

CHEMBIOCHEM

Supporting Information

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Cellular Internalisation of an Inositol Phosphate Visualised by Using Fluorescent InsP₅

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cbic_201300583_sm_miscellaneous_information.pdf

Supporting Information

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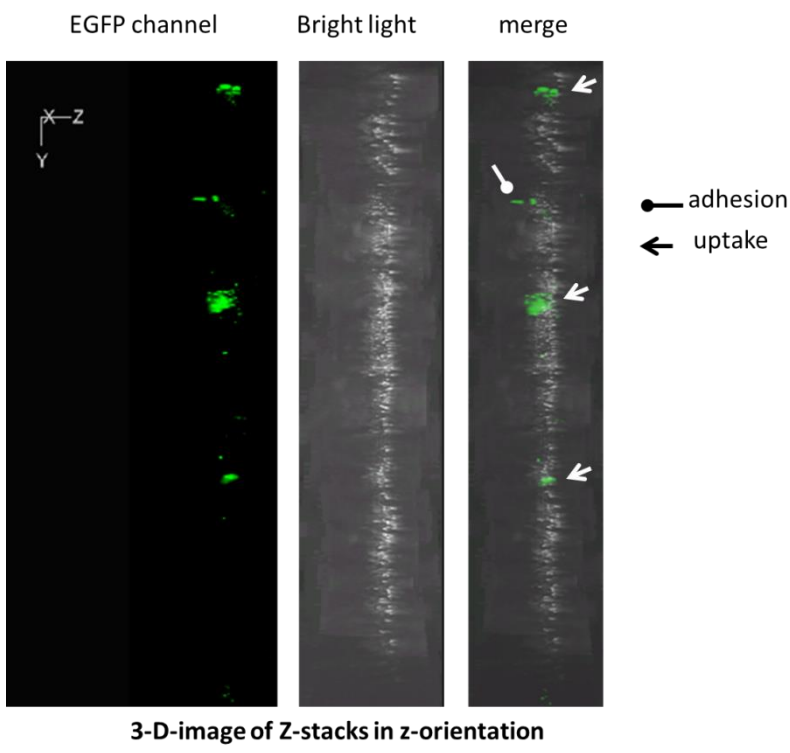
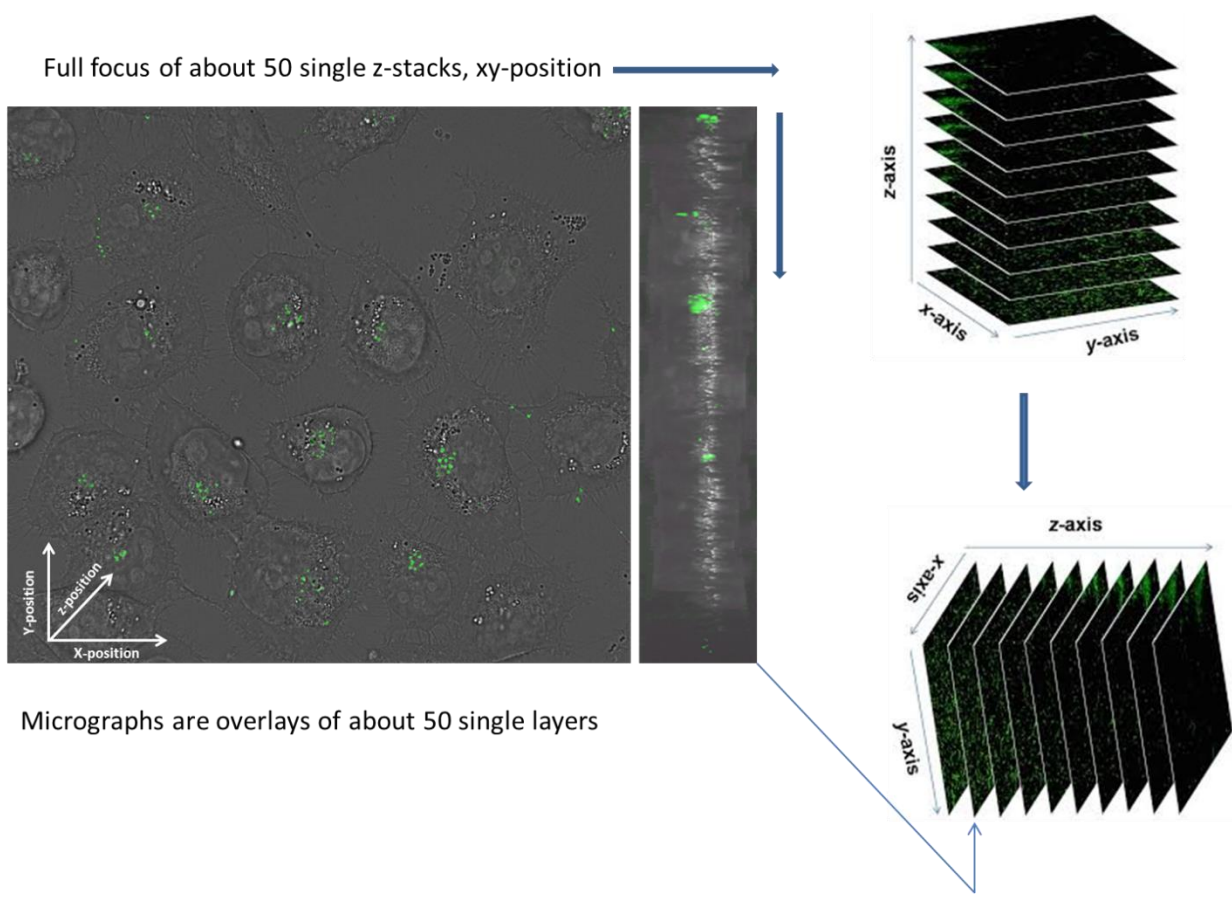


Figure S1. H1299 cells were treated with **5** (20 μM) for 16 h. Washed cells were fixed and analysed by fluorescence microscopy. Z-stacks and 3D-reconstitutions of Z-stacks were performed and uptake of **5** was analysed in micrographs that were rotated in the Z-axis by 90°. Green dots at the outer layer were defined as adhesion (● —) and dots in the middle of the cell layer as uptake (◀).

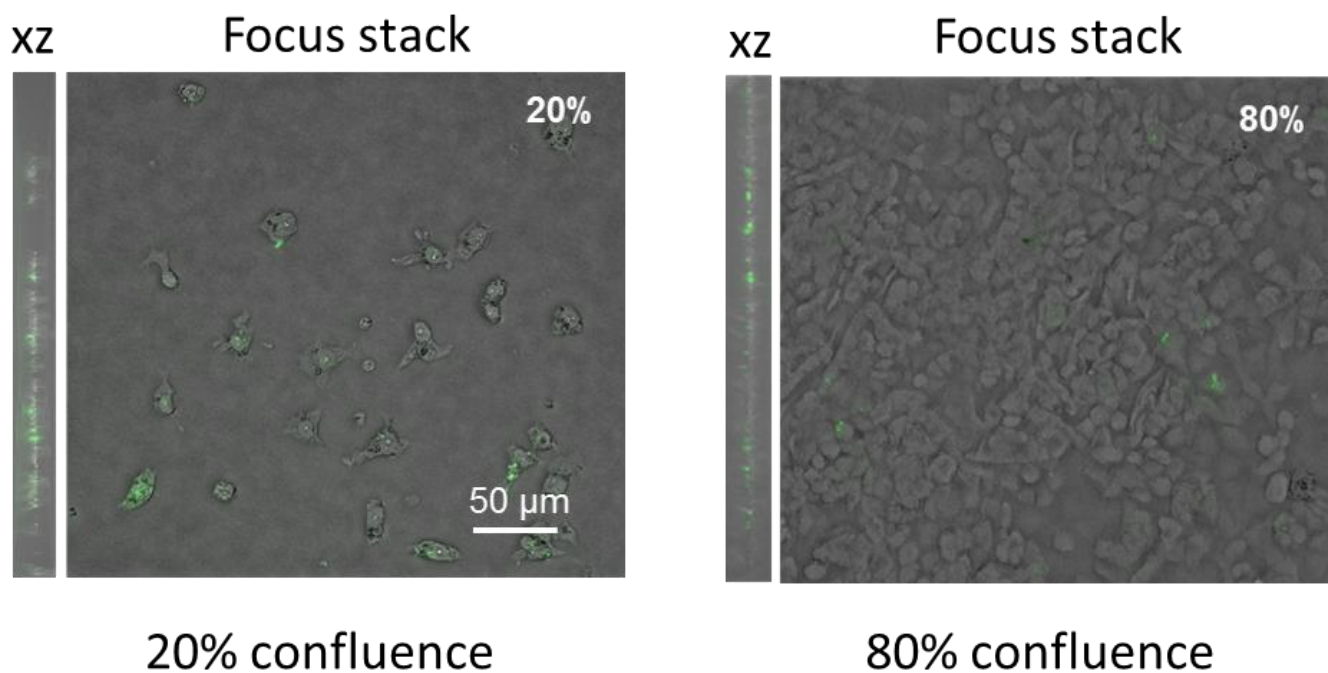


Figure S2. Uptake of FAM-InsP₅ (**5**) in H1299 cells. H1299 cells were grown to 20% or 80% confluence and treated with **5** (20 μM). After 16 h, washed cells were with paraformaldehyde and embedded in Fluoromount-G medium. Z-stacks of bright and fluorescence light micrographs were performed. Shown are the xz-layer and focus stackings of bright and fluorescence light overlays in a magnification of 10 x 20. Five micrographs per series (20% or 80% confluence) were performed and whole cell numbers and numbers of green fluorescent cells of the middle cell layers were counted to calculate % cellular uptake of **5**.

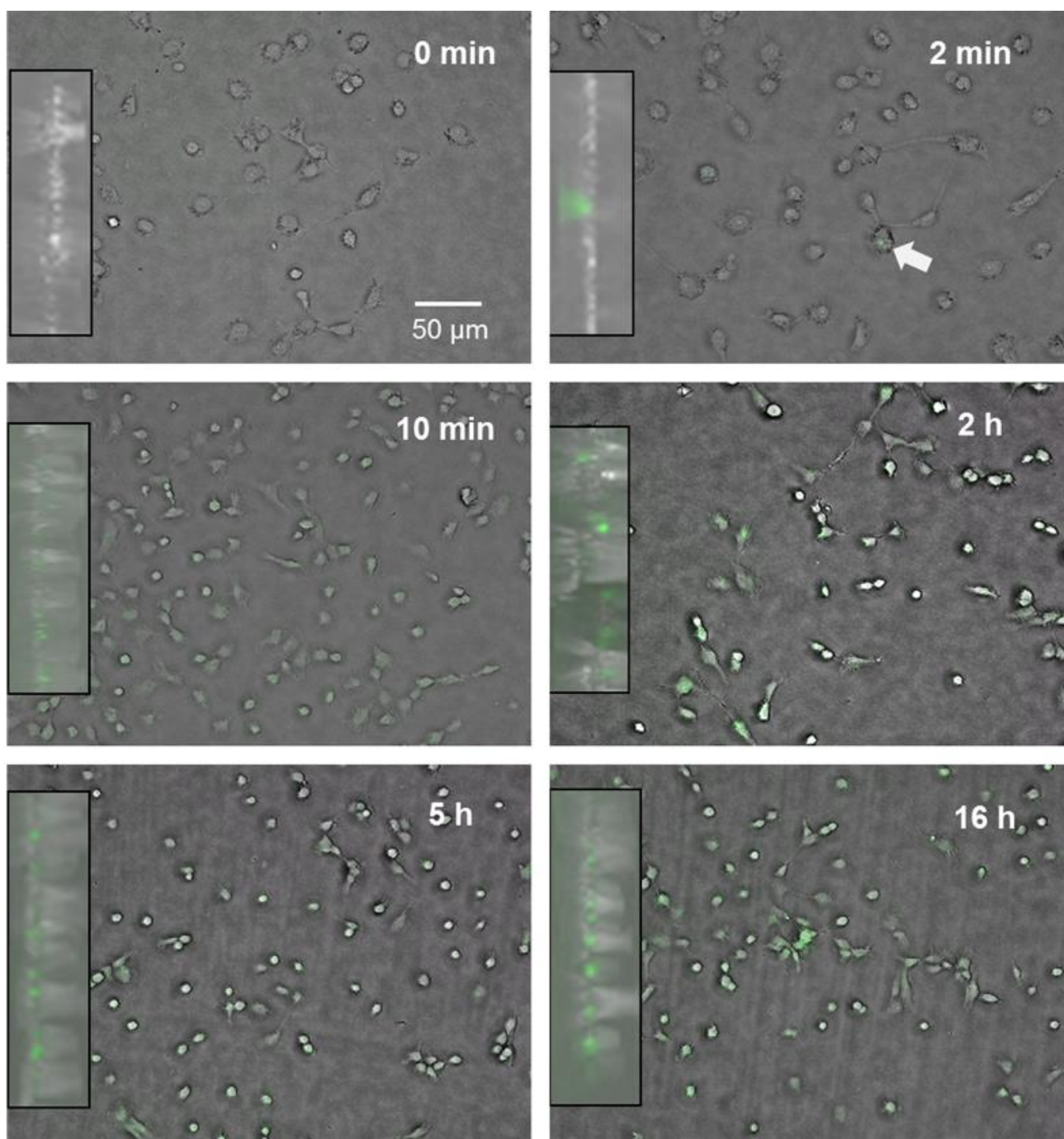
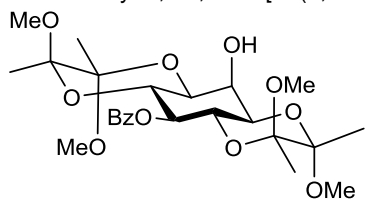
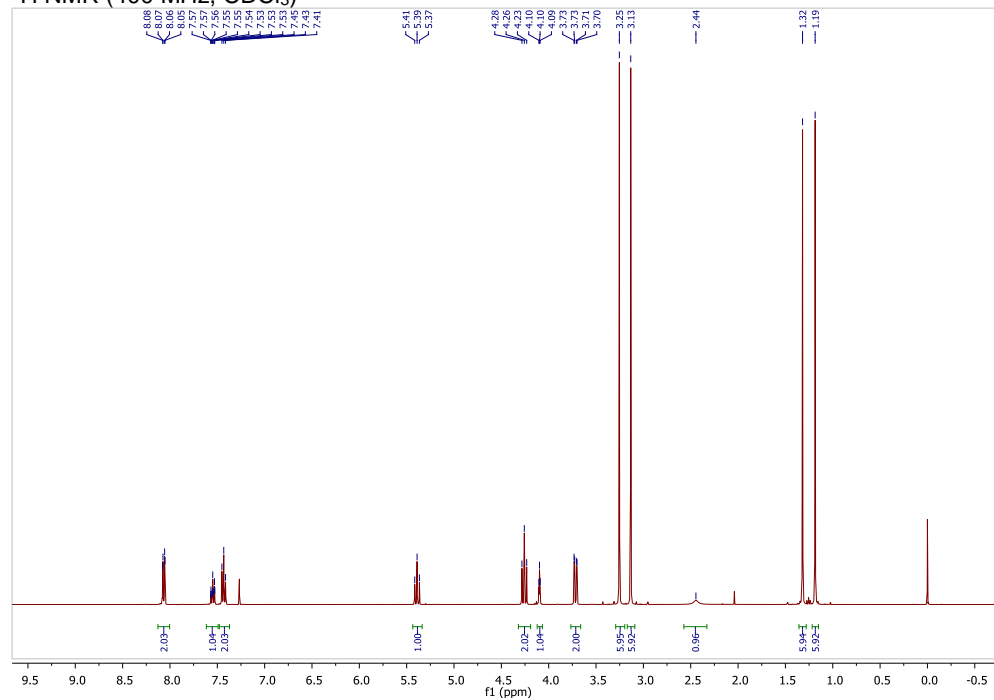


Figure S3. Time dependence of FAM-InsP₅ (**5**) uptake into H1299 cells. H1299 cells were grown to 50% confluence and treated with 20 μM **5** for the time points as indicated in the figure. Then, the washed cells were embedded in Fluoromount-G medium and uptake of **5** was determined as described above. To illustrate uptake over time, the xz-layers of representative micrographs are shown in higher magnification (left panels). After 2 min of incubation, only cell surface aggregates of **5** were visible (indicated by an arrow) and after 10 min, the first intracellular aggregates of **5** were detected. For each time point, at least five micrographs were evaluated, and % cellular uptake of **5** was calculated as before.

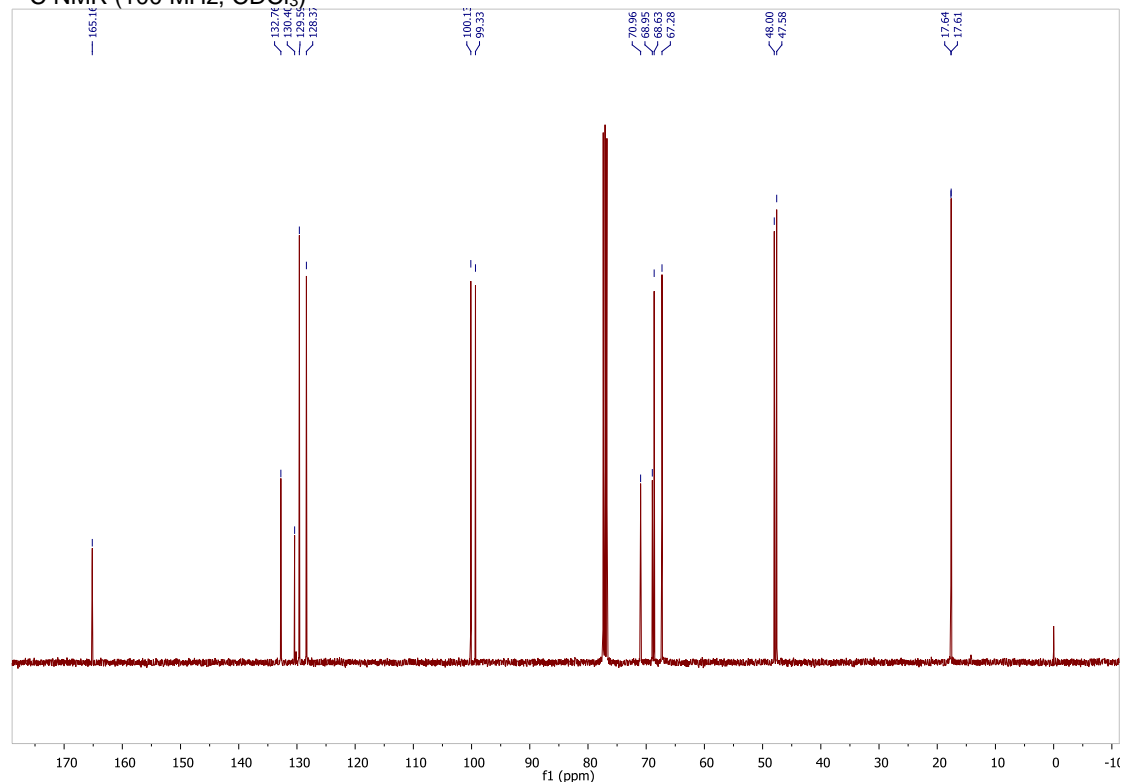
5-O-Benzoyl-1,6:3,4-bis-[O-(2,3-dimethoxybutane-2,3-diyl)]-myo-inositol (7)



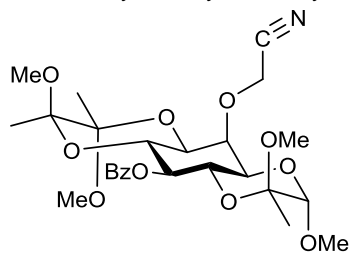
¹H NMR (400 MHz, CDCl₃)



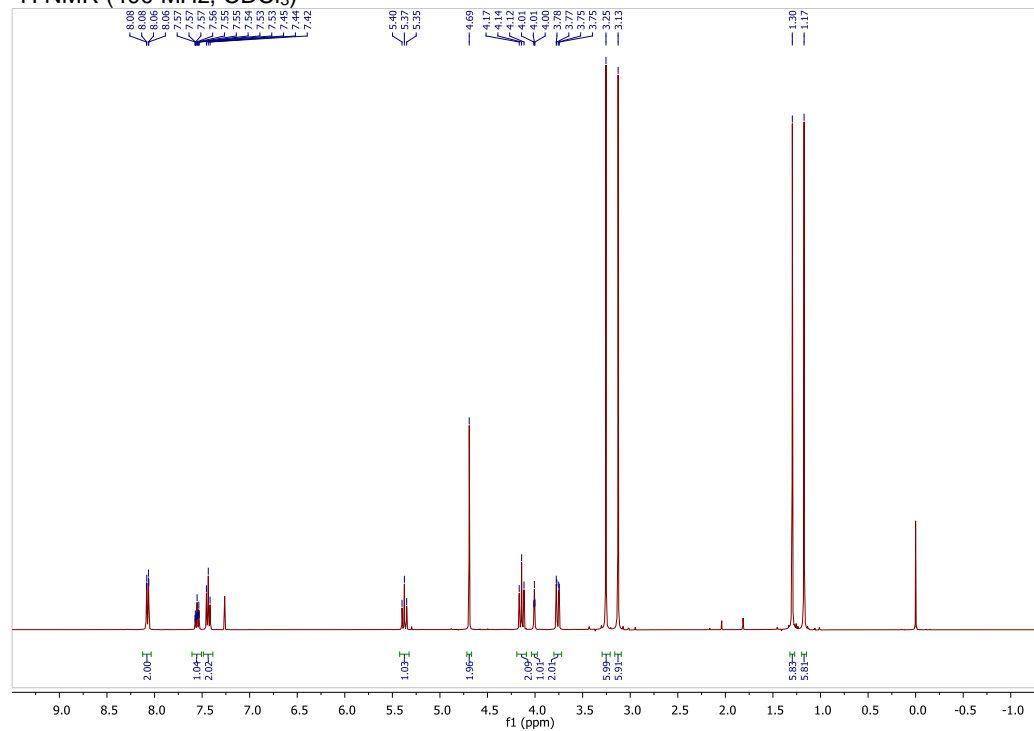
¹³C NMR (100 MHz, CDCl₃)



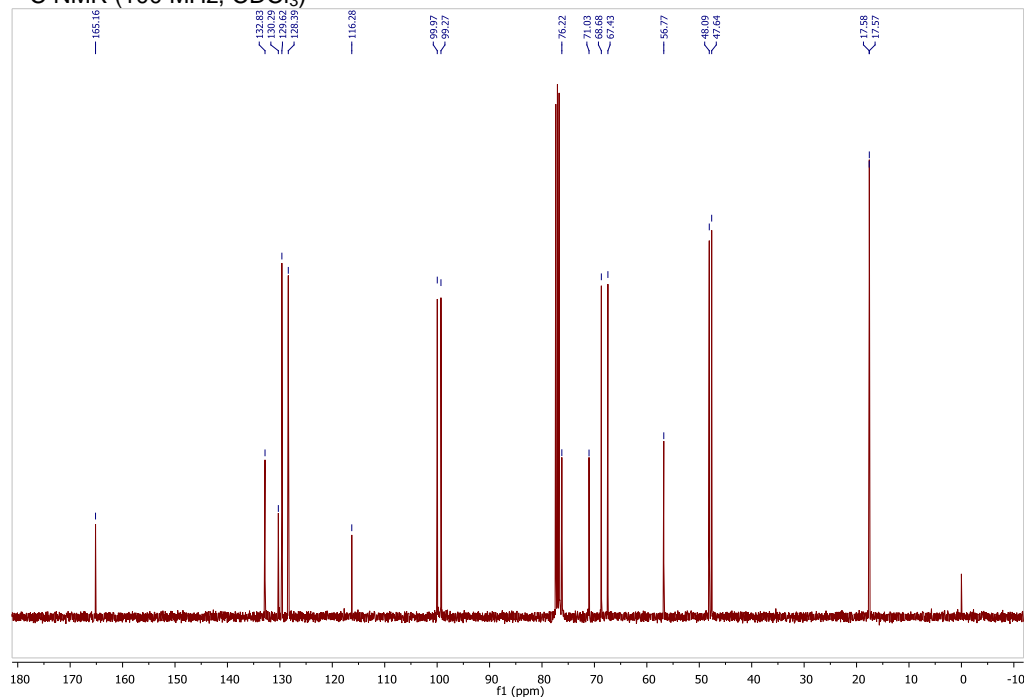
5-O-Benzoyl-2-O-cyanomethyl-1,6:3,4-bis-[O-(2,3-dimethoxybutane-2,3-diyl)]-myo-inositol (**8**)



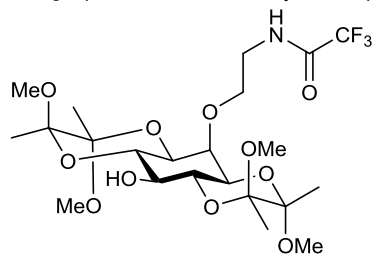
$^1\text{H NMR}$ (400 MHz, CDCl_3)



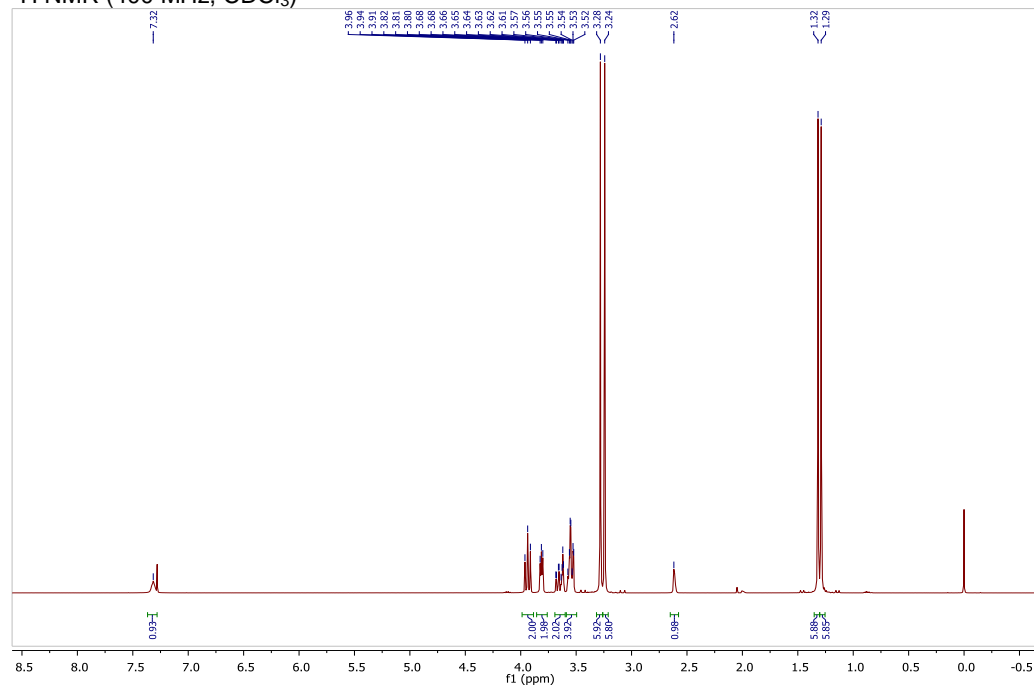
$^{13}\text{C NMR}$ (100 MHz, CDCl_3)



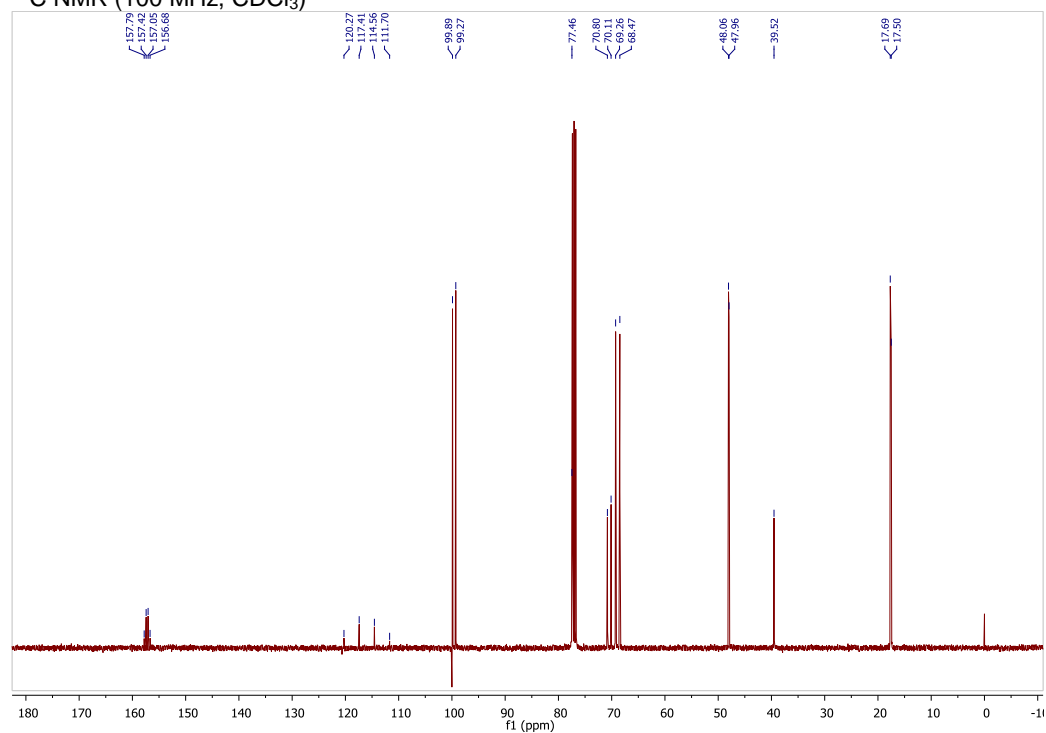
2-O-[2-(2,2,2-Trifluoroacetyl)amino]ethyl]-1,6:3,4-bis-[O-(2,3-dimethoxybutane-2,3-diy)]-myo-inositol (**9**)



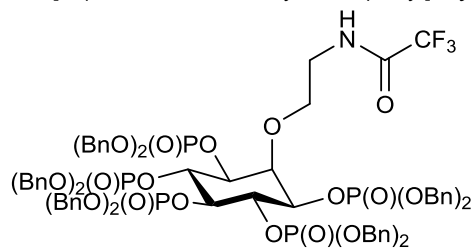
¹H NMR (400 MHz, CDCl₃)



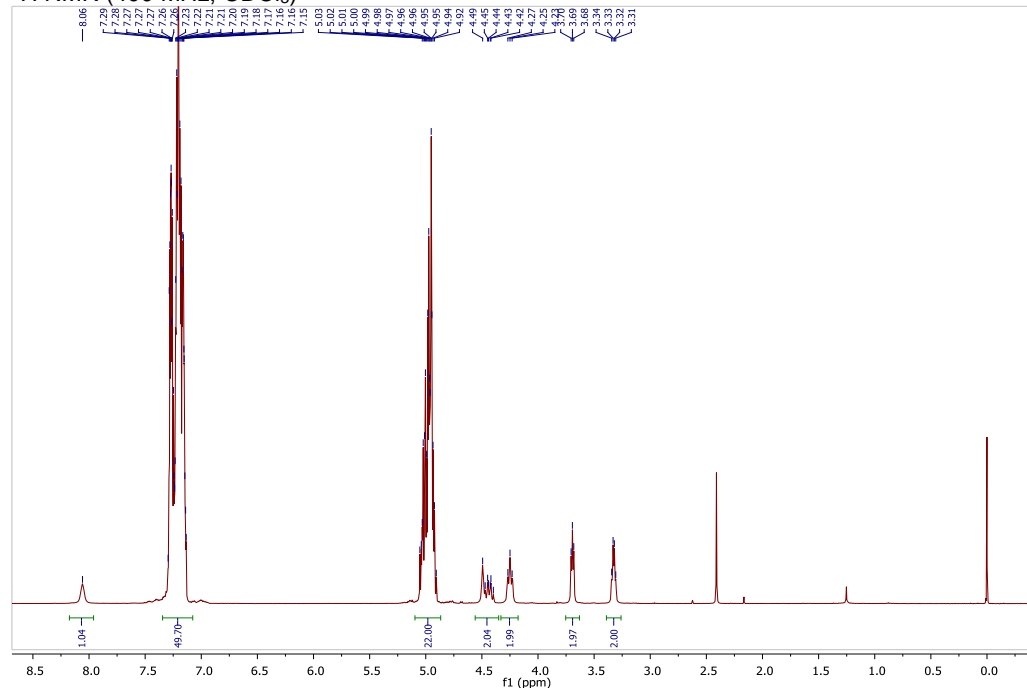
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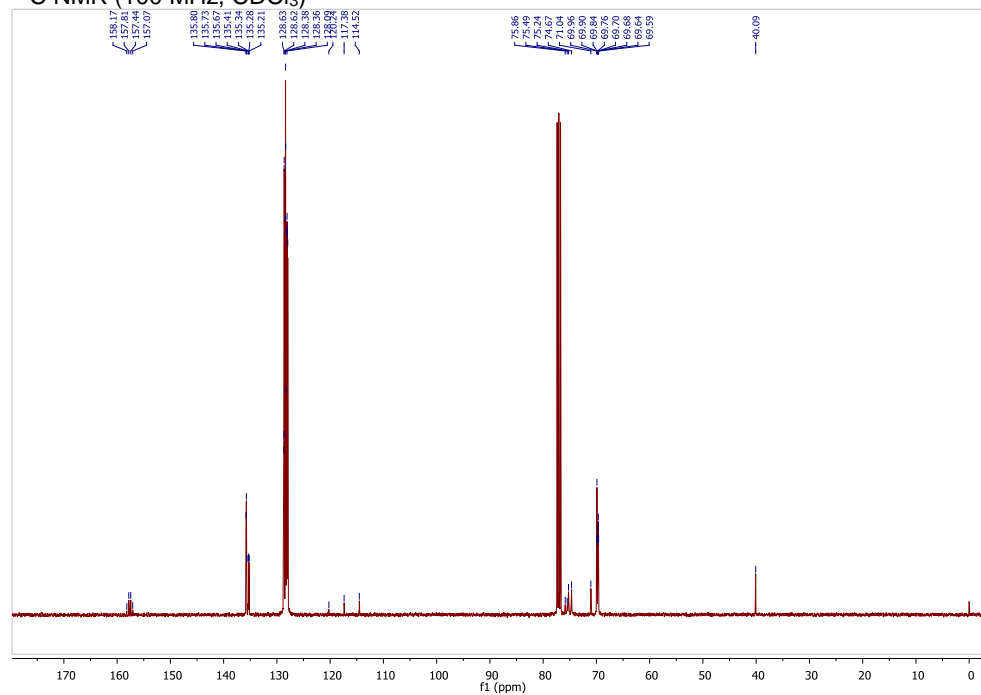
2-O-[2-(2,2,2-Trifluoroacetyl)amino]ethyl]-*myo*-inositol 1,3,4,5,6-pentakis (dibenzylphosphate) (**10**)



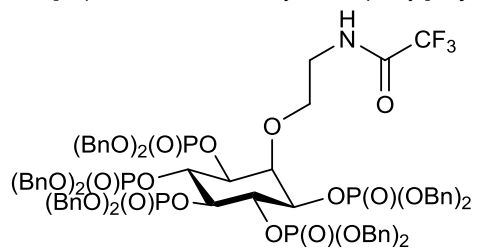
¹H NMR (400 MHz, CDCl₃)



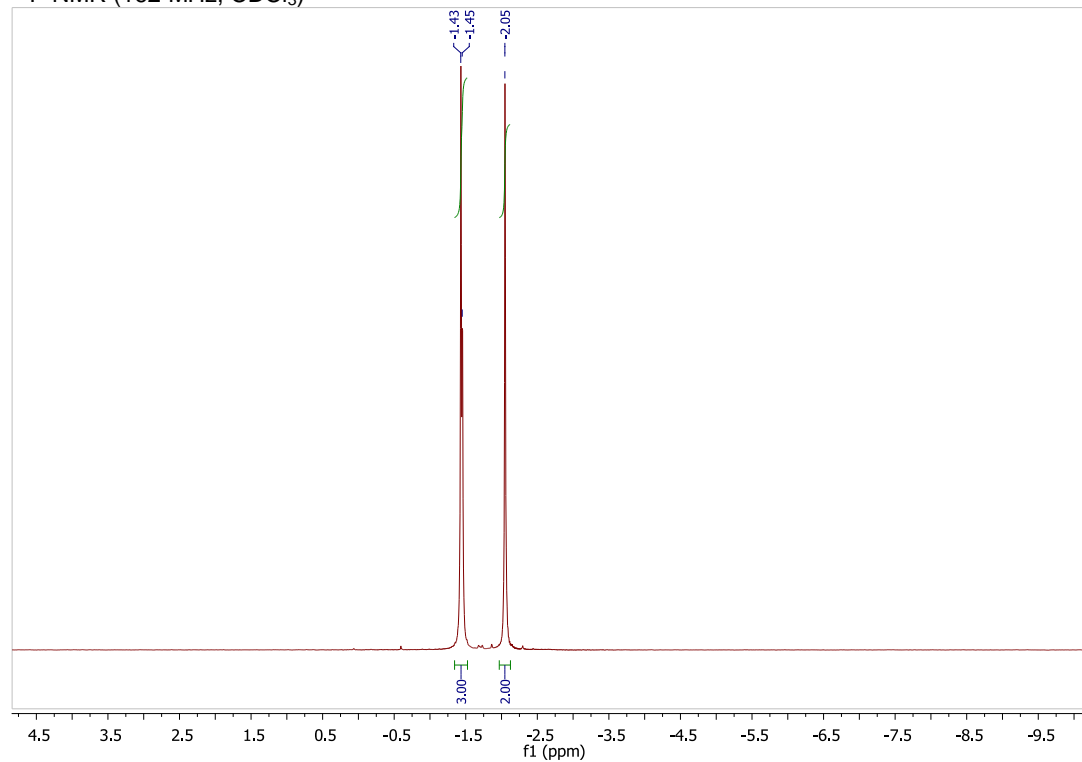
¹³C NMR (100 MHz, CDCl₃)



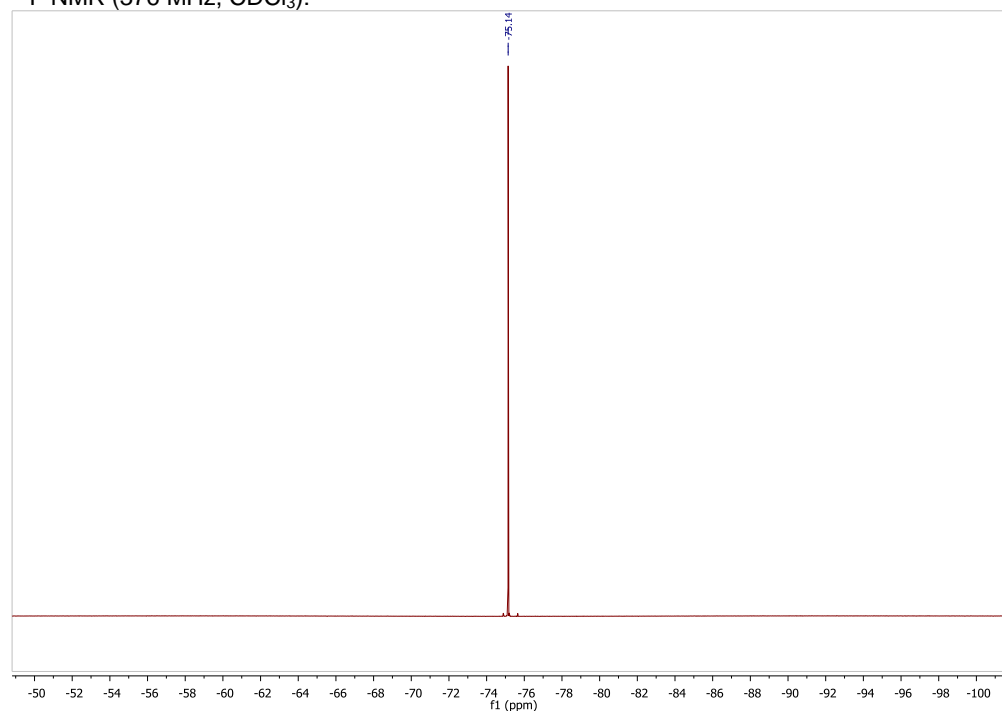
2-O-[2-(2,2,2-Trifluoroacetyl)amino]ethyl]-*myo*-inositol 1,3,4,5,6-pentakis (dibenzylphosphate) (**10**)



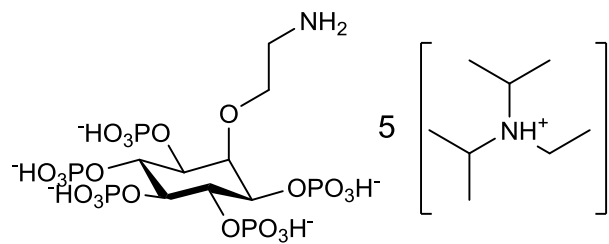
^{31}P NMR (162 MHz, CDCl_3)



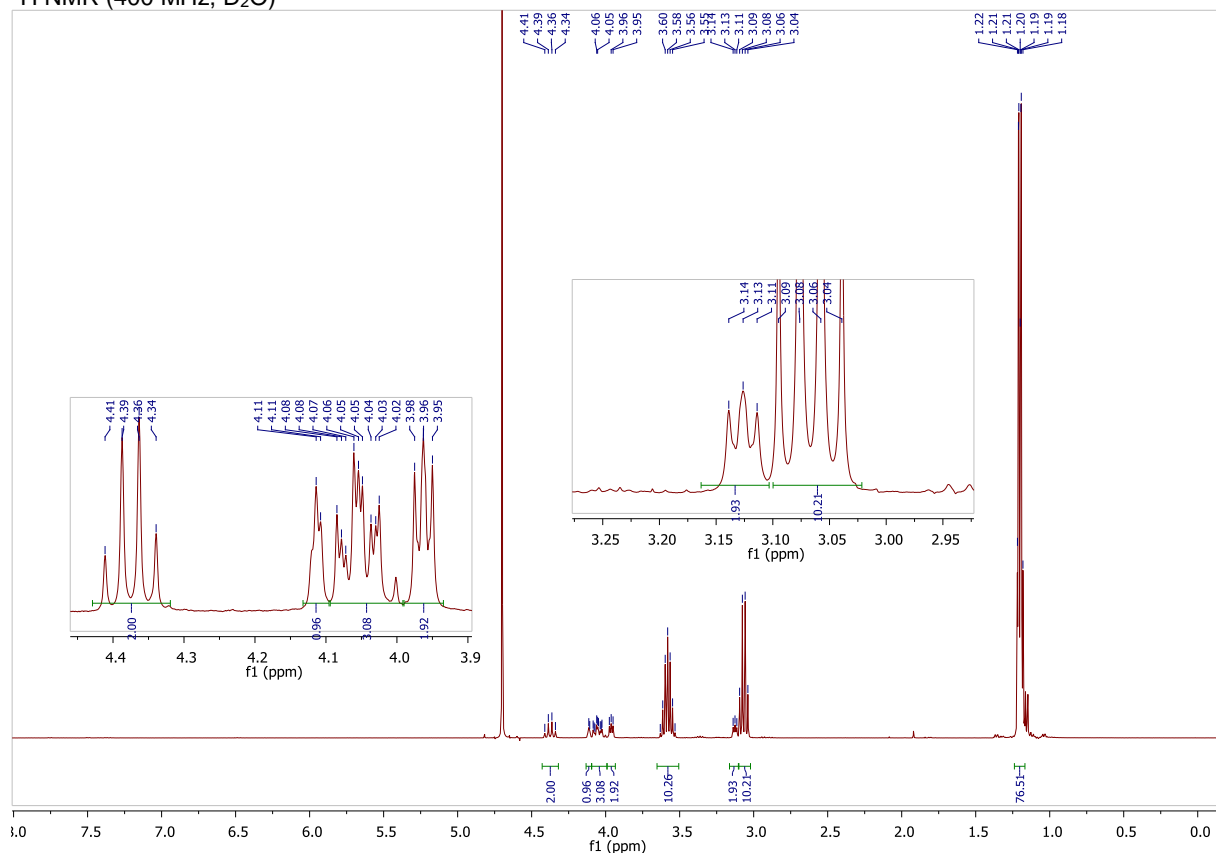
^{19}F NMR (376 MHz, CDCl_3):



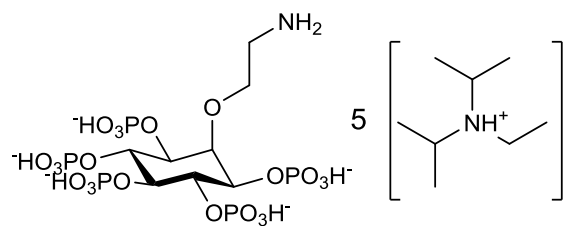
2-O-(2-Aminoethyl)-*myo*-inositol 1,3,4,5,6-pentakisphosphate (4)



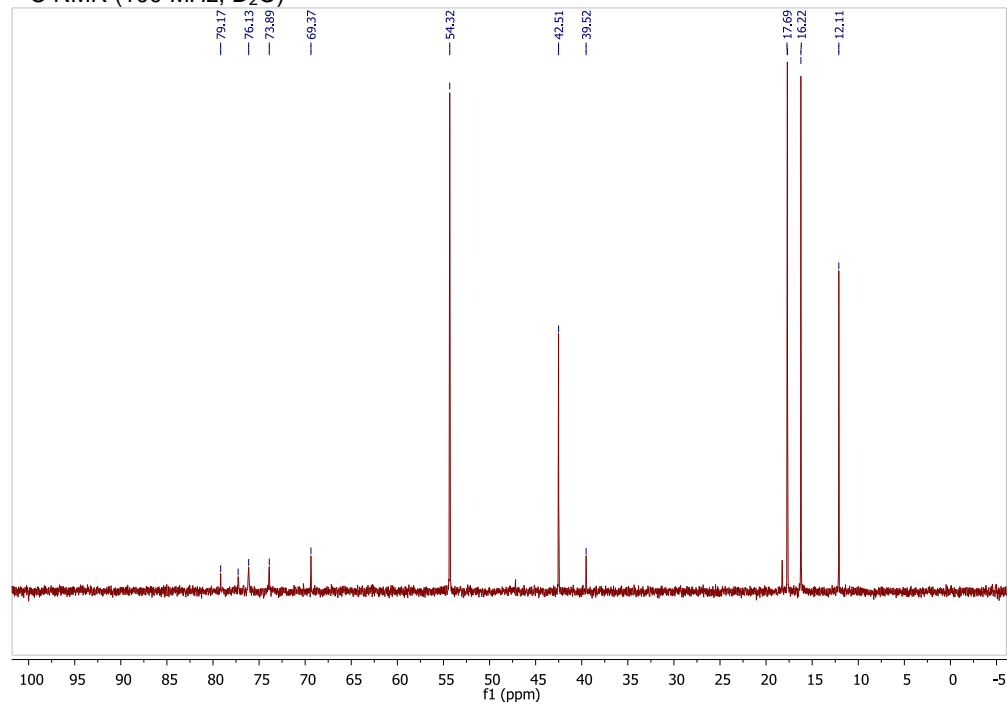
^1H NMR (400 MHz, D_2O)



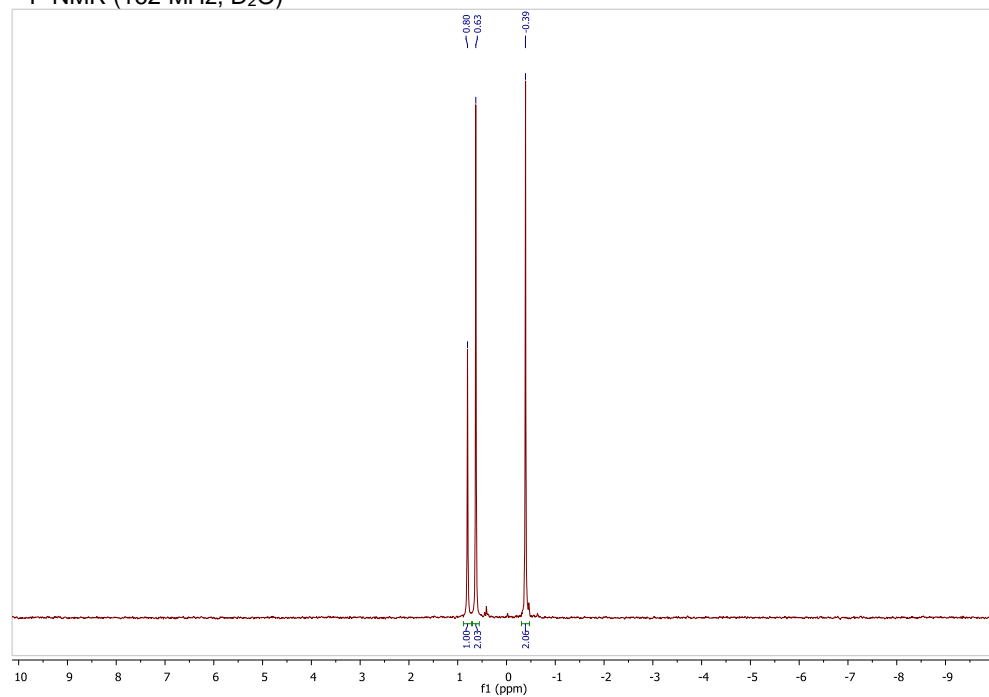
2-O-(2-Aminoethyl)-*myo*-inositol 1,3,4,5,6-pentakisphosphate (4)



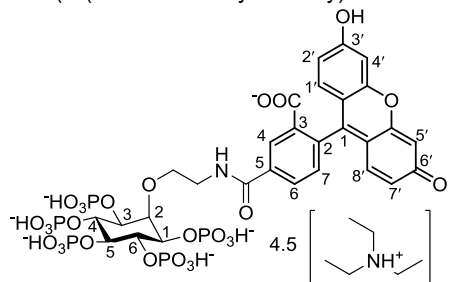
^{13}C NMR (100 MHz, D_2O)



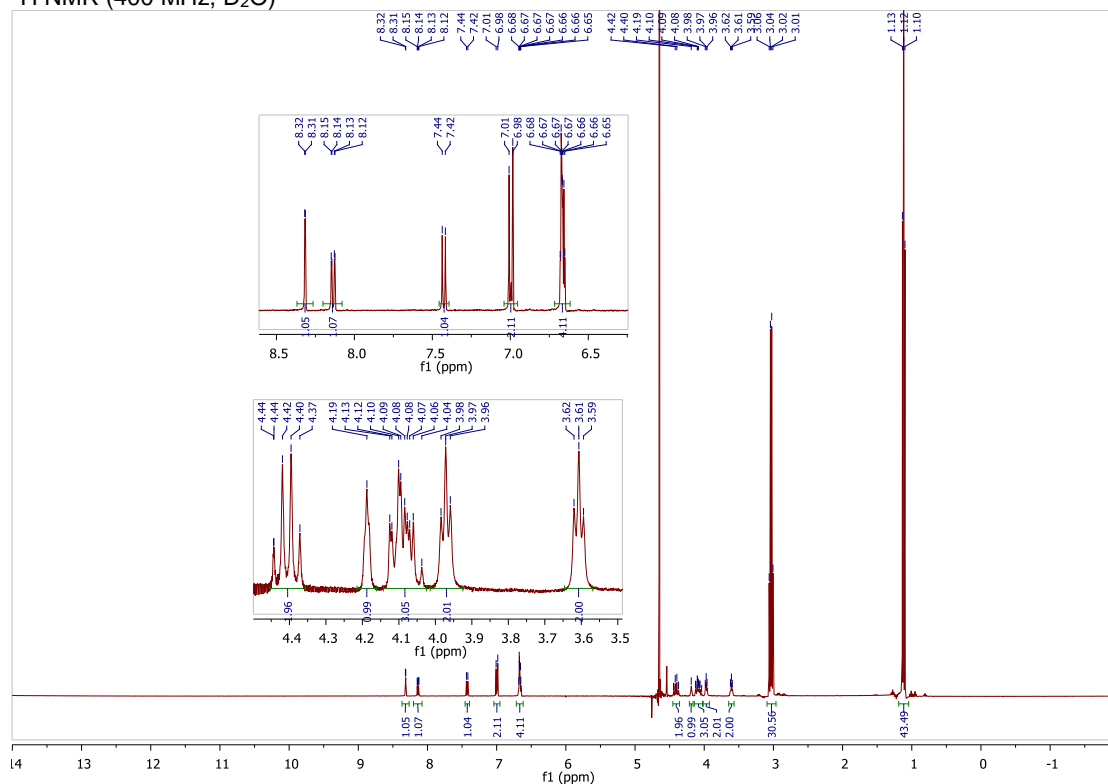
^{31}P NMR (162 MHz, D_2O)



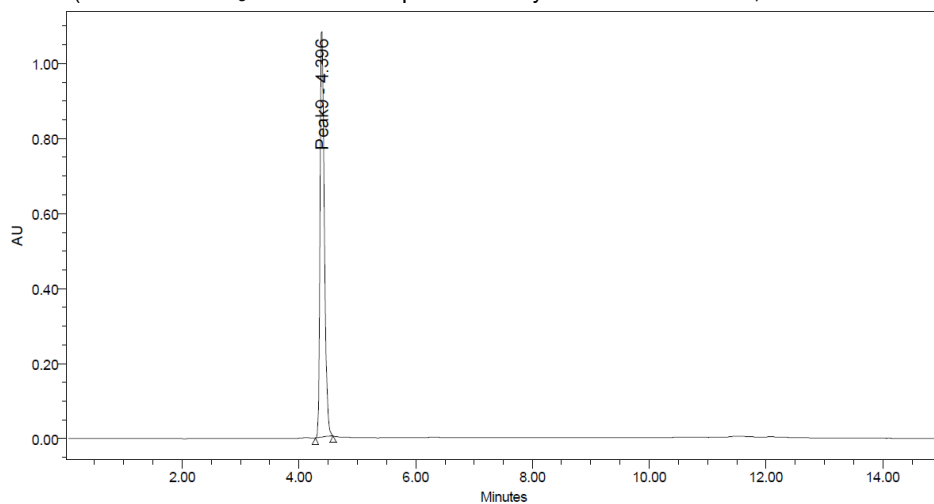
2-O-(2-(5-Fluoresceinylcarboxy)-aminoethyl)-myo-inositol 1,3,4,5,6-pentakisphosphate (5)



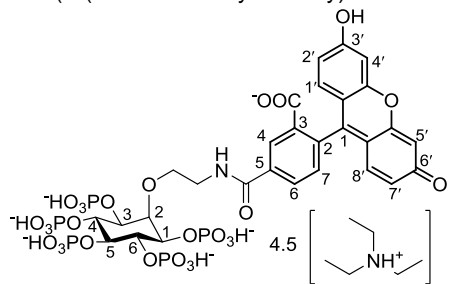
^1H NMR (400 MHz, D_2O)



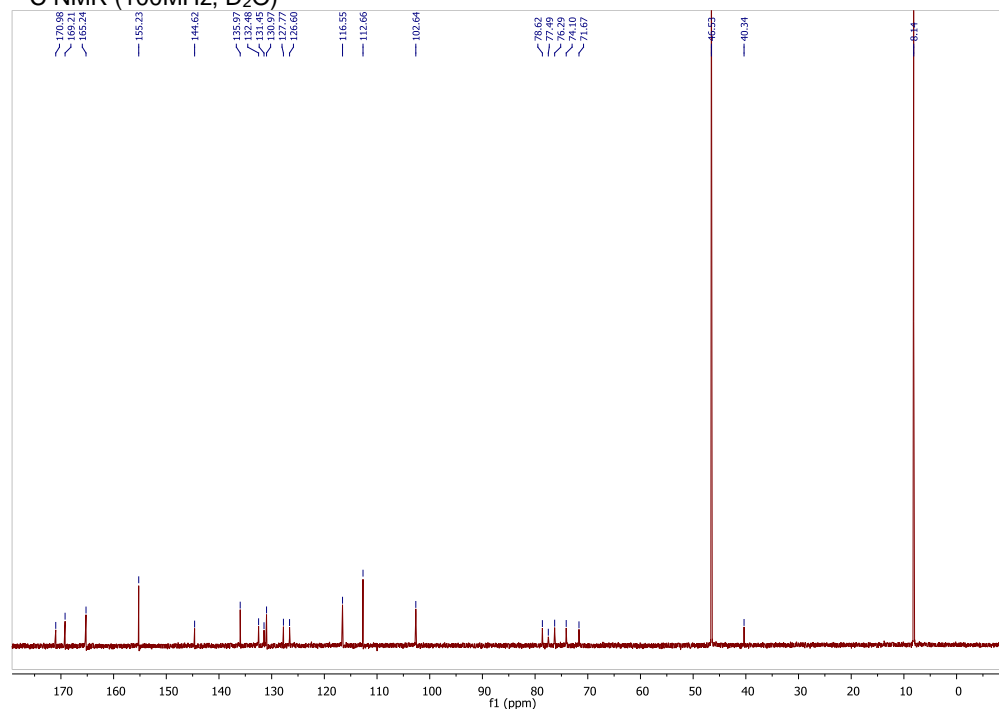
HPLC (5% to 70% CH_3CN in 0.1 M aqueous triethylammonium acetate, detection at 254 nm).



2-O-(2-(5-Fluoresceinylcarboxy)-aminoethyl)-*myo*-inositol 1,3,4,5,6-pentakisphosphate (5)



¹³C NMR (100MHz, D₂O)



³¹P NMR (109 MHz, CD₃OD)

