SUPPLEMENTARY INFORMATION

Hippocampal Memory Traces Are Differentially Modulated By Experience, Time,

And Adult Neurogenesis

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INVENTORY OF SUPPLEMENTAL INFORMATION

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Figure S1. ArcCreER^{T2} x R26R-STOP-floxed-EYFP recombination efficiency. (A) Mice were injected with TAM and then administered a 1- or 4-shock CFC paradigm 5 h later. Mice were sacrificed 1 h following the start of CFC and tissue was processed for Cre recombinase and Arc. (B) To assess recombination efficiency, confocal microscopy was then used to assess co-localization of Arc and Cre-recombinase in the DG. Over 99% of Arc⁺ cells expressed nuclear Cre recombinase, regarßdless of the CFC protocol administered. (C) Representative image of Cre recombinase and Arc. (D) Mice were injected with vehicle (Veh) and then administered the contextual fear conditioning (CFC) paradigm 5 h later. Mice were sacrificed 5 days later and tissue was processed for EYFP. (E) $ArcCreER^{T2} \times R26R$ -STOP-floxed-EYFP exhibit low recombination in the DG, both dorsally and ventrally (dorsal average: 4.994 EYFP⁺ cells per section; ventral average: 4.611 EYFP⁺ cells per DG section). (F-I) Representative images of EYFP⁺ cells in the DG following a Veh injection. Arrows indicate EYFP⁺ cells. (J) Binding of TAM releases CreER^{T2} from heat shock complexes in the cytoplasm, allowing for liganddependent translocation to the nucleus, where it can direct recombination between the loxP sites. Representative images of Cre recombinase staining in Veh-injected and TAMinjected mice. Error bars represent + SEM. See also Figure 1.

Figure S2. EYFP expression throughout the brain following 1-shock CFC. (A) Mice were injected with TAM and 1-shock CFC paradigm was administered 5 h later. Mice were sacrificed 5 days following the CFC training. (**B-M**) Representative images throughout the brain of EYFP expression. S: subiculum, VC: visual cortex, AC: auditory cortex, DMC: dorsomed hypothal nu, compact, DG: dentate gyrus, CA3: Cornu

Ammonis region 3, CPu: caudate putamen, DP: dorsal peduncular cortex, IL: infralimbic cortex, PrL: prelimibic cortex, CE: central amygdaloid, BLA: basolateral amygdala. See also Figure 2.

Figure S3. Arc and c-fos expression patterns differ dramatically in CA3. (A-B) 10X and 20X images, respectively, of EYFP and Arc expression in the HPC. While Arc labeling in the DG is relatively somatic, the labeling becomes primarily dendritic in CA3. (C-D) 10X and 20X images, respectively, of EYFP and c-fos expression in the HPC. cfos labeling is relatively somatic, not only in the DG, but in CA3 as well, making colocalization studies in CA3 much easier by using c-fos as a marker of activity.

See also Figure 3.

Figure S4. *In vivo* optogenetic inhibition of CA2 does not impair expression of initially encoded memory in ArcCreER^{T2} x R26R-CAG-STOP-floxed-Arch-GFP mice. (A) Genetic design. (B) Representative fiber tracts in the DG, CA3, and CA2 implanted mice, and their respective coordinates. (C) Representative trace of DG Arch-GFP⁺ neurons. (D) Arch-GFP expression does not differ in mice labeled in context A or in context C. (n = 3-4 mice per group). (E) Experimental design. Mice were implanted with fiber optics into mainly CA2 and allowed >2 weeks to recover. Mice were then injected with TAM and administered 4-shock CFC 5 h later. Two weeks later, mice were placed back into context A for 6 min. Two days later mice were placed into a novel context B for 6 min. (F-G) ArcCreER^{T2}(+) and (-) did not differ in fear expression during or following laser inhibition in either the training context A or novel context B during the first minute of light ON or the first minute of light OFF (minute 1 (light ON) versus minute 4 (light OFF): repeated-measures ANOVA, genotype effect, F(1,19) = 0.002, p =

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0.96; light ON/OFF effect, F(1,19) = 67.603, p < 0.0001; genotype x light ON/OFF, F(1,19) = 2.795, p = 0.11) (*t*-tests for light ON and for light OFF: p's > 0.05). Minutes 1-3 (light ON) versus minutes 4-6 (light OFF) were also analyzed and gave similar results (repeated-measures ANOVA, genotype effect, F(1,19) = 0.398, p = 0.54; light ON/OFF effect, F(1,19) = 74.947, p < 0.0001; genotype x light ON/OFF, F(1,19) = 0.224, p =0.64). (n = 9-12 mice per group). Error bars represent \pm SEM. See also Figure 5.

Figure S5. In vivo optogenetic inhibition of CA3 impairs expression of initially encoded memory in ArcCreER^{T2} x R26R-CAG-STOP-floxed-eNpHR3.0-EYFP mice. (A) Genetic design. (B-D) Representative EYFP images of the HPC, DG, and CA3 at 2 weeks post TAM injection. (E) Representative eNpHR3.0-EYFP⁺ CA3 pyramidal cell. (F-G) In CA3, in vitro photostimulation resulted in complete inhibition of APs. Voltage (upper trace) and current clamp (lower trace) recording of an eNpHR3.0-EYFP⁺ CA3 neuron. In vitro photostimulation resulted in 94.58 ± 17.23 pA steady state current in eNpHR3.0-EYFP⁺ CA3 neurons, which corresponds to -18.01 ± 1.73 mV hyperpolarization (n = 3 cells per group). (H) Experimental design. Mice were implanted with fiber optics into CA3 and allowed >2 weeks to recover. Mice were then injected with TAM and administered 4-shock CFC 5 h later. Two weeks following the CFC training, mice were placed back into the training context A for 6 min. The first 3 min of context A exposure were with light ON, with the following 3 min were with light OFF. (I) ArcCreER^{T2}(+) mice expressed significantly less fear during laser inhibition when compared with ArcCreER^{T2}(-) mice (minute 1 (light ON) versus minute 4 (light OFF): repeated-measures ANOVA, genotype effect, F(1,16) = 5.278, p = 0.04; light ON/OFF

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effect, F(1,16) = 3.912; p = 0.07; genotype x light ON/OFF, F(1,16) = 5.073, p = 0.04). Minutes 1-3 (light ON) versus minutes 4-6 (light OFF) were also analyzed and gave similar results (repeated-measures ANOVA, genotype effect, F(1,16) = 5.176, p = 0.04; light ON/OFF effect, F(1,16) = 1.360, p = 0.2606; genotype x light ON/OFF, F(1,16) = 2.523, p = 0.13; planned comparison for light ON, p = 0.0085, planned comparison for light OFF, p = 0.24). ArcCreER^{T2}(+) and (-) did not differ in fear expression following laser inhibition (p = 0.95). (n = 8-10 mice per group). * p < 0.05. Error bars represent \pm SEM. See also Figure 5.

Figure S6. Social defeat results in a depressive-like phenotype in ArcCreER^{T2} x

R26R-STOP-floxed-EYFP mice. (A-B) Body weight did not differ before the start of and following social defeat (SD). (C) In the DI paradigm, SD mice exhibited significantly less approaches to the CD-1 aggressor (p = 0.04) and significantly more approaches to the empty enclosure (p < 0.01). (D) The interaction quotient was significantly less in SD mice (p = 0.02). (E-F) In NOR paradigm, general activity and investigation declined across exposures 1-4 for Ctrl and SD mice. (G) SD mice investigated the novel object more than Ctrl mice (p = 0.02). (H) In the EPM, SD mice spend less time in the open arms (p = 0.02). (I-L) In the OF, no differences were detected between Ctrl and SD mice. (M) In the TST, SD mice displayed increased immobility [F(1,18) = 8.614, p < 0.01]. (N) TAM does not alter the 1-shock CFC impairment in SD mice. Context-elicited freezing was significantly reduced in SD mice. [F(1,17) = 4.593, p < 0.05]. (O) The number of BrdU⁺ cells was significantly less in SD mice in the most ventral DG section (p < 0.01). (n = 4-13 mice / group). * p < 0.05. ** p < 0.01. Error bars represent \pm SEM. See also Figure 7.

Figure S7. Example fear conditioning traces during memory encoding and expression in ArcCreER^{T2} x R26R-CAG-STOP-floxed-eNpHR3.0 mice. (A) ArcCreER^{T2}(-) and (+) mice did not differ in freezing behavior during the training experience (genotype: F(1,17) = 0.355, p = 0.56; time: F(1,5) = 9.853, p = <0.0001; time x genotype interaction: F(1,5) = 0.768, p = 0.58). (B) Optogenetic inhibition of eNpHR3.0-YFP⁺ CA3 neurons impaired expression of the corresponding fear memory in context A in ArcCreER^{T2}(+) mice when compared with ArcCreER^{T2}(-) mice (Data shown as bar graph in Figure S07). (n = 8-10 mice per group). Error bars represent \pm SEM. See also Figure 5.