

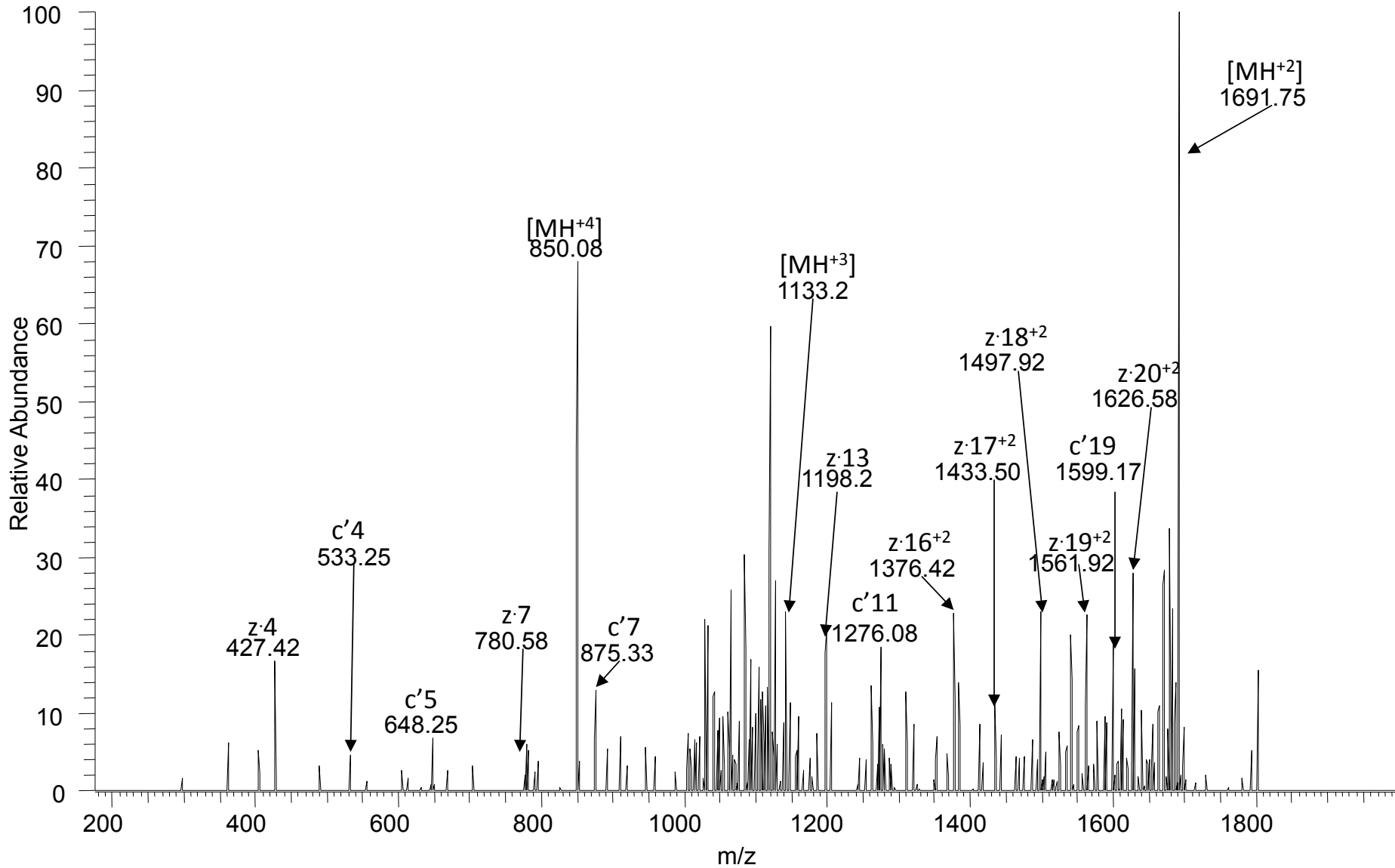
Supplementary table 10A: Glycopeptides identified using ETD in *Acinetobacter baumannii* (strain ATCC19606)

Uniprot accession number	Protein name	parent mass	charge	Precursor MH+	glycan mass	glycan composition	peptide mass	peptide	Site localisation	Page
DOC6Q2_ACIBA	Putative uncharacterized protein [HMPREF0010_00432] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]"	850.14	4	3397.52	1030.36	HexNAc-Hex ₂ -HexNAc-300	2367.160	³⁴ EEEQDKVETAVSEPQPQKPAK ⁵³	S ⁴⁵⁺	3
DOC6Q2_ACIBA	Putative uncharacterized protein [HMPREF0010_00432] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]"	866.41	3	2597.19	988.34	HexNAc-Hex ₂ -HexNAc-258	1608.859	⁴⁰ VETAVSEPQPQKPAK ⁵¹	S ⁴⁵⁺	4
DOC7T4_ACIBA	Putative uncharacterized protein [HMPREF0010_00814] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	964.42	3	2891.25	1030.36	HexNAc-Hex ₂ -HexNAc-300	1860.893	²⁸ KEEAAQAGQDAASTAVADK ⁴⁶	S ⁴⁰⁺	5
DOC8B9_ACIBA	Uncharacterized protein [F911_02370] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	1007.05	3	3019.13	1030.36	HexNAc-Hex ₂ -HexNAc-300	1988.766	²⁸ NDGM(+16)HEASDPATSHDM(+16)NK ⁴⁵	S ³⁵⁺	6
DOC8B9_ACIBA	Uncharacterized protein [F911_02370] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	755.78	4	3020.11	1030.36	HexNAc-Hex ₂ -HexNAc-300	1989.753	²⁸ N(+1)DGM(+16)HEASDPATSHDM(+16)NK ⁴⁵	S ³⁵⁺	7
DOC8B9_ACIBA	Uncharacterized protein [F911_02370] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	755.04	4	3017.15	1044.38	HexNAc-Hex ₂ -HexNAc-314	1972.765	²⁸ NDGM(+16)HEASDPATSHDMNK ⁴⁵	S ³⁵⁺	8
DOCAB8_ACIBA	Uncharacterized protein [F911_03368] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	657.32	3	1969.94	1030.36	HexNAc-Hex ₂ -HexNAc-300	939.562	³⁰² AKPASAPAVK ³¹¹	S ³⁰⁶⁺	9

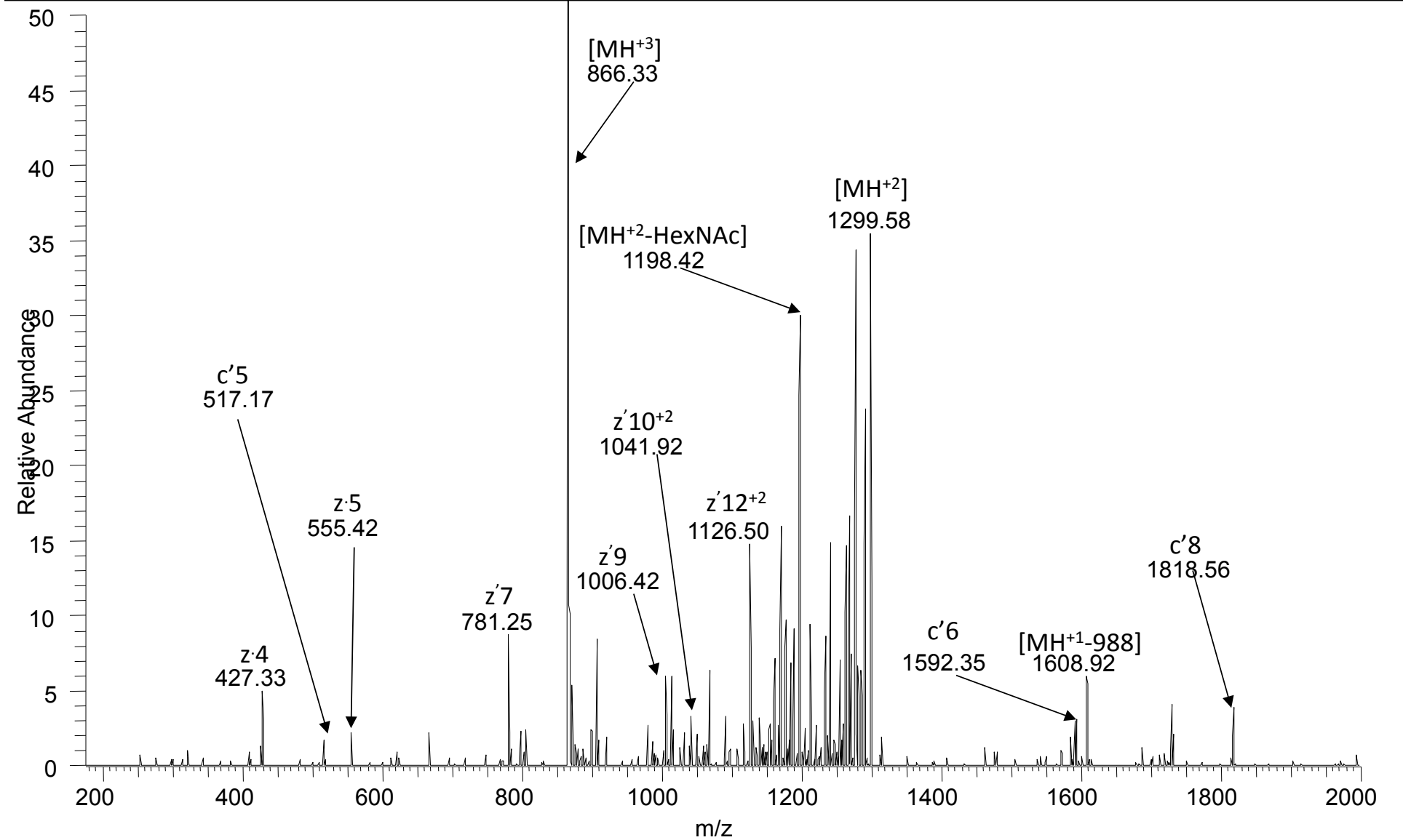
DOCAB8_ ACIBA	Uncharacterized protein [F911_03368] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	643.31	3	1927.91	988.35	HexNAc-Hex ₂ - HexNAc-258	939.562	³⁰² AKPASAPAVK ³¹¹	S ³⁰⁶⁺	10
DOCAB8_ ACIBA	Uncharacterized protein [F911_03368] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	1128.16	3	3382.47	1030.36	HexNAc-Hex ₂ - HexNAc-300	2352.114	¹¹⁶ KTEASAAAATEQQDSFDAQVQR ¹³⁶	S ¹²⁰	11
DOCDQ9_ ACIBA	Uncharacterized protein [F911_00789] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	846.38	3	2536.10	1030.36	HexNAc-Hex ₂ - HexNAc-300	1505.744	⁴⁰ STAKDEQPASSASVK ⁵⁴	S ^{51/50+}	12
DOCEI7_A CIBA	ToIA (Uncharacterized protein) [F911_02604] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	855.70	3	2565.10	1030.36	HexNAc-Hex ₂ - HexNAc-300	1534.734	⁵² QAASDIATATDNASAK ⁶⁷	S ⁵⁵⁺	13
DOCEI7_A CIBA	ToIA (Uncharacterized protein) [F911_02604] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	951.74	3	2853.20	988.35	HexNAc-Hex ₂ - HexNAc-300	1864.848	⁴⁸ SAGDQAASDIATATDNASAK ⁶⁷	S ⁵⁵⁺	14
DOCEI7_A CIBA	ToIA (Uncharacterized protein) [F911_02604] [Acinetobacter baumannii ATCC 19606 = CIP 70.34]	965.74	3	2895.21	1030.36	HexNAc-Hex ₂ - HexNAc-300	1864.852	⁴⁸ SAGDQAASDIATATDNASAK ⁶⁷	S ⁵⁵⁺	15

Supplementary table 10A: Identifications are labeled according to the corresponding Uniprot accession number and protein name. The parent m/z, charge state, deconvoluted mass, glycan mass, peptide mass, peptide sequence and glycan site attachment are provided for each identified glycopeptide. All identified glycopeptide ETD spectra are provided on the corresponding pages

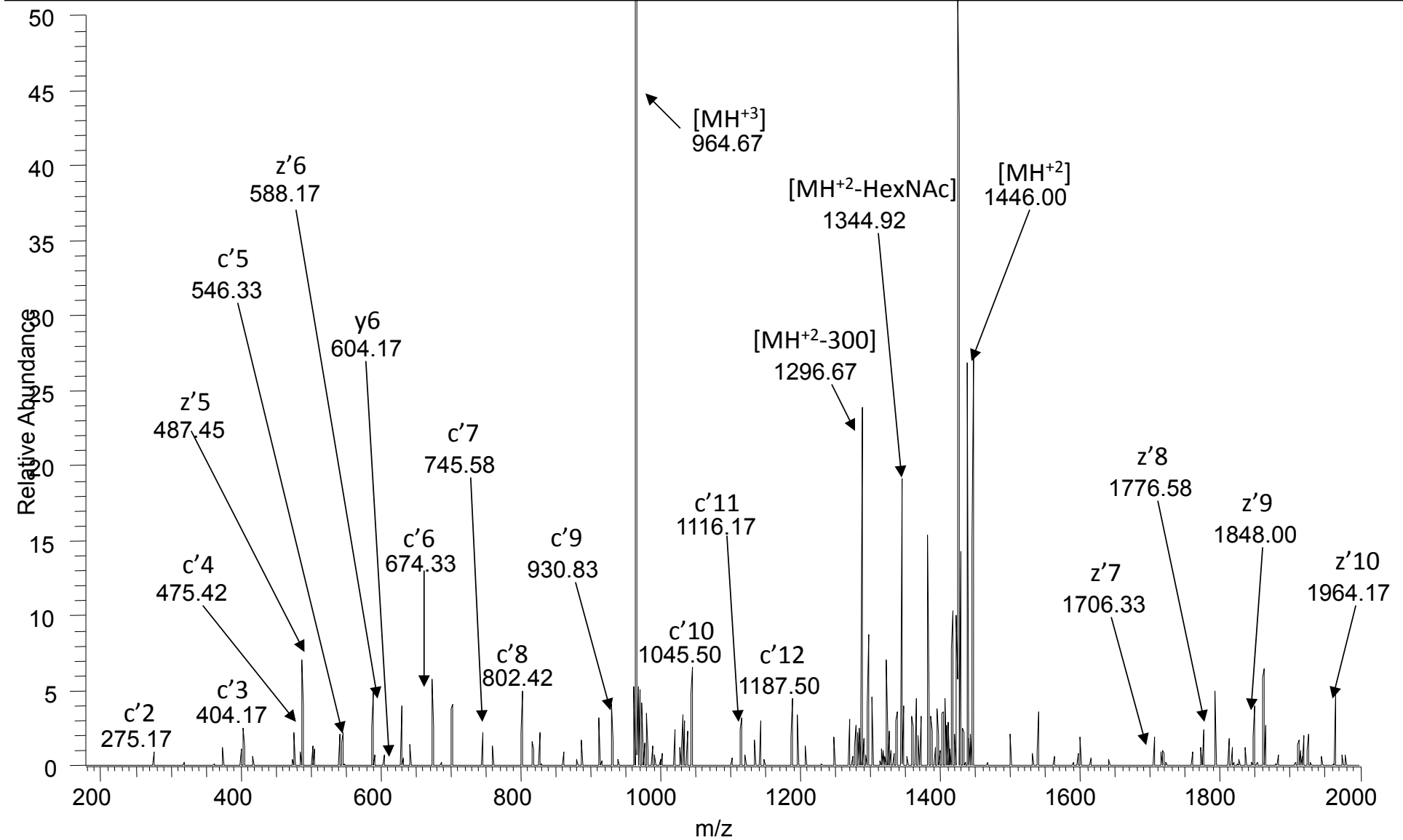
acinetobacter baumannii 19606		$^{34}\text{E} \text{E} \text{E} \text{Q} \text{D} \text{K} \text{V} \text{E} \text{T} \text{A} \text{V} \text{S} \text{E} \text{P} \text{Q} \text{P} \text{Q} \text{K} \text{P} \text{A} \text{K}^{53}$ c4 c5 c7 c11 z20 z19 z18 z17 z16 z13 z7 z4
Precursor charge:	+4	
Precursor m/z:	850.13525	
Peptide mass (MH+):	2367.15954	



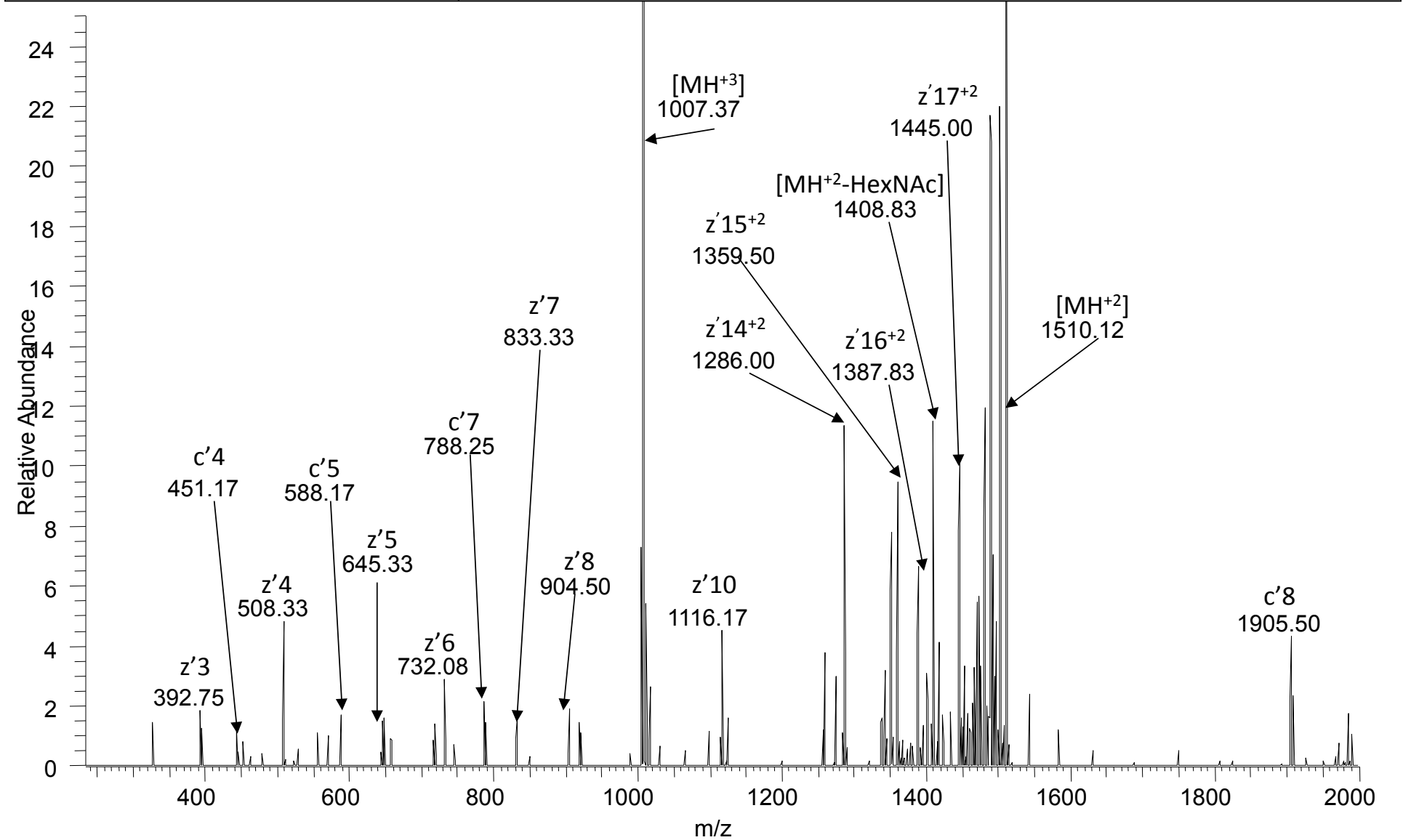
acinetobacter baumannii 19606		^{40}V E T $\left\{ \begin{array}{l} \text{A} \text{ V} \\ \text{z}12 \end{array} \right\} \left\{ \begin{array}{l} \text{S} \text{ E} \\ \text{z}10 \end{array} \right\} \left\{ \begin{array}{l} \text{P} \text{ Q} \\ \text{z}9 \end{array} \right\} \left\{ \begin{array}{l} \text{P} \text{ Q} \\ \text{z}7 \end{array} \right\} \left\{ \begin{array}{l} \text{K} \text{ P} \\ \text{z}5 \end{array} \right\} \left\{ \begin{array}{l} \text{A} \text{ K} \\ \text{z}4 \end{array} \right\}^{51}$
Precursor charge:	+3	
Precursor m/z:	866.40688	
Peptide mass (MH ⁺):	2597.1948	



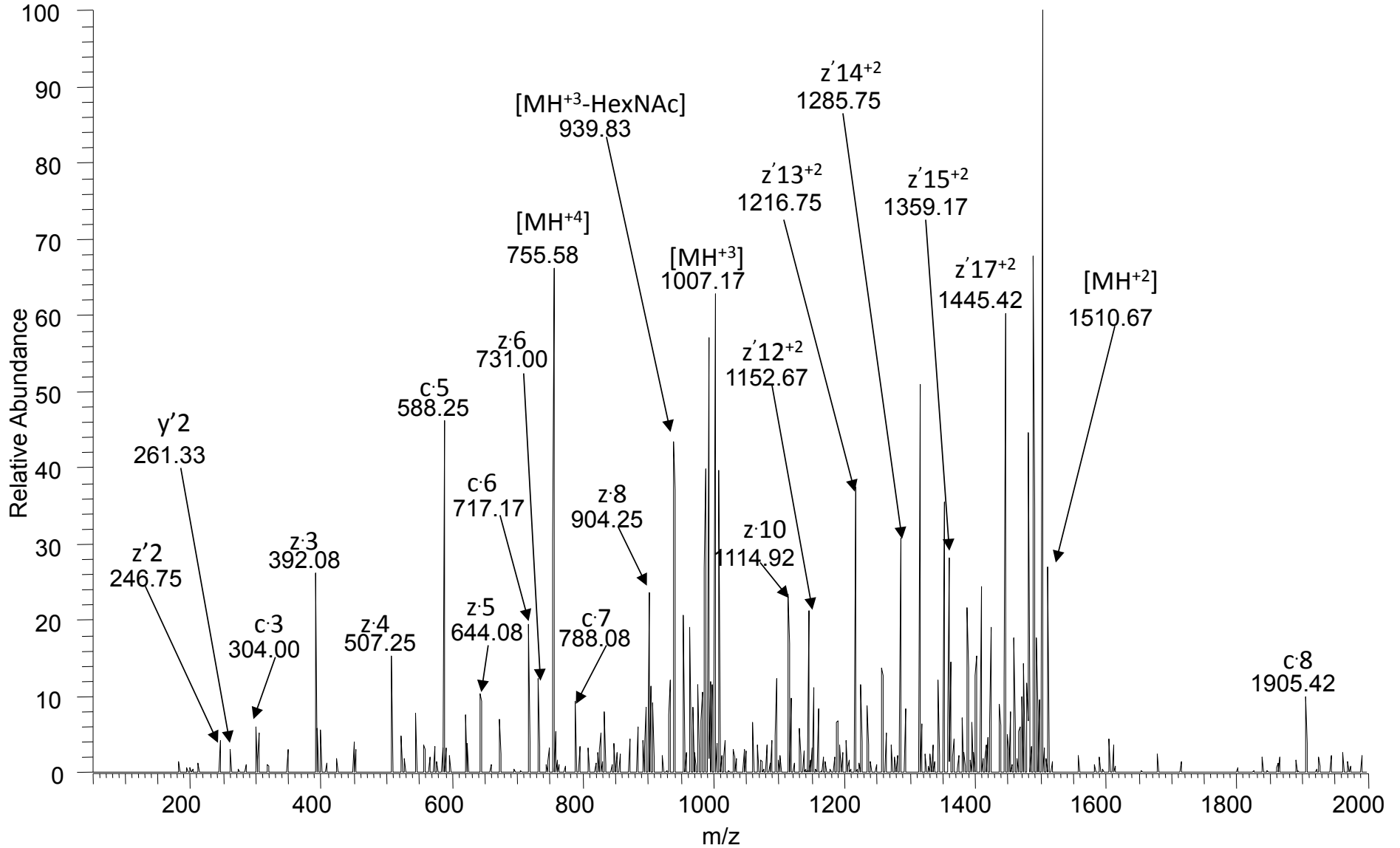
acinetobacter baumannii 19606		c_2 c_3 c_4 c_5 c_6 c_8 c_9 c_{10} c_{11} c_{12} ^{28}K E E A A Q A G Q D A A S T A V A D K 46 z_{10} z_9 z_8 z_7 z_6 z_5											
Precursor charge:	+3												
Precursor m/z:	964.42273												
Peptide mass (MH ⁺):	2891.2523												



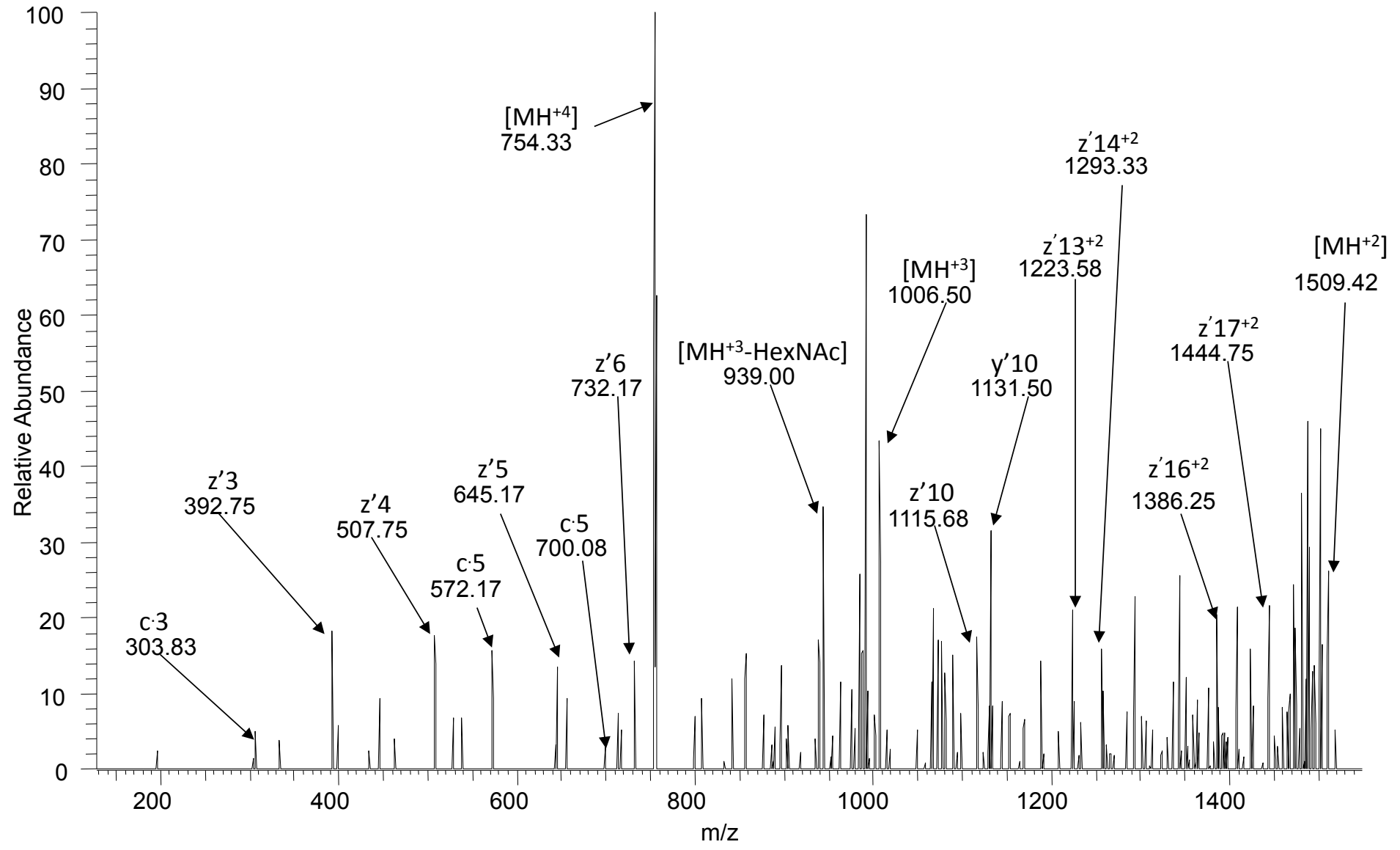
acinetobacter baumannii 19606		^{28}N D G M(+16) H E A S D P A T S H D M(+16) N K 45	
Precursor charge:	+3		
Precursor m/z:	1007.04865		
Peptide mass (MH ⁺):	3019.13007		



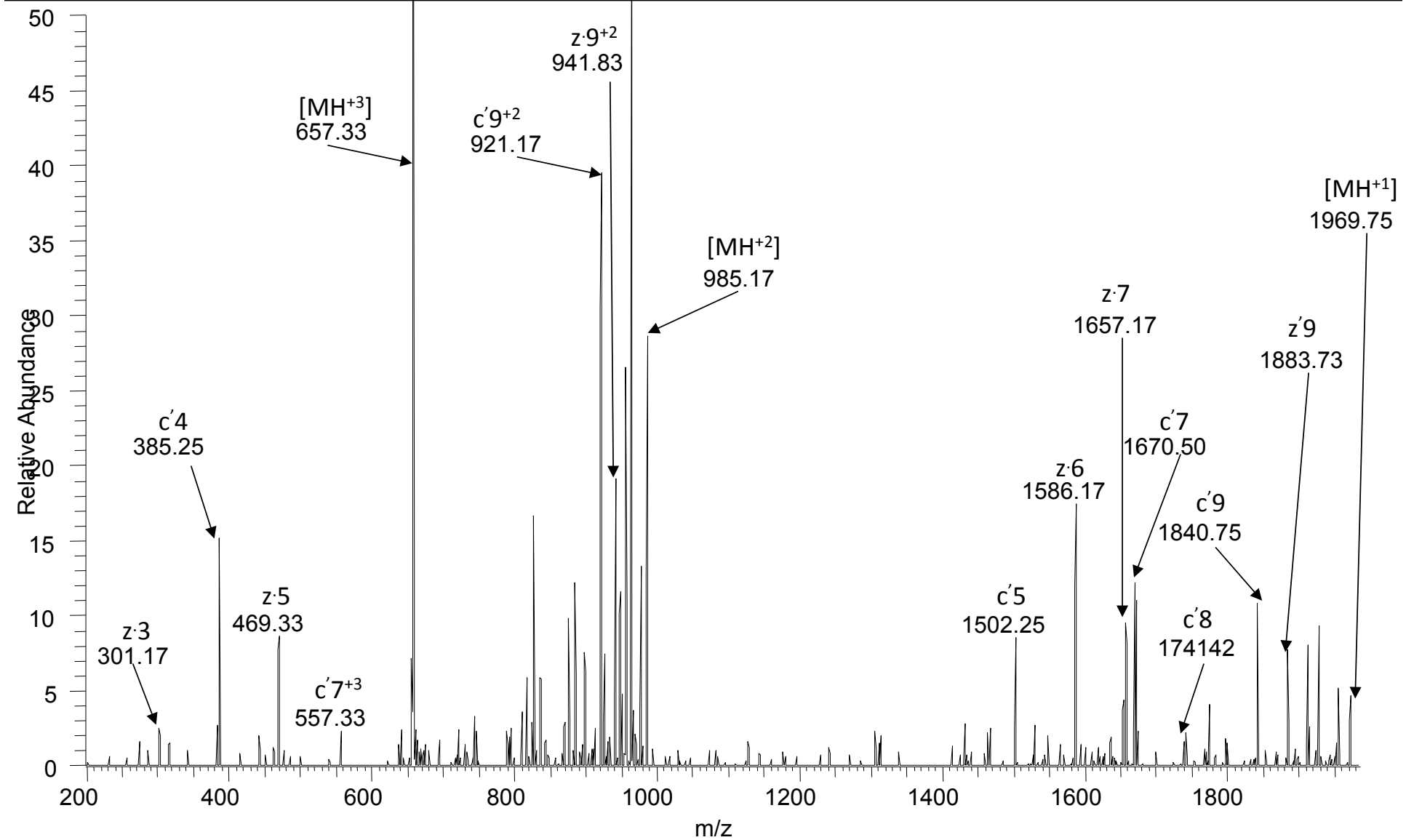
acinetobacter baumannii 19606		$^{28}\text{N}(+1)$	
Precursor charge:	+4		
Precursor m/z:	755.78442		
Peptide mass (MH+):	3020.11386		



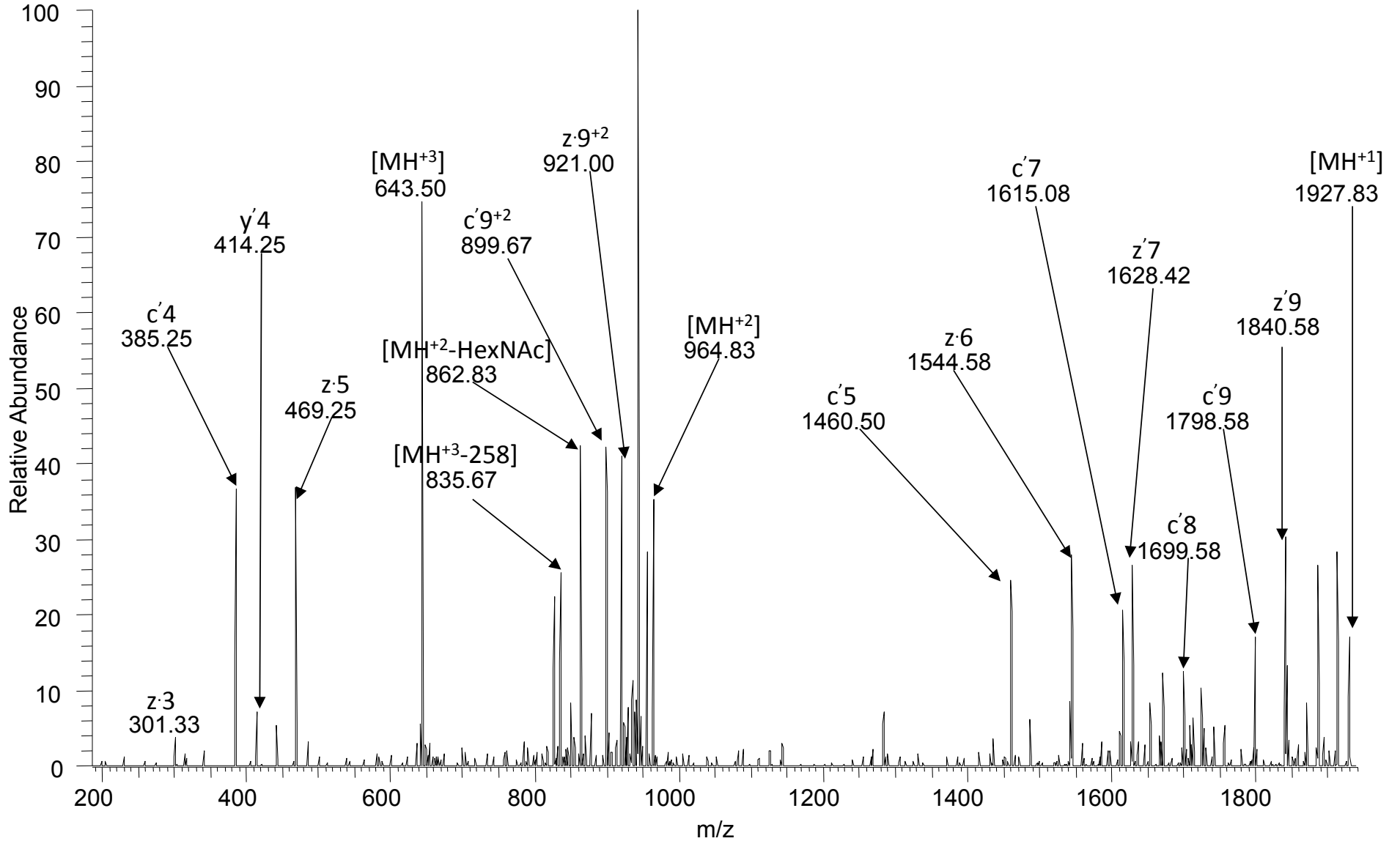
acinetobacter baumannii 19606		^{28}N D G M(+16) H E A S D P A T S H D M(+16) N K ⁴⁵	
Precursor charge:	+4	c3	c5 c6
Precursor m/z:	755.04230	z17	z14 z13
Peptide mass (MH+):	3017.14538		z10/y10 z6 z5 z4 z3



acinetobacter baumannii 19606														
Precursor charge:	+3	302		A	K	P	A	S	A	P	A	V	K	311
Precursor m/z:	657.3183			z9		z7		z6		z5		z3		
Peptide mass (MH+):	1969.94													



acinetobacter baumannii 19606			
Precursor charge:	+3	302 A $\left\{ \begin{array}{l} c4 \\ z9 \end{array} \right\}$ K P $\left\{ \begin{array}{l} c4 \\ z7 \end{array} \right\}$ A $\left\{ \begin{array}{l} c5 \\ z6 \end{array} \right\}$ S $\left\{ \begin{array}{l} c7 \\ z5 \end{array} \right\}$ A P $\left\{ \begin{array}{l} c8 \\ z3 \end{array} \right\}$ A $\left\{ \begin{array}{l} c9 \end{array} \right\}$ V $\left\{ \begin{array}{l} c8 \\ c9 \end{array} \right\}$ K 311	
Precursor m/z:	643.30884		
Peptide mass (MH ⁺):	1927.91064		



acinetobacter baumannii 19606	
Precursor charge: +3	
Precursor m/z: 1128.16248	
Peptide mass (MH ⁺): 3382.47156	

¹¹⁶K T E A S

^{c3} | ^{c4} |

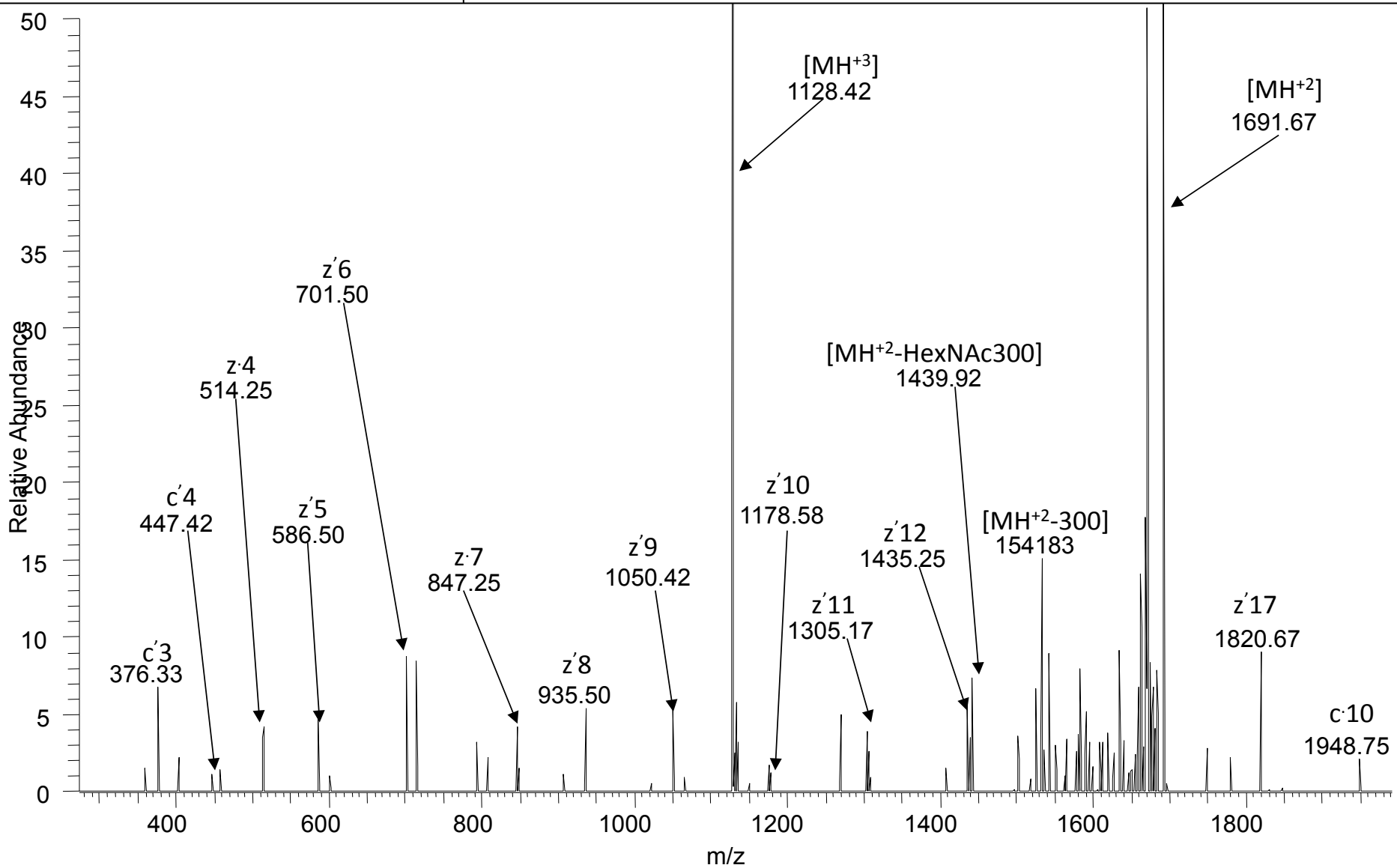
^{z17} | A A A A T

^{c10} |

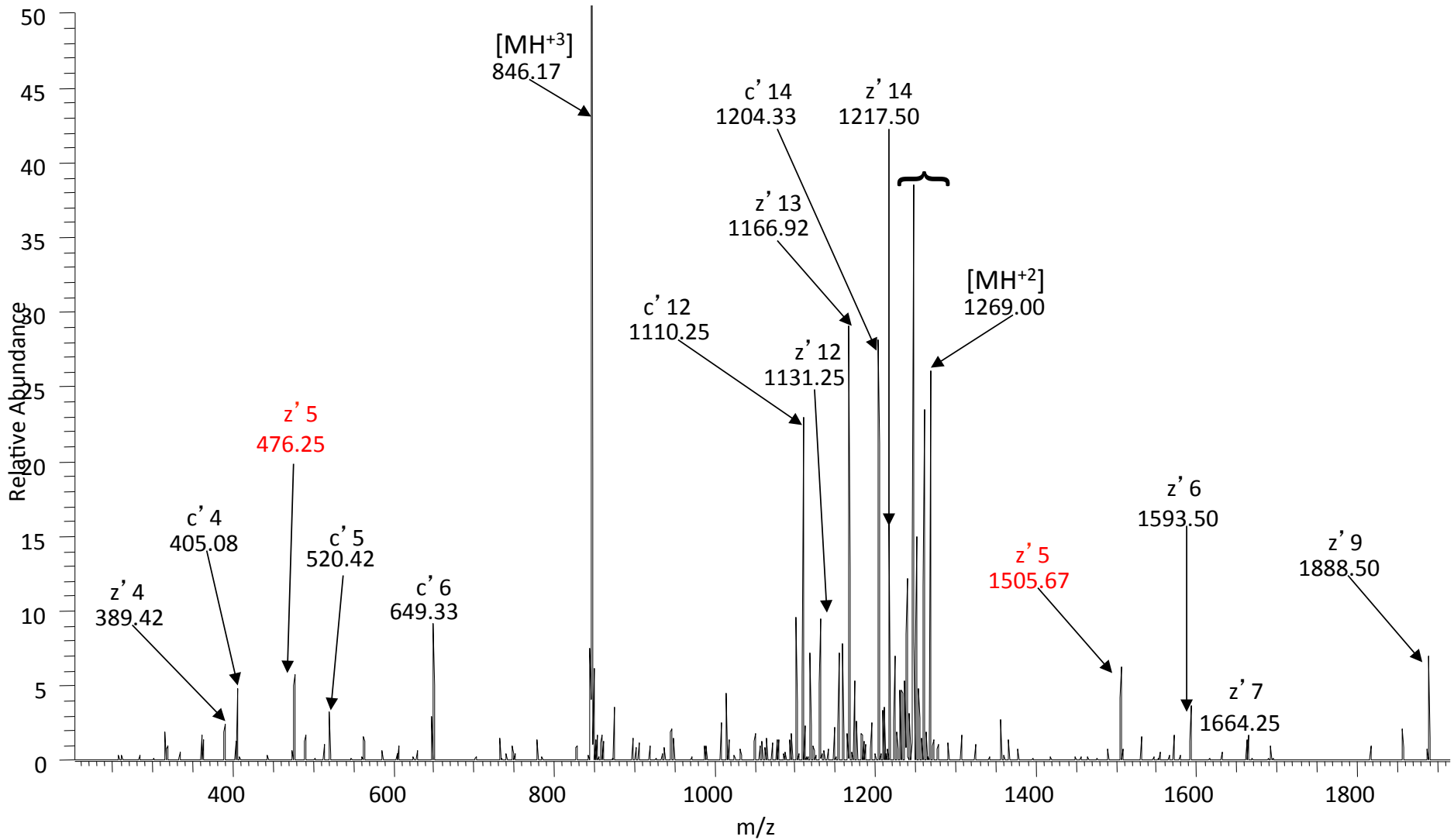
^{z12} | ^{z11} | ^{z10} | ^{z9} | ^{z8} | ^{z7} |

^{z6} | ^{z5} | ^{z4}

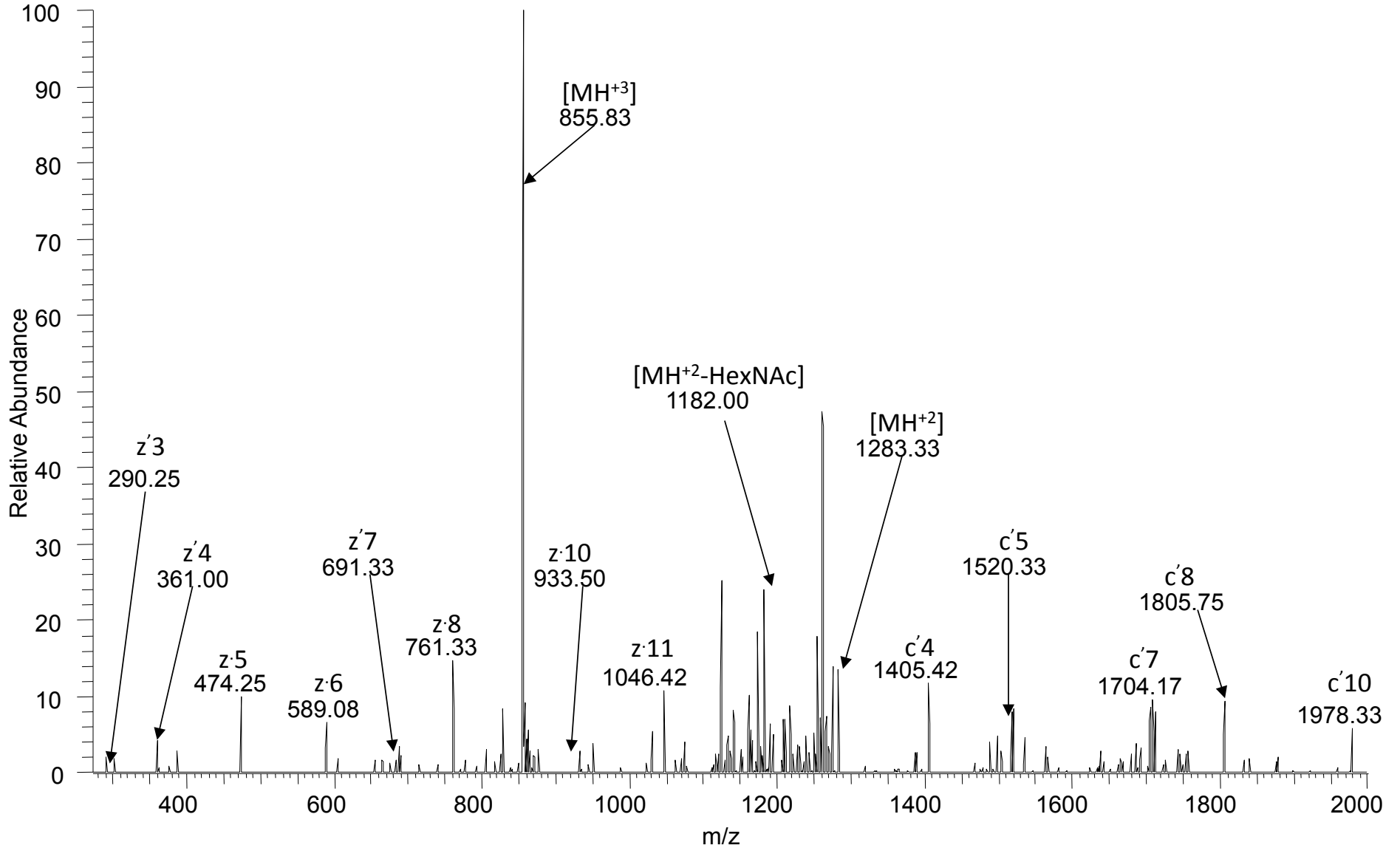
E Q Q D S F D A Q V Q R¹³⁶



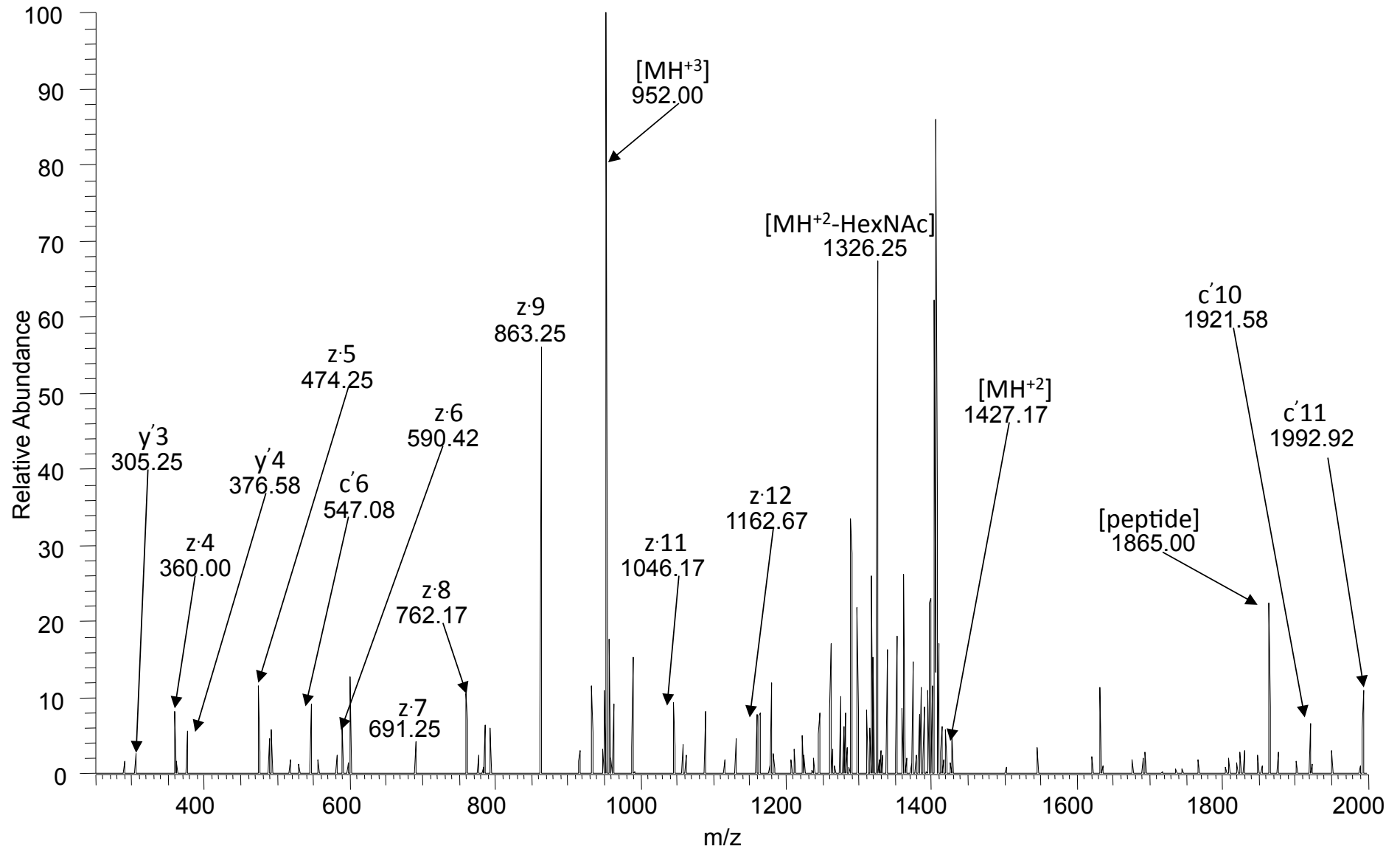
acinetobacter baumannii 19606	
Precursor charge: +3	50 S T A K D E Q P A S S A S V K 54
Precursor m/z: 846.37618	z14 z13 z12 z9 z7 z6 z5 z4
Peptide mass (MH+): 2536.1046	c4 c5 c6 c12 c13 c14



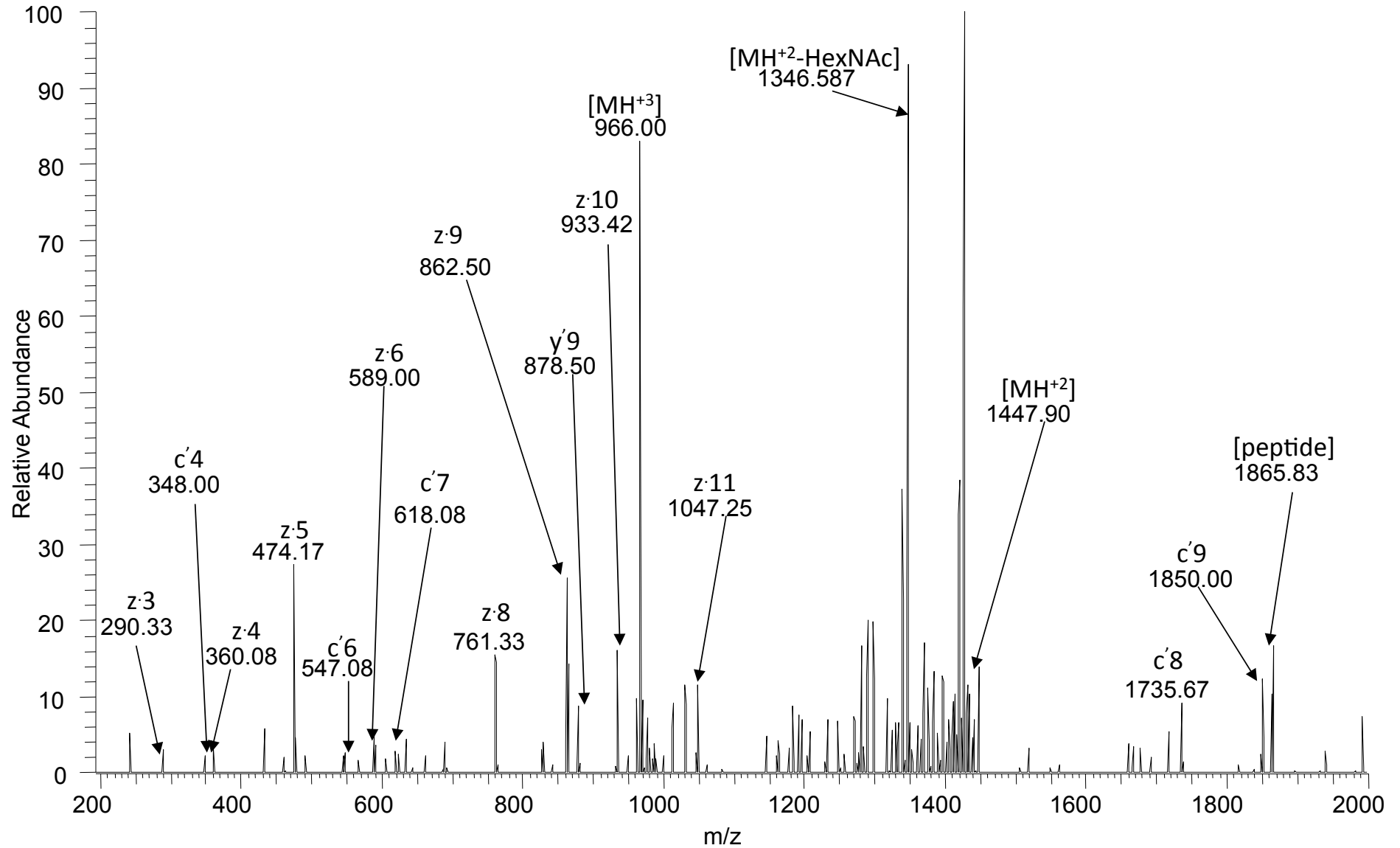
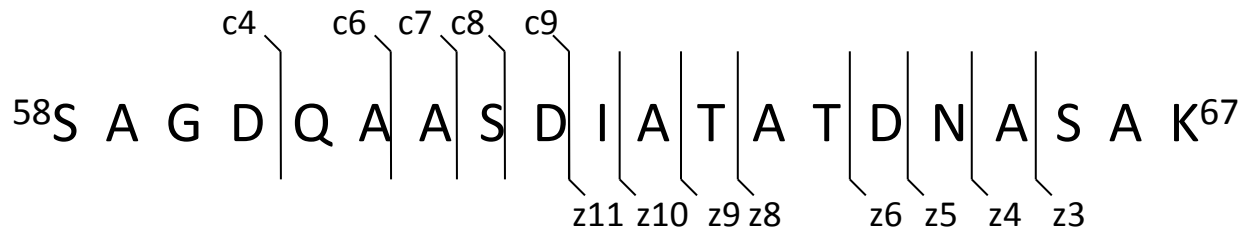
acinetobacter baumannii 19606		$c4$ $c5$ $c7$ $c8$ $c10$ ^{52}Q A A S D I A T A T D N A S A K^{67} $z11$ $z10$ $z8$ $z7$ $z6$ $z5$ $z4$	
Precursor charge:	+3		
Precursor m/z:	855.70392		
Peptide mass (MH+):	2565.09588		



acinetobacter baumannii 19606		^{58}S A G D Q A $\left\{ \begin{array}{l} \text{c6} \\ \text{z12} \end{array} \right.$ A S $\left\{ \begin{array}{l} \text{c10} \\ \text{z11} \end{array} \right.$ A T $\left\{ \begin{array}{l} \text{c11} \\ \text{z9} \end{array} \right.$ A T $\left\{ \begin{array}{l} \text{z8} \\ \text{z7} \end{array} \right.$ D $\left\{ \begin{array}{l} \text{z6} \\ \text{z5} \end{array} \right.$ N $\left\{ \begin{array}{l} \text{z4} \\ \text{y3} \end{array} \right.$ S A K^{67}	
Precursor charge:	+3		
Precursor m/z:	951.73907		
Peptide mass (MH ⁺):	2853.19762		

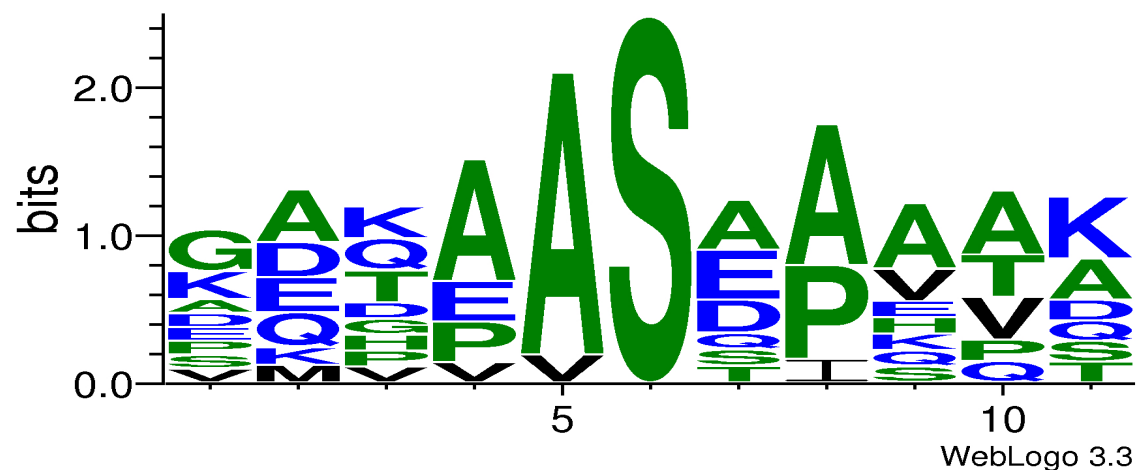


acinetobacter baumannii 19606	
Precursor charge: +3	
Precursor m/z: 965.74292	
Peptide mass (MH ⁺): 2895.21398	



Supplementary Table 10B: Glycosylation site localisation in *Acinetobacter baumannii* (strain ATCC19606)

Uniprot accession number	Site localisation	five amino acid each side of the site
D0C6C0_ACIBA	S ₁₈₆	EQKAASEPEQK
D0C6Q2_ACIBA	S ₄₅ ⁺	VETAVSEPQPQ
D0C7T4_ACIBA	S ₄₀ ⁺	GQDAASTAVAD
D0C8B9_ACIBA	S ₃₅ ⁺	GMHEASDPATS
D0CAB8_ACIBA	S ₃₀₆ ⁺	KAKPASAPAVK
D0CA69_ACIBA	S ₃₄	AAGAASEAVAA
D0CA69_ACIBA	S ₇₈	SDVAASAAH--
D0CAB8_ACIBA	S ₁₂₀ ⁺	KKTEASAAAAT
D0CDA9_ACIBA	S ₁₂₃	PAPVASQAK--
D0CDQ9_ACIBA	S ₄₉ ⁺	DEQPASSASVK
D0CEI7_ACIBA	S ₅₅ ⁺	GDQAASDIATA



Supplementary table 10B: Glycosylation site localization in *A. baumannii* ATCC19606. Identifications are labeled according to the corresponding Uniprot accession number within strain ATCC19606. The site and residue of localization are denoted with + indicating sites localized by ETD. To enable assessment of the local environment of glycosylation five amino acids either side of the site of modification are provided.