Supplemental materials:

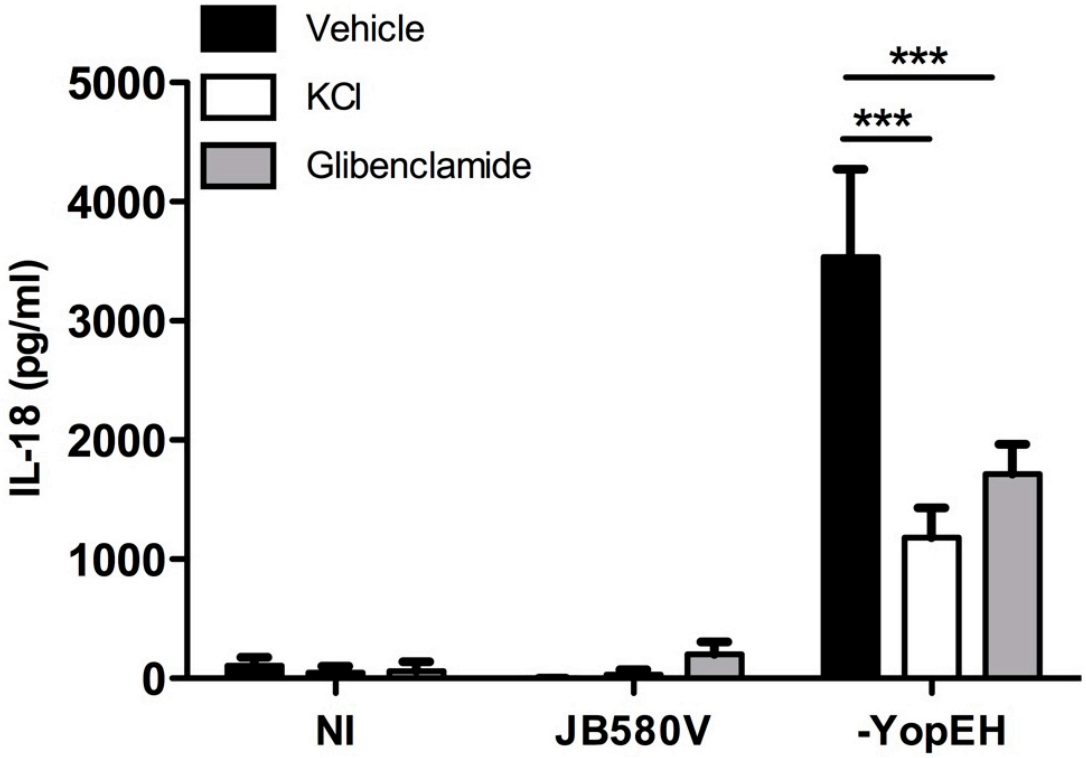

Figure S1. IL-18 secretion is dependent on K^+ efflux. (Related to Figure 2) Caco-2 cells were treated with 50μM glibenclamide, 130mM KCl, or DMSO for 30 min before infection with wild type or *yopEH* mutant *Y. enterocolitica*. Cell culture supernatants were collected post infection and analyzed for secreted IL-18 by ELISA. The data represent the mean ± SD of triplicate samples from two independent experiments. *$p < 0.001$ versus vehicle.

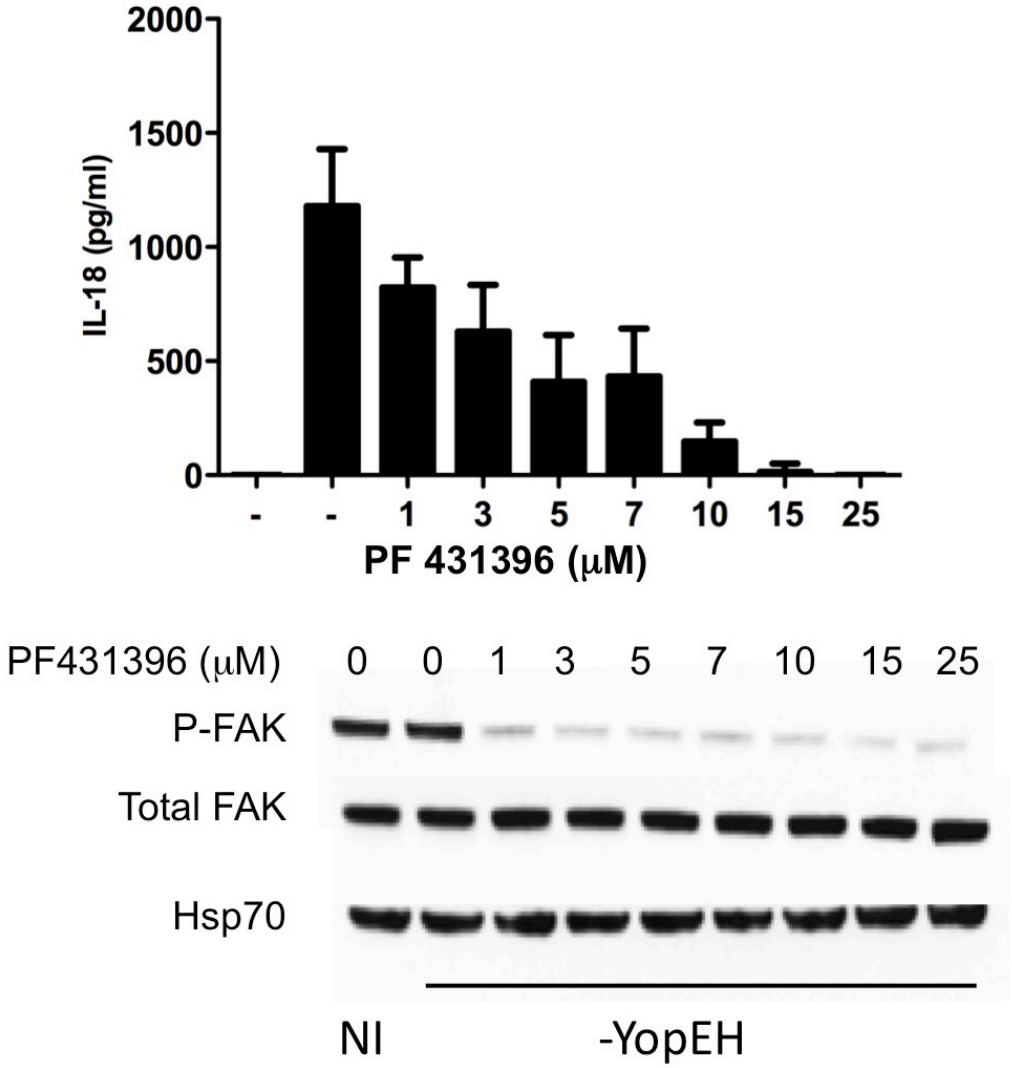

Figure S2. IL-18 secretion is dependent on the activity of FAK. (Related to Figure 5) Caco-2 cells were pre-treated with increasing concentrations of the FAK inhibitor PF-431396 or DMSO (0μM) for 1h and then infected with indicated bacterial strains at an MOI of 10. After 8 h, ELISA determined the concentration of IL-18 in the conditioned media. Cells were also lysed and subjected to immunoblot analysis of phospho-FAK, total FAK, and the loading control Hsp70. Data represents mean ± SD of two independent experiments. *$p < 0.001$.

Table S I: Bacterial strains for infection studies

Strain	Description	Source or Reference
JB580V	Wild-type *Y. enterocolitica* 8081v -*yenR*(r^-m^+)	(Kinder et al., 1993)
JB580C	JB580V cured of virulence plasmid pYVe8081; lacks Ysc/Yop T3SS	(Pepe and Miller, 1993)
-YopP	JB580V *yopP*	(Bose et al., 2012)
-YopE	JB580V *yopE*	This work
-YopH	JB580V *yopH*	This work
-YopEH	JB580V *yopEH*	This work
-YopQ	JB580V *yopQ*	This work
-Inv	JB580V *inv*	gift from Virginia Miller (UNC Chapel Hill)
JB580C -Inv	JB580C *inv*	This work
-YopEH -Inv	JB580V *yopEH inv*	This work
STm	*Salmonella enterica* subsp. *enterica* (*ex* Kauffmann and Edwards) Le Minor and Popoff serovar Typhimurium	(ATCC® 14028™)
E. coli Inv	HB101 (pMV101) *E. coli* expressing *Y.enterocolitica* Invasin	(Pepe and Miller, 1990); gift from Virginia Miller (UNC Chapel Hill)
E. coli vector	HB101 pMV101	(Pepe and Miller, 1990);
E. coli InvD760A	*E. coli* expressing inv with D760A	This work
E. coli InvD809A	*E. coli* expressing inv with D809A	This work

Table S II: Primers used in this study:

Oligo sequence 5'-3'	
YopE- Upstream Forward: GTCGACGATGAGGTTTGTAGCCGTTC Reverse: TTAATTAACTTAGTGGGAAAATAGCCGG	YopE- Downstream Forward: TTAATTAAGTGATATGGTGACTAGTCCTGC Reverse: CCCTGCCTCTTACTCTACTTC
YopH- Upstream Forward: GTCGACGATAATACCCCCAGGTTGA Reverse: TTAATTAA GGTGAGCCGTGTATTTTCAT	YopH-Downstream Forward: TTAATTAAGGACAAGGGCGACCATTATT Reverse: CTGGCACCTATCAATGAGAAGG
YopQ- Upstream Forward: GTCGACGATGCAGAACTGGCGAATAC Reverse: TTAATTAAGAGATCTTCCGGCATACTGAC	YopQ- Downstream Forward: TTAATTAACTACTCTCAATGAGCTTCCCATG Reverse: GTAGTGTCCGTGAATACTGTCC

Primers used for RT-PCR

Oligo sequence 5'-3'

GAPDH
Forward: GTCAGTGGTGGACCTGACCT
Reverse: AGGGGTCTACATGGCAACTG
Caspase-1
Forward: ATCCGTTCCATGGGTGAAGGTACA
Reverse: CAAATGCCTCCAGCTCTGTA
NLRP3
Forward: TTCTTTCTGTTTGCTGAGTTTTTG
Reverse: TTCCTGGCATATCACAGTGG
Primers used for invasin site-directed mutagenesis
Oligo sequence 5'-3'
Inv D760A
Forward: GCCAGCGCACTGCCTTTACTGCGCTGATC
Reverse: GATCAGCGCAGTAAAGGCAGTGCGCTGGC
Inv D809A
Forward : GACCAGTACAGCCTTTGTCACTATGG
Reverse: CCATAGTGACAAAGGCTGTACTGGTC