 MHz, CDCl₃) δ 7.52 (dd, 1H, *J* = 7.8, 1.7 Hz), 7.34 (ddd, 1H, *J* = 8.3, 7.4, 1.7 Hz), 7.04 (ddd, 1H, *J* = 7.6, 7.6, 1.1 Hz), 6.93 (dd, 1H, *J* = 8.3, 1.1 Hz), 4.81 (dd, 1H, *J* = 11.5, 3.9 Hz),

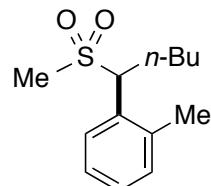
3.87 (s, 3H), 2.58 (s, 3H), 2.40 (dd, 1H, J = 13.5, 9.6, 6.9, 3.9 Hz), 2.05 (dd, 1H, J = 13.5, 11.5, 9.4, 5.3 Hz), 1.38–1.24 (m, 2H), 1.23–1.12 (m, 2H), 0.84 (t, 3H, J = 7.3 Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 157.6, 130.1, 129.2, 121.7, 121.6, 110.9, 60.4, 55.9, 38.6, 28.7, 25.9, 22.4, 13.9.

FT-IR (neat) 3009, 2957, 2872, 1601, 1587, 1494, 1464, 1440, 1412, 1380, 1296, 1247, 1192, 1164, 1137, 1090, 1051, 1025, 956, 792, 755 cm^{-1} .

MS (ESI) m/z (M^++Na) calcd for $\text{C}_{13}\text{H}_{20}\text{NaO}_3\text{S}$: 279, found: 279.

$[\alpha]^{25}_{\text{D}} = +61^\circ$ ($c = 1.00$, CHCl_3).



(S)-1-Methyl-2-(1-(methylsulfonyl)pentyl)benzene (Table 4, Entry 10). 1-Bromo-1-(methylsulfonyl)pentane (160 mg, 0.700 mmol) and *o*-tolylzinc iodide (0.910 mmol) were used. The product was purified by column chromatography (15%→20% ethyl acetate/hexanes). Colorless oil. First run: 137 mg (81%, 97% ee). Second run: 134 mg (80%, 97% ee).

The ee was determined by HPLC on a CHIRALPAK AS-H column (10% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 18.4 min (minor), 28.0 min (major).

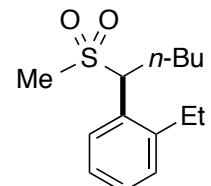
^1H NMR (500 MHz, CD_2Cl_2) δ 7.52–7.50 (m, 1H), 7.30–7.23 (m, 3H), 4.37 (dd, 1H, J = 11.4, 3.7 Hz), 2.61 (s, 3H), 2.40 (s, 3H), 2.37–2.31 (m, 1H), 2.12–2.04 (m, 1H), 1.40–1.24 (m, 2H), 1.24–1.10 (m, 2H), 0.84 (t, 3H, J = 7.3 Hz).

^{13}C NMR (126 MHz, CD_2Cl_2) δ 138.4, 132.0, 131.2, 129.0, 127.9, 127.1, 65.0, 38.7, 29.1, 28.5, 22.8, 20.3, 13.9.

FT-IR (neat) 3025, 2957, 2931, 2872, 1493, 1464, 1411, 1380, 1294, 1224, 1208, 1177, 1138, 1113, 1051, 958, 825, 796, 771, 736 cm^{-1} .

MS (ESI) m/z (M^++Na) calcd for $\text{C}_{13}\text{H}_{20}\text{NaO}_2\text{S}$: 263, found: 263.

$[\alpha]^{25}_{\text{D}} = +24.2^\circ$ ($c = 0.99$, CHCl_3).



(S)-1-Ethyl-2-(1-(methylsulfonyl)pentyl)benzene (Table 4, Entry 11). 1-Bromo-1-(methylsulfonyl)pentane (160 mg, 0.700 mmol), (2-ethylphenyl)zinc iodide (1.40 mmol), $\text{NiCl}_2 \cdot \text{glyme}$ (30.8 mg, 0.140 mmol), and (*R,R*)-L1 (60.9 mg, 0.182 mmol) were used. The product was purified by column chromatography (15% ethyl acetate/hexanes). Light-yellow oil. First run: 145 mg (81%, 98% ee). Second run: 146 mg (82%, 98% ee).

The ee was determined by HPLC on a CHIRALPAK AS-H column (10% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 13.1 min (minor), 22.1 min (major).

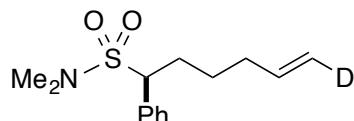
^1H NMR (500 MHz, CD_2Cl_2) δ 7.52–7.51 (m, 1H), 7.34–7.26 (m, 3H), 4.41 (dd, 1H, J = 11.2, 3.9 Hz), 2.82–2.75 (m, 1H), 2.73–2.66 (m, 1H), 2.62 (s, 3H), 2.35 (dddd, 1H, J = 13.5, 11.0, 5.7, 3.8 Hz), 2.12–2.04 (m, 1H), 1.41–1.20 (m, 3H), 1.23 (t, 3H, J = 7.6 Hz), 1.19–1.09 (m, 1H), 0.85 (t, 3H, J = 7.2 Hz).

^{13}C NMR (126 MHz, CD_2Cl_2) δ 144.4, 131.2, 129.6, 129.1, 127.9, 126.9, 64.5, 38.8, 29.3, 28.7, 26.2, 23.0, 15.7, 13.9.

FT-IR (neat) 3063, 3026, 2960, 2932, 2873, 1491, 1453, 1411, 1379, 1294, 1218, 1176, 1138, 1113, 1061, 958, 831, 797, 757 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{14}\text{H}_{22}\text{NaO}_2\text{S}$: 277, found: 277.

$[\alpha]^{25}_{\text{D}} = +24.1^\circ$ ($c = 1.01$, CHCl_3).



(*S,E*)-*N,N*-Dimethyl-1-phenylhex-5-ene-1-sulfonamide-6-*d* (eq 3). White solid.

The ee was determined by HPLC on a CHIRALCEL OD-H column (1% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 13.9 min (major), 17.4 min (minor).

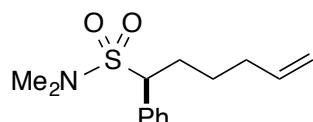
^1H NMR (500 MHz, CDCl_3) δ 7.42–7.34 (m, 5H), 5.70 (dt, 1H, J = 17.0, 6.5 Hz), 4.95 (dt, 1H, J = 17.1, 1.6 Hz), 4.09 (dd, 1H, J = 11.2, 3.9 Hz), 2.53 (s, 6H), 2.32 (dddd, 1H, J = 13.9, 10.3, 6.4, 3.9 Hz), 2.20–2.12 (m, 1H), 2.10–1.98 (m, 2H), 1.36–1.22 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 137.8, 134.0, 129.6, 129.0, 128.9, 115.0 (t, J = 24 Hz), 67.8, 37.8, 33.3, 29.5, 26.2.

FT-IR (neat) 3088, 3065, 3024, 2926, 2860, 2822, 2261, 1623, 1496, 1480, 1456, 1436, 1326, 1292, 1256, 1200, 1140, 1064, 1043, 984, 970, 917, 906, 822, 799, 778, 745 cm^{-1} .

MS (EI) m/z ($\text{M}^+ - \text{SO}_2\text{NMe}_2$) calcd for $\text{C}_{12}\text{H}_{14}\text{D}$: 160, found: 160.

$[\alpha]^{25}_{\text{D}} = -34^\circ$ ($c = 0.99$, CHCl_3); 96% ee.



(*S*)-*N,N*-Dimethyl-1-phenylhex-5-ene-1-sulfonamide (Figure 1). White solid.

The ee was determined by HPLC on a CHIRALCEL OD-H column (1% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 14.1 min (major), 17.8 min (minor).

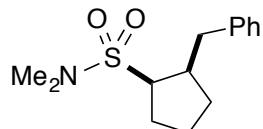
^1H NMR (500 MHz, CDCl_3) δ 7.42–7.34 (m, 5H), 5.70 (ddt, 1H, J = 17.0, 10.3, 6.7 Hz), 4.98–4.92 (m, 2H), 4.09 (dd, 1H, J = 11.2, 3.9 Hz), 2.53 (s, 6H), 2.32 (dddd, 1H, J = 14.1, 10.3, 6.3, 3.9 Hz), 2.20–2.12 (m, 1H), 2.10–1.98 (m, 2H), 1.36–1.22 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 138.0, 134.0, 129.6, 129.0, 128.9, 115.3, 67.8, 37.8, 33.4, 29.5, 26.2.

FT-IR (neat) 3067, 3033, 2934, 2908, 2868, 2821, 1640, 1497, 1480, 1455, 1417, 1329, 1282, 1199, 1141, 1063, 1043, 993, 966, 916, 906, 870, 814, 781, 746, 735 cm⁻¹.

MS (EI) *m/z* (M⁺–SO₂NMe₂) calcd for C₁₂H₁₅: 159, found: 159.

[α]²⁵_D = -33° (c = 0.82, CHCl₃); 97% ee.



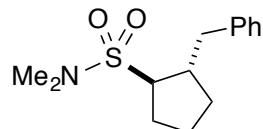
syn-2-Benzyl-N,N-dimethylcyclopentane-1-sulfonamide (Figure 1). White solid.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.30–7.26 (m, 2H), 7.20–7.16 (m, 3H), 3.54 (ddd, 1H, *J* = 8.7, 8.7, 6.3 Hz), 3.31–3.25 (m, 1H), 2.89 (s, 6H), 2.60–2.52 (m, 2H), 2.15–2.07 (m, 1H), 2.04–1.97 (m, 1H), 1.95–1.87 (m, 1H), 1.66–1.44 (m, 3H).

¹³C NMR (126 MHz, CD₂Cl₂) δ 141.8, 129.4, 128.6, 126.2, 62.8, 44.7, 37.8, 35.7, 29.7, 26.8, 22.7.

FT-IR (neat) 3084, 3060, 3024, 2922, 2874, 2850, 2806, 1602, 1583, 1495, 1473, 1452, 1332, 1273, 1195, 1136, 1073, 1058, 1029, 958, 845, 822, 727 cm⁻¹.

MS (EI) *m/z* (M⁺) calcd for C₁₄H₂₁NO₂S: 267, found: 267.



anti-2-Benzyl-N,N-dimethylcyclopentane-1-sulfonamide (Figure 1). Colorless oil.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.32–7.28 (m, 2H), 7.22–7.19 (m, 3H), 3.20 (ddd, 1H, *J* = 8.9, 6.1, 6.1 Hz), 2.97 (dd, 1H, *J* = 12.6, 4.9 Hz), 2.78 (s, 6H), 2.65–2.53 (m, 2H), 2.08–1.95 (m, 2H), 1.82–1.74 (m, 1H), 1.73–1.61 (m, 2H), 1.42–1.35 (m, 1H).

¹³C NMR (126 MHz, CD₂Cl₂) δ 140.7, 129.6, 128.7, 126.5, 64.5, 43.6, 41.3, 37.8, 32.1, 28.5, 24.9.

FT-IR (neat) 3084, 3060, 3025, 2917, 2849, 1602, 1583, 1494, 1461, 1453, 1435, 1315, 1199, 1136, 1082, 1059, 1029, 960, 882, 849, 733 cm⁻¹.

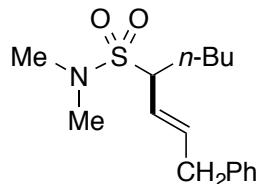
MS (EI) *m/z* (M⁺) calcd for C₁₄H₂₁NO₂S: 267, found: 267.

IV. Enantioselective Alkenylations

General Procedure. Cp₂ZrHCl (Schwartz's reagent; 258 mg, 1.00 mmol) was added to an oven-dried 4-mL vial equipped with a magnetic stir bar, and then the vial was capped with a PTFE-lined septum cap. The vial was evacuated and refilled with nitrogen (three cycles). 1,2-Dimethoxyethane (1.00 ml) was added to the vial, followed by the alkyne (1.00 mmol). The reaction mixture was stirred at r.t. for 1.5 h, at which time it had become homogenous. An oven-dried 20-mL vial equipped with a magnetic stir bar was charged with NiCl₂·glyme (11.0 mg, 0.050 mmol), (3*R*,8*S*)-L6 (23.3 mg, 0.065 mmol), and the electrophile (0.500 mmol). The vial

was sealed with a PTFE-lined septum cap, placed under vacuum, and then filled with nitrogen. This evacuation-refill cycle was repeated three times. 1,2-Dimethoxyethane (2.57 mL) was added, and the mixture was stirred at r.t. for 1 h. The solution of the nucleophile was transferred by syringe over 2 min to the vial that contained the electrophile. The reaction mixture was stirred at r.t. for 24 h, and then the reaction was quenched by the addition of ethanol (0.50 mL). The solution was filtered through a pad of silica (eluted with Et₂O). The filtrate was concentrated, and the resulting residue was purified by column chromatography.

A second run was conducted with (3*S*,8*R*)-L6.



(*S,E*)-*N,N*-Dimethyl-1-phenyloct-2-ene-4-sulfonamide (Table 5, Entry 1). 1-Bromo-*N,N*-dimethylpentane-1-sulfonamide (129 mg, 0.500 mmol) and (*E*)-(3-phenylprop-1-en-1-yl)zirconium reagent (1.00 mmol) were used. The product was purified by column chromatography on silica gel (15% ethyl acetate/hexanes). Colorless oil. First run: 116 mg (79%, 91% ee). Second run: 122 mg (83%, 90% ee).

The ee was determined by HPLC on a CHIRALCEL OJ-H column (5% *i*-PrOH/hexanes, 1.0 mL/min) with *t*_r = 18.2 min (major), 20.9 min (minor).

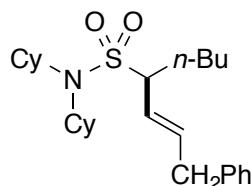
¹H NMR (500 MHz, CDCl₃) δ 7.32–7.28 (m, 2H), 7.24–7.20 (m, 1H), 7.18–7.15 (m, 2H), 5.87 (dd, 1H, *J* = 15.3, 6.9, 6.9, 0.4 Hz), 5.44 (ddd, 1H, *J* = 15.3, 9.7, 1.5, 1.5 Hz), 3.57 (ddd, 1H, *J* = 10.5, 10.5, 3.5 Hz), 3.49–3.39 (m, 2H), 2.82 (s, 6H), 2.04–1.97 (m, 1H), 1.75–1.68 (m, 1H), 1.41–1.18 (m, 4H), 0.89 (t, 3H, *J* = 7.1 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 139.3, 136.9, 128.7, 128.6, 126.5, 125.1, 65.8, 39.1, 38.3, 28.8, 28.7, 22.4, 14.0.

FT-IR (neat) 3061, 3027, 2954, 2930, 2871, 1663, 1603, 1494, 1453, 1379, 1329, 1281, 1198, 1139, 1076, 1062, 1029, 966, 804, 747, 730 cm⁻¹.

MS (EI) *m/z* (M⁺–SO₂NMe₂) calcd for C₁₄H₁₉: 187, found: 187.

[α]_D²⁵ = +22.9° (c = 1.01, CHCl₃).



(*S,E*)-*N,N*-Dicyclohexyl-1-phenyloct-2-ene-4-sulfonamide (Table 5, Entry 2). 1-Bromo-*N,N*-dicyclohexylpentane-1-sulfonamide (197 mg, 0.500 mmol) and (*E*)-(3-phenylprop-1-en-1-yl)zirconium reagent (1.00 mmol) were used. The product was purified by column chromatography on silica gel (5% Et₂O/hexanes) and then on C-18 silica gel (10%→100%.

acetonitrile/water). Viscous light-yellow oil. First run: 179 mg (83%, 95% ee). Second run: 180 mg (83%, 94% ee).

The ee was determined by HPLC on a CHIRALPAK AD-H column (1% *i*-PrOH/hexanes, 0.6 mL/min) with $t_r = 16.6$ min (major), 17.7 min (minor).

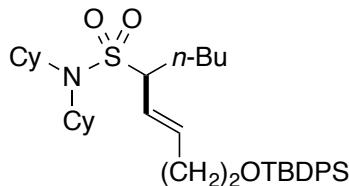
^1H NMR (500 MHz, CD_2Cl_2) δ 7.31–7.28 (m, 2H), 7.22–7.17 (m, 3H), 5.80 (dd, 1H, $J = 15.3, 7.4, 6.0, 0.4$ Hz), 5.42 (ddd, 1H, $J = 15.4, 9.8, 1.5, 1.5$ Hz), 3.48–3.38 (m, 2H), 3.33 (dd, 1H, $J = 10.8, 10.0, 3.2$ Hz), 3.17–3.10 (m, 2H), 2.04–1.97 (m, 1H), 1.79–1.56 (m, 15H), 1.41–1.16 (m, 8H), 1.08 (qt, 2H, $J = 13.1, 3.4$ Hz), 0.89 (t, 3H, $J = 7.2$ Hz).

^{13}C NMR (126 MHz, CD_2Cl_2) δ 140.1, 136.6, 128.9, 128.8, 126.6, 126.0, 69.0, 58.4, 39.2, 34.0, 33.1, 29.5, 29.3, 26.99, 26.97, 25.8, 22.7, 14.1.

FT-IR (neat) 3084, 3062, 3027, 2931, 2855, 1603, 1495, 1466, 1453, 1401, 1381, 1322, 1274, 1256, 1188, 1164, 1139, 1108, 1074, 1047, 1028, 981, 895, 854, 823, 750 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{26}\text{H}_{41}\text{NNaO}_2\text{S}$: 454, found: 454.

$[\alpha]^{25}_D = -6.9^\circ$ ($c = 1.02$, CHCl_3).



(*S,E*)-1-((tert-Butyldiphenylsilyl)oxy)-*N,N*-dicyclohexylnon-3-ene-5-sulfonamide (Table 5, Entry 3). 1-Bromo-*N,N*-dicyclohexylpentane-1-sulfonamide (197 mg, 0.500 mmol) and (*E*)-(*4*-((*tert*-butyldiphenylsilyl)oxy)but-1-en-1-yl)zirconium reagent (1.00 mmol) were used. The product was purified by column chromatography on silica gel (3% ethyl acetate/hexanes) and then on C-18 silica gel (10% → 100% acetonitrile/water). Viscous light-yellow oil. First run: 254 mg (81%, 95% ee). Second run: 259 mg (83%, 94% ee).

The ee was determined by HPLC on a CHIRALPAK AD-H column (0.5% *i*-PrOH/hexanes, 0.8 mL/min) with $t_r = 14.6$ min (minor), 17.9 min (major).

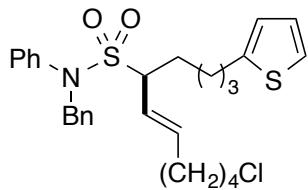
^1H NMR (500 MHz, CDCl_3) δ 7.67–7.64 (m, 4H), 7.45–7.41 (m, 2H), 7.40–7.36 (m, 4H), 5.69 (ddd, 1H, $J = 15.5, 6.4, 6.4$ Hz), 5.42 (ddd, 1H, $J = 15.5, 9.7, 1.4, 1.4$ Hz), 3.75–3.68 (m, 2H), 3.26 (ddd, 1H, $J = 10.6, 9.5, 3.2$ Hz), 3.15–3.09 (m, 2H), 2.39–2.29 (m, 2H), 2.08–2.01 (m, 1H), 1.76–1.57 (m, 14H), 1.39–1.14 (m, 9H), 1.12–1.02 (m, 2H), 1.05 (s, 9H), 0.86 (t, 3H, $J = 7.2$ Hz).

^{13}C NMR (126 MHz, CDCl_3) δ 135.7, 135.6, 134.3, 133.9, 133.8, 129.79, 129.78, 127.79, 127.78, 126.10, 69.5, 63.2, 58.2, 36.0, 33.9, 32.8, 29.3, 29.0, 26.9, 26.7, 25.5, 22.5, 19.3, 14.0.

FT-IR (neat) 3071, 3048, 2931, 2856, 1590, 1471, 1453, 1428, 1389, 1323, 1257, 1221, 1188, 1164, 1138, 1110, 1048, 1028, 998, 980, 939, 895, 854, 822, 764, 738 cm^{-1} .

MS (ESI) m/z ($\text{M}^+ + \text{Na}$) calcd for $\text{C}_{37}\text{H}_{57}\text{NNaO}_3\text{SSi}$: 646, found: 646.

$[\alpha]^{25}_D = +1.7^\circ$ ($c = 0.99$, CHCl_3).



(*S,E*)-*N*-Benzyl-11-chloro-*N*-phenyl-1-(thiophen-2-yl)undec-6-ene-5-sulfonamide (Table 5, Entry 4). *N*-Benzyl-1-bromo-*N*-phenyl-5-(thiophen-2-yl)pentane-1-sulfonamide (239 mg, 0.500 mmol) and (*E*)-(6-chlorohex-1-en-1-yl)zirconium reagent (1.00 mmol) were used. The product was purified by column chromatography (first purification: 5% ethyl acetate/hexanes; second purification: 15% cyclopentyl methyl ether/hexanes). Viscous light-yellow oil. First run: 165 mg (64%, 80% ee). Second run: 156 mg (60%, 81% ee).

The ee was determined by HPLC on a CHIRALPAK AD-H column (10% *i*-PrOH/hexanes, 0.8 mL/min) with t_r = 17.5 min (major), 23.8 min (minor).

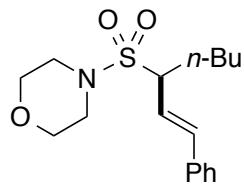
^1H NMR (500 MHz, CD_2Cl_2) δ 7.32–7.28 (m, 2H), 7.27–7.19 (m, 8H), 7.12 (dd, 1H, J = 5.1, 1.2 Hz), 6.91 (dd, 1H, J = 5.1, 3.4 Hz), 6.77 (dddd, 1H, J = 3.3, 1.0, 1.0, 1.0 Hz), 5.80 (ddd, 1H, J = 15.3, 6.8, 6.8 Hz), 5.46 (dddd, 1H, J = 15.4, 9.7, 1.5, 1.5 Hz), 4.99 (d, 1H, J = 15.1 Hz), 4.67 (d, 1H, J = 15.1 Hz), 3.60–3.55 (m, 1H), 3.58 (t, 2H, J = 6.6 Hz), 2.87–2.76 (m, 2H), 2.26–2.13 (m, 2H), 2.04 (dddd, 1H, J = 13.6, 9.9, 6.3, 3.4 Hz), 1.86–1.72 (m, 3H), 1.72–1.56 (m, 4H), 1.50–1.41 (m, 1H), 1.32–1.22 (m, 1H).

^{13}C NMR (126 MHz, CD_2Cl_2) δ 145.6, 139.7, 139.0, 137.3, 129.4, 129.3, 128.7, 128.6, 127.9, 127.8, 127.0, 124.5, 124.2, 123.2, 66.7, 56.7, 45.4, 32.5, 32.2, 31.6, 29.9, 29.2, 26.5, 26.2.

FT-IR (neat) 3064, 3032, 2933, 2860, 1595, 1493, 1454, 1337, 1216, 1145, 1093, 1065, 1028, 976, 916, 862, 775 cm^{-1} .

MS (ESI) m/z (M $^+$ +Na) calcd for $\text{C}_{28}\text{H}_{34}\text{ClNNaO}_2\text{S}_2$: 538, found: 538.

$[\alpha]^{25}_D$ = -23.0° (c = 1.03, CHCl_3).



(*S,E*)-4-((1-Phenylhept-1-en-3-yl)sulfonyl)morpholine (Table 5, Entry 5). 4-((1-Bromopentyl)sulfonyl)morpholine (150 mg, 0.500 mmol), and (*E*)-styrylzirconium reagent (1.00 mmol) were used. The product was purified by column chromatography (15% ethyl acetate/hexanes). White solid. First run: 110 mg (68%, 97% ee). Second run: 110 mg (68%, 95% ee).

The ee was determined by HPLC on a CHIRALPAK AS-H column (10% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 18.7 min (minor), 30.1 min (major).

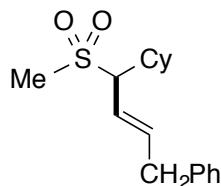
^1H NMR (500 MHz, CDCl_3) δ 7.42–7.39 (m, 2H), 7.38–7.34 (m, 2H), 7.32–7.29 (m, 1H), 6.62 (d, 1H, J = 15.9 Hz), 6.06 (dd, 1H, J = 15.9, 9.8 Hz), 3.70–3.62 (m, 5H), 3.38–3.30 (m, 4H), 2.17–2.10 (m, 1H), 1.87–1.79 (m, 1H), 1.45–1.22 (m, 4H), 0.90 (t, 3H, J = 7.0 Hz).

¹³C NMR (126 MHz, CDCl₃) δ 137.0, 135.7, 129.0, 128.7, 126.7, 122.7, 67.1, 67.0, 46.8, 29.0, 28.8, 22.4, 14.0.

FT-IR (neat) 2958, 2923, 2859, 1450, 1339, 1324, 1260, 1148, 1114, 1073, 955, 743 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₇H₂₅NNaO₃S: 346, found: 346.

[α]²⁵_D = -82° (c = 0.98, CHCl₃).



(S,E)-(4-Cyclohexyl-4-(methylsulfonyl)but-2-en-1-yl)benzene (Table 5, Entry 6).

(Bromo(methylsulfonyl)methyl)cyclohexane (179 mg, 0.700 mmol), (*E*)-(3-phenylprop-1-en-1-yl)zirconium reagent (1.40 mmol), and (*R,R*)-L1 (30.4 mg, 0.091 mmol) were used. The product was purified by column chromatography (15% ethyl acetate/hexanes). Light-yellow oil. First run: 109 mg (53%, 93% ee). Second run: 98 mg (48%, 93% ee).

The ee was determined by HPLC on a CHIRALPAK IB-3 column (5% *i*-PrOH/hexanes, 1.0 mL/min) with t_r = 13.0 min (minor), 22.9 min (major).

¹H NMR (500 MHz, CDCl₃) δ 7.33–7.30 (m, 2H), 7.25–7.21 (m, 1H), 7.18–7.16 (m, 2H), 5.89 (ddd, 1H, *J* = 15.2, 6.9, 6.9 Hz), 5.71 (dddd, 1H, *J* = 15.3, 10.4, 1.4, 1.4 Hz), 3.48 (d, 2H, *J* = 6.9 Hz), 3.29 (dd, 1H, *J* = 10.4, 3.8 Hz), 2.76 (s, 3H), 2.32 (tq, 1H, *J* = 11.9, 3.5 Hz), 2.08–2.02 (m, 1H), 1.78–1.72 (m, 2H), 1.70–1.61 (m, 2H), 1.40–1.26 (m, 2H), 1.23–1.07 (m, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 139.2, 139.0, 128.8, 128.6, 126.6, 122.6, 73.2, 39.8, 39.3, 36.0, 32.2, 28.9, 26.4, 26.1, 26.0.

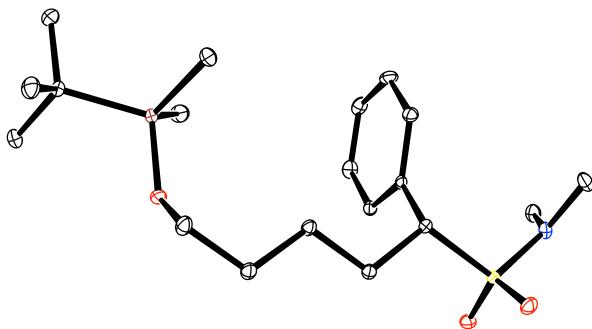
FT-IR (neat) 3083, 3060, 3026, 2927, 2852, 1660, 1602, 1494, 1452, 1411, 1351, 1295, 1240, 1173, 1133, 1077, 1029, 978, 894, 852, 784, 751, 700 cm⁻¹.

MS (ESI) *m/z* (M⁺+Na) calcd for C₁₇H₂₄NaO₂S: 315, found: 315.

[α]²⁵_D = +60.8° (c = 1.00, CHCl₃).

V. Determination of Absolute Stereochemistry

Product from entry 7 of Table 2 (run with (S,S)-L1). (R)-5-((tert-Butyldimethylsilyl)oxy)-N,N-dimethyl-1-phenylpentane-1-sulfonamide. A crystal suitable for X-ray crystallography was grown by vapor diffusion with dichloromethane and pentane.



A suitable crystal of $C_{19}H_{35}NO_3SSi$ was selected for analysis. All measurements were made on a Bruker SMART 1000 CCD with filtered Mo-K α radiation at a temperature of 100 K. Using Olex2,⁶ the structure was solved with the ShelXS⁷ structure solution program using Direct Methods and refined with the ShelXL⁷ refinement package using Least Squares minimization. The absolute stereochemistry was determined on the basis of the absolute structure parameter.

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- (6) Dolomanov, O. V.; Bourhis, L. J.; Gildea, R. J.; Howard, J. A. K.; Puschmann, H. *J. Appl. Crystallogr.* **2009**, *42*, 339–341.
(7) Sheldrick, G. M. *Acta Cryst.* **2008**, *A64*, 112–122.

Table S–1. Crystal data and structure refinement for crystal01.

Identification code	crystal01		
Empirical formula	$C_{19}H_{35}NO_3SSi$		
Formula weight	385.63		
Temperature	100 K		
Wavelength	0.71073 Å		
Crystal system	Monoclinic		
Space group	$P2_1$		
Unit cell dimensions	$a = 5.9209(6)$ Å	$\alpha = 90^\circ$.	
	$b = 10.6607(12)$ Å	$\beta = 99.2230(10)^\circ$.	
	$c = 17.0647(19)$ Å	$\gamma = 90^\circ$.	
Volume	$1063.2(2)$ Å ³		
Z	2		
Density (calculated)	1.205 Mg/m ³		
Absorption coefficient	0.226 mm ⁻¹		
F(000)	420		
Crystal size	$0.4 \times 0.4 \times 0.1$ mm ³		
Theta range for data collection	1.209 to 29.107°.		
Index ranges	$-7 \leq h \leq 8, -13 \leq k \leq 14, -22 \leq l \leq 23$		
Reflections collected	16770		
Independent reflections	5160 [R(int) = 0.0236]		
Completeness to theta = 25.000°	100.0 %		
Absorption correction	Semi-empirical from equivalents		
Max. and min. transmission	1.0000 and 0.9257		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	5160 / 1 / 233		
Goodness-of-fit on F ²	1.098		
Final R indices [I>2sigma(I)]	R1 = 0.0279, wR2 = 0.0669		
R indices (all data)	R1 = 0.0308, wR2 = 0.0690		
Absolute structure parameter	0.02(2)		
Largest diff. peak and hole	0.325 and -0.163 e/Å ⁻³		

Table S–2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal01. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
S(1)	-4031(1)	5489(1)	517(1)	14(1)
Si(1)	2618(1)	1014(1)	3465(1)	14(1)
O(1)	-3746(3)	5244(1)	-291(1)	20(1)
O(2)	-6212(2)	5232(1)	755(1)	19(1)
O(3)	648(2)	432(2)	2769(1)	19(1)
N(1)	-3503(3)	6967(2)	689(1)	16(1)
C(1)	-4272(4)	7610(2)	1361(1)	20(1)
C(2)	-1459(4)	7506(2)	435(1)	22(1)
C(3)	-1857(3)	4582(2)	1132(1)	12(1)
C(4)	-2416(3)	3172(2)	1032(1)	15(1)
C(5)	-552(3)	2381(2)	1521(1)	16(1)
C(6)	-952(4)	967(2)	1416(1)	18(1)
C(7)	820(4)	204(2)	1957(1)	21(1)
C(8)	-1531(3)	5030(2)	1983(1)	12(1)
C(9)	508(3)	5621(2)	2311(1)	17(1)
C(10)	842(3)	6027(2)	3095(1)	21(1)
C(11)	-835(4)	5849(2)	3558(1)	21(1)
C(12)	-2858(4)	5251(2)	3243(1)	20(1)
C(13)	-3211(3)	4849(2)	2456(1)	16(1)
C(14)	1116(4)	2120(2)	4054(1)	24(1)
C(15)	4837(4)	1893(2)	3029(1)	27(1)
C(16)	3960(3)	-312(2)	4110(1)	15(1)
C(17)	5647(4)	216(2)	4811(1)	23(1)
C(18)	2103(4)	-1074(2)	4431(1)	22(1)
C(19)	5248(4)	-1178(2)	3614(1)	24(1)

Table S–3. Bond lengths [\AA] and angles [$^\circ$] for crystal01.

S(1)-O(1)	1.4394(15)
S(1)-O(2)	1.4405(15)
S(1)-N(1)	1.6247(18)
S(1)-C(3)	1.8048(19)
Si(1)-O(3)	1.6471(14)
Si(1)-C(14)	1.865(2)
Si(1)-C(15)	1.863(2)
Si(1)-C(16)	1.886(2)
O(3)-C(7)	1.426(2)
N(1)-C(1)	1.469(3)
N(1)-C(2)	1.466(3)
C(3)-C(4)	1.543(3)
C(3)-C(8)	1.512(3)
C(4)-C(5)	1.526(3)
C(5)-C(6)	1.532(3)
C(6)-C(7)	1.517(3)
C(8)-C(9)	1.396(3)
C(8)-C(13)	1.391(3)
C(9)-C(10)	1.390(3)
C(10)-C(11)	1.378(3)
C(11)-C(12)	1.387(3)
C(12)-C(13)	1.393(3)
C(16)-C(17)	1.537(3)
C(16)-C(18)	1.537(3)
C(16)-C(19)	1.535(3)
O(1)-S(1)-O(2)	118.93(9)
O(1)-S(1)-N(1)	107.45(9)
O(1)-S(1)-C(3)	106.10(9)
O(2)-S(1)-N(1)	106.79(9)
O(2)-S(1)-C(3)	108.81(9)
N(1)-S(1)-C(3)	108.40(9)
O(3)-Si(1)-C(14)	106.44(9)
O(3)-Si(1)-C(15)	111.35(9)
O(3)-Si(1)-C(16)	108.79(9)

C(14)-Si(1)-C(16)	110.66(10)
C(15)-Si(1)-C(14)	108.96(11)
C(15)-Si(1)-C(16)	110.57(10)
C(7)-O(3)-Si(1)	127.79(13)
C(1)-N(1)-S(1)	121.23(14)
C(2)-N(1)-S(1)	118.06(14)
C(2)-N(1)-C(1)	114.98(17)
C(4)-C(3)-S(1)	109.70(13)
C(8)-C(3)-S(1)	111.03(13)
C(8)-C(3)-C(4)	113.95(16)
C(5)-C(4)-C(3)	110.75(15)
C(4)-C(5)-C(6)	113.27(16)
C(7)-C(6)-C(5)	112.19(16)
O(3)-C(7)-C(6)	110.46(17)
C(9)-C(8)-C(3)	119.76(17)
C(13)-C(8)-C(3)	121.38(17)
C(13)-C(8)-C(9)	118.86(18)
C(10)-C(9)-C(8)	120.46(18)
C(11)-C(10)-C(9)	120.29(19)
C(10)-C(11)-C(12)	119.91(19)
C(11)-C(12)-C(13)	120.06(19)
C(8)-C(13)-C(12)	120.41(19)
C(17)-C(16)-Si(1)	109.87(14)
C(17)-C(16)-C(18)	109.15(17)
C(18)-C(16)-Si(1)	110.21(14)
C(19)-C(16)-Si(1)	109.22(14)
C(19)-C(16)-C(17)	109.34(17)
C(19)-C(16)-C(18)	109.03(17)

Table S-4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal01. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

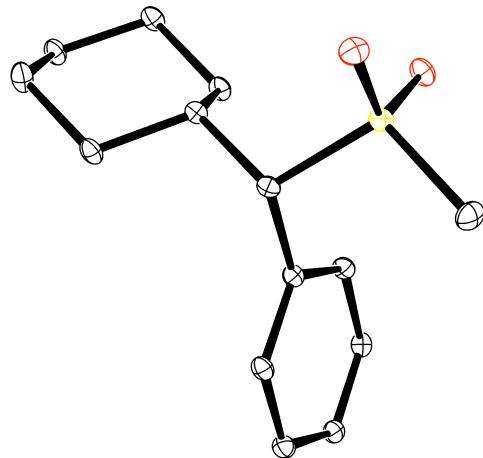
	U11	U22	U33	U23	U13	U12
S(1)	14(1)	14(1)	12(1)	1(1)	-1(1)	0(1)
Si(1)	14(1)	12(1)	15(1)	2(1)	1(1)	-1(1)
O(1)	26(1)	21(1)	13(1)	0(1)	-2(1)	0(1)
O(2)	14(1)	19(1)	23(1)	1(1)	-1(1)	-1(1)
O(3)	21(1)	20(1)	14(1)	2(1)	-2(1)	-6(1)
N(1)	20(1)	13(1)	16(1)	2(1)	4(1)	1(1)
C(1)	24(1)	14(1)	22(1)	0(1)	5(1)	3(1)
C(2)	27(1)	17(1)	24(1)	2(1)	8(1)	-6(1)
C(3)	12(1)	13(1)	11(1)	1(1)	1(1)	2(1)
C(4)	18(1)	13(1)	12(1)	-2(1)	1(1)	0(1)
C(5)	19(1)	14(1)	13(1)	-1(1)	-1(1)	2(1)
C(6)	24(1)	15(1)	14(1)	-1(1)	-2(1)	0(1)
C(7)	27(1)	15(1)	18(1)	-1(1)	-1(1)	4(1)
C(8)	15(1)	10(1)	12(1)	1(1)	1(1)	1(1)
C(9)	14(1)	20(1)	18(1)	1(1)	3(1)	0(1)
C(10)	18(1)	20(1)	22(1)	-5(1)	-5(1)	-1(1)
C(11)	30(1)	19(1)	14(1)	-3(1)	0(1)	5(1)
C(12)	24(1)	21(1)	16(1)	0(1)	8(1)	2(1)
C(13)	18(1)	14(1)	15(1)	1(1)	2(1)	0(1)
C(14)	23(1)	20(1)	29(1)	-5(1)	0(1)	4(1)
C(15)	23(1)	28(1)	29(1)	12(1)	2(1)	-8(1)
C(16)	16(1)	14(1)	15(1)	2(1)	1(1)	1(1)
C(17)	23(1)	25(1)	20(1)	5(1)	-4(1)	-2(1)
C(18)	24(1)	17(1)	24(1)	5(1)	4(1)	-2(1)
C(19)	24(1)	21(1)	29(1)	1(1)	5(1)	6(1)

Table S-5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal01.

	x	y	z	U(eq)
H(1A)	-5587	7185	1497	30
H(1B)	-4669	8461	1216	30
H(1C)	-3064	7603	1809	30
H(2A)	-208	7483	869	33
H(2B)	-1761	8359	270	33
H(2C)	-1070	7027	-1	33
H(3)	-413	4726	934	15
H(4A)	-3873	3002	1201	18
H(4B)	-2546	2944	476	18
H(5A)	-473	2592	2077	19
H(5B)	911	2591	1368	19
H(6A)	-2463	763	1529	22
H(6B)	-902	741	869	22
H(7A)	586	-681	1840	25
H(7B)	2340	428	1861	25
H(9)	1649	5743	2003	20
H(10)	2205	6421	3308	25
H(11)	-611	6130	4081	26
H(12)	-3980	5118	3558	24
H(13)	-4578	4458	2246	19
H(14A)	483	2801	3721	36
H(14B)	2183	2444	4490	36
H(14C)	-93	1686	4254	36
H(15A)	5788	1311	2802	40
H(15B)	5761	2371	3437	40
H(15C)	4107	2449	2624	40
H(17A)	6824	681	4613	35
H(17B)	6325	-463	5136	35
H(17C)	4846	759	5121	35
H(18A)	1374	-556	4777	32
H(18B)	2791	-1786	4721	32

H(18C)	986	-1356	3996	32
H(19A)	4214	-1474	3160	37
H(19B)	5860	-1880	3931	37
H(19C)	6475	-722	3439	37

Product from entry 2 of Table 3: (S)-(Cyclohexyl(methylsulfonyl)methyl)benzene (from a reaction using (R,R)-L1). A crystal suitable for X-ray crystallography was grown by vapor diffusion with dichloromethane and pentane.



A suitable crystal of C₁₄H₂₀O₂S was selected for analysis. All measurements were made on a Bruker APEX-II CCD with filtered Mo-K α radiation at a temperature of 100 K. Using Olex2,⁶ the structure was solved with the ShelXS⁷ structure solution program using Direct Methods and refined with the ShelXL⁷ refinement package using Least Squares minimization. The absolute stereochemistry was determined on the basis of the absolute structure parameter.

Table S–6. Crystal data and structure refinement for crystal03.

Identification code	crystal03		
Empirical formula	$C_{14}H_{20}O_2S$		
Formula weight	252.36		
Temperature	100.15 K		
Wavelength	0.71073 Å		
Crystal system	Orthorhombic		
Space group	$P2_12_12_1$		
Unit cell dimensions	$a = 6.2283(3)$ Å	$\alpha = 90^\circ$.	
	$b = 13.7866(7)$ Å	$\beta = 90^\circ$.	
	$c = 15.3937(8)$ Å	$\gamma = 90^\circ$.	
Volume	$1321.81(12)$ Å ³		
Z	4		
Density (calculated)	1.268 Mg/m ³		
Absorption coefficient	0.233 mm ⁻¹		
F(000)	544		
Crystal size	0.62 x 0.16 x 0.09 mm ³		
Theta range for data collection	1.983 to 33.731°.		
Index ranges	$-9 \leq h \leq 9, -21 \leq k \leq 20, -23 \leq l \leq 23$		
Reflections collected	39471		
Independent reflections	4910 [R(int) = 0.0621]		
Completeness to theta = 25.000°	100.0 %		
Absorption correction	Semi-empirical from equivalents		
Max. and min. transmission	1.0000 and 0.8575		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	4910 / 0 / 155		
Goodness-of-fit on F ²	1.141		
Final R indices [I>2sigma(I)]	R1 = 0.0540, wR2 = 0.1115		
R indices (all data)	R1 = 0.0741, wR2 = 0.1179		
Absolute structure parameter	0.01(3)		
Largest diff. peak and hole	0.692 and -0.385 e/Å ⁻³		

Table S–7. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal03. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
S(1)	7502(1)	3757(1)	4892(1)	17(1)
O(1)	6971(3)	3206(1)	5662(1)	21(1)
O(2)	9662(3)	4136(1)	4836(1)	21(1)
C(1)	5566(4)	4731(2)	4817(2)	14(1)
C(2)	5963(4)	5465(2)	5570(2)	14(1)
C(3)	7759(4)	6208(2)	5418(2)	17(1)
C(4)	8064(4)	6849(2)	6224(2)	20(1)
C(5)	5993(4)	7371(2)	6464(2)	21(1)
C(6)	4161(4)	6651(2)	6587(2)	23(1)
C(7)	3868(4)	5994(2)	5790(2)	19(1)
C(8)	5392(4)	5139(2)	3904(2)	15(1)
C(9)	7131(4)	5552(2)	3461(2)	17(1)
C(10)	6869(4)	5905(2)	2620(2)	18(1)
C(11)	4895(4)	5840(2)	2207(2)	18(1)
C(12)	3168(4)	5434(2)	2644(2)	19(1)
C(13)	3419(4)	5081(2)	3483(2)	17(1)
C(14)	7027(5)	3018(2)	3977(2)	23(1)

Table S–8. Bond lengths [\AA] and angles [$^\circ$] for crystal03.

S(1)-O(1)	1.4458(19)
S(1)-O(2)	1.4461(19)
S(1)-C(1)	1.809(2)
S(1)-C(14)	1.763(3)
C(1)-H(1)	1.0000
C(1)-C(2)	1.557(3)
C(1)-C(8)	1.518(3)
C(2)-H(2)	1.0000
C(2)-C(3)	1.535(3)
C(2)-C(7)	1.533(3)
C(3)-H(3A)	0.9900
C(3)-H(3B)	0.9900
C(3)-C(4)	1.536(3)
C(4)-H(4A)	0.9900
C(4)-H(4B)	0.9900
C(4)-C(5)	1.523(4)
C(5)-H(5A)	0.9900
C(5)-H(5B)	0.9900
C(5)-C(6)	1.524(4)
C(6)-H(6A)	0.9900
C(6)-H(6B)	0.9900
C(6)-C(7)	1.536(4)
C(7)-H(7A)	0.9900
C(7)-H(7B)	0.9900
C(8)-C(9)	1.401(3)
C(8)-C(13)	1.391(4)
C(9)-H(9)	0.9500
C(9)-C(10)	1.393(3)
C(10)-H(10)	0.9500
C(10)-C(11)	1.387(4)
C(11)-H(11)	0.9500
C(11)-C(12)	1.386(4)
C(12)-H(12)	0.9500
C(12)-C(13)	1.390(4)
C(13)-H(13)	0.9500

C(14)-H(14A)	0.9800
C(14)-H(14B)	0.9800
C(14)-H(14C)	0.9800
O(1)-S(1)-O(2)	116.89(12)
O(1)-S(1)-C(1)	106.82(11)
O(1)-S(1)-C(14)	108.23(11)
O(2)-S(1)-C(1)	110.34(10)
O(2)-S(1)-C(14)	108.51(13)
C(14)-S(1)-C(1)	105.43(13)
S(1)-C(1)-H(1)	105.6
C(2)-C(1)-S(1)	109.22(16)
C(2)-C(1)-H(1)	105.6
C(8)-C(1)-S(1)	112.41(17)
C(8)-C(1)-H(1)	105.6
C(8)-C(1)-C(2)	117.36(18)
C(1)-C(2)-H(2)	107.1
C(3)-C(2)-C(1)	115.9(2)
C(3)-C(2)-H(2)	107.1
C(7)-C(2)-C(1)	109.8(2)
C(7)-C(2)-H(2)	107.1
C(7)-C(2)-C(3)	109.60(18)
C(2)-C(3)-H(3A)	109.5
C(2)-C(3)-H(3B)	109.5
C(2)-C(3)-C(4)	110.6(2)
H(3A)-C(3)-H(3B)	108.1
C(4)-C(3)-H(3A)	109.5
C(4)-C(3)-H(3B)	109.5
C(3)-C(4)-H(4A)	109.4
C(3)-C(4)-H(4B)	109.4
H(4A)-C(4)-H(4B)	108.0
C(5)-C(4)-C(3)	111.3(2)
C(5)-C(4)-H(4A)	109.4
C(5)-C(4)-H(4B)	109.4
C(4)-C(5)-H(5A)	109.5
C(4)-C(5)-H(5B)	109.5
C(4)-C(5)-C(6)	110.9(2)

H(5A)-C(5)-H(5B)	108.1
C(6)-C(5)-H(5A)	109.5
C(6)-C(5)-H(5B)	109.5
C(5)-C(6)-H(6A)	109.2
C(5)-C(6)-H(6B)	109.2
C(5)-C(6)-C(7)	111.9(2)
H(6A)-C(6)-H(6B)	107.9
C(7)-C(6)-H(6A)	109.2
C(7)-C(6)-H(6B)	109.2
C(2)-C(7)-C(6)	110.9(2)
C(2)-C(7)-H(7A)	109.5
C(2)-C(7)-H(7B)	109.5
C(6)-C(7)-H(7A)	109.5
C(6)-C(7)-H(7B)	109.5
H(7A)-C(7)-H(7B)	108.1
C(9)-C(8)-C(1)	123.1(2)
C(13)-C(8)-C(1)	118.2(2)
C(13)-C(8)-C(9)	118.7(2)
C(8)-C(9)-H(9)	119.9
C(10)-C(9)-C(8)	120.2(2)
C(10)-C(9)-H(9)	119.9
C(9)-C(10)-H(10)	119.7
C(11)-C(10)-C(9)	120.5(2)
C(11)-C(10)-H(10)	119.7
C(10)-C(11)-H(11)	120.3
C(12)-C(11)-C(10)	119.4(2)
C(12)-C(11)-H(11)	120.3
C(11)-C(12)-H(12)	119.8
C(11)-C(12)-C(13)	120.4(2)
C(13)-C(12)-H(12)	119.8
C(8)-C(13)-H(13)	119.6
C(12)-C(13)-C(8)	120.8(2)
C(12)-C(13)-H(13)	119.6
S(1)-C(14)-H(14A)	109.5
S(1)-C(14)-H(14B)	109.5
S(1)-C(14)-H(14C)	109.5
H(14A)-C(14)-H(14B)	109.5

H(14A)-C(14)-H(14C) 109.5

H(14B)-C(14)-H(14C) 109.5

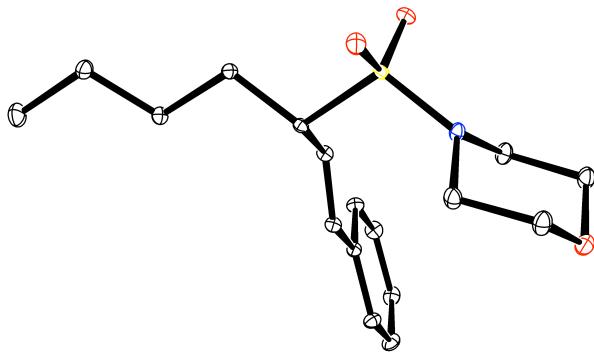
Table S–9. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal03. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U11	U22	U33	U23	U13	U12
S(1)	19(1)	13(1)	18(1)	1(1)	-1(1)	-1(1)
O(1)	26(1)	17(1)	20(1)	2(1)	-1(1)	-2(1)
O(2)	17(1)	16(1)	29(1)	2(1)	0(1)	0(1)
C(1)	11(1)	15(1)	17(1)	1(1)	-1(1)	-3(1)
C(2)	14(1)	11(1)	18(1)	2(1)	-1(1)	-2(1)
C(3)	11(1)	17(1)	24(1)	-2(1)	0(1)	-3(1)
C(4)	15(1)	16(1)	28(1)	-2(1)	-3(1)	-2(1)
C(5)	18(1)	15(1)	29(1)	-4(1)	-1(1)	2(1)
C(6)	17(1)	22(1)	31(2)	-6(1)	4(1)	0(1)
C(7)	12(1)	18(1)	26(1)	-2(1)	0(1)	0(1)
C(8)	15(1)	12(1)	18(1)	1(1)	0(1)	0(1)
C(9)	14(1)	17(1)	20(1)	0(1)	-1(1)	-3(1)
C(10)	19(1)	14(1)	20(1)	1(1)	4(1)	-1(1)
C(11)	22(1)	14(1)	18(1)	0(1)	-1(1)	3(1)
C(12)	18(1)	18(1)	21(1)	-2(1)	-3(1)	2(1)
C(13)	12(1)	17(1)	21(1)	0(1)	1(1)	-2(1)
C(14)	32(2)	14(1)	22(1)	-3(1)	-2(1)	-1(1)

Table S–10. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal03.

	x	y	z	U(eq)
H(1)	4140	4428	4942	17
H(2)	6373	5077	6093	17
H(3A)	7392	6620	4912	21
H(3B)	9117	5865	5286	21
H(4A)	8533	6442	6719	23
H(4B)	9203	7333	6109	23
H(5A)	5613	7837	6000	25
H(5B)	6210	7741	7009	25
H(6A)	2812	7011	6695	28
H(6B)	4457	6244	7103	28
H(7A)	2725	5513	5908	22
H(7B)	3415	6392	5286	22
H(9)	8494	5592	3736	20
H(10)	8051	6192	2327	21
H(11)	4728	6071	1629	22
H(12)	1806	5398	2368	23
H(13)	2229	4797	3773	20
H(14A)	7233	3400	3447	34
H(14B)	5551	2773	3996	34
H(14C)	8032	2471	3980	34

Product from entry 5 of Table 5 (run with (3*R*,8*S*)-L6). (*S,E*)-4-((1-Phenylhept-1-en-3-yl)sulfonyl)morpholine. A crystal suitable for X-ray crystallography was grown by vapor diffusion with Et₂O and pentane.



A suitable crystal of C₁₇H₂₅NO₃S was selected for analysis. All measurements were made on a Bruker APEX-II CCD with filtered Mo-K α radiation at a temperature of 100 K. Using Olex2,⁶ the structure was solved with the ShelXS⁷ structure solution program using Direct Methods and refined with the ShelXL⁷ refinement package using Least Squares minimization. The absolute stereochemistry was determined on the basis of the absolute structure parameter.

Table S–11. Crystal data and structure refinement for crystal02.

Identification code	crystal02		
Empirical formula	$C_{17}H_{25}NO_3S$		
Formula weight	323.44		
Temperature	100 K		
Wavelength	0.71073 Å		
Crystal system	Monoclinic		
Space group	$P2_1$		
Unit cell dimensions	$a = 12.8350(6)$ Å	$\alpha = 90^\circ$.	
	$b = 5.6272(3)$ Å	$\beta = 113.133(2)^\circ$.	
	$c = 12.9187(6)$ Å	$\gamma = 90^\circ$.	
Volume	$858.03(7)$ Å ³		
Z	2		
Density (calculated)	1.252 Mg/m ³		
Absorption coefficient	0.201 mm ⁻¹		
F(000)	348		
Crystal size	0.5 x 0.12 x 0.12 mm ³		
Theta range for data collection	1.714 to 31.552°.		
Index ranges	-18<=h<=18, -8<=k<=8, -19<=l<=19		
Reflections collected	57330		
Independent reflections	5731 [R(int) = 0.0344]		
Completeness to theta = 25.242°	100.0 %		
Absorption correction	Semi-empirical from equivalents		
Max. and min. transmission	1.0000 and 0.8839		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	5731 / 1 / 200		
Goodness-of-fit on F ²	1.073		
Final R indices [I>2sigma(I)]	R1 = 0.0258, wR2 = 0.0683		
R indices (all data)	R1 = 0.0273, wR2 = 0.0695		
Absolute structure parameter	0.019(11)		
Largest diff. peak and hole	0.444 and -0.190 e/Å ⁻³		

Table S–12. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal02. $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	$U(\text{eq})$
S(1)	4726(1)	7787(1)	3347(1)	12(1)
O(1)	4294(1)	10123(2)	2942(1)	19(1)
O(2)	5535(1)	7480(2)	4482(1)	18(1)
O(3)	6358(1)	4985(2)	1135(1)	21(1)
N(1)	5344(1)	6838(2)	2534(1)	14(1)
C(1)	3537(1)	5903(2)	3176(1)	11(1)
C(2)	3067(1)	6643(2)	4055(1)	13(1)
C(3)	2154(1)	4910(2)	4061(1)	14(1)
C(4)	1833(1)	5268(3)	5067(1)	18(1)
C(5)	944(1)	3484(3)	5081(1)	25(1)
C(6)	2671(1)	6069(2)	1991(1)	13(1)
C(7)	2415(1)	4264(2)	1261(1)	13(1)
C(8)	1574(1)	4329(2)	93(1)	12(1)
C(9)	797(1)	6196(2)	-325(1)	16(1)
C(10)	21(1)	6194(3)	-1441(1)	19(1)
C(11)	9(1)	4336(3)	-2156(1)	20(1)
C(12)	764(1)	2453(3)	-1748(1)	20(1)
C(13)	1539(1)	2453(2)	-630(1)	16(1)
C(14)	4862(1)	7376(2)	1320(1)	18(1)
C(15)	5820(1)	7255(3)	909(1)	21(1)
C(16)	6847(1)	4566(3)	2317(1)	21(1)
C(17)	5947(1)	4555(3)	2806(1)	19(1)

Table S–13. Bond lengths [\AA] and angles [$^\circ$] for crystal02.

S(1)-O(1)	1.4424(10)
S(1)-O(2)	1.4360(9)
S(1)-N(1)	1.6356(11)
S(1)-C(1)	1.7985(12)
O(3)-C(15)	1.4267(18)
O(3)-C(16)	1.4241(17)
N(1)-C(14)	1.4733(16)
N(1)-C(17)	1.4694(17)
C(1)-C(2)	1.5379(17)
C(1)-C(6)	1.5001(15)
C(2)-C(3)	1.5265(17)
C(3)-C(4)	1.5241(18)
C(4)-C(5)	1.525(2)
C(6)-C(7)	1.3364(17)
C(7)-C(8)	1.4711(16)
C(8)-C(9)	1.4021(17)
C(8)-C(13)	1.3993(17)
C(9)-C(10)	1.3937(16)
C(10)-C(11)	1.391(2)
C(11)-C(12)	1.393(2)
C(12)-C(13)	1.3954(17)
C(14)-C(15)	1.5194(19)
C(16)-C(17)	1.5189(19)
O(1)-S(1)-N(1)	106.16(6)
O(1)-S(1)-C(1)	107.85(6)
O(2)-S(1)-O(1)	120.05(6)
O(2)-S(1)-N(1)	106.10(6)
O(2)-S(1)-C(1)	107.05(6)
N(1)-S(1)-C(1)	109.34(6)
C(16)-O(3)-C(15)	109.98(11)
C(14)-N(1)-S(1)	120.64(9)
C(17)-N(1)-S(1)	118.32(9)
C(17)-N(1)-C(14)	113.32(11)
C(2)-C(1)-S(1)	108.03(8)

C(6)-C(1)-S(1)	109.94(8)
C(6)-C(1)-C(2)	112.83(10)
C(3)-C(2)-C(1)	110.77(10)
C(4)-C(3)-C(2)	112.63(10)
C(3)-C(4)-C(5)	112.19(12)
C(7)-C(6)-C(1)	123.45(11)
C(6)-C(7)-C(8)	125.69(11)
C(9)-C(8)-C(7)	122.55(11)
C(13)-C(8)-C(7)	118.95(11)
C(13)-C(8)-C(9)	118.50(11)
C(10)-C(9)-C(8)	120.60(12)
C(11)-C(10)-C(9)	120.30(13)
C(10)-C(11)-C(12)	119.73(12)
C(11)-C(12)-C(13)	119.96(12)
C(12)-C(13)-C(8)	120.90(12)
N(1)-C(14)-C(15)	107.67(11)
O(3)-C(15)-C(14)	111.21(11)
O(3)-C(16)-C(17)	111.08(11)
N(1)-C(17)-C(16)	108.16(11)

Table S–14. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal02. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U11	U22	U33	U23	U13	U12
S(1)	12(1)	11(1)	11(1)	-1(1)	4(1)	-3(1)
O(1)	24(1)	11(1)	23(1)	0(1)	11(1)	-2(1)
O(2)	15(1)	26(1)	12(1)	-3(1)	2(1)	-7(1)
O(3)	22(1)	22(1)	21(1)	0(1)	12(1)	1(1)
N(1)	16(1)	16(1)	13(1)	3(1)	7(1)	2(1)
C(1)	10(1)	10(1)	11(1)	0(1)	3(1)	-1(1)
C(2)	12(1)	12(1)	13(1)	-1(1)	5(1)	0(1)
C(3)	11(1)	16(1)	14(1)	1(1)	4(1)	0(1)
C(4)	18(1)	20(1)	19(1)	-2(1)	10(1)	-1(1)
C(5)	22(1)	29(1)	28(1)	2(1)	15(1)	-4(1)
C(6)	11(1)	13(1)	12(1)	1(1)	2(1)	0(1)
C(7)	11(1)	14(1)	12(1)	0(1)	2(1)	0(1)
C(8)	11(1)	14(1)	11(1)	-2(1)	3(1)	-2(1)
C(9)	15(1)	16(1)	14(1)	-1(1)	3(1)	1(1)
C(10)	15(1)	21(1)	16(1)	2(1)	2(1)	2(1)
C(11)	16(1)	29(1)	12(1)	-2(1)	2(1)	-4(1)
C(12)	18(1)	26(1)	15(1)	-7(1)	6(1)	-3(1)
C(13)	13(1)	18(1)	17(1)	-4(1)	5(1)	0(1)
C(14)	18(1)	23(1)	13(1)	4(1)	7(1)	3(1)
C(15)	24(1)	23(1)	19(1)	4(1)	12(1)	1(1)
C(16)	18(1)	24(1)	23(1)	4(1)	10(1)	4(1)
C(17)	21(1)	16(1)	22(1)	7(1)	12(1)	4(1)

Table S–15. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for crystal02.

	x	y	z	U(eq)
H(1)	3809	4223	3327	13
H(2A)	3691	6680	4811	15
H(2B)	2742	8262	3881	15
H(3A)	1471	5115	3359	16
H(3B)	2431	3264	4072	16
H(4A)	1536	6898	5045	22
H(4B)	2520	5104	5770	22
H(5A)	764	3776	5741	37
H(5B)	1241	1868	5118	37
H(5C)	257	3664	4395	37
H(6)	2285	7535	1747	16
H(7)	2810	2813	1519	15
H(9)	800	7474	156	19
H(10)	-503	7467	-1715	23
H(11)	-513	4352	-2920	24
H(12)	752	1170	-2231	24
H(13)	2051	1161	-356	19
H(14A)	4520	8982	1187	22
H(14B)	4266	6206	911	22
H(15A)	5514	7563	89	25
H(15B)	6385	8503	1288	25
H(16A)	7412	5819	2689	25
H(16B)	7245	3017	2467	25
H(17A)	5409	3231	2480	23
H(17B)	6304	4338	3632	23

VI. ^1H NMR Spectra

JC9121A C0C13

exp20 PROTON

SAMPLE	DATE	2012	SAT MODE	PRESATURATION
SOLVENT	CDCl ₃	WET	SPECIAL	n
VNMRSYS/DATA/JC012~	1A-CDC13/PROTON01	TEMP	NOT USED	n
ACQUISITION	8000.0	GAIN	NOT USED	n
SW	2.500	SPIN	20	0.008
AT	pw90	HST	9.700	9.700
NP	40000	ALFA	6.600	6.600
FB	not used	FLAGS		
D1	32	i1	n	
DT	1.000	in	n	
NT	16	dp	y	
CT	16	hs	nn	
TRANSMITTER		PROCESSING		
TN	H1	LB	0.20	
STRFQ	499.708	FN	NOT USED	
TOF	499.7	DISPLAY		
TPWR	61	SP	-0.1	
PW	4.850	WP	4996.8	
DECOPPLER	C13	RFL	4630.1	
DN	0	RFP	3627.9	
D0F	0	RP	3.9	
DM	NNN	1P	5.2	
DECPWAVE	35	PLOT		
DPPWR	32258	WC	250	
DMFW	VS	SC	0	
A1	TH	49	54	

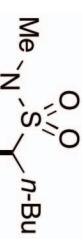
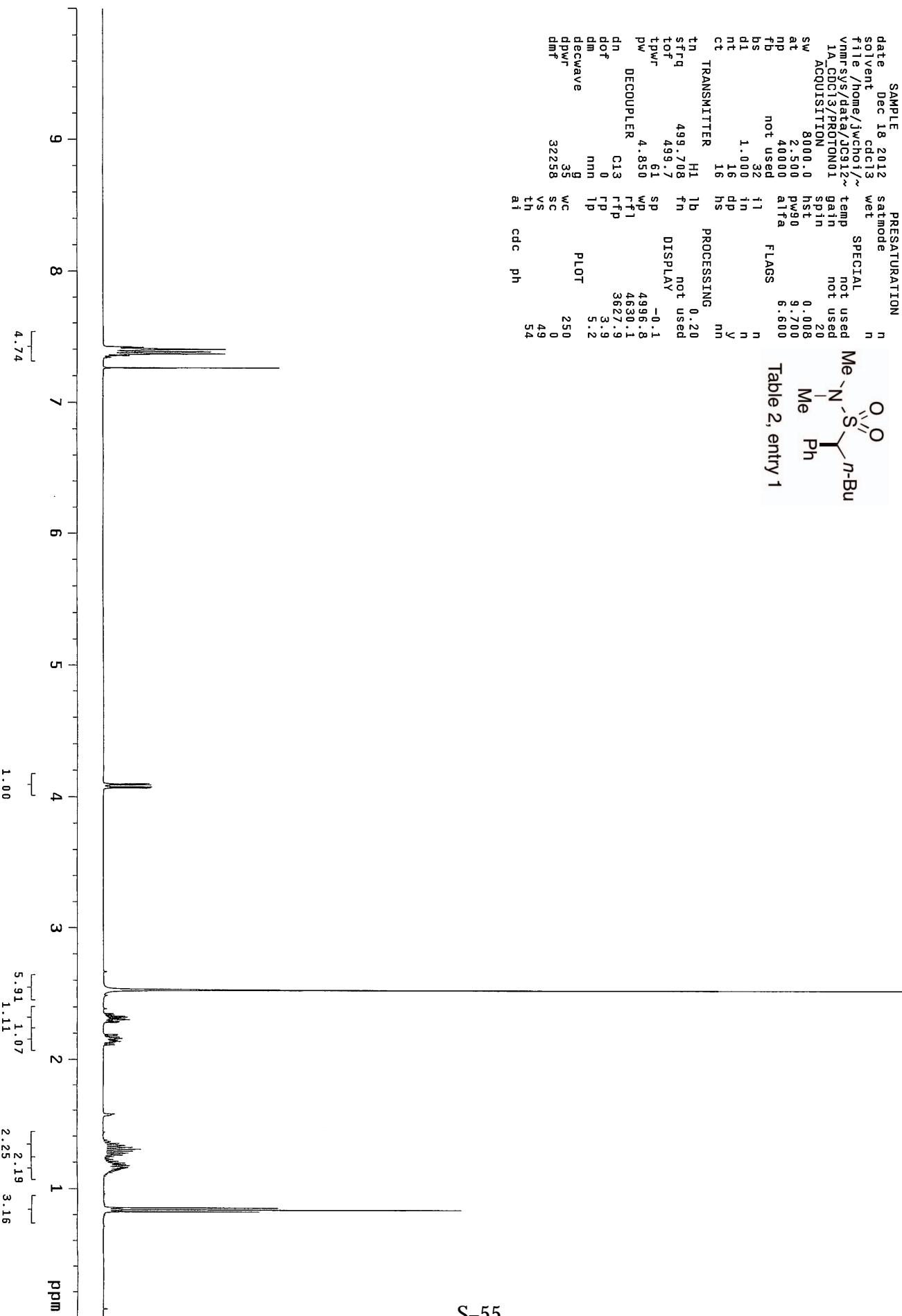


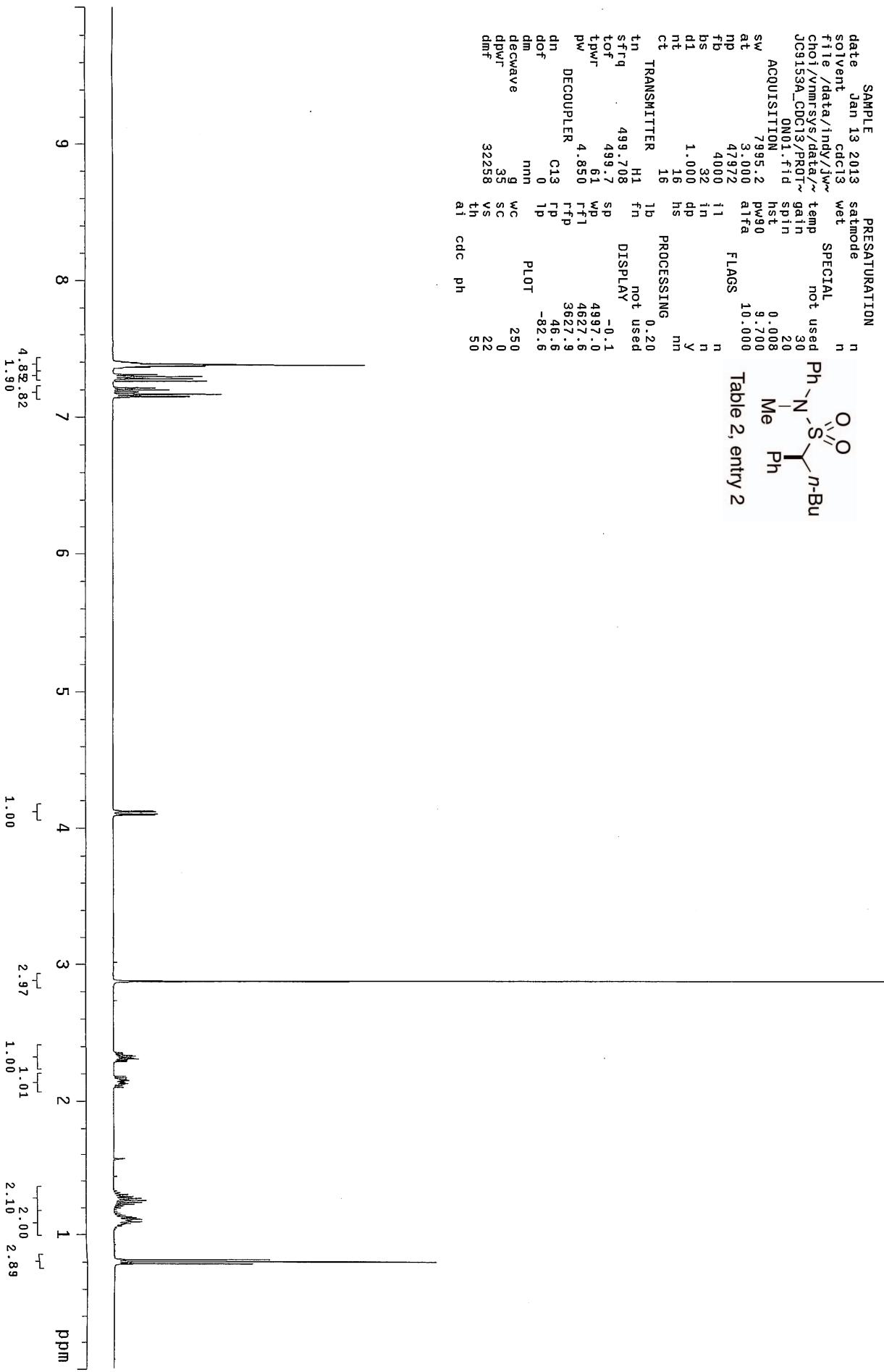
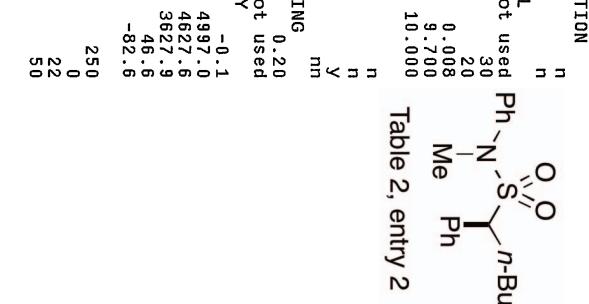
Table 2, entry 1



JC9153A CDC13

exp23 PROTON

date	SAMPLE	PRESATURATION
Jan 13 2013	c13	satmode
solvent	/data/indy/jwv	wet
f1le	/data/indy/jwv	SPECIAL
choi/vnmrjsys/data/	temp	not used
JC9153A_CDC13.PROT~	ON01.fid	gain
sw	3.000	30
at	7.995.2	20
np	47.972	0
fb	4.000	0.008
bs	32	9.700
d1	1.000	hs
nt	16	a1fa
ct	16	10.000
TRANSMITTER		FLAGS
t1n	H1	i1
sfi-q	499.708	n
tof	499.7	in
tpwr	61	n
pw	4.850	dp
DECOUPLER	C13	y
dn	r1p	nn
dof	0	nn
dm	nnn	
dewave	g	
dprf	35	
dmtf	32258	
	vs	
	th	
	ai	
	cdc	
	ph	



JC9149B 1H CDCl₃

exp23 PROTON

	SAMPLE	PRESATURATION
date	Jan 13 2013	satmode
solvent	ccl3	wet
f1le	/data/indy/jw~	SPECIAL
choi/vnmrsys/data/~	temp	not used
JC9149B.1H_COC13/P~	gain	30
ROTOND4.fid	spin	20
ACQUISITION	hst	0.008
sw	pwg0	9.700
at	7995.2	10.000
np	3.000	a1fa
fb	47972	47972
bs	4000	i1
d1	32	i1
nt	1.000	dp
ct	16	hs
TRANSMITTER	16	PROCESSING
tn	1b	0.20
sfrq	499.708	fn
tof	499.7	DISPLAY
tpwr	499.7	-0.1
pw	4.850	sp
DECOUPLER	4.850	wp
dn	rfp	4627.8
dof	C13	3677.9
dm	0	433.5
decwave	nnn	-78.8
dpwr	35	PLOT
dmpf	32258	250
	vs	0
	th	116
	ai	32
	cdc	ppm
	ph	

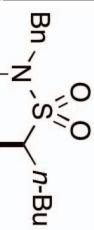
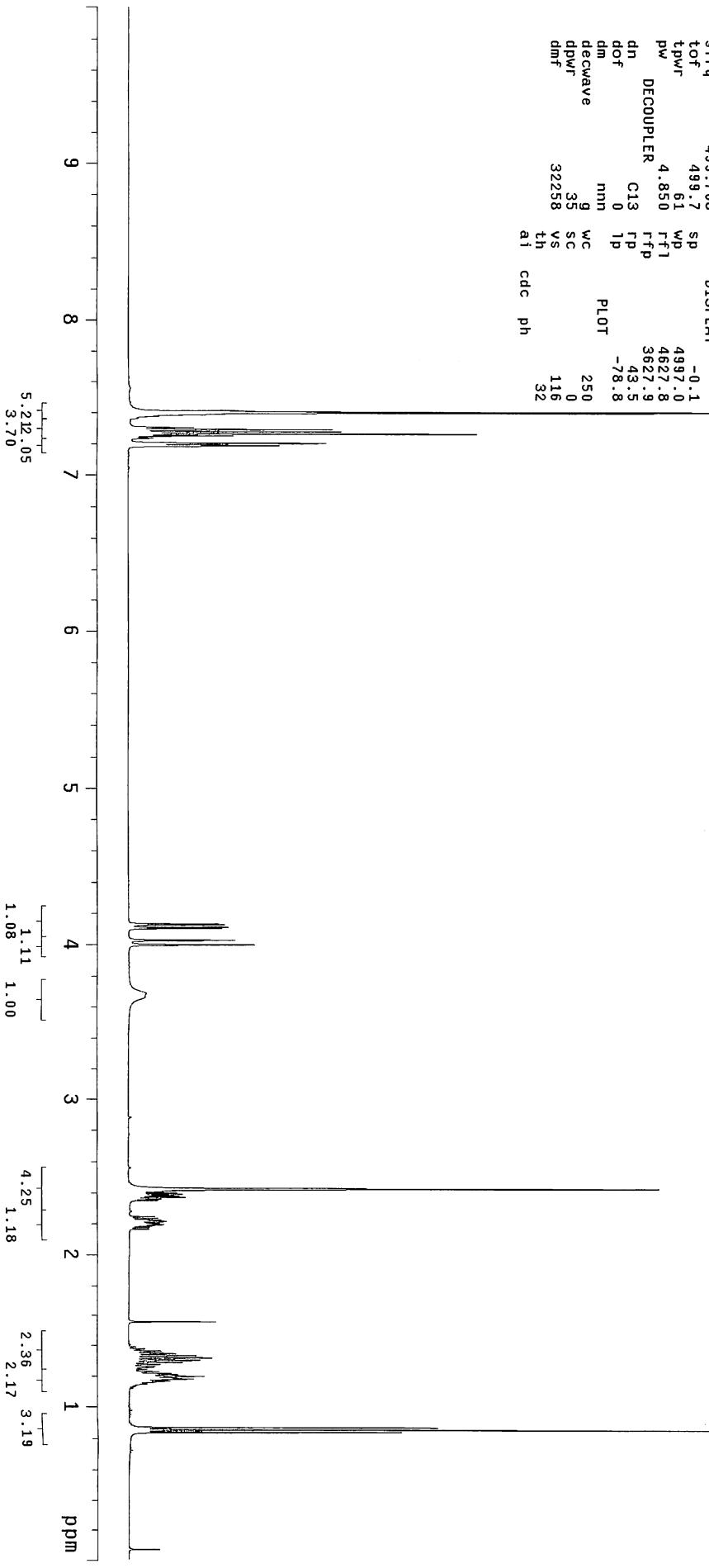


Table 2, entry 3



JC9147A 1H CDCl₃

exp23 PROTON

SAMPLE	3	2013	PRESATURATION
date	Jan	solvent	satmode
file	/data/indy/JW~	cdcl ₃	wet
choi/vnmrsys/data/~		SPECIAL	n
JC9147A	1H	CDCl ₃ /P~	not used
ROTOND1.fid		gain	46
ACQUISITION		spin	2.0
sw	7995.2	hst	0.008
at	3.000	pw90	2.0
np	47972	alfa	9.700
fb	4000	FLAGS	10.000
bs	32	i1	n
d1	1.000	in	n
nt	16	dp	y
ct	16	hs	mn

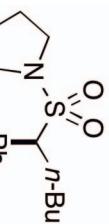
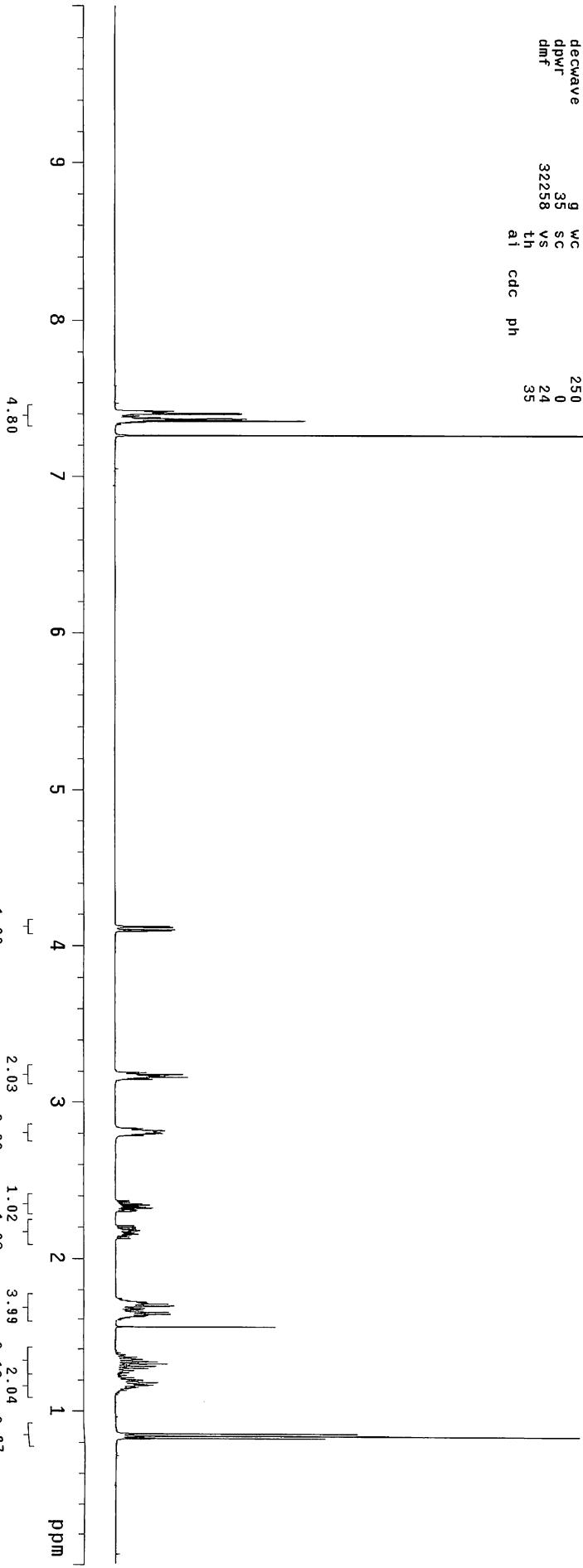


Table 2, entry 4

TRANSMITTER	1b	PROCESSING	0.20
tn	H1	fn	not used
sfrq	499.708	DISPLAY	-0.1
tof	499.7	sp	499.7.0
tpwr	61	wp	4622.6
pw	4.850	rff1	3622.9
DECOUPLER		rfp	47.0
dn	C13	rp	-86.6
dof	0	1p	250
din	nnn	PLOT	24
decwave	g	wc	35
dpower	35	sc	
dinf	32258	vs	
th		th	
ai		cdc	
		ph	



JC9155 1H CDC13

exp23 PROTON

SAMPLE Jan 17 2013 satmode n
solvent cdc13 wet n
file /data/indy/jw~ SPECIAL n
choi/vnmrsys/data/~ temp not used
JC9155_1H_CDC13/PR~ gain 44
OTON01.fid spin 20
ACQUISITION 7995.2 pw90 0.008
sw 3.000 aifa 9.700
at 47972 hst 10.000

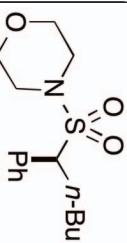
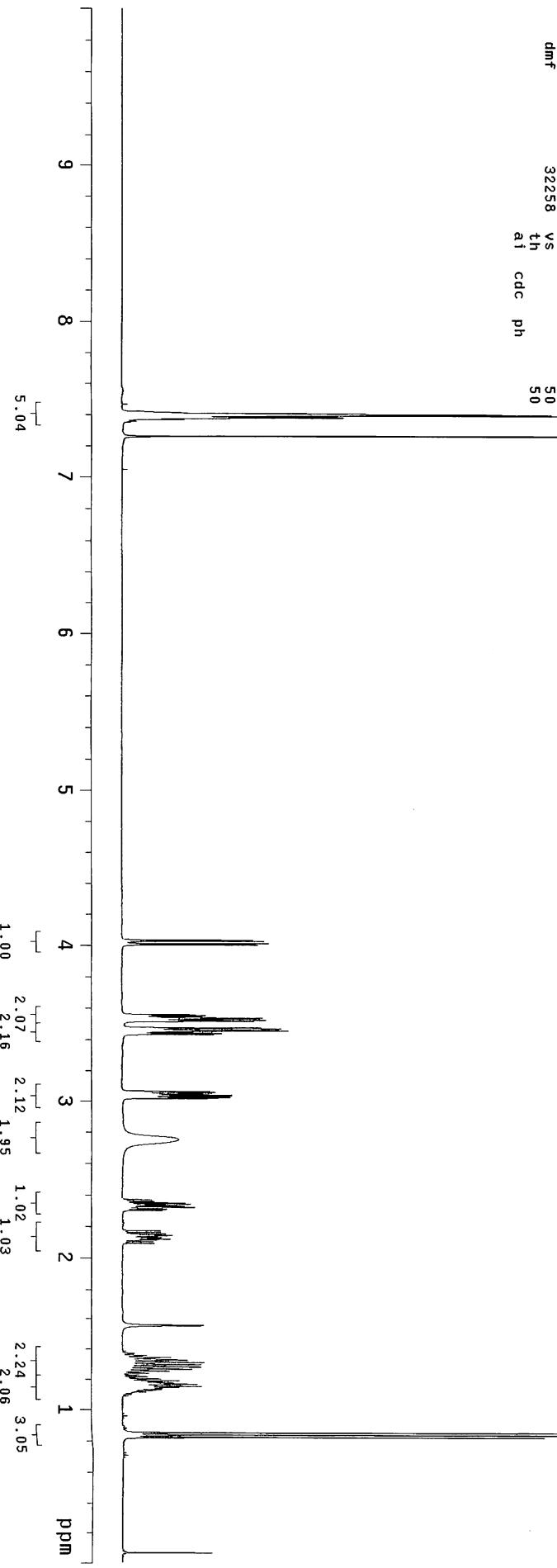


Table 2, entry 5

PRESATURATION n
tn 1b not used 0.20
sfrq 439.708 fn
tfrq 439.7 sp DISPLAY -0.1
tof 499.7 wp 4997.0
tpwr 61 rrf1 4622.8
pw 4.850 rfp 3627.9
DECOUPLER C13 rp 47.0
dn 0 tp PLOT -86.3
dof nnn
dm g wc 250
decwave sc 0
dpwr 35 vs 50
dnf 32258 th 50
ai cdc ph



exp24 PROTON

SAMPLE	May 14 2013	PRESATURATION	n
solvent	cdcl3	satmode	n
file	/indy/jwchoi/	wet	n
vnmrsys	/data/JJC100~	SPECIAL	n
93B_1H_CDC13/PROTO~	N01.fid	not used	n
ACQUISITION	gain	32	20
sw	temp	0	0.08
8000.0	hst	9	9.900
at	pw90	9	9.900
3.000	alra	10.000	
pp	FLAGS		
fb	i1	n	
bs	in	n	
d1	dp	y	
nt	16	hs	nn
ct	16	PROCESSING	0.20
tn	H1	fn	not used
sfrq	499.708	DISPLAY	-0.1
tof	499.7	sp	4996.8
tpwr	61	wp	4930.1
pw	4.950	r _f 1	3927.9
DECOUPLER	C13	r _f p	6.8
dn	0	r _p	-79.6
dof	0	1p	
dm	nnn	PLOT	250
decwave	w40_autox7~	wc	
dppr	991	sc	0
dmtf	41	vs	65
	32258	th	13
		ai	
		cdc	
		ph	

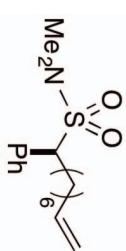
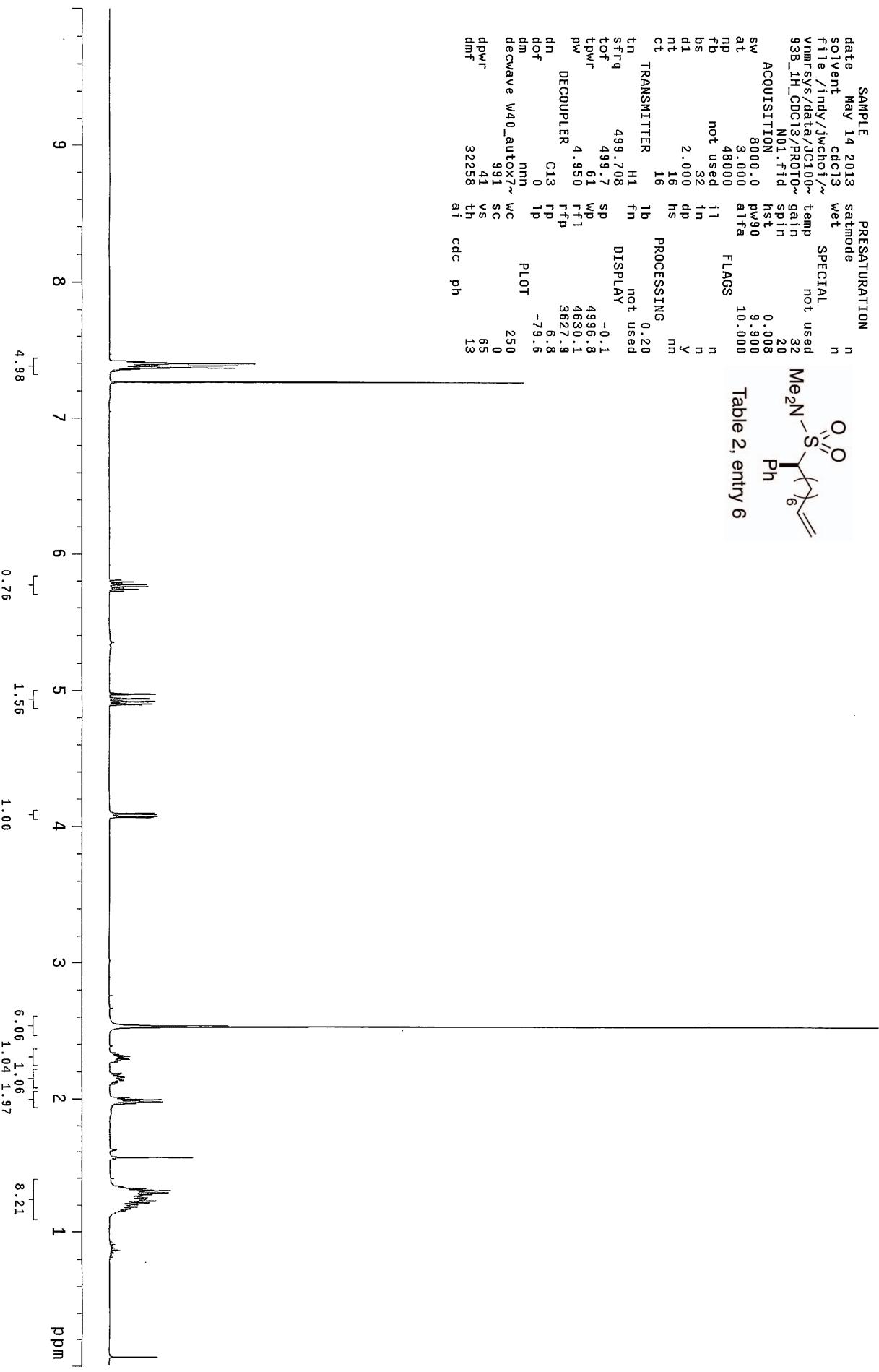


Table 2, entry 6



PM2-21A 1H CDC13

exp30 PROTON

SAMPLE	21	2013	PRESATURATION	n
date	Oct	cd13	satmode	wet
solvent			SPECIAL	n
file	/indv/jwchoi/	~	not used	
vnmrssr	/data/PM2-2-	temp	26	2.0
1A_1H_CDC13/PROTON~	01.fid	spin	0.008	
ACQUISITION		hst	9	9.00
sw	8000.0	pw90	10.000	
rt	3.000	alfa		
np	48000	FLAGS		
fb	not used	i1	n	
bs	32	in	n	
d1	2.000	dp	y	
nt	16	hs	nn	
ct	16			
TRANSMITTER		PROCESSING	0.20	
tn	H1	1b	not used	
sfrq	499.698	fn		
tof	499.7	DISPLAY	-499.9	
tpwr	61	sp	5496.6	
pw	4.950	wp	4632.1	
DECOUPLER		r _{f1}	3827.8	
dn	C13	r _{f2}	-87.1	
dof	0	1p	-73.6	
din	non	PLOT		
decwave	w40_autox7~	wc	250	
dppr	991	sc	0	
dmpf	41	vs	16	
	32258	th	25	
		ai	cddc	ph

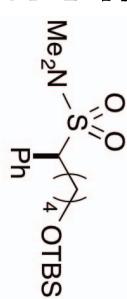
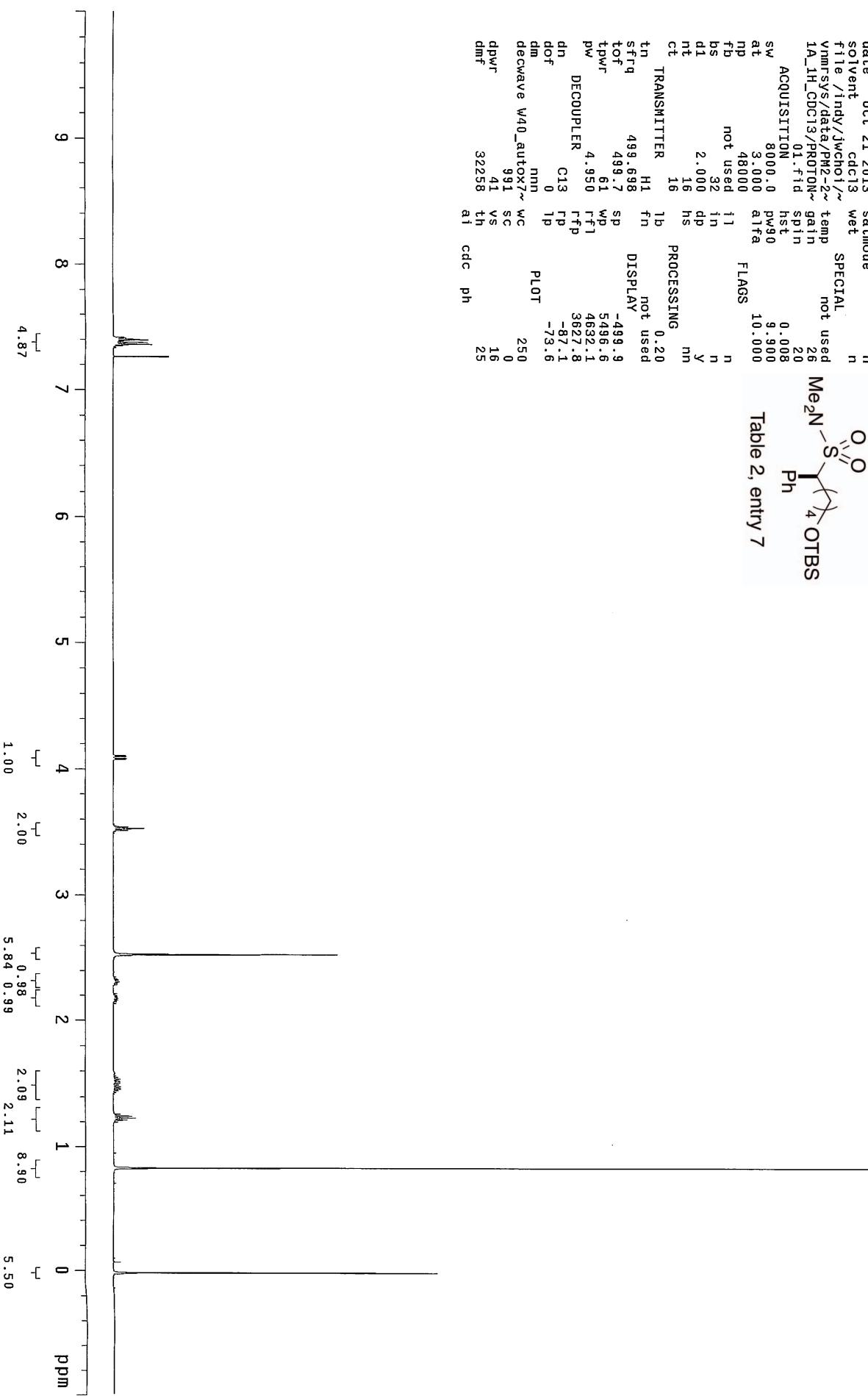


Table 2, entry 7



JC9221B 1H CDCl3

exp23 PROTON

date	SAMPLE	PRESATURATION
Feb 22	CDCl3	satnmode
solvent		n
file	/data/indy/jw~	
choi/vnmrsys/data/~	temp	SPECIAL
JC9221B_1H_CDC13/P~	gain	not used
ROTNO1.fid	spin	30
ACQUISITION	hst	20
sw	pw90	0.008
at	8000.0	9.700
np	3.000	10.000
fb	a1fa	
bs		
d1		
rt		
ct		
TRANSMITTER	32	FLAGS
tn	1b	0
sfrq	H1	20
	fn	not used
tof	499.708	
tpwr	499.7	DISPLAY
pw	61	-0.1
rf1	4.850	
DECOUPLER	rfp	
dn	C13	496.8
dof	rp	4630.1
dm	0	3627.9
decwave	tp	46.0
dpwr	nnn	-72.7
dinf	g	
	wc	
	sc	
	vs	
	th	
	ai	
	cdc	
	ph	

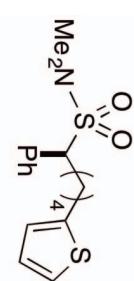
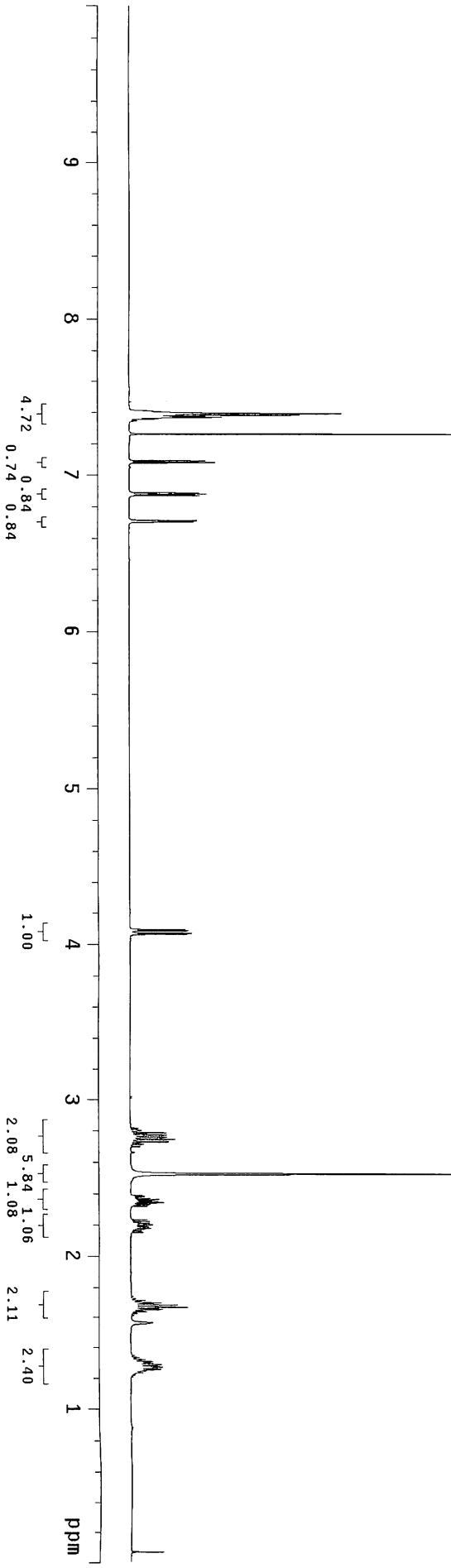


Table 2, entry 8



JC12043A CDC13

exp57 PROTON

SAMPLE date Jun 18 2014 solvent cdcl₃ PRESATURATION satmod e n
 file /indy/heinisc~ h/vnmr/sys/data/c1~ wet n
 2043H_CDC13/PROTON~ temp not used n
 01.fid spin 0.008 20
 ACQUISITION gain 9.900 25
 sw 8000.0 pw90 spin 9.900 20
 at 3.000 alfa 10.000
 np 48000
 fb not used i1 1 20
 bs 32 in n
 d1 2.000 dp in y
 nt 16 hs nn
 ct 16
 TRANSMITTER tn H1 f1 0.20
 tn 499.689 not used
 sfrq 499.7 DISPLAY -0.2
 tof 499.7 sp 4996.8
 tpowr 61 wp 4635.7
 pw 4.350 rfp 3627.7
 DECOUPLER C13 rfp -166.7
 dh C13 rfp -78.7
 dof 0 1p
 dm mn PLOT 250
 decwave w40_autox7~ wc 0
 g91 sc 64
 dpwr 41 vs 25
 dmF 32258 th ai
 ai cdc ph

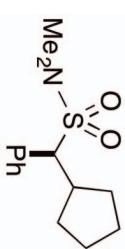
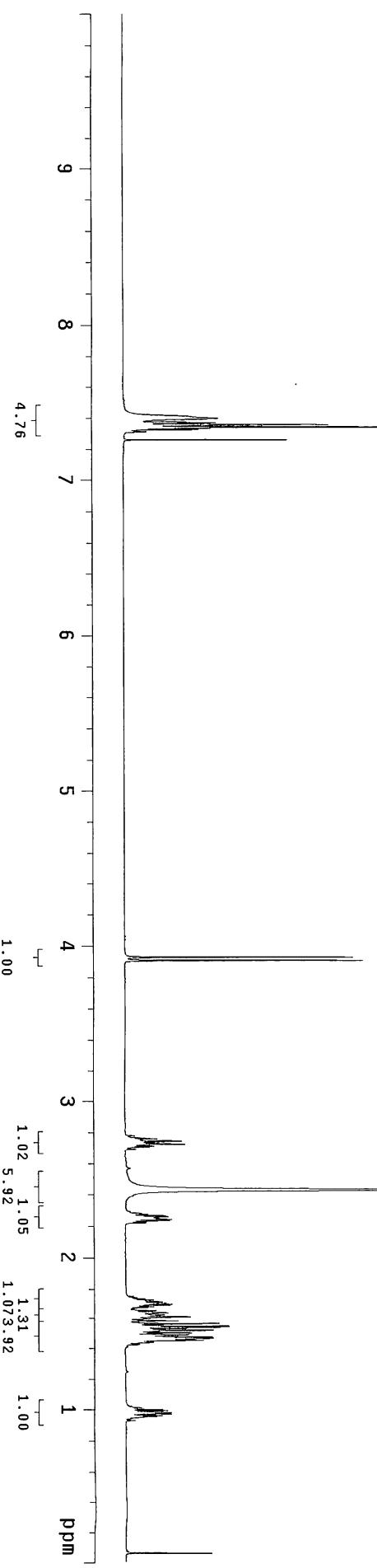


Table 2, entry 9



JC911B 1H CDCl₃

exp23 PROTON

date	SAMPLE	PRESATURATION
Dec 9 2012	saturnode	n
solvent	cdcl ₃	wet
file /data/indy/jmr/choi/vnmrsys/data/~	SPECIAL	n
JC911B 1H CDCl ₃ .P~	gain	20
ROTON3.fid	not used	0.008
ACQUISITION	spin	9.700
sw	hst	6.600
at	pw90	9.700
np	alpha	6.600
fb	not used	0.20
bs	i1	n
d1	in	n
nt	1.000	y
ct	dp	nn
TRANSMITTER	hs	nn
tn	1b	0.20
sfrq	H1	fn
tn	499.708	not used
tfrq	499.7	DISPLAY
tof	499.7	-0.1
tpwr	61	wp
pw	4.850	rff1
DECOUPLER	C13	rfp
dn	0	rp
dof	0	1p
dif	mn	PLOT
decwave	g	250
dpwr	35	sc
dif	32.258	vs
	th	37
	ai	54
	cdc	1.00
	ph	3.10

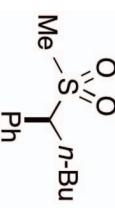
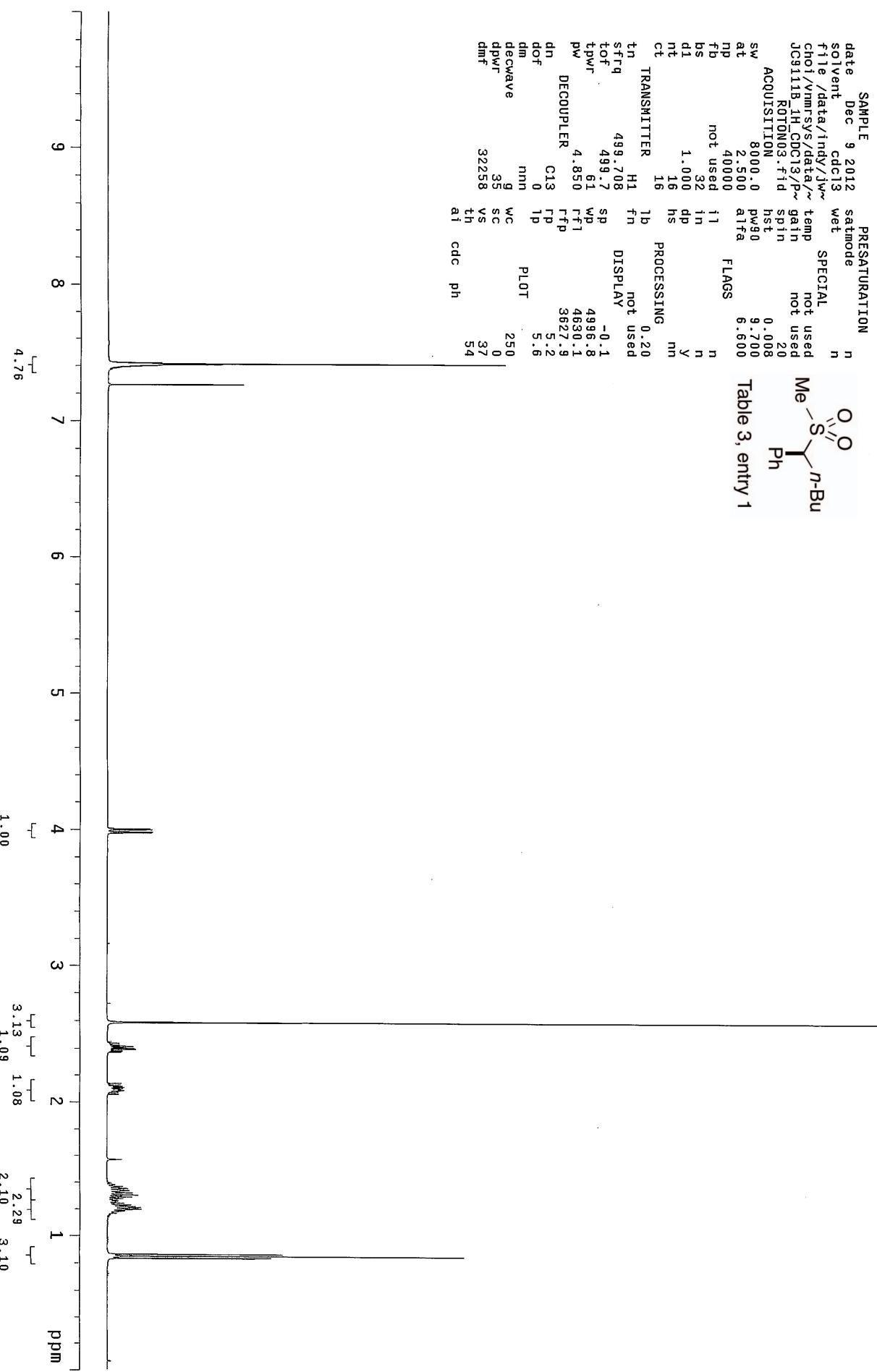


Table 3, entry 1



JC10185 CDC13

exp23 PROTON

SAMPLE Jul 19 2013 satmode
solvent cdc13 wet
file /jndy/jwchoi/~/
vnmrsys/data/JC101/~
85_CDCl3/PROTON01.
fid gain 32
temp 20
SPECIAL n
not used
ACQUISITION spin 32
8000.0 0.008
pw0 9.900
at 3.000 10.000
np 48000 atra
fb 2.000 dpp
bs 32 i1
d1 16 in
nt 16 hs
ct 16 PROCESSING mn
TRANSMITTER 0.20
tn 1b fn not used
strq 499.698 DISPLAY -0.1
tof 499.7 sp 4996.8
tpwr 61 wp 1001.8
pw 4.950 rfp -76.4
DECOUPLER C13 rfp 0
dn 0 tp -76.7
dof PLOT
dm nnn
decwave w40_autoX7~ wc 250
dpwr 991 sc 0
dimf 41 vs 59
32258 th 13
ai cdc ph

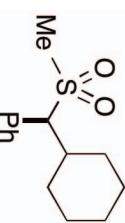
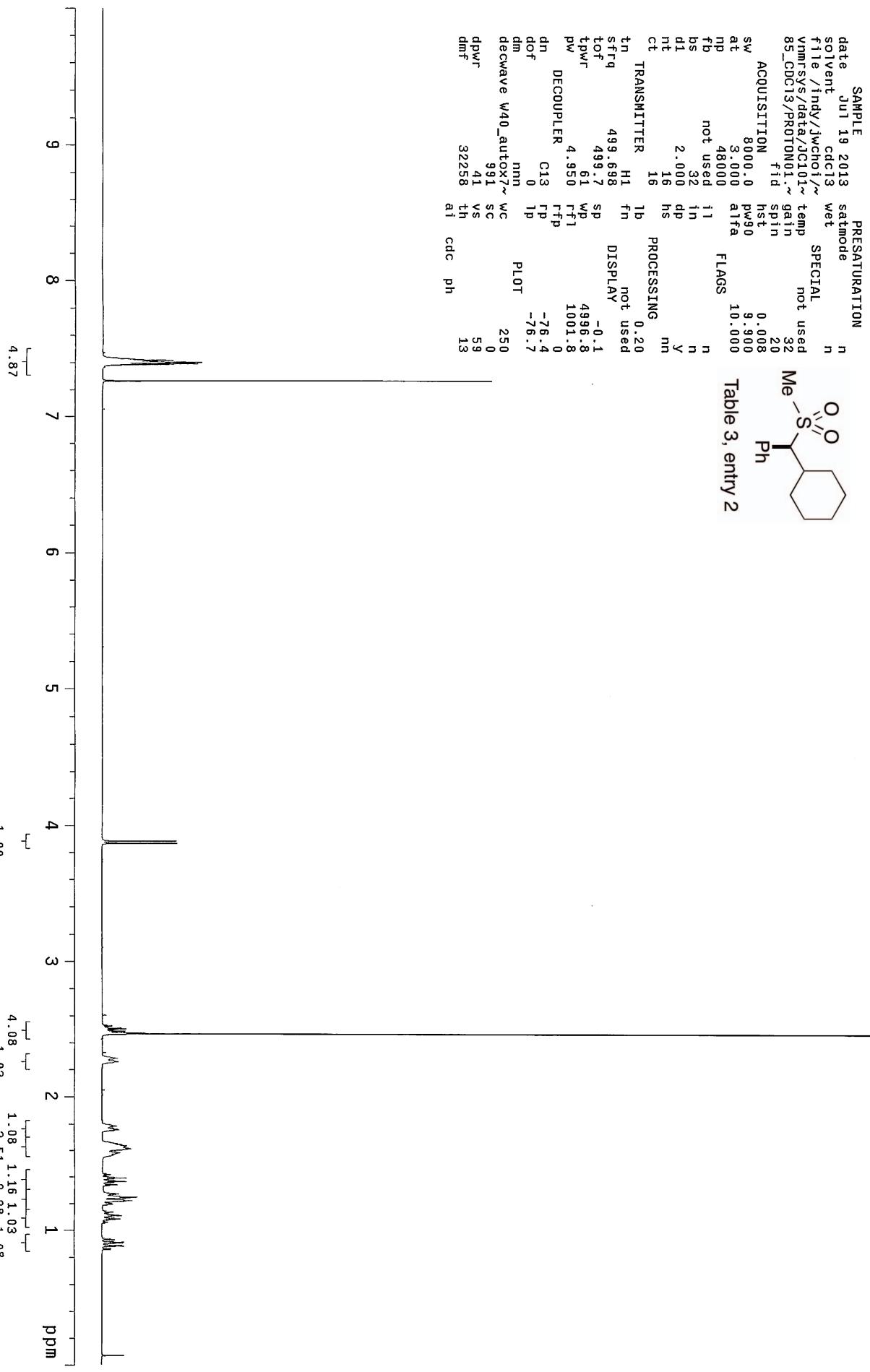


Table 3, entry 2



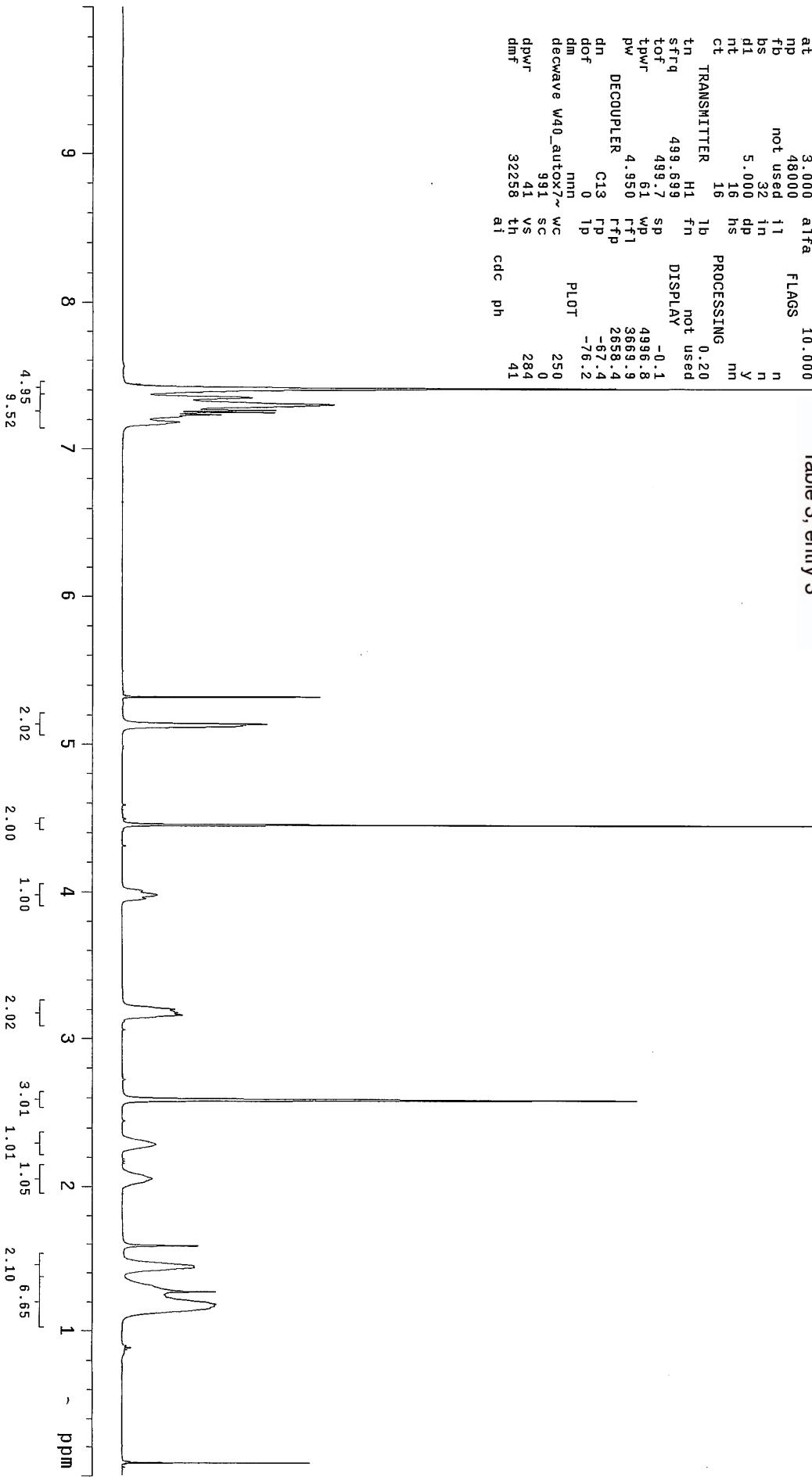
JC101B1 CD2C12

exp2 PROTON

	SAMPLE	PRESATURATION	
date	Aug 7 2013	satmodo	n
solvent	cd2c12	wet	n
file	/indy/jwchoi/~/vnmrssr/data/JC101/~/	SPECIAL	
81_CD2C12/PROTON12~		not used	
ACQUISITION	fid	spin	20
sw	800.0	hst	0.008
at	3.000	pw90	9.900
np	48000	alpha	10.000
fb	not used	i1	n
bs	32	in	n
d1	5.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	0.20
tn	H1	fn	not used
sfrq	499.699	DISPLAY	-0.1
tof	499.7	sp	4996.8
tpwr	61	wp	3669.9
pw	4.950	rfl	2658.4
DECOUPLER		rfp	-67.4
dn	C13	rp	-76.2
dof	0	1p	
d0m	nmn	PLOT	
decwave	w40_autox7~	wc	250
dppr	991	sc	0
dmpf	32258	vs	284
	41	th	41
	cdd	ph	



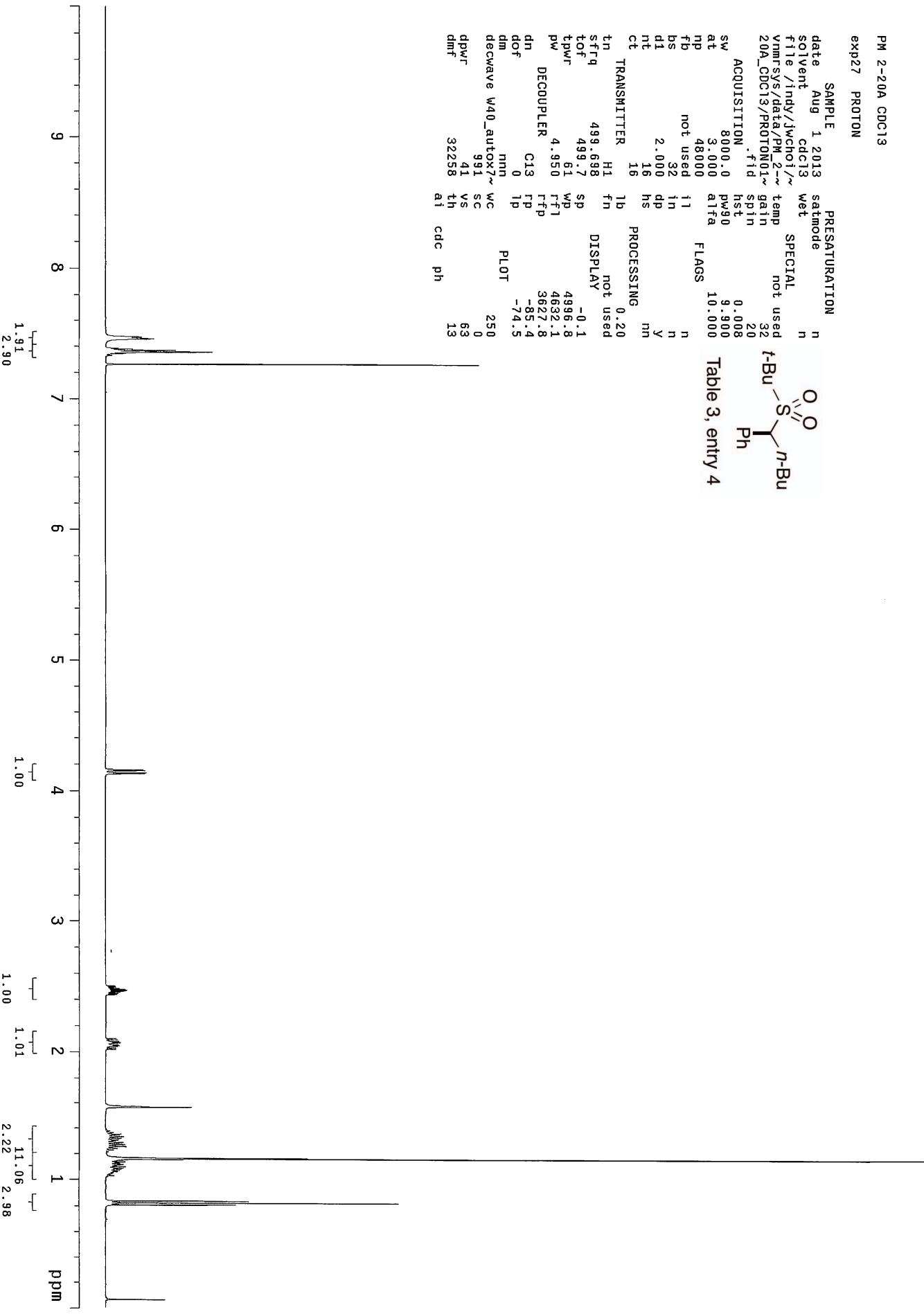
Table 3, entry 3



PM 2-20A CDC13

exp27 PROTON

SAMPLE PRESATURATION
date Aug 1 2013 satmode n
solvent cdc13 wet n
file /indy/jwchoi/~/ SPECIAL n
vnmrsys/data/PM2-~ temp not used
20A_CDCC13/PROTON01~ gain 32 t-Bu-S(=O)(=O)-n-Bu
ACQUISITION fid 20
sw 8000.0 pw90 9,900 Ph
at 3.000 a1fa 10.000 Table 3, entry 4
np 48000
fb not used i1 0.008
bs 32 in n
d1 2.000 dp y
nt 16 hs nn
ct 16
TRANSMITTER 1b PROCESSING 0.20
tn H1 fn not used
sf1q 499.698 DISPLAY -0.1
tof 499.7 sp
tpwr 6.1 wp 4996.8
pw 4.950 rfp 4633.1
DECOUPLER C13 rfp 3621.8
dn 0 rp -85.4
dof 0 tp -74.5
dm nnn PLOT 250
decwave w40_autox7~ wc
991 sc 0
dpwr 41 vs 63
dmf 32258 th 13
ai cdc ph



JC9119A CDC13

exp30 PROTON

SAMPLE	PRESATURATION
date Dec 14 2012	saturate
solvent cdc13	wet
file /data/indy/jw ^r	SPECIAL
choi/vnmrsys/data/~	
JC9119A_CDC13/PROT~	
ON01.fid	temp
ACQUISITION	gain
sw 800.0	not used
at 2.500	20
np 40000	0.008
fb	9.700
bs	6.600
d1	
nt	
ct	
TRANSMITTER	FLAGS
tn H1	i1
sfrq 499.708	in
tof 499.7	dp
tpwr 6.1	hs
pw 4.850	
DECOUPLER	PROCESSING
dn C13	0.20
dof 0	not used
dm nnn	DISPLAY
decwave g	-0.1
dpwr 35	4996.8
dmf 32258	4660.1
ai vs	3677.9
th	4.4
cac	2.1
ph	

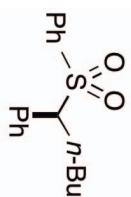
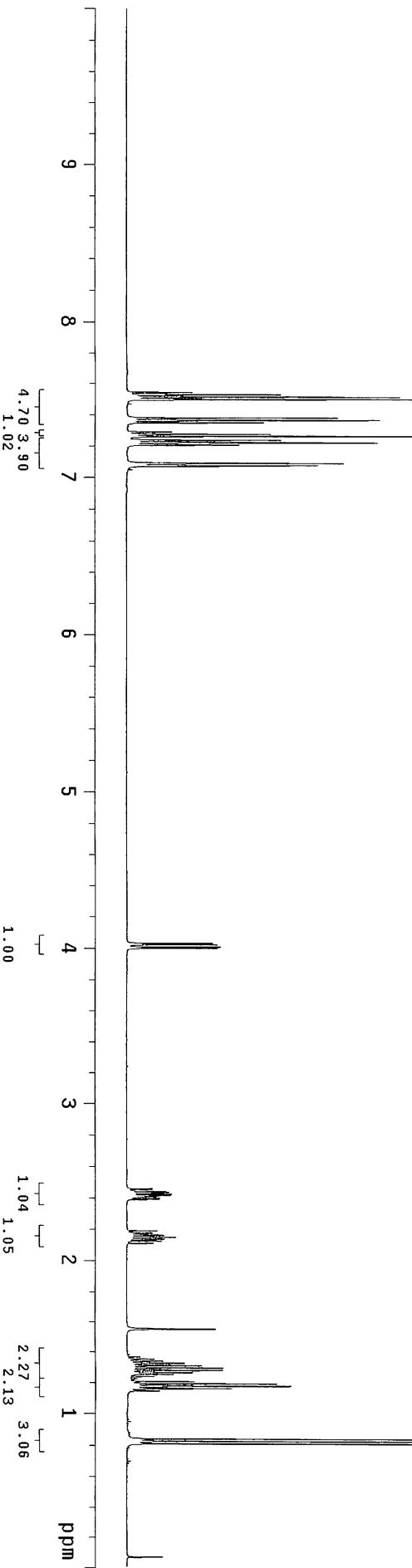


Table 3, entry 5



JC9229 CDC13

exp23 PROTON

SAMPLE	Feb 16 2013	PRESATURATION	n
solvent	ccl ₃	wet	n
file	/data/indy/jew	SPECIAL	
choi/vnmrsys/data/~/		not used	
JC9229_1H_13C_CDC1~	gain	30	
3/PROTON2.fid	spin	20	
ACQUISITION	hst	0.008	
sw	pw90	9.700	
at	3.000	aifa	10.000
np	48.000	FLAGS	
fb	not used	i1	
bs	32	in	n
d1	1.000	dp	n
nt	16	hs	y
ct	16	PROCESSING	nn
tn	H1	1b	0.20
sfrq	499.708	fn	not used
tof	499.7	DISPLAY	-0.1
tpwr	61	sp	4986.8
pw	4.850	r _{f1}	4630.1
DECOUPLER	C13	r _{fp}	3627.9
dn	0	r _p	43.7
dof	0	1p	-71.3
din	nnn	PLOT	
decwave	gg		250
dppr	sc	vs	0
dinf	35	th	45
		ai	50
		cdc	
		ph	

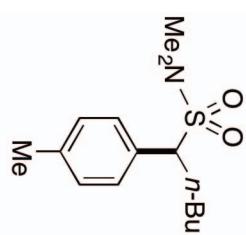
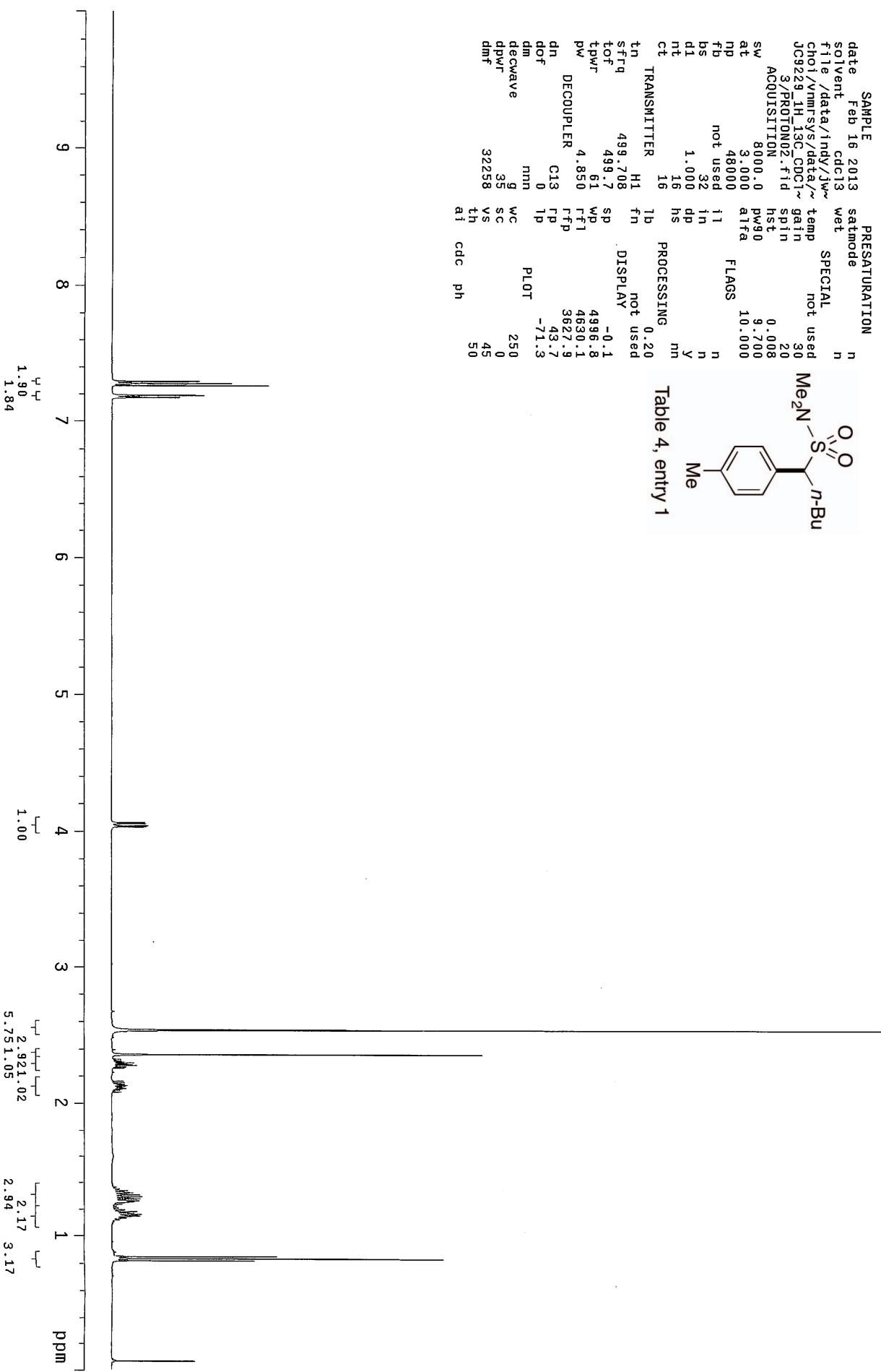
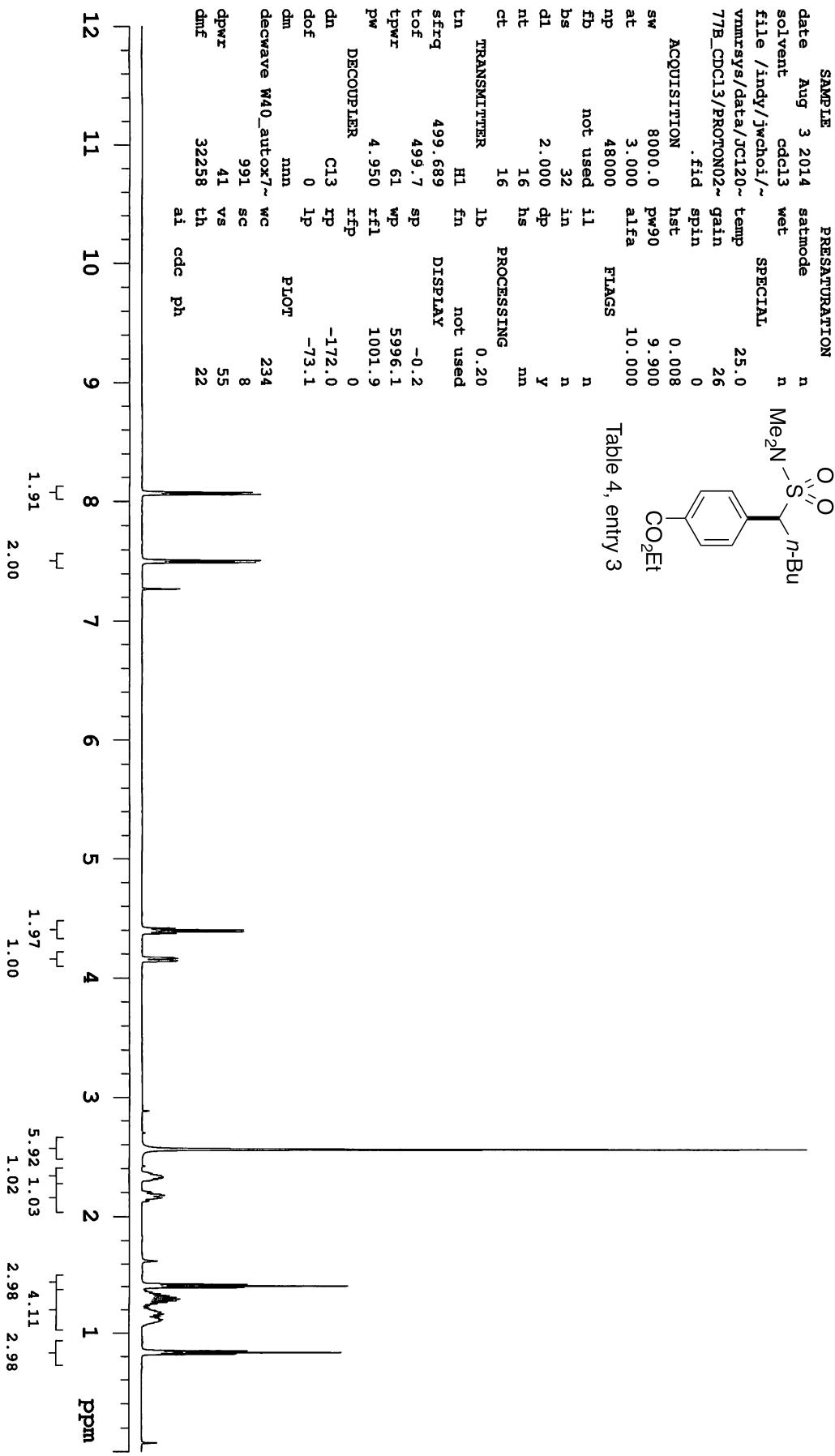


Table 4, entry 1



exp64 PROTON

SAMPLE	PRESATURATION	n	O
date	Aug 3 2014	satmode	n
solvent	cdcl3	wet	Me ₂ N-S(=O)(=O)-n-Bu
file	/indy/jchoi/-vnmrsys/data/JC120-	temp	SPECIAL
77B_CDCl ₃ /PROTON2~	gain	25.0	
ACQUISITION	.fid	spin	26
sw	8000.0	hst	0.008
at	3.000	pw90	9.900
np	48000	alfa	10.000
fb	not used	i1	n
bs	32	in	n
d1	2.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	0.20
TRANSMITTER	H1	fn	not used
sfrq	499.689	DISPLAY	
tof	499.7	sp	-0.2
tpwr	61	wp	5996.1
pw	4.950	rfl	1001.9
DECOPPLER	rfp	r0	0
dn	C13	rp	-172.0
dof	0	1p	-73.1
dm	num	PLOT	
deswave	W40_autos7~	wc	234
dpwr	991	sc	8
dmf	32258	th	55
		ai	cdc ph



JC9159B CDC13

exp23 PROTON

SAMPLE PRESATURATION
date Jan 17 2013 satmode

solvent cdc13 wet n
 file /data/inj/jw~
 choi/vminsys/data/~ temp SPECIAL not used n
 JC9159B_CDCl3/PROT~ gain 30 Me₂N-S-C(=O)-n-Bu
 ONO₁ fin sin

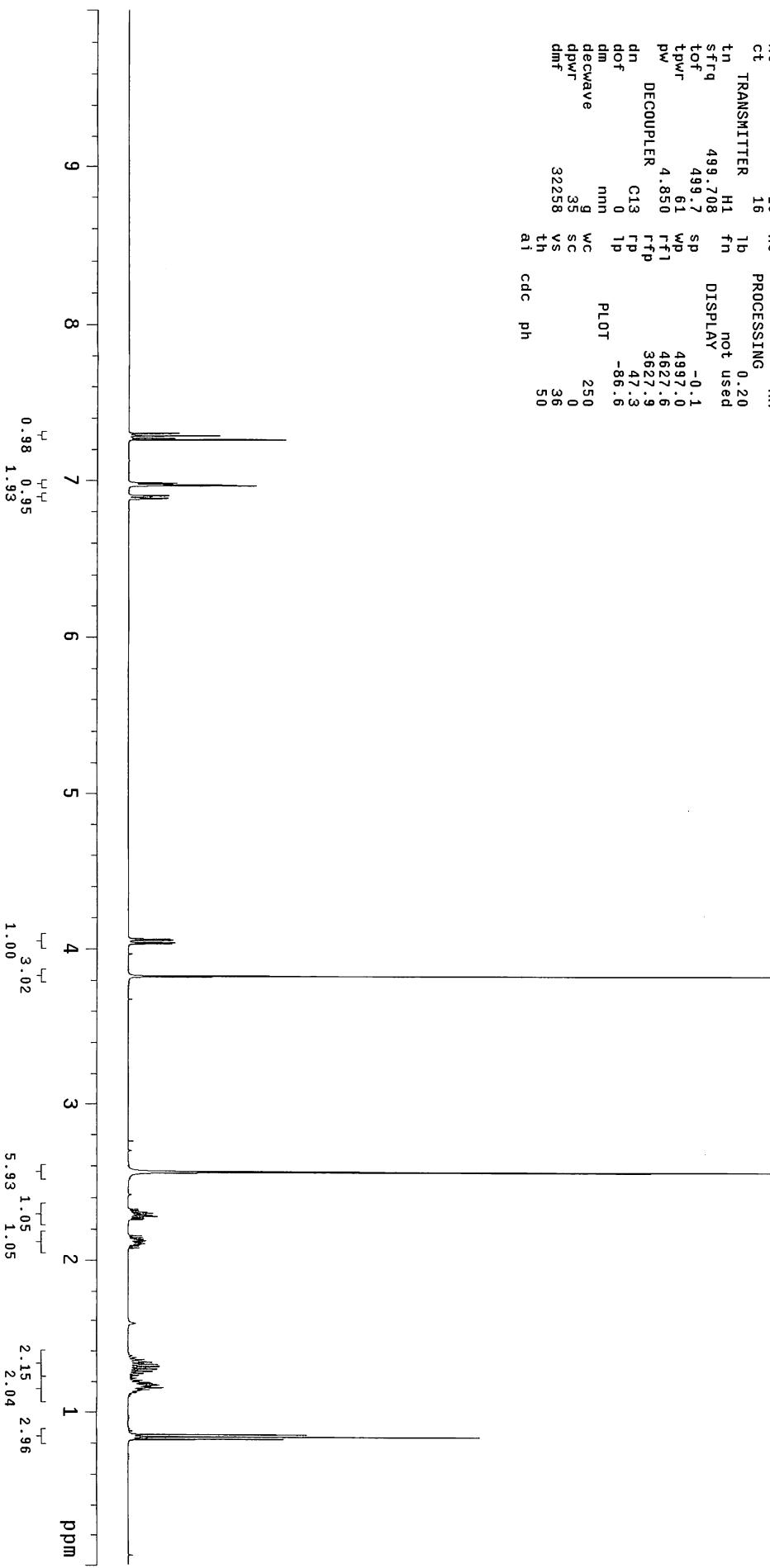
	ACQUISITION	SW	SPW	HST	0.008
	/995.2	3.000	pwg0	9.700	10.000
	47972	400	alfa	n	n
bs	32	in		y	
d1	1.000	dp		n	
nt	16	hs		n	

FLAGS

Table 4, entry 4



Table 4, entry 4



JC10223B CDC13

exp28 PROTON

SAMPLE	saturnode	PRESATURATION	
date	Aug 6 2013	n	
solvent	cde13	wet	
file	/indy/jwchoi/		
vmrsvs	data/JC102/		
23B_CDCl3/PROTON01~	temp	SPECIAL	
	gain	not used	
ACQUISITION	f1d	32	
sw	hst	2.0	
at	pw90	0	
pp	3.000	0.008	
nt	a1fa	9	
fb	48.000	9.900	
bs	not used	10.000	
d1	i1	Me ₂ N-S(=O)(=O)-n-Bu	
nt	in		
ct	16	FLAGS	
tn	hs	nn	
sfrq	499.698	DISPLAY	
tof	499.7	-0.1	
tpwr	61	496.8	
pw	4.950	4632.1	
DECOUPLER	r1f	3627.8	
dn	C13	r1p	-84.4
dof	0	1p	-75.3
din	nnn	PLOT	250
decwave	w40_autox7~	wc	0
d1pwr	991	sc	24
d1mf	32258	vs	50
	th		
	ai		
	cdc		
	ph		

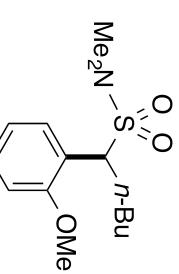
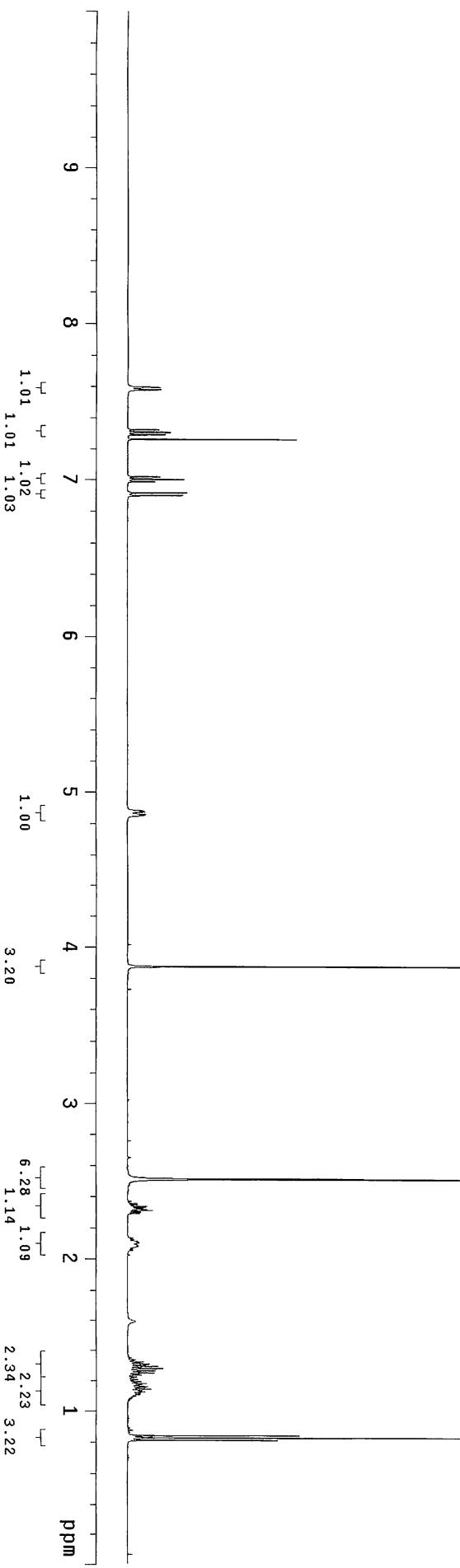


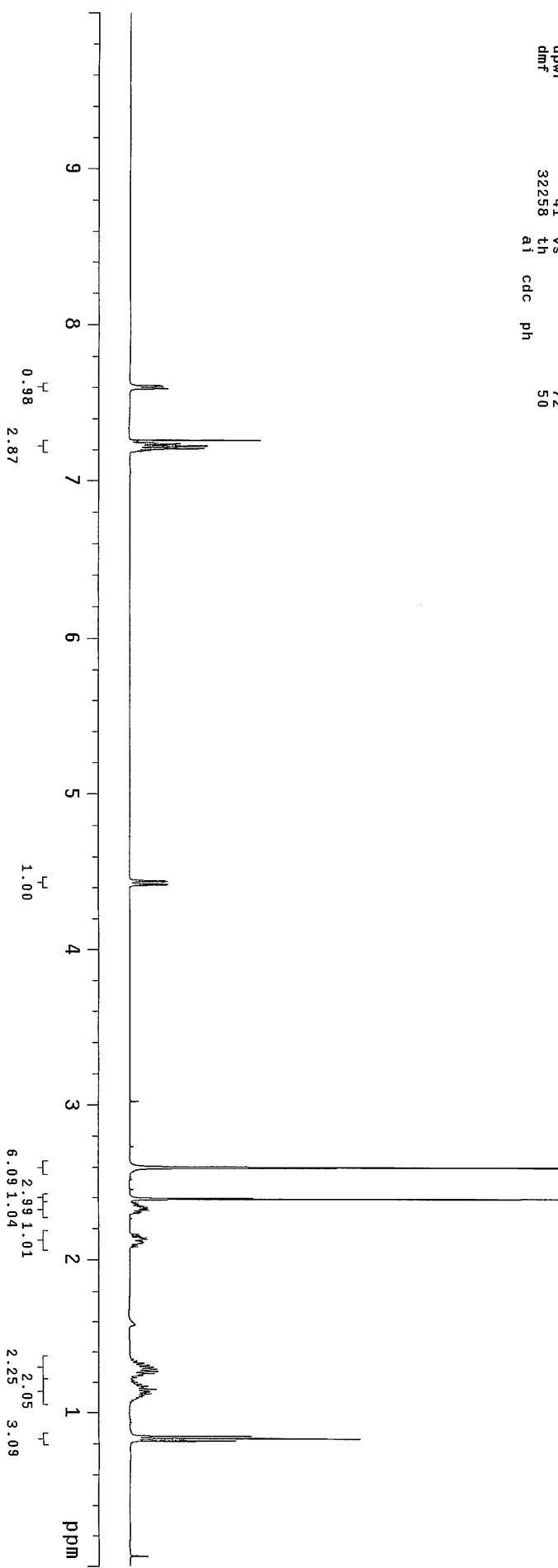
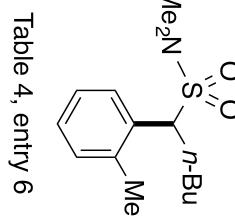
Table 4, entry 5



JC10209B CDC13

exp28 PROTON

	SAMPLE	PRESATURATION
date	Aug 9 2013	saturation
solvent	cdcl3	wet
f1e	/indy/jwchoi/~/	
vmrsys	data&JC102~	
09B_CDCl3/PROTON01~	gain	20
at	f1d	20
sw	temp	0.008
ACQUISITION	hst	20
np	pw90	9.900
d1	a1fa	10.000
nt	48.000	
ct	FLAGS	
fb	i1	n
bs	in	n
d1	2.000	y
nt	16	m
TRANSMITTER	16	PROCESSING
tn	1b	0.20
sfrq	H1	not used
499.698	fn	
tof	499.7	DISPLAY
tprt	sp	-0.1
4.950	61	4996.8
pw	wp	4632.1
DECOUPLER	rf1	3627.8
dn	rfp	-81.2
dof	C13	-81.2
din	0	-72.5
decwave	nnn	PLOT
w40_autox7~	wc	250
dpwr	991	0
dinr	sc	72
32258	41	50
ai	vs	
	th	
	cddc	
	ph	



JC11021 CD2C12

exp30 PROTON

SAMPLE	date	Oct 16 2013	PRESATURATION	n
	solvent	cd2c12	satnode	n
	file	/indy/jwchoi/~/	wet	n
Vnmrsss	21_CD2C12/PROTON01~	temp	SPECIAL	
sw	8000.0	gain	not used	
at	3.000	spin	28	
np	4800	hst	20	
fb	not used	pwg0	0	
bs	32	9	0.008	
d1	2.000	9	900	
nt	16	alfa	10.000	
ct	16	FLAGS	10	
TRANSMITTER	1b	PROCESSING	0.20	
tn	H1	fn	not used	
sfrq	499.699	DISPLAY	-0.1	
tof	499.7	sp		
tpwr	61	wp	4996.8	
pw	4.950	r _f 1	3668.9	
DECOUPLER	C13	r _f p	268.4	
dn	0	rp	-85.2	
dof	0	tp	-72.4	
dm	nnn	PLOT		
decwave	w40_autox7~	wc	250	
dpwr	991	sc	0	
dmf	32258	vs	27	
		th	50	
		ai		
		cdc		
		ph		

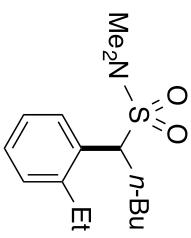
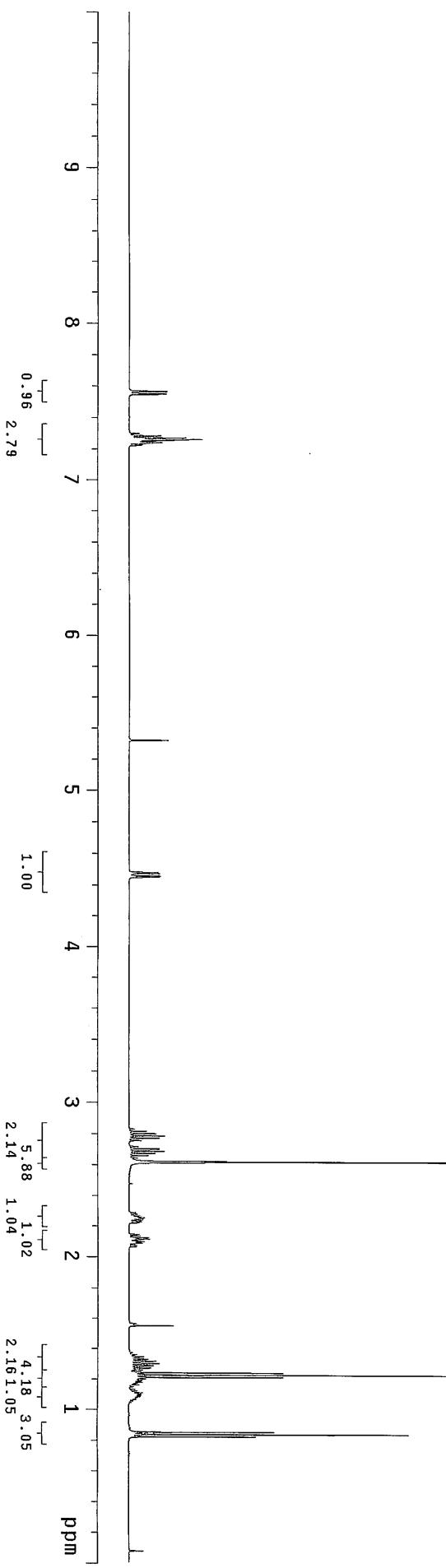


Table 4, entry 7



JC10143B C0C13

exp23 PROTON

SAMPLE	JUL 20 2013	PRESATURATION	n
SOLVENT	CDCl ₃	SATMODE	wet
FILE	/indy/jwchoi/~/	SPECIAL	n
43B_C0C13/PROTON01.~	temp	not used	n
ACQUISITION	.fid	32	20
SW	8000.0	SPIN	0.008
AT	3.000	HST	9.900
PP	48000	PW90	10.000
FB	not used	ALFA	
BS	32	FLAGS	
D1	2.000	I1	n
NT	16	IN	n
CT	16	DP	y
TRANSMITTER	H1	PROCESSING	nn
TN	499.698	1B	0.20
SFRQ	499.7	FN	not used
TOF	499.7	DISPLAY	-0.1
TPWR	61	WP	4996.8
PW	4.950	RFP1	4632.1
DECOPPLER	C13	RFP	3627.8
DN	0	RP	-83.7
D0F	0	1P	-75.2
DIN	0	PLOT	250
DECWAVE	W40_AUTOX7.~	WC	0
DPOWER	991	SC	60
DNP1	41	VS	50
	32258	TH	
		AI	
		CDC	
		PH	

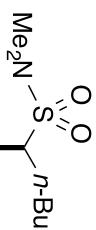
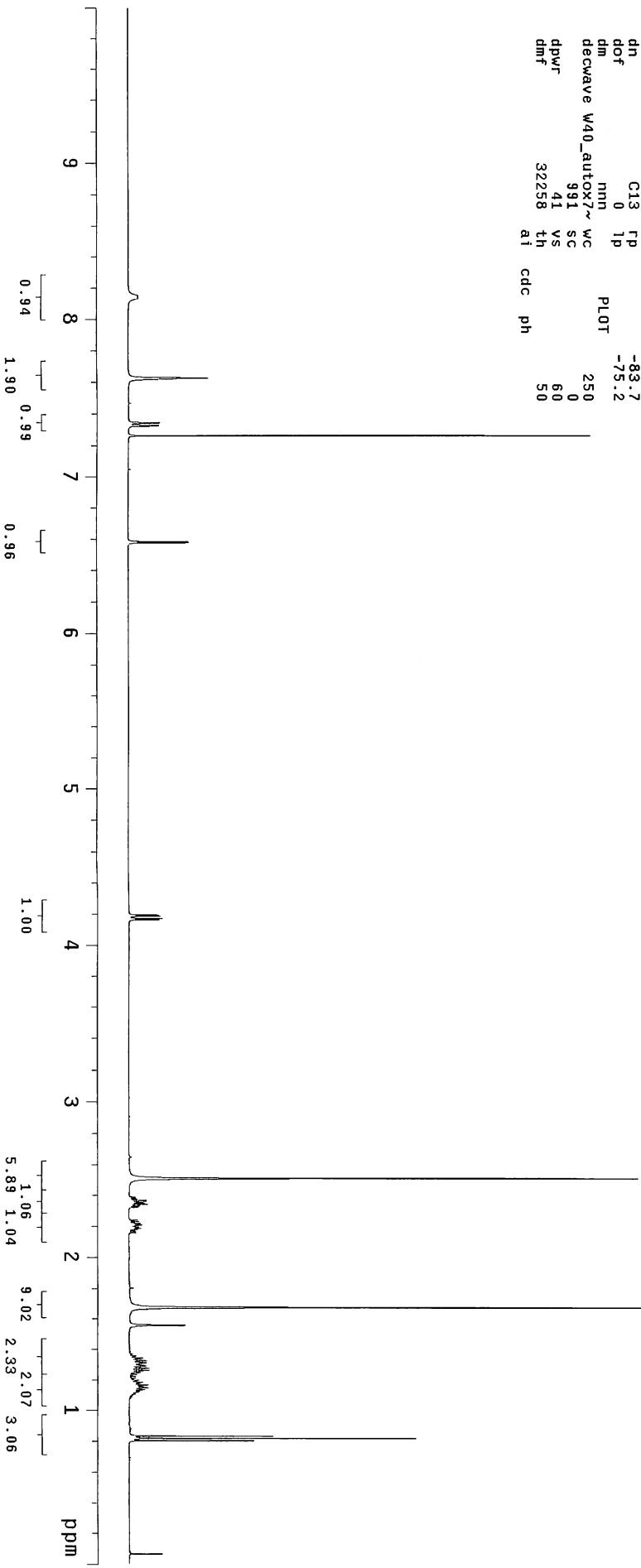


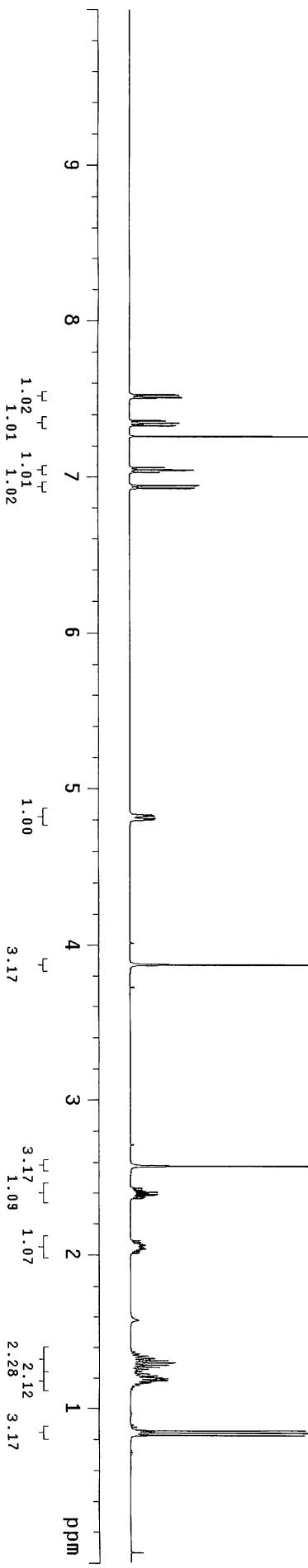
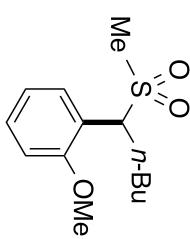
Table 4, entry 8



JC10219B CDC13

exp26 PROTON

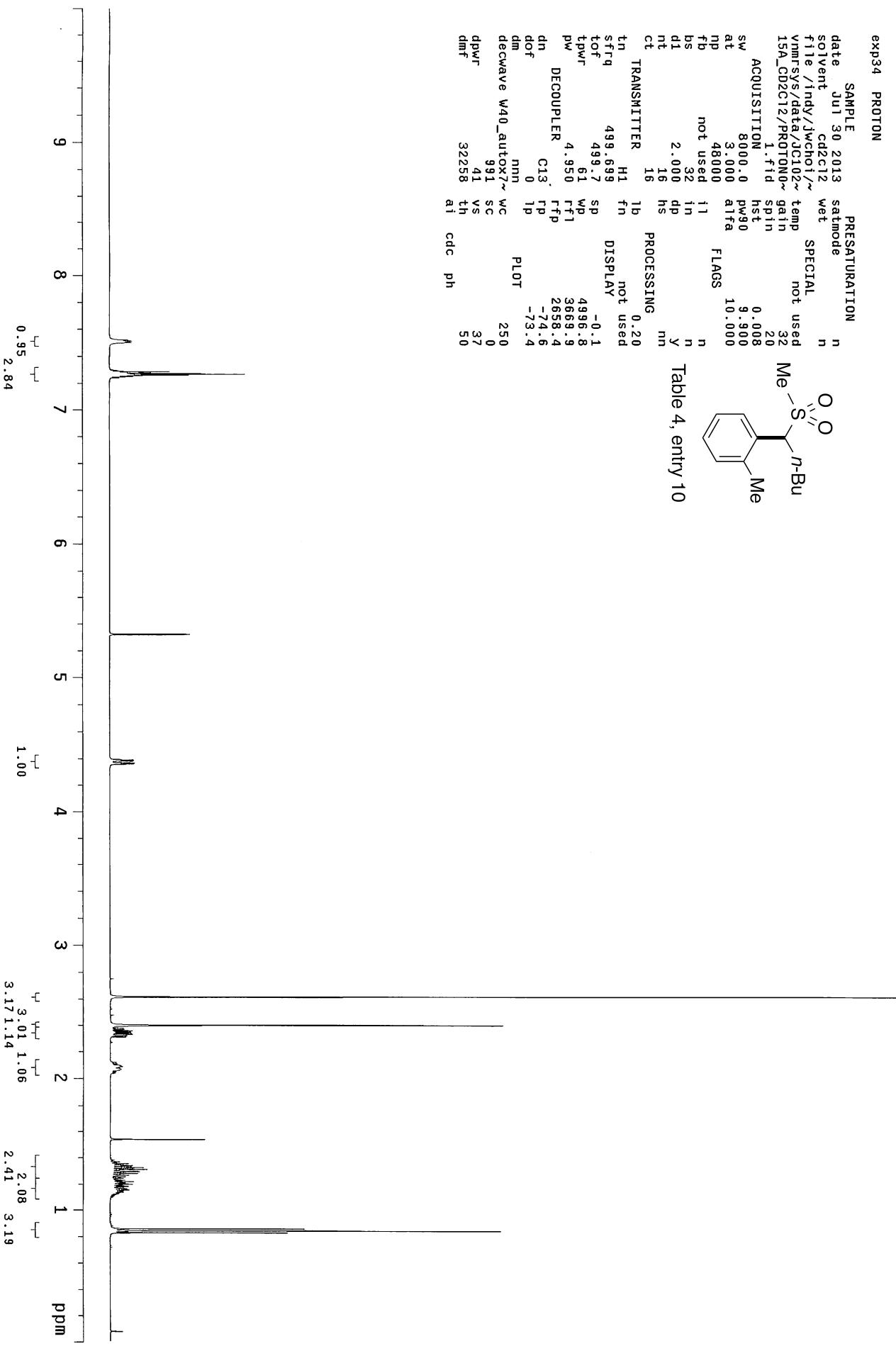
SAMPLE	1	2013	PRESATURATION	n
date	Aug	cdcl3	satmonde	n
solvent			wet	n
f1e	/indy/jwchoi/	~	SPECIAL	
vmrsys	data&JC102~		not used	
19B_CDCl3/PROTON01~		gain	32	
		spin	20	
		hst	0	
ACQUISITION		pw90	9.008	
sw	8000.0		9.900	
at	3.000	a1fa	10.000	
np	48000			
fb		FLAGS		
bs	not used	i1	n	
d1	32	in	n	
nt	16	dp	y	
ct	16	hs	nn	
TRANSMITTER		PROCESSING	0-20	
tn		1b	not used	
sfrq	499.698	fn	DISPLAY	
tof	499.7	sp	-0.1	
tprt	61	wp	4996.8	
pw	4.950	r _{f1}	4632.1	
DECOUPLER		r _{fp}	3627.8	
dn		C13	-83.0	
dof	0	r _p	-73.0	
dm		tp		
decwave	w40_autoX7~	NNN		
dpwr	991	WC	250	
dimf	41	SC	0	
	32258	VS	28	
		TH	50	
		ai	cddc	
			ph	



JC10215A CD2C12

exp34 PROTON

SAMPLE	Ju1 30 2013	PRESATURATION	n
solvent	cd2c12	satmode	n
file	/Indy/jwchoi/~/vnmrssr/data/JC102/	SPECIAL	n
15A_CD2C12/PROTON0~	gain	not used	32
ACQUISITION	1.fid	spin	2.0
sw	8000.0	hst	0.008
at	3.000	pw90	9.900
pp	48000	alfa	10.000
fb	not used	FLAGS	Me - S - O - O - n-Bu
bs	32	i1	Me
d1	2.000	in	Me
nt	16	dp	Table 4, entry 10
ct	16	hs	nn
TRANSMITTER	1b	PROCESSING	0.20
tn	H1	fn	not used
sfrq	499.699	DISPLAY	-0.1
tof	499.7	sp	4996.8
tpwr	61	wp	3669.9
pw	4.950	r _f 1	2658.4
DECOUPLER	C13	r _f p	-74.6
dn	13	r _p	-73.4
dof	0	1p	
din	nnn	PLOT	250
decwave	w40_autox7~	wc	991
dppr	41	sc	0
dmpf	32258	vs	37
		th	50
		ai	
		cdc	
		ph	

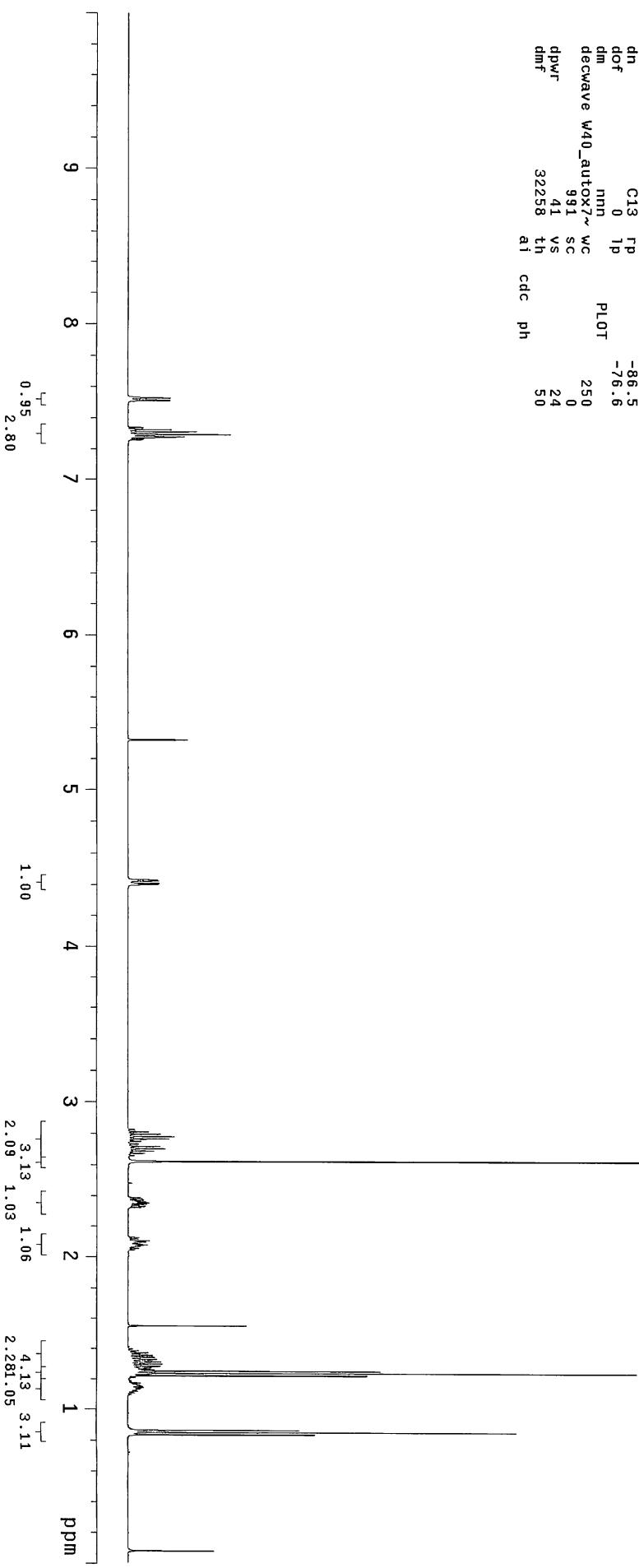
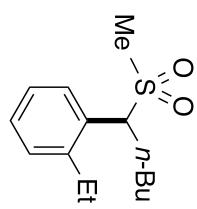


JC11023A 1H CD2C12

exp30 PROTON

SAMPLE date Oct 17 2013 solvent c6d6 file /'indy/jwchoi/~/ vnmrsys/data/JC110~/ 23A_1H_CD2C12/PROT~ 0N01.fid ACQUISITION sw 8000.0 at 3.000 np 48000 fb not used bs 32 d1 2.000 nt 16 ct 16 TRANSMITTER tn H1 fn not used sfrq 499.698 DISPLAY -0.1 tof 499.7 sp wp pw 4.950 rrf1 rfp DECOUPLER dn C13 rfp dof 0 Tp dm nnn decwave w40_autox7~ wc 991 sc 0 dpwr 41 vs 24 dm 32258 th ai cdc ph

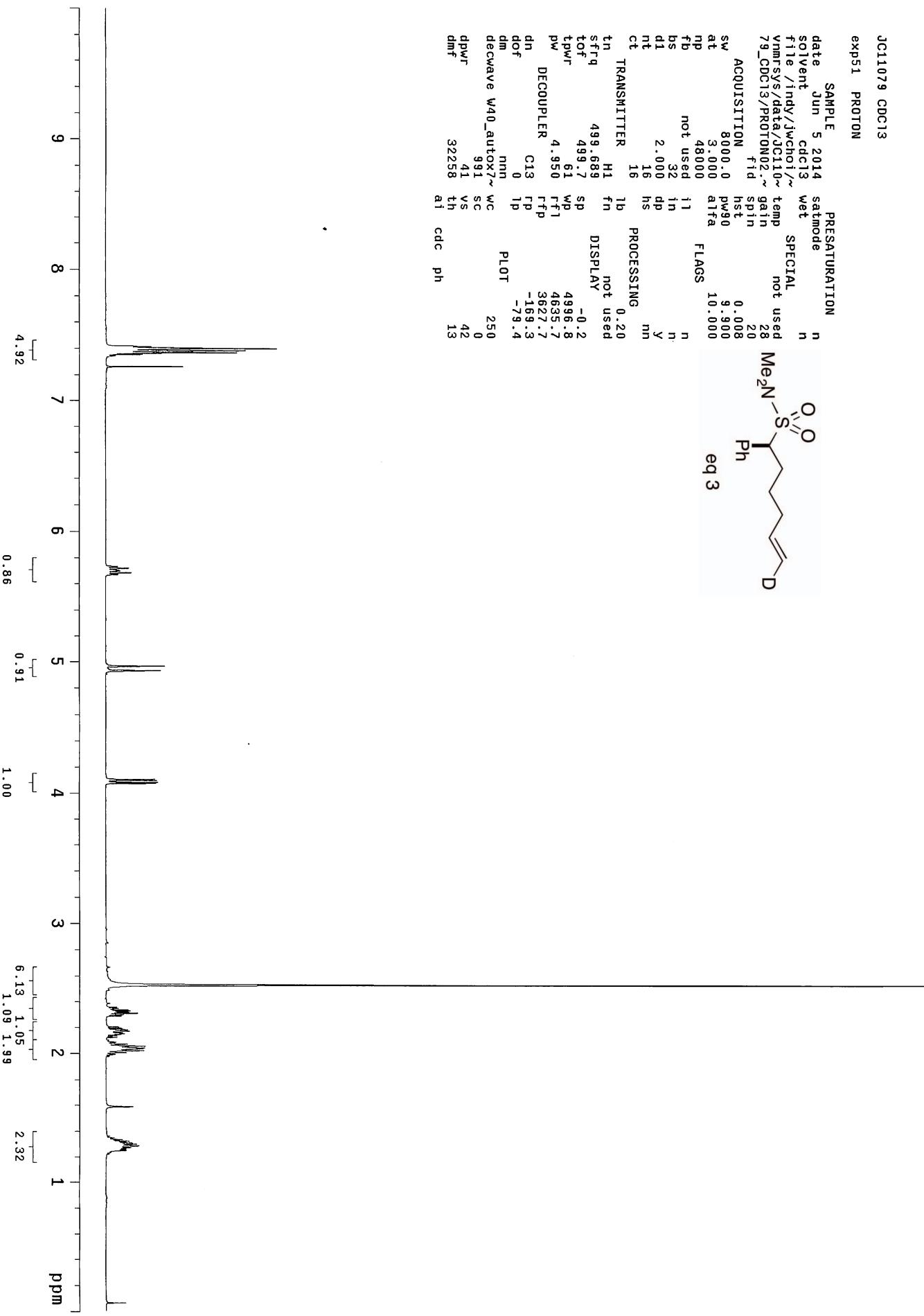
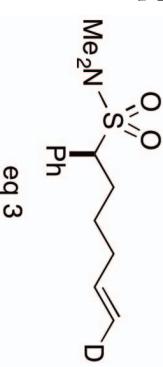
Table 4, entry 11



JC11079 CDC13

exp51 PROTON

SAMPLE	Jun 5 2014	PRESATURATION	n
date	cdd13	satmode	n
solvent	wet	SPECIAL	n
file	/indy/jwchoi/		
vnmrsys	data/JC110~		
79_CDC13/PROTON02.~	temp	not used	
ACQUISITION	fid	gain	28
sw	8000.0	spin	20
at	3.000	hst	0.008
np	48000	pw90	9.900
fb	not used	a1fa	10.000
bs	32	FLAGS	
d1	2.000	i1	0
nt	16	in	n
ct	16	dp	n
TRANSMITTER	H1	hs	y
tn	499.689	PROCESSING	nn
sfrq	499.689	0.20	
tof	499.7	not used	
tpwr	6.1	DISPLAY	-0.2
pw	4.950	4996.8	
DECOUPLER	C13	4635.7	
dn	rp	3623.7	
dof	0	-169.3	
dm	tp	-79.4	
decwave	wnn	PLOT	
dpwr	991	250	
dinf	41	sc	
	32258	vs	0
		42	
		th	13
		ai	
		cdc	
		ph	



JC10045B1 CDC13

exp51 PROTON

SAMPLE date Jun 3 2014 solvent cdc13 file /indy/jwchoi/~/vnmrsys/data/JC100~ 45B1_CDC13/PROTON0~

ACQUISITION 1.fid gain 28 spin 20 wet n SPECIAL not used

sw 8000.0 pw90 0.008

at 3.00 hst 9.900

np 48000 a1fa 10.000

fb not used i1 n

bs 32 in n

d1 2.000 dp y

nt 16 hs nn

ct 16 PROCESSING 0.20

TRANSMITTER tn H1 fn not used

sfrq 499.689 DISPLAY -0.2

tfrq 499.7 sp

tpwr 6.1 wp 4996.8

pw 4.950 rfp 4635.3

DECOUPLER dn C13 rp 3627.7

dof 0 tp -167.5

dm nnn -79.6

decwave w40_autox7~ wc 250

dpwr 9.91 sc 0

dmpw 4.1 vs 47

ai 32258 th 25

cddc ph

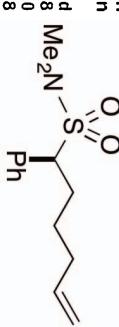
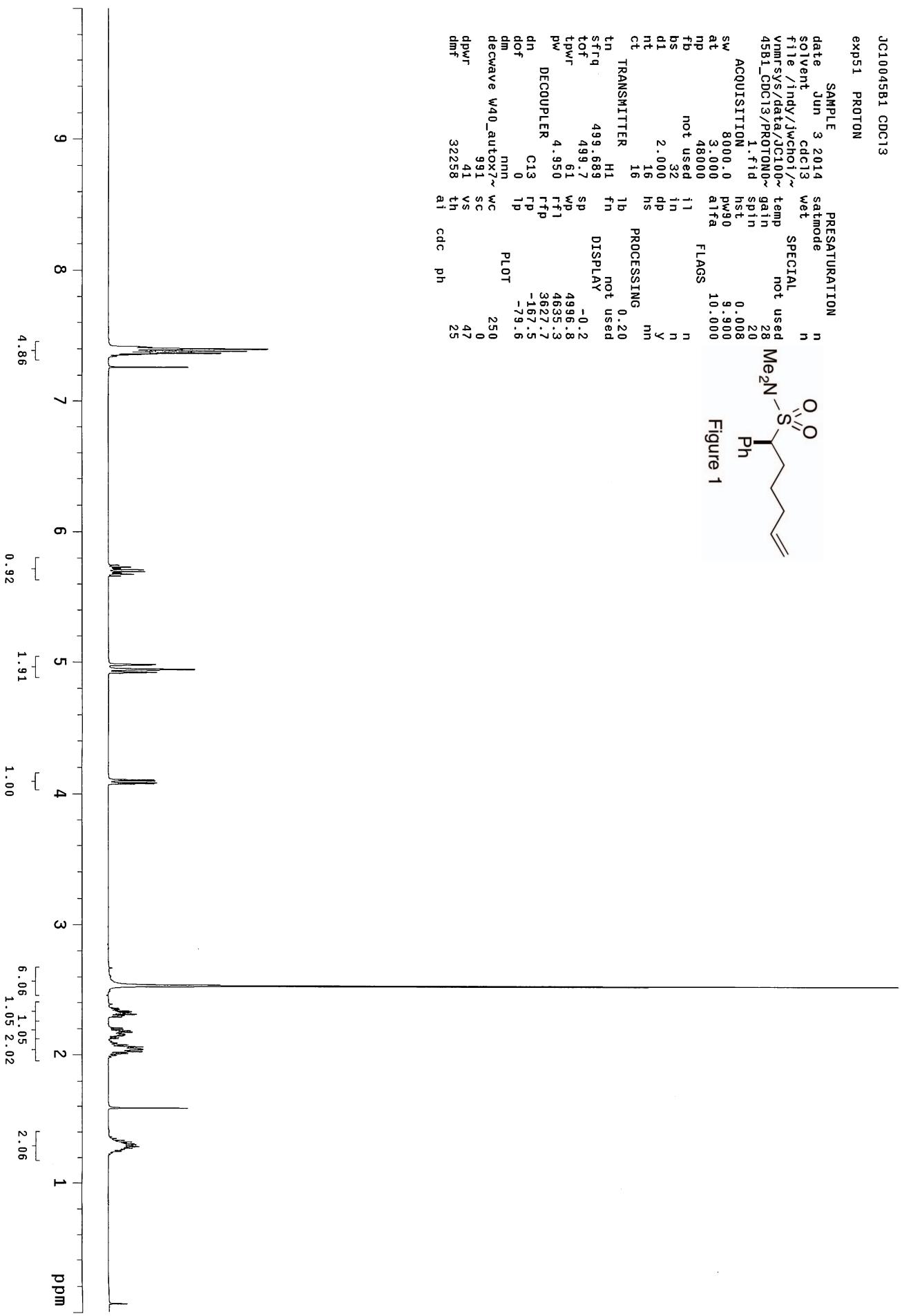
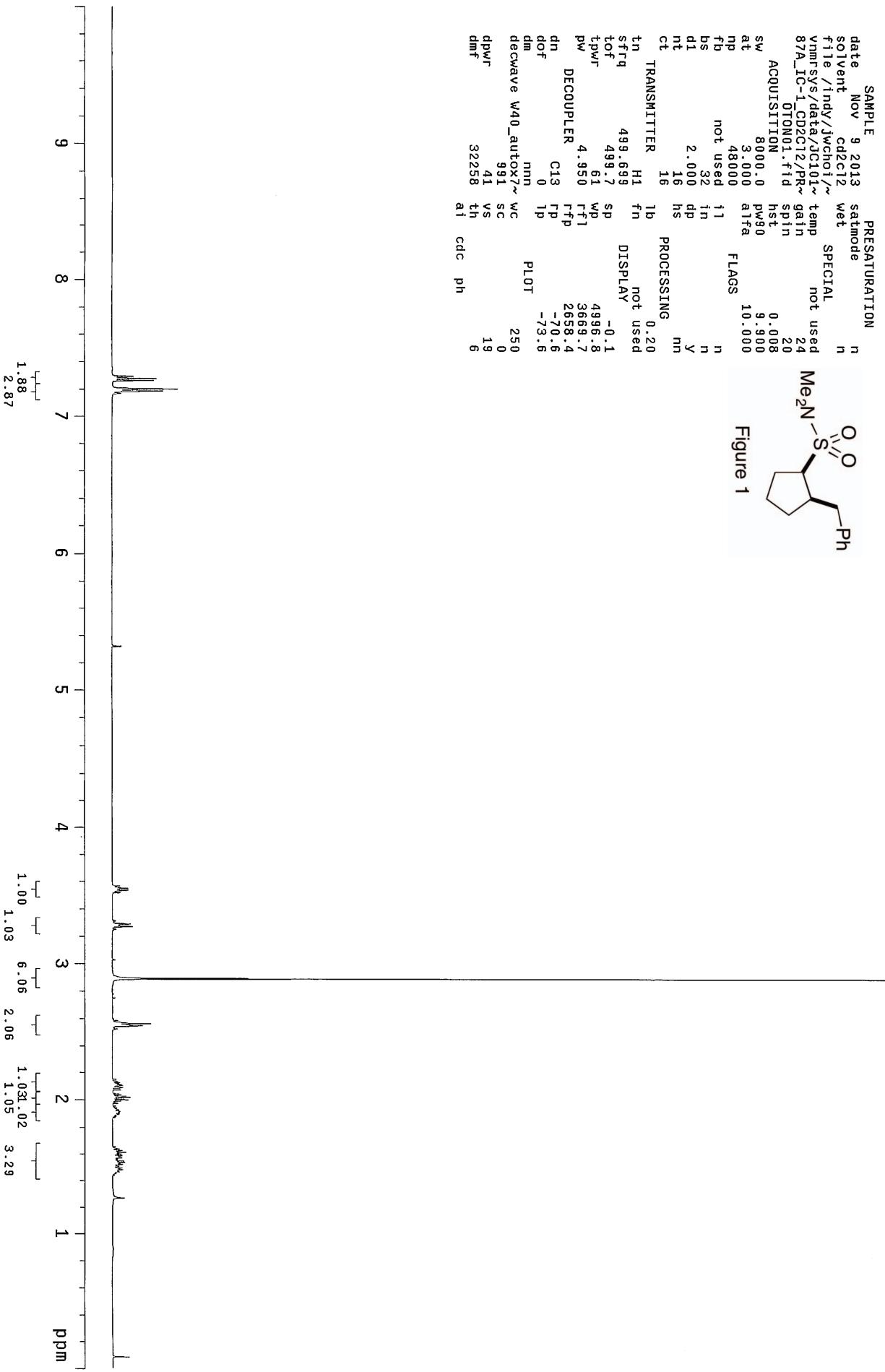
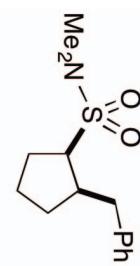


Figure 1



exp1 PROTON

date	SAMPLE	PRESATURATION
Nov	9 2013	satmode
solvent	cd2c12	wet
file	/indy/jwchoi/~/	SPECIAL
vmmrssys	data/JCI01/~/	not used
87A_IC-1_CD2C12/PR~	OTON01.fid	gain
ACQUISITION	OTON01.fid	spin
sw	8000.0	0 008
at	3.000	9 900
np	48000	10.000
fb	not used	a1fa
bs	32	FLAGS
d1	2.000	i1
nt	16	in
ct	16	dp
TRANSMITTER	Tb	hs
tn	H1	PROCESSING
sfrq	f1	0.20
t0f	499.699	not used
tpwr	499.7	DISPLAY
pw	4.950	-0.1
DECOUPLER	r _{f1}	4996.8
dn	r _{f1}	3669.7
dof	C13	2658.4
din	rp	-70.6
decwave	0	-73.6
dppwr	Tp	PLOT
dinr	nnn	250
decwave	wc	991
dppwr	sc	0
dinr	41	19
decwave	th	6
dinr	32258	a1
	cdc	cdc
	ph	ph



exp1 PROTON

SAMPLE	Nov 10 2013	PRESATURATION	n
date	cd2c12	satmode	n
solvent	/Indy/Juchoi/~/	SPECIAL	n
file	vnmrsys	temp	not used
87A_IA-1_CD2C12/PR~	OTON1.fid	gain	24
ACQUISITION		spin	20
sw	8000.0	0.008	
at	3.000	9.900	
np	48000	pw90	
fb	not used	alfa	10.000
bs	32	FLAGS	
d1	2.000	i1	n
nt	16	in	n
ct	16	dp	y
TRANSMITTER		hs	nn
tn	1b	PROCESSING	0.20
srrq	H1	fn	not used
t_pwr	499.699	DISPLAY	-0.1
pw	4.950	sp	4996.8
DECOUPLER	C13	r _{f1}	360.2
dn	0	r _{fp}	268.4
dof	0	rp	-66.3
din	0	tp	-80.4
decwave	w40_autox7~	PLOT	250
dppr	991	sc	0
dinr	32258	vs	26
		th	6
		ai	
		cdc	
		ph	

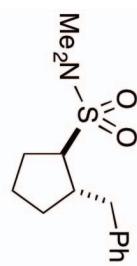
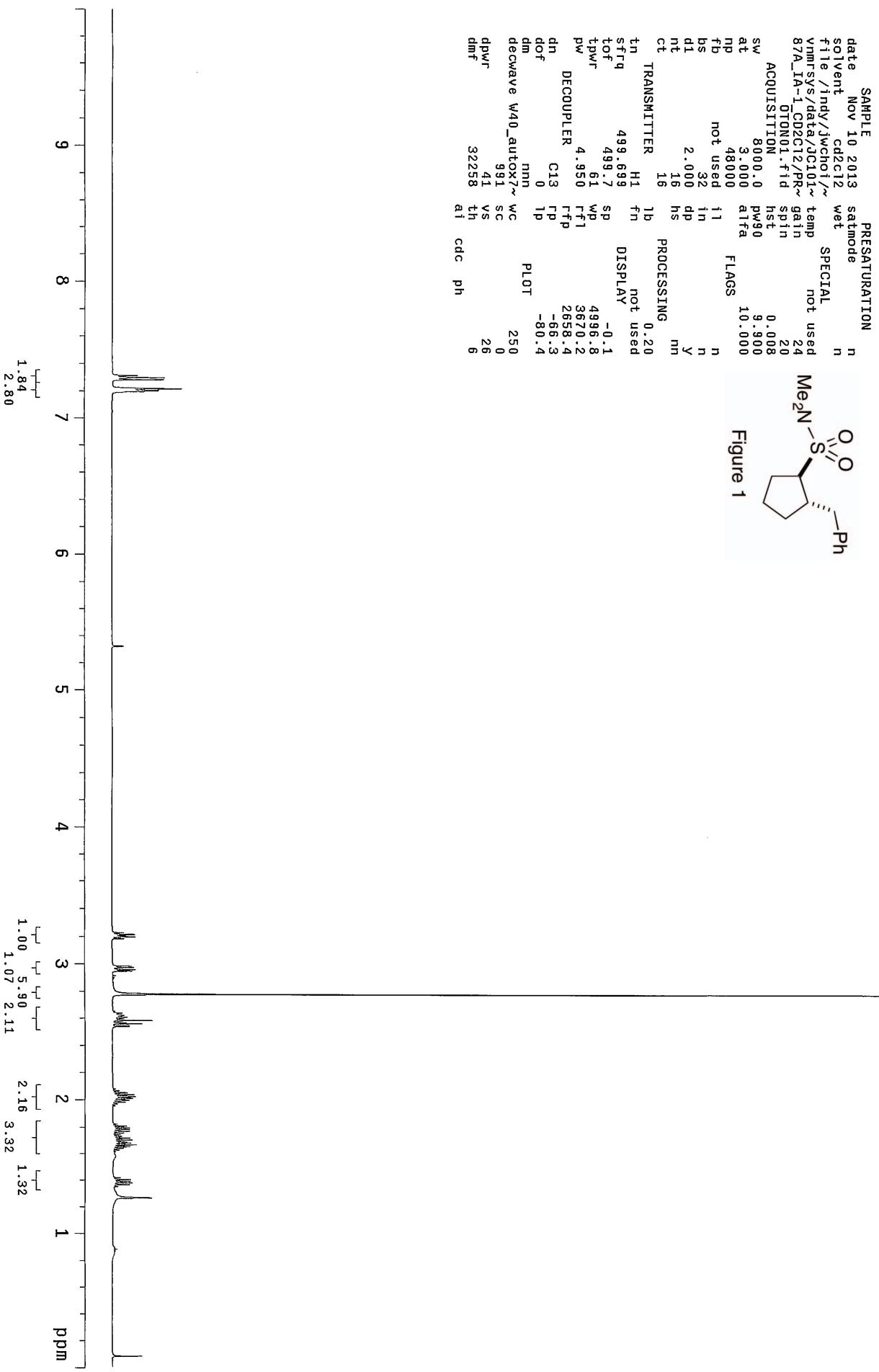
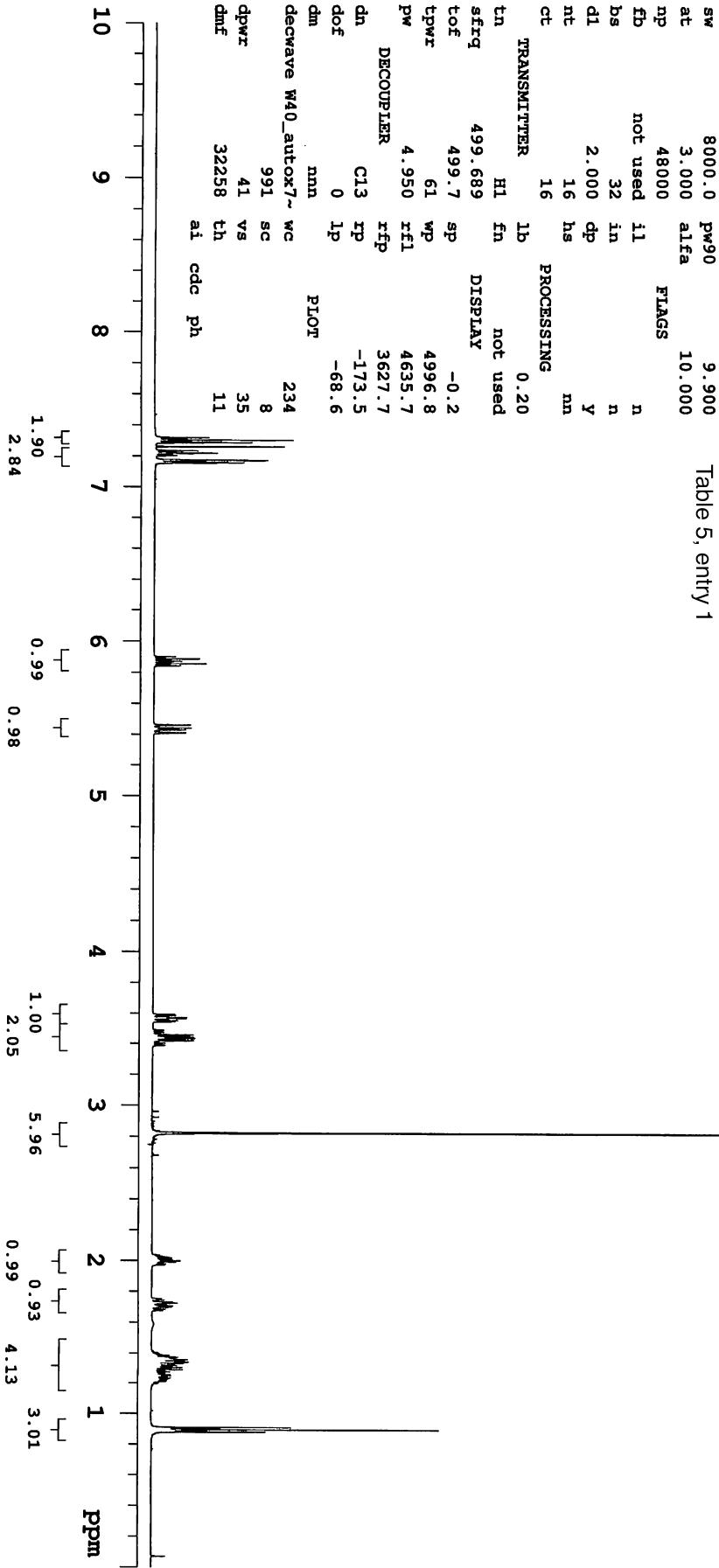


Figure 1



exp64 PROTON

SAMPLE	PRESATURATION	n	O O
date Aug 1 2014	satmode	n	Me - N - S - n-Bu
solvent cdc13	wet		Me
file /indy/jwchoi/~/vnmrsys/data/JC120-~	SPECIAL	25.0	CH ₂ Ph
75B_CDCl3/PROTON01~	gain	28	
.fid	spin	20	
ACQUISITION	hst:	0.008	
sw 8000.0	pw90	9.900	Table 5, entry 1
at 3.000	alfa	10.000	
np 48000	FLAGS		
fb not used	i1	n	
bs 32	in	n	
d1 2.000	dp	y	
nt 16	hs	nn	
ct 16	PROCESSING	0.20	
tn H1	fn	not used	
sfrq 499.689	DISPLAY		
tof 499.7	SP	-0.2	
tpwr 61	WP	4996.8	
pw 4.950	rfl	4635.7	
DECOUPLER	r _{fp}	3627.7	
dn C13	r _p	-173.5	
dof 0	lp	-68.6	
dm nnn	PLOT		
decwave W40_autox7~	wc	234	
dppwr 991	sc	8	
dmf 32258	th	35	
	ai	11	
	cdc	ph	



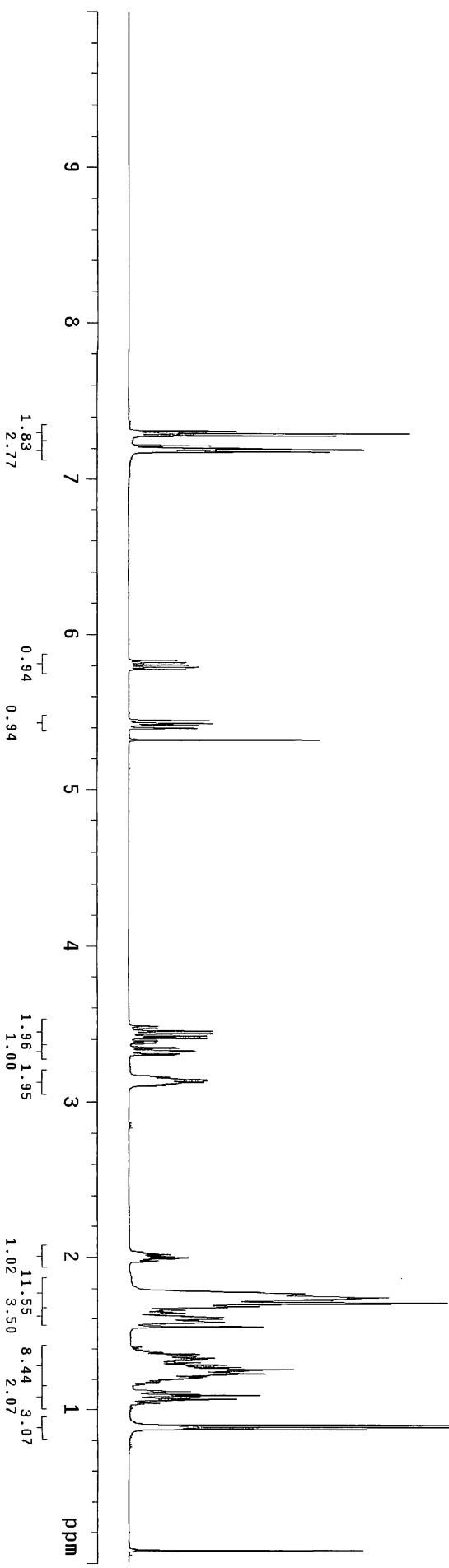
JC10275 CD2C12

exp30 PROTON

SAMPLE	SAMPLE	PRESATURATION
date Sep 5 2013	satmode	n
solvent cd2cl2	wet	n
file /indry/jwchoi/~	SPECIAL	
vnmrsys/data/JC102~	not used	
75_CD2Cl2/PROTON1~	gain	28
fid	size	20

Table 5, entry 2

	ACQUISITION	PROCESSING	DISPLAY	PLOT	FLAGS
sw	8000-0	hst	9	-	0
at	3.000	pwg0	10	-	0
np	48000	alfa	11	-	0
fb		not used	12	-	0
bs		in	13	-	0
d1	32	2.000	14	-	0
nt		dp	15	-	0
ct		16	16	-	0
TRANSMITTER		1b	1b	-	0
tn	499.699	H1	f1	not used	0
sfrq					0
tn	499.699				0
tof	499.7	sp	49	-	0
pw	6.61	wp	36	-	0
pw	4.950	r _{f1}	36	-	0
DECOUPLER		r _{f1}	26	-	0
dn	C13	r _p	26	-	0
dof	0	1p	-	-	0
dm	nnn				0
decwave	w40_autox7~	wc			0
dpwr	991	sc			0
dpwr	41	vs			0
dmf	32258	th			0
ai		cde			0



JC11025A 1H CDCl3

exp1 PROTON

	SAMPLE	PRESATURATION	
date	Oct 23 2013	satmode	n
solvent	cdcl3	wet	
file	/indy/jwchoi/~/	SPECIAL	n
vmrsys	vmrsys/data/tc110~	not used	
25A_1H_CDCl3/PROT~	N04_fid	gain	26
sw	8000.0	spin	20
ACQUISITION	pw90	hst	0.008
at	3.000	alfa	9.900
np	4800.0	FLAGS	10.000
fb	not used	i1	n
bs	32	in	n
d1	2.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	0.20
TRANSMITTER	1b	H1	not used
tn	499.698	fn	
sfrq	499.698	DISPLAY	-0.1
tof	499.7	sp	
tpwr	61	wp	4996.8
pw	4.950	r ^f 1	4632.1
DECOPPLER	C13	r ^p	3627.8
dn	0	1p	-87.5
dof		PLOT	-72.5
dm		nnn	
decwave	w40_autox7~	wc	250
dpwr	g91	sc	0
dmf	41	vs	37
	32258	th	13
	ai	cac	ph

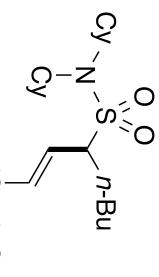
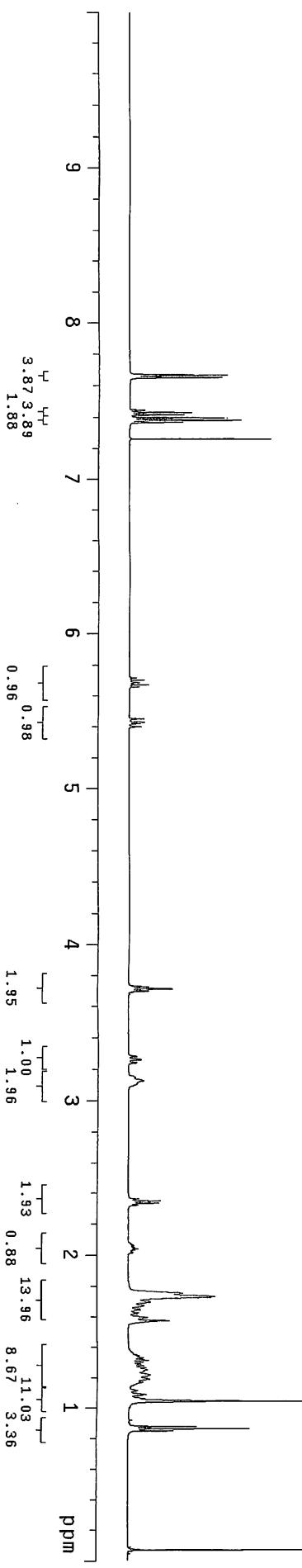


Table 5, entry 3



JC10279A CD2C12

exp30 PROTON

	SAMPLE	PRESATURATION
date	Sep 16 2013	satmode
solvent	cd2c12	wet
file	/indy/jwchoi/	
vnmrsy	/data/JC102~	
79A_CD2C12/PROTON0~	temp	SPECIAL
1.fid	gain	not used
ACQUISITION	1.	spin
sw	8000.0	24
at	3.000	20
np	48000	0.008
fb	not used	0
bs	32	9
d1	2.000	900
nt	16	10.000
ct	16	a1fa
TRANSMITTER	1b	FLAGS
tn	H1	i1
sfrq	499.699	in
tof	499.7	sp
tpwr	61	wp
pw	4.950	r ^f 1
DECOUPLER	C13	r ^f p
dn	0	rp
dof	0	1p
din	nnn	PLOT
decwave	w40_autox7~	250
dpwr	991	wc
dinf	41	250
	32258	vs
		80
		38
		ai
		cadc
		ph

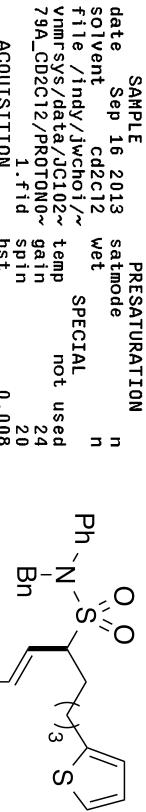
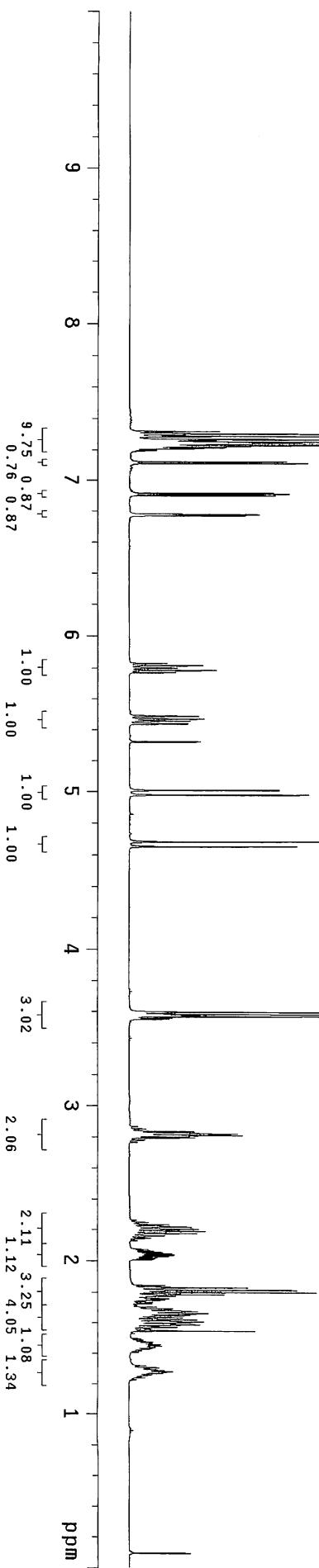


Table 5, entry 4



JC10281A 1H CDC13

exp2 PROTON

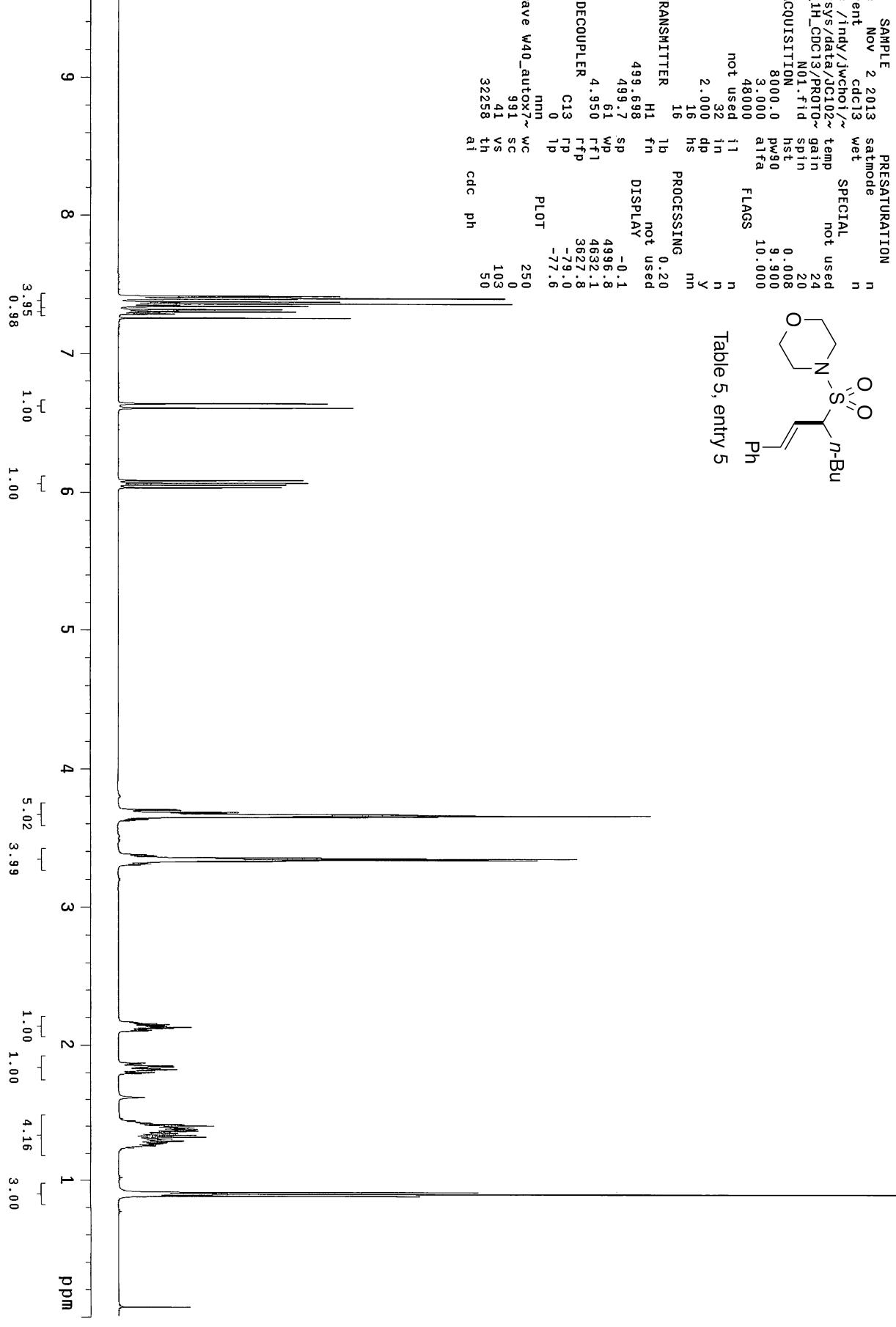
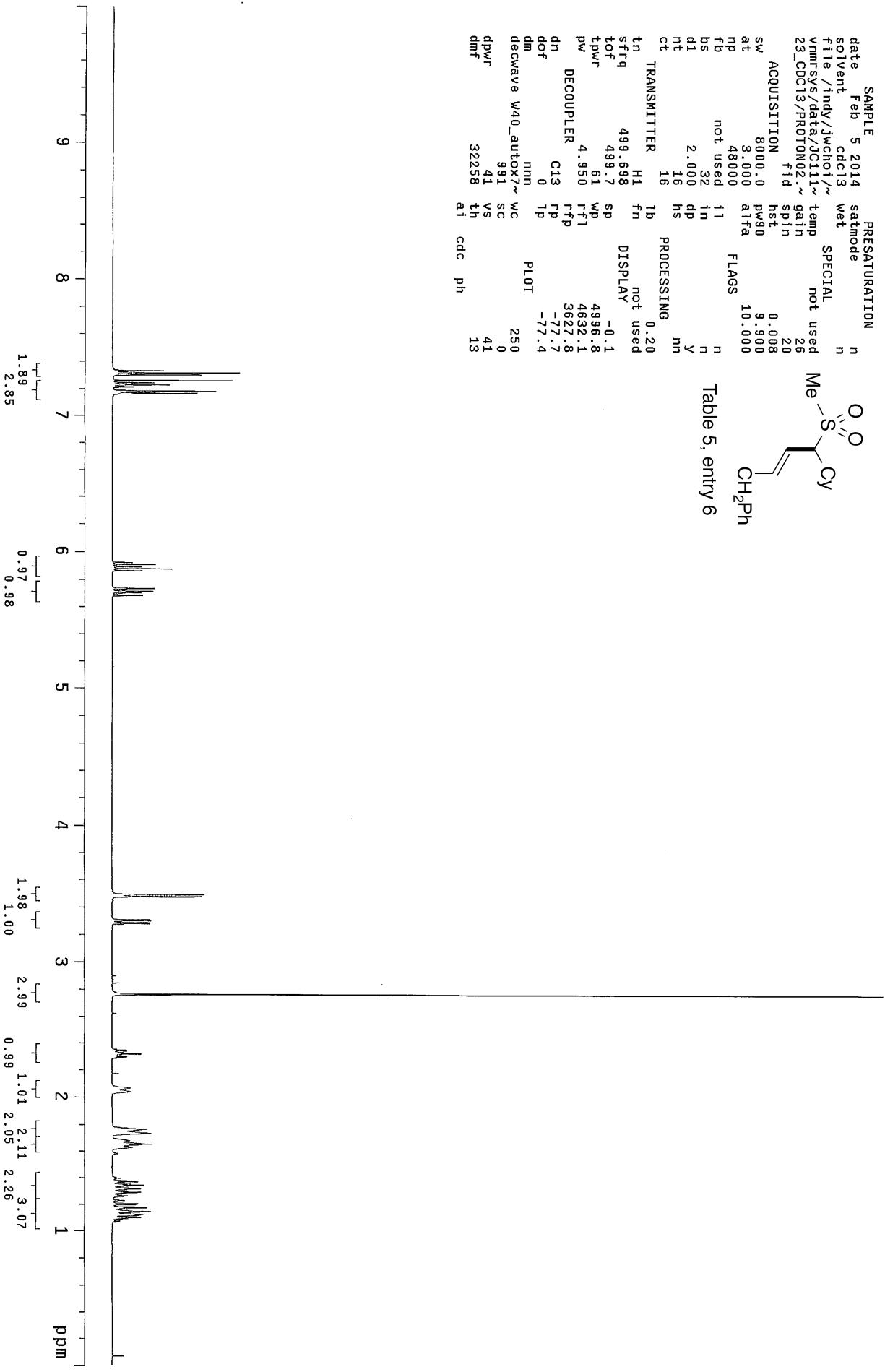


Table 5, entry 5

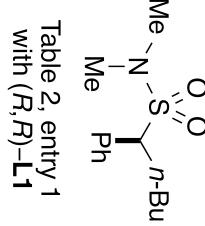
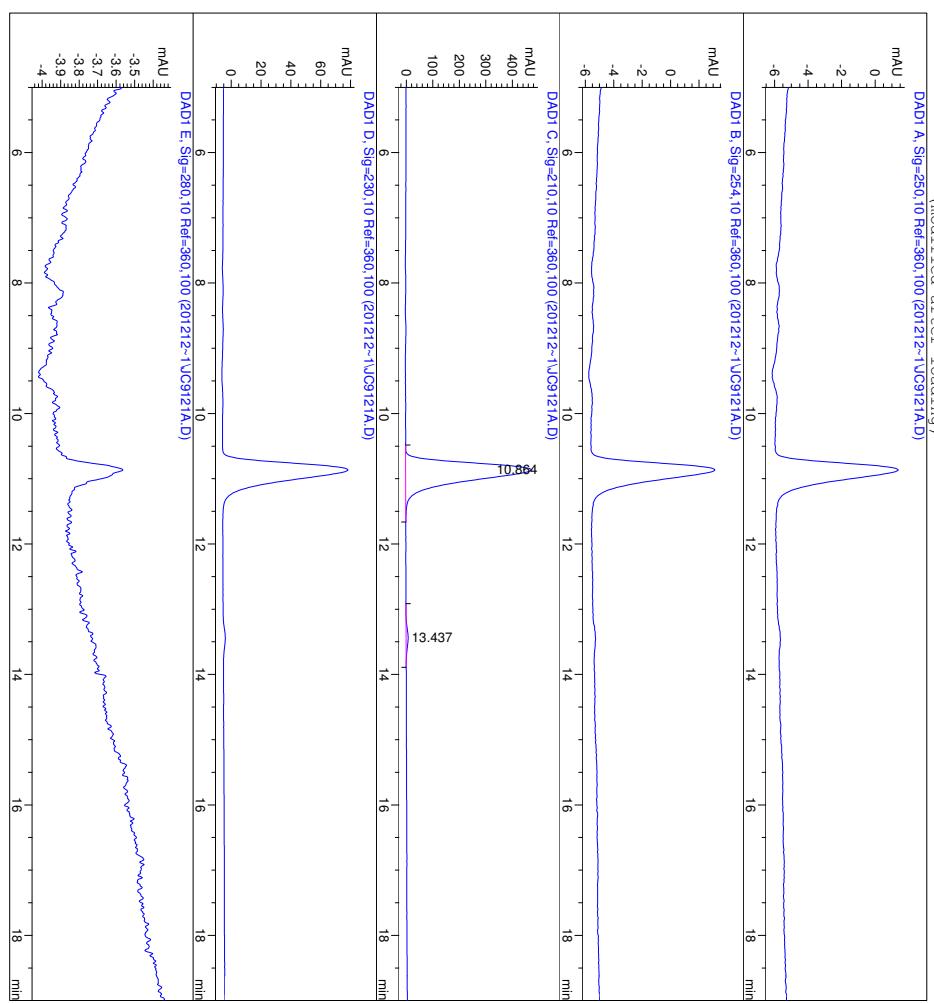
JC11123 CDC13

exp37 PROTON

	SAMPLE	PRESATURATION	
date	Feb 5 2014	satmode	n
solvent	cdcl3	wet	n
file	/indy/jwchoi/`	SPECIAL	
vmrssys	data/JC111`	temp	not used
23_CDCl3/PROTON02.^~	fid	gain	26
ACQUISITION	fid	spin	20
sw	8000.0	hst	0.008
at	3.000	pw90	9.900
np	48000	a1fa	10.000
fb	not used	FLAGS	Me
bs	i1		$\text{S}(\text{O})\text{O}$
d1	in		CY
nt	2.000		CH ₂ Ph
ct	16	PROCESSING	Table 5, entry 6
TRANSMITTER	1b	0-2.0	
tn	H1	fn	
sfrq	499.698	not used	
tof	499.7	DISPLAY	
tpwr	49.61	-0.1	
pw	4.950	wp	4996.8
DECOUPLER	rff1	rff1	4652.1
dn	C13	rfp	3677.8
dof	0	rp	-77.7
dif	0	tp	-77.4
decwave	nnn	PLOT	250
w40_autox7.^~	wc		
dpwr	991	sc	0
dif	32258	vs	41
		th	13
		ai	
		cdc	
		ph	



VII. HPLC Data



Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU:s] [mAU] %
 1 10.864 BB 0.2815 18388.73828 473.84717 98.0169
 2 13.437 PV 0.2756 173.77281 7.93276 1.9831

Totals : 8762.51109 481.77993

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

=====
 Signal 5: DAD1 E, Sig=280,10 Ref=360,100
 **** End of Report ****

S-90

Data File C:\HPCHEM\1\DATA\201212-1\JC9121A.D

Sample Name: JC9121A

Data File C:\HPCHEM\1\DATA\201212-1\JC9121A.D

Sample Name: JC9121A

=====
 Injection Date : 12/18/2012 2:33:11 PM Seq. Line : 1
 =====

=====
 Sample Name : JC9121A Location : Vial 4
 =====

=====
 Acq. Operator : CE Inj : 1
 =====

=====
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 =====

=====
 Different Inj Volume from Sequence ! Actual Inj Volume : 5 μ l
 =====

=====
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01.40.M
 =====

=====
 Last changed : 12/18/2012 2:48:17 PM by CE
 =====

=====
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02.20.M
 =====

=====
 Last changed : 7/30/2014 4:56:50 PM by MK
 =====

=====
 (modified after loading)
 =====

=====
 Signal
 =====

=====
 Multiplier : 1.0000
 =====

=====
 Dilution 1.0000
 =====

=====
 Use Multiplier & Dilution Factor with ISTDS
 =====

=====
 Signal 1: DAD1 A, Sig=250,10 Ref=360,100
 =====

=====
 Signal 2: DAD1 B, Sig=254,10 Ref=360,100
 =====

=====
 Signal 3: DAD1 C, Sig=210,10 Ref=360,100
 =====

=====
 Area Percent Report
 =====

=====
 =====

Data File C:\HPCHEM\1\DATA\201212-1\JC9121B.D

Sample Name: JC9121B

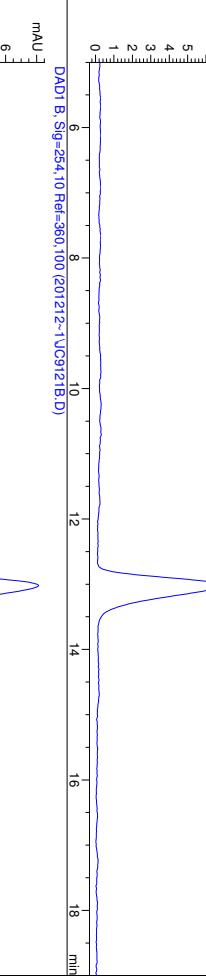
Data File C:\HPCHEM\1\DATA\201212-1\JC9121B.D

Sample Name: JC9121B

=====
 Injection Date : 12/18/2012 3:04:38 PM
 Sample Name : JC9121B
 Location : Vial 5
 Acq. Operator : CE
 Acq. Instrument : Instrument 1
 Inj : 1
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01-30.M
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Last changed : 4/7/2011 2:40:51 AM by CC
 Analysis Method : C:\HPCHEM\1\METHODS\AN00540.M
 Last changed : 8/2/2014 8:3:12 PM by MK
 (modified after loading)

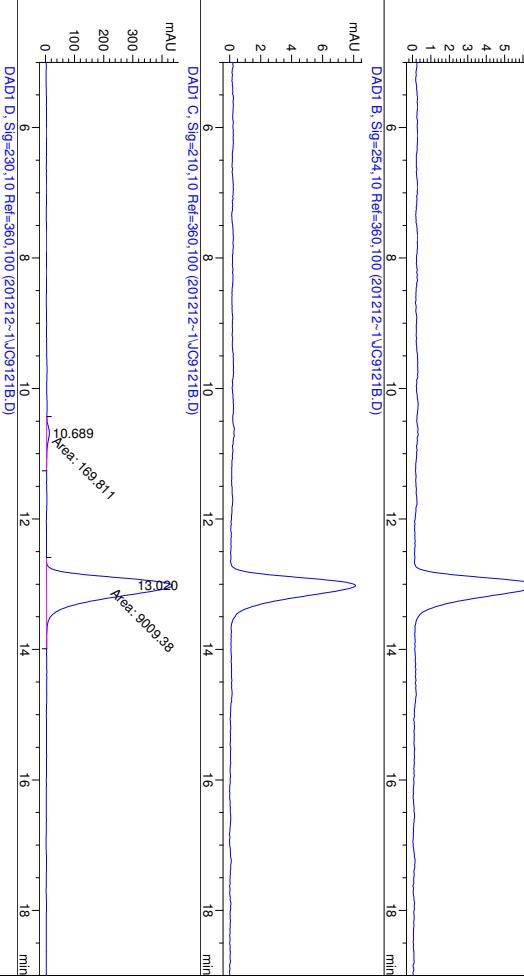
DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9121B.D)

mAU



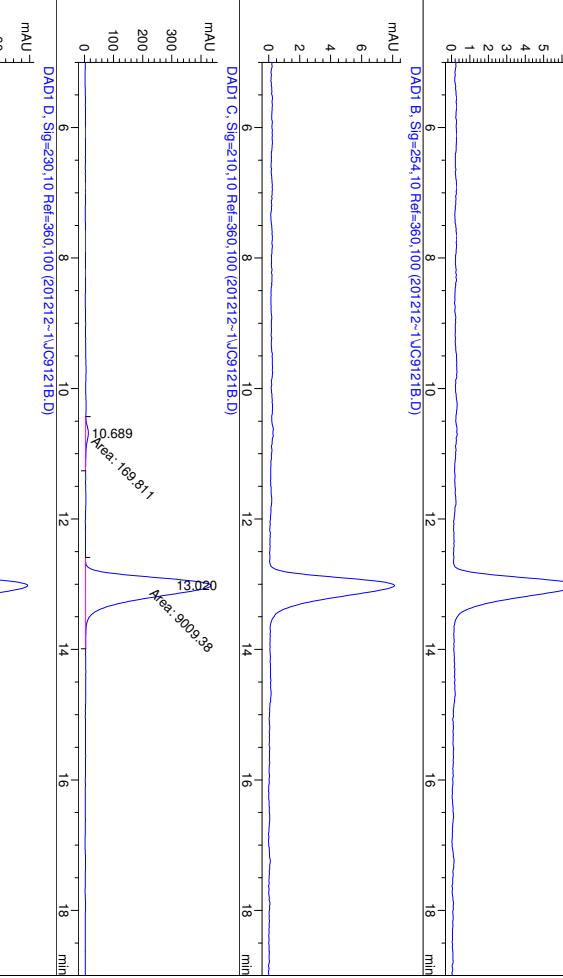
Signal 2: DAD1 B, Sig=254,10 Ref=360,100

mAU



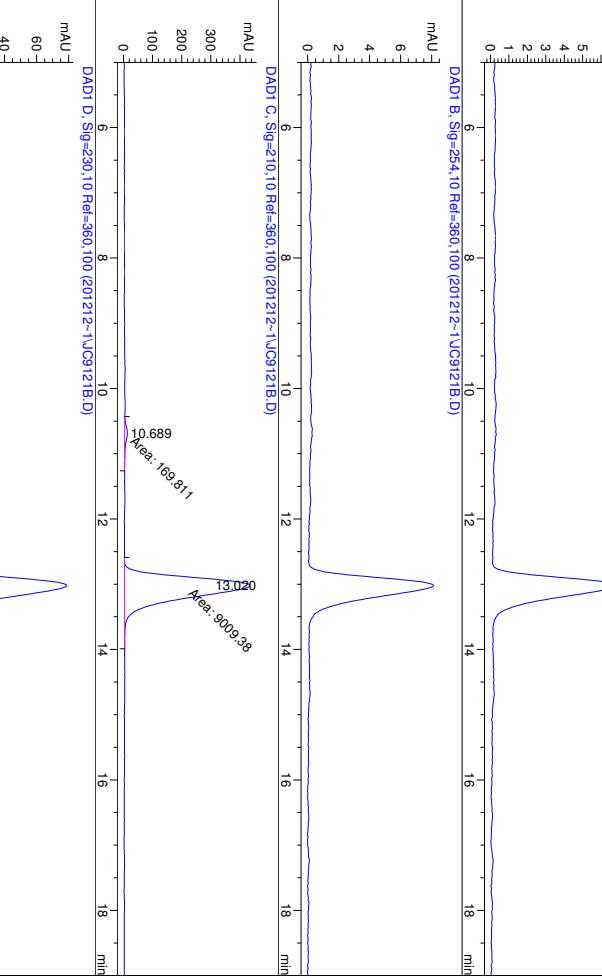
Signal 3: DAD1 C, Sig=210,10 Ref=360,100

mAU



Signal 4: DAD1 D, Sig=230,10 Ref=360,100

mAU



Signal 5: DAD1 E, Sig=280,10 Ref=360,100

mAU

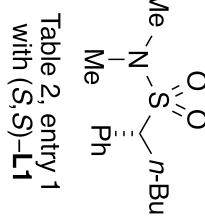
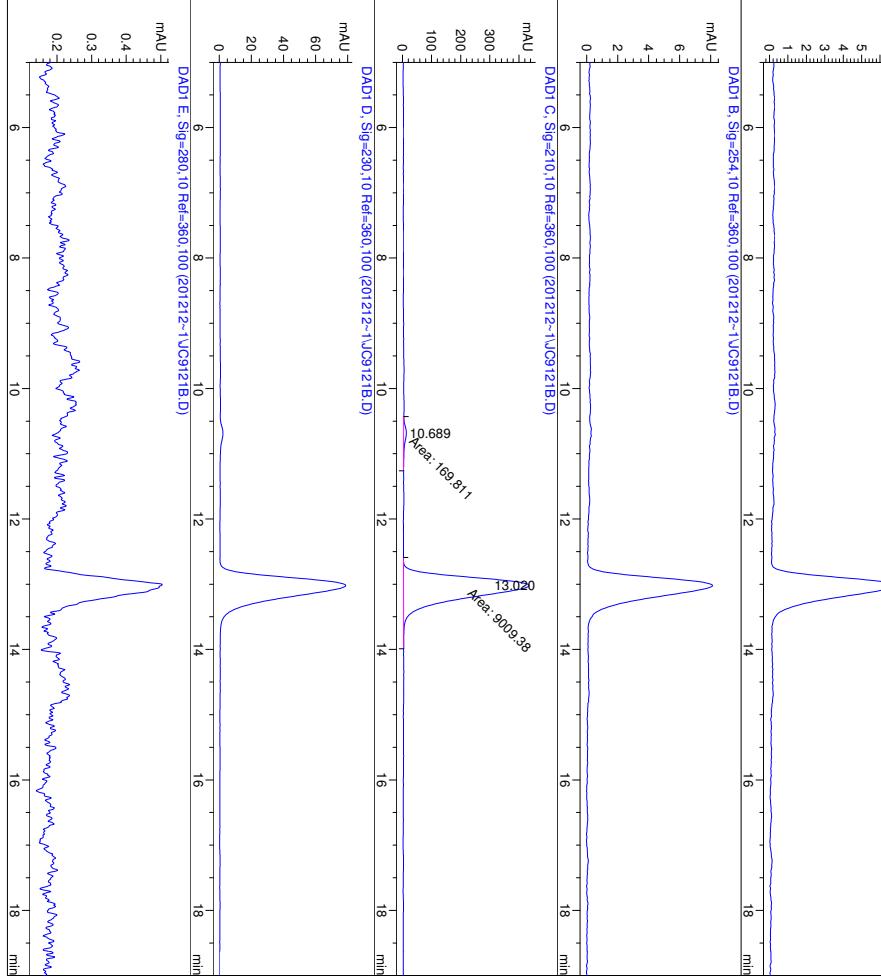


Table 2, entry 1
with (*S,S*)-L1

Peak #	Ret Time [min]	Type	Width [min]	Area [mAU]	Height [mAU]	Area %
1	10.689	FM	0.2978	169.81142	9.50306	1.8500
2	13.020	MM	0.3481	9909.37891	431.37000	98.1500

Totals : 9179.19032 440.87395

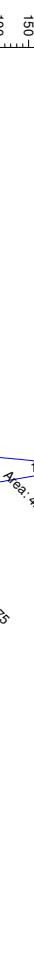
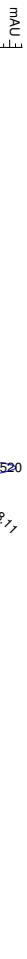
Results obtained with enhanced integrator!

Signal 4: DAD1, Sig=230,10 Ref=360,100

=====
 *** End of Report ***
 =====

Injection Date : 1/13/2013 11:37:20 PM Seq. Line : 2
 Sample Name : JC9153A Location : Vial 3
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 3 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-02-40.M
 Last changed : 8/9/2012 4:54:01 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 4:55:19 PM by MK
 (modified after loading)

DAD1 A, Sig=250,10 Ref=360,100 (201301~1\JC9153A1.D)



Signal 1: DAD1 A, Sig=250,10 Ref=360,100
 Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %
 1 10.520 MF 0.3390 4288.11475 210.79334 96.5968
 2 11.708 FM 0.3773 151.07475 6.67396 3.4032
 Totals : 4439.18950 217.46631

Results obtained with enhanced integrator!

Signal 2: DAD1 B, Sig=254,10 Ref=360,100
 Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %
 1 10.520 MF 0.3389 2415.18042 118.77331 96.5410
 2 11.707 FM 0.3774 86.53488 3.82184 3.4590
 Totals : 2501.71530 122.59315

Results obtained with enhanced integrator!

Signal 3: DAD1 C, Sig=210,10 Ref=360,100
 Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %
 1 10.519 MF 0.3557 2.14168e4 1003.47882 96.1911
 2 11.708 FM 0.3883 848.05426 36.40364 3.8089
 Totals : 2.22639e4 1039.88247

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100
 Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %
 1 10.520 BB 0.3125 1.22823e4 606.00146 96.7746
 2 11.709 BB 0.3369 409.36346 18.58926 3.2254
 Totals : 1.26917e4 624.59073

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100
 Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %
 1 10.520 BB 0.3125 1.22823e4 606.00146 96.7746
 2 11.709 BB 0.3369 409.36346 18.58926 3.2254
 Totals : 1.26917e4 624.59073

Results obtained with enhanced integrator!

DAD1 E, Sig=280,10 Ref=360,100 (201301~1\JC9153A1.D)
 mAU

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

***** End of Report *****

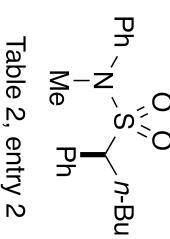
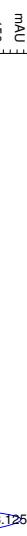


Table 2, entry 2
with (R, R)-L1

=====
 Injection Date : 1/13/2013 9:26:32 PM
 Sample Name : JC9153B
 Location : Vial 4
 Acq. Operator : CE
 Acq. Instrument : Instrument 1
 Inj : 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 3 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-02-30.M
 Last changed : 4/19/2010 4:08:56 PM by sz
 Analysis Method : C:\HPCHEM\1\METHODS\AN00540.M
 Last changed : 8/1/2014 11:08:22 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201301~1\JC9153B1.D)



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.026	BP	0.3146	113.62238	5.33121	2.4632
2	13.125	BB	0.3752	4499.24658	185.38203	97.5368
Totals :				2610.51468	190.71324	

Results obtained with enhanced integrator!

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.023	MM	0.3532	59.19446	2.79296	2.2676
2	13.125	MM	0.4060	2551.31982	104.72879	97.7324

Totals : 2610.51468 190.71324

Results obtained with enhanced integrator!

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.025	MM	0.3401	560.64459	27.47764	2.3671
2	13.125	MM	0.4189	2.31246e4	919.98969	97.6329

Totals : 2.36502e4 947.46732

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.030	BP	0.3344	358.89542	16.20137	2.7223
2	13.125	BB	0.3746	1.28247e4	529.39978	97.2777

Totals : 1.31836e4 545.60115

Results obtained with enhanced integrator!

Signal	DAD1 E, Sig=280,10 Ref=360,100 (201301~1\JC9153B1.D)
5	0

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====

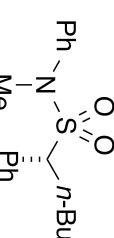


Table 2, entry 2
with (S,S)-L1

Data File C:\HPCHEM\1\DATA\201301~1\JC9149A2.D

Sample Name: JC9149A

Data File C:\HPCHEM\1\DATA\201301~1\JC9149A2.D

Sample Name: JC9149A

=====
 Injection Date : 1/5/2013 12:33:05 AM
 Sample Name : JC9149A
 Location : Vial 6
 Acq. Operator : CE
 Inj : 1
 Acq. Instrument : Instrument 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AD-02-40.M
 Last changed : 1/4/2013 10:05:27 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 4:58:11 PM by MK
 (modified after loading)

=====
 Seq. Line : 6
 Location : Vial 6
 Inj : 1
 O O
 $\text{Bn}-\text{N}-\text{S}-\text{n-Bu}$
 Me
 Ph

DAD1, Sig=250,10 Ref=360,100 (201301~1\JC9149A2.D)

mAU



Signal 2: DAD1, Sig=254,10 Ref=360,100

Signal 3: DAD1, C, Sig=210,10 Ref=360,100

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Totals :

1.24443e4

294.61277

mAU



Signal 2: DAD1, Sig=254,10 Ref=360,100

Signal 3: DAD1, C, Sig=210,10 Ref=360,100

Signal 4: DAD1, D, Sig=230,10 Ref=360,100

Totals :

1.24443e4

294.61277

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

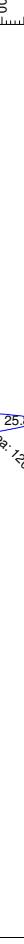
=====
 *** End of Report ***
 =====

Table 2, entry 3
with (*R*,*R*)-L1

=====
 Area Percent Report
 =====

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU]	Height [mAU]	Area %
1	25.848	MM	0.7041	1.12040e4	285.00970	96.7508
2	28.891	MM	0.7018	404.33865	9.60306	3.2492

mAU



Signal 2: DAD1, D, Sig=230,10 Ref=360,100

Signal 3: DAD1, D, Sig=230,10 Ref=360,100

Signal 4: DAD1, D, Sig=230,10 Ref=360,100

Totals :

1.24443e4

294.61277

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====

mAU



Signal 2: DAD1, E, Sig=280,10 Ref=360,100

Signal 3: DAD1, E, Sig=280,10 Ref=360,100

Signal 4: DAD1, E, Sig=280,10 Ref=360,100

Totals :

1.24443e4

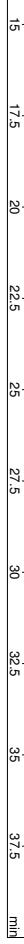
294.61277

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====

mAU



Signal 2: DAD1, E, Sig=280,10 Ref=360,100

Signal 3: DAD1, E, Sig=280,10 Ref=360,100

Signal 4: DAD1, E, Sig=280,10 Ref=360,100

Totals :

1.24443e4

294.61277

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====

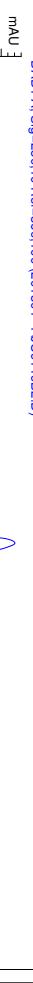
Data File C:\HPCHEM\1\DATA\201301~1\JC9149B2.D

Sample Name: JC9149B

Sample Name: JC9149B

=====
Injection Date : 1/5/2013 1:14:21 AM Seq. Line : 7
Sample Name : JC9149B Location : Vial 7
Acq. Operator : CE Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
Acq. Method : C:\HPCHEM\1\METHODS\AD-02-40.M
Last changed : 1/4/2013 10:05:27 PM by CE
Analysis Method : C:\HPCHEM\1\METHODS\AD00540.M
Last changed : 8/1/2014 11:12:30 PM by MK
(modified after loading)

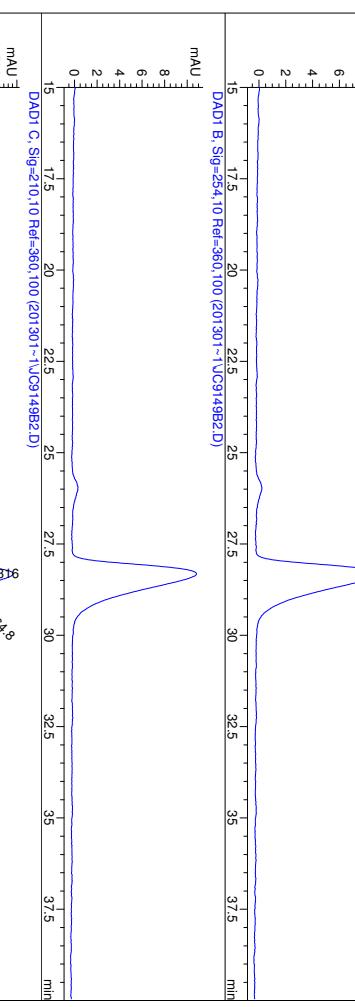
DAD1, Sig=250,10 Ref=360,100 (201301-1\JC9149B2.D)



mAU

0
4
8
12
16
20
24
28
32
36

min



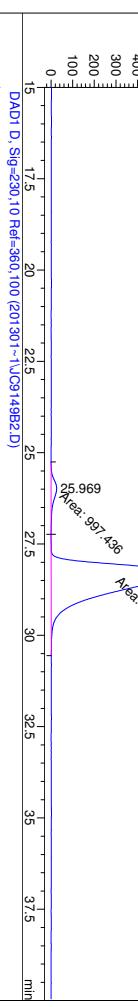
DAD1 B, Sig=254,10 Ref=360,100 (201301-1\JC9149B2.D)



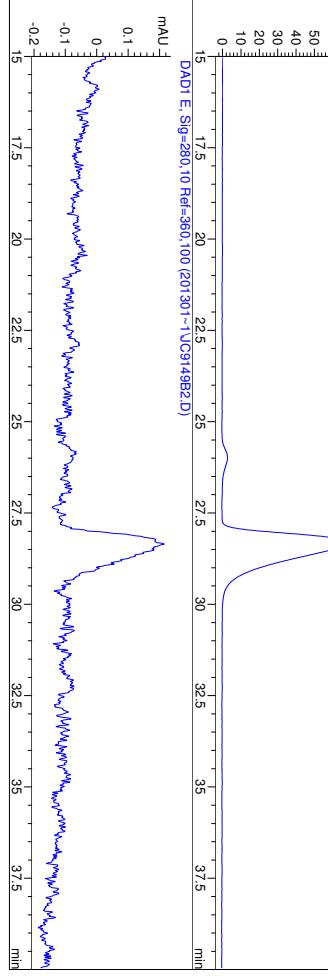
mAU

0
2
4
6
8
10
12
14
16
18
20
22
24
26
28
30
32
34
36
38

min



DAD1 D, Sig=230,10 Ref=360,100 (201301-1\JC9149B2.D)



DAD1 E, Sig=280,10 Ref=360,100 (201301-1\JC9149B2.D)

mAU

0
15
30
45
60
75
90
105
120
135
150
165
180
195
210
225
240
255
270
285
300
315
330
345
360
375

min

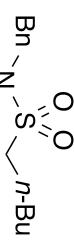


Table 2, entry 3
with (S,S)-L1

Data File C:\HPCHEM\1\DATA\201301~1\JC9149B2.D

Sample Name: JC9149B

=====
Area Percent Report

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDS

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	25.969	MM	0.6423	997.43585	25.88220	3.3740
2	28.316	MM	0.8177	2.85648e4	582.20422	96.6260

Totals : 2.95622e4 608.08643

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

=====
*** End of Report ***
=====

Injection Date : 1/3/2013 6:22:31 PM Seq. Line : 3
 Sample Name : JC9147B Location : Vial 5

Acq. Operator : CE Inj : 1

Acq. Instrument : Instrument 1 Inj Volume : 5 μ l

Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l

Acq. Method : C:\HPCHEM\1\METHODS\AD-02-30.M

Last changed : 1/3/2013 5:29:04 PM by CE

Analysis Method : C:\HPCHEM\1\METHODS\AD00540.M

Last changed : 8/1/2014 11:14:15 PM by MK

(modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201301~1\JC9147B.D)

mAU

8
6
4
2
0

7.5 10 12.5 15 17.5 20 22.5 25 27.5 min

DAD1 B, Sig=254,10 Ref=360,100 (201301~1\JC9147B.D)

mAU

8
6
4
2
0

7.5 10 12.5 15 17.5 20 22.5 25 27.5 min

DAD1 C, Sig=210,10 Ref=360,100 (201301~1\JC9147B.D)

mAU

10
8
6
4
2
0

7.5 10 12.5 15 17.5 20 22.5 25 27.5 min

DAD1 D, Sig=230,10 Ref=360,100 (201301~1\JC9147B.D)

mAU

500
400
300
200
100
0

7.5 10 12.5 15 17.5 20 22.5 25 27.5 min

DAD1 E, Sig=280,10 Ref=360,100 (201301~1\JC9147B.D)

mAU

80
60
40
20
0

7.5 10 12.5 15 17.5 20 22.5 25 27.5 min

Signal 2: DAD1, Sig=250,10 Ref=360,100

Signal Percent Report

Sorted By Signal

Multiplexer : 1.0000

Dilution 1.0000

Use Multiplier & Dilution Factor with ISTDS

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU]	Height [mAU]	Area %
1	14.388	MM	0.3734	1.43203e4	639.37732	98.1776
2	20.773	MM	0.4779	265.93488	9.27401	1.8224

Totals : 1.45922e4 648.65133

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

***** End of Report *****

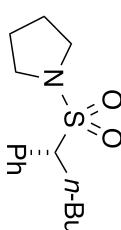


Table 2, entry 4 with (S,S)-L1

=====
 Injection Date : 1/17/2013 6:42:19 PM
 Sample Name : JC9155
 Location : Vial 3
 Acq. Operator : CE
 Acq. Instrument : Instrument 1
 Inj : 1
 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-03-30.M
 Last changed : 12/31/2012 2:58:25 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 15:02:27 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201301~1\JC9155.D)



mAU

DAD1, Sig=250,10 Ref=360,100 (201301~1\JC9155.D)

mAU

=====
 Area Percent Report
 =====

O
O
N
S
Ph
n-Bu

Table 2, entry 5
with (*R*,*R*)-L1

Signal 2: DAD1, B, Sig=254,10 Ref=360,100

Signal 3: DAD1, C, Sig=210,10 Ref=360,100

Signal 4: DAD1, D, Sig=230,10 Ref=360,100

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.872	MM	0.45172	9.29207e4	1080.68958	98.9997
2	16.710	MM	0.4686	295.95520	10.52571	1.0003

Totals : 2.95866e4 1091.21529

Results obtained with enhanced integrator!

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.873	MM	0.43615	55.96.80615	213.91138	99.0331
2	16.712	MM	0.4778	54.64594	1.90619	0.9669

Totals : 55.1.45210 215.81756

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====

Data File C:\HPCHEM\1\DATA\201212-1\JC9139B1.D

Sample Name: JC9139B

Data File C:\HPCHEM\1\DATA\201212-1\JC9139B1.D

Sample Name: JC9139B

=====
 Injection Date : 12/31/2012 3:20:37 PM Seq. Line : 2
 Sample Name : JC9139B Location : Vial 3
 Acq. Operator : JC9139B Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-03-30.M
 Last changed : 12/31/2012 2:58:25 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/1/2014 11:17:34 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9139B1.D)



mAU

10

8

6

4

2

0

5

7.5

10

12.5

15

17.5

20

22.5

25

min

DAD1, Sig=254,10 Ref=360,100 (2012-1\JC9139B1.D)

mAU

12.5

10

7.5

5

2.5

0

5

7.5

10

12.5

15

17.5

20

22.5

25

min

DAD1, Sig=254,10 Ref=360,100 (2012-1\JC9139B1.D)

mAU

200

0

5

10

12.5

15

17.5

20

22.5

25

min

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9139B1.D)

mAU

120

100

80

60

40

20

0

5

7.5

10

12.5

15

17.5

20

22.5

25

min

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9139B1.D)

mAU

1

0.8

0.6

0.4

0.2

0

5

7.5

10

12.5

15

17.5

20

22.5

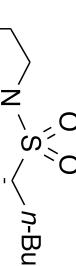
25

min

=====
 Injection Date : 12/31/2012 3:20:37 PM Seq. Line : 2
 Sample Name : JC9139B Location : Vial 3
 Acq. Operator : JC9139B Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-03-30.M
 Last changed : 12/31/2012 2:58:25 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/1/2014 11:17:34 PM by MK
 (modified after loading)

=====
 Signal 2: DAD1, Sig=254,10 Ref=360,100
 Signal 3: DAD1, C, Sig=210,10 Ref=360,100
 Signal 1: DAD1, A, Sig=250,10 Ref=360,100
 Signal 2: DAD1, B, Sig=254,10 Ref=360,100
 Signal 3: DAD1, C, Sig=210,10 Ref=360,100
 Signal 4: DAD1, D, Sig=230,10 Ref=360,100
 Signal 5: DAD1, E, Sig=280,10 Ref=360,100
 Results obtained with enhanced integrator!

Table 2, entry 5
with (S,S)-L1



=====
 **** End of Report ****

Data File C:\HPCHEM\1\DATA\201305~1\JCX093A.D

Sample Name: JC10093A

Data File C:\HPCHEM\1\DATA\201305~1\JCX093A6.D

Sample Name: JC10093M

===== 5/19/2013 5:16:09 PM Seq. Line : 2
Injection Date : 5/19/2013 5:16:09 PM Location : Vial 41
Sample Name : JC10093A

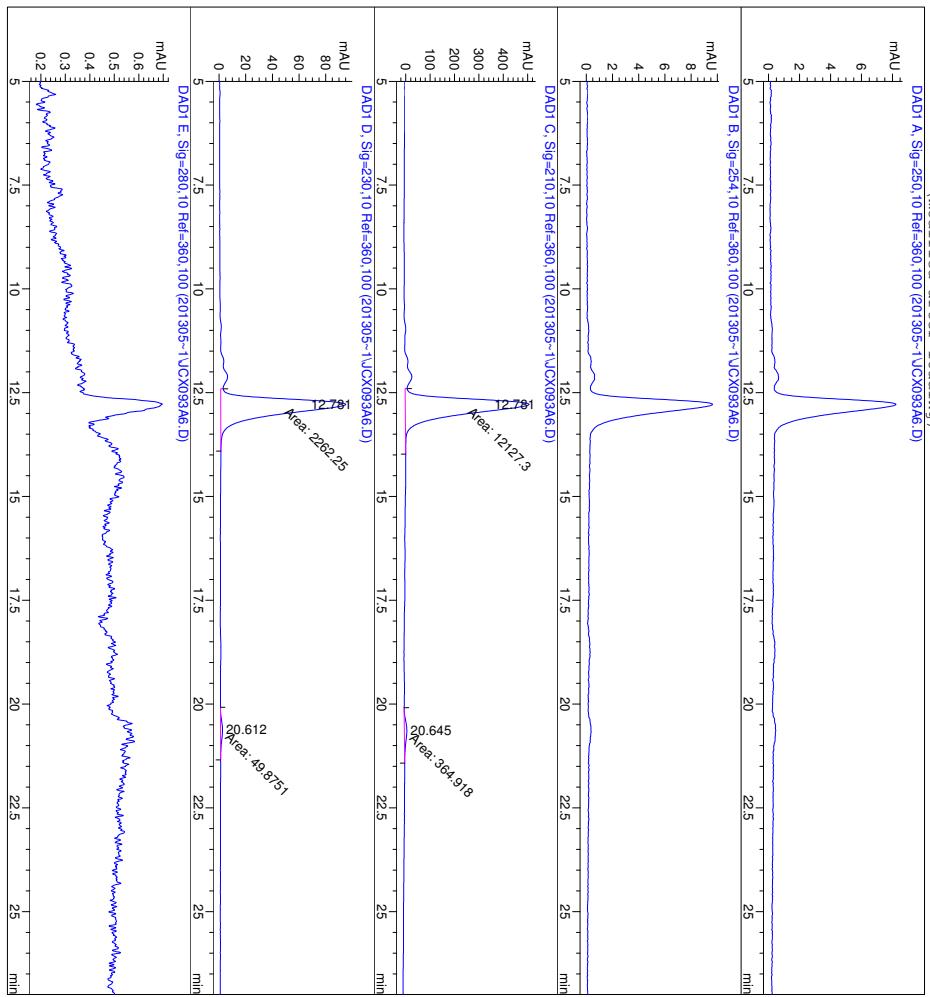
Acq. Instrument : Instrument 1 Inj Volume : 15 µl
Different Inj Volume from Sequence ! Actual Inj Volume : 5 µl
Acq. Method : C:\HPCHEM1\METHODS\0D-01-40.M
Last changed : 12/18/2012 2:31:36 PM by CE

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDss

DAD1 A, Sig=250,10 Rel=360,100 (201305~1)UCX093A6.D
.....
(modified after loading)

Signal 1: DAD1 A, Sig=250, 10 Ref=360, 1000

Table 2 entry 6



```

Totals :          1.24922e4    518.86157

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100
          Peak Retiming Type   Width      Area      Height     Area
          # [min]           [min] [mAU/s] [mAU] %-----|-----|
          -----+-----+-----+-----+-----+-----+
          1 12.781 FM       0.4017 2262.25317 93.85459 97.8429
          2 20.612 MM       0.5479 49.87514 1.51723 2.1571
          -----+-----+-----+-----+-----+-----+
Totals :          2312.12832 95.37182

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100
          =====

```

Table 2, entry 6

S-100

Data File C:\HPCHEM\1\DATA\201305-1\JCX093B6.D

Sample Name: JC10093B

Data File C:\HPCHEM\1\DATA\201305-1\JCX093B6.D

Sample Name: JC10093B

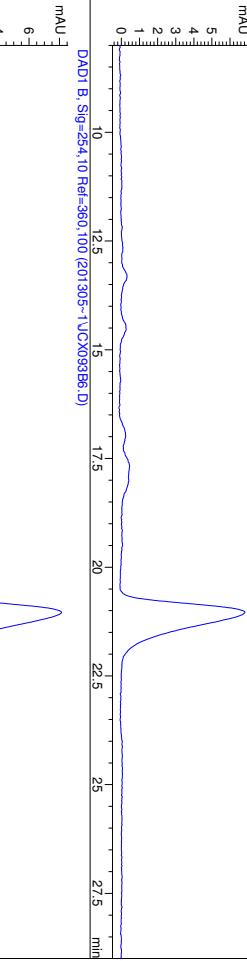
=====
Injection Date : 5/19/2013 5:57:27 PM Seq. Line : 3
Sample Name : JC10093B Location : Vial 42
Acq. Operator : CE Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
Acq. Method : C:\HPCHEM\1\METHODS\OD-01-40.M Dilution
Last changed : 12/18/2012 2:31:36 PM by CE
Analysis Method : C:\HPCHEM\1\METHODS\VA005-40.M
Last changed : 8/2/2014 9:01:58 PM by MK
(modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201305-1\JCX093B6.D)

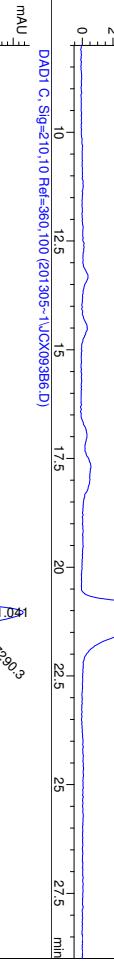
mAU



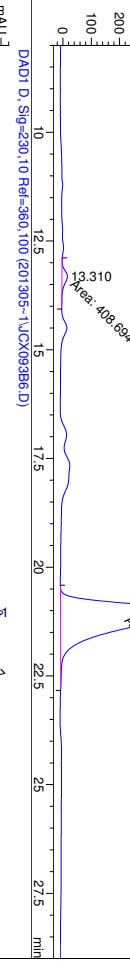
mAU



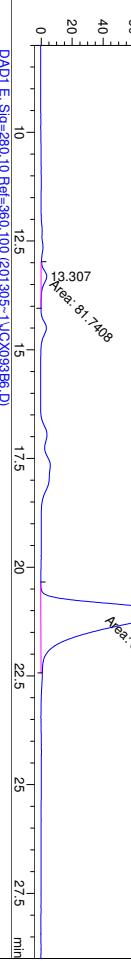
mAU



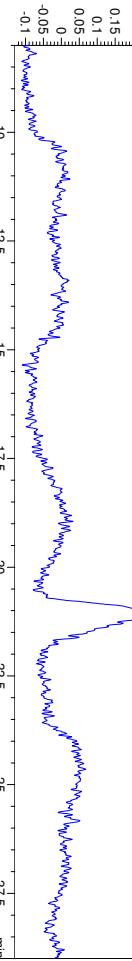
mAU



mAU



mAU



mAU

=====
Area Percent Report
=====

O O
 $\backslash \backslash$
 $\backslash \backslash$
Me₂N—S—C(=O)₂Ph

Table 2, entry 6
with (S,S)-L1

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDS

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.310	MM	0.3540	408.69366	19.24129	2.3091
2	21.041	MM	0.6492	1.72903e4	443.91245	97.6909

Totals : 1.76990e4 463.15373

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.307	FM	0.3703	81.74083	3.67898	2.4967
2	21.041	MF	0.6541	319.27197	81.33752	97.5033

Totals : 3274.01280 85.01650

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====
*** End of Report ***
=====

Data File C:\HPCHEM\1\DATA\201310~1\PM2-21A1.D

Sample Name: PM2-21A

Data File C:\HPCHEM\1\DATA\201310~1\PM2-21A1.D

Sample Name: PM2-21

===== 10/23/2013 1:57:18 PM ===== Seq. Line : 2 T o c + : v1.13

===== Area Percent Report =====

Acq.	Instrument	Instrument 1
Different	Inj	Volume from Sequence !
Acq.	Method	C:\HPCHEM\1\METHODS\OD-02-30.M
Last changed		9/13/2013 12:42:31 PM by MK

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=250, 10 Ref=360, 100

mAU

mAU

$$\text{Me}_2\text{N} \sim \text{S} \begin{array}{c} \backslash \\ / \end{array} \text{C} \begin{array}{c} \backslash \\ / \end{array} \text{O} \text{O}$$

Ph

OTBS

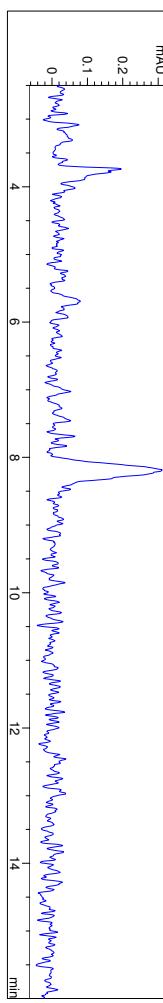
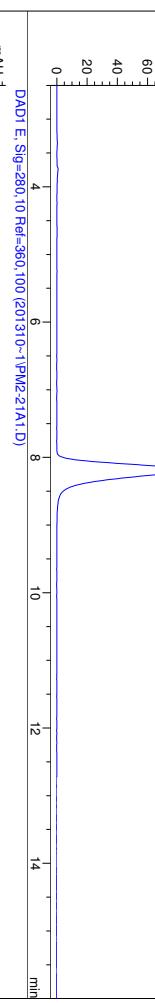
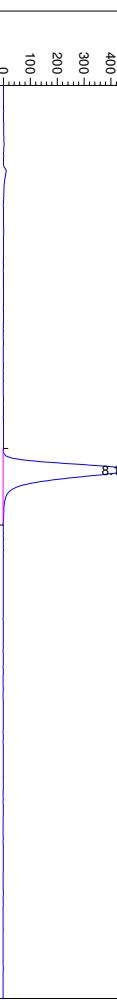
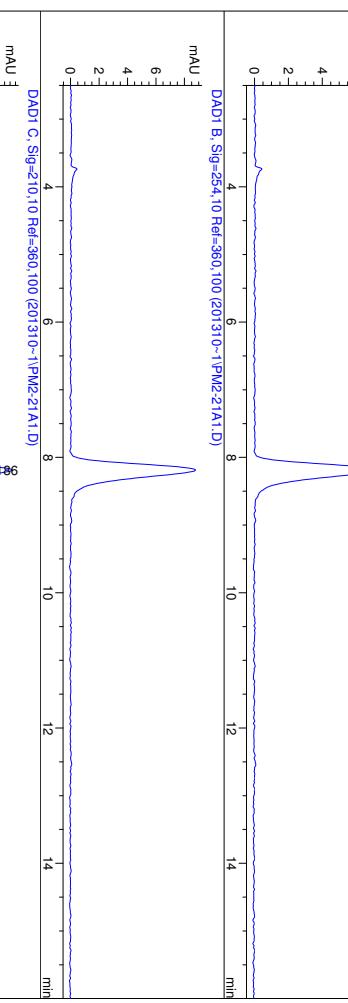


Table 2, entry 7
with (R,R) -L1

Results obtained with enhanced integrator!

Signal 4: BABI B, Sig=230, 10 Ret=360, 100

Signal 5: DAD1 E, Sig=280, 10 Ref=360, 100

— — — — —

S-102

Data File C:\HPCHEM\1\DATA\201310-1\PM2-21B1.D

Sample Name: PM2-21B

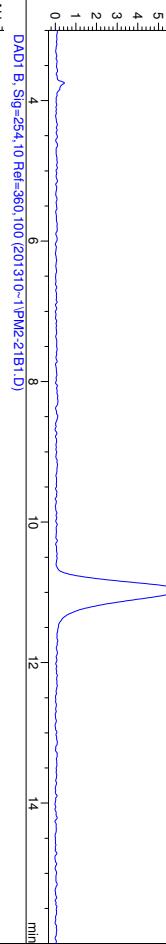
Sample Name: PM2-21B

=====
 Injection Date : 10/23/2013 2:28:36 PM
 Sample Name : PM2-21B
 Location : Vial 14
 Acq. Operator : MK
 Inj. Inj. : 1
 Acq. Instrument : Instrument 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\VOB-02-30.M
 Last changed : 9/13/2013 12:42:31 PM by MK
 Analysis Method : C:\HPCHEM\1\METHODS\A005-40.M
 Last changed : 8/1/2014 11:25:57 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201310-1\PM2-21B1.D)

mAU

5
4
3
2



10
9
8
7
6
5
4
3
2

min

DAD1 B, Sig=254,10 Ref=360,100 (201310-1\PM2-21B1.D)

mAU

6
5
4
3
2

10
9
8
7
6
5
4
3
2

min

DAD1 C, Sig=210,10 Ref=360,100 (201310-1\PM2-21B1.D)

mAU

6
5
4
3
2

10
9
8
7
6
5
4
3
2

min

DAD1 D, Sig=230,10 Ref=360,100 (201310-1\PM2-21B1.D)

mAU

300
200
100
0

10
9
8
7
6
5
4
3
2

min

DAD1 E, Sig=280,10 Ref=360,100 (201310-1\PM2-21B1.D)

mAU

50
40
30
20
10
0

10
9
8
7
6
5
4
3
2

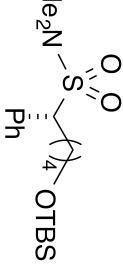
min

Data File C:\HPCHEM\1\DATA\201310-1\PM2-21B1.D

Sample Name: PM2-21B

=====
 Area Percent Report
 =====
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS



Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU]	Height [mAU]	Area %
1	8.233	MM	0.2711	55.66857	3.42192	0.7273
2	10.969	MM	0.3284	7598.58594	385.60574	99.2727

Totals : 7654.25451 389.02787

Results obtained with enhanced integrator!

=====
 *** End of Report ***
 =====

Table 2, entry 7
with (S,S)-L1

Data File C:\HPCHEM\1\DATA\201302-1\JC9221A1.D

Sample Name: JC9221A

Data File C:\HPCHEM\1\DATA\201302-1\JC9221A1.D

Sample Name: JC9221A

=====
Injection Date : 2/22/2013 11:28:27 AM Seq. Line : 2
Sample Name : JC9221A Location : Vial 9
Acq. Operator : CE Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
Acq. Method : C:\HPCHEM\1\METHODS\OD-05-40.M
Last changed : 4/7/2011 5:39:35 PM by CC
Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
Last changed : 7/30/2014 5:09:46 PM by MK
(modified after loading)

=====
Signal Line : 2
Location : Vial 9
Inj : 1
Inj Volume : 15 μ l
Actual Inj Volume : 5 μ l
=====

=====
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDS

DAD1, Sig=250,10 Ref=360,100 (201302-1\JC9221A1.D)

DAD1, Sig=254,10 Ref=360,100 (201302-1\JC9221A1.D)

Signal 2: DAD1, Sig=254,10 Ref=360,100

Signal 3: DAD1, C, Sig=210,10 Ref=360,100

Signal 1: DAD1, A, Sig=250,10 Ref=360,100

Totals :

1,35859e4 399.23533

Results obtained with enhanced integrator!

Signal 4: DAD1, D, Sig=230,10 Ref=360,100

Peak RetTime Type Width Area Height Area %

[min] [min] [mAU*s] [mAU] [mAU]

1 18.257 MM 0.5628 1.2986e4 382.84482 95.1621

2 22.785 MM 0.6684 657.27887 16.39051 4.8379

Totals :

1,20901e4 356.56774

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

=====
**** End of Report ****

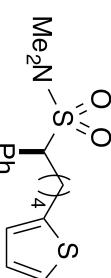
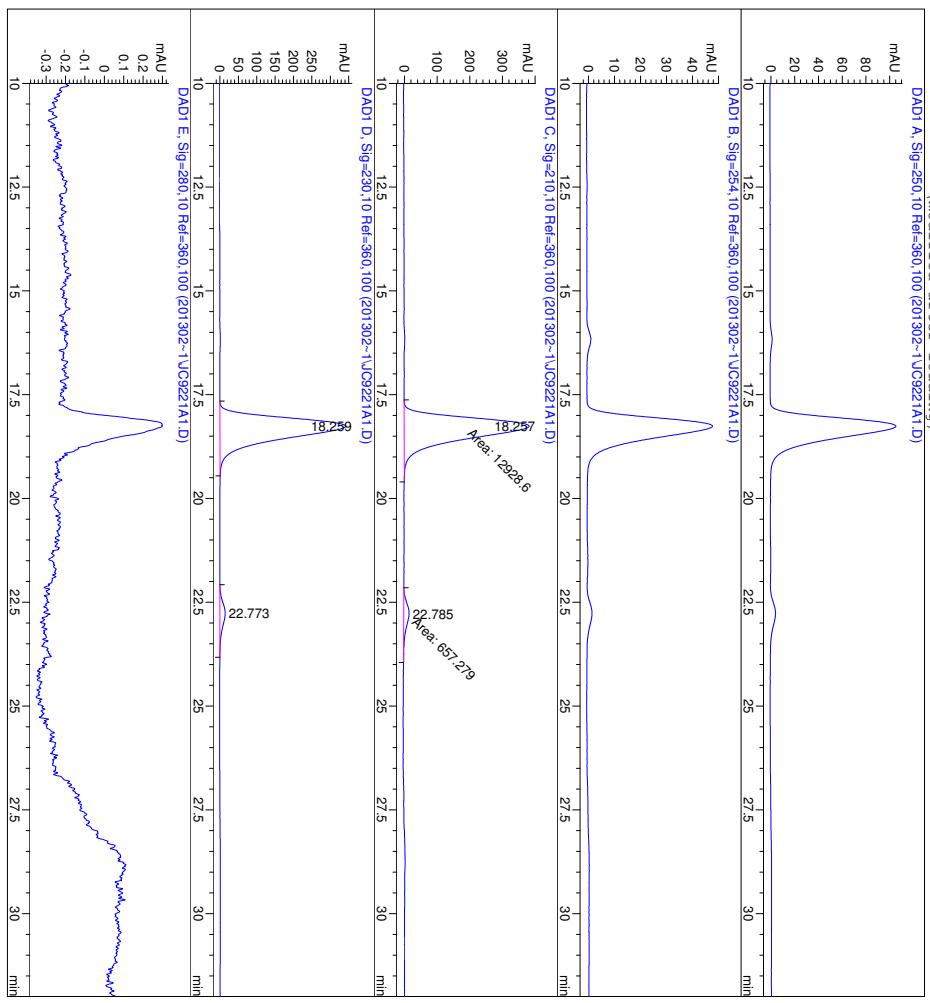


Table 2, entry 8
with (R,R)-L1



Data File C:\HPCHEM\1\DATA\201302~1\JC9221B1.D

Sample Name: JC9221B

Data File C:\HPCHEM\1\DATA\201302~1\JC9221B1.D

Sample Name: JC9221H

Injection Date : 2/22/2013 12:09:44 PM Seq. Line : 3
Run Time : 00:00:00 Job No. : 10

Area Percent Report

Sample Name : 000221
Acq. Operator : CE
Acq. Instrument : Instrument 1
Different Inj Volume from Sequence ! Actual Sequence
Acq. Method : C:\HPCHEM1\METHODS\OD-05-40.MCC
Last changed : 7/27/2011 5:39:35 PM by CC

Sorted By	:	Signal
Multiplier	:	1.0000
Dilution	:	1.0000
Use Multiplier & Dilution Factor with ISTDs	:	

$$\text{Mn}(\text{N}_\text{3})_\text{2}\text{S}_\text{2}\text{O}_\text{8}$$

(modified after loading)
DAD1 A, Sig=250,10 Ref=360,100 (201302-1\UC9221B1.D)

Signal 1: DAD1 A, Sig=250, 10 Ref=360, 100

Signal 2: DAD1 B, Sig=254, 10 Ref=360, 100

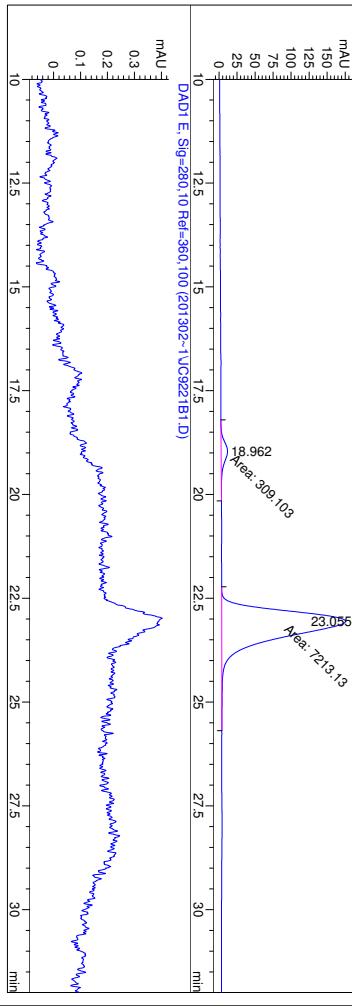
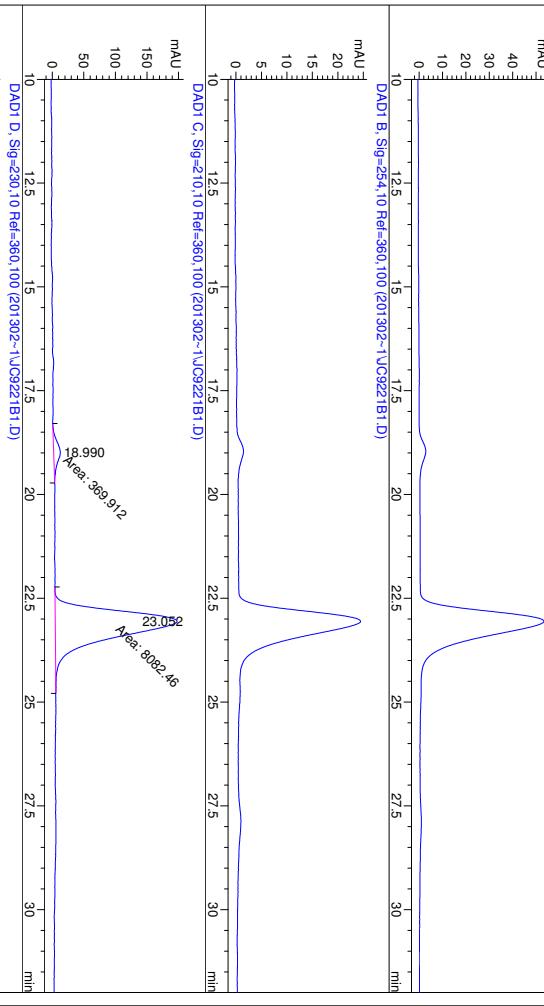


Table 2, entry 8
with (S,S) -L1

S-105

Data File C:\HPCHEM\1\DATA\GROUP\JC12043A.D

Sample Name: JC12043A

Sample Name: JC12043A

=====
 Injection Date : 6/16/2014 7:47:41 PM
 Sample Name : JC12043A
 Location : Vial 11
 Acq. Operator : MK
 Inj. Inj. : 1
 Acq. Instrument : Instrument 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 6 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01-30.M
 Last changed : 6/16/2014 7:51:29 PM by MK
 (modified after loading)
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 6:15:41:33 PM by MK
 (modified after loading)

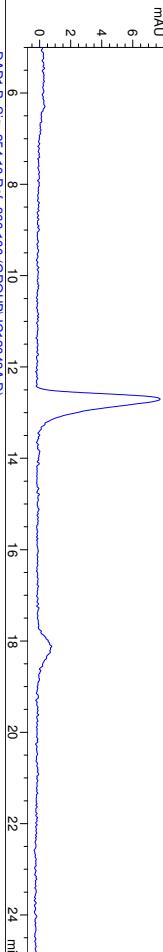
=====
 Sample Name: JC12043A

=====
 Data File C:\HPCHEM\1\DATA\GROUP\JC12043A.D
 =====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS

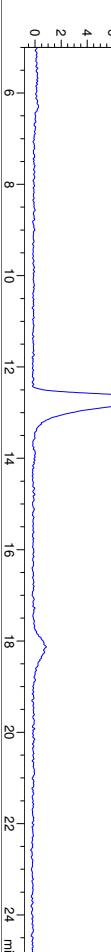
Signal 1: DAD1 A, Sig=250,10 Ref=360,100
 (modified after loading)

DAD1 A, Sig=250,10 Ref=360,100 (GROUP\JC12043A.D)



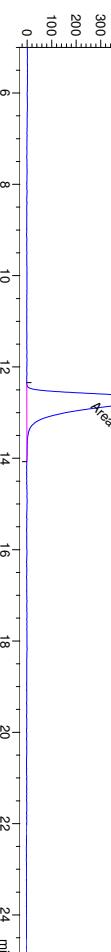
mAU

DAD1 B, Sig=254,10 Ref=360,100 (GROUP\JC12043A.D)



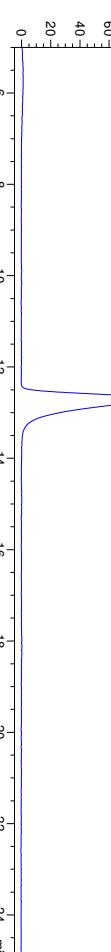
mAU

DAD1 C, Sig=210,10 Ref=360,100 (GROUP\JC12043A.D)



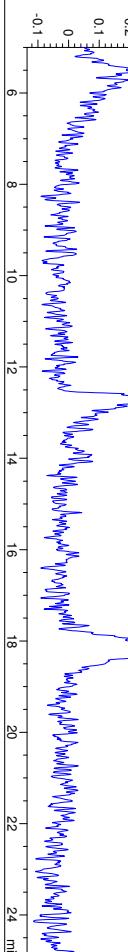
mAU

DAD1 D, Sig=230,10 Ref=360,100 (GROUP\JC12043A.D)



mAU

DAD1 E, Sig=280,10 Ref=360,100 (GROUP\JC12043A.D)



mAU

12.700

Area: 11488.9

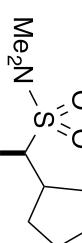


Table 2, entry 9
with (R,R)-L1

Peak #	Retention Time [min]	Width [min]	Area [mAU]	Height [mAU]	Area %
1	12.700	0.3740	1.14889e4	512.01752	100.0000

Totals :

1.14889e4 512.01752

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

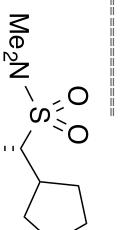
===== *** End of Report *** =====

=====
 Injection Date : 6/16/2014 8:19:00 PM
 Sample Name : JC12043B
 Location : Vial 12
 Acq. Operator : MK
 Inj. Inj. : 1
 Acq. Instrument : Instrument 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 6 μ l
 Last changed : 6/16/2014 7:51:29 PM by MK
 (modified after loading)
 Analysis Method : C:\HPCHEM\1\METHODS\OD-01-30.M
 Last changed : 8/1/2014 11:32:53 PM by MK
 (modified after loading)

=====
 Seq. Line : 5
 Location : Vial 12
 Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS

=====
 Signal 1: DAD1 A, Sig=250,10 Ref=360,100 (GROUP\JC12043B.D)
 mAU
 4
 3
 2
 1
 0
 6 8 10 12 14 16 18 20 22 24 min
 DAD1 B, Sig=254,10 Ref=360,100 (GROUP\JC12043B.D)

=====
 Signal 2: DAD1 B, Sig=254,10 Ref=360,100
 Signal 3: DAD1 C, Sig=210,10 Ref=360,100
 mAU
 5
 4
 3
 2
 1
 0
 6 8 10 12 14 16 18 20 22 24 min
 DAD1 C, Sig=210,10 Ref=360,100 (GROUP\JC12043B.D)



=====
 Signal 4: DAD1 D, Sig=230,10 Ref=360,100
 mAU
 250
 200
 150
 100
 50
 0
 6 8 10 12 14 16 18 20 22 24 min
 DAD1 D, Sig=230,10 Ref=360,100 (GROUP\JC12043B.D)

=====
 Signal 5: DAD1 E, Sig=280,10 Ref=360,100
 mAU
 40
 30
 20
 10
 0
 6 8 10 12 14 16 18 20 22 24 min
 DAD1 E, Sig=280,10 Ref=360,100 (GROUP\JC12043B.D)

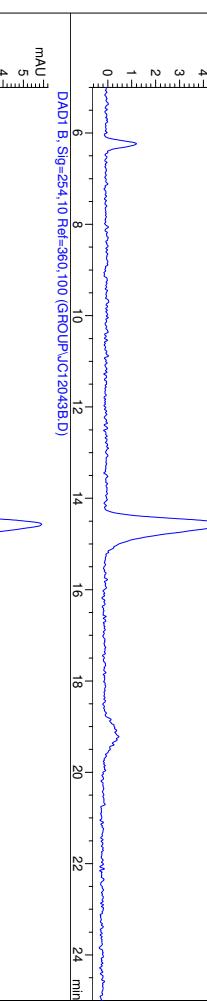


Table 2, entry 9
with (S,S)-L1

=====
 Injection Date : 12/7/2012 3:31:19 PM
 Sample Name : JC911A
 Location : Vial 35
 Acq. Operator : CE
 Acq. Instrument : Instrument 1
 Inj : 1
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-05-40.M
 Last changed : 4/7/2011 5:39:35 PM by CC
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 6:56:15 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC911A.D)

DAD1 A, Sig=250,10 Ref=360,100 (2012-1\JC911A.D)

mAU

2.5
2
1.5
1
0.5
0

12 14 16 18 20 22 24 26 min

DAD1 B, Sig=254,10 Ref=360,100 (2012-1\JC911A.D)

DAD1 B, Sig=254,10 Ref=360,100 (2012-1\JC911A.D)

mAU

3
2
1
0.5
0

12 14 16 18 20 22 24 26 min

DAD1 C, Sig=210,10 Ref=360,100 (2012-1\JC911A.D)

DAD1 C, Sig=210,10 Ref=360,100 (2012-1\JC911A.D)

mAU

10
12
14
16
18
20
22
24
26 min

DAD1 D, Sig=230,10 Ref=360,100 (2012-1\JC911A.D)

DAD1 D, Sig=230,10 Ref=360,100 (2012-1\JC911A.D)

mAU

150
100
50
0

12 14 16 18 20 22 24 26 min

DAD1 E, Sig=280,10 Ref=360,100 (2012-1\JC911A.D)

DAD1 E, Sig=280,10 Ref=360,100 (2012-1\JC911A.D)

mAU

10
12
14
16
18
20
22
24
26 min

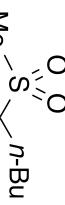


Table 3, entry 1
with (*R,R*)-L1

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

Totals :

5951.13913 214.59150

Results obtained with enhanced integrator!

***** End of Report *****

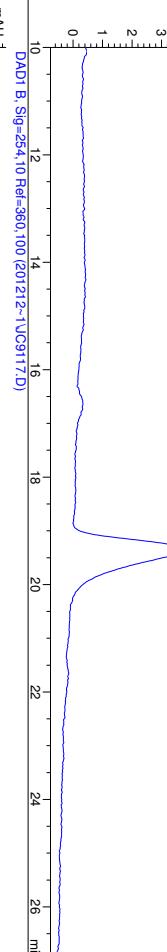
=====
 Injection Date : 12/14/2012 4:31:29 PM Seq. Line : 2
 Sample Name : JC9117 Location : Vial 21
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-05-40.M
 Last changed : 4/7/2011 5:39:35 PM by CC
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/1/2014 11:34:56 PM by MK
 (modified after loading)
 =====

DAD1 A, Sig=250,10 Ref=360,100 (2012-1\JC9117.D)



Signal 2: DAD1 B, Sig=254,10 Ref=360,100
Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Table 3, entry 1
with (*S,S*)-L1

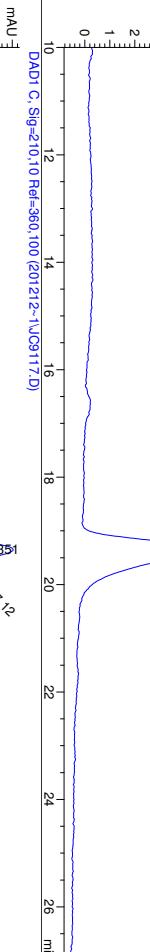


Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.679	MM	0.4290	240.29202	9.33470	2.9100
2	19.351	MM	0.5532	8017.11914	241.52242	97.0900

Totals : 8257.4116 250.85712

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100



=====
 Signal 5: DAD1 E, Sig=280,10 Ref=360,100
 =====

*** End of Report ***



S-109

Data File C:\HPCHEM\1\DATA\201307-1\JC10185.D

Sample Name: JC10185

Data File C:\HPCHEM\1\DATA\201307-1\JC10185.D

Sample Name: JC10185

=====
 Injection Date : 7/20/2013 9:37:03 AM Seq. Line : 34
 Sample Name : JC10185 Location : Vial 82
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AD-04-30.M
 Last changed : 11/29/2010 7:04:08 PM by JTM
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 6:15:59:22 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201307-1\JC10185.D)

mAU



Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU]	Height [mAU]	Area %
1	16.698	MM	0.2409	22.95708	1.58850	0.2416
2	25.782	MM	0.5562	9479.26074	284.07181	99.7584

Totals : 9502.21782 285.66031

Results obtained with enhanced integrator!

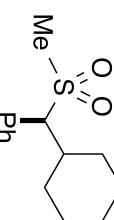
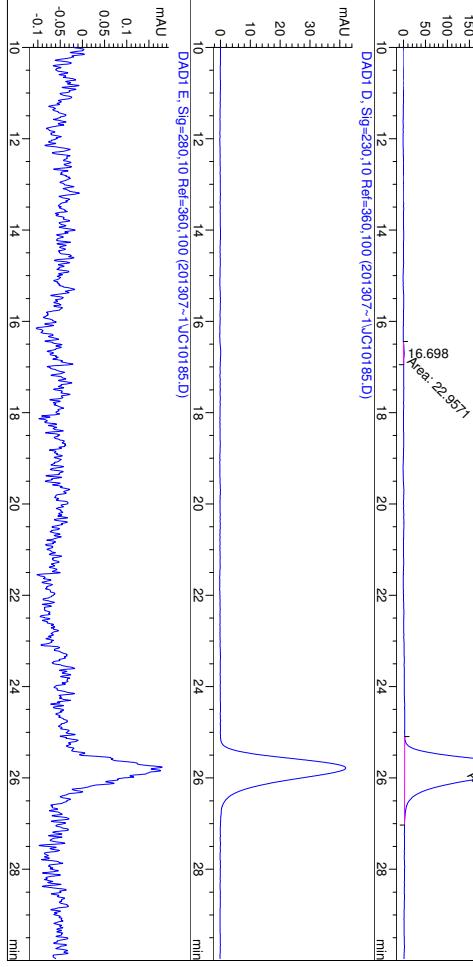
Signal 4: DAD1 D, Sig=230,10 Ref=360,100

mAU



Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====

Table 3, entry 2
with (R,R)-L1

mAU

min

Data File C:\HPCHEM\1\DATA\201307-1\JC10119.D

Sample Name: JC10119B

Data File C:\HPCHEM\1\DATA\201307-1\JC10119.D

Sample Name: JC10119B

Injection Date : 7/20/2013 9:05:45 AM Seq. Line : 33
Sample Name : JC10119B Location : Vial 81
Acq. Operator : CE Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
Acq. Method : C:\HPCHEM\1\METHODS\AD-04-30.M
Last changed : 11/29/2010 7:04:08 PM by JTM
Analysis Method : C:\HCCEM\1\METHODS\AD00540.M
Last changed : 8/1/2014 11:37:12 PM by MK
(modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201307-1\JC10119.D)

DAD1 A, Sig=250,10 Ref=360,100 (201307-1\JC10119.D)

mAU

8

6

4

2

0

10

12

14

16

18

20

22

24

26

28

min

DAD1 B, Sig=254,10 Ref=360,100 (201307-1\JC10119.D)

mAU

8

6

4

2

0

10

12

14

16

18

20

22

24

26

28

min

DAD1 C, Sig=210,10 Ref=360,100 (201307-1\JC10119.D)

mAU

10

8

6

4

2

0

10

12

14

16

18

20

22

24

26

28

min

DAD1 D, Sig=230,10 Ref=360,100 (201307-1\JC10119.D)

mAU

500

400

300

200

100

0

10

12

14

16

18

20

22

24

26

28

min

DAD1 E, Sig=280,10 Ref=360,100 (201307-1\JC10119.D)

mAU

60

40

20

0

10

12

14

16

18

20

22

24

26

28

min



Table 3, entry 2
with (S,S)-L1

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.579	BB	0.3588	2140.25293	90.87717	100.0000

Totals : 2140.25293

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

***** End of Report ***

Data File C:\HPCHEM\1\DATA\201307~1\JC10177.D

595

Data File C:\HPCHEM\1\DATA\201307~1\JC10177.D

Sample Name: JC1017

=====
Injection Date : 7/11/2013 9:50:27 AM Seg. Line : --- 2
=====

```

Acq. Operator : CE
Acq. Instrument : Instrument 1
Different Inj volume from Sequence !
Acq. Method : C:\HCCHEM\METHODS\NOD-20-60.M
Last changed : 6/28/2013 4:31:36 PM by CE

```

Sorted By	:	Signal
Multiplier	:	1.0000
Dilution	:	1.0000
Use Multiplier & Dilution Factor with ISTDS		

$$\text{Me}-\text{S}(\text{O})-\text{CH}_2-\text{CH}_2-\text{N}(\text{Bn})\text{Cbz}$$

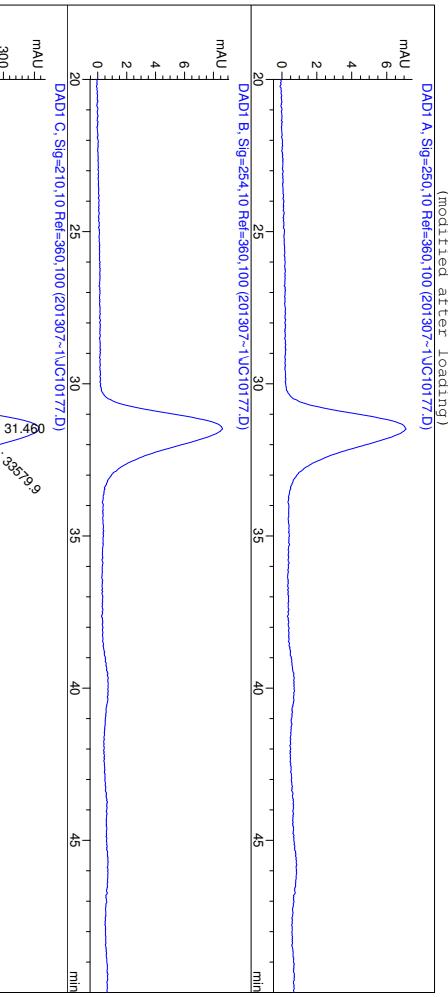
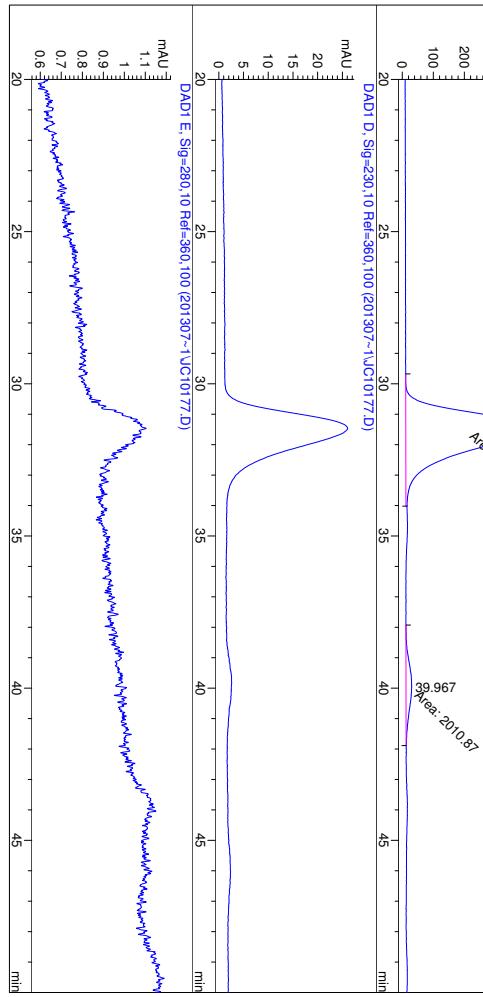


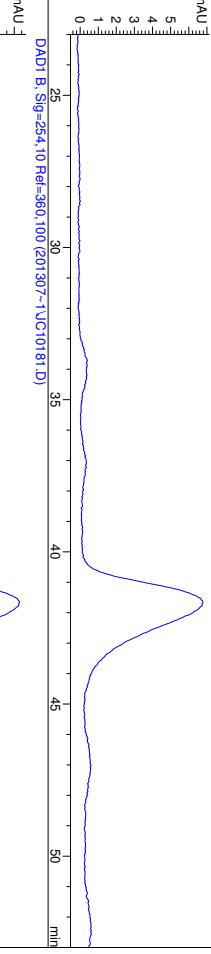
Table 3, entry 3
with (R,R) -L1



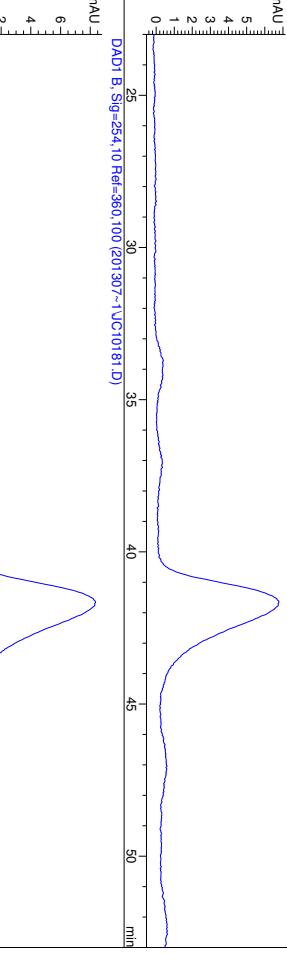
Instrument 1 7/30/2014 7:02:26 PM MF

Injection Date : 7/20/2013 7:21:29 AM Seq. Line : 30
 Sample Name : JC10181 Location : Vial 10
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\VOB-20-60.M
 Last changed : 6/28/2013 4:31:36 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AD00540.M
 Last changed : 8/1/2014 11:39:47 PM by MK
 (modified after loading)

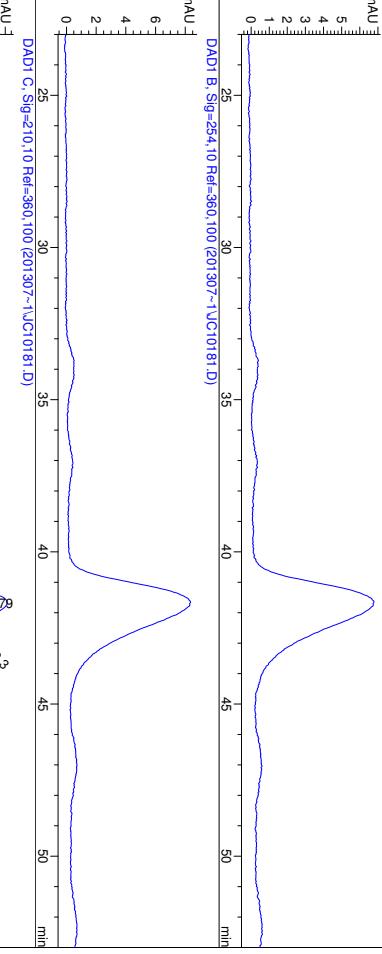
DAD1 A, Sig=250,10 Ref=360,100 (201307-1\JC10181.D)



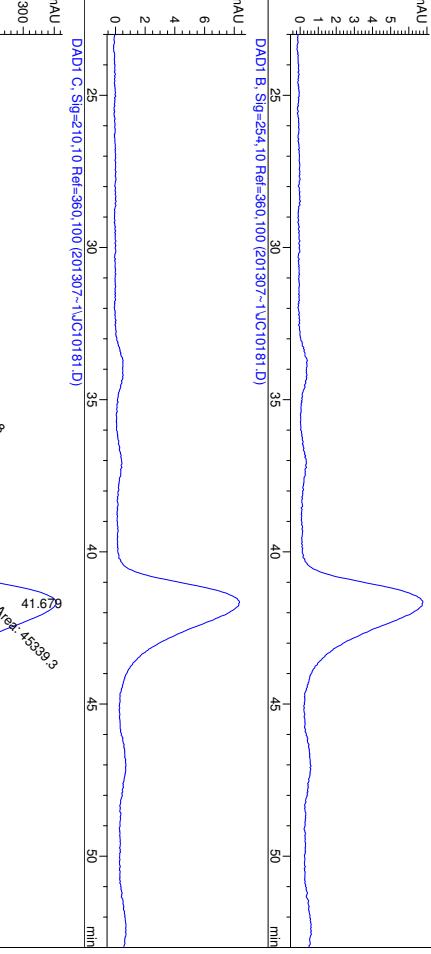
DAD1 B, Sig=254,10 Ref=360,100 (201307-1\JC10181.D)



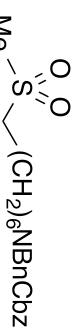
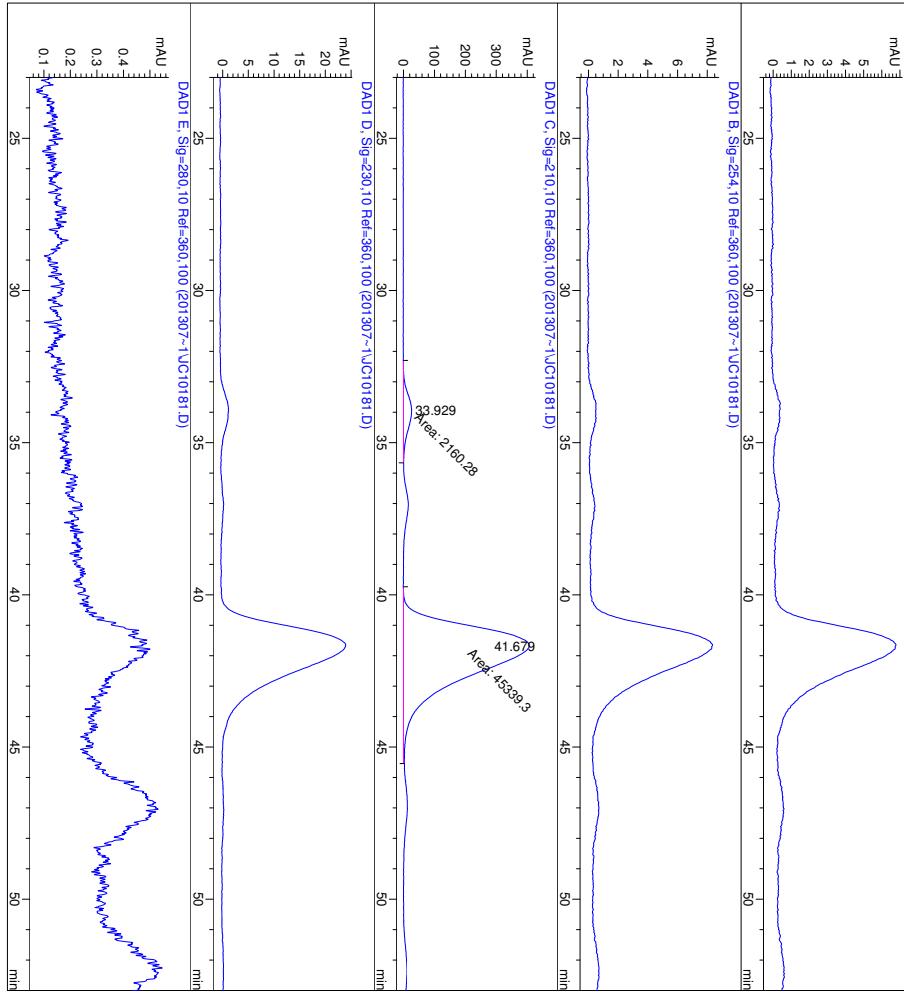
DAD1 C, Sig=210,10 Ref=360,100 (201307-1\JC10181.D)



DAD1 D, Sig=230,10 Ref=360,100 (201307-1\JC10181.D)



DAD1 E, Sig=280,10 Ref=360,100 (201307-1\JC10181.D)

Table 3, entry 3
with (S,S)-L1

Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %
 1 33.929 MF 1.3461 21.60.27637 26.74717 4.5480
 2 41.679 MF 1.8592 4.53393e4 406.43207 95.4520

Totals : 4.74996e4 433.17924

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***

Data File C:\HPCHEM\1\DATA\201308-1\PM2-20A1.D

Sample Name: PM2-20A

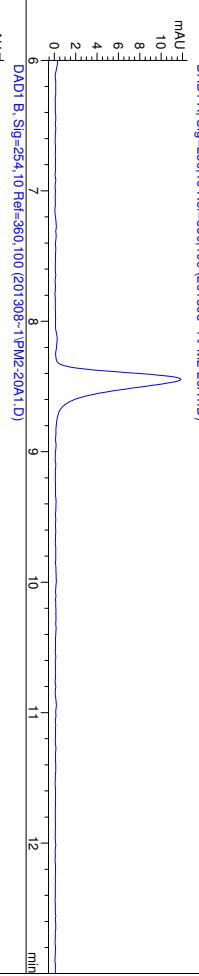
Data File C:\HPCHEM\1\DATA\201308-1\PM2-20A1.D

Sample Name: PM2-20A

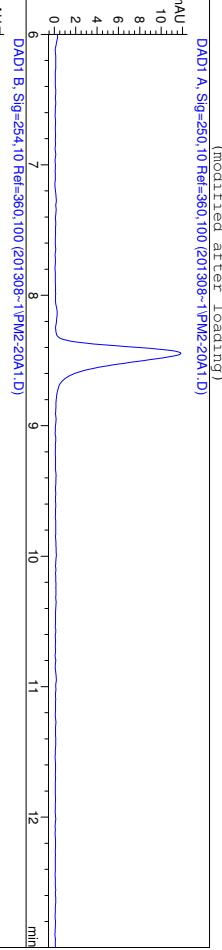
=====
Injection Date : 8/2/2013 10:41:10 AM Seq. Line : 2
Sample Name : PM2-20A Location : Vial 77
Acq. Operator : CE Inj. Inj. : 1
Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 3 μ l
Acq. Method : C:\HPCHEM\1\METHODS\IB-01-20.M
Last changed : 3/19/2012 12:43:39 PM by CE
Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
Last changed : 7/30/2014 7:04:50 PM by MK
(modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\PM2-20A1.D)

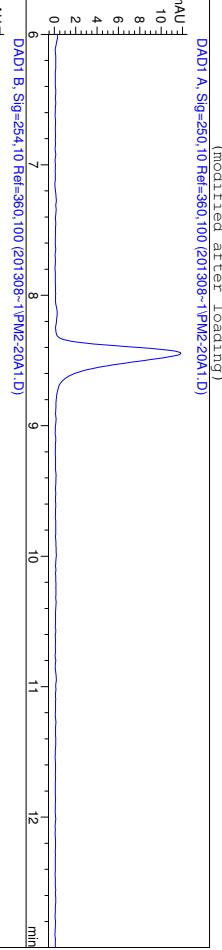
mAU



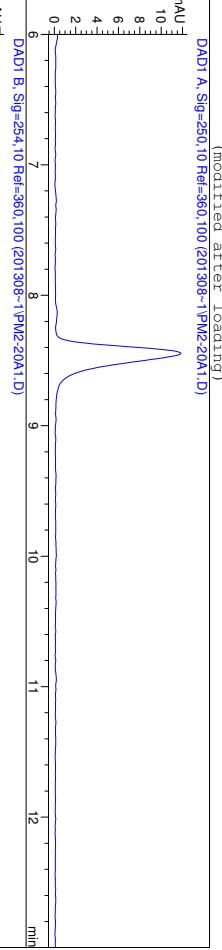
mAU



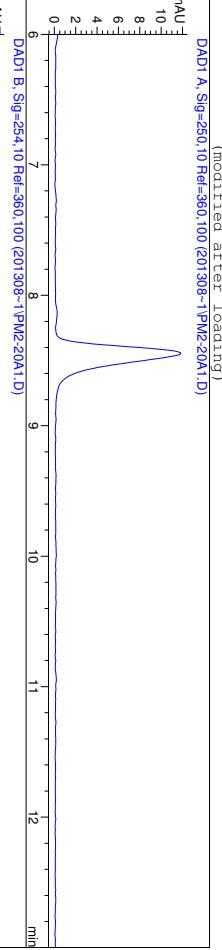
mAU



mAU



mAU



mAU

=====
Area Percent Report
=====

Sample Name: PM2-20A

Sample Name: PM2-20A

=====
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDS
=====

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Totals :

6529.72577 701.04255

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak RetTime Type Width Area Height Area %

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %	
1	8.444	FM		0.1553	252.45752	99.5957	
2	9.930	FM		0.1630	9.55209	9.7556e-1	0.4043

Totals :

2362.51303 253.43408

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====

=====
*** End of Report ***
=====

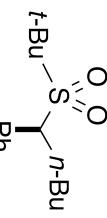


Table 3, entry 4
with (*R*,*R*)-L1

Data File C:\HPCHEM\1\DATA\201308-1\PM2-20B1.D

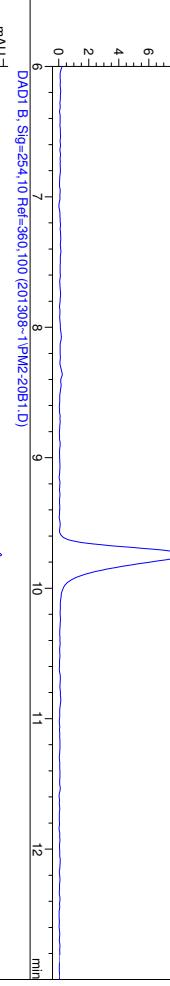
Sample Name: PM2-20B

Sample Name: PM2-20B

=====
 Injection Date : 8/2/2013 11:02:26 AM
 Sample Name : PM2-20B
 Location : Vial 78
 Acq. Operator : CE
 Acq. Instrument : Instrument 1
 Inj : 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 3 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\IB-01-20.M
 Last changed : 3/19/2012 12:43:39 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\IA005-40.M
 Last changed : 8/1/2014 11:41:35 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\PM2-20B1.D)

mAU



Data File C:\HPCHEM\1\DATA\201308-1\PM2-20B1.D

Sample Name: PM2-20B

=====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS

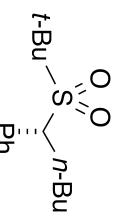


Table 3, entry 4
with (S,S)-L1

Signal 2: DAD1, Sig=254,10 Ref=360,100

Signal 3: DAD1, C, Sig=210,10 Ref=360,100

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.381	MM	0.1398	35.99226	4.28965	0.6915
2	9.743	MM	0.1683	5168.82764	511.87228	99.3085

Totals : 5204.81990 516.16193

Results obtained with enhanced integrator!

Signal 4: DAD1, D, Sig=230,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.373	VB	0.1312	22.56680	2.60208	1.2052
2	9.743	MM	0.1686	1849.8377	182.89543	98.7948

Totals : 1872.40457 185.49751

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====

Data File C:\HPCHEM\1\DATA\201212-1\JC9119A.D

Sample Name: JC9119A

Data File C:\HPCHEM\1\DATA\201212-1\JC9119A.D

Sample Name: JC9119A

=====
 Injection Date : 12/14/2012 5:34:16 PM
 Sample Name : JC9119A
 Location : Vial 22
 Acq. Operator : CE
 Inj. Inj. : 1
 Acq. Instrument : Instrument 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 3 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01.40.M
 Last changed : 8/17/2010 4:39:48 PM by JTM
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02.20.M
 Last changed : 7/30/2014 7:15:44 PM by MK
 (modified after loading)

=====

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9119A.D)

mAU

15

10

5

0

8 10 12 14 16 18 20 22 24 min

DAD1, B, Sig=254,10 Ref=360,100 (2012-1\JC9119A.D)

=====
 mAU

15

10

5

0

8 10 12 14 16 18 20 22 24 min

DAD1, C, Sig=210,10 Ref=360,100 (2012-1\JC9119A.D)

=====
 mAU

15

10

5

0

8 10 12 14 16 18 20 22 24 min

DAD1, D, Sig=230,10 Ref=360,100 (2012-1\JC9119A.D)

=====
 mAU

15

10

5

0

8 10 12 14 16 18 20 22 24 min

DAD1, E, Sig=280,10 Ref=360,100 (2012-1\JC9119A.D)

=====
 mAU

125

100

75

50

25

0

8 10 12 14 16 18 20 22 24 min

DAD1, E, Sig=280,10 Ref=360,100 (2012-1\JC9119A.D)

=====
 Results obtained with enhanced integrator!

=====
 Signal 4: DAD1, D, Sig=230,10 Ref=360,100

=====
 Peak RetTime Type Width Area Height Area %

#
 [min] [min] [mAU*s] [mAU] [%]

1 13.582 MW 0.4179 1.21576e4 484.81223 92.7100

2 18.672 MW 0.5410 955.98328 29.44996 7.2900

Totals : 1.31136e4 514.26218

=====
 Results obtained with enhanced integrator!

=====
 Signal 4: DAD1, D, Sig=230,10 Ref=360,100

=====
 Peak RetTime Type Width Area Height Area %

#
 [min] [min] [mAU*s] [mAU] [%]

1 13.582 MW 0.4166 3389.75903 155.62090 92.9020

2 18.670 MW 0.5358 297.19092 9.24424 7.0980

Totals : 4186.94995 164.86513

=====
 Results obtained with enhanced integrator!

=====
 Signal 5: DAD1, E, Sig=280,10 Ref=360,100

=====
 **** End of Report ****

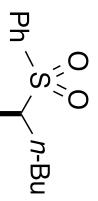


Table 3, entry 5
with (R,R) -L1

Data File C:\HPCHEM\1\DATA\201212-1\JC9119B.D

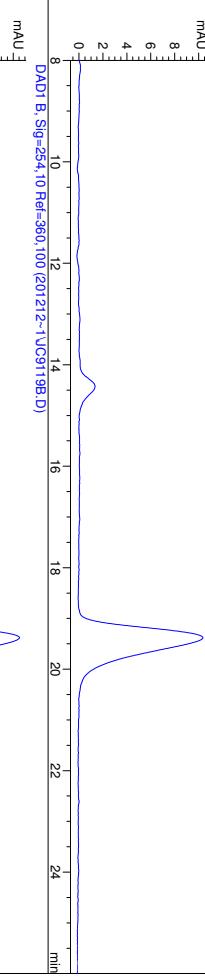
Sample Name: JC9119B

Data File C:\HPCHEM\1\DATA\201212-1\JC9119B.D

Sample Name: JC9119B

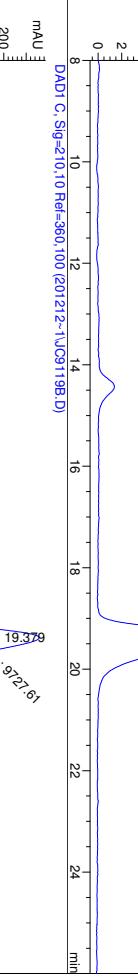
=====
 Injection Date : 12/14/2012 6:15:28 PM
 Sample Name : JC9119B
 Location : Vial 23
 Acq. Operator : CE
 Inj : 1
 Acq. Instrument : Instrument 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 3 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-0140.M
 Last changed : 8/17/2010 4:39:48 PM by JTM
 Analysis Method : C:\HPCHEM\1\METHODS\AD00540.M
 Last changed : 8/2/2014 10:20:42 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9119B.D)



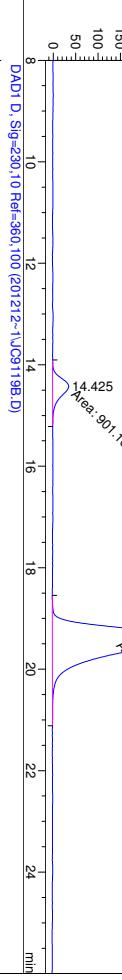
mAU

DAD1, Sig=254,10 Ref=360,100 (2012-1\JC9119B.D)



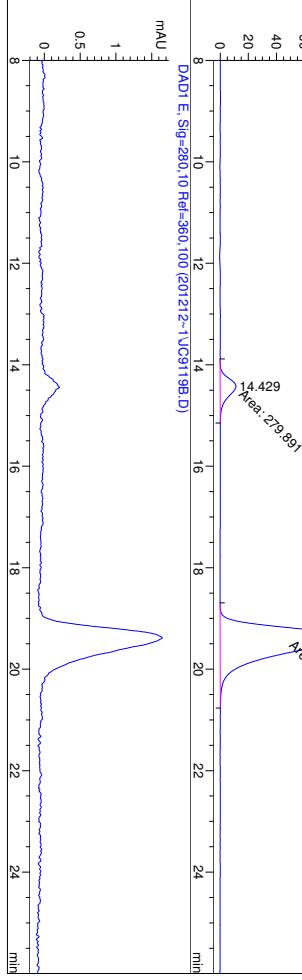
mAU

DAD1, Sig=254,10 Ref=360,100 (2012-1\JC9119B.D)



mAU

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9119B.D)



mAU

DAD1, Sig=250,10 Ref=360,100 (2012-1\JC9119B.D)



Table 3, entry 5
with (S,S)-L1

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.425	MW	0.4183	901.16510	35.90579	8.4785
2	19.379	MW	0.5770	972.60840	280.98239	91.5215

Totals : 1.06288e4 316.88818

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.429	MW	0.4146	279.89063	11.25042	8.3390
2	19.381	MW	0.5735	3076.50317	89.40532	91.6610

Totals : 3356.39380 100.65574

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

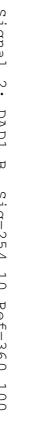
=====
 *** End of Report ***
 =====

=====
 Injection Date : 2/16/2013 5:08:58 PM
 Sample Name : JC9225
 Location : Vial 7
 Acq. Operator : CE
 Acq. Instrument : Instrument 1
 Inj : 1
 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-02-30.M
 Last changed : 2/16/2013 4:45:43 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 7:16:00 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201302-1\JC9225.D)



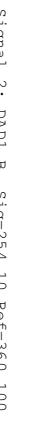
Signal 2: DAD1 B, Sig=254,10 Ref=360,100



Signal 3: DAD1 C, Sig=210,10 Ref=360,100

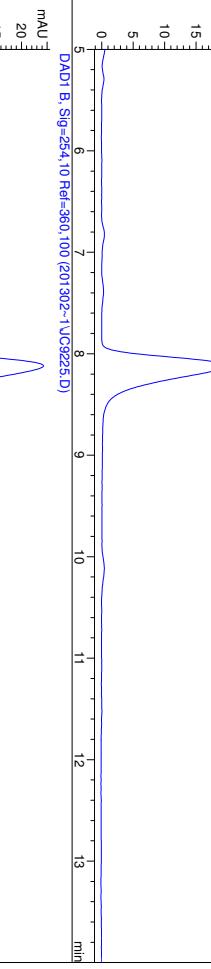


Signal 1: DAD1 A, Sig=250,10 Ref=360,100



Totals :

1.65402e4 1034.63302



Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100



Peak RetTime Type Width Area Height Area %

#	[min]	[min]	[mAU*s]	[mAU]	
1	8.119	0.2655	1.6168e4	729.36182	97.9659
2	10.121	0.2835	253.56413	14.90512	2.1361

Totals :

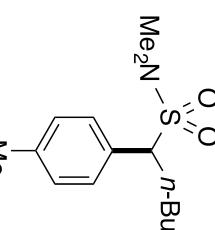
1.18703e4 744.26694

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100



=====
 *** End of Report ***
 =====



Signal 2: DAD1 B, Sig=254,10 Ref=360,100



Signal 3: DAD1 C, Sig=210,10 Ref=360,100



Signal 1: DAD1 A, Sig=250,10 Ref=360,100



Totals :

1.65402e4 1034.63302

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100



Peak RetTime Type Width Area Height Area %

#	[min]	[min]	[mAU*s]	[mAU]	
1	8.119	0.2655	1.6168e4	729.36182	97.9639
2	10.121	0.2835	253.56413	14.90512	2.1361

Totals :

1.18703e4 744.26694

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

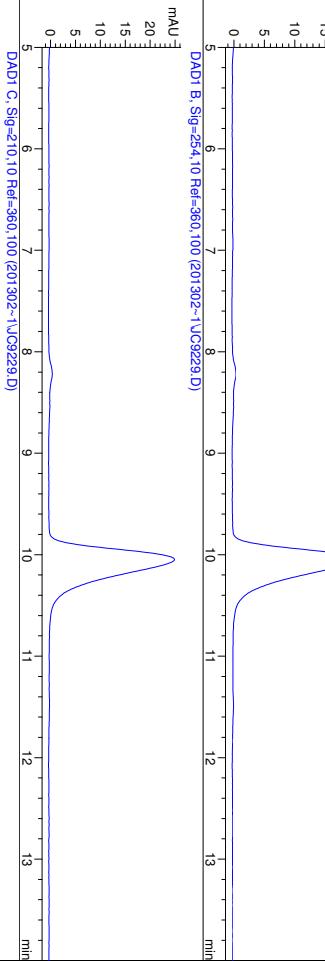


=====
 *** End of Report ***
 =====

=====
 Injection Date : 2/16/2013 5:40:14 PM Seq. Line : 3
 Sample Name : JC9229 Location : Vial 8
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-02-30.M Dilution
 Last changed : 2/16/2013 4:45:43 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AD00540.M
 Last changed : 8/1/2014 1:49:11 PM by MK
 (modified after loading)
 =====

DAD1, Sig=250,10 Ref=360,100 (201302-1\JC9229.D)

mAU

15
10
5
0

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

mAU

15
10
5
0

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

mAU

20
15
10
5
0

Totals : 2.04249e4 1092.26965

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

mAU

800
600
400
200
0

Peak RetTime Type Width Area Height Area %

#

[min]

[min]

[mAU*s]

[mAU]

%

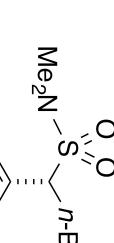
Table 4, entry 1 with (S,S)-L1					
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]
1	8.212	MW	0.2489	330.94400	22.15661
2	10.047	MW	0.3138	1.43211e4	760.68982

Totals : 1.46520e4 782.84643

Results obtained with enhanced integrator!

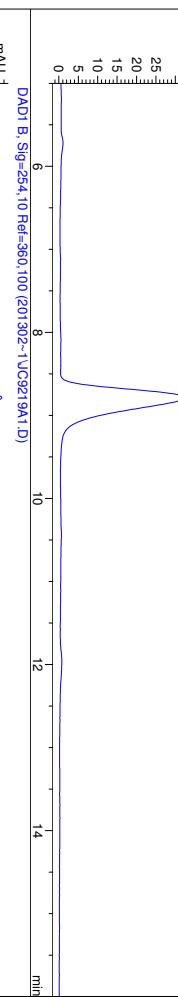
Signal 5: DAD1 E, Sig=280,10 Ref=360,100

mAU

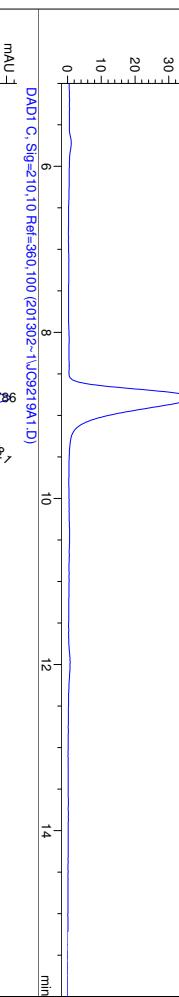
500
400
300
200
100
0

=====
 Injection Date : 2/11/2013 7:19:54 PM Seq. Line : 6
 Sample Name : JC9219A Location : Vial 5
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Last changed : 2/11/2013 7:37:52 PM by CE
 (modified after loading)
 Analysis Method : C:\HPCHEM\1\METHODS\OD-03-40.M
 Last changed : 7/30/2014 7:18:31 PM by MK
 (modified after loading)

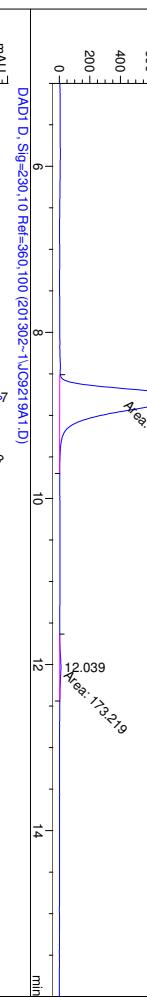
DAD1 A, Sig=250,10 Ref=360,100 (201302-1\JC9219A1.D)



DAD1 B, Sig=254,10 Ref=360,100 (201302-1\JC9219A1.D)



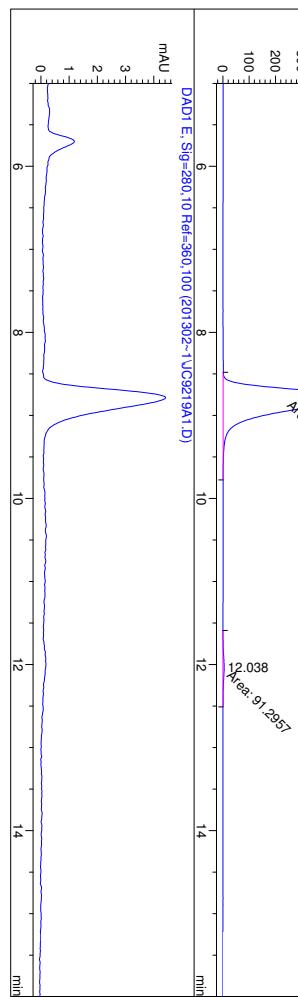
DAD1 C, Sig=210,10 Ref=360,100 (201302-1\JC9219A1.D)



DAD1 D, Sig=230,10 Ref=360,100 (201302-1\JC9219A1.D)



DAD1 E, Sig=280,10 Ref=360,100 (201302-1\JC9219A1.D)



=====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.786	FM	0.2972	1.46781e4	823.10712	98.8336
2	12.039	FM	0.3704	173.21939	7.79425	1.1664

Totals : 1.48513e4 830.90137

Results obtained with enhanced integrator!

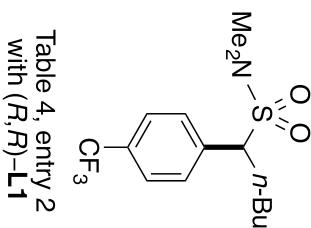
Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.787	FM	0.2959	8493.53320	478.45052	98.9365
2	12.038	FM	0.3593	91.29572	4.23473	1.0635

Totals : 8584.82892 482.68535

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100



Data File C:\HPCHEM\1\DATA\GROUP\J12077A1.D
Sample

Name: JC12077A

Data File C:\HPCHEM\1\DATA\GROUP\J12077A1.D

Sample Name: JC12077L

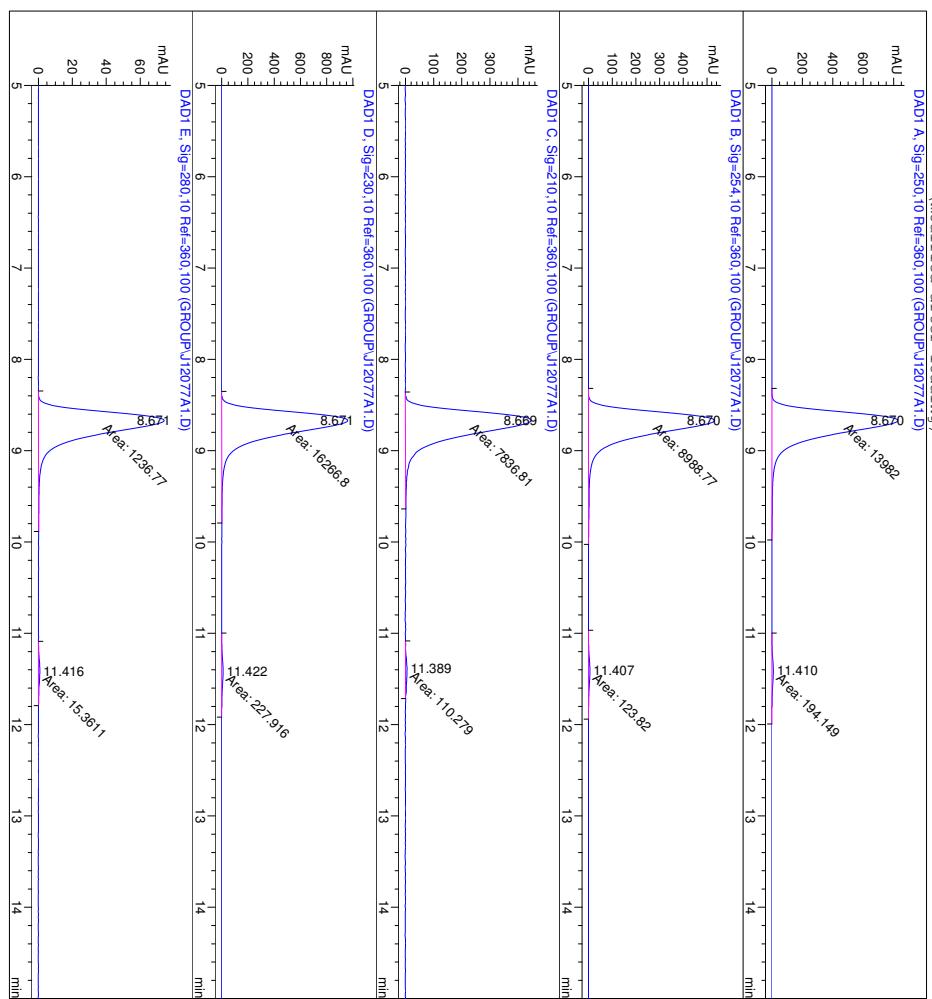
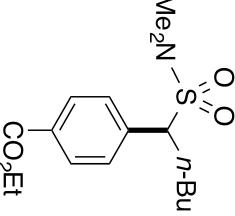


Table 4, entry 3
with (R,R) -L1

CC(C)(C)N(C)S(C)(C)C(C)C[C@H](C)c1ccccc1COC(=O)C



-122

O₂
Me₂N-
Table 4
with ((

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor With ISIDs

Signal 1: DADI A, Sig=250, 10 Ref=360, 100

Peak RetTime	Type	Width	Area	Height	Area %
#	[min]	[min]	[mAU*s]	[mAU]	
1	8.670 MM	0.2815	1.3982e4	827.71404	98.6305
2	11.410 MM	0.3566	194.14923	9.07509	1.3695

Totals : 1.41762e4 836.77913

Results obtained with enhanced integrator!

Signal 2: DADI B, Sig=254, 10 Ref=360, 100

Peak RetTime	Type	Width	Area	Height	Area
#	[min]	[min]	[mAU*s]	[mAU]	%
1	8.670 MM	0.2817	8988.7744	531.88164	98.6442
2	11.407 MM	0.3539	123.82027	5.83146	1.3588

Totals : 9112.59371 537.71610

Results obtained with enhanced integrator!

Signal 3: DADI C, Sig=210, 10 Ref=360, 100

Peak RetTime	Type	Width	Area	Height	Area
#	[min]	[min]	[mAU*s]	[mAU]	%
1	8.669 MM	0.2922	7836.80311	447.05707	98.6123
2	11.389 MM	0.3057	110.27897	6.01225	1.3877

Totals : 7947.08707 453.05932

Results obtained with enhanced integrator!

Signal 4: DADI D, Sig=230, 10 Ref=360, 100

Peak RetTime	Type	Width	Area	Height	Area
#	[min]	[min]	[mAU*s]	[mAU]	%
1	8.671 MM	0.2825	1.62668e4	959.65015	98.6183
2	11.422 MM	0.3601	227.91589	10.54932	1.3817

Totals : 1.64948e4 970.19947

Results obtained with enhanced integrator!

Signal 5: DADI E, Sig=280, 10 Ref=360, 100

Peak RetTime	Type	Width	Area	Height	Area
#	[min]	[min]	[mAU*s]	[mAU]	%
1	8.671 MM	0.2779	1236.76501	74.16629	98.7732
2	11.416 MM	0.3130	15.36111	8.18057e-1	1.2268

Totals : 1252.12613 74.98435

Results obtained with enhanced integrator!

Area	%
98.612	1.387
98.618	1.381
98.773	1.226

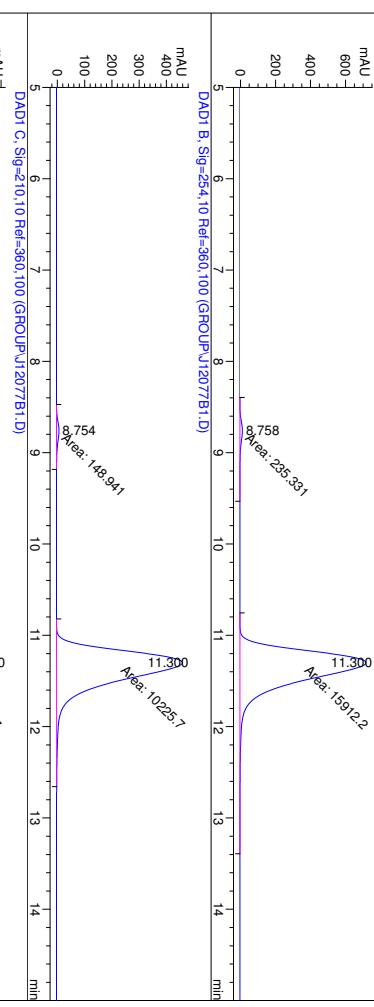
$$-\frac{1}{7} \quad -\frac{3}{7} \quad -\frac{1}{7}$$

Digitized by srujanika@gmail.com

Instrument 1 8/2/2014 12:59:23 PM MK

Injection Date : 8/2/2014 11:46:00 AM Seq. Line : 3
 Sample Name : JC12077B Location : Vial 18
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\VOD-05-40.M
 Last changed : 12/28/2013 3:19:26 PM by MK
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/2/2014 12:15:10 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (GROUP\J12077B1.D)



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.758	MM	0.2769	235.33102	14.16449	1.4574
2	11.300	MM	0.3687	1.59122e4	719.25024	98.5644

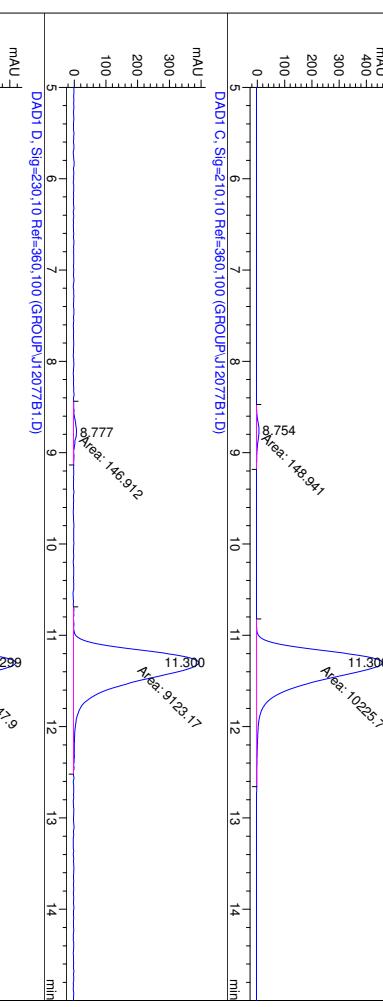
Totals :

1.61475e4

733.41473

Results obtained with enhanced integrator!

Signal 2: DAD1 B, Sig=254,10 Ref=360,100



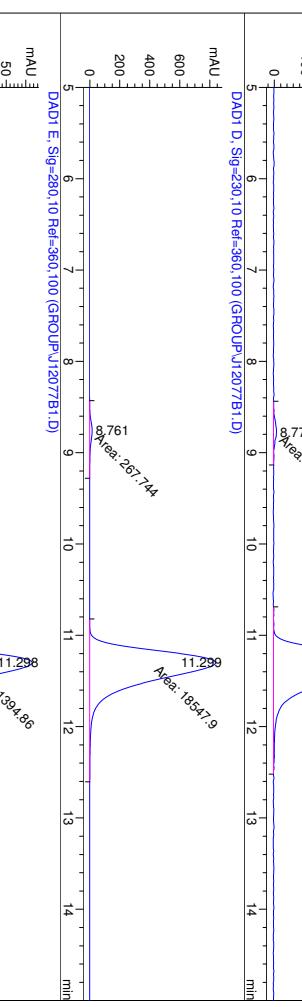
Totals :

1.03746e4

471.52147

Results obtained with enhanced integrator!

Signal 3: DAD1 C, Sig=210,10 Ref=360,100



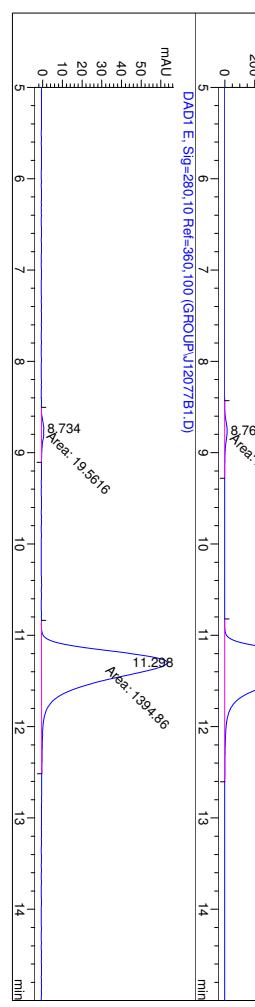
Totals :

9270.08516

406.07240

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100



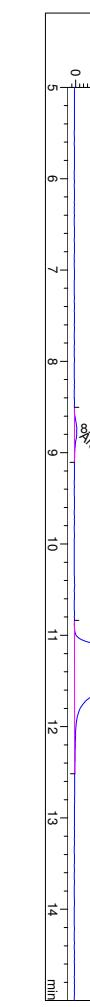
Totals :

1.88156e4

856.77717

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100



Totals :

1414.41723

65.17344

Results obtained with enhanced integrator!

**** End of Report ****

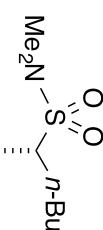


Table 4, entry 3
with (S,S)-L1

Data File C:\HPCHEM\1\DATA\201301~1\JC9159B1.D

Sample Name: JC9159B

Data File C:\HPCHEM\1\DATA\201301~1\JC9159B1.D

Sample Name: JC9159B

=====
 Injection Date : 1/17/2013 6:11:02 PM
 Sample Name : JC9159B
 Location : Vial 5
 Acq. Operator : CE
 Acq. Instrument : Instrument 1
 Inj : 1
 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-03-30.M
 Last changed : 12/31/2012 2:58:25 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/1/2014 11:54:48 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201301~1\JC9159B1.D)

mAU

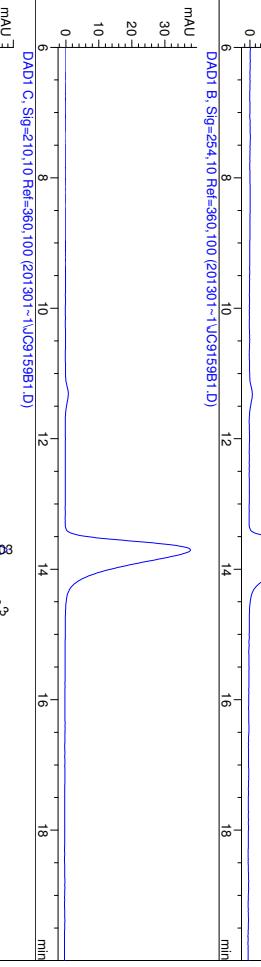
20

15

10

5

0



30

20

10

0

6

8

10

12

14

16

18

20

22

24

26

28

30

min

=====
 Area Percent Report
 =====

=====
 Signal
 =====

Sorted By
 =====

Multiplexer
 =====

1.0000
 =====

1.0000
 =====

Dilution
 =====

Use Multiplier & Dilution Factor with ISTDS
 =====

=====
 Signal 1: DAD1 A, Sig=250,10 Ref=360,100
 =====

Signal 2: DAD1 B, Sig=254,10 Ref=360,100
 =====

Signal 3: DAD1 C, Sig=210,10 Ref=360,100
 =====

=====
 Peak RetTime Type Width Area Height Area %
 =====

[min] [min] [mAU*s] [mAU] [%]
 =====

1 11.314 MWI 0.3282 703.80688 35.73879 2.2207
 =====

2 13.703 MWI 0.4525 3.09893e4 1141.28613 97.7793
 =====

Totals :
 =====

3.16931e4 1177.02492
 =====

Results obtained with enhanced integrator!
 =====

Signal 4: DAD1 D, Sig=230,10 Ref=360,100
 =====

Peak RetTime Type Width Area Height Area %
 =====

[min] [min] [mAU*s] [mAU] [%]
 =====

1 11.310 MWI 0.3180 250.68091 13.13707 1.8132
 =====

2 13.703 MWI 0.4130 1.35748e4 547.84503 98.1868
 =====

Totals :
 =====

1.38255e4 560.98210
 =====

Results obtained with enhanced integrator!
 =====

Signal 5: DAD1 E, Sig=280,10 Ref=360,100
 =====

Peak RetTime Type Width Area Height Area %
 =====

[min] [min] [mAU*s] [mAU] [%]
 =====

1 11.311 MWI 0.3191 109.32001 5.70986 1.8014
 =====

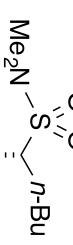
2 13.703 MWI 0.4053 559.26662 245.05873 98.1986
 =====

Totals :
 =====

6068.60663 250.76859
 =====

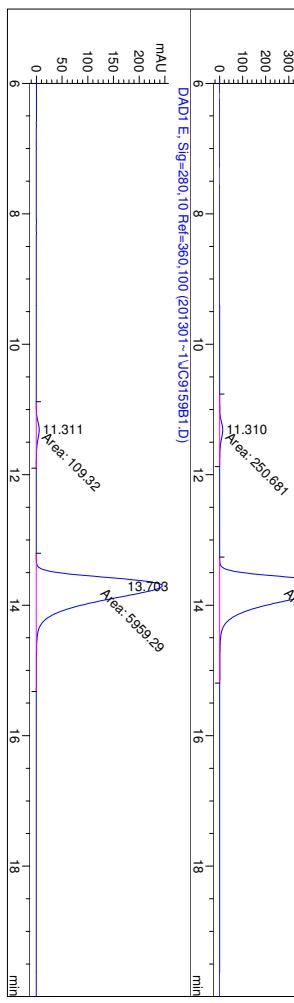
Results obtained with enhanced integrator!
 =====

=====
 *** End of Report ***
 =====



**Table 4, entry 4
with (S,S)-L1**

S-125



Data File C:\HPCHEM\1\DATA\201308-1\JC10223A.D

Sample Name: JC10223A

Data File C:\HPCHEM\1\DATA\201308-1\JC10223A.D

Sample Name: JC10223A

=====
Injection Date : 8/5/2013 11:54:59 PM Seq. Line : 2
Sample Name : JC10223A Location : Vial 94
Acq. Operator : MK Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
Acq. Method : C:\HPCHEM\1\METHODS\AS-05-40.M
Last changed : 4/7/2011 5:40:42 PM by CC
Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
Last changed : 7/30/2014 7:21:45 PM by MK
(modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10223A.D)



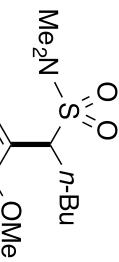
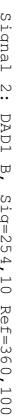
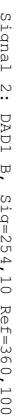
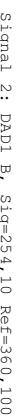
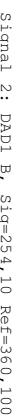
Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Signal 5: DAD1 E, Sig=280,10 Ref=360,100



**Table 4, entry 5
with (R,R)-L1**

Peak #	Retention Time [min]	Type	Width [min]	Area [MAU*s]	Height [MAU]	Area %
1	16.642	MM	0.4629	8169.60059	294.14471	98.0914
2	19.296	MM	0.5049	158.95808	5.24689	1.9086

Totals : 8328.55867 299.39160

Results obtained with enhanced integrator!

Signal 4: DAD1, Sig=230,10 Ref=360,100

Signal 5: DAD1, Sig=280,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [MAU*s]	Height [MAU]	Area %
1	16.641	MM	0.4629	8169.60059	294.14471	98.0914
2	19.296	MM	0.5049	158.95808	5.24689	1.9086

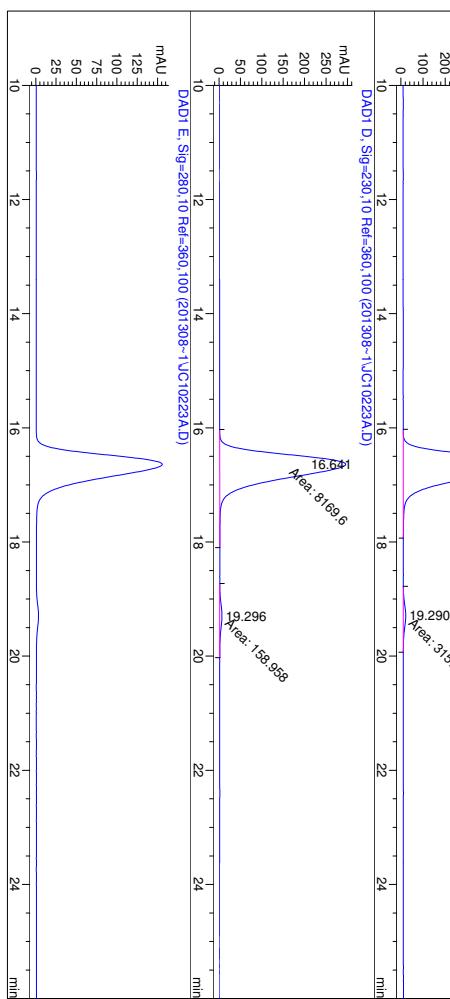
Totals : 8328.55867 299.39160

Results obtained with enhanced integrator!

Signal 5: DAD1, Sig=280,10 Ref=360,100

=====
*** End of Report ***
=====

S-126



DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223A.D)

Injection Date : 8/6/2013 12:36:17 AM Seq. Line : 3
 Sample Name : JC10223B Location : Vial 95
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AS-05-40.M Dilution :
 Last changed : 4/7/2011 5:40:42 PM by CC Use Multiplier & Dilution Factor with ISTDS
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/1/2014 11:56:57 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10223B.D)



mAU

12

10

8

6

4

2

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=254,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

15

10

5

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=210,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

15

10

5

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=230,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

200

100

0

10

20

30

40

50

60

70

80

90

100

120

100

50

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

22

24

min

DAD1, Sig=280,10 Ref=360,100 (201308-1\JC10223B.D)

mAU

125

100

75

50

25

0

10

12

14

16

18

20

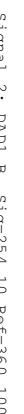
22

=====
 Injection Date : 8/9/2013 1:39:02 AM Seq. Line : 6
 Sample Name : JC10207A Location : Vial 42
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-02-30.M Dilution
 Last changed : 2/16/2013 4:45:43 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AD00540.M
 Last changed : 8/2/2014 1:04:33 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10207A.D)



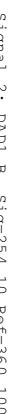
Signal 2: DAD1, B, Sig=254,10 Ref=360,100



Signal 3: DAD1, C, Sig=210,10 Ref=360,100



Signal 4: DAD1, D, Sig=230,10 Ref=360,100



Signal 5: DAD1, E, Sig=280,10 Ref=360,100

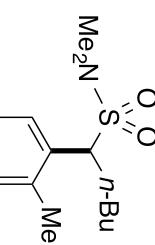


Table 4, entry 6
with (*R,R*)-L1

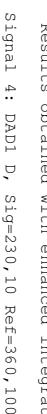
Peak #	Retention Time [min]	Type	Width [min]	Area [MAU*s]	Height [MAU]	Area %
1	9.763	MM	0.3255	2.90178e4	1485.83618	98.2944
2	11.939	MM	0.2984	503.50601	28.12491	1.7056

Totals : 2.95213e4 1513.96109

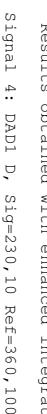
Results obtained with enhanced integrator!



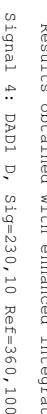
Signal 2: DAD1, B, Sig=254,10 Ref=360,100



Signal 3: DAD1, C, Sig=210,10 Ref=360,100



Signal 4: DAD1, D, Sig=230,10 Ref=360,100

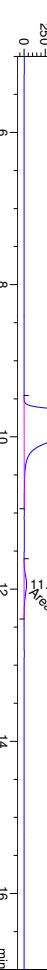


Signal 5: DAD1, E, Sig=280,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [MAU*s]	Height [MAU]	Area %
1	9.764	MM	0.2880	9872.96777	571.38434	98.4903
2	11.937	MM	0.2972	151.33591	8.48584	1.5097

Totals : 1.00243e4 579.87018

Results obtained with enhanced integrator!



Signal 2: DAD1, B, Sig=254,10 Ref=360,100



Signal 3: DAD1, C, Sig=210,10 Ref=360,100



Signal 4: DAD1, D, Sig=230,10 Ref=360,100



Signal 5: DAD1, E, Sig=280,10 Ref=360,100

***** End of Report *****

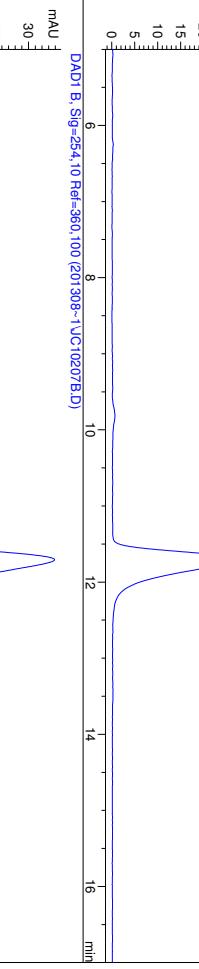
=====
 Injection Date : 8/9/2013 2:10:20 AM Seq. Line : 7
 Sample Name : JC10207B Location : Vial 43
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-02-30.M
 Last changed : 2/16/2013 4:45:43 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AD00540.M
 Last changed : 8/1/2014 1:59:19 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10207B.D)



Signal 2: DAD1, Sig=250,10 Ref=360,100

DAD1 B, Sig=254,10 Ref=360,100 (201308-1\JC10207B.D)



Signal 3: DAD1, C, Sig=210,10 Ref=360,100

DAD1 C, Sig=210,10 Ref=360,100 (201308-1\JC10207B.D)



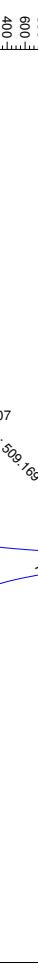
Signal 4: DAD1, D, Sig=230,10 Ref=360,100

DAD1 D, Sig=230,10 Ref=360,100 (201308-1\JC10207B.D)



Signal 5: DAD1, E, Sig=280,10 Ref=360,100 (201308-1\JC10207B.D)

DAD1 E, Sig=280,10 Ref=360,100 (201308-1\JC10207B.D)



Results obtained with enhanced integrator!

Signal 4: DAD1, D, Sig=230,10 Ref=360,100

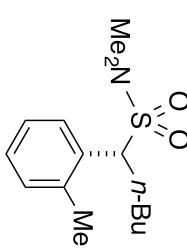
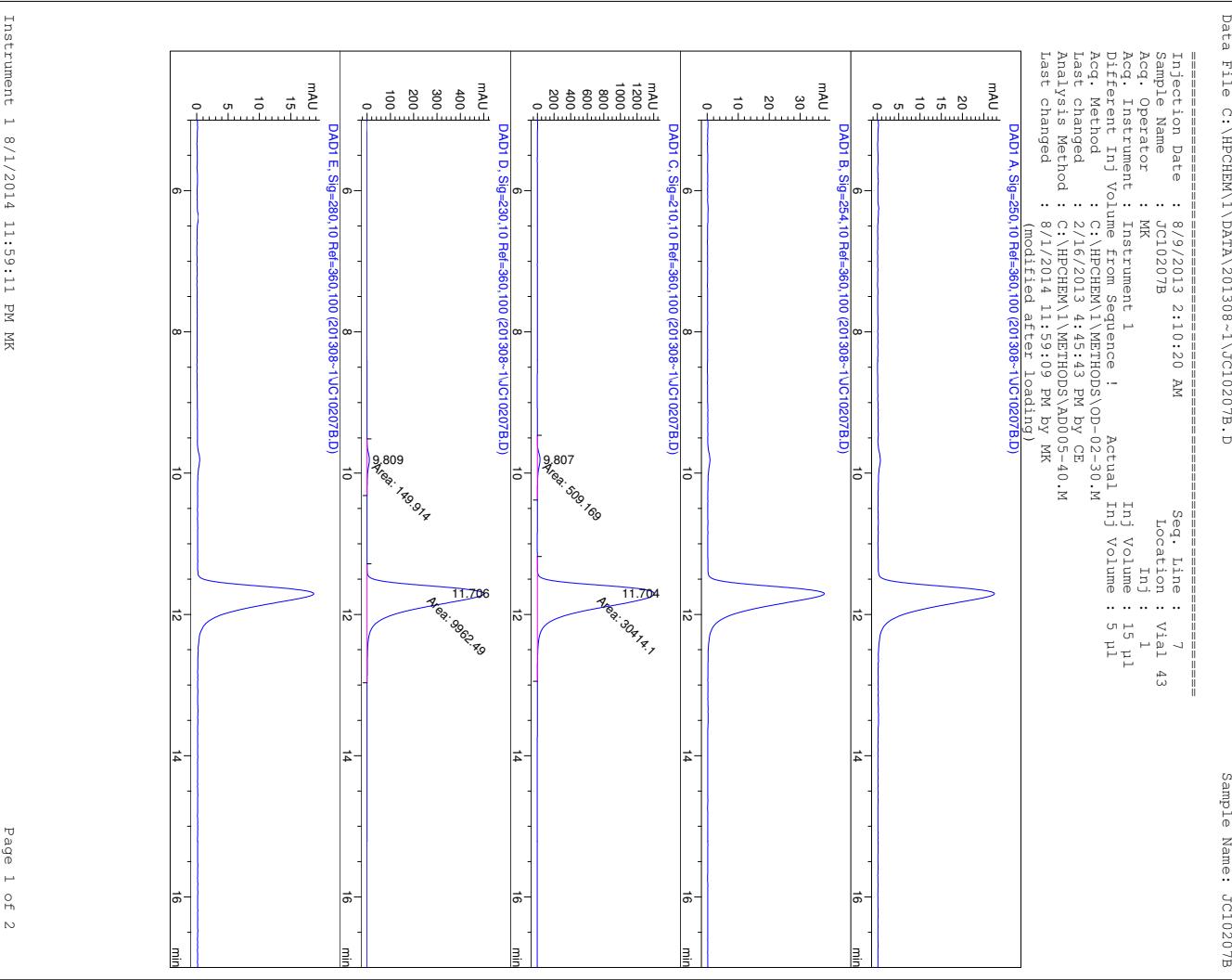
Peak #	Retention Time [min]	Type	Width [min]	Area [MAU*s]	Height [mAU]	Area %
1	9.809	MW	0.2538	149.91431	9.84457	1.4825
2	11.706	MW	0.3315	9362.49023	500.87515	98.5175

Totals : 1.01124e4 510.71973

Results obtained with enhanced integrator!

Signal 5: DAD1, E, Sig=280,10 Ref=360,100

===== *** End of Report *** =====

Table 4, entry 6
with (S,S)-L1

=====
 Injection Date : 10/16/2013 10:17:19 PM Seq. Line : 3
 Sample Name : JC11021 Location : Vial 12
 Acq. Operator : TMB Inj : 1
 Acq. Instrument : Instrument 1
 Acq. Method : C:\HPCHEM\METHODS\IC-1540.M Inj Volume : 5 μ l
 Last changed : 5/17/2012 7:45:15 PM by JTM
 Analysis Method : C:\HPCHEM\METHODS\ODH-0440.M
 Last changed : 7/30/2014 7:44:57 PM by TMB
 (modified after last loading)

DAD1 A, Sig=254,4 Ref=360,100 (201310-1\JC11021.D)

mAU

8
6
4
2
0

10

DAD1 B, Sig=254,16 Ref=360,100 (201310-1\JC11021.D)

mAU

8
6
4
2
0

10

DAD1 C, Sig=210,8 Ref=360,100 (201310-1\JC11021.D)

mAU

10
8
6
4
2
0

12.5

15

17.5

20

22.5

25

27.5

30

min

0

10

20

22.5

25</

Data File C:\HPCHEM\1\DATA\201310-1\JC10297B.D

Sample Name: JC10297B

Data File C:\HPCHEM\1\DATA\201310-1\JC10297B.D

Sample Name: JC10297B

=====
Injection Date : 10/16/2013 9:36:03 PM Seq. Line : 2
Sample Name : JC10297B Location : Vial 11
Acq. Operator : TMB Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
Acq. Method : C:\HPCHEM\METHODS\1540.M
Last changed : 5/17/2012 7:45:15 PM by JTM
Analysis Method : C:\HPCHEM\METHODS\ODH-0440.M
Last changed : 8/2/2014 12:24:05 AM by TMB
(modified after loading)

=====
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

=====
Signal 2: DAD1 B, Sig=254,16 Ref=360,100



=====

DAD1 A, Sig=254,4 Ref=360,100 (201310-1\JC10297B.D)

mAU

10

8

6

4

2

0

10 12.5 15 17.5 20 22.5 25 27.5 30 min

DAD1 B, Sig=254,16 Ref=360,100 (201310-1\JC10297B.D)

mAU

10

8

6

4

2

0

10 12.5 15 17.5 20 22.5 25 27.5 30 min

DAD1 B, Sig=254,16 Ref=360,100 (201310-1\JC10297B.D)

mAU

10

8

6

4

2

0

10 12.5 15 17.5 20 22.5 25 27.5 30 min

DAD1 C, Sig=210,8 Ref=360,100 (201310-1\JC10297B.D)

mAU

10

8

6

4

2

0

10 12.5 15 17.5 20 22.5 25 27.5 30 min

DAD1 D, Sig=230,16 Ref=360,100 (201310-1\JC10297B.D)

mAU

10

8

6

4

2

0

10 12.5 15 17.5 20 22.5 25 27.5 30 min

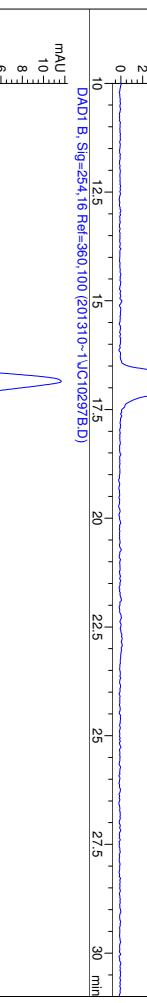
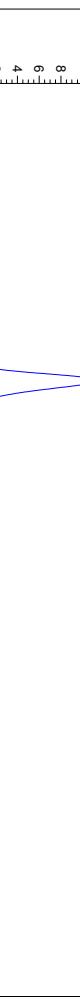
=====
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Signal 2: DAD1 B, Sig=254,16 Ref=360,100

Signal 3: DAD1 C, Sig=210,8 Ref=360,100

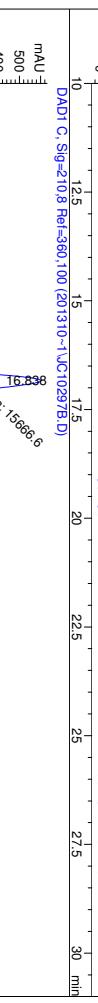
Signal 4: DAD1 D, Sig=230,16 Ref=360,100

Signal 5: DAD1 E, Sig=280,16 Ref=360,100



Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,16 Ref=360,100



Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,16 Ref=360,100

=====

Table 4, entry 7 with (S,S)-L1

=====

Peak Retention Time Width Area Height Area %

#	[min]	[min]	[mAU*s]	[mAU]	%
1	16.838	MM	0.4396	4621.76465	175.22726
2	22.878	MM	0.5719	66.18513	1.92889

=====

End of Report ****

Injection Date : 6/14/2013 11:25:16 PM Seq. Line : 3
 Sample Name : JC10133B Location : Vial 31
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 2 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\ND0-02-40.M
 Last changed : 2/22/2013 10:06:44 AM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\LA005-40.M
 Last changed : 8/2/2014 12:07:02 AM by MK
 (modified after loading)

DAD1 A, Sig=250,10 Ref=360,100 (201306-1\JCX133B1.D)



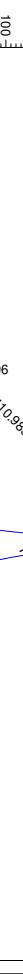
DAD1 B, Sig=254,10 Ref=360,100 (201306-1\JCX133B1.D)



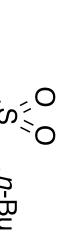
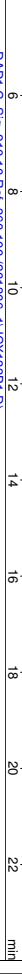
DAD1 C, Sig=254,10 Ref=360,100 (201306-1\JCX133B1.D)



DAD1 D, Sig=230,10 Ref=360,100 (201306-1\JCX133B1.D)



DAD1 E, Sig=280,10 Ref=360,100 (201306-1\JCX133B1.D)



Signal 1: DAD1 A, Sig=250,10 Ref=360,100
 Signal 2: DAD1 B, Sig=254,10 Ref=360,100
 Signal 3: DAD1 C, Sig=254,10 Ref=360,100

Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %

#	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.708	MM	0.4004	459.7119	19.13779	4.8354
2	15.305	MM	0.5404	9047.59863	279.01901	95.1646

Totals : 9507.31055 298.15680

Results obtained with enhanced integrator!

Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %

#	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.706	MM	0.4003	410.98477	17.11033	4.8296
2	15.305	MM	0.5407	8098.67578	249.64102	95.1704

Totals : 8099.66055 266.75135

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %

#	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.708	MM	0.4001	523.60632	21.81178	4.9454
2	15.306	MM	0.5401	1.00641e4	310.58426	95.0546

Totals : 1.05877e4 332.39604

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %

#	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.708	MM	0.4006	1306.40112	54.35100	4.8897
2	15.305	MM	0.5438	2.54111e4	778.85779	95.1103

Totals : 2.67175e4 833.20879

Results obtained with enhanced integrator!

Signal 6: DAD1 F, Sig=280,10 Ref=360,100

Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %

#	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.710	MM	0.3975	110.80473	4.64648	4.8097
2	15.305	MM	0.5379	2192.98193	67.95478	95.1903

Totals : 2303.78667 72.60126

Results obtained with enhanced integrator!

Signal 7: DAD1 G, Sig=280,10 Ref=360,100

Peak RetTime Type Width Area Height Area
 # [min] [min] [mAU*s] [mAU] %

#	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.710	MM	0.3975	110.80473	4.64648	4.8097
2	15.305	MM	0.5379	2192.98193	67.95478	95.1903

Totals : 2303.78667 72.60126

Results obtained with enhanced integrator!
 *** End of Report ***

Data File C:\HPCHEM\1\DATA\201308-1\JC10219A.D

Sample Name: JC10219A

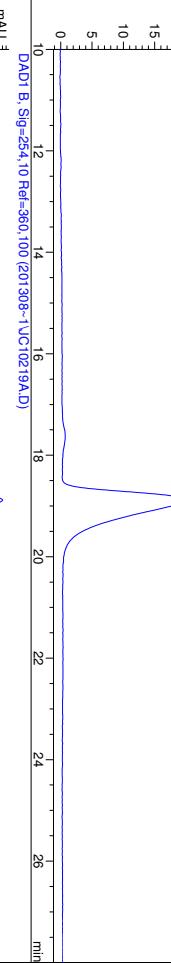
Data File C:\HPCHEM\1\DATA\201308-1\JC10219A.D

Sample Name: JC10219A

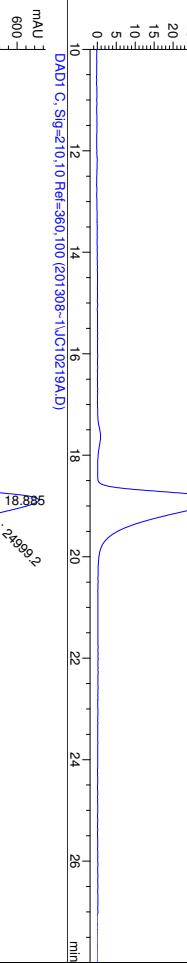
=====
Injection Date : 8/1/2013 10:52:58 AM Seq. Line : 2
Sample Name : JC10219A Location : Vial 79
Acq. Operator : CE Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
Acq. Method : C:\HPCHEM\1\METHODS\OD-05-40.M
Last changed : 4/7/2011 5:39:35 PM by CC
Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
Last changed : 7/30/2014 7:32:15 PM by MK
(modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10219A.D)

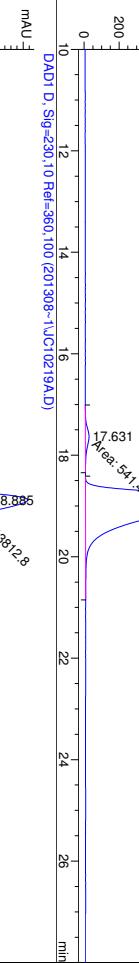
mAU



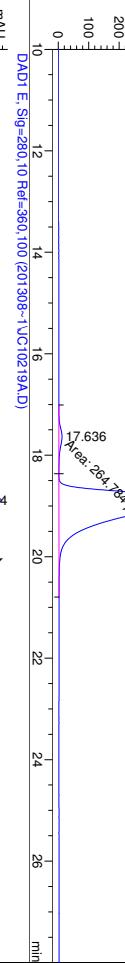
mAU



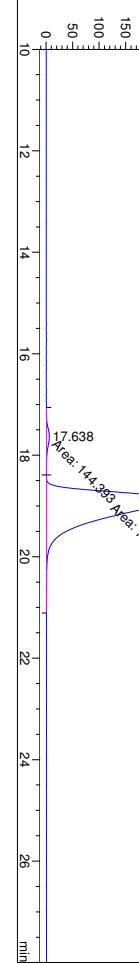
mAU



mAU



mAU



mAU



=====
Area Percent Report
=====
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDS

Signal 2: DAD1, Sig=254,10 Ref=360,100

Signal 3: DAD1, C, Sig=210,10 Ref=360,100

Signal 4: DAD1, D, Sig=230,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.631	MM		0.4328	2.541.48834	20.85328
2	18.885	MM		0.5805	2.49999.2	17.76105

Totals : 2.55407e4 738.61432

Results obtained with enhanced integrator!

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.636	MM		0.4283	2.641.78415	10.30480
2	18.885	MM		0.5576	1.38128e4	412.84863

Totals : 1.40776e4 423.15343

Results obtained with enhanced integrator!

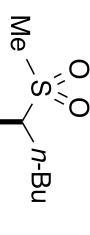
Signal 5: DAD1, E, Sig=280,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.638	MM		0.4283	144.39316	5.61829
2	18.884	MM		0.5489	7802.81	236.90977

Totals : 7947.20712 242.52807

Results obtained with enhanced integrator!

=====
*** End of Report ***
=====



**Table 4, entry 9
with (R,R)-L1**

Injection Date : 8/1/2013 11:34:20 AM Seq. Line : 3
 Sample Name : JC10219B Location : Vial 80
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-05-40.M
 Last changed : 4/7/2011 5:39:35 PM by CC
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/2/2014 12:10:46 AM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10219B.D)

mAU



mAU



mAU

15

10

5

0

10

15

20

25

30

35

40

45

50

55

60

65

70

75

80

85

90

95

100

105

110

115

120

125

130

135

140

145

150

155

160

165

170

175

180

185

190

195

200

205

210

215

220

225

230

235

240

245

250

255

260

265

270

275

280

285

290

295

300

305

308

310

312

314

316

318

320

322

324

326

328

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730

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750

752

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756

758

760

762

764

766

768

770

772

774

776

778

780

782

784

786

788

790

792

794

796

798

800

802

804

806

808

810

Data File C:\HPCHEM\1\DATA\201308-1\JC10215A.D

Sample Name: JC10215A

Data File C:\HPCHEM\1\DATA\201308-1\JC10215A.D

Sample Name: JC10215A

=====
 Injection Date : 8/1/2013 2:28:49 AM
 Sample Name : JC10215A
 Location : Vial 79
 Acq. Operator : CE
 Inj. Inj. : 1
 Acq. Instrument : Instrument 1
 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AS-10-40.M
 Last changed : 4/6/2011 9:32:30 PM by CC
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 7:34:55 PM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10215A.D)



Signal 2: DAD1, B, Sig=254,10 Ref=360,100



Signal 3: DAD1, C, Sig=210,10 Ref=360,100



Signal 4: DAD1, D, Sig=230,10 Ref=360,100



Signal 5: DAD1, E, Sig=280,10 Ref=360,100



Totals :

2.56535e4 491.44314

Results obtained with enhanced integrator!

=====
 Area Percent Report
 =====

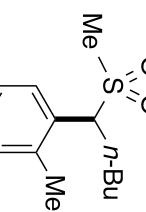
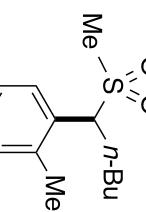
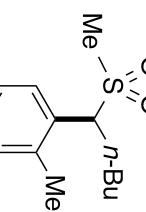
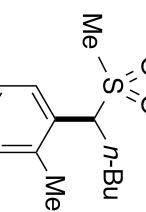
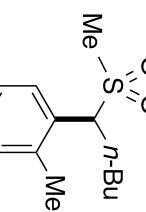
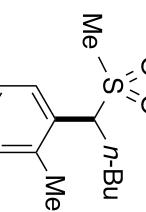
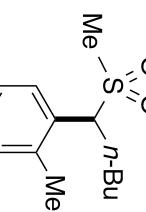
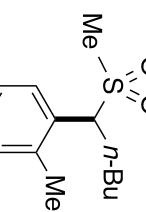
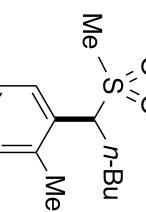
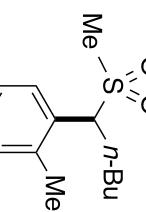
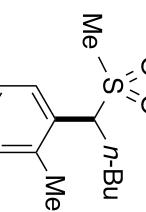
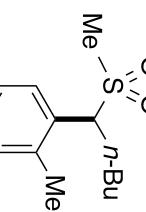
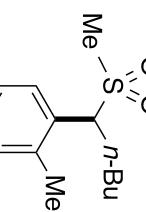
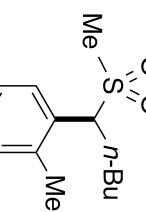
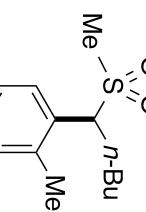
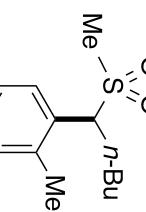
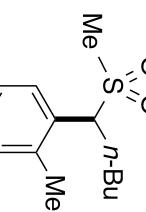
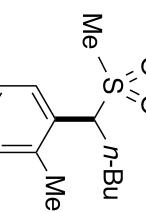
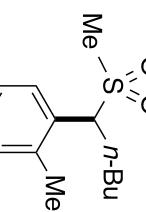
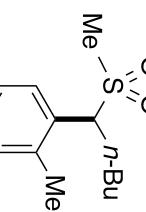
Sorted By
 =====

Multiplexer : Signal
 =====

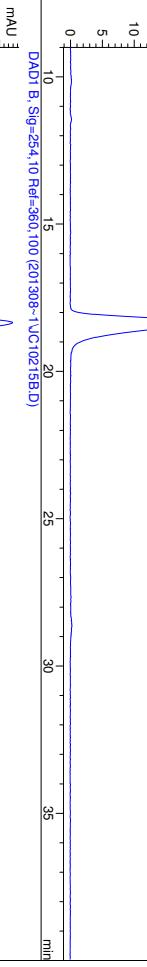
Dilution : 1.0000
 =====

Use Multiplier & Dilution Factor with ISTDS
 =====

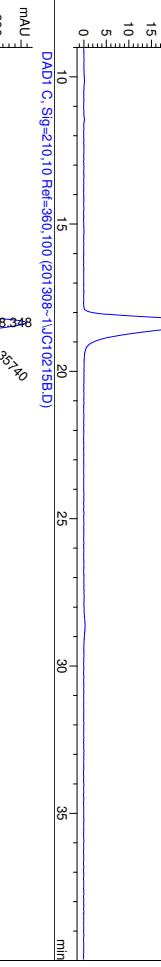
=====
 End of Report ***
 =====



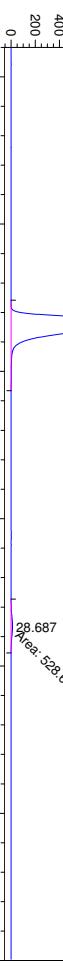
=====
 Injection Date : 8/1/2013 3:10:07 AM Seq. Line : 19
 Sample Name : JC10215B Location : Vial 80
 Acq. Operator : CE Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AS-10-40.M Dilution :
 Last changed : 4/6/2011 9:32:30 PM by CC Use Multiplier & Dilution Factor with ISTDS
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/2/2014 12:12:31 AM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201308-1\JC10215B.D)**mAU**15
10
5
0

mAU

20
15
10
5
0

mAU

10
15
20
25
30
35
min

mAU

10
15
20
25
30
35
min

=====
 Area Percent Report
 =====
 Signal # : 1
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS
 =====

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [MAU*s]	Height [MAU]	Area %
1	18.348	MM	0.5711	3.5740e4	1042.94275	98.5423
2	28.687	MM	0.7471	528.68359	11.79463	1.4577

Totals : 3.62687e4 1054.73737

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

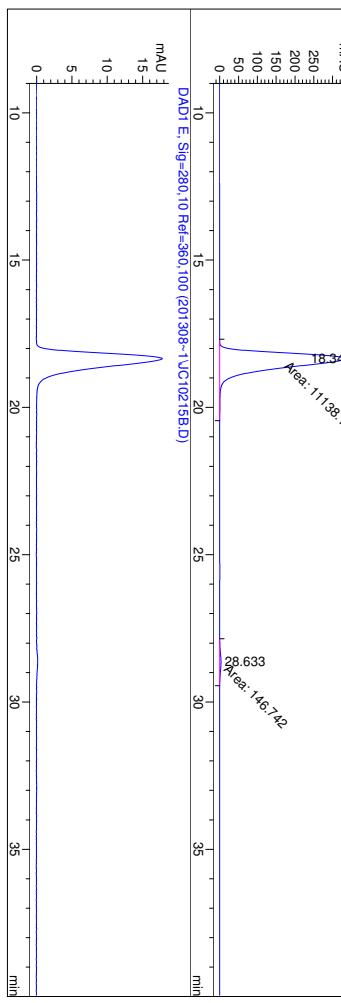
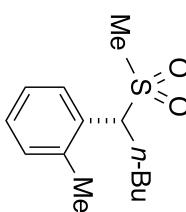
Peak #	Retention Time [min]	Type	Width [min]	Area [MAU*s]	Height [MAU]	Area %
1	18.346	MM	0.5543	1.11381e4	334.92871	98.6997
2	28.633	MM	0.6968	146.74203	3.50967	1.3003

Totals : 1.12849e4 338.43838

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====
 *** End of Report ***
 =====



mAU

10
15
20
25
30
35

=====
 Injection Date : 10/16/2013 10:37:14 PM Seq. Line : 2
 Sample Name : JC11023A Location : Vial 91
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AS-10-40.M
 Last changed : 9/3/2013 3:07:57 PM by MK
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/2/2014 12:15:46 AM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201310-1\JC11023A.D)



mAU

0

2

4

6

8

10

15

20

25

30

min

DAD1, Sig=254,10 Ref=360,100 (201310-1\JC11023A.D)

mAU

0

2

4

6

8

10

15

20

25

30

min

DAD1, Sig=210,10 Ref=360,100 (201310-1\JC11023A.D)

mAU

0

2

4

6

8

10

15

20

25

30

min

DAD1, Sig=230,10 Ref=360,100 (201310-1\JC11023A.D)

mAU

0

10

20

30

40

50

60

70

80

90

100

110

120

130

140

150

160

170

180

190

200

210

220

230

240

250

260

270

280

290

300

310

320

330

340

350

360

370

380

390

400

410

420

430

440

450

460

470

480

490

500

510

520

530

540

550

560

570

580

590

600

610

620

630

640

650

660

670

680

690

700

710

720

730

740

750

760

770

780

790

800

810

820

830

840

850

860

870

880

890

900

910

920

930

940

950

960

970

980

990

1000

1010

1020

1030

1040

1050

1060

1070

1080

1090

1100

1110

1120

1130

1140

1150

1160

1170

1180

1190

1200

1210

1220

1230

1240

1250

1260

1270

1280

1290

1300

1310

1320

1330

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1350

1360

1370

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1470

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1490

1500

1510

1520

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1600

1610

1620

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1940

1950

1960

1970

1980

1990

2000

2010

2020

2030

2040

2050

2060

2070

2080

2090

2100

2110

2120

2130

2140

2150

2160

2170

2180

2190

2200

2210

2220

2230

2240

2250

2260

2270

2280

2290

2300

2310

2320

2330

2340

2350

2360

2370

2380

2390

2400

2410

2420

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2490

2500

2510

2520

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2570

2580

2590

2600

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2690

2700

2710

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2740

2750

2760</

Data File C:\HPCHEM\1\DATA\201310-1\JC11023B.D

Sample Name: JC11023B

Data File C:\HPCHEM\1\DATA\201310-1\JC11023B.D

Sample Name: JC11023B

=====
 Injection Date : 10/16/2013 11:18:31 PM Seq. Line : 3
 Sample Name : JC11023B Location : Vial 92
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AS-10-40.M
 Last changed : 9/3/2013 3:07:57 PM by MK
 Analysis Method : C:\HPCHEM\1\METHODS\AN005-40.M
 Last changed : 8/2/2014 12:15:46 AM by MK
 (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201310-1\JC11023B.D)

mAU

8

6

4

2

0

5

10

15

20

25

30

min

DAD1, Sig=254,10 Ref=360,100 (201310-1\JC11023B.D)

mAU

10

8

6

4

2

0

5

10

15

20

25

30

min

DAD1, Sig=210,10 Ref=360,100 (201310-1\JC11023B.D)

mAU

10

8

6

4

2

0

5

10

15

20

25

30

min

DAD1, Sig=230,10 Ref=360,100 (201310-1\JC11023B.D)

mAU

10

8

6

4

2

0

5

10

15

20

25

30

min

DAD1, Sig=280,10 Ref=360,100 (201310-1\JC11023B.D)

mAU

150

100

50

0

5

10

15

20

25

30

min

DAD1, Sig=280,10 Ref=360,100 (201310-1\JC11023B.D)

mAU

50

100

150

200

250

300

350

400

450

500

550

600

650

700

750

800

850

900

950

1000

1050

1100

1150

1200

1250

1300

1350

1400

1450

1500

1550

1600

1650

1700

1750

1800

1850

1900

1950

2000

2050

2100

2150

2200

2250

2300

2350

2400

2450

2500

2550

2600

2650

2700

2750

2800

2850

2900

2950

3000

3050

3100

3150

3200

3250

3300

3350

3400

3450

3500

3550

3600

3650

3700

3750

3800

3850

3900

3950

4000

4050

4100

4150

4200

4250

4300

4350

4400

4450

4500

4550

4600

4650

4700

4750

4800

4850

4900

4950

5000

5050

5100

5150

5200

5250

5300

5350

5400

5450

5500

5550

5600

5650

5700

5750

5800

5850

5900

5950

6000

6050

6100

6150

6200

6250

6300

6350

6400

6450

6500

6550

6600

6650

6700

6750

6800

6850

6900

6950

7000

7050

7100

7150

7200

7250

7300

7350

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7500

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7600

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7900

7950

8000

8050

8100

8150

8200

8250

8300

8350

8400

8450

8500

8550

8600

8650

8700

8750

8800

8850

8900

8950

9000

9050

9100

9150

9200

9250

9300

9350

9400

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9500

9550

9600

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9700

9750

9800

9850

9900

9950

10000

10050

10100

10150

10200

10250

10300

10350

10400

10450

10500

10550

10600

10650

10700

10750

10800

10850

10900

10950

11000

11050

11100

11150

11200

11250

11300

11350

11400

Data File C:\HPCHEM\1\DATA\201401~1\JC11079L.D

Sample Name: JC11079 LINEAR

Data File C:\HPCHEM\1\DATA\201401~1\JC11079L.D

Sample Name: JC11079 LINEAR

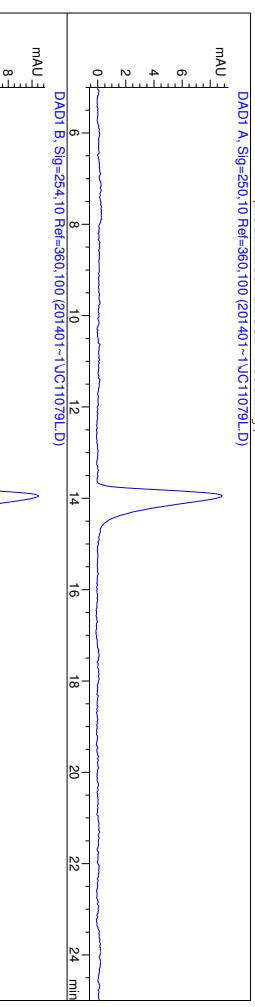
=====
 Injection Date : 1/31/2014 11:15:03 PM Seq. Line : 9
 Sample Name : JC11079 LINEAR Location : Vial 10
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01-40.M
 Last changed : 12/28/2013 4:11:34 PM by MK
 Analysis Method : C:\HPCHEM\1\METHODS\AA005-40.M
 Last changed : 8/2/2014 11:58:47 PM by MK
 (modified after loading)

=====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS

Signal 1: DAD1 A, Sig=250, 10 Ref=360, 100
 Signal 2: DAD1 B, Sig=254, 10 Ref=360, 100
 Signal 3: DAD1 C, Sig=210, 10 Ref=360, 100
 Signal 4: DAD1 D, Sig=230, 10 Ref=360, 100
 Signal 5: DAD1 E, Sig=280, 10 Ref=360, 100

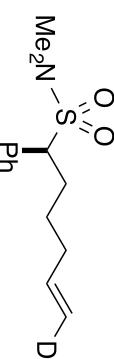
Totals : 1.44586e4 579.72513



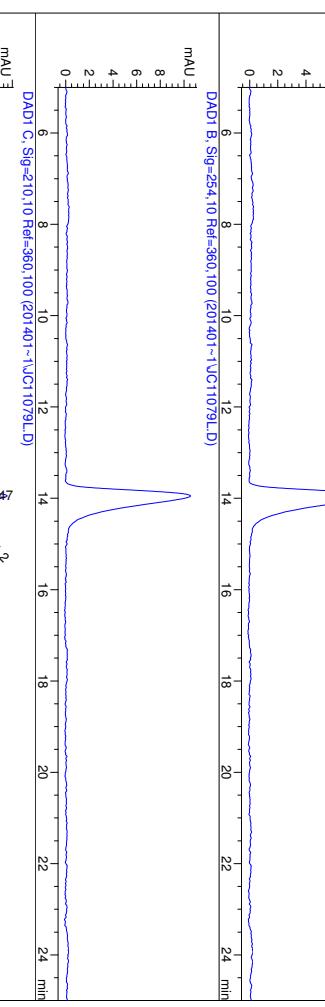
Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230, 10 Ref=360, 100

=====
 *** End of Report ***
 =====



eq 3
with $(R,R)\text{-L1}$



Signal 5: DAD1 E, Sig=280, 10 Ref=360, 100



S-140

Data File C:\HPCHEM\1\DATA\GROUP\J12075A5.D
Sample Name: J1C12075A

Data File C:\HPCHEM\1\DATA\GROUP\J12075A5.D
Sample Name: J1C12075A

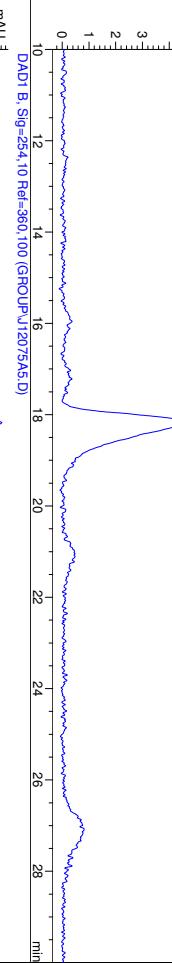
Sample Name: J1C12075A

=====
Injection Date : 8/1/2014 10:06:30 PM Seq. Line : 2
Sample Name : J1C12075A Location : Vial 19
Acq. Operator : MK Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 4 μ l
Acq. Method : C:\HPCHEM\1\METHODS\00J-05-30.M
Last changed : 8/1/2014 10:12:37 PM by MK
(modified after loading)
Analysis Method : C:\HPCHEM\1\METHODS\AA0005-40.M
Last changed : 8/2/2014 11:25:11:4 PM by MK
(modified after loading)

=====
Injection Date : 8/1/2014 10:06:30 PM Seq. Line : 2
Sample Name : J1C12075A Location : Vial 19
Acq. Operator : MK Inj : 1
Acq. Instrument : Instrument 1 Inj Volume : 15 μ l
Different Inj Volume From Sequence ! Actual Inj Volume : 4 μ l
Acq. Method : C:\HPCHEM\1\METHODS\00J-05-30.M
Last changed : 8/1/2014 10:12:37 PM by MK
(modified after loading)
Analysis Method : C:\HPCHEM\1\METHODS\AA0005-40.M
Last changed : 8/2/2014 11:25:11:4 PM by MK
(modified after loading)

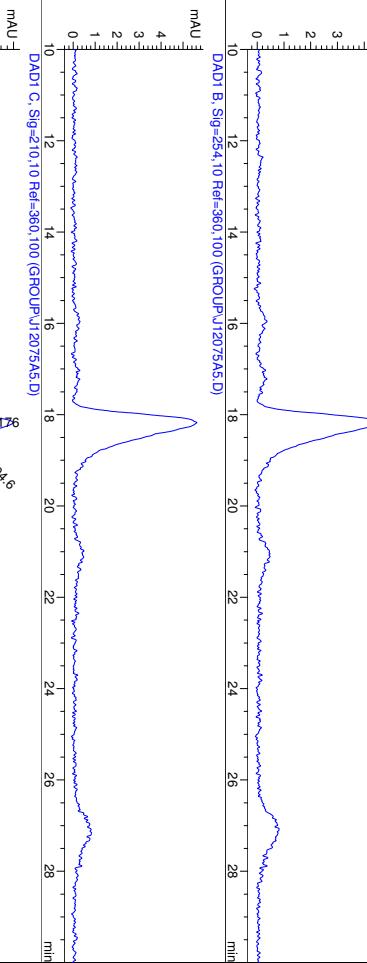
DAD1 A, Sig=250,10 Ref=360,100 (GROUP\J12075A5.D)

mAU



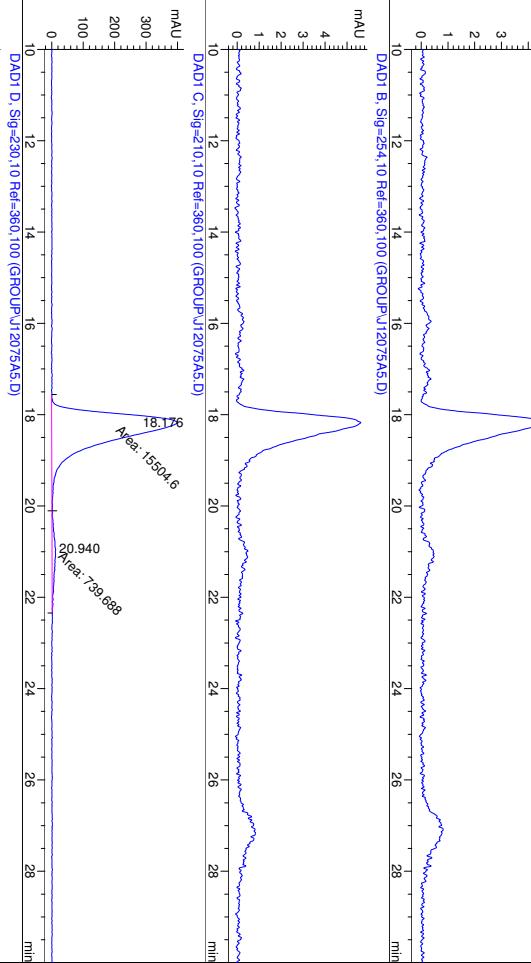
Signal 2: DAD1 B, Sig=254,10 Ref=360,100

mAU



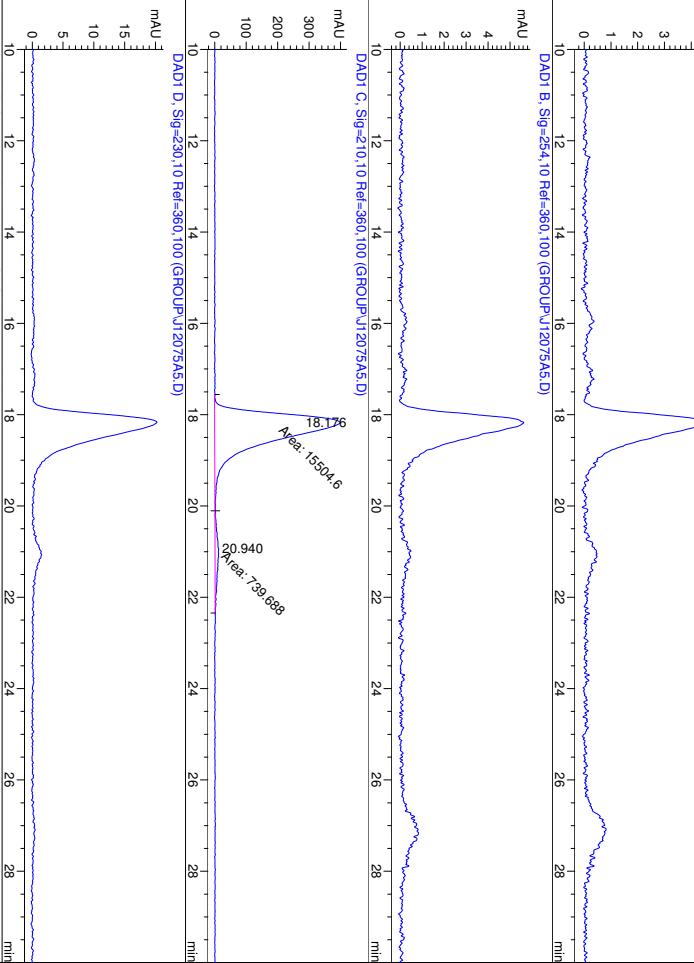
Signal 3: DAD1 C, Sig=210,10 Ref=360,100

mAU



Signal 4: DAD1 D, Sig=230,10 Ref=360,100

mAU



Signal 5: DAD1 E, Sig=280,10 Ref=360,100

mAU

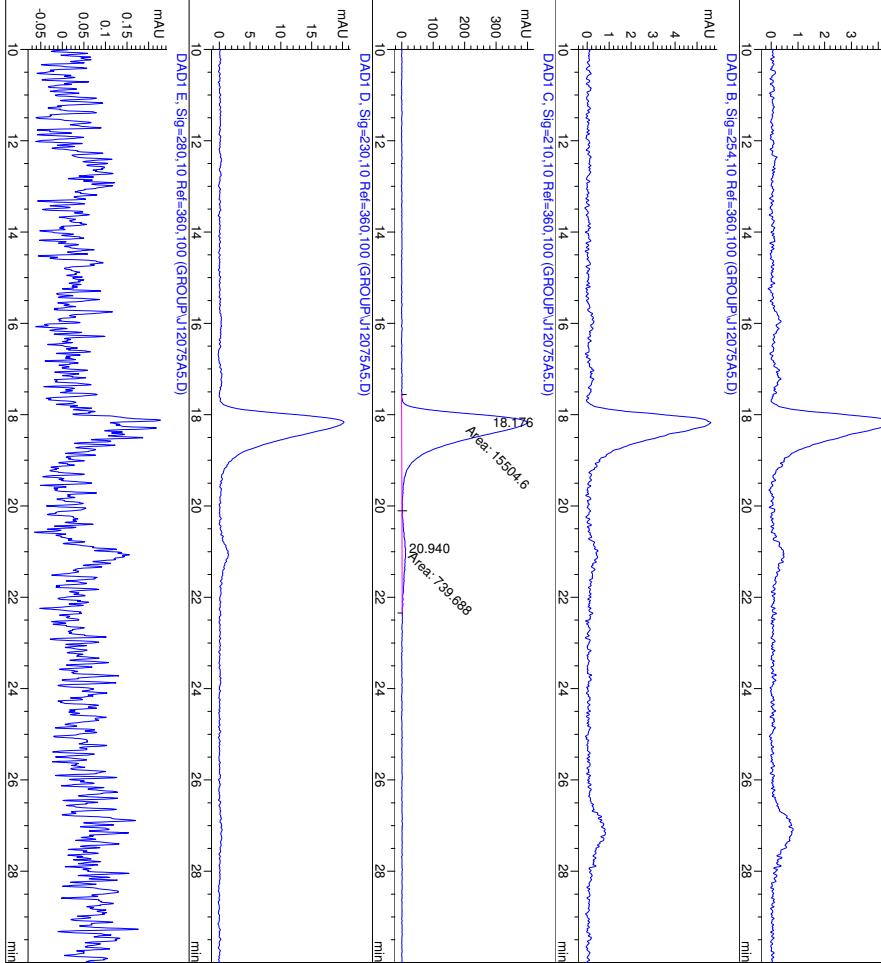


Table 5, entry 1
with ($3R,8S$)-L6

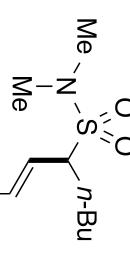


Table 5, entry 1
with ($3R,8S$)-L6

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.176	MP	0.6470	1.5506e4	399.41086	95.4465
2	20.940	FM	1.0783	739.68774	11.43283	4.5535

Totals : 1.62443e4 410.84368

Results obtained with enhanced integrator!

Signal 4: DAD1, Sig=230,10 Ref=360,100

=====
*** End of Report ***
=====

Data File C:\HPCHEM\1\DATA\GROUP\J12075B5.D

Sample Name: JCT12075B

=====
 Injection Date : 8/1/2014 10:47:48 PM
 Sample Name : JC12075B
 Location : Vial 20
 Acq. Operator : MK
 Acq. Instrument : Instrument 1
 Inj : 1
 Different Inj Volume From Sequence ! Actual Inj Volume : 4 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\00-J-05-30.M
 Last changed : 8/1/2014 10:12:37 PM by MK
 (modified after loading)
 Analysis Method : C:\HPCHEM\1\METHODS\A0005-40.M
 Last changed : 8/2/2014 12:51:14 PM by MK
 (modified after loading)

mAU

DAD1 A, Sig=250,10 Ref=360,100 (GROUP J12075B5.D)

Sample Name: JC12075B

mAU

DAD1 B, Sig=254,10 Ref=360,100 (GROUP J12075B5.D)

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

mAU

DAD1 C, Sig=210,10 Ref=360,100 (GROUP J12075B5.D)

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

mAU

DAD1 D, Sig=230,10 Ref=360,100 (GROUP J12075B5.D)

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

mAU

DAD1 E, Sig=280,10 Ref=360,100 (GROUP J12075B5.D)

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

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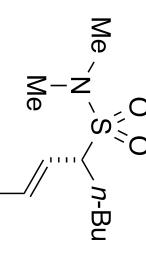


Table 5, entry 1
with $(3S,8R)$ -L6

Peak #	Retention Time [min]	Width [min]	Area [mAU]	Height [mAU]	Area %
1	18.563	0.6055	800.43811	22.03275	5.2285
2	20.658	1.4329	1.45087e4	168.75829	94.7715

Totals :

1,53092e4 190.79103

Results obtained with enhanced integrator!

Signal 4: DAD1, Sig=230,10 Ref=360,100

=====
 **** End of Report ****
 =====

Data File C:\HPCHEM\1\DATA\201309-1\JC10275.D

Sample Name: JC10275

Data File C:\HPCHEM\1\DATA\201309-1\JC10275.D

Sample Name: JC10275

=====
 Injection Date : 9/4/2013 1:38:06 PM
 Sample Name : JC10275
 Location : Vial 9
 Acq. Operator : MK
 Acq. Instrument : Instrument 1
 Inj : 1
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Last changed : 11/14/2012 10:09:31 PM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\JC-ADOLIA.M
 Last changed : 8/3/2014 12:13:42 AM by MK
 (modified after loading)

DAD1 A, Sig=250,10 Ref=360,100 (201309-1\JC10275.D)



Signal 2: DAD1 B, Sig=254,10 Ref=360,100



Signal 3: DAD1 C, Sig=210,10 Ref=360,100



Signal 4: DAD1 D, Sig=230,10 Ref=360,100



Signal 5: DAD1 E, Sig=280,10 Ref=360,100



=====
 Area Percent Report
 =====

Sample Name: JC10275

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS

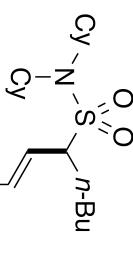
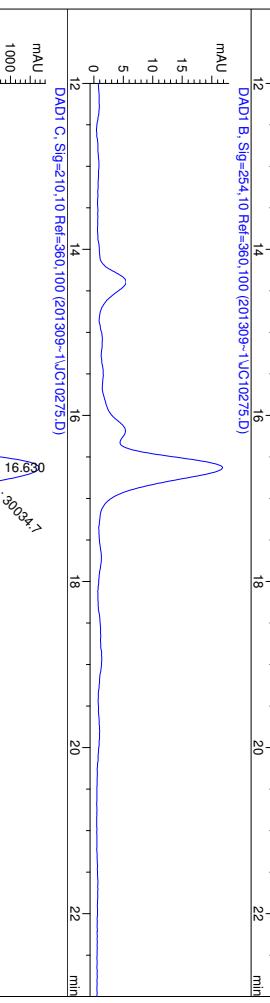


Table 5, entry 2
with (3*R*,8*S*)-L6

CH₂Ph

=====
 **** End of Report ****



Totals : 3.08045e4 1303.61983
 Results obtained with enhanced integrator!
 Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====
 **** End of Report ****

Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D

Sample Name: J11025A

Sample Name: J11025A

=====
 Injection Date : 10/22/2013 5:01:30 PM
 Sample Name : J11025A
 Location : Vial 9
 Acq. Operator : MK
 Acq. Instrument : Instrument 1
 Inj : 1
 Different Inj Volume From Sequence ! Actual Inj Volume : 3 μ l
 Last changed : 10/22/2013 9:47:57 AM by MK
 Analysis Method : C:\HPCHEM\1\METHODS\Anal05-40.M
 Last changed : 7/30/2014 10:35:34 PM by MK
 (modified after loading)

=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====

=====
 Seq. Line : 8
 Location : Vial 9
 Inj : 1
 Acq. Method : C:\HPCHEM\1\METHODS\Anal05-40.M
 Last changed : 10/22/2013 9:47:57 AM by MK
 Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 10:35:34 PM by MK
 (modified after loading)

=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====

=====
 Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDS

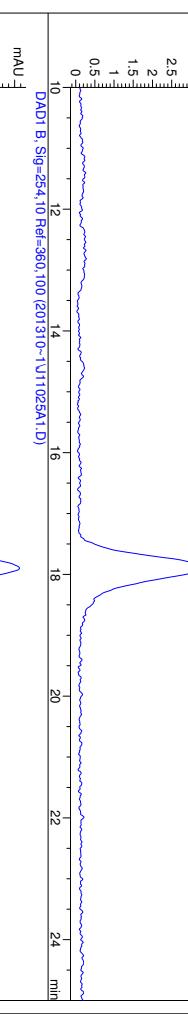
=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====

=====
 DAD1 A, Sig=250,10 Ref=360,100 (201310-1\J11025A1.D)

=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====



Totals : 7952.09590 252.36462

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====

=====
 DAD1 B, Sig=254,10 Ref=360,100 (201310-1\J11025A1.D)

=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====

=====
 DAD1 C, Sig=210,10 Ref=360,100 (201310-1\J11025A1.D)

=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====

=====
 DAD1 D, Sig=230,10 Ref=360,100 (201310-1\J11025A1.D)

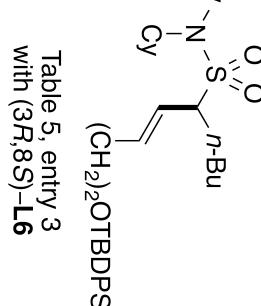
=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====

=====
 DAD1 E, Sig=280,10 Ref=360,100 (201310-1\J11025A1.D)

=====
 Data File C:\HPCHEM\1\DATA\201310-1\J11025A1.D
 =====

=====
 Area Percent Report
 =====



Data File C:\HPCHEM\1\DATA\201310~1\J11025B1.D

Sample Name: JC11025B

Data File C:\HPCHEM\1\DATA\201310~1\J11025B1.D

Sample Name: JC11025

===== Injection Date : 10/22/2013 5:42:45 PM Seq. Line : 9
===== Cntr No.: rc11025b Tocatn: vial 10

```

sample_name : UCL1042
Acq_ Operator : MK
Acq_ Instrument : Instrument 1
Different Inj Volume from Sequence ! Actual
Acq_ Method : C:\HPCHEM\METHODS\VAD005-40.M

```

Sorted BY Multiplier	:	Signal 1.0000
Dilution	:	1.0000

Last changed : 8/2/2014 12:21:07 AM by M
(modified after loading)

Signal 1: DAD1 A, Sig=250, 10 Ref=360, 10

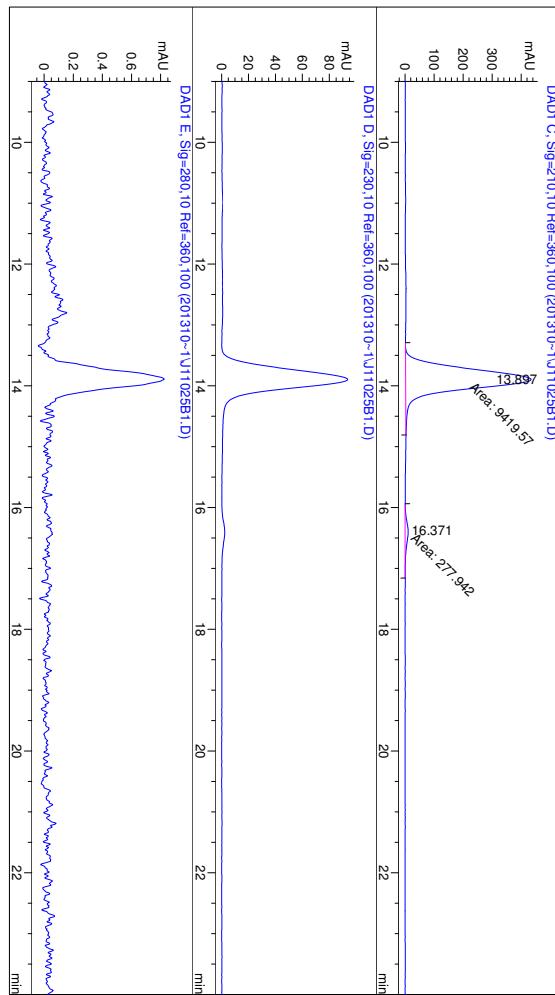
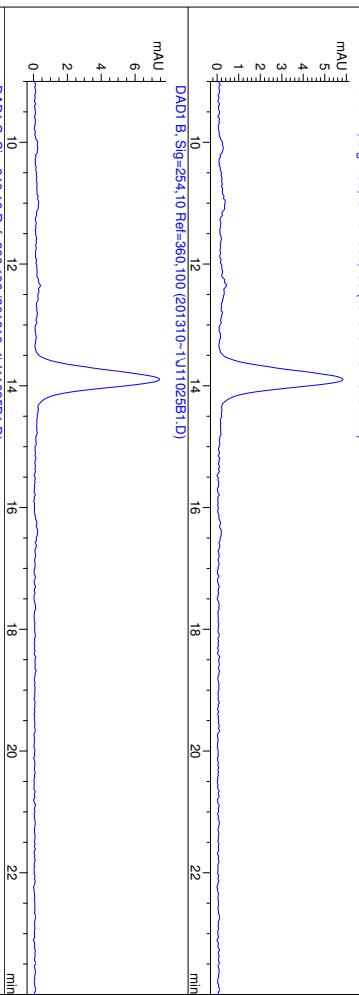
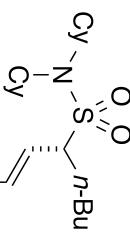


Table 5, entry 3
with $(3S,8R)$ -L6



Signal 2: DAD1_B, Sig=254,10 Ref=360,100
 Signal 3: DAD1_C, Sig=210,10 Ref=360,100

#	[min]	[min]	[mAU*s]	[mAU]
1	13.897	MM	0.3334	9.619.56641
2	16.371	MM	0.4602	27.7.94183
				10.06597

Totals : 9697.50824 442
Results obtained with enhanced integration

Signal 4: DAD1 D, Sig=230, 10 Ref=360, 100

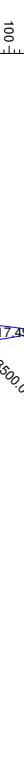
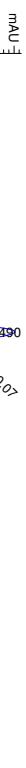
Signal 5: DAD1 E, Sig=280, 10 Ref=360, 100

— — — — —

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Injection Date : 9/16/2013 1:56:44 PM Seq. Line : 9
 Sample Name : JC10279B Location : Vial 95
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 1 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 1 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\AD-10-40.M
 Last changed : 7/12/2012 11:12:25 AM by CE
 Analysis Method : C:\HPCHEM\1\METHODS\AD005-40.M
 Last changed : 8/3/2014 12:46:07 AM by MK
 (modified after loading)

DAD1 A, Sig=250,10 Ref=360,100 (201309-1\JC10279B.D)



Results obtained with enhanced integrator!

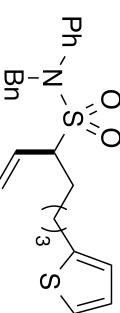
Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.490	MM	0.4828	3500.07324	120.82352	90.0647
2	23.842	MM	0.6752	386.10425	9.53050	9.9353
Totals :				3886.17749	130.35402	

Results obtained with standard integrator!

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.490	MM	0.4828	3500.07324	120.82352	90.0647
2	23.842	MM	0.6752	386.10425	9.53050	9.9353
Totals :				3886.17749	130.35402	



**Table 5, entry 4
with (3*R*,8*S*)-**L6****

Peak RetTime Type Width Area Height Area
[min] [min] [mAU*s] [mAU] %
1 17.490 MM 0.4828 3500.07324 120.82352 90.0647
2 23.842 MM 0.6752 386.10425 9.53050 9.9353

Results obtained with enhanced integrator!
 Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.492	MM	0.4910	1.55605e4	528.16541	90.1217
2	23.818	MM	0.6559	1705.25134	43.32966	9.8765

Totals : 1.72657e4 571.49507
 Results obtained with enhanced integrator!
 Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.491	MM	0.4814	1.00163e4	346.80835	90.2137
2	23.839	MM	0.6694	1086.56787	27.05270	9.7863

Totals : 1.11029e4 373.86105
 Results obtained with standard integrator!
 Signal 5: DAD1 E, Sig=280,10 Ref=360,100

=====
*** End of Report ***

Injection Date : 2/5/2014 11:58:40 PM Seq. Line : 2
 Sample Name : JC11123 Location : Vial 31
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\IB-05-40.M Dilution :
 Last changed : 7/6/2013 9:54:10 AM by CE Analysis Method : C:\HPCHEM\1\METHODS\OD-02-20.M
 Last changed : 7/30/2014 10:42:05 PM by MK (modified after loading)

DAD1, Sig=250,10 Ref=360,100 (201402-1\JC11123.D)



DAD1 B, Sig=254,10 Ref=360,100 (201402-1\JC11123.D)



mAU



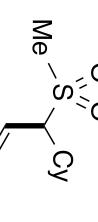
mAU



mAU



mAU



**Table 5, entry 6
with (*R*,*R*)-L1**

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.030	MF	0.2092	10.729430	8.54649	2.9057
2	22.909	MF	0.4974	38.85.25415	120.13020	97.0943

Totals : 4.48962e4 1316.61627

Results obtained with enhanced integrator!

Signal 4: DAD1, Sig=230,10 Ref=360,100

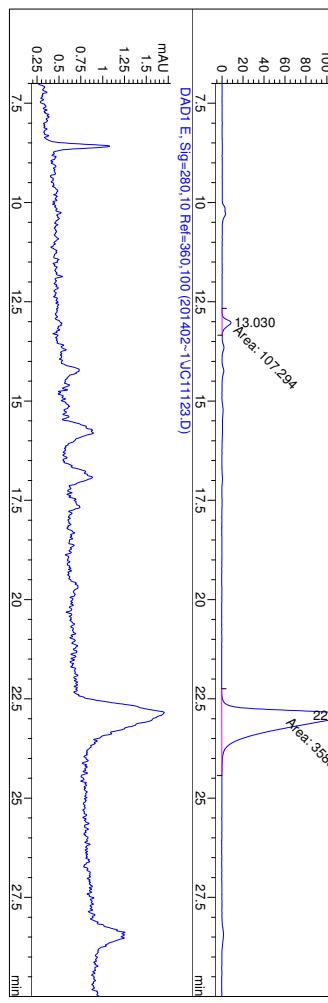
Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.030	MF	0.2092	10.729430	8.54649	2.9057
2	22.909	MF	0.4974	38.85.25415	120.13020	97.0943

Totals : 3692.54845 128.67670

Results obtained with enhanced integrator!

Signal 5: DAD1, Sig=280,10 Ref=360,100

***** End of Report *****



mAU

Data File C:\HPCHEM\1\DATA\201402-1\JC11113A.D

Sample Name: JC11113

Data File C:\HPCHEM\1\DATA\201402-1\JC11113A.D

Sample Name: JC11113

Injection Date : 2/6/2014 12:39:55 AM Seq. Line : 3
 Sample Name : JC11113 Location : Vial 32
 Acq. Operator : MK Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 μ l
 Different Inj Volume From Sequence ! Actual Inj Volume : 5 μ l
 Acq. Method : C:\HPCHEM\1\METHODS\IB-05-40.M Dilution :
 Last changed : 7/6/2013 9:54:10 AM by CE C:\HPCHEM\1\METHODS\AD005-40.M
 Analysis Method : 8/2/2014 12:13:33 AM by MK Last changed :
 (modified after loading)

DAD1 A, Sig=250,10 Ref=360,100 (201402-1\JC11113A.D)

mAU



mAU



mAU



mAU



mAU



mAU



mAU



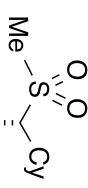
mAU



mAU



mAU

CH₂PhTable 5, entry 6
with (S,S)-L1

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.829	MF	0.2556	12.9761914	96.1899	
2	23.784	FM	0.3747	955.53381	42.49660	3.8101

Totals :

2.50787e4 1340.11574

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	Retention Time [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.830	MF	0.2556	1843.23779	120.16679	97.0832
2	23.795	FM	0.3695	55.37868	2.49811	2.9168

Totals : 1898.61647 122.66490

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

***** End of Report *****

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