

Supplementary Figure 1: Voltage dependence of the co-translocational unfolding of Trx V5-C109-oligo(dC)₃₀ (C-terminus first)¹ and Trx oligo(dC)₃₀-C1S-V5 (N-terminus first) within the α HL pore. (a) Voltage dependence of the rate of step 2 \rightarrow 3: (O) V5-C109-oligo(dC)₃₀; (Δ) oligo(dC)₃₀-C1S-V5. (b) Voltage dependence of the rate of step 3 \rightarrow 4: (O) V5-C109-oligo(dC)₃₀; (\Box) oligo(dC)₃₀-C1S-V5, fast pathway; (Δ) oligo(dC)₃₀-C1S-V5, slow pathway. (c) Voltage dependence of the rate of step 4 \rightarrow 1: (O) V5-C109-oligo(dC)₃₀; (Δ) oligo(dC)₃₀-C1S-V5. Each data point for the N-terminus-first measurements is derived from an exponential fit to data obtained from at least 3 different pores (n = 3).



Supplementary Figure 2: Urea-unfolding curves measured by circular dichroism. (a) Trx S1C-V5, the solid line represents the fit to a two-state model (values from the fit are given in the main text). (b) Trx \bigtriangledown 6G-V5, the solid line represents the fit to a two-state model (values from the fit are given in the main text). Both experiments were carried out in 1 mM DTT, 2 M KCl, pH 7.2, 22 °C. Ellipticity was measured at 222 nm.



Supplementary Figure 3: Dwell time distributions in level 3 for N-terminus threading into the α HL pore. (a) For Trx $oligo(dC)_{30}$ -S1C-V5, the two unfolding pathways are equally populated. (b) N-terminus threading of Trx $oligo(dC)_{30}$ -P22A-V23I-V5. In this case, most of the Trx molecules go through the slow pathway for step 3 \rightarrow 4. Both distributions were fitted to a double exponential function (black line). The dashed lines show the deconvolution into single exponentials. Data from at least 3 different pores were used to construct each histogram.



Supplementary Figure 4: Kinetics of steps $2\rightarrow 3$ and $3\rightarrow 4$ in the presence of urea. (a) Step $2\rightarrow 3$, +140 mV. (b) Step $2\rightarrow 3$, +120 mV. (c) Step $2\rightarrow 3$, +100 mV. (d) Step $3\rightarrow 4$, +140 mV. (e) Step $3\rightarrow 4$, +120 mV. (f) Step $3\rightarrow 4$, +100 mV. Each data set was fitted to a straight line and the average of slopes between voltages used to derive the kinetic m-values. Each point was obtained from data from at least 3 different pores.



Supplementary Figure 5: Current measurements for the translocation of Trx oligo(dC)₃₀-S1C-V5 through the α HL pore (N-terminus first). (a) Four events are displayed: Event (i) has a total duration of >100 s and a noisy level 3. Events (ii) and (iii) have total durations of <5 s and normal levels 3. Event (iv) has a total duration of >100 s and a normal level 3. (b) Zoom showing the noisy level 3 of event (i). Events with a noisy level 3 represented 40% of the total number of events analysed (91% of the total: see the text). The events show similar kinetics, whether level 3 is noisy or not (Supplementary Fig. 6). Level 3 occasionally switched from a noisy state to the normal state, and vice versa. We attribute the differences in noise to different arrangements of the polypeptide when threaded within the pore.



Figure 6: Dwell time distributions from events that show a noisy level 3 after N-terminus threading. (a) Histogram of dwell times in level 2 obtained at +140 mV. Solid line, fit to a single exponential distribution with rate $k_{2-3} = 323 \pm 17 \text{ s}^{-1}$. (b) Histogram of dwell times in level 3 obtained at +140 mV. Solid line, fit to a double exponential distribution with rates $k_{3a-4} = 0.26 \pm 0.03 \text{ s}^{-1}$ and $k_{3b-4} = 0.0054 \pm 0.0003 \text{ s}^{-1}$. (c) Histogram of dwell times in level 4 obtained at +140 mV; solid line, fit to a single exponential distribution with rate $k_{4-1} = 107 \pm 6 \text{ s}^{-1}$. Data from at least 10 different pores were used to construct each histogram.

Supplementary Reference

1.- Rodriguez-Larrea, D. and Bayley, H. Multistep protein unfolding during nanopore translocation. *Nat. Nanotechnol.* **8**, 288-95 (2013).