

Supplementary data

Arabidopsis membrane-associated acyl-CoA-binding protein ACBP1 is involved in stem cuticle formation

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Supplementary Figure S1.

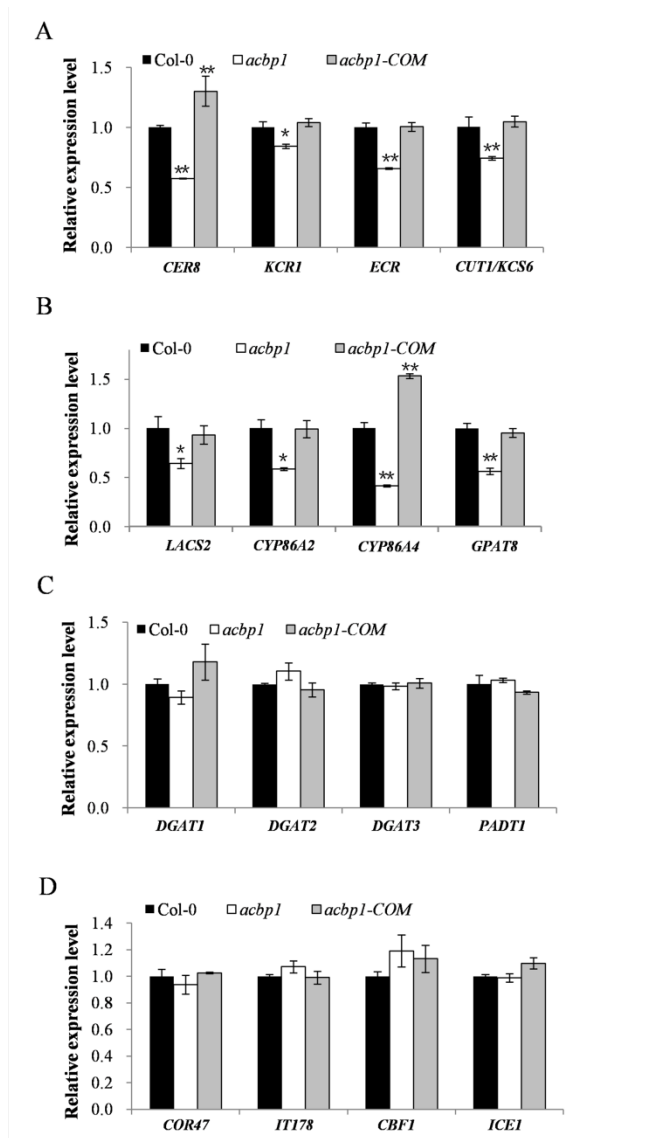


Figure S1. Expression analysis of wax biosynthetic genes (A), cutin biosynthetic genes (B), and genes with no implication on cuticle formation (triacylglycerol biosynthetic genes (C) and cold-related genes (D)) in stems of Col-0, the *acbp1* mutant and *acbp1-COM* line. Asterisks denote significant differences from the wild type (* $P < 0.05$, ** $P < 0.01$). Values are means \pm SE (n = 3).

Supplementary Figure S2.

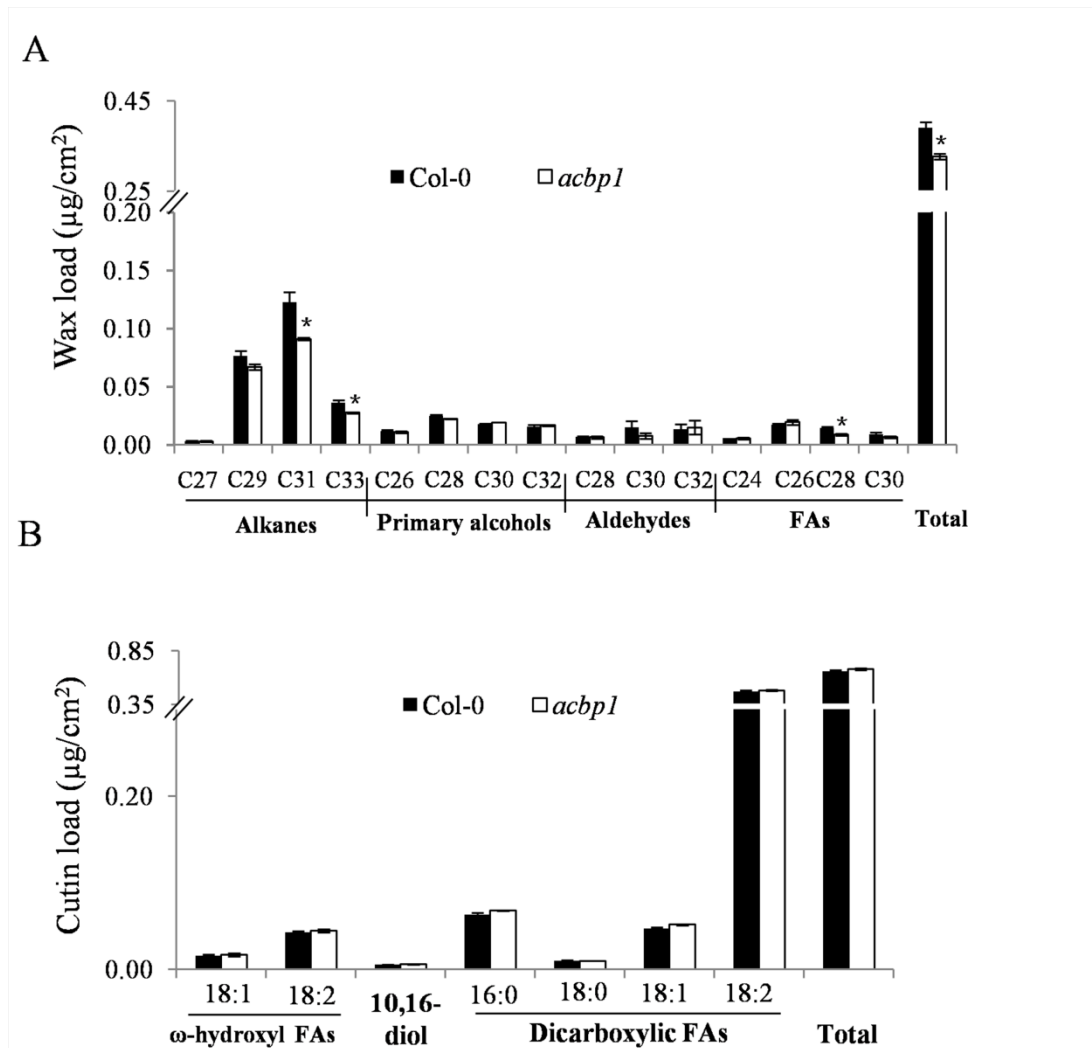


Figure S2. Cuticular wax (A) and cutin monomer (B) composition and amount in leaves of Col-0 and the *acbp1* mutant. Four-week-old leaves were used for wax and cutin analysis by gas chromatography (GC)-flame ionization detector and GC-mass spectrometry. FA, fatty acid; 10,16-diol, C16-10,16-dihydroxyl fatty acids. Asterisks denote significant differences from the wild type (* $P < 0.05$). Values are means \pm SE (n = 3).

Supplementary Table S1. Sequences of gene-specific primers for qRT-PCR.

Name	Sequence (5'→3')	Target gene
ML1124	CCCGCTATGTATGTCGC	At3g18780 (<i>ACTIN2</i>)
ML1125	AAGGTCAAGACGGAGGAT	
ML1824	AGGCAAACATCACCACAATAGG	At1g68530 (<i>CUT1/KCS6</i>)
ML1825	CTCGTCGATCACCGCTCTG	
ML1826	AGCACCTTTGAGCCCTGAGA	At2g47240 (<i>CER8</i>)
ML1827	AACCGCTGGAATACCGACTG	
ML1828	ATTCCACGCGGTTTCCTCTTC	At3g55360 (<i>ECR</i>)
ML1829	GGCTTTCCATCTTTTCCATCAA	
ML1832	ACCCACTTGGCTCCTCATCTC	At1g67730 (<i>KCRI</i>)
ML1833	GCTTTACCGATTCCGCTCTGTTG	
ML1917	TAATACACGTAAGCAAGAAT	At4g00360 (<i>CYP86A2</i>)
ML1918	GTCGTGATGAGCCAGAAAAACC	
ML1919	GGCTCTTCCAACGCAAAAACC	At1g01600 (<i>CYP86A4</i>)
ML1920	CGAACGCCAATCCGCAAATG	
ML1933	GATTGTGCCAGTAGCGATGAA	At4g00400 (<i>GPAT8</i>)
ML1934	TCTTGCCACCACCGTTGAC	
ML1937	AACGCAGCAATTTCCGGTCCAG	At1g49430 (<i>LACS2</i>)
ML1938	AATGTCGCCACGCCAGTATCC	
ML2026	GGAGGGCGAGAGAGAGTCCACTT	At2g19450 (<i>DGAT1</i>)
ML2027	CCAATCTCGCAGCGATCTGAAC	
ML2028	CCGAGCTGAGGAACATTCAAATCA	At3g51520 (<i>DGAT2</i>)
ML2029	CGACCATATTTGCTACGATGATCGA	
ML2030	GGTCTTGATCCTGAAGCTGGTTTG	At1g48300 (<i>DGAT3</i>)
ML2031	TCAGTCATCTTCTTCATGGCTTTGG	
ML2032	TGCTGAAGCAAAGGATGTTGCA	At5g13640 (<i>PDAT1</i>)
ML2033	CCTTCTCCGGTGACCAATCAAG	
ML1716	GTCCACGCCGTTGGTTGT	At1g20440 (<i>COR47</i>)
ML1717	AATCCCCTTCTTCTCCTCCG	
ML1718	TTCCACCAGGGACAAAGG	At5g52310 (<i>LTI78</i>)
ML1719	CATCGTGTCCGTAAGAGGC	
ML1785	GGACTTCCAACCGCTGAG	At4g25490 (<i>CBF1</i>)
ML1786	CAAGCCGAGTCAGCGAAGTT	
ML1901	GATTCTTGCTGCTCGGTCACT	At3g26744 (<i>JCE1</i>)
ML1902	CCATTAGCAGGACTACCAAAAACC	

The genes implicated in wax biosynthesis tested in qRT-PCR were *CUT1/KCS6*, *CER8*, *ECR* and *KCRI*. The genes implicated in cutin biosynthesis tested were

CYP86A2, *CYP86A4*, *GPAT8* and *LACS2*. The genes with no implication in cuticle formation tested included triacylglycerol biosynthetic genes (*DGAT1*, *DGAT2*, *DGAT3* and *PADT1*) and cold-related genes (*COR47*, *LTI78*, *CBF1* and *ICE1*).

Supplementary Table S2. Mass-to-charge ratios (m/z) of cutin compounds in mass spectrometry.

Cutin compound	Mass-to-charge ratios (m/z)
C17:0-FA (internal standard)	284
C15:0 cycloketone (internal standard)	282
C18:1-FA	296
C18:2-FA	294
C10,16-dihydroxyl fatty acids	343
C16:0-DCA	314
C18:0-DCA	342
C18:1- DCA	339
C18:2-DCA	337

FA, fatty acid; DCA, dicarboxylic fatty acid.