SUPPLEMENTARY METHODS: Plasmid construction

Low (centromeric, pJNY347) and high (2μ , pJNY339) copy plasmids containing *LHP1* (with promoter and terminator regions, amplified from genomic DNA) were constructed in pRS315 (77) and pRS425 (78) respectively. *LHP1*^{F51A} was generated by PCR. 2 fragments were amplified from plasmid pJNY339 using T7 promoter primer plus BSAI51Aup (5'-

CCAGGGTCTCGCGCAACGCCCTGTCATATGGAAAGTTG; Bsal site in italics altered codon 51 underlined) and T3 promoter primer plus BSAI51Ado (5'-CCAGGGTCTCTTGCGCACAACAGCG). These fragments were digested with Bsal, ligated and cloned into pRS425. Cloned fragments were sequenced to confirm accuracy.

SUPPLEMENTARY TABLE S1: Yeast strains used in this study.

Strain	Genotype	Reference/Source
JDY3	MATα, trp1, ura3, ade2, his3, lys2	(59)
JDY8	sec65∆::HIS3, [rho-], otherwise as JDY3	(79)
JDY44	srp21∆::HIS3, [rho-], otherwise as JDY3	(59)
JDY66	srp14∆::HIS3, [rho-], otherwise as JDY3	(59)
JDY97	srp68∆::HIS3, [rho-], otherwise as JDY3	(59)
JDY99	srp72∆::HIS3, [rho-], otherwise as JDY3	(59)
JDY103	scr1∆::HIS3, [rho-], otherwise as JDY3	(59)
JDY335	MATα, srp14::HIS3, [rho-], trp1-1, ura3-1, ade2-1, his3-11, -15, leu2-3, -112, can1-100	This study
JDY483	MAT a , <i>trp1-1</i> , <i>ura3-1</i> , <i>ade2-1</i> , <i>his3-11</i> , <i>-15</i> , <i>leu2-3</i> , <i>-112</i> , <i>can1-100</i> , <i>scr1Δ::TRP1</i> , pRS316- <i>SCR1</i>	(57)
JDY596	srp72::GFP-kanMX3, otherwise as JDY3	This study
JDY618	MAT α , mtr4-1, ura3-52, lys2-801, pep4::HIS3, prb1- Δ 1-6R, GAL+	Alan Tartakoff
JDY721	MAT a , ura3-1, ade2-1, his3-11, -15, leu2-3, -112, can1-100, trp1-1::(TRP1, GFP-MS2)	This study
JDY726	MAT a , ura3-1, ade2-1, his3-11, -15, leu2-3, -112, can1-100, trp1-1::(TRP1, GFP-MS2), scr1::MS2X6-HIS3	This study
JDY895	MAT a , his3 Δ 1, leu2 Δ 0, met15 Δ 0, ura3 Δ 0	(80)
JDY898	<i>trf4∆::kanMX6</i> , <i>HisMX6-pGAL-3HA::trf5</i> , otherwise as JDY895	(14)
JDY899	air2∆::kanMX4, otherwise as JDY895	(11)

JDY900 JDY901 JDY906 JDY910 JDY911	air1Δ::kanMX4, otherwise as JDY895 air1Δ::kanMX4, air2Δ::natMX6, otherwise as JDY895 rrp6Δ::HIS3MX6, otherwise as JDY483 lhp1Δ::HIS3MX6, otherwise as JDY483	(11) (11) This study
JDY906 JDY910	JDY895 rrp6∆::HIS3MX6, otherwise as JDY483	
JDY910	•	This study
	<i>Ihp1∆::HIS3MX6</i> , otherwise as JDY483	
JDY911		This study
	<i>trf4∆::HIS3MX6</i> , otherwise as JDY483	This study
JDY912	<i>trf5∆::HIS3MX6</i> , otherwise as JDY483	This study
JDY974	rex1∆::kanMX6 otherwise as JDY895	Euroscarf
JDY978	<i>lsm1∆::kanMX4</i> , otherwise as JDY895	Jean Beggs
JDY981	<i>Ism6∆:: hphMX6</i> , otherwise as JDY895	Jean Beggs
JDY982	<i>Ism7∆::hphMX6</i> , otherwise as JDY895	Jean Beggs
JDY983	<i>lsm5∆::hphMX6</i> , otherwise as JDY895	Jean Beggs
JDY984	<i>lsm4∆::kanMX4</i> , otherwise as JDY895	Jean Beggs
JDY990	natNT2-pGAL(S)::mtr4, otherwise as JDY483	This study
JDY1015	natNT2-pGAL(L)::rrp44, otherwise as JDY483	This study
JDY1018	natNT2-pGAL(L)::Ihp1, otherwise as JDY483	This study
JDY1028	rex1∆::natNT2, otherwise as JDY483	This study
JDY1042	<i>Ihp1∆::HIS3MX6</i> , <i>rex1∆::natMX6</i> , otherwise as JDY483	This study
JDY1058	natNT2-pGAL(L)::Ihp1, trf4∆::HIS3MX6, otherwise as JDY483	This study
JDY1083	natNT2-pGAL(S)::mtr3, otherwise as JDY483	This study
JDY1084	natNT2-pGAL(L)::rrp42, otherwise as JDY483	This study
JDY1093	srp21::GFP-HIS3MX6, otherwise as JDY895	This study
JDY1156	$air1\Delta$:: $kanMX6$, $air2\Delta$:: $natMX6$, $lhp1\Delta$:: $HIS3MX6$, otherwise as JDY895	This study
JDY1199	<i>Ihp1∆::natNT2</i> otherwise as JDY335	This study
JDY1201	rrp6∆::natNT2 otherwise as JDY335	This study
JDY1203	<i>trf4∆::kanMX6</i> otherwise as JDY335	This study
JDY1239	$rex1\Delta$:: $kanMX4$, $srp14\Delta$:: $HIS3$, $[rho-]$, $ura3$, $his3$, $leu2$ *	This study
JDY1241	rrp44∆::kanMX6, pRS315-rrp44-exo-HTP, srp14∆:HIS3, ura3, his3, leu2 *	This study

CS-66	<i>rrp44∆::kanMX6</i> , pRS315- <i>RRP44-HTP</i> , otherwise as JDY895	(3)
CS-77	<i>rrp44::kanMX6</i> , pRS315- <i>rrp44-exo-HTP</i> , otherwise as JDY895	(3)
CS-31	trf4::HTP-URA3, otherwise as JDY895	(50)

^{*} The mating type of these strains has not been determined.

SUPPLEMENTAL TABLE S2 The 3' terminus of scR1.

The 3' terminus of scR1 was inferred by reverse transcription-PCR after ligating a 3'-blocked DNA oligonucleotide onto RNA isolated from the cells indicated. PCR products were sequenced directly and, where indicated, cloned and sequenced individually. The genomic sequence at the 3' end of *SCR1* is:

...CATATTTTTAACA... the run of 6 thymidines comprising the pol III terminator

Genotype	RT-PCR product	Cloned RT-PCR products	No. of clones
wild type	CAUAUUUU(U)*	CAUAUUUU	11
• •		CAUAUUUUU	6
		CAUAUUUUU	1
		CAUAUUU	1
		CAUA	1
Ihp1∆	CAUAU(U/A)(U/A)(U/A)	CAUAAAAA	2
,	, ,, ,, ,,	CAUAU	2
		CAUAUAAA	1
		CAUAUU	3
		CAUAUUA	2
		CAUAUUAA	1
		CAUAUUAAA	4
		CAUAUUU	4
		CAUAUUUAA	4
		CAUAUUUU	1
Ihp1 ^{F51A}	CAUAU(U/A)(U/A)(U/A)		
rex1∆	CAUAUÙUUÛ(U)	CAUAUUUU	1
TOXTE	(_ ,	CAUAUUUU	8
		CAUAUUUUU	4
Ihp1 Δ , rex1 Δ	CAUAUU(U/A)(U/A)	CAUAU	6
пртд, техтд		CAUAUAAA	2
		CAUAUAAAA	1
		CAUAUU	2
		CAUAUUU	1
		CAUAUUUAA	1
trf4∆	CAUAU(UUUU)	AAUUGUGC ₍₄₂₀₎ [†]	1
UITA		CCAUCAGG ₍₄₆₂₎	1
		UUGGCGGU ₍₄₅₃₎	3
		CAUAU	2
		CAUAUA	1
		CAUAUU	2
		CAUAUUU	5
		CAUAUUUU	6
		CAUAUUUU	2
		CAUAUUUUA	1
		CAUAUUUUUU	2
		CAUAUUUUUUAACAUUUU [¶]	1
Ib = 4 4 4 4 4 4	CALIALICIDALICID	CUCGCACA ₍₄₇₅₎	1
Ihp1 Δ , $trf4\Delta$	CAUAU(U)(U)(U)		
		GUGUGUC ₍₅₀₇₎	1
		CAUAAAA	1
		CAUAU	10
		CAUAUU	1
		CAUAUUU	2
		CAUAUUUU	4

	T	T 2	
Ihp1 $∆$, $rrp6∆$	CAUAU(U)(U)	CAUAA	3
		CAUAAA	1
		CAUAAAA	1
		CAUAAAAA	1
		CAUAU	1
		CAUAUA	1 1
		CAUAUAA	1
		CAUAUAAA	1
			-
	0.411411	CAUAUUU	1
air1 Δ , air2 Δ	CAUAU	CAU	1
		CAUA	4
		CAUAU	10
		CAUAUAA	1
		CAUAUU	2
		CAUAUUU	1
		CAUAUUUU	3
air1∆	CAUAUUUU(U)		
air2∆	CAUAUUUU(U)		+
	` '	CALIAAAA	+
rrp6∆	CAUAU(U/A)(U/A)	CAUAAAA	1
		CAUAU	3
		CAUAUAAA	2
		CAUAUU	2
		CAUAUUAA	2
		CAUAUUU	1
		CAUAUUUU	1
rrp6 ^{D238N}	CAUAU(U/A)(U/A)	CAUAAAAA	1
'		CAUAU	4
		CAUAUAAA	2
		CAUAUUAAA	1
		CAUAUUUUU	l i
		CAUAUUUUU	1
rrp44-exo	CAUAU(U)(U)	CAU	1
11p44-6x0	CAOAO(O)(O)	CAUA	1
		CAUAAAA	3
		CAUAU	10
		CAUAUU	2
		CAUAUUAA	1
		CAUAUUU	4
		CAUAUUUU	2
		CAUAUUUUAA	1
		CAUAUUUU	5
rrp44-endo	CAUAUUUU(U)		
rrp44-exo,	CAUAU(U)(U)	CAUA	1
rrp6∆	()(-)	CAUAU	1
.,,,,,		CAUAUU	1
		CAUAUUUU	1
		CAUAUUUUU	
10.00 4.4	CALIALIIIIIIIIII		+ -
Ism4∆	CAUAUUUU(U)		
Ism5∆	CAUAUUUU(U)		
Ism6∆	CAUAUUUU(U)		
Ism7∆	CAUAUUUU(U)		
	` '	•	

^{*} Bracketed nucleotides indicate heterogeneity of the 3' end with either alternative nucleotides at a given position, or that the sequence indicated the presence of a

particular nucleotide (such as the 5th U in wild type scR1) in only part of the cDNA product.

[¶] this clone extends beyond the terminator of scR1 into the next stretch of T's in the template.

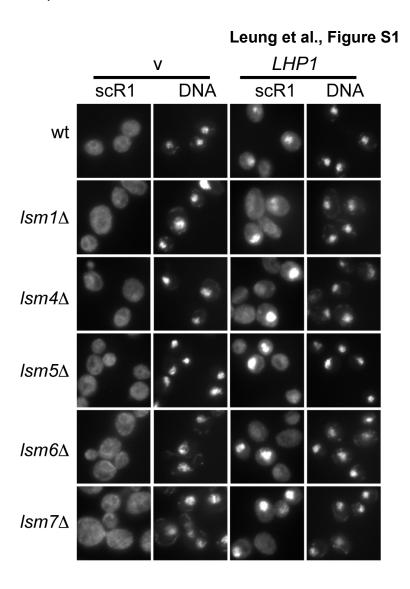


Figure S1 The nuclear LSM complex is not required for correct localisation of scR1, or for its nuclear accumulation on over-expression of Lhp1.

Fluorescence *in situ* hybridisation (FISH) was carried out on the cells indicated, which contained high copy (2μ) plasmids that were either empty (v) or contained *LHP1*, using fluorescently labelled probe scR1A (scR1). DNA was stained with DAPI (DNA). Lsm1 is exclusively part of the cytoplasmic Lsm complex, and provided a control, whereas Lsm4-7 are components of both the cytoplasmic and

[†]numbers in brackets refer to final nucleotide of scR1 present in the clone sequenced.

nuclear Lsm complexes. Individual channels are shown. Fluorescence images of cells were captured as described (Materials and Methods).

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