Visible Light-Promoted Metal-Free sp³-C–H Fluorination

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Figure S1. The UV-vis spectra of acetophenone in acetonitrile at different concentrations.



Figure S2. The UV-vis spectra of 5 mol % acetophenone catalyst alone in acetonitrile (4 mM) (red), and of the reaction mixture (5 mol % acetophenone, 1.0 equiv ethylbenzene, 2.0 equiv Selectfluor) in acetonitrile (blue).



Figure S2. The UV-vis spectra of 5 mol % benzophenone catalyst alone in acetonitrile (4 mM) (red), and of the reaction mixture (5 mol % benzophenone, 1.0 equiv ethylbenzene, 2.0 equiv Selectfluor) in acetonitrile (blue).



Figure S3. The UV-vis spectra of 5 mol % 9-fluorenone catalyst alone in acetonitrile (4 mM) (red), and of the reaction mixture (5 mol % 9-fluorenone, 1.0 equiv ethylbenzene, 2.0 equiv Selectfluor) in acetonitrile (blue).



Figure S5. The UV-vis spectrum of cyclopentenone (27) in 2-propanol (50 mM).



Figure S6. The UV-vis spectra of (*Z*)-enal **30** in methylene chloride (50 mM) (red), and (*E*)-enal **32** in methylene chloride (50 mM) (blue).



Figure S7. Acetophenone-catalyzed fluorination of cyclooctane under CFL-irradiation is a light-dependent reaction.



Figure S8. C–H abstraction/conjugate addition of cyclopentenone (**27**) under CFL-irradiation is a light-dependent reaction.



Figure S9. Time-course NMR spectra of the Norrish type II cleavage and the Norrish–Yang cyclization of valerophenone (**22**) under CFL-irradiation.



Figure S10. The emission spectra of the 19 W CFL used.



Figure S10. The emission spectra of the 9 W violet LED bulb used.



Figure S12. The emission spectra of the RPR lamps used in Table 1 (provided by Southern New England Ultraviolet Company).



Figure S13. The transmission spectra of the Asahi longpass filters used in Table 1 (provided by Asahi Spectra).

Materials and Methods

General Information. All reactions were performed in glassware under argon. Organic solutions were concentrated by rotary evaporator at ca. 30 mmHg unless otherwise noted. Flash column chromatography was performed as described by Still^{S1}, employing EMD silica gel 60 (230–400 mesh ASTM). TLC analyses were performed on EMD 250 μ m Silica Gel 60 F₂₅₄ plates and visualized by quenching of UV fluorescence (λ_{max} 254 nm), or by staining ceric ammonium molybdate. ¹H and ¹³C NMR spectra were recorded on a Varian Inova-600, Inova-500, or Inova-400. Chemical shifts for ¹H and ¹³C NMR spectra are reported in ppm (δ) relative to the ¹H and ¹³C signals in the solvent (CDCl₃: 7.26, 77.00 ppm; C₆D₆: δ 7.16, 128.06 ppm; CD₃CN: δ 1.94, 1.32 ppm) and the multiplicities are presented as follows: s = singlet, d = doublet, t = triplet, q = quartet, hept = heptet, m = multiplet. Crude ¹H and ¹⁹F NMR spectra were recorded on an Inova-400 in CD₃CN using fluorobenzene (¹⁹F NMR δ –114.930) as an external standard. Mass spectra were acquired on an Agilent 6120 Single Quadrupole LC/MS or Agilent 7820A GC/5975 MSD. UV-vis spectra was collected on Shimadzu UV-Visible Spectrophotometer (UV-1601).

Materials. Acetophenone (analytical standard, $\geq 99.5\%$ GC), and acetonitrile (anhydrous, 99.8%) was purchased from Sigma-Aldrich. Selectfluor (98+%) was purchased from Alfa Aesar. Fluorobenzene ($\geq 99.5\%$) was purchased from Fluka. Acetonitrile-*d*3 (D, 99.8%) was purchased from Cambridge Isotope Laboratories, Inc. A 19 W EcoSmart Daylight CFL Bulb was used for the photoreaction. RPR-3000Å, RPR-3500Å, RPR-4190Å, Rayonet RMR-200 chamber reactor, and RMA-500 Merry-Go-Round were purchased from Southern New England Ultraviolet Company. 9-W (3 × 3W) 380–385nm violet LED flashlight was purchased from LED wholesalers. The 385 nm, and 400 nm longpass filters were purchased from Asahi Spectra USA Inc.

General procedure for the acetophenone-catalyzed $C(sp^3)$ –H fluorination. To a 4 mL clear vial charged with Selectfluor (70.9 mg, 0.2 mmol, 1.0 equiv) were added anhydrous acetonitrile (2.0 mL), acetophenone (1.2 mg, 0.01 mmol), and the reaction substrate (0.3 mmol, 1.5 equiv). The reaction mixture was degassed by *Freeze-Pump-Thaw* cycles for three times and irradiated with a 19 W CFL 2–5 cm away from the reaction at room temperature (27±2 °C). The crude yield was determined by NMR using fluorobenzene as an external standard. The reaction mixture was then poured into diethyl ether (20 mL), filtrated, concentrated and purified by silica gel flash column chromatography using diethyl ether/pentane as the eluent.

^{S1} W. C. Still, M. Kahn and A. Mitra, J. Org. Chem. **1978**, 43, 2923.

	+ C ₆ D ₁₂ (5- d ₁₂) (20 equiv)	hv (CFL) 5 mol % acetophenone 1.0 equiv Selectfluor		+ C ₆ D ₁₁ F (6- d ₁₁) % : 42%)
(5) (20 equiv)		CH ₃ CN, 27 °C, 3 h <i>k</i> _H / <i>k</i> _D = 1.4	6) (6) (58% : 4	

The intermolecular kinetic isotope effect study. To a 4 mL clear vial charged with Selectfluor (35.4 mg, 0.1 mmol, 1.0 equiv) were added anhydrous acetonitrile (1.0 mL), acetopheneone (0.6 mg, 0.005 mmol) cyclohexane (168.4 mg, 20.0 mmol, 20.0 equiv), and cyclohexane- d_{12} (192.5 mg, 1.0 mmol, 20.0 equiv). The reaction mixture was degassed by *Freeze-Pump-Thaw* cycles for three times and irradiated with a 19 W CFL 2–5 cm away from the reaction at room temperature for 3 h. The ratio of **6**:**6**- d_{11} was determined by ¹⁹F NMR on a Varian Inova-400 NMR instrument to be 58:42.



Norrish type II cleavage and Norrish–Yang cyclization reaction of valerophenone (22) under CFL irradiation. To a 4 mL clear vial charged with valerophenone (22) (8.1 mg, 0.05 mmol) was added acetonitrile-*d*3 (0.5 mL, redistilled from calcium hydride and degassed by *freeze-pump-thaw* cycles for three times) in glovebox. The reaction solution was transfer to a NMR tube (Wilmad-Lab Glass, 527PP, 5mm OD). The NMR tube was taken out from the glovebox, irradiated with a 19 W CFL at room temperature, and monitored by a Varian Inova-400 NMR instrument. After 40 h, the solution was concentrated and the residue was purified by silica gel chromatography (3% diethyl ether/pentane) to recover valerophenone (22) (0.8 mg, 10% yield) and give acetophenone (23) (3.5 mg, 58% yield), *trans*-2-methyl-1-phenylcyclobutanol (25) (0.8 mg, 10% yield), and *cis*-2-methyl-1-phenylcyclobutanol (26) (0.6 mg, 7% yield).⁸² 23: ¹H NMR (400 MHz, CD₃CN) δ 7.96–7.98 (m, 2H), 7.59–7.63 (m, 1H), 7.48–7.53 (m, 2H), 2.57 (s, 3H); 25: ¹H NMR (400 MHz, CD₃CN) δ 7.43–7.45 (m, 2H), 7.32–7.36 (m, 2H), 7.20–7.25 (m, 1H), 3.14 (s, 1H), 2.60–2.69 (m, 1H), 2.35–2.42 (m, 1H), 2.08–2.13 (m, 1H), 1.96–2.00 (m, 1H), 1.67–1.76 (m, 1H), 1.08 (d, *J* = 7.0 Hz, 3H). 26: ¹H NMR (400 MHz, CD₃CN) δ 7.43–7.45 (s, 1H), 2.65 (ddd, *J* = 7.4, 7.9 Hz, 2H), 7.25 (d, *J* = 7.3 Hz, 1H), 3.46 (s, 1H), 2.65 (ddd, *J* = 11.7, 9.0, 3.1 Hz, 1H), 2.53–2.59 (m, 1H), 2.12–2.22 (m, 1H), 2.01–2.09 (m, 1H), 1.22–1.31 (m, 1H), 0.52 (d, *J* = 7.0 Hz, 3H).



Photolytic C–H abstraction/conjugate addition of 2-cyclopentenone 27 under CFL-irradiaion. To a 4 mL clear vial charged with 2-cyclopentenone **27** (8.2 mg, 0.1 mmol) was added anhydrous 2-propanol (2.0 mL), and the reaction mixture was degassed three times by *Freeze-Pump-Thaw* cycles and stirred at room temperature under CFL for 40 h. After remove the 2-propanol, the residue was purified by flash column chromatography (30% ethyl acetate/hexanes) to give **29** (12.2 mg, 86% yield) as a colorless oil:

⁵² F. D. Lewis and T. A. Hilliard, J. Am. Chem. Soc. 1972, 94, 3852.

 $R_f = 0.3$ (60% ethyl acetate/hexanes).^{S3 1}H NMR (400 MHz, CDCl₃) δ 2.37 (dd, J = 18.7, 8.6 Hz, 1H), 2.13–2.31 (m, 4H), 2.03–2.10 (m, 1H), 1.71–1.82 (m, 1H), 1.47 (brs, 1H), 1.25 (d, J = 11.7 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 219.3, 70.8, 47.7, 40.0, 39.0, 28.6, 27.8, 23.8; MS(ESI) calcd for C₈H₁₄O₂Na (M+Na)⁺ 165.1, found 165.1.



Photolytic [2+2] cycloaddition and E/Z isomerization of enal 30 under CFL-irradiation. To a 4 mL clear vial charged with (Z)-4-(cinnamyloxy)but-2-enal **30** (20.4 mg, 0.1 mmol) was added anhydrous DCM (2.0 mL), and the reaction mixture was degassed three times by *Freeze-Pump-Thaw* cycles and stirred at room temperature under CFL for 24h. After remove the DCM, the residue was purified by flash column chromatography (10% ethyl acetate/hexanes) to give 31 (6.2 mg, 31% yield) and 32 (7.1 mg, 35% yield).^{S4} **30**: ^IH NMR (400 MHz, CDCl₃) δ 10.09 (d, J = 6.7 Hz, 1H), 7.38–7.41 (m, 2H), 7.31–7.35 (m, 2H), 7.24–7.28 (m, 1H), 6.66 (t, J = 5.7 Hz, 2H), 6.29 (dt, J = 16.0, 6.2 Hz, 1H), 6.64– 6.51 (m, 1H), 4.23 (dd, J = 6.1, 1.2 Hz, 4H), 6.62–6.67 (m, 2H), 6.62 (d, J = 5.5 Hz, 1H), 6.08 (ddt, J = 10.8, 6.7, 1.8 Hz, 1H), 4.55 (dd, J = 5.5, 2.0 Hz, 3H), 4.23 (dd, J = 6.1, 1.2 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 191.4, 147.6, 136.3, 133.3, 129.7, 128.6, 127.9, 126.5, 125.0, 71.6, 66.8; MS(ESI) calcd for $C_{13}H_{15}O_2$ (M+H)⁺ 203.1, found 203.1. **31**: ¹H NMR (600 MHz, CDCl₃) δ 9.26 (s, 1H), 7.31–7.33 (m, 2H), 7.21–7.24 (m, 3H), 4.03 (d, J = 9.8 Hz, 1H), 3.96 (d, J = 9.5 Hz, 1H), 3.69 (dd, J = 10.7, 5.0 Hz, 1H), 3.63 (dd, J = 9.4, 5.5 Hz, 1H), 3.58 (dd, J = 9.6, 5.3 Hz, 1H), 3.55 (dd, J = 5.5, 5.6 Hz, 1H), 3.27 (dd, J = 10.7, 4.9 Hz, 1H), 3.13 (dd, J = 5.5, 5.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 202.0, 139.2, 128.8, 127.7, 127.0, 73.7, 73.2, 52.6, 45.4, 43.7, 36.1; MS(ESI) calcd for $C_{13}H_{15}O_2$ (M+H)⁺ 203.1, found 203.1. **32**: ¹H NMR (400 MHz, CDCl₃) δ 9.58 (d, J = 7.9 Hz, 1H), 7.37–7.40 (m, 2H), 7.30–7.33 (m, 2H), 7.22–7.26 (m, 1H), 6.84 (dt, J = 15.8, 4.1 Hz, 1H), 6.62 (d, J = 16.0 Hz, 1H), 6.38 (ddt, J = 15.7, 7.9, 1.9 Hz, 1H), 6.27 (dt, J = 15.9, 6.0 Hz, 1H), 4.29 (dd, J = 4.1, 1.9 Hz, 2H), 4.21 (dd, J = 6.1, 1.4 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 193.2, 153.0, 136.3, 133.0, 131.7, 128.6, 127.8, 126.5, 125.0, 71.5, 68.4; MS(ESI) calcd for $C_{13}H_{15}O_2$ (M+H)⁺ 203.1, found 203.1.



When (*E*)-4-(cinnamyloxy)but-2-enal **32** was used as the starting material under the same conditions, The reaction was slow and most of the **32** (65% yield) was recovered after 1d.

^{S3} J. F. Gil, D. J. Ramón and M. Yus, *Tetrahedron*, **1994**, *50*, 3437.

^{S4} C. Ko, J. B. Feltenberger, S. K. Ghosh and R. P. Hsung, Org. Lett. 2008, 10, 1971.

Characterization Data

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1-Fluorocyclohexane (6).^{S 5} Prepared from cyclohexane (25.3 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (15 h) to give 6 (76% NMR yield). ¹⁹F NMR (376 MHz, CD₃CN) δ –172.38 (brs, 1F).

1-Fluorocycloheptane (7).^{S 6} Prepared from cycloheptane (29.5 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (6 h) to give 7 (85% NMR yield). ¹⁹F NMR (376 MHz, CD₃CN) δ –163.78 (m, 1F).

1-Fluorocyclooctane (8).^{S6} Prepared from cyclooctane (33.7 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (3 h) to give 8 (82 % NMR yield). ¹⁹F NMR (376 MHz, C_6D_6) δ -159.23 (brs, 1F).

1-Fluorocyclodecane (9).^{S7} Prepared from cyclodecane (42.1 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (15 h), purified by flash column chromatography (pentane), and concentrated by rotary evaporator at ca. 100 mm Hg to give 9 (23.1 mg, 73% yield) as a colorless oil: $R_f = 0.5$ (pentane). ¹H NMR (400 MHz, C₆D₆) δ 4.65 (dtt, J = 46.6, 7.8, 4.1 Hz, 1H), 1.63–1.89 (m, 4H), 1.42–1.51 (m, 2H), 1.27–1.38 (m, 12H); ¹³C NMR (100

9 7.8, 4.1 Hz, 1H), 1.63–1.89 (m, 4H), 1.42–1.51 (m, 2H), 1.27–1.38 (m, 12H); ¹³C NMR (100 MHz, C₆D₆) δ 93.5 (d, J = 165.8 Hz), 31.1 (d, J = 21.9 Hz), 25.4, 25.1, 24.0, 21.8 (d, J = 7.7 Hz); ¹⁹F NMR (376 MHz, C₆D₆) δ –165.63 (dtt, J = 46.5, 26.5, 13.7 Hz, 1F); MS(EI) calcd for C₁₀H₁₉F (M)⁺ 158.1, found 158.1.

F **1-Fluorocyclododecane** (10).^{S 8} Prepared from cyclododecane (50.5 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (15 h), purified by flash column chromatography (pentane), and concentrated by rotary evaporator at ca. 100 mm Hg to give 10 (27.9 mg, 75% yield) as a colorless oil: $R_f = 0.5$ (pentane). ¹H NMR (400 MHz, C₆D₆) δ 4.59 (dddd, J = 47.3, 11.5, 7.2, 4.7 Hz, 1H), 1.55–1.73 (m, 4H), 1.31–1.38 (m, 2H), 1.19 (brs, 16H); ¹³C NMR (100 MHz, C₆D₆) δ 91.8 (d, J = 167.0 Hz), 30.3 (d, J = 21.2 Hz), 24.4, 24.2, 23.6, 23.5, 20.9 (d, J = 6.7 Hz); ¹⁹F NMR (376 MHz, C₆D₆) δ –176.13 (dddd, J = 47.3, 23.6, 15.4, 8.2 Hz, 1F); MS(EI) calcd for C₁₂H₂₃F (M)⁺ 186.2, found 186.1.

exo-2-Fluoronorbornane (11).^{S6} Prepared from norborane (28.9 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (24 h) to give *exo*-2-fluoronorbornane 11 (58% NMR yield) and *endo*-2-fluoronorbornane (4% NMR yield). *exo*-2-fluoronorbornane 11: ¹⁹F NMR (376 MHz, CD₃CN) δ –160.29 (m, 1F); *endo*-2-fluoronorbornane: ¹⁹F NMR (376 MHz, CD₃CN) δ –190.00 (m, 1F).

^{S5} R. D. Chambers, A. M. Kenwright, M. Parsons, G. Sandford and J. S. Moilliet, J. Chem. Soc., Perkin Trans. 1, 2002, 2190.

^{S6} H.-J. Schneider, W. Gschwendtner, D. Heiske, V. Hoppen and F. Thomas, *Tetrahedron*, **1977**, *33*, 1769.

⁸⁷ S. Bloom, C. R. Pitts, D. C. Miller, N. Haselton, M. G. Holl, E. Urheim and T. Lectka, *Angew. Chem. Int. Ed.* 2012, *51*, 10580.

^{S8} Y. Amaoka, M. Nagatomo and M. Inoue, Org. Lett. 2013, 15, 2160.



3-Fluoropropanoic acid (12). Prepared from propanoic acid (22.2 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (40 h) to give **12** (<5% NMR yield). ¹⁹F NMR (376 MHz, CD₃CN) δ –219.68 (tt, *J* = 47.6, 26.8 Hz, 1F).

F O **Benzyl 3-fluorobutanoate** (13S).^{S9} Prepared from butyric acid (26.4 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (15 h) to give 3-fluorobutyric acid (70% NMR yield). To the reaction mixture were added *N*,*N*-diisopropylethylamine (77.5 mg, 3.0 equiv) and benzyl bromide (68.4 mg, 2.0 equiv). The reaction mixture was stirred at room temperature for 12 h, then poured into diethyl ether (20 mL), filtrated, concentrated and purified by silica gel flash column chromatography (10% ethyl acetate/beyapes) to give **13S** (21.7

^{13S} (77.5 mg, 3.0 equiv) and benzyl bromide (68.4 mg, 2.0 equiv). The reaction mixture was stirred at room temperature for 12 h, then poured into diethyl ether (20 mL), filtrated, concentrated and purified by silica gel flash column chromatography (10% ethyl acetate/hexanes) to give **13S** (21.7 mg, 55% yield) as a colorless oil: $R_f = 0.3$ (20% ethyl acetate/hexanes). ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.39 (m, 5H), 5.02–5.22 (m, 1H), 5.16 (s, 2H), 2.78 (ddd, J = 15.8, 14.2, 7.9 Hz, 1H), 2.59 (ddd, J = 28.3, 15.8, 4.8 Hz, 1H), 1.40 (dd, J = 23.8, 6.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 169.9, 135.6, 128.6, 128.3, 128.2, 87.0 (d, J = 167.2 Hz), 66.6, 41.9 (d, J = 24.0 Hz), 20.8 (d, J = 22.3 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –172.54 (ddqd, J = 47.6, 28.3, 23.7, 14.2 Hz); MS(ESI) calcd for C₁₁H₁₃FO₂Na (M+Na)⁺ 219.1, found 219.1.

S-Fluoro-3-methylbutanoic acid (14).^{S10} Prepared from isovaleric acid (30.6 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (15 h), purified by flash column chromatography (20% ethyl acetate/hexanes), and concentrated by rotary evaporator at ca. 100 mm Hg to give 14 (19.5 mg, 81% yield) as a colorless oil: $R_f = 0.2$ (30% ethyl acetate/hexanes). ¹H NMR (400 MHz, C₆D₆) δ 10.68 (brs, 1H), 2.36 (d, J = 15.8 Hz, 2H), 1.23 (d, J = 21.3 Hz, 6H); ¹³C NMR (100 MHz, C₆D₆) δ 176.6 (d, J = 8.9 Hz), 92.6 (d, J = 170.1 Hz), 45.8 (d, J = 26.6 Hz), 26.7 (d, J = 24.0 Hz); ¹⁹F NMR (376 MHz, C₆D₆) δ –134.48 (heptt, J = 21.3, 15.8 Hz, 1F); MS(ESI) calcd for C₅H₉FO₂Na (M+Na)⁺ 143.1, found 143.1.

 $\mathbf{o} = \left(\mathbf{f} \\ \mathbf{f}$



(2*R*)-*N*-Phthaloyl-3-fluorovaline methyl ester (16).^{S 12} Prepared from (*S*)-*N*-phthaloylvaline methyl ester (52.2 mg, 0.2 mmol, 1.0 equiv) with selectfluor (106.5 mg, 0.3 mmol, 1.5 equiv) catalyzed by acetophenone (4.8 mg, 0.02 mmol, 20 mol%) according to the general procedure (48 h) and purified by flash column chromatography

(10% ethyl acetate/hexanes) to give **16** (47.6 mg, 85% yield) as a colorless oil: $R_f = 0.2$ (20% ethyl acetate/hexanes). ¹H NMR (400 MHz, CDCl₃) δ 7.89 (dd, J = 5.4, 3.1 Hz, 2H), 7.76 (dd, J = 5.4, 3.1 Hz, 2H), 4.98 (d, J = 10.2 Hz, 1H), 3.72 (s, 3H), 1.69 (d, J = 22.9 Hz, 3H), 1.55 (d, J = 22.4 Hz,

^{S9} K.-Y. Kim, B. C. Kim, H. B. Lee and H. Shin, J. Org. Chem. 2008, 73, 8106.

^{S10} R. Keck and J. Rétey, *Helv Chim Acta* **1980**, *63*, 769.

^{S11} W. Liu, X. Huang, M.-J. Cheng, R. J. Nielsen, W. A. Goddard III and J. T. Groves, *Science* **2012**, *337*, 1322.

^{S12} J.-B. Xia, Y. Ma and C. Chen, Org. Chem. Front. 2014, 1, 468.

3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.3, 166.6 (d, J = 10.5 Hz), 134.3, 131.6, 123.7, 95.6 (d, J =172.9 Hz), 57.7 (d, J = 25.9 Hz), 52.5, 26.3 (d, J = 22.8 Hz), 23.7 (d, J = 23.7 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –136.53 (dqg, J = 10.2, 22.6, 22.6 Hz, 1F); MS(ESI)⁺ calcd for C₁₄H₁₄FNO₄Na (M+Na)⁺ 302.1, found 302.1.



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1-(2-Fluoropropan-2-yl)-4-methyl-7-oxabicyclo[2.2.1]heptane (17).^{S12} Prepared from 1,4-Cineole (30.9 mg, 0.2 mmol, 1.0 equiv) with selectfluor (106.5 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (20 h), purified by flash column chromatography (2% diethyl ether/pentane), and concentrated by rotary evaporator at ca. 100 mm Hg to give 17 (25.1 mg, 73% yield) as a colorless oil: $R_f = 0.4$ (5% diethyl ether/pentane). ¹H NMR (400 MHz, C_6D_6) δ 1.80 (ddd, J = 12.9, 10.6, 4.0 Hz, 2H), 1.24–1.43 (m, 15H); ¹³C NMR (100 MHz, C_6D_6) δ 94.9 (d, J = 172.0 Hz), 89.7 (d, J = 21.1 Hz), 83.7, 37.2, 32.5 (d, J = 5.3 Hz), 23.5 (d, J = 25.6 Hz), 21.3; ¹⁹F NMR (376 MHz, C_6D_6) δ –146.22 (hept, J = 21.4 Hz, 1F); MS(EI) calcd for $C_{10}H_{17}FO(M)^+$ 172.1,

found 172.1.



(2R,5R)-2-(2-Fluoropropan-2-yl)-5-methylcyclohexanone (18).^{S12} Prepared from Lmenthone (15.4 mg, 0.1 mmol, 1.0 equiv) with selectfluor (53.1 mg, 0.15 mmol, 1.5 equiv) in CH₃CN (1.5 mL) according to the general procedure (30 h), purified by preparative TLC (plate pretreated with ammonium (7.0 M in methanol, 1 mL) in hexane (50 mL)) (5% ethyl acetate/hexanes), and concentrated by rotary evaporator at ca. 100 mm

Hg to give **18** (12.3 mg, 71% yield). ¹H NMR (400 MHz, C_6D_6) δ 2.30 (dt, J = 12.8, 6.3 Hz, 1H), 2.09– 2.18 (m, 2H), 1.55 (d, J = 23.2 Hz, 3H), 1.45 (d, J = 22.8 Hz, 3H), 1.32–1.47 (m, 3H), 1.18–1.28 (m, 1H), 0.77–0.89 (m, 1H), 0.61 (d, J = 6.0 Hz, 3H); ¹³C NMR (100 MHz, C₆D₆) δ 207.9 (d, J = 11.0 Hz), 95.3 (d, J = 165.5 Hz), 58.9 (d, J = 22.5 Hz), 51.2 (d, J = 4.3 Hz), 35.8, 33.7, 27.8 (d, J = 4.7 Hz), 27.6 (d, J = 23.3 Hz), 22.3 (d, J = 23.4 Hz), 22.2; ¹⁹F NMR (376 MHz, C₆D₆) δ –133.73 (heptd, J = 23.0, 7.4Hz, 1F); MS(EI) calcd for $C_{10}H_{17}FO(M)^+$ 172.1, found 172.1.

Fluorohexanes (F-19).^{S 13} Prepared from hexane (86.2 mg, 1.0 mmol, 5.0 equiv) according to the general procedure (15 h) to give 2-fluorohexane F-19a (50% NMR yield), 3-fluorohexane F-19b (22% NMR yield), and 1-fluorohexane F-19c (3% NMR F-19 vield). **F-19a**: ¹⁹F NMR (376 MHz, CD₃CN) δ –172.17 (ddqd, J = 48.3, 27.9, 24.1, 17.1 Hz, 1F); **F-19b**: ¹⁹F NMR (376 MHz, CD₃CN) δ –181.79 (m, 1F); **F-19c**: ¹⁹F NMR (376 MHz, CD₃CN) δ –218.46 (tt, *J* = 47.6, 25.1 Hz, 1F).



Fluorohexan-2-ones (F-20). Prepared from 2-hexanone (30.0 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (48 h) to give 5-fluorohexan-2-one F-20a (34% NMR yield), and 4-fluorohexan-2-one F-20b (26% NMR yield). F-20a: ¹⁹F NMR (376 MHz, CD₃CN) δ –174.13 (m, 1F); **F-20b**: ¹⁹F NMR (376 MHz, CD₃CN) δ –181.77 (m, 1F).



Fluorosclareolides (F-21).^{S11} Prepared from sclareolide (30.0 mg, 0.3 mmol, 1.5 equiv) according to the general procedure (24 h) to give 2α-fluorosclareolide F-21a (59% NMR yield), 2β-fluorosclareolide F-21b (6% NMR yield) and 3αfluorosclareolide **F-21c** (18% NMR yield). Purified by flash column chromatography (10% ethyl acetate/hexanes) to give a mixture of all three isomers (43.0 mg, 80% yield) as a colorless solid: $R_f = 0.3$ (20% ethyl acetate/hexanes). F-21a: ¹⁹F NMR $(376 \text{ MHz}, \text{CDCl}_3) \delta -179.85 \text{ (ddddd}, J = 48.0, 11.0, 11.0, 5.5, 5.5 \text{ Hz}, 1\text{F}); \text{F-21b}:$

^{S13} I. Bucsi, B. Török, A. I. Marco, G. Rasul, G. K. S. Prakash and G. A. Olah, J. Am. Chem. Soc., 2002, 124, 7728.

¹⁹F NMR (376 MHz, CDCl₃) δ –171.12 (m, 1F); **F-21c**: ¹⁹F NMR (376 MHz, CDCl₃) δ –187.33 (ddd, J = 46.3, 46.5, 14.6 Hz, 1F).











f1 (ppm)









2,2739



110 100 90 f1 (ppm) ò 130 120 80 70 60 30 20 10 200 190 180 170 160 150 140 50 40

F. O.

135



-172.3 -172.4 -172.5 -172.6 -172.7 f1 (ppm)





S27



-50 -130 -140 f1 (ppm) -70 -100 -110 -120 -150 -160 -200 -210 -60 -80 -90 -170 -180 -190 -220











^{100 90} f1 (ppm) <u>110</u>





-20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 -230 -240 -250 f1 (ppm)



^{-140 -150} f1 (ppm) -40 -110 -120 -160 -170 -210 -50 -60 -70 -80 -90 -100 -130 -180 -190 -200 -220 -230 -240 -25(



S36

-11, 914 -11, 948 -11, 1864 -11, 1864 -11, 1864 -11, 187

















