

Supporting Information
Bioorganic and Medicinal Chemistry Letters

Imidazo[1,2-*a*]pyridine-based peptidomimetics as inhibitors of Akt

Young B. Kim, Chang Won Kang, Sujeewa Ranatunga, Hua Yang,
Said M. Sebti, and Juan R. Del Valle*

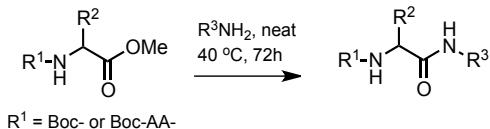
Drug Discovery Department, H. Lee Moffitt Cancer Center and Research Institute

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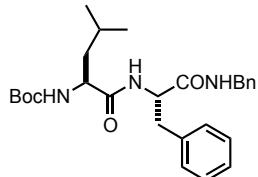
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EXPERIMENTAL PROCEDURES

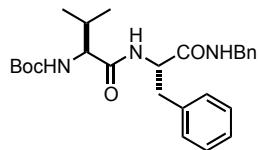
General synthesis notes. Unless stated otherwise, reactions were performed in flame-dried glassware under a positive pressure of argon or nitrogen gas using dry solvents. Commercial grade reagents and solvents were used without further purification except where noted. Diethyl ether, toluene, dimethylformamide dichloromethane, and tetrahydrofuran were purified by a Glass Contour column-based solvent purification system. Other anhydrous solvents were purchased directly from chemical suppliers. Thin-layer chromatography (TLC) was performed using silica gel 60 F254 pre-coated plates (0.25 mm). Flash chromatography was performed using silica gel (60 μ m particle size). The purity of all compounds was judged by TLC analysis (single spot/two solvent systems) using a UV lamp, CAM (ceric ammonium molybdate), ninhydrin, or basic KMnO₄ stain(s) for detection purposes. 1D and 2D NMR spectra were recorded on a 400 or 500 MHz spectrometer. Proton chemical shifts are reported as δ values relative to residual signals from deuterated solvents (CDCl₃ or DMSO-*d*₆).



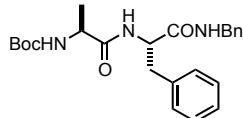
General procedure for C-terminal amidation of dipeptide methyl esters. A solution of the N-Boc α -amino methyl ester or N-Boc dipeptide methyl ester (500 μ mol) was dissolved in the desired amine and heated to 40 °C in a sealed tube and stirred until TLC indicated consumption of the starting material (3 to 4 days). The crude residue was diluted with EtOAc and washed with 1M aq. HCl. The organic layer was dried over Na₂SO₄, concentrated, and purified by flash chromatography over silica gel (EtOAc/Hexanes) to afford the desired amides.



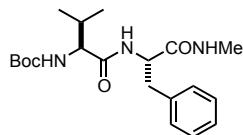
Boc-Leu-Phe-NHBn. Yield: 81%; ¹H NMR (400 MHz, CDCl₃) δ 7.27 – 7.13 (m, 9H), 7.07 (d, *J* = 6.8 Hz, 2H), 6.87 – 6.84 (m, 1H), 6.80 (d, *J* = 8.1 Hz, 1H), 4.91 (d, *J* = 5.6 Hz, 1H), 4.75 (dd, *J* = 14.7, 7.0 Hz, 1H), 4.32 – 4.29 (m, 2H), 4.06 – 4.02 (m, 1H), 3.17 (dd, *J* = 13.6, 5.8 Hz, 1H), 3.05 (dd, *J* = 13.6, 7.3 Hz, 1H), 1.53 – 1.49 (m, 2H), 1.31 (s, 9H), 0.87 (dd, *J* = 7.2, 6.7 Hz, 5.4H), 0.82 (dd, *J* = 6.1, 3.2 Hz, 0.6H; ¹³C NMR (101 MHz, CDCl₃) δ 172.5, 170.6, 155.9, 137.9, 136.6, 129.5, 128.7, 128.6, 127.8, 127.3, 127.0, 80.5, 54.1, 53.8, 43.5, 41.1, 38.1, 28.3, 24.8, 23.1, 21.8; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₂₇H₃₈N₃O₄ 468.2857, found 468.2849



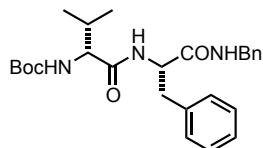
Boc-Val-Phe-NHBn. Yield: 70%; ¹H NMR (400 MHz, CDCl₃) δ 7.48 (d, *J* = 8.2 Hz, 1H), 7.37 – 7.35 (m, 1H), 7.24 – 7.11 (m, 8H), 6.97 (d, *J* = 5.5 Hz, 2H), 5.42 (d, *J* = 8.1 Hz, 1H), 4.89 (q, *J* = 7.5 Hz, 1H), 4.31 (dd, *J* = 14.9, 6.2 Hz, 1H), 4.12 (dd, *J* = 14.8, 5.2 Hz, 1H), 4.04 (t, *J* = 7.6 Hz, 1H), 3.10 – 3.04 (m, 2H), 1.96 (dq, *J* = 13.5, 6.8 Hz, 1H), 1.36 (s, 9H), 0.87 – 0.76 (m, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 171.9, 170.9, 156.0, 138.0, 136.6, 129.5, 128.6, 128.4, 127.6, 127.1, 126.8, 79.7, 60.1, 54.3, 43.2, 38.6, 30.9, 28.4, 19.3, 18.0; HRMS (ESI-TOF) (*m/z*) [MNa]⁺ calcd for C₂₆H₃₅N₃O₄Na 476.2520, found 476.2540.



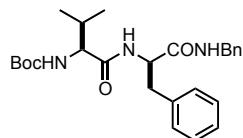
Boc-Ala-Phe-NHBn. Yield: 80%; ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 8.43 (t, $J = 5.5$ Hz, 1H), 7.89 (d, $J = 8.3$ Hz, 1H), 7.30 – 7.17 (m, 9H), 7.12 (d, $J = 7.2$ Hz, 2H), 7.00 (d, $J = 7.1$ Hz, 1H), 4.52 (dd, $J = 14.3, 8.0$ Hz, 1H), 4.25 (t, $J = 5.7$ Hz, 2H), 3.93 – 3.86 (m, 1H), 3.00 (dd, $J = 13.7, 5.6$ Hz, 1H), 2.88 (dd, $J = 13.4, 8.6$ Hz, 1H), 1.35 (s, 7.7H), 1.33 (s, 1.3H), 1.08 (d, $J = 7.2$ Hz, 2.7H), 0.94 (d, $J = 7.2$ Hz, 0.3H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ 172.5, 170.7, 155.2, 139.0, 137.6, 129.3, 128.2, 128.1, 127.1, 126.7, 126.3, 78.3, 53.8, 50.1, 42.0, 37.7, 28.2, 18.1; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{24}\text{H}_{32}\text{N}_3\text{O}_4$ 426.2387, found 426.2386.



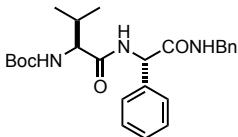
Boc-Val-Phe-NHMe. Yield: 87%; ^1H NMR (400 MHz, CDCl_3) δ 7.33 – 7.12 (m, 5H), 6.47 – 6.43 (m, 1H), 6.35 – 6.31 (m, 1H), 4.86 – 4.88 (m, 1H), 4.68 – 4.65 (m, 1H), 3.85 – 3.83 (m, 1H), 3.15 – 3.06 (m, 1H), 3.07 (dd, $J = 13.4, 6.7$ Hz, 1H), 2.71 (d, $J = 4.2$ Hz, 3H), 2.11 – 2.08 (m, 1H), 1.38 (s, 9H), 0.89 (d, $J = 6.7$ Hz, 3H), 0.78 (d, $J = 5.6$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.4, 171.2, 156.4, 136.7, 129.3, 128.8, 127.1, 80.6, 60.7, 54.0, 38.0, 30.3, 28.3, 26.3, 19.4, 17.6; HRMS (ESI-TOF) (m/z) [MNa] $^+$ calcd for $\text{C}_{20}\text{H}_{31}\text{N}_3\text{O}_4\text{Na}$ 400.2207, found 400.2229.



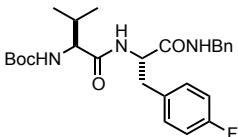
Boc-(D)Val-Phe-NHBn. Yield: 78%; ^1H NMR (400 MHz, CDCl_3) δ 7.28 – 7.14 (m, 10H), 7.09 (d, $J = 7.0$ Hz, 2H), 6.73 (d, $J = 8.4$ Hz, 1H), 5.26 (d, $J = 7.7$ Hz, 1H), 4.83 (dd, $J = 15.1, 7.2$ Hz, 1H), 4.38 (dd, $J = 15.0, 5.8$ Hz, 1H), 4.28 (dd, $J = 14.9, 5.7$ Hz, 1H), 3.72 (t, $J = 7.3$ Hz, 1H), 3.12 (d, $J = 6.9$ Hz, 2H), 1.88 (dt, $J = 13.3, 6.7$ Hz, 1H), 1.35 (s, 9H), 0.76 (dd, $J = 13.4, 6.7$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.1, 170.7, 156.2, 138.1, 136.7, 129.4, 128.8, 128.6, 127.6, 127.3, 127.0, 80.1, 60.9, 54.4, 43.4, 38.0, 30.6, 28.4, 19.2, 18.2; HRMS (ESI-TOF) (m/z) [MNa] $^+$ calcd for $\text{C}_{26}\text{H}_{35}\text{N}_3\text{O}_4$ 476.2520, found 476.2540.



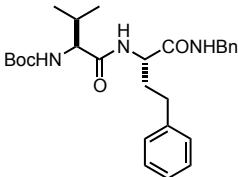
Boc-Val-(D)Phe-NHBn. Yield: 73%; ^1H NMR (400 MHz, CDCl_3) δ 7.31 (dd, $J = 12.9, 7.2$ Hz, 1H), 7.26 – 7.14 (m, 9H), 7.06 (t, $J = 9.7$ Hz, 1H), 6.97 (d, $J = 8.6$ Hz, 1H), 5.41 (d, $J = 7.9$ Hz, 1H), 4.86 (dd, $J = 15.3, 7.4$ Hz, 1H), 4.31 (t, $J = 5.7$ Hz, 2H), 3.78 (t, $J = 7.5$ Hz, 1H), 3.09 (dt, $J = 13.7, 6.9$ Hz, 2H), 1.87 (dq, $J = 13.4, 6.6$ Hz, 1H), 1.35 (s, 9H), 0.78 – 0.68 (m, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.2, 170.8, 156.2, 138.0, 136.7, 129.4, 128.7, 128.5, 127.5, 127.2, 127.0, 80.0, 60.7, 54.4, 43.3, 38.0, 30.7, 28.4, 19.2, 18.1; HRMS (ESI-TOF) (m/z) [MNa] $^+$ calcd for $\text{C}_{26}\text{H}_{35}\text{N}_3\text{O}_4$ 476.2520, found 476.2541.



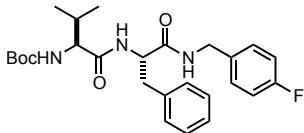
Boc-Val-Phg-NHBn. Yield: 84%; ^1H NMR (400 MHz, CDCl_3) δ 7.43 (d, $J = 7.4$ Hz, 0.4H), 7.30 (d, $J = 7.4$ Hz, 0.6H), 7.08 – 6.81 (m, 9H), 6.72 (d, $J = 4.8$ Hz, 2H), 5.42 (d, $J = 7.1$ Hz, 1H), 5.02 (d, $J = 8.4$ Hz, 0.5H), 4.89 (d, $J = 8.3$ Hz, 0.5H), 4.07 – 3.87 (m, 2H), 3.80 – 3.65 (m, 1H), 1.73 – 1.52 (m, 1H), 1.05 (d, $J = 10.0$ Hz, 9H), 0.49 (ddd, $J = 32.2, 19.4, 6.6$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.7, 170.0, 156.0, 137.8, 128.9, 128.9, 128.6, 128.2, 127.6, 127.5, 127.4, 127.2, 79.9, 59.9, 57.0, 43.6, 31.2, 28.4, 19.4, 17.8; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{26}\text{H}_{36}\text{N}_3\text{O}_4$ 440.2544, found 440.2517



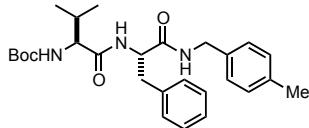
Boc-Val-(4-F)Phe-NHBn. Yield: 82%; ^1H NMR (400 MHz, CDCl_3) δ 7.27 – 7.13 (m, 5H), 7.11 – 6.98 (m, 4H), 6.87 (dd, $J = 10.9, 6.4$ Hz, 2H), 5.23 (d, $J = 7.6$ Hz, 1H), 4.80 (dd, $J = 15.1, 7.3$ Hz, 1H), 4.33 (dd, $J = 14.7, 6.0$ Hz, 1H), 4.19 (dd, $J = 14.8, 5.1$ Hz, 1H), 3.96 (t, $J = 6.9$ Hz, 1H), 3.13 – 2.92 (m, 2H), 2.00 (dd, $J = 13.5, 6.8$ Hz, 1H), 1.35 (s, 9H), 0.83 (dd, $J = 15.1, 6.8$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.8, 170.6, 163.2, 160.7, 156.1, 137.9, 132.2, 131.0, 128.6, 127.7, 127.4, 115.6, 115.4, 80.2, 60.4, 54.2, 43.4, 37.6, 30.7, 28.4, 19.3, 17.9; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{26}\text{H}_{35}\text{N}_3\text{O}_4\text{F}$ 472.2606, found 472.2583.



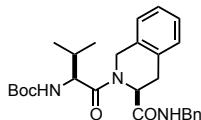
Boc-Val-homoPhe-NHBn. Yield: 62% (15% RSM); ^1H NMR (400 MHz, CDCl_3) δ 7.39 – 6.94 (m, 12H), 5.15 (d, $J = 7.6$ Hz, 1H), 4.55 (d, $J = 6.3$ Hz, 1H), 4.36 (dd, $J = 36.7, 14.9$ Hz, 2H), 3.95 – 3.92 (m, 1H), 2.61 (d, $J = 7.1$ Hz, 2H), 2.16 – 2.14 (m, 1H), 2.02 (dd, $J = 7.8, 5.6$ Hz, 2H), 1.39 (s, 9H), 0.85 (t, $J = 8.4$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.0, 171.2, 156.1, 141.0, 138.1, 128.7, 128.6, 128.5, 127.8, 127.5, 126.2, 80.2, 60.3, 53.0, 43.6, 34.0, 31.9, 30.8, 28.4, 19.4, 18.0; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{27}\text{H}_{38}\text{N}_3\text{O}_4$ 468.2857, found 468.2852.



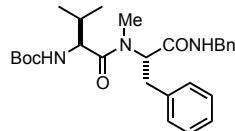
Boc-Val-Phe-NH(4-F)Bn. Yield: 66% (20% RSM); ^1H NMR (400 MHz, CDCl_3) δ 7.29 – 7.09 (m, 7H), 7.03 – 6.91 (m, 2H), 6.90 – 6.81 (m, 2H), 5.21 (d, $J = 7.3$ Hz, 1H), 4.83 (dd, $J = 15.2, 7.3$ Hz, 1H), 4.28 (dd, $J = 14.9, 6.0$ Hz, 1H), 4.14 (dd, $J = 14.9, 5.0$ Hz, 1H), 3.96 (t, $J = 6.6$ Hz, 1H), 3.12 (dd, $J = 13.6, 6.4$ Hz, 1H), 3.05 (dd, $J = 13.4, 7.7$ Hz, 1H), 2.05 – 1.95 (m, 1H), 1.33 (s, 9H), 0.84 (d, $J = 6.8$ Hz, 3H), 0.80 (d, $J = 6.7$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.7, 170.8, 163.2, 160.8, 156.2, 136.5, 133.8, 129.4, 128.7, 127.0, 115.4, 80.2, 60.4, 54.2, 42.6, 38.3, 30.7, 28.3, 19.3, 17.8; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{26}\text{H}_{35}\text{N}_3\text{O}_4$ 472.2606, found 472.2605.



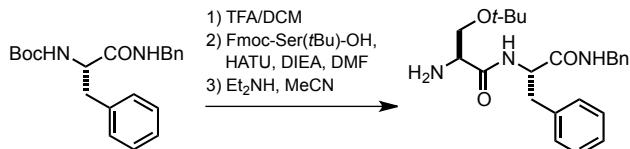
Boc-Val-Phe-NHMe. Yield: 61% (18% RSM); ^1H NMR (400 MHz, CDCl_3) δ 7.28 – 6.86 (m, 11H), 5.08 (d, J = 7.3 Hz, 1H), 4.82 – 4.73 (m, 1H), 4.28 (dd, J = 14.6, 5.7 Hz, 1H), 4.23 – 4.13 (m, 1H), 3.94 – 3.92 (m, 1H), 3.12 (dd, J = 13.4, 6.1 Hz, 1H), 3.04 (dd, J = 13.4, 7.6 Hz, 1H), 2.28 (s, 3H), 2.06 – 1.96 (m, 1H), 1.35 (s, 9H), 0.86 (d, J = 6.8 Hz, 3H), 0.79 (d, J = 6.7 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.6, 170.5, 156.1, 136.9, 136.6, 134.8, 129.4, 129.2, 128.7, 127.8, 127.0, 80.2, 60.4, 54.2, 43.3, 38.4, 30.6, 28.3, 21.2, 19.4, 17.7; HRMS (ESI-TOF) (m/z) $[\text{MH}]^+$ calcd for $\text{C}_{27}\text{H}_{38}\text{N}_3\text{O}_4$ 468.2857, found 468.2861.



Bo-Val-Tic-NHBn. Yield: 53% (34% RSM); ^1H NMR (400 MHz, CDCl_3) δ 7.73 (s, 1H), 7.26 – 7.24 (m, 6H), 7.09 – 7.06 (m, 1H), 6.99 – 6.92 (m, 1H), 6.32 (s, 1H), 5.22 (d, J = 9.1 Hz, 0.5H), 5.07 (dd, J = 18.4, 12.1 Hz, 1.5H), 4.95 (d, J = 5.1 Hz, 0.5H), 4.84 (d, J = 14.9 Hz, 0.5H), 4.56 – 4.52 (m, 1.5H), 4.40 – 4.32 (m, 1.5H), 4.26 (t, J = 7.6 Hz, 0.5H), 4.18 – 4.10 (m, 0.5H), 3.66 – 3.58 (m, 0.5H), 3.38 (dd, J = 15.7, 6.0 Hz, 0.5H), 3.03 (td, J = 15.3, 6.1 Hz, 1H), 1.98 – 1.96 (m, 1H), 1.41 (s, 4.5H), 1.36 (s, 4.5H), 1.04 – 0.94 (m, 4.5H), 0.83 (d, J = 6.8 Hz, 1.5H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.6, 168.9, 156.7, 138.0, 132.7, 132.6, 128.8, 128.5, 128.1, 127.6, 127.2, 127.0, 126.5, 80.6, 56.4, 54.0, 43.8, 43.7, 31.7, 30.5, 28.5, 19.9, 17.1; HRMS (ESI-TOF) (m/z) $[\text{MH}]^+$ calcd for $\text{C}_{27}\text{H}_{36}\text{N}_3\text{O}_4$ 466.2700, found 466.2698.

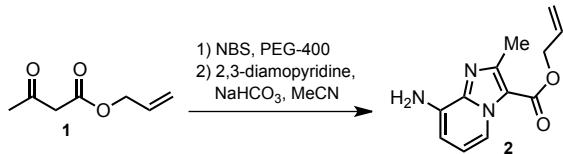


Boc-Val-(N-Me)Phe-NHBn. Yield: 39% (37% RSM); ^1H NMR (400 MHz, CDCl_3) δ 8.05 (t, J = 5.8 Hz, 0.5H), 7.39 – 7.22 (m, 9H), 7.15 (dd, J = 7.4, 1.7 Hz, 1H), 6.52 (t, J = 5.6 Hz, 0.5H), 5.35 (t, J = 7.9 Hz, 0.6H), 5.16 (dd, J = 22.8, 9.1 Hz, 1H), 4.86 (t, J = 7.2 Hz, 0.4H), 4.48 – 4.32 (m, 2.6H), 4.19 (dd, J = 8.8, 6.7 Hz, 0.4H), 3.60 – 3.52 (m, 0.5H), 3.44 (dd, J = 14.0, 8.3 Hz, 0.5H), 3.13 (s, 1.5H), 3.07 – 2.92 (m, 2.5H), 1.87 (dq, J = 13.3, 6.7 Hz, 0.5H), 1.48 (s, 4.4H), 1.39 (d, J = 7.7 Hz, 0.6H), 1.33 (d, J = 5.3 Hz, 4H), 1.25 – 1.15 (m, 0.4H), 1.03 – 1.01 (m, 0.2H), 0.96 (d, J = 6.7 Hz, 0.2H), 0.92 – 0.83 (m, 4.2H), 0.71 (d, J = 6.7 Hz, 1.2H), 0.47 (d, J = 6.7 Hz, 0.2H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.9, 169.4, 157.1, 138.0, 137.7, 129.4, 128.7, 128.6, 127.6, 127.5, 126.9, 80.7, 62.6, 54.9, 43.4, 34.1, 31.2, 29.8, 28.4, 19.6, 17.7; HRMS (ESI-TOF) (m/z) $[\text{MH}]^+$ calcd for $\text{C}_{27}\text{H}_{38}\text{N}_3\text{O}_4$ 468.2857, found 490.2835.

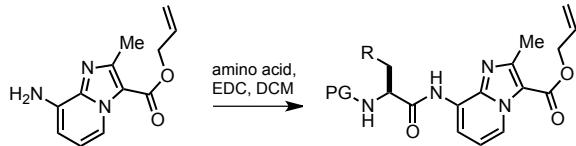


2-Amino-N-(1-benzylcarbamoly-(S)-2-phenyl-ethyl)-(S)-3-tbutoxyl-propionamide. A solution of Boc-Phe-NHBn (86 mg, 0.243 mmol) in 4 mL of 25% TFA/ DCM was stirred for 2 h at RT. The solvent was concentrated under reduced pressure. The crude material was dried under reduced pressure. The amine-TFA salt intermediate was dissolved in 5 mL DMF and treated with Fmoc-Ser(*t*-Bu)-OH (186 mg, 0.49 mmol), HATU (184 mg, 0.49 mmol), and DIEA (188 mg, 1.46 mmol). After stirring at RT for 12 h, the reaction was

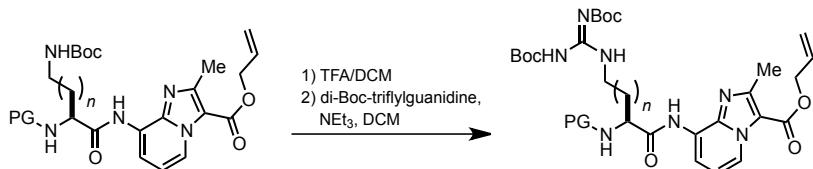
concentrated under reduced pressure. The residue was diluted with EtOAc and washed with 1M aq. HCl, sat. aq. NaHCO₃, and brine. The organic layer was evaporated, and the crude material was stirred for 2 h in 10 mL DEA/MeCN (1:1) for Fmoc deprotection. The solvent was removed under reduced pressure and the residue was purified by silica gel flash chromatography (1% to 5% MeOH/CHCl₃, gradient) to give the product as colorless oil. (77 mg, 0.194 mmol, 90% for 3 steps): ¹H NMR (400 MHz, CDCl₃) δ 7.82 (d, *J* = 8.3 Hz, 1H), 7.32 – 7.17 (m, 8H), 7.09 (d, *J* = 7.2 Hz, 1H), 6.43 (t, *J* = 7.7 Hz, 1H), 4.66 (q, *J* = 7.5 Hz, 1H), 4.36 (ddd, *J* = 39.5, 14.8, 5.7 Hz, 2H), 3.44 – 3.35 (m, 2H), 3.31 – 3.24 (m, 1H), 3.11 (d, *J* = 7.1 Hz, 2H), 1.65 (s, 2H), 1.10 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 173.6, 170.7, 137.7, 136.8, 129.4, 128.6, 128.5, 127.6, 127.4, 126.9, 73.5, 63.7, 55.2, 54.4, 43.4, 38.0, 27.4; HRMS (ESI-TOF) *m/z* [M + H]⁺ calcd for C₂₃H₃₂N₃O₃ 398.2438, found 398.2438.



H-IP(Me)-OAllyl. A mixture of allyl acetoacetate (71.0 mg, 0.50 mmol) and solid *N*-bromosuccinimide (97.9 mg, 0.55 mmol) in PEG-400 (2 mL) was stirred at RT until TLC indicated consumption of the starting material (30 – 60 min). The reaction mixture was diluted with water and extracted with Et₂O. The combined organic layers were washed with water and brine, dried over Na₂SO₄, then filtered and concentrated. The crude residue was then dissolved in 2.5 mL MeCN and treated with 2,3-diaminopyridine (54.6 mg, 0.50 mmol) and NaHCO₃ (46.2 mg, 0.55 mmol). The reaction mixture was heated to 90 °C in a sealed tube and stirred for 15 h. The solution was then cooled to RT, diluted with EtOAc, filtered through a pad of celite, and concentrated. Purification by flash chromatography over silica gel (50 % EtOAc/Hexanes) afforded 2 as a tan solid (67% yield for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 8.71 – 8.69 (m, 1H), 6.77 (t, *J* = 7.2 Hz, 1H), 6.54 (ddd, *J* = 3.8, 2.2, 0.7 Hz, 1H), 6.14 – 5.96 (m, 1H), 5.44 – 5.38 (m, 1H), 5.31 – 5.27 (m, 1H), 4.86 (dt, *J* = 5.6, 1.4 Hz, 2H), 4.56 (bs, 2H), 2.70 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 161.2, 151.1, 140.2, 135.1, 132.5, 118.3, 118.3, 114.8, 113.5, 106.6, 64.9, 16.8; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₁₂H₁₄N₃O₂ 232.1081, found 232.1111.

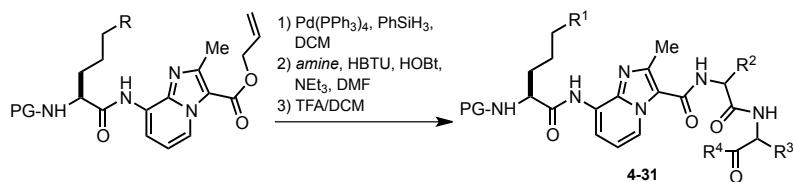


General procedure for N-terminal amidation of the IP scaffold. A solution of H-IP(Me)-OAllyl (0.433 mmol) in 5 mL DCM was treated with the desired N-protected amino acid (0.866 mmol) and EDC·HCl (0.866 mmol). After stirring at RT for 15 h, the reaction was concentrated under reduced pressure and purified by flash chromatography over silica gel (75 % EtOAc/Hexanes) afforded the desired products.



General procedure for guanidinylation of amine side chains. Guanidine-containing analogs were prepared by treatment of the intermediate Boc-protected tripeptide mimics with 20% TFA/DCM for 4 h followed by concentration under reduced pressure. The residue was diluted with EtOAc, concentrated under reduced pressure, and dried under vacuum. The crude residue was then dissolved in 20 mL DCM and treated with NEt₃ (1.25 mmol) and N,N'-di-Boc triflylguanidine (0.374 mmol). After stirring at RT for 18 h, the reaction was

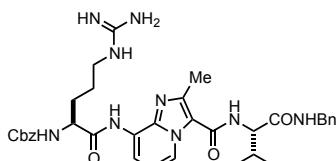
concentrated under reduced pressure and purified by flash chromatography over silica gel (50 % EtOAc/Hexanes) to afford the di-Boc guanidine derivatives.



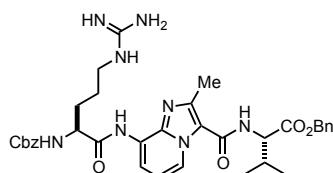
General procedure for synthesis of inhibitors 4-31. A mixture of the allyl ester subunit (1.00 mmol), $\text{Pd}(\text{PPh}_3)_4$ (0.050 mmol), and PhSiH (3.00 mmol) in DCM (50 mL) was stirred at RT until TLC indicated consumption of the starting material (30 – 60 min). The reaction mixture was concentrated under reduced pressure and purified by flash chromatography over silica gel (5 % MeOH/CHCl₃) afforded the desired carboxylic acids.

A solution of above carboxylic acid (0.098 mmol) in DMF (4 mL) was treated with HBTU (0.117 mmol), HOEt (0.019 mmol), NEt₃ (0.392 mmol), and the appropriate C-terminal dipeptide amide (or amino amide) building block (0.117 mmol). After stirring at RT for 20 h, the reaction was concentrated under reduced pressure. The residue was diluted with EtOAc and washed with sat. aq. NH₄Cl. The organic layer was dried over Na₂SO₄ and evaporated. Purification by flash column chromatography over silica gel (1% MeOH/CHCl₃) afforded the desired coupled products.

A solution of above product (0.029 mmol) was treated with 20% TFA/DCM (2.4 mL). After stirring RT 12 h, the reaction was concentrated under reduced pressure. The residue was purified by flash column chromatography over silica gel (85/16/2 CHCl₃/MeOH/NH₄OH) afforded the desired products.

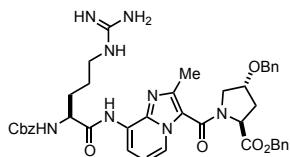


Cbz-Arg-IP(Me)-Val-NHBn (4). Yield: 64%; mp 161 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.00 (s, 1H), 8.70 (t, *J* = 5.8 Hz, 1H), 8.65 (d, *J* = 6.9 Hz, 1H), 8.09 (d, *J* = 7.6 Hz, 1H), 7.93 – 7.82 (m, 2H), 7.64 (t, *J* = 4.0 Hz, 1H), 7.42 – 7.20 (m, 11H), 7.17 (s, 1H), 7.04 (s, 1H), 7.01 (t, *J* = 7.3 Hz, 1H), 5.07 (q, *J* = 12.5 Hz, 2H), 4.52 – 4.42 (m, 1H), 4.41 (t, *J* = 7.7 Hz, 1H), 4.35 (d, *J* = 5.8 Hz, 2H), 3.17 – 3.07 (m, 2H), 2.64 (s, 3H), 2.21 – 2.09 (m, 1H), 1.89 – 1.73 (m, 1H), 1.70 – 1.50 (m, 3H), 1.02 – 0.92 (m, 6H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.5, 171.4, 161.1, 158.9, 157.1, 156.6, 144.5, 139.8, 139.0, 137.3, 128.8, 128.7, 128.3, 128.1, 127.7, 127.2, 126.6, 122.2, 117.7, 113.6, 66.1, 58.9, 55.4, 42.5, 40.8, 30.8, 29.1, 25.7, 19.9, 19.0, 15.9; HRMS (ESI-TOF) *m/z* [MH]⁺ calcd for C₃₅H₄₄N₉O₅ 670.3460, found 670.3454.

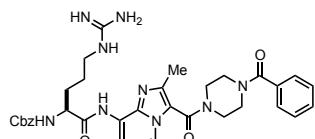


Cbz-Arg-IP(Me)-Val-OBn (5). Yield: 13%; ¹H NMR (400 MHz, CD₃OD) δ 8.59 (d, *J* = 6.9 Hz, 1H), 8.14 (d, *J* = 7.5 Hz, 1H), 8.10 (d, *J* = 7.8 Hz, 1H), 7.88 (d, *J* = 8.4 Hz, 1H), 7.74 (d, *J* = 8.3 Hz, 1H), 7.58 – 7.44 (m, 2H), 7.42 – 7.25 (m, 8H), 6.96 (t, *J* = 7.3 Hz, 1H), 5.23 (dd, *J* = 47.9, 12.1 Hz, 2H), 5.13 (s, 2H), 4.58 (t, *J* = 5.5 Hz, 1H), 4.44 – 4.38 (m, 1H), 3.23 (t, *J* = 7.1 Hz, 2H), 2.63 (s, 3H), 2.31 (dq, *J* = 13.6, 6.8 Hz, 1H), 1.87 – 1.62 (m, 4H), 1.05 (d, *J* = 1.9 Hz, 3H), 1.03 (d, *J* = 1.9 Hz, 3H); ¹³C NMR (101 MHz, CD₃OD) δ 171.9, 171.5, 161.8, 157.2, 144.4, 135.7, 128.3, 128.2, 128.1, 127.7, 127.5, 126.9, 125.8, 125.5, 122.5, 117.2, 115.1, 113.3, 110.0,

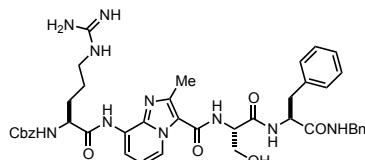
105.0, 66.6, 66.5, 58.5, 55.4, 40.5, 30.3, 28.8, 25.0, 18.2, 17.5, 13.9; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C₃₅H₄₃N₈O₆ 671.3306, found 671.3284.



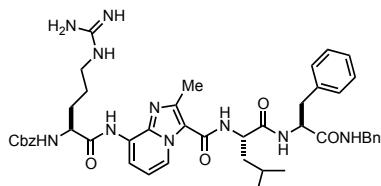
Cbz-Arg-IP(Me)-Hyp(Bn)-OBn (6). Yield: 8%; ¹H NMR (400 MHz, CD₃OD) δ 8.14 (d, J = 6.9 Hz, 1H), 7.42 – 7.09 (m, 16H), 6.99 (t, J = 7.3 Hz, 1H), 5.28 – 5.15 (m, 2H), 5.14 (s, 2H), 4.56 (d, J = 11.5 Hz, 1H), 4.42 (s, 1H), 4.27 (s, 1H), 3.73 (d, J = 11.8 Hz, 1H), 3.52 (s, 1H), 3.23 (t, J = 6.6 Hz, 2H,), 2.72 – 2.60 (m, 1H), 2.48 (d, J = 9.3 Hz, 1H), 2.40 (s, 3H), 2.22 – 2.11 (m, 1H), 2.08 – 1.95 (m, 1H), 1.87 – 1.56 (m, 4H); HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C₄₂H₄₆N₉O₇ 775.3662, found 775.3558.



Cbz-Arg-IP(Me)-Hyp(Bn)-OBn (7). Yield: 22%; ¹H NMR (400 MHz, CD₃OD) δ 8.26 (d, J = 6.9 Hz, 1H), 8.12 (d, J = 7.5 Hz, 1H), 7.53 – 7.43 (m, 5H), 7.40 – 7.17 (m, 5H), 6.93 (t, J = 7.3 Hz, 1H), , 5.12 (s, 2H), 4.41 (dd, J = 8.5, 5.0 Hz, 1H), 3.96 – 3.47 (m, 8H), 3.22 (t, J = 6.9 Hz, 2H), 2.49 (s, 3H), 1.87 – 1.64 (m, 4H); ¹³C NMR (101 MHz, CD₃OD) δ 171.9, 171.3, 162.0, 157.3, 157.2, 145.8, 142.6, 136.6, 134.9, 130.0, 128.4, 128.0, 127.6, 127.5, 126.8, 125.6, 122.1, 116.3, 115.0, 113.3, 66.5, 55.4, 40.5, 28.8, 25.0, 13.2; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C₃₄H₄₀N₉O₅ 654.3147, found 654.3140.

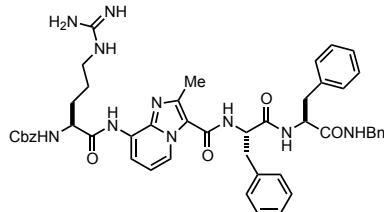


Cbz-Arg-IP(Me)-Ser-Phe-NHBn (8). Yield: 65%; mp 129 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.00 (s, 1H), 8.68 (d, J = 6.9 Hz, 1H), 8.53 (t, J = 5.9 Hz, 1H), 8.35 (d, J = 8.3 Hz, 1H), 8.09 (d, J = 7.7 Hz, 1H), 7.89 (d, J = 7.6 Hz, 1H), 7.76 (d, J = 7.7 Hz, 1H), 7.61 (t, J = 4.0 Hz, 1H), 7.47 – 7.12 (m, 17H), 7.03 (s, 1H), 6.96 (t, J = 7.3 Hz, 1H), 5.20 (s, 1H), 5.07 (q, J = 12.5 Hz, 2H), 4.67 – 4.52 (m, 2H), 4.51 – 4.41 (m, 1H), 4.28 (d, J = 6.0 Hz, 2H), 3.78 – 3.65 (m, 2H), 3.18 – 3.01 (m, 3H), 2.89 (dd, J = 13.4, 9.1 Hz, 1H), 2.61 (s, 3H), 1.87 – 1.75 (m, 1H), 1.71 – 1.49 (m, 3H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.5, 171.1, 170.2, 160.9, 158.8, 158.5, 157.1, 156.6, 144.9, 139.4, 139.0, 138.1, 137.3, 129.6, 128.8, 128.7, 128.5, 128.3, 128.1, 127.5, 127.2, 126.7, 126.6, 117.4, 113.6, 66.1, 62.1, 55.7, 55.4, 54.7, 42.5, 40.8, 37.9, 29.1, 25.7, 15.9; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C₄₂H₄₉N₁₀O₇ 805.3780, found 805.3802.

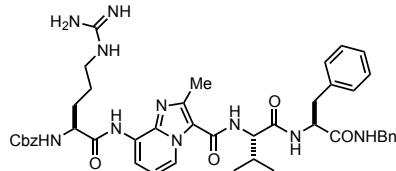


Cbz-Arg-IP(Me)-Leu-Phe-NHBn (9). Yield: 63%; mp 129 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.97 (s, 1H), 8.54 – 8.48 (m, 2H), 8.28 (d, J = 8.1 Hz, 1H), 8.07 (d, J = 7.6 Hz, 1H), 8.00 (d, J = 8.1 Hz, 1H), 7.92 – 7.85 (m, 1H), 7.32 – 7.25 (m, 13H), 7.17 – 7.12 (m, 4H), 7.04 (s, 1H), 6.95 – 6.90 (m, 1H), 5.07 (q, J = 12.5 Hz, 2H), 4.60 (dd, J = 14.3, 8.4 Hz, 1H), 4.54 – 4.39 (m, 2H), 4.27 (d, J = 5.8 Hz, 2H), 3.12 (d, J = 6.5 Hz, 2H), 3.07 –

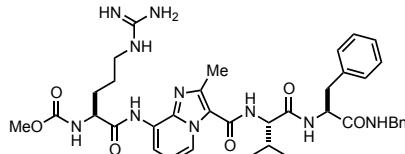
3.01 (m, 1H), 2.95 (s, 2H), 2.89 (dd, $J = 9.2, 4.4$ Hz, 1H), 2.57 (s, 3H), 1.85 – 1.78 (m, 1H), 1.58 – 1.63 (m, 3H), 1.51 – 1.46 (m, 1H), 1.25 – 1.20 (m, 1H), 0.90 (dd, $J = 6.3, 2.3$ Hz, 4.8H), 0.81 (t, $J = 5.5$ Hz, 1.2H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ 171.9, 170.8, 160.6, 158.3, 158.0, 156.7, 156.2, 144.2, 139.1, 138.6, 137.7, 136.9, 131.5, 129.2, 128.4, 128.2, 128.0, 127.7, 127.0, 126.7, 126.3, 118.7, 117.3, 115.7, 113.1, 65.6, 55.0, 54.1, 51.7, 42.0, 37.6, 28.7, 25.3, 24.5, 23.2, 22.8, 22.0, 21.5, 15.4; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{45}\text{H}_{55}\text{N}_{10}\text{O}_6$ 831.4301, found 831.4277.



Cbz-Arg-IP(Me)-Phe-Phe-NHBn (10). Yield: 76%; mp 110 °C; ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 9.96 (s, 1H), 8.73 (d, $J = 8.6$ Hz, 0.5H), 8.62 (t, $J = 5.9$ Hz, 0.5H), 8.54 (t, $J = 6.0$ Hz, 0.5H), 8.48 (d, $J = 8.1$ Hz, 0.5H), 8.39 (t, $J = 7.2$ Hz, 1H), 8.05 (dd, $J = 7.4, 3.9$ Hz, 1H), 7.96 (d, $J = 8.1$ Hz, 1H), 7.88 (d, $J = 7.2$ Hz, 1H), 7.65 (s, 1H), 7.39 – 7.13 (m, 22H), 7.05 (s, 1H), 6.90 (t, $J = 7.3$ Hz, 0.5H), 6.86 (t, $J = 7.3$ Hz, 0.5H), 5.06 (q, $J = 12.5$ Hz, 2H), 4.78 – 4.75 (m, 1H), 4.63 (dd, $J = 14.2, 8.1$ Hz, 1H), 4.47 – 4.40 (m, 1H), 4.38 – 4.33 (m, 1H), 4.29 (d, $J = 6.1$ Hz, 1H), 3.17 – 2.69 (m, 7H), 2.31 (s, 3H), 1.78 – 1.74 (m, 1H), 1.66 – 1.53 (m, 2H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ 172.1, 171.1, 171.0, 170.8, 160.5, 158.4, 158.1, 156.7, 156.2, 144.2, 139.1, 138.5, 137.9, 137.6, 136.9, 129.3, 128.4, 128.3, 128.1, 127.9, 127.7, 127.1, 126.8, 126.4, 126.1, 121.8, 117.1, 113.0, 65.7, 55.0, 54.5, 54.3, 42.1, 40.4, 37.8, 37.0, 28.7, 25.3, 15.1; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{48}\text{H}_{53}\text{N}_{10}\text{O}_6$ 865.4144, found 865.4108.

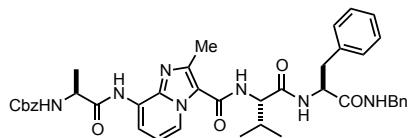


Cbz-Arg-IP(Me)-Val-Phe-NHBn (11). Yield: 54%; mp 159 °C; ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 9.98 (s, 1H), 8.57 (d, $J = 6.8$ Hz, 1H), 8.53 (t, $J = 6.0$ Hz, 1H), 8.34 (d, $J = 8.2$ Hz, 1H), 8.06 (d, $J = 7.6$ Hz, 1H), 7.89 (d, $J = 7.6$ Hz, 1H), 7.77 (d, $J = 8.2$ Hz, 1H), 7.69 (d, $J = 8.4$ Hz, 1H), 7.39 – 7.11 (m, 18H), 6.92 (t, $J = 7.2$ Hz, 1H), 5.01 (dd, $J = 12.0, 19.2$ Hz, 2H), 4.62 (m, 1H), 4.44 (m, 1H), 4.35 (t, $J = 7.4$ Hz, 1H), 4.24 (d, $J = 2.8$ Hz, 2H), 3.10 (m, 2H), 2.99 (dd, $J = 5.0, 13.8$ Hz, 1H), 2.84 (dd, $J = 10.0, 12.0$ Hz, 1H), 2.58 (s, 3H), 2.07 (m, 1H), 1.80 (s, 1H), 1.59 (m, 3H), 0.84 (t, $J = 7.0$ Hz, 5.6H), 0.72 (d, $J = 6.7$ Hz, 0.2H), 0.62 (d, $J = 6.8$ Hz, 0.2H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ 172.2, 170.9, 170.8, 160.6, 158.9, 158.6, 156.9, 156.3, 144.1, 139.1, 138.6, 137.7, 137.0, 129.3, 128.4, 128.2, 128.1, 127.7, 127.1, 126.7, 126.3, 121.9, 118.6, 117.3, 115.6, 113.2, 65.7, 58.4, 55.1, 54.2, 42.1, 37.8, 30.5, 28.8, 25.4, 19.4, 18.4, 15.3; HRMS (ESI-TOF) (m/z) [MH] $^+$ calcd for $\text{C}_{44}\text{H}_{53}\text{N}_{10}\text{O}_6$ 817.4144, found 817.4127.

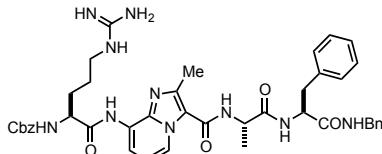


Moc-Arg-IP(Me)-Val-Phe-NHBn (12). Yield: 38%; mp 146 °C; ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 9.98 (s, 1H), 8.59 (dd, $J = 6.9, 1.0$ Hz, 1H), 8.53 (t, $J = 6.0$ Hz, 1H), 8.34 (t, $J = 6.3$ Hz, 1H), 8.07 (dd, $J = 7.6, 0.8$ Hz, 1H), 7.75 – 7.63 (m, 3H), 7.30 – 7.10 (m, 12H), 7.04 (s, 1H), 7.00 – 6.91 (m, 1H), 4.64 (td, $J = 8.6, 6.0$ Hz, 1H), 4.45 (d, $J = 4.1$ Hz, 1H), 4.37 (dd, $J = 8.3, 7.1$ Hz, 1H), 4.26 (d, $J = 5.9$ Hz, 2H), 3.57 (s, 3H), 3.12 (dd, $J = 12.5, 6.5$ Hz, 2H), 3.02 (dd, $J = 13.7, 5.8$ Hz, 1H), 2.92 – 2.83 (m, 1H), 2.59 (s, 3H), 2.08 (dt, $J = 14.9, 6.8$ Hz, 1H), 1.79 – 1.74 (m, 1H), 1.66 – 1.51 (m, 3H), 0.87 (t, $J = 6.3$ Hz, 5.6H), 0.72 (d, $J = 6.7$ Hz, 0.2H), 0.62 (d, $J = 6.9$ Hz, 0.2H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ 172.2, 170.9, 170.7, 160.5, 158.4, 158.1, 156.8, 156.7, 144.1,

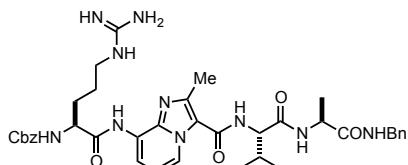
139.1, 138.6, 137.7, 129.2, 128.2, 128.1, 127.0, 126.7, 126.3, 126.2, 121.9, 117.2, 113.1, 58.3, 54.9, 54.1, 51.7, 42.0, 37.8, 30.5, 28.8, 25.3, 19.4, 18.4, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₃₈H₄₉N₁₀O₆ 741.3831, found 741.3876.



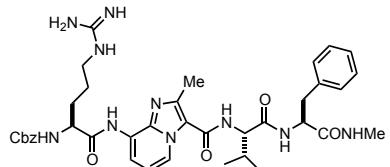
Cbz-Ala-IP(Me)-Val-Phe-NHBn (13). Yield: 29%; mp 160 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.94 (s, 1H), 8.59 (dd, *J* = 6.9, 1.0 Hz, 1H), 8.54 (t, *J* = 5.9 Hz, 1H), 8.35 (d, *J* = 8.3 Hz, 1H), 8.09 (d, *J* = 7.5 Hz, 1H), 7.90 (d, *J* = 7.2 Hz, 1H), 7.74 (d, *J* = 8.3 Hz, 1H), 7.39 – 7.11 (m, 15H), 6.97 (t, *J* = 7.3 Hz, 1H), 5.10 – 5.02 (m, 2H), 4.69 – 4.61 (m, 1H), 4.48 (dd, *J* = 14.3, 7.1 Hz, 1H), 4.37 (dd, *J* = 8.4, 7.1 Hz, 1H), 4.27 (d, *J* = 6.0 Hz, 2H), 3.03 (dd, *J* = 13.6, 5.9 Hz, 1H), 2.93 – 2.84 (m, 1H), 2.59 (s, 3H), 2.08 (dt, *J* = 13.7, 6.8 Hz, 1H), 1.34 (d, *J* = 7.1 Hz, 3H), 0.87 (dd, *J* = 13.0, 6.6 Hz, 5.6H), 0.73 (d, *J* = 6.5 Hz, 0.2H), 0.62 (d, *J* = 6.8 Hz, 0.2H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.8, 170.8, 170.7, 162.3, 160.5, 156.0, 139.1, 137.7, 137.0, 129.2, 128.4, 128.3, 128.2, 128.1, 127.9, 127.7, 127.0, 126.8, 126.7, 126.3, 126.2, 121.9, 117.3, 113.4, 65.6, 58.4, 54.1, 51.0, 42.0, 37.8, 30.5, 19.4, 18.4, 17.8, 15.4; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₁H₄₆N₇O₆ 732.3504, found 732.3483.



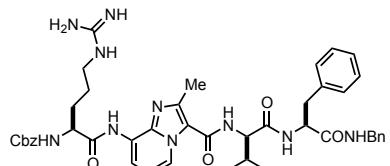
Cbz-Arg-IP(Me)-Ala-Phe-NHBn (14). Yield: 49%; mp 157 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (s, 1H), 8.62 (dd, *J* = 6.9, 1.0 Hz, 1H), 8.52 (t, *J* = 6.0 Hz, 1H), 8.27 (d, *J* = 8.2 Hz, 1H), 8.08 (d, *J* = 7.6 Hz, 1H), 7.98 (d, *J* = 8.3 Hz, 1H), 7.89 (d, *J* = 7.7 Hz, 1H), 7.72 (dt, *J* = 8.3, 1.0 Hz, 1H), 7.67 (t, *J* = 5.4 Hz, 1H), 7.54 (ddd, *J* = 8.3, 5.9, 0.9 Hz, 1H), 7.44 – 7.31 (m, 5H), 7.27 – 7.12 (m, 10H), 7.05 (s, 1H), 6.96 – 6.91 (m, 1H), 5.07 (q, *J* = 12.5 Hz, 2H), 4.60 – 4.42 (m, 3H), 4.36 – 4.25 (m, 2H), 3.15 – 3.08 (m, 2H), 3.05 (dd, *J* = 13.6, 5.6 Hz, 1H), 2.91 (dd, *J* = 13.6, 8.8 Hz, 1H), 2.59 (s, 3H), 1.82 – 1.79 (m, 1H), 1.69 – 1.54 (m, 3H), 1.33 (d, *J* = 7.1 Hz, 2.5H), 1.15 (d, *J* = 7.0 Hz, 0.5H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.1, 170.8, 160.4, 158.5, 158.2, 156.8, 156.2, 144.4, 139.1, 138.6, 137.7, 137.0, 129.3, 128.2, 128.1, 127.7, 127.0, 126.7, 126.3, 124.5, 122.0, 119.2, 117.1, 113.1, 109.7, 65.7, 55.0, 54.3, 48.7, 42.1, 37.6, 28.7, 25.3, 21.1, 18.1, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₂H₄₉N₁₀O₆ 789.3831, found 789.3847.



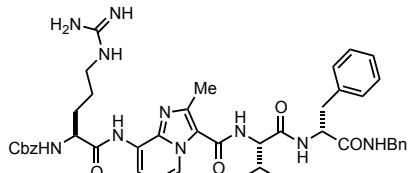
Cbz-Arg-IP(Me)-Val-Ala-NHBn (15). Yield: 42%; mp 127 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.13 (s, 1H), 8.96 – 8.94 (m, 1H), 8.61 (d, *J* = 6.9 Hz, 1H), 8.49 – 8.41 (m, 3H), 8.35 (d, *J* = 7.2 Hz, 1H), 8.07 (d, *J* = 7.5 Hz, 1H), 7.97 (d, *J* = 7.6 Hz, 1H), 7.81 (d, *J* = 8.5 Hz, 2H), 7.38 – 7.19 (m, 10H), 6.95 (t, *J* = 7.3 Hz, 1H), 5.06 (q, *J* = 12.6 Hz, 2H), 4.46 – 4.40 (m, 2H), 4.39 – 4.34 (m, 1H), 4.27 (dd, *J* = 12.4, 7.3 Hz, 2H), 3.08 – 3.05 (m, 2H), 2.64 (s, 3H), 2.15 (dt, *J* = 13.5, 6.7 Hz, 1H), 1.82 – 1.80 (m, 1H), 1.63 – 1.58 (m, 3H), 1.27 (d, *J* = 7.0 Hz, 3H), 0.96 (dd, *J* = 12.3, 6.8 Hz, 6H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.3, 172.1, 170.6, 167.1, 162.2, 160.7, 157.4, 156.3, 144.1, 139.3, 138.6, 137.0, 128.4, 128.3, 127.9, 127.7, 127.0, 126.7, 126.3, 117.2, 113.1, 112.9, 79.2, 65.6, 58.1, 54.9, 48.4, 42.0, 30.6, 25.1, 19.5, 18.4, 18.3, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₃₈H₄₉N₁₀O₆ 741.3831, found 741.3805.



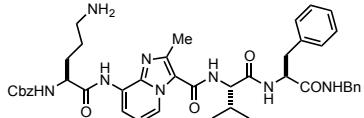
Cbz-Arg-IP(Me)-Val-Phe-NHMe (16). Yield: 36%; mp 136 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (s, 1H), 8.60 – 8.56 (m, 1H), 8.25 (d, *J* = 8.3 Hz, 1H), 8.07 (d, *J* = 7.5 Hz, 1H), 7.93 (d, *J* = 4.7 Hz, 1H), 7.87 (d, *J* = 7.4 Hz, 1H), 7.71 (t, *J* = 5.5 Hz, 1H), 7.67 (d, *J* = 8.6 Hz, 1H), 7.37 – 7.29 (m, 5H), 7.24 – 7.15 (m, 5H), 7.12 – 7.05 (m, 3H), 6.98 (t, *J* = 7.3 Hz, 1H), 5.05 (q, *J* = 12.6 Hz, 2H), 4.52 – 4.40 (m, 2H), 4.31 (t, *J* = 7.4 Hz, 1H), 3.10 (d, *J* = 6.1 Hz, 2H), 2.95 (dd, *J* = 13.7, 5.2 Hz, 1H), 2.79 (dd, *J* = 13.6, 9.3 Hz, 1H), 2.56 (dd, *J* = 13.4, 7.3 Hz, 6H), 2.06 (dt, *J* = 13.7, 6.7 Hz, 1H), 1.82 – 1.75 (m, 1H), 1.68 – 1.47 (m, 3H), 0.84 (dd, *J* = 9.7, 6.8 Hz, 5.6H), 0.66 (d, *J* = 6.8 Hz, 0.2H), 0.60 (d, *J* = 6.8 Hz, 0.2H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.5, 171.7, 171.0, 161.0, 158.8, 158.5, 157.2, 156.6, 144.5, 139.0, 138.2, 137.3, 129.6, 128.8, 128.4, 128.1, 126.6, 122.3, 119.1, 117.6, 116.1, 113.6, 66.1, 58.8, 55.4, 54.4, 38.2, 30.8, 29.1, 25.9, 25.7, 19.8, 18.8, 15.9; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₃₈H₄₉N₁₀O₆ 741.3831, found 741.3820.



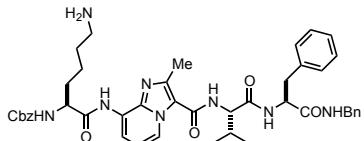
Cbz-Arg-IP(Me)-(D)Val-Phe-NHBn (17). Yield: 54%; mp 160 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.99 (s, 1H), 8.66 (d, *J* = 8.5 Hz, 1H), 8.59 (t, *J* = 6.0 Hz, 1H), 8.44 (dd, *J* = 6.9, 0.9 Hz, 1H), 8.06 (d, *J* = 7.5 Hz, 1H), 7.89 (t, *J* = 7.9 Hz, 2H), 7.76 (t, *J* = 5.5 Hz, 1H), 7.38 – 7.11 (m, 18H), 6.83 (t, *J* = 7.3 Hz, 1H), 5.06 (dd, *J* = 12.5 Hz, 2H), 4.67 – 4.38 (m, 1H), 4.48 – 4.42 (m, 1H), 4.37 (d, *J* = 6.0 Hz, 2H), 4.23 (t, *J* = 7.6 Hz, 1H), 3.20 (dd, *J* = 13.6, 3.9 Hz, 1H), 3.12 (dd, *J* = 12.4, 6.4 Hz, 2H), 2.80 (dd, *J* = 13.5 Hz, 1H), 2.60 (s, 3H), 1.98 – 1.88 (m, 1H), 1.84 – 1.76 (m, 1H), 1.68 – 1.53 (m, 3H), 0.74 (d, *J* = 6.2 Hz, 3H), 0.62 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.2, 171.4, 171.1, 161.1, 158.8, 158.5, 156.8, 156.3, 144.6, 139.1, 138.6, 138.1, 137.0, 129.2, 128.4, 128.3, 128.1, 127.7, 126.8, 126.7, 126.3, 121.6, 118.6, 117.1, 115.7, 113.1, 65.7, 59.3, 55.0, 54.7, 42.1, 37.4, 29.9, 28.8, 25.4, 19.2, 18.6, 15.6; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₄H₅₃N₁₀O₆ 817.4144, found 817.4177.



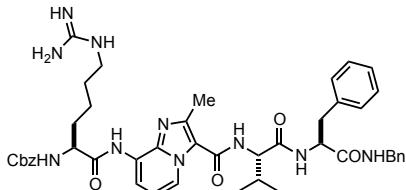
Cbz-Arg-IP(Me)-Val-(D)Phe-NHBn (18). Yield: 54%; mp 105 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (s, 1H), 8.66 (d, *J* = 8.5 Hz, 1H), 8.58 (t, *J* = 6.0 Hz, 1H), 8.45 (d, *J* = 6.9 Hz, 1H), 8.05 (d, *J* = 7.7 Hz, 1H), 7.87 (t, *J* = 7.9 Hz, 2H), 7.70 (t, *J* = 5.3 Hz, 1H), 7.39 – 7.06 (m, 17H), 7.30 – 7.25 (m, 1H), 6.84 (t, *J* = 7.3 Hz, 1H), 5.06 (dd, *J* = 12.5 Hz, 2H), 4.67 – 4.57 (m, 1H), 4.50 – 4.41 (m, 1H), 4.37 (d, *J* = 6.0 Hz, 2H), 4.24 (t, *J* = 7.5 Hz, 1H), 3.25 – 3.06 (m, 3H), 2.85 – 2.74 (m, 1H), 2.58 (s, 3H), 1.99 – 1.86 (m, 1H), 1.84 – 1.72 (m, 1H), 1.70 – 1.52 (m, 3H), 0.73 (d, *J* = 6.7 Hz, 3H), 0.62 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.1, 171.4, 171.1, 161.0, 158.6, 158.3, 1569.8, 156.2, 144.5, 139.1, 138.6, 138.1, 137.0, 129.2, 128.4, 128.3, 128.1, 127.9, 127.7, 126.8, 126.7, 126.3, 121.7, 117.0, 113.1, 65.8, 59.2, 55.0, 54.6, 42.1, 40.4, 37.4, 30.0, 28.7, 25.3, 19.2, 18.6, 15.6; HRMS (ESI-TOF) (*m/z*) [MNa]⁺ calcd for C₄₄H₅₂N₁₀O₆Na 839.3964, found 839.3969.



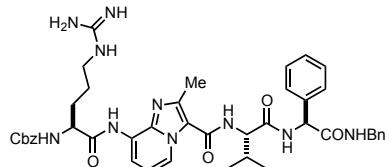
Cbz-Orn-IP(Me)-Val-Phe-NHBn (19). Yield: 42%; mp 147 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 8.56 (dd, *J* = 14.7, 6.4 Hz, 2H), 8.36 (d, *J* = 8.2 Hz, 1H), 8.08 (d, *J* = 7.7 Hz, 1H), 7.95 – 7.82 (m, 1H), 7.71 (d, *J* = 8.2 Hz, 1H), 7.41 – 7.09 (m, 16H), 6.95 (t, *J* = 7.3 Hz, 1H), 5.06 (q, *J* = 12.7 Hz, 2H), 4.64 (dd, *J* = 14.4, 8.4 Hz, 1H), 4.42 – 4.32 (m, 3H), 4.26 (d, *J* = 6.0 Hz, 2H), 3.02 (dd, *J* = 13.7, 5.8 Hz, 1H), 2.88 (m, 2H), 2.59 (s, 3H), 2.55 (t, *J* = 6.8 Hz, 2H), 2.09 (dd, *J* = 13.6, 6.8 Hz, 1H), 1.82 – 1.75 (m, 1H), 1.69 – 1.56 (m, 1H), 1.51 – 1.43 (m, 2H), 0.87 (t, *J* = 6.9 Hz, 5.4H), 0.73 (d, *J* = 6.6 Hz, 0.3H), 0.61 (d, *J* = 6.9 Hz, 0.3H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.5, 170.9, 170.7, 160.6, 156.2, 144.1, 139.1, 138.6, 137.7, 137.0, 129.2, 128.4, 128.2, 128.1, 127.9, 127.7, 127.3, 127.0, 126.7, 126.3, 121.8, 117.2, 113.2, 112.9, 65.6, 58.4, 55.3, 54.1, 42.0, 41.1, 37.8, 30.9, 29.4, 29.2, 19.4, 18.4, 15.6; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₃H₅₁N₈O₆ 775.3926, found 775.3939.



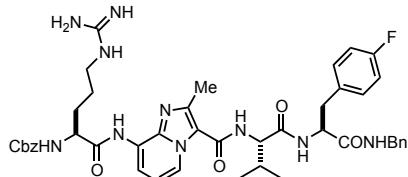
Cbz-Lys-IP(Me)-Val-Phe-NHBn (20). Yield: 36%; mp 165 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 8.66 (d, *J* = 8.9 Hz, 0.4H), 8.62 – 8.48 (m, 1.6H), 8.42 (d, *J* = 6.9 Hz, 0.3H), 8.35 (d, *J* = 8.1 Hz, 0.7H), 8.08 – 8.02 (m, 1H), 7.89 – 7.78 (m, 1H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.40 – 7.11 (m, 15H), 6.94 (t, *J* = 7.3 Hz, 0.8H), 6.82 (t, *J* = 7.3 Hz, 0.2H), 5.06 (q, *J* = 12.5 Hz, 2H), 4.64 (dd, *J* = 14.4, 8.7 Hz, 1H), 4.30 (m, 5H), 3.02 (dd, *J* = 13.6, 5.6 Hz, 1H), 2.88 (dd, *J* = 13.6, 8.9 Hz, 1H), 2.59 (s, 3H), 2.55 – 2.51 (m, 2H), 2.08 (dd, *J* = 13.2, 6.4 Hz, 1H), 1.79 – 1.65 (m, 1H), 1.62 – 1.58 (m, 1H), 1.39 – 1.32 (m, 5H), 1.25 – 1.16 (m, 1H), 0.87 (t, *J* = 6.9 Hz, 4.2H), 0.73 (d, *J* = 6.7 Hz, 0.6H), 0.61 (d, *J* = 6.7 Hz, 0.6H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.5, 170.9, 170.7, 160.6, 156.3, 144.1, 139.1, 138.6, 137.7, 137.0, 129.2, 128.4, 128.3, 128.2, 128.1, 127.9, 127.7, 127.0, 126.8, 126.7, 126.3, 117.2, 113.2, 104.6, 65.6, 58.4, 55.6, 54.1, 42.0, 41.1, 37.8, 32.3, 31.4, 30.5, 23.0, 19.4, 18.4, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₄H₅₄N₈O₆ 789.4083, found 789.4087.



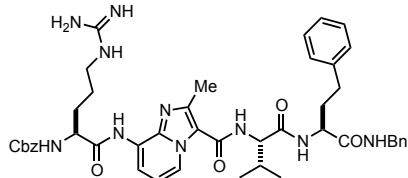
Cbz-Har-IP(Me)-Val-Phe-NHBn (21). Yield: 59%; mp 153 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.00 (s, 1H), 8.58 (d, *J* = 6.9 Hz, 1H), 8.54 (t, *J* = 5.9 Hz, 1H), 8.35 (d, *J* = 8.3 Hz, 1H), 8.08 (d, *J* = 7.5 Hz, 1H), 7.86 (d, *J* = 7.7 Hz, 1H), 7.71 (d, *J* = 8.6 Hz, 1H), 7.65 (t, *J* = 5.4 Hz, 1H), 7.40 – 7.11 (m, 17H), 7.06 (s, 1H), 6.95 (t, *J* = 7.3 Hz, 1H), 5.06 (q, *J* = 12.5 Hz, 2H), 4.64 (dd, *J* = 14.3, 8.5 Hz, 1H), 4.41 – 4.35 (m, 2H), 4.26 (d, *J* = 5.9 Hz, 2H), 3.11 – 3.06 (m, 2H), 3.02 (dd, *J* = 13.7, 5.7 Hz, 1H), 2.88 (dd, *J* = 13.8, 8.8 Hz, 1H), 2.59 (s, 3H), 2.11 – 2.07 (m, 1H), 1.81 – 1.77 (m, 1H), 1.68 – 1.62 (m, 1H), 1.48 – 1.42 (m, 4H), 0.88 (t, *J* = 6.8 Hz, 5.4H), 0.73 (d, *J* = 6.7 Hz, 0.3H), 0.62 (d, *J* = 6.8 Hz, 0.3H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.3, 170.8, 170.7, 160.6, 158.5, 158.2, 156.8, 156.3, 144.1, 139.1, 138.6, 137.7, 137.0, 129.2, 128.4, 128.2, 128.1, 127.7, 127.0, 126.7, 126.3, 118.7, 117.2, 115.7, 113.2, 65.8, 58.3, 55.4, 54.1, 42.0, 40.6, 37.8, 31.2, 30.5, 28.2, 22.9, 19.4, 18.4, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₅H₅₅N₁₀O₆ 831.4301, found 831.4278.



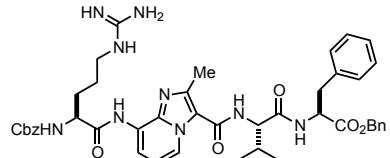
Cbz-Arg-IP(Me)-Val-Phg-NHBn (22). Yield: 49%; mp 154 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.00 (s, 1H), 8.91 (d, *J* = 8.0 Hz, 1H), 8.86 (t, *J* = 6.0 Hz, 1H), 8.63 – 8.56 (m, 1H), 8.08 (d, *J* = 7.6 Hz, 1H), 7.95 – 7.84 (m, 2H), 7.75 – 7.70 (m, 1H), 7.50 – 7.42 (m, 2H), 7.39 – 7.27 (m, 8H), 7.25 – 7.18 (m, 7H), 7.08 (s, 1H), 6.96 – 6.91 (m, 1H), 5.58 (dd, *J* = 15.2, 7.9 Hz, 1H), 5.07 (q, *J* = 12.6 Hz, 2H), 4.54 (t, *J* = 7.8 Hz, 1H), 4.50 – 4.41 (m, 1H), 4.31 – 4.27 (m, 2H), 3.12 (d, *J* = 6.1 Hz, 2H), 2.64 (s, 3H), 2.18 – 2.13 (m, 1H), 1.84 – 1.79 (m, 1H), 1.72 – 1.50 (m, 3H), 0.94 (ddd, *J* = 11.4, 6.6, 4.3 Hz, 6H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.2, 171.0, 170.7, 169.8, 160.9, 158.5, 156.3, 156.2, 144.3, 139.0, 138.8, 138.6, 137.0, 128.4, 128.3, 128.3, 127.9, 127.7, 127.2, 127.0, 126.8, 126.2, 121.8, 117.2, 113.2, 65.7, 58.4, 56.3, 55.0, 42.1, 40.4, 30.5, 28.8, 25.3, 19.4, 18.6, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₃H₅₁N₁₀O₆ 803.3988, found 803.3962.



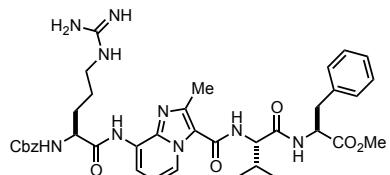
Cbz-Arg-IP(Me)-Val-(4-F)Phe-NHBn (23). Yield 44%; mp 139 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.00 (s, 1H), 8.60 – 8.52 (m, 2H), 8.36 (d, *J* = 8.3 Hz, 1H), 8.08 (d, *J* = 7.6 Hz, 1H), 7.89 (d, *J* = 7.5 Hz, 1H), 7.70 (d, *J* = 6.9 Hz, 1H), 7.39 – 7.18 (m, 12H), 7.14 (d, *J* = 6.9 Hz, 2H), 7.08 – 6.98 (m, 4H), 6.95 (t, *J* = 7.3 Hz, 1H), 5.07 (q, *J* = 12.5 Hz, 2H), 4.66 – 4.58 (m, 1H), 4.49 – 4.41 (m, 1H), 4.36 (t, *J* = 7.7 Hz, 1H), 4.27 (d, *J* = 5.8 Hz, 2H), 3.12 (d, *J* = 6.0 Hz, 2H), 3.01 (dd, *J* = 13.7, 5.7 Hz, 1H), 2.85 (dd, *J* = 13.6, 9.3 Hz, 1H), 2.58 (s, 3H), 2.07 (dt, *J* = 13.6, 6.8 Hz, 1H), 1.85 – 1.79 (m, 1H), 1.68 – 1.52 (m, 3H), 0.88 (t, *J* = 6.9 Hz, 5.4H), 0.73 (d, *J* = 6.7 Hz, 0.3H), 0.63 (d, *J* = 6.7 Hz, 0.3H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.2, 170.7, 162.2, 160.6, 159.8, 158.6, 156.8, 156.2, 144.1, 139.1, 138.6, 136.9, 133.8, 131.1, 128.4, 128.2, 127.7, 127.1, 126.8, 126.2, 118.6, 117.2, 115.7, 114.6, 113.2, 65.7, 58.3, 55.0, 54.2, 42.0, 40.4, 37.0, 30.5, 28.8, 25.3, 19.4, 18.5, 15.4; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₄H₅₂FN₁₀O₆ 835.4050, found 835.4032.



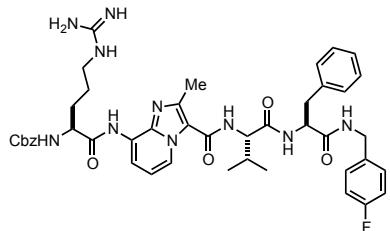
Cbz-Arg-IP(Me)-Val-homoPhe-NHBn (24). Yield: 52%; mp 162 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.00 (s, 1H), 8.60 (d, *J* = 6.9 Hz, 1H), 8.49 (t, *J* = 6.0 Hz, 1H), 8.38 (d, *J* = 7.9 Hz, 1H), 8.06 (d, *J* = 7.6 Hz, 1H), 7.92 (d, *J* = 8.4 Hz, 1H), 7.89 (d, *J* = 7.6 Hz, 1H), 7.70 (t, *J* = 5.6 Hz, 1H), 7.38 – 7.22 (m, 13H), 7.20 – 7.14 (m, 4H), 7.06 (s, 1H), 6.92 (t, *J* = 7.3 Hz, 1H), 5.07 (q, *J* = 12.5 Hz, 2H), 4.49 – 4.40 (m, 2H), 4.35 (dd, *J* = 8.4, 5.0 Hz, 1H), 4.29 (d, *J* = 5.9 Hz, 2H), 3.12 (dd, *J* = 12.3, 6.3 Hz, 2H), 2.69 – 2.53 (m, 5H), 2.18 (dq, *J* = 13.7, 6.7 Hz, 1H), 1.96 – 1.87 (m, 3H), 1.69 – 1.53 (m, 3H), 0.98 (dd, *J* = 6.7, 2.9 Hz, 6H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.2, 171.3, 171.0, 160.8, 158.5, 158.2, 156.8, 156.2, 144.2, 141.3, 139.4, 138.6, 136.9, 128.4, 128.3, 128.3, 127.7, 127.1, 126.8, 126.2, 125.9, 121.7, 117.4, 113.1, 112.9, 65.7, 58.6, 55.0, 52.5, 42.1, 34.0, 31.4, 30.3, 28.8, 25.3, 19.5, 18.6, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₅H₅₄N₁₀O₆ 831.4306, found 831.4325.



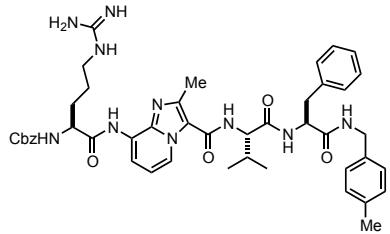
Cbz-Arg-IP(Me)-Val-Phe-OBn (25). Yield: 43%; mp 64 °C; ^1H NMR (400 MHz, CD₃OD) δ 1 H NMR (400 MHz, CD₃OD) δ 8.73 – 8.66 (m, 2H), 8.14 (d, J = 7.5 Hz, 1H), 7.63 (d, J = 8.6 Hz, 1H), 7.40 – 7.10 (m, 13H), 7.08 (d, J = 7.2 Hz, 1H), 7.02 (t, J = 7.2 Hz, 1H), 5.13 (s, 4H), 4.78 (dd, J = 14.2, 8.6 Hz, 1H), 4.45 – 4.38 (m, 2H), 3.27 – 3.15 (m, 3H), 2.99 (dd, J = 13.8, 9.0 Hz, 1H), 2.16 – 1.96 (m, 2H), 1.87 – 1.64 (m, 3H), 1.01 – 0.92 (m, 6H); ^{13}C NMR (101 MHz, CD₃OD) δ 172.1, 172.0, 171.1, 161.5, 160.9, 157.3, 157.2, 143.6, 136.6, 135.5, 134.8, 128.9, 128.1, 128.1, 128.0, 127.9, 127.7, 127.5, 126.4, 125.3, 123.0, 117.1, 116.1, 113.6, 66.7, 66.5, 58.7, 55.4, 53.9, 40.5, 37.0, 30.8, 28.8, 25.0, 18.4, 17.6, 13.9; HRMS (ESI-TOF) m/z [M H]⁺ calcd for C₄₄H₅₂N₉O₇ 818.3984, found 818.3948.



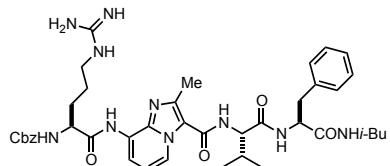
Cbz-Arg-IP(Me)-Val-Phe-OMe (26). Yield: 65%; mp 127 °C; ^1H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (s, 1H), 8.61 (dd, J = 11.8, 4.8 Hz, 2H), 8.09 (d, J = 7.4 Hz, 1H), 7.88 (d, J = 7.7 Hz, 1H), 7.69 – 7.64 (m, 2H), 7.39 – 7.30 (m, 6H), 7.26 – 7.13 (m, 6H), 7.05 (s, 1H), 7.02 – 6.97 (m, 1H), 5.07 (q, J = 12.5 Hz, 2H), 4.54 (ddd, J = 9.1, 7.5, 5.8 Hz, 1H), 4.50 – 4.37 (m, 2H), 3.59 (s, 3H), 3.16 – 3.02 (m, 3H), 3.01 – 2.88 (m, 1H), 2.57 (s, 3H), 2.10 (dq, J = 13.6, 6.8 Hz, 1H), 1.78 (bs, 1H), 1.65 – 1.58 (m, 3H), 0.93 (dd, J = 6.7, 3.5 Hz, 5.6H), 0.77 (d, J = 6.7 Hz, 0.2H), 0.70 (d, J = 6.8 Hz, 0.2H); ^{13}C NMR (101 MHz, (CD₃)₂SO) δ 172.1, 171.8, 171.1, 160.4, 158.1, 156.8, 156.2, 144.1, 138.6, 137.2, 136.9, 129.1, 128.4, 128.2, 127.9, 127.7, 126.5, 126.2, 121.9, 117.2, 113.2, 65.7, 57.8, 55.0, 53.6, 51.8, 40.4, 36.6, 30.6, 28.7, 25.3, 19.3, 18.4, 15.4; HRMS (ESI-TOF) (m/z) [MH]⁺ calcd for C₃₈H₄₈N₉O₇ 742.3677, found 742.3709.



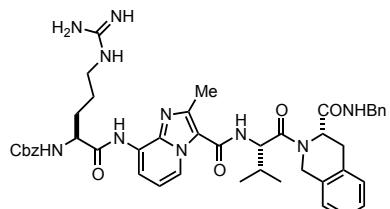
Cbz-Arg-IP(Me)-Val-Phe-NH(4-F)Bn (27). Yield: 64%; mp 142 °C; ^1H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (s, 1H), 8.61 – 8.58 (m, 1H), 8.54 (t, J = 5.9 Hz, 1H), 8.35 (d, J = 8.2 Hz, 1H), 8.08 (d, J = 7.6 Hz, 1H), 7.89 (d, J = 7.6 Hz, 1H), 7.67 (dd, J = 11.9, 7.1 Hz, 2H), 7.39 – 7.28 (m, 5H), 7.27 – 7.13 (m, 9H), 7.10 – 7.03 (m, 3H), 6.96 (t, J = 7.3 Hz, 1H), 5.07 (q, J = 12.5 Hz, 2H), 4.62 (dd, J = 14.4, 8.4 Hz, 1H), 4.50 – 4.42 (m, 1H), 4.41 – 4.34 (m, 1H), 4.24 (d, J = 6.0 Hz, 2H), 3.12 (dd, J = 12.4, 6.4 Hz, 2H), 3.01 (dd, J = 13.7, 5.9, 1H), 2.93 – 2.81 (m, 1H), 2.59 (s, 3H), 2.09 (dq, J = 13.6, 6.7 Hz, 1H), 1.83 – 1.80 (m, 1H), 1.64 – 1.58 (m, 3H), 0.90 – 0.85 (m, 5.6H), 0.73 (d, J = 6.7 Hz, 0.2H), 0.63 (d, J = 6.7 Hz, 0.2H); ^{13}C NMR (101 MHz, (CD₃)₂SO) δ 172.1, 170.8, 162.3, 160.5, 159.9, 158.4, 158.1, 156.7, 156.2, 144.1, 138.6, 137.6, 136.9, 135.3, 129.2, 129.0, 128.9, 128.4, 128.1, 127.7, 126.3, 121.9, 117.2, 115.0, 113.1, 65.7, 58.3, 55.0, 54.1, 41.3, 40.4, 37.7, 30.5, 28.7, 25.3, 19.4, 18.4, 15.5; HRMS (ESI-TOF) (m/z) [MH]⁺ calcd for C₄₄H₅₂FN₁₀O₆ 835.4050, found 835.4027.



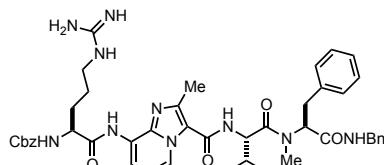
Cbz-Arg-IP(Me)-Val-Phe-NH(4-Me)Bn (28). Yield: 52%; mp 161 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (s, 1H), 8.59 (d, *J* = 6.9 Hz, 1H), 8.47 (t, *J* = 5.9 Hz, 1H), 8.32 (d, *J* = 8.3 Hz, 1H), 8.08 (d, *J* = 7.6 Hz, 1H), 7.89 (d, *J* = 7.6 Hz, 1H), 7.69 (d, *J* = 8.6 Hz, 1H), 7.63 (t, *J* = 5.5 Hz, 1H), 7.41 – 7.28 (m, 5H), 7.29 – 7.12 (m, 7H), 7.05 – 6.98 (m, 5H), 6.96 (t, *J* = 7.3 Hz, 1H), 5.07 (q, *J* = 12.5 Hz, 2H), 4.63 (dd, *J* = 14.3, 8.6 Hz, 1H), 4.45 (d, *J* = 4.4 Hz, 1H), 4.42 – 4.33 (m, 1H), 4.21 (d, *J* = 6.0 Hz, 2H), 3.12 (d, *J* = 6.0 Hz, 2H), 3.01 (dd, *J* = 13.7, 5.6 Hz, 1H), 2.86 (dd, *J* = 13.6, 9.0 Hz, 1H), 2.59 (s, 3H), 2.24 (s, 3H), 2.09 (dd, *J* = 13.5, 6.8 Hz, 1H), 1.82 – 1.79 (m, 1H), 1.68 – 1.49 (m, 3H), 0.88 (t, *J* = 6.8 Hz, 5.6H), 0.72 (d, *J* = 6.6 Hz, 0.2H), 0.62 (d, *J* = 6.7 Hz, 0.2H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 171.1, 169.7, 169.7, 159.6, 157.5, 157.2, 155.7, 155.2, 143.1, 137.6, 136.7, 135.9, 135.0, 134.7, 128.2, 127.7, 127.4, 127.1, 126.7, 126.0, 125.3, 120.9, 116.2, 112.2, 111.9, 64.7, 57.4, 54.0, 53.1, 40.8, 39.4, 36.8, 29.5, 27.7, 24.3, 19.7, 18.4, 17.4, 14.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₅H₅₅N₁₀O₆ 831.4301, found 831.4266.



Cbz-Arg-IP(Me)-Val-Phe-NHiBu (29). Yield: 63%; mp 159 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.99 (s, 1H), 8.61 (d, *J* = 6.9 Hz, 1H), 8.27 (d, *J* = 8.3 Hz, 1H), 8.09 (d, *J* = 7.6 Hz, 1H), 7.97 (t, *J* = 5.9 Hz, 1H), 7.89 (d, *J* = 7.6 Hz, 1H), 7.75 – 7.70 (m, 2H), 7.39 – 7.30 (m, 5H), 7.26 – 7.17 (m, 6H), 7.15 – 7.06 (m, 2H), 6.99 (t, *J* = 7.3 Hz, 1H), 5.07 (q, *J* = 12.5 Hz, 2H), 4.58 (dd, *J* = 14.5, 8.7 Hz, 1H), 4.50 – 4.42 (m, 1H), 4.40 – 4.30 (m, 1H), 3.15 – 3.09 (m, 2H), 2.90 – 2.78 (m, 4H), 2.59 (s, 3H), 2.07 (dt, *J* = 13.6, 6.8 Hz, 1H), 1.83 – 1.78 (m, 1H), 1.62 (td, *J* = 13.3, 6.7 Hz, 4H), 0.87 (t, *J* = 7.1 Hz, 6H), 0.76 (dd, *J* = 6.6, 5.1 Hz, 5.4H), 0.71 (d, *J* = 6.7 Hz, 0.3H), 0.63 (d, *J* = 6.7 Hz, 0.3H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.2, 170.7, 170.6, 160.5, 158.3, 156.8, 156.2, 144.0, 138.6, 137.8, 137.0, 129.2, 128.4, 128.0, 127.7, 126.2, 121.9, 117.3, 113.2, 104.6, 65.7, 58.4, 55.0, 54.1, 46.1, 38.0, 30.5, 28.8, 28.0, 25.3, 20.1, 20.0, 19.4, 18.4, 15.5; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₁H₅₅N₁₀O₆ 783.4301, found 783.4292.

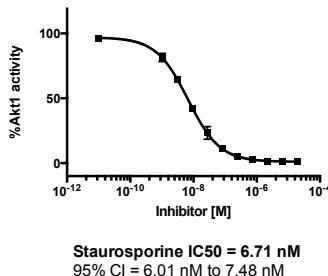


Cbz-Arg-IP(Me)-Val-Tic-NHBn (30). Yield: 68%; mp 43 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (d, *J* = 8.1 Hz, 1H), 8.58 (d, *J* = 7.0 Hz, 1H), 8.46 – 8.43 (m, 1H), 8.11 – 8.00 (m, 2H), 7.88 (d, *J* = 7.1 Hz, 1H), 7.69 (s, 1H), 7.33 (dd, *J* = 19.4, 9.3 Hz, 6H), 7.21 (dd, *J* = 9.7, 5.3 Hz, 7H), 7.13 – 7.05 (m, 4H), 6.95 (dd, *J* = 12.9, 5.8 Hz, 1H), 6.87 (bs, 0.8H), 5.33 (bs, 0.2H), 5.13 – 4.99 (m, 3H), 4.93 (t, *J* = 6.0 Hz, 1H), 4.75 (d, *J* = 15.2 Hz, 0.7H), 4.45 (bs, 1H), 4.35 (d, *J* = 17.5 Hz, 0.3H), 4.27 – 4.15 (m, 1.5H), 3.95 (dd, *J* = 15.3, 5.1 Hz, 0.4H), 3.13 (d, *J* = 5.9 Hz, 3H), 2.66 (s, 1H), 2.56 (s, 2H), 2.30 (dd, *J* = 13.6, 6.8 Hz, 0.6H), 2.24 – 2.16 (m, 0.4H), 2.05 (d, *J* = 52.2 Hz, 0.3H), 1.79 – 1.75 (m, 1H), 1.68 – 1.59 (m, 3H), 1.29 (d, *J* = 46.4 Hz, 0.7H), 1.12 – 0.96 (m, 6H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.1, 171.0, 170.7, 169.2, 160.6, 158.3, 156.2, 156.8, 144.2, 139.4, 139.0, 138.6, 136.9, 134.3, 133.7, 132.5, 128.4, 128.1, 127.9, 127.7, 127.1, 126.7, 126.6, 126.0, 121.8, 117.1, 113.2, 79.2, 65.7, 55.0, 54.6, 53.8, 45.7, 41.9, 31.5, 30.1, 28.7, 25.3, 19.6, 18.3, 15.3; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₅H₅₃N₁₀O₆ 829.4144, found 829.4174.



Cbz-Arg-IP(Me)-Val-(N-Me)Phe-NHBn (31). Yield: 33%; mp 46 °C; ¹H NMR (400 MHz, (CD₃)₂SO) δ 9.98 (s, 1H), 8.53 – 8.47 (m, 1H), 8.42 – 8.36 (m, 1H), 8.08 (d, *J* = 7.6 Hz, 1H), 8.00 (d, *J* = 8.5 Hz, 0.4H), 7.90 (d, *J* = 7.6 Hz, 1H), 7.82 (d, *J* = 8.3 Hz, 0.6H), 7.77 (t, *J* = 5.1 Hz, 1H), 7.38 – 7.29 (m, 6H), 7.25 – 7.20 (m, 5H), 7.18 – 7.14 (m, 3H), 7.12 – 7.09 (m, 2H), 7.07 – 6.91 (m, 3H), 5.33 (dd, *J* = 9.6, 6.2 Hz, 0.5H), 5.10 – 4.98 (m, 2.5H), 4.71 (t, *J* = 8.1 Hz, 0.6H), 4.51 – 4.42 (m, 1.4H), 4.33 (dd, *J* = 15.3, 6.2 Hz, 1H), 4.22 (dd, *J* = 15.1, 5.8 Hz, 1H), 3.91 (dd, *J* = 15.1, 5.3 Hz, 0.6H), 3.40 (dd, *J* = 14.0, 4.1 Hz, 0.4H), 3.23 (dd, *J* = 14.4, 6.0 Hz, 1H), 3.15 – 3.10 (m, 3H), 3.03 – 2.95 (m, 1H), 2.88 – 2.84 (m, 1H), 2.58 – 2.53 (m, 3H), 2.09 – 2.06 (m, 1H), 1.88 – 1.75 (m, 1H), 1.69 – 1.52 (m, 3H), 0.93 (dd, *J* = 13.6, 6.7 Hz, 3.5H), 0.69 (d, *J* = 6.8 Hz, 1H), 0.64 (d, *J* = 6.6 Hz, 0.2H), 0.52 (d, *J* = 6.6 Hz, 1H), 0.46 (d, *J* = 6.8 Hz, 0.2H); ¹³C NMR (101 MHz, (CD₃)₂SO) δ 172.1, 172.1, 171.1, 169.7, 160.4, 158.6, 156.8, 144.2, 139.3, 138.8, 137.8, 137.0, 129.3, 128.8, 128.5, 128.4, 128.2, 128.1, 127.9, 127.8, 127.0, 127.0, 126.7, 126.2, 121.8, 118.6, 117.0, 115.7, 113.1, 112.9, 104.6, 65.4, 55.1, 54.3, 42.1, 34.2, 32.2, 30.1, 28.7, 25.3, 19.5, 19.2, 18.5, 17.7, 15.4; HRMS (ESI-TOF) (*m/z*) [MH]⁺ calcd for C₄₅H₅₅N₁₀O₆ 831.4301, found 831.4290.

In vitro hotspot kinase assay. Inhibition of Akt1-3 kinase activity was performed at Reaction Biology Corporation using the “HotSpot” assay platform. The substrate Crosstide (GRPRTSSFAEG) was prepared in base reaction buffer containing 20 mM Hepes (pH 7.5), 10 mM MgCl₂, 1 mM EGTA, 0.02% Brij35, 0.02 mg/ml BSA, 0.1 mM Na₃VO₄, 2 mM DTT, 1% DMSO to achieve a final substrate concentration of 20 μM. N-terminal His₆-tagged, recombinant human kinase purified from insect cells was added to achieve a final enzyme concentration of 2.5 nM. Compounds were delivered as DMSO stock solutions in varying concentrations and ³³P-ATP (specific activity 0.01 μCi/μl final) was added into the reaction mixture to initiate the reaction. The kinase reaction was incubated for 120 min at rt and the reactions spotted onto P81 ion exchange paper. Dose-response data points are an average of N = 3 experiments and curves were fit using GraphPad Prism software and a variable slope 4-parameter model. A control inhibitor, staurosporine, was determined to have an IC₅₀ of 6.71 nM:



Inhibition of seven different AGC family kinases by compound **11** was performed at Reaction Biology Corporation using the same HotSpot assay platform. Inhibition of these kinases by 10 μM **11** is provided below:

Kinase	%inhibition at 10 μM
PDK1	0
PKA	54
PKCa	7
LATS1	0
MSK1	65
ROCK1	11
RSK1	49

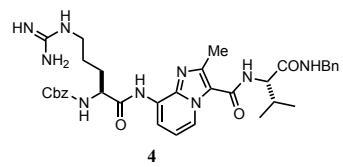
Western blot kinase assay. The effect of **11** on Akt1 kinase activity was determined by using immunoprecipitated Akt protein from HEK293 cells transfected with Myc-tagged wild-type Akt1. The non-radioactive in vitro Akt kinase assay was performed in the presence of 200 μ M ATP with GSK3 β fusion protein as a substrate, according to the manufacturer's instructions (Cell Signaling Assay Kit #9840), except that Myc antibody was used in place of immobilized Akt antibody to immunoprecipitate Akt. The phosphorylation of GSK3 β (S9) is detected by western blot analysis with anti-phospho-GSK3 β (S9) antibody (#5558, Cell Signaling) and GSK3 β antibody (#5558, and #9315, Cell Signaling).

Peptide digestion studies. The stability of **11** and control peptides (Cbz-RTSVF-NHBn and Crosstide) to degradation by chymotrypsin and pronase was determined by integration of HPLC peaks relative to an internal standard sample of Fmoc-Phe-OH. A 1 mM sample of analyte in pH 7.4 ammonium bicarbonate buffer (with 2% DMSO) was incubated at 37°C in the presence of chymotrypsin or pronase (0.1 mg/mL). Aliquots at various timepoints were analyzed by RP-HPLC (C_{18} 4 x 150 mm column with acetonitrile/water + 0.1% formic acid eluent, 1 mL/min flow rate). Integrated analyte peaks normalized to internal standard were plotted as a percentage of analyte amount in the absence of enzymes.

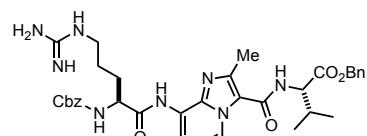
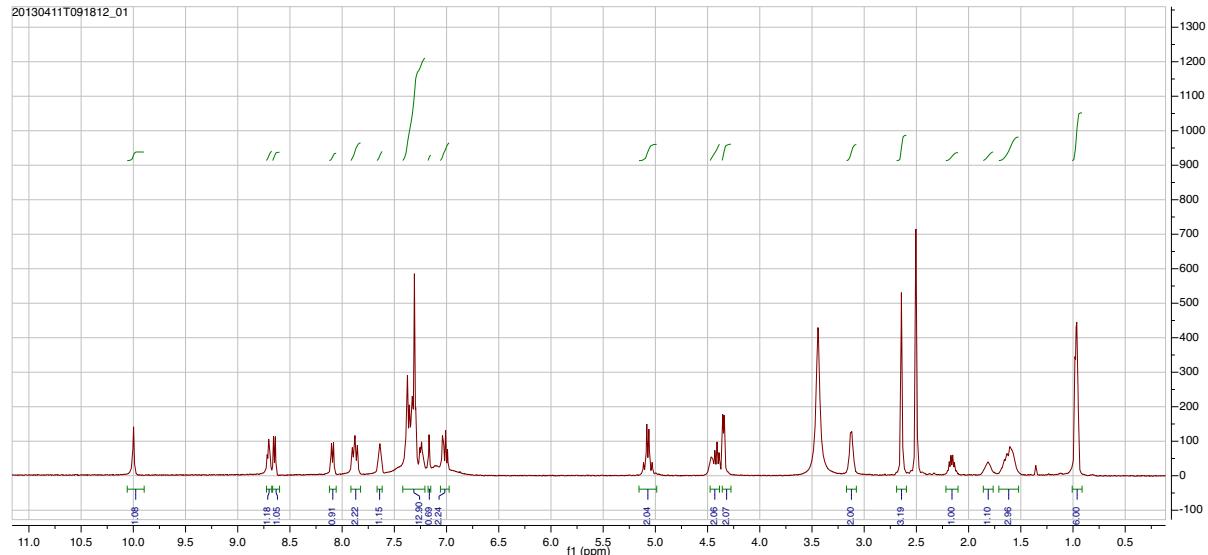
Computational docking studies. The three-dimensional structural coordinates for the activated Akt/protein kinase B ternary complex with glycogen synthase kinase-3 beta (GSK3)-peptide (Ac-GRPRTTSFAE-NH3) and AMP-PNP (Phosphoaminophosphonic acid - adenylate ester) were obtained from the Protein Databank (PDB ID: 1O6K) [1]. After assigning bond orders and atom types, the original hydrogen atoms were removed and then added with an assumed physiological pH of 7.4 for predicting protonation states. Partial atomic charges were assigned using the OPLS-2005 force field [2]. These procedures were performed with the Protein Preparation Wizard [3] from Schrödinger, Inc. Energy minimization was carried out with the Impact Refinement module [4] using the OPLS-2005 force field and truncated newtonian method [5]. Minimization was concluded when the root-mean-square deviation (RMSD) reached a maximum cutoff of 0.30 Å. This step allows hydrogen atoms to be freely minimized, while allowing for heavy-atom movement to relax strained bonds, angles, and clashes. This procedure minimizes potential crystal artifacts that may be present in the PDB input structure. X-ray structures served as reference structures for the calculation of the RMSD. The GSK3 ligand in the crystal structure was used to determine the location of a docking grid box and was then removed along with the AMP-PNP co-crystallized ligand prior to grid generation. The grid size was expanded to fit the length of the GSK3 ligand and all thiol and hydroxyl groups in the grid where allowed to rotate. The rigid receptor docking of the GSK3 control and BE10 was carried out using the Glide [6], version 6.2, program in the extra-precision (XP) mode [7]. No constraints were applied for all docking studies. For both ligands, a maximum of five poses were saved after the docking process.

- [1] Yang, Jing, et al. "Crystal structure of an activated Akt/protein kinase B ternary complex with GSK3-peptide and AMP-PNP." *Nature Structural & Molecular Biology* 9.12 (2002): 940-944.
- [2] Banks, Jay L., et al. "Integrated modeling program, applied chemical theory (IMPACT)." *Journal of computational chemistry* 26.16 (2005): 1752-1780.
- [3] Maestro v9.7, Schrödinger, Inc.: Portland, OR
- [4] Impact version 6.2, Schrödinger, LLC, New York, NY, 2009
- [5] Dembo, Ron S., and Trond Steihaug. "Truncated-newtono algorithms for large-scale unconstrained optimization." *Mathematical Programming* 26.2 (1983): 190-212.
- [6] Glide, version 6.2, Schrödinger, LLC, New York, NY, 2014.
- [7] Friesner, R.A.; Murphy, R.B.; Repasky, M.P.; Frye, L.L.; Greenwood, J.R.; Halgren, T.A.; Sanschagrin, P.C.; Mainz, D.T., "Extra Precision Glide: Docking and Scoring Incorporating a Model of Hydrophobic Enclosure for Protein-Ligand Complexes," *J. Med. Chem.*, 2006, 49, 6177–619

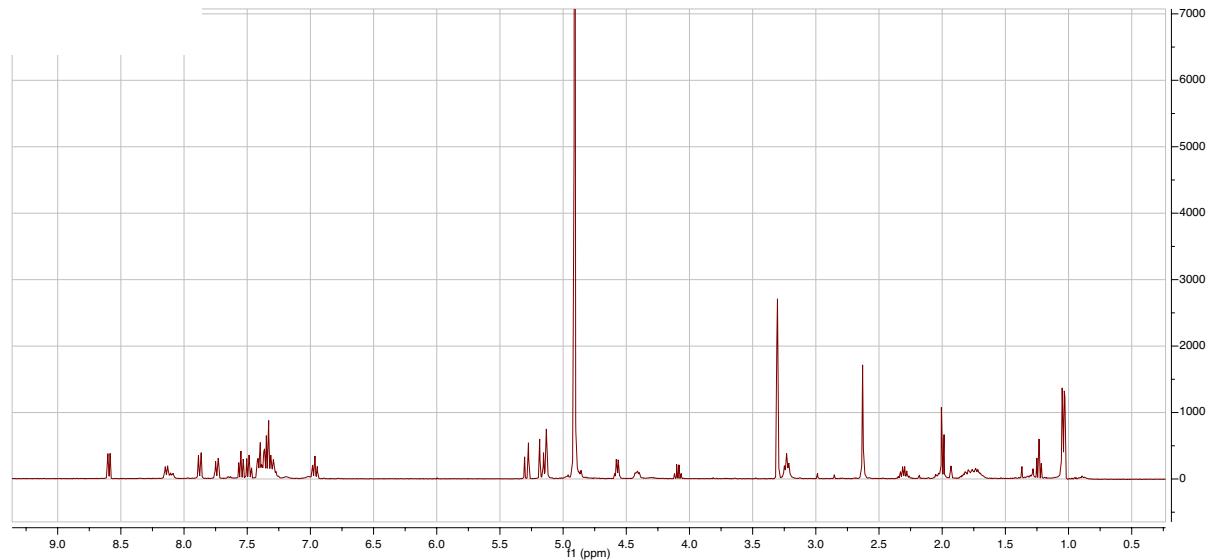
¹H NMR SPECTRA FOR COMPOUNDS 4-31.

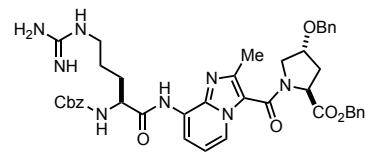


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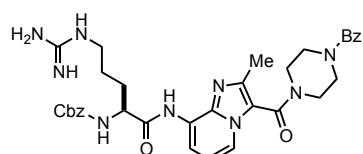
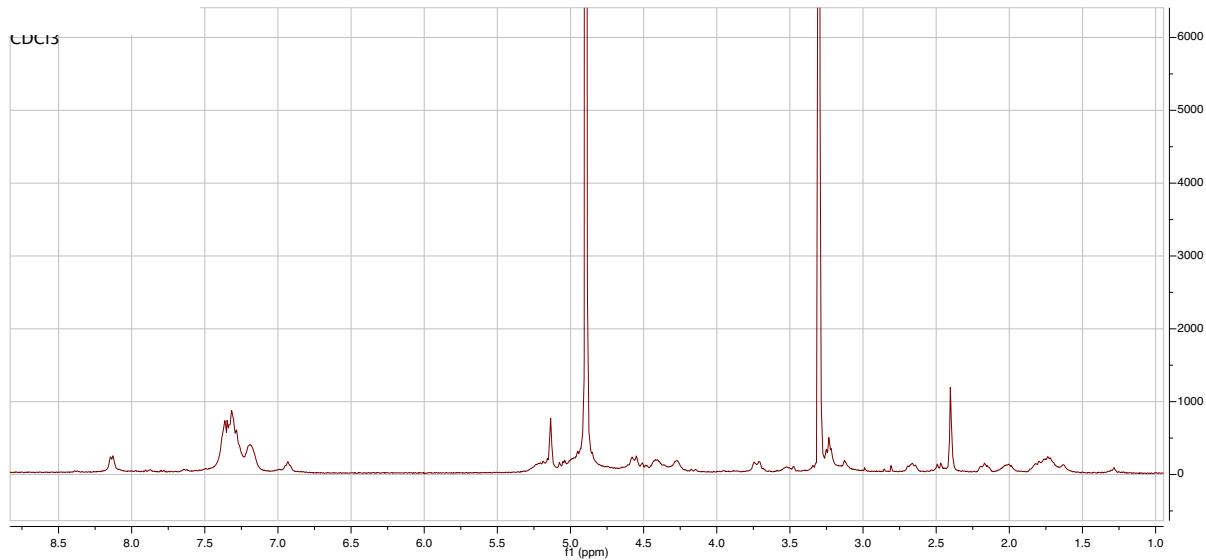


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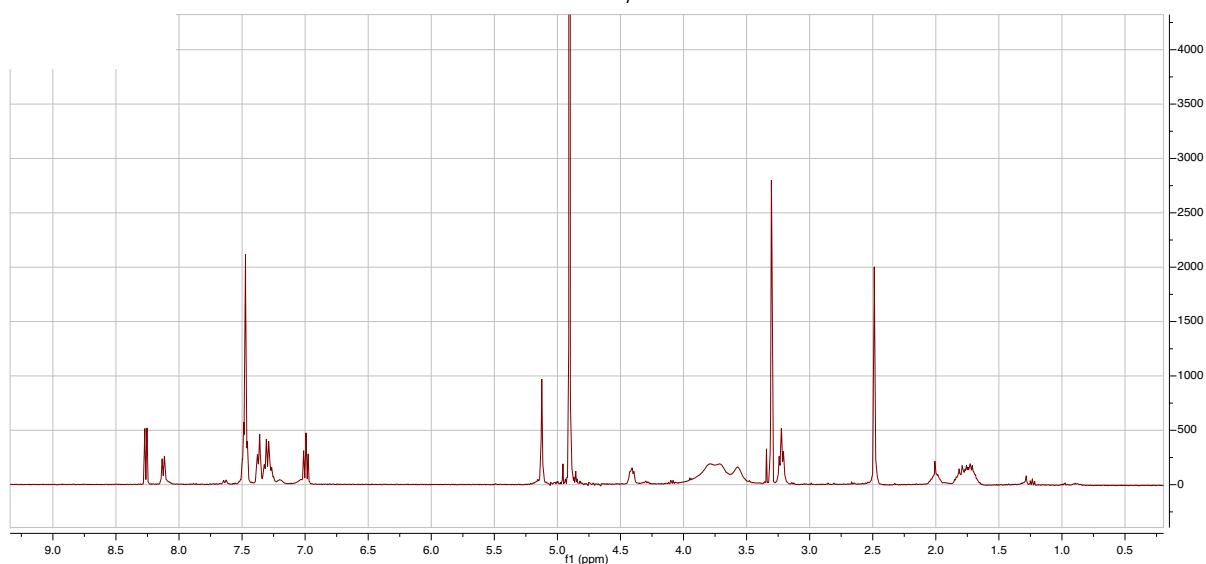


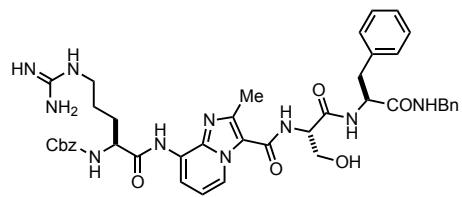


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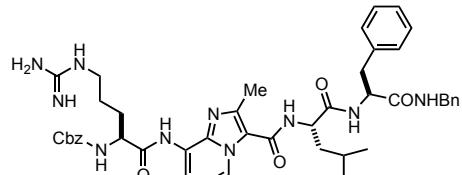
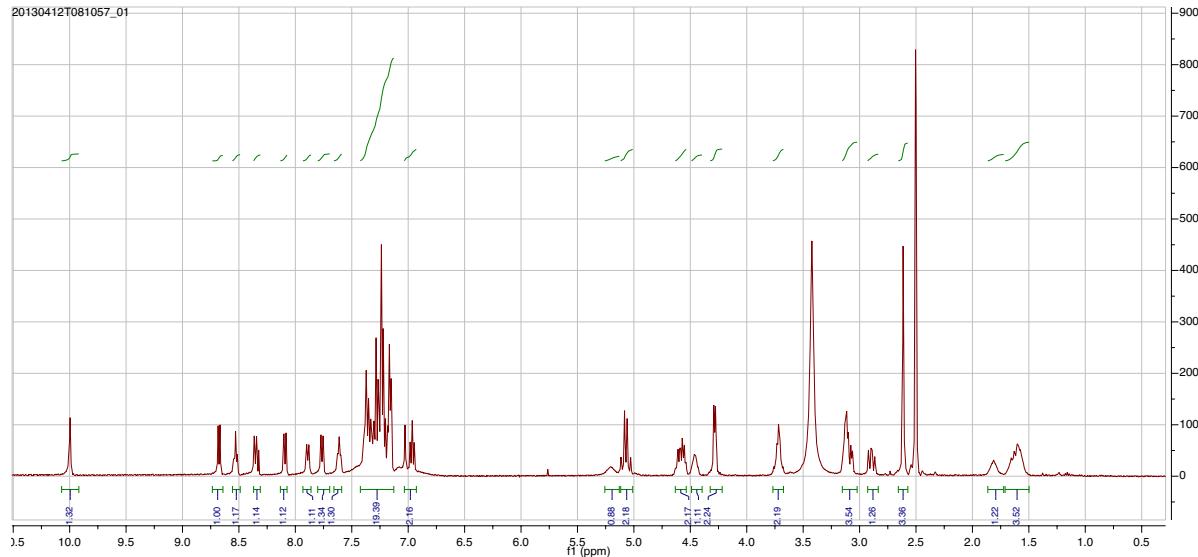


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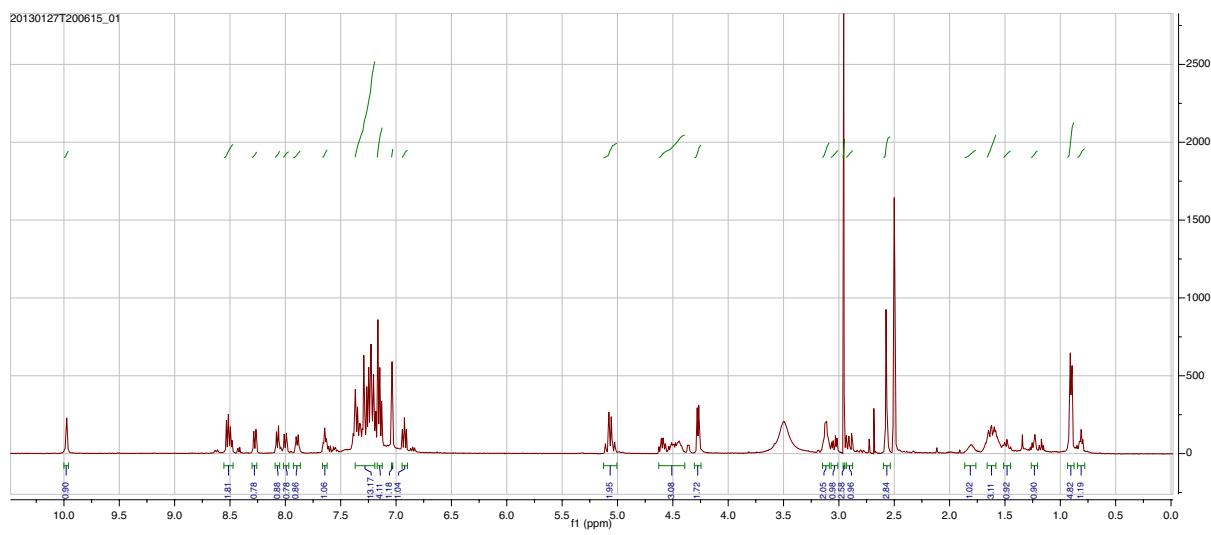


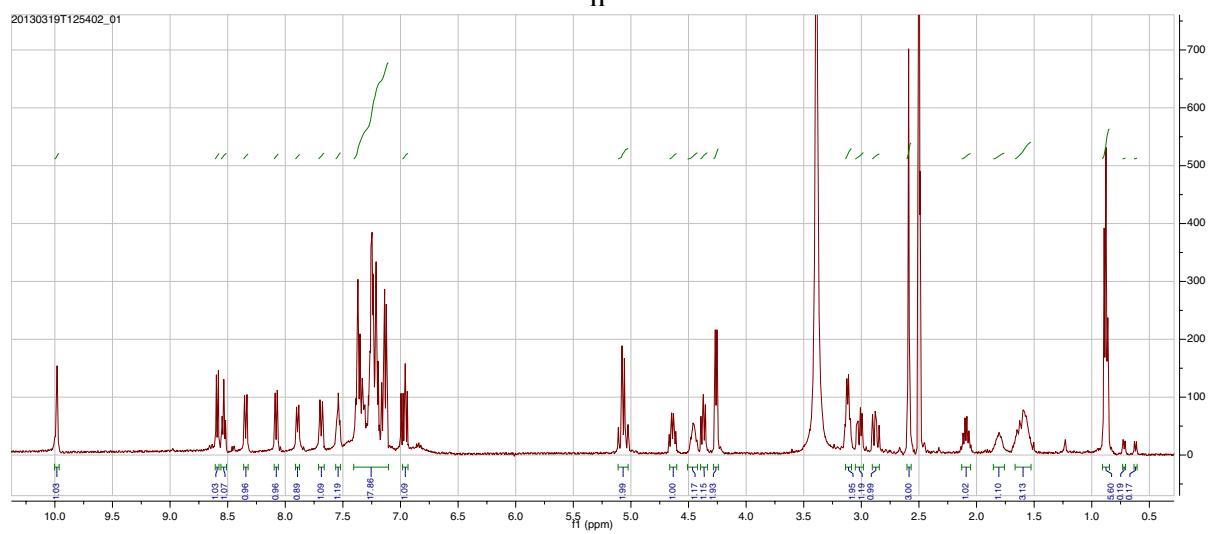
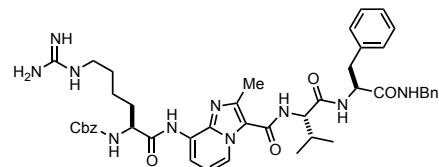
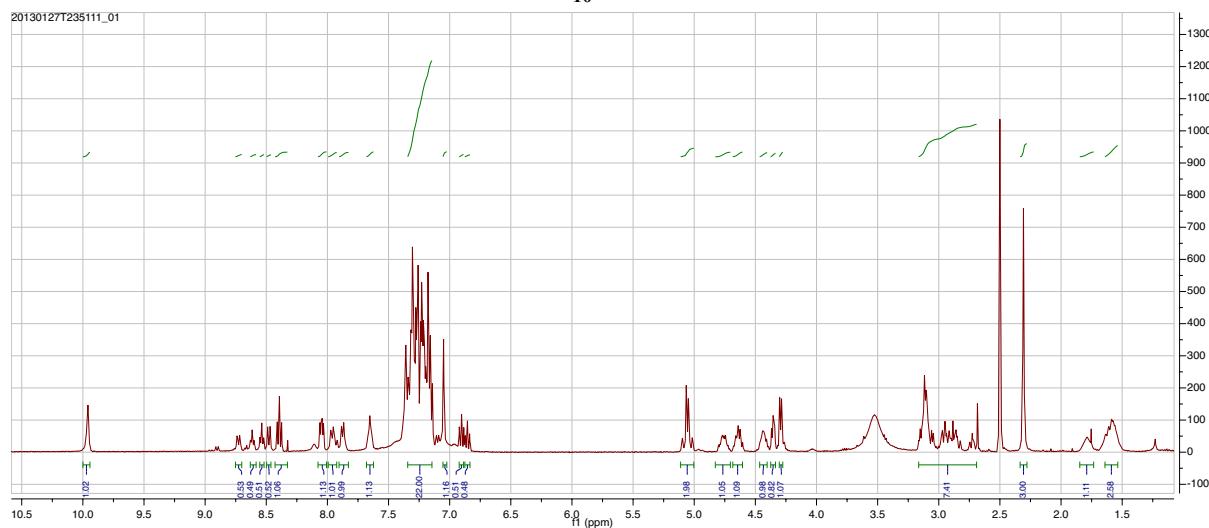
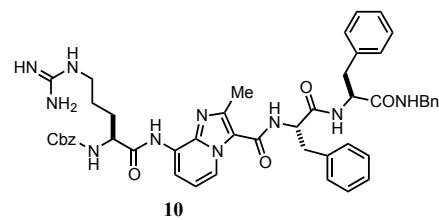


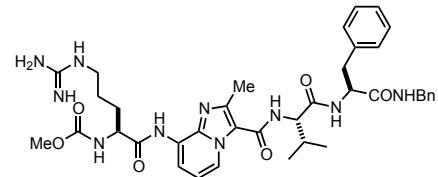
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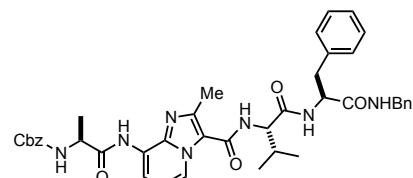
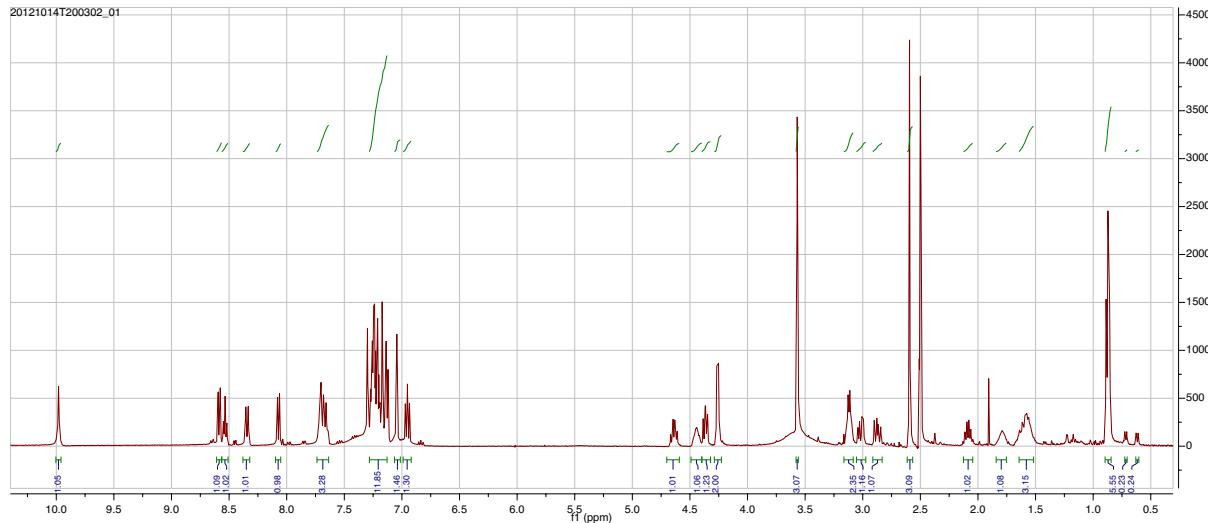
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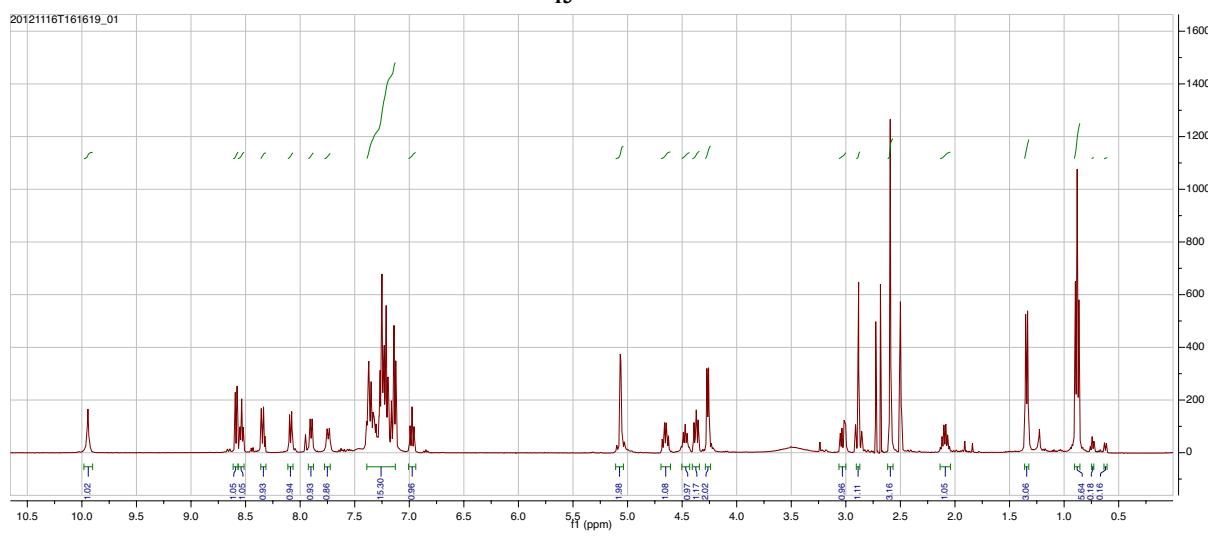


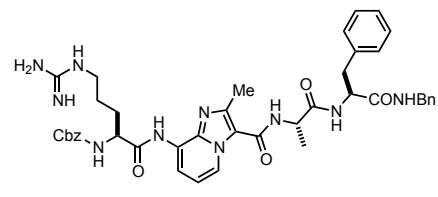


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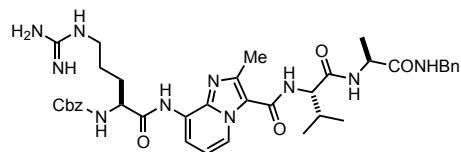
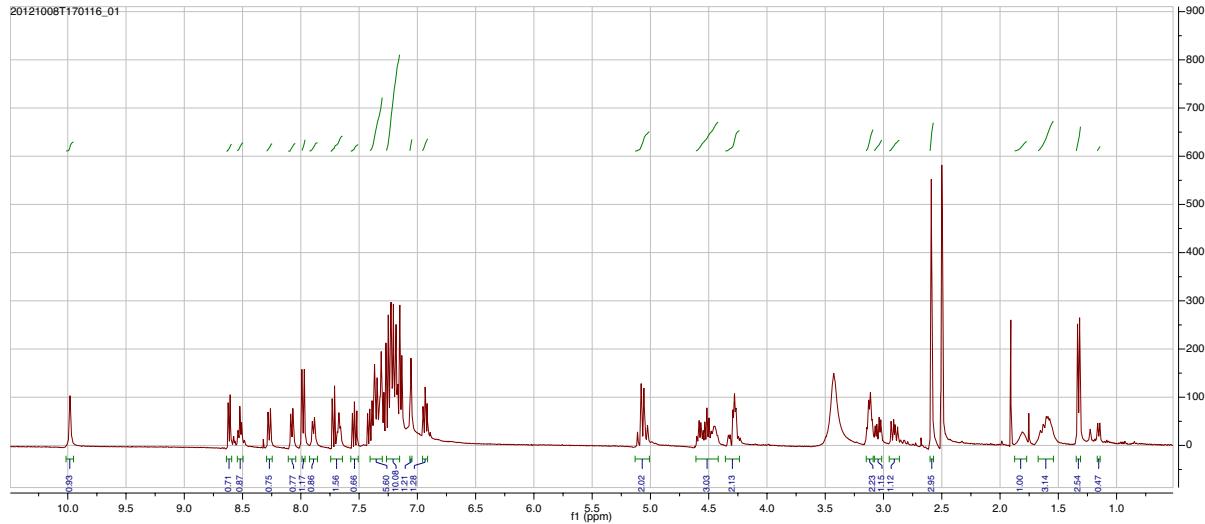


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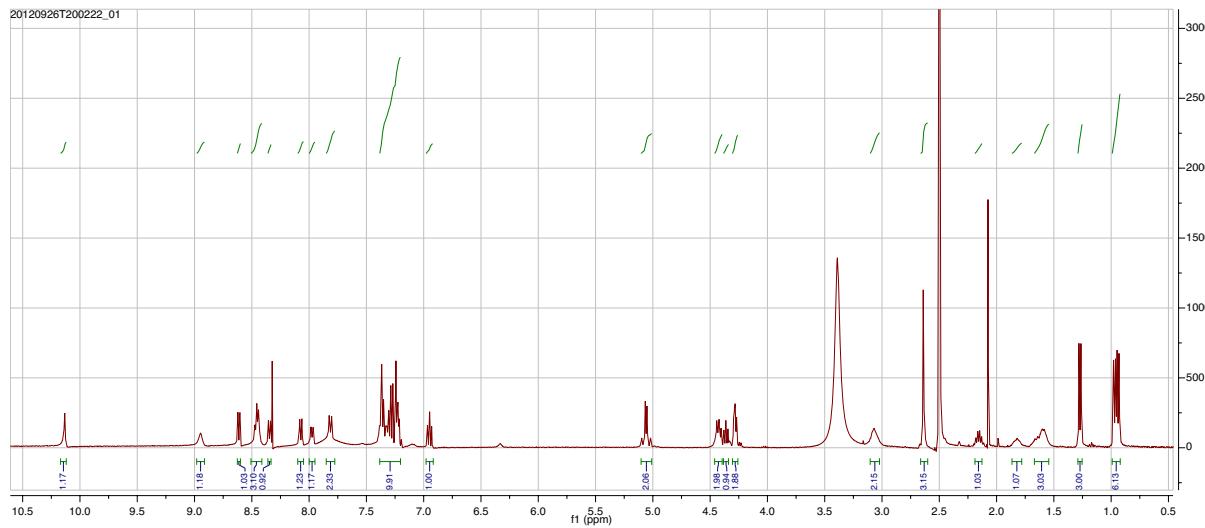


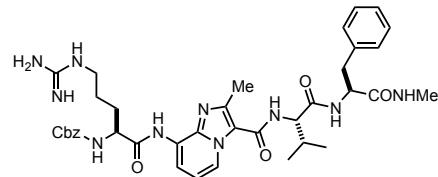


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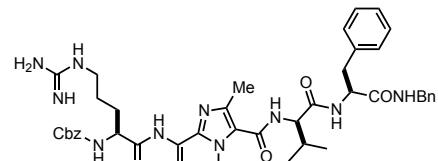
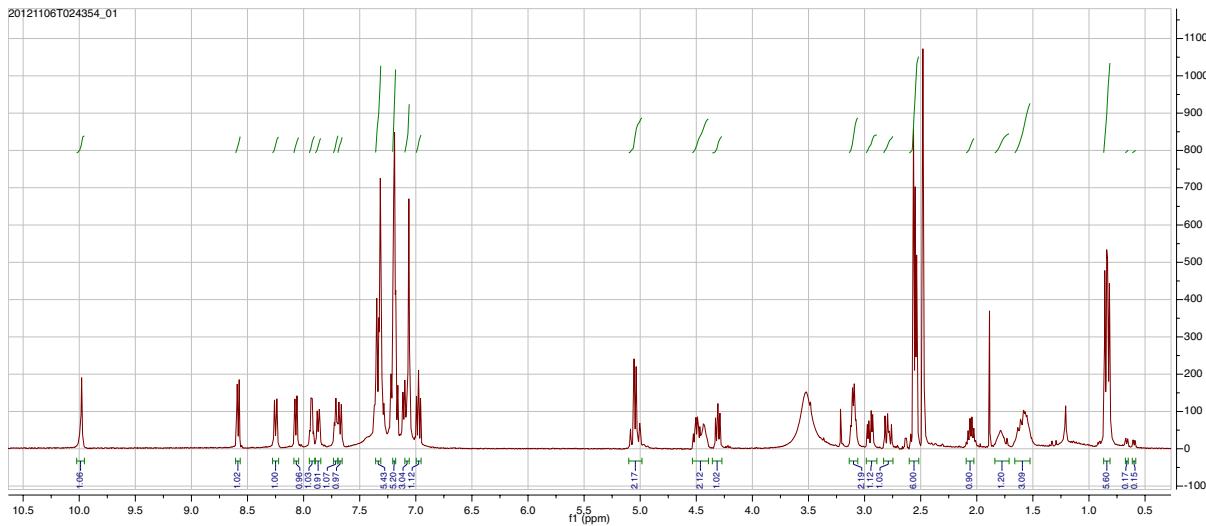


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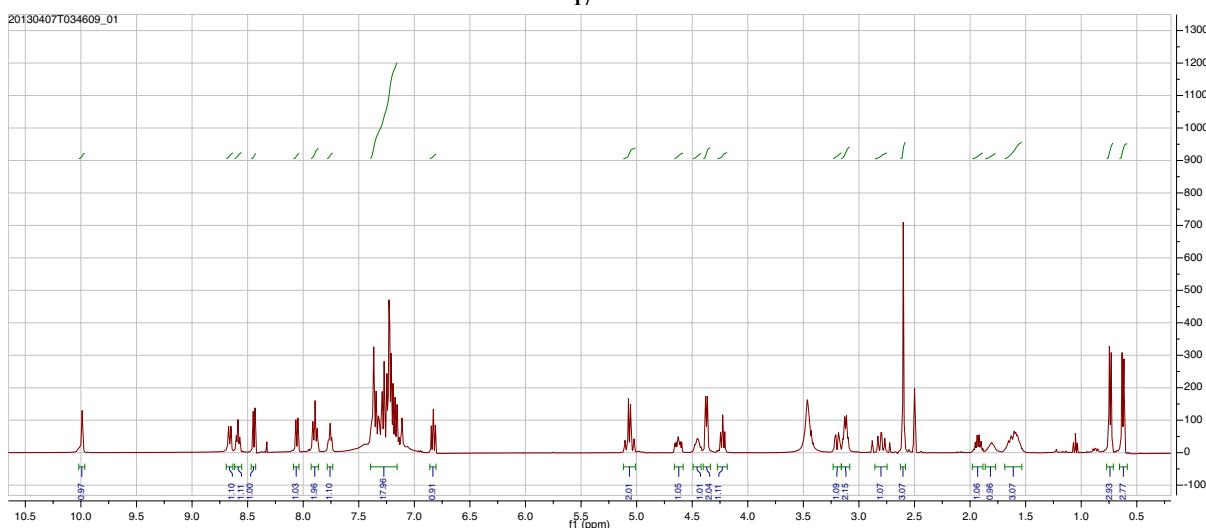


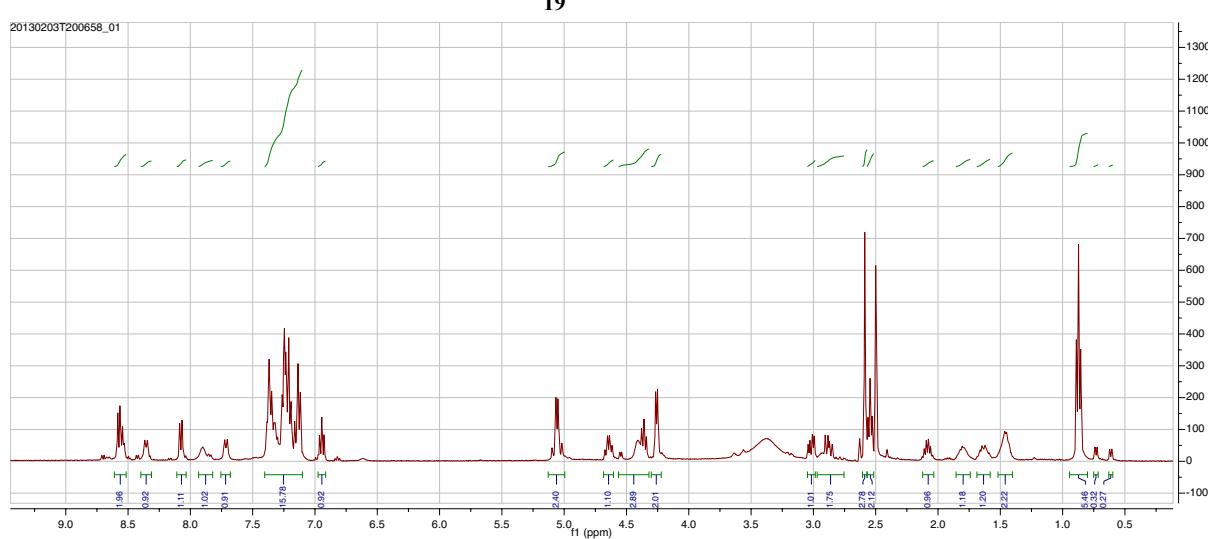
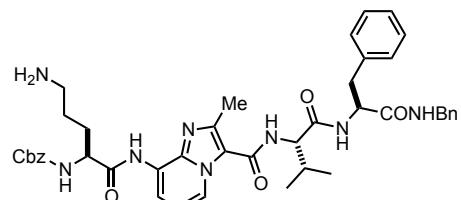
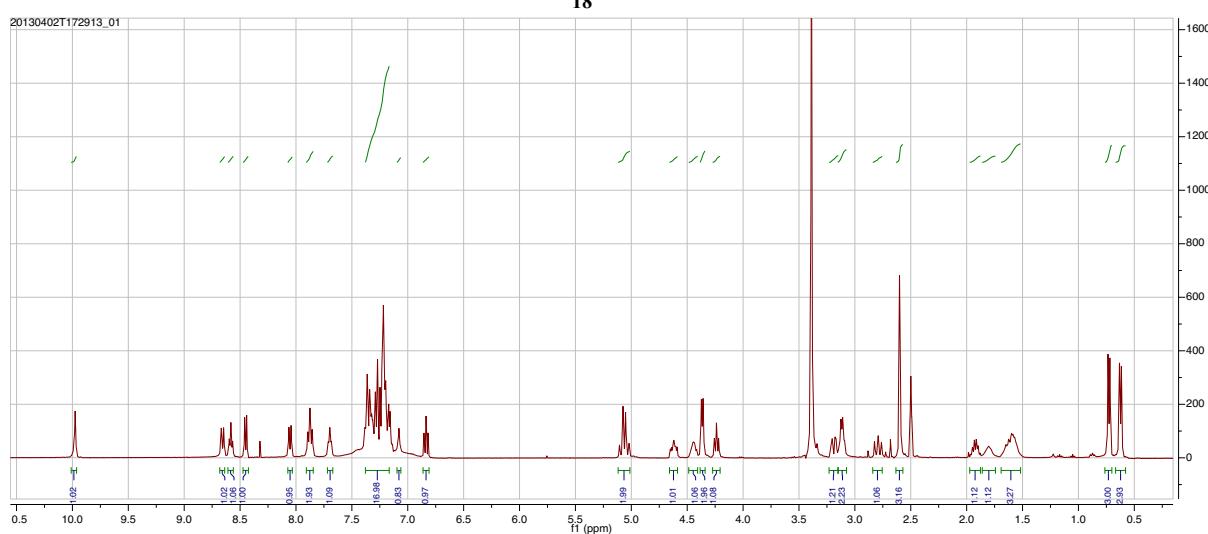
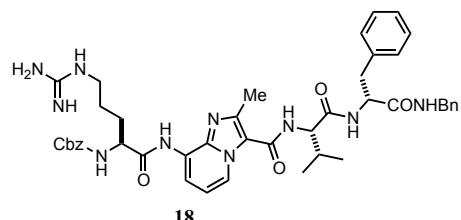


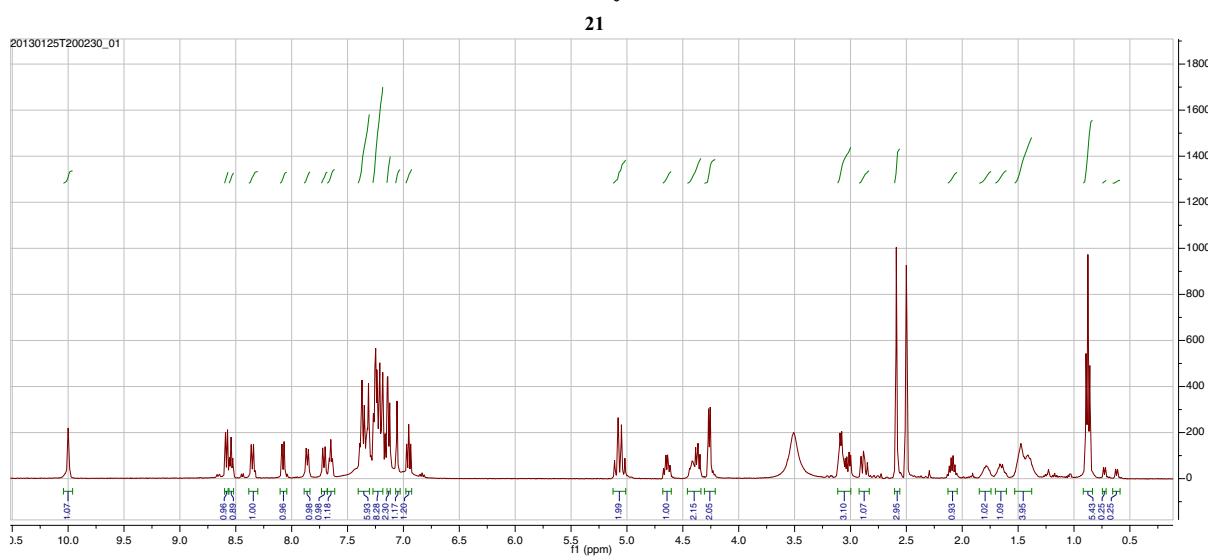
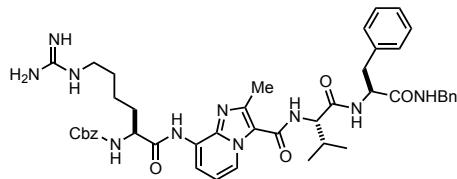
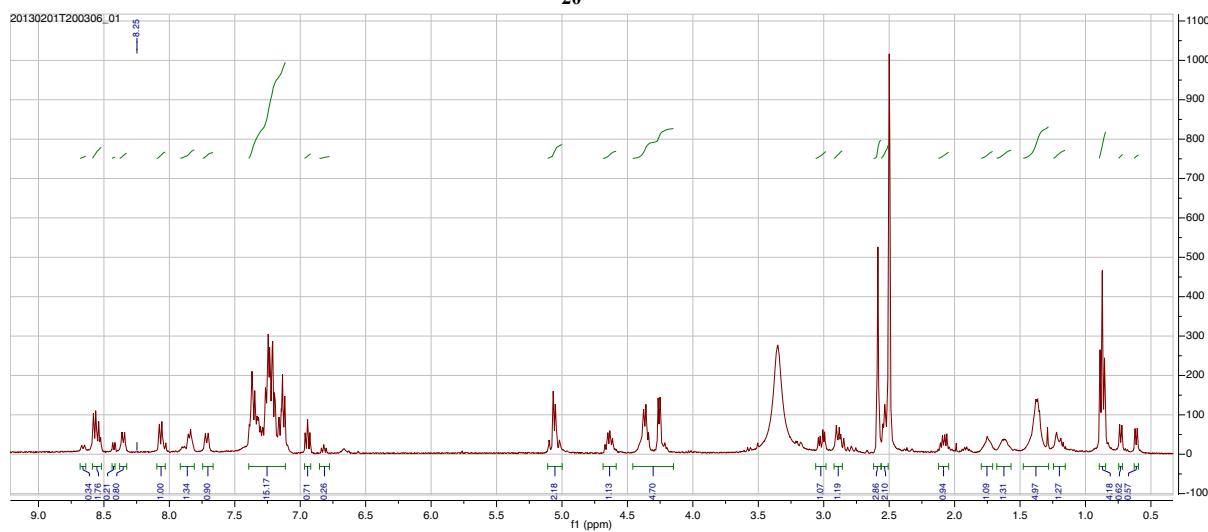
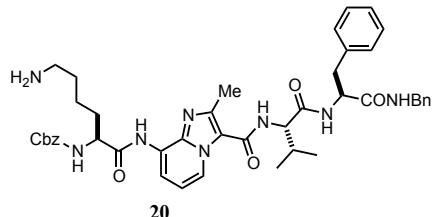
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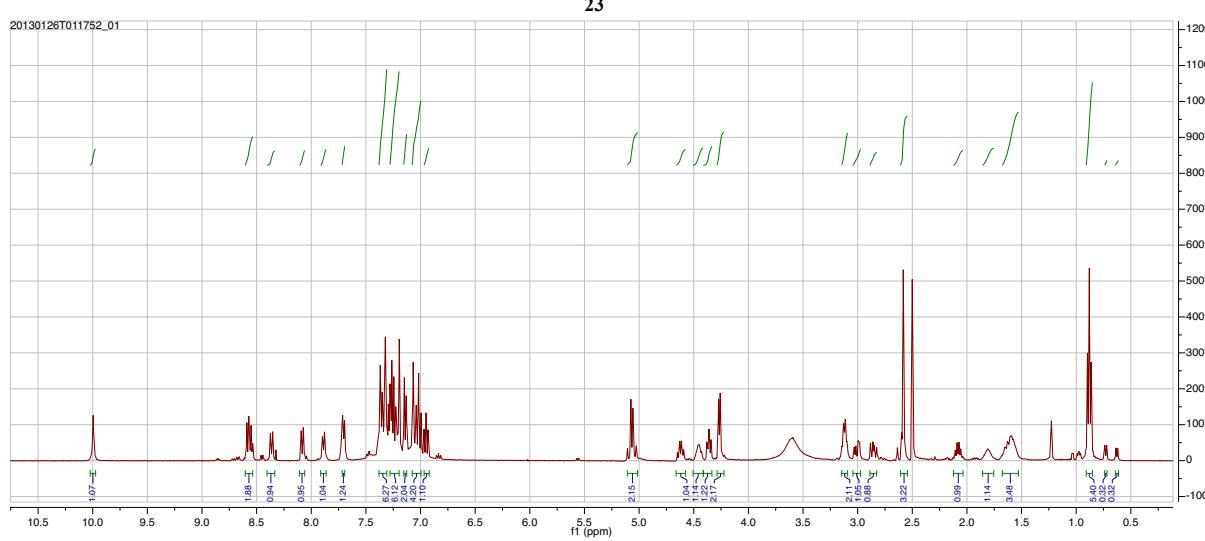
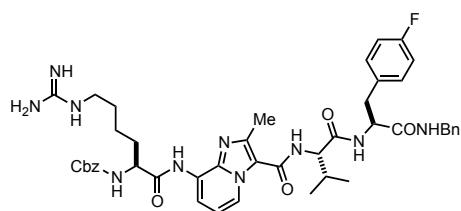
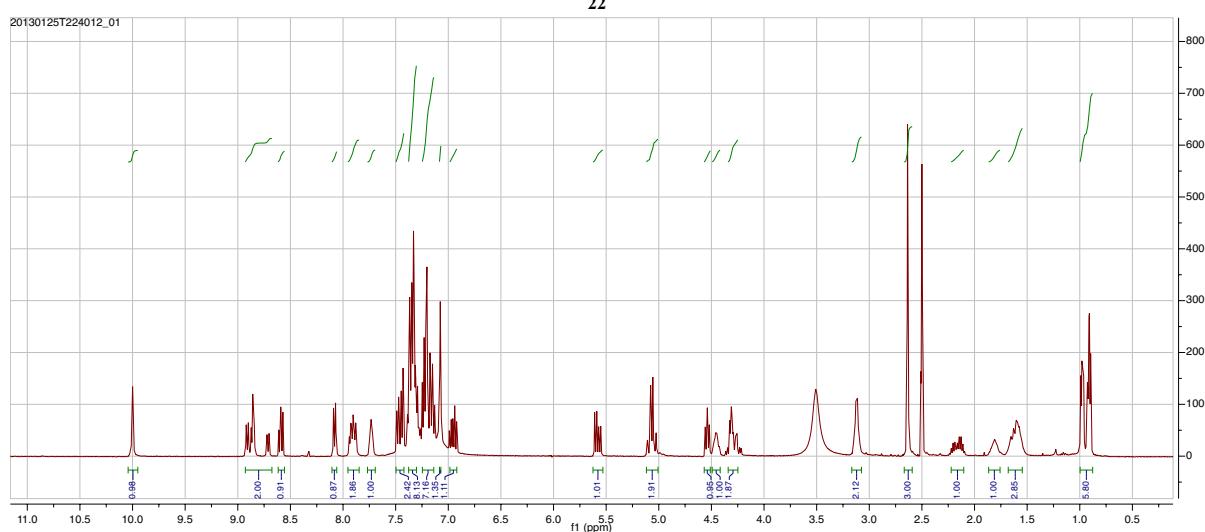
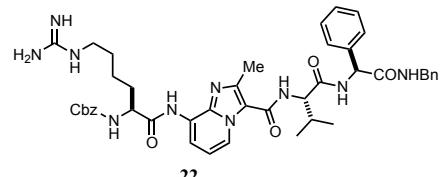


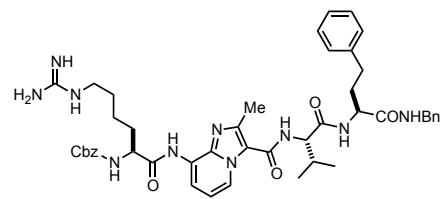
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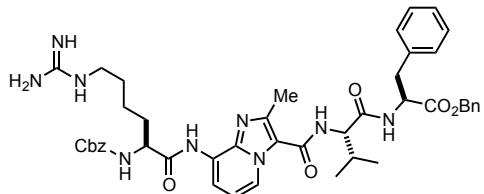
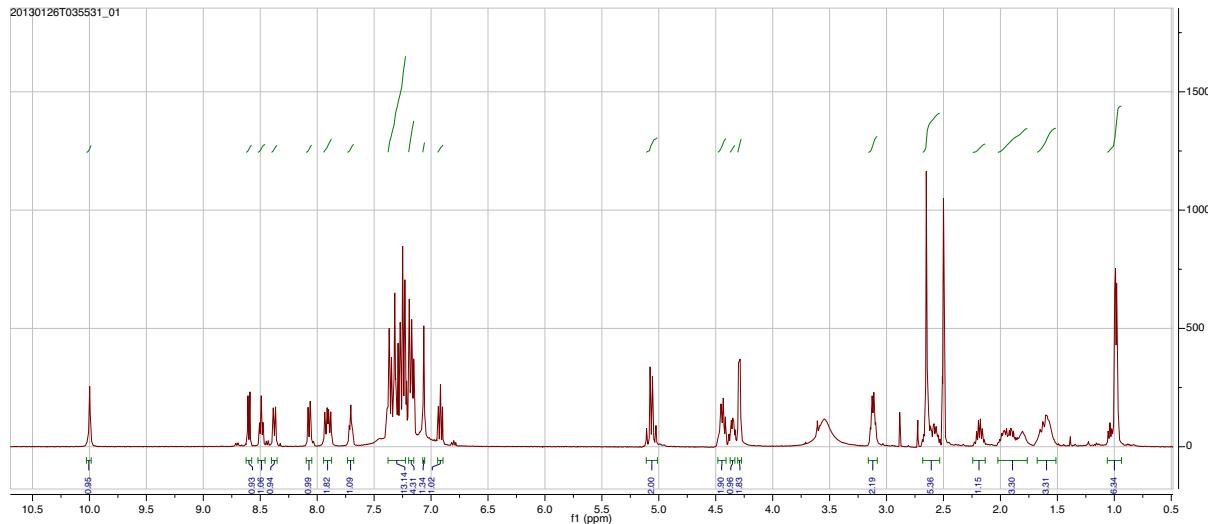






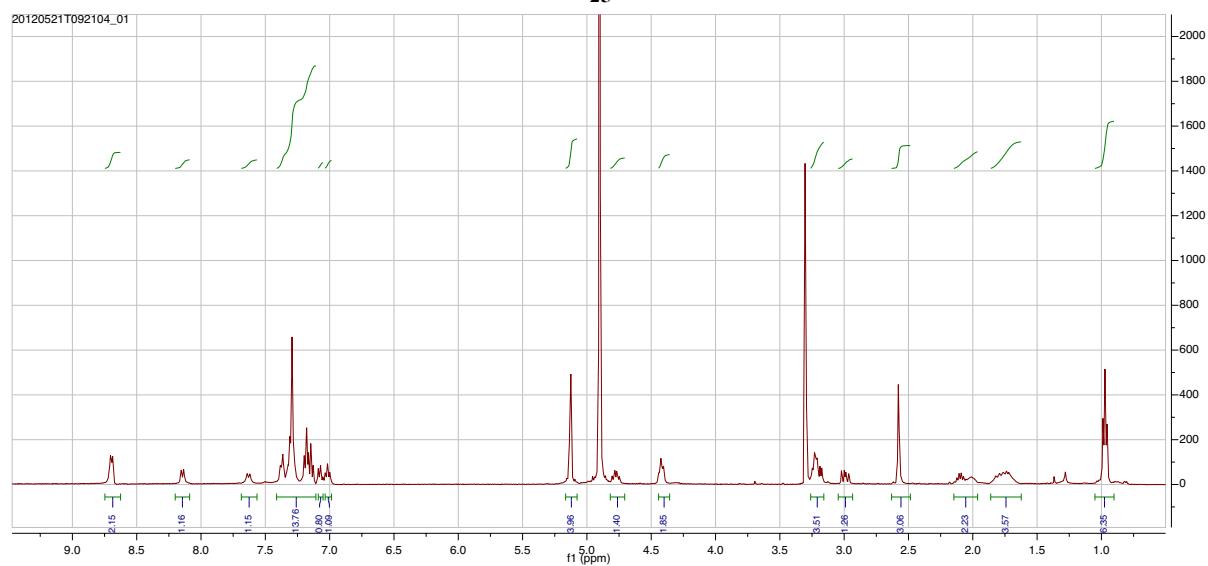
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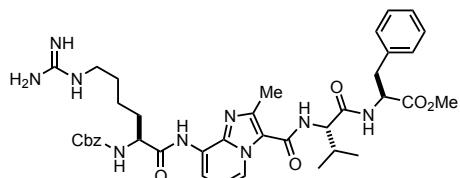
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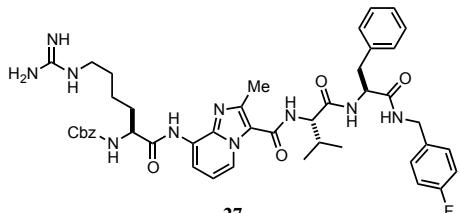
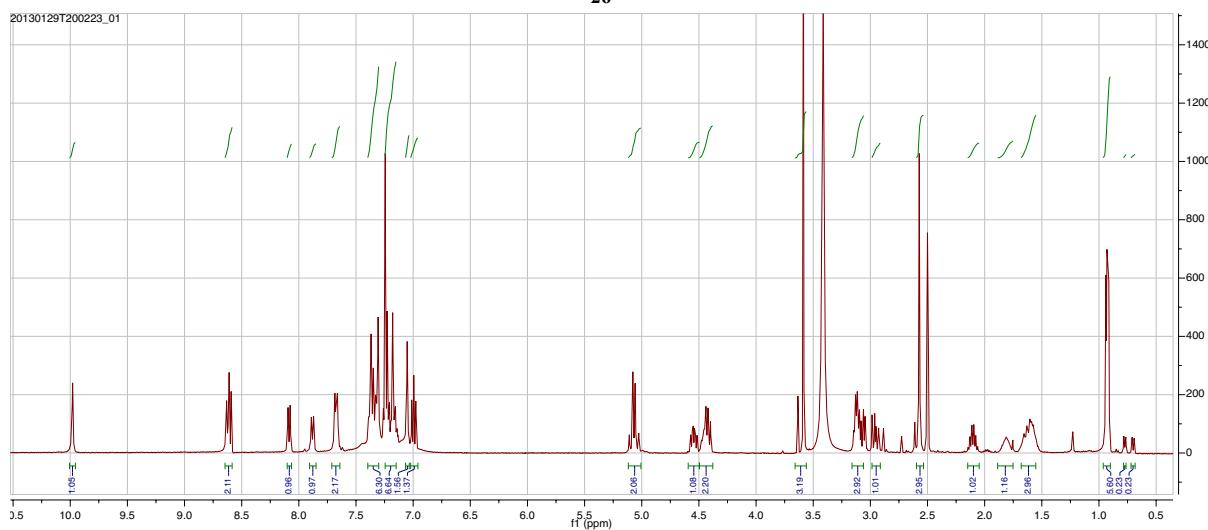
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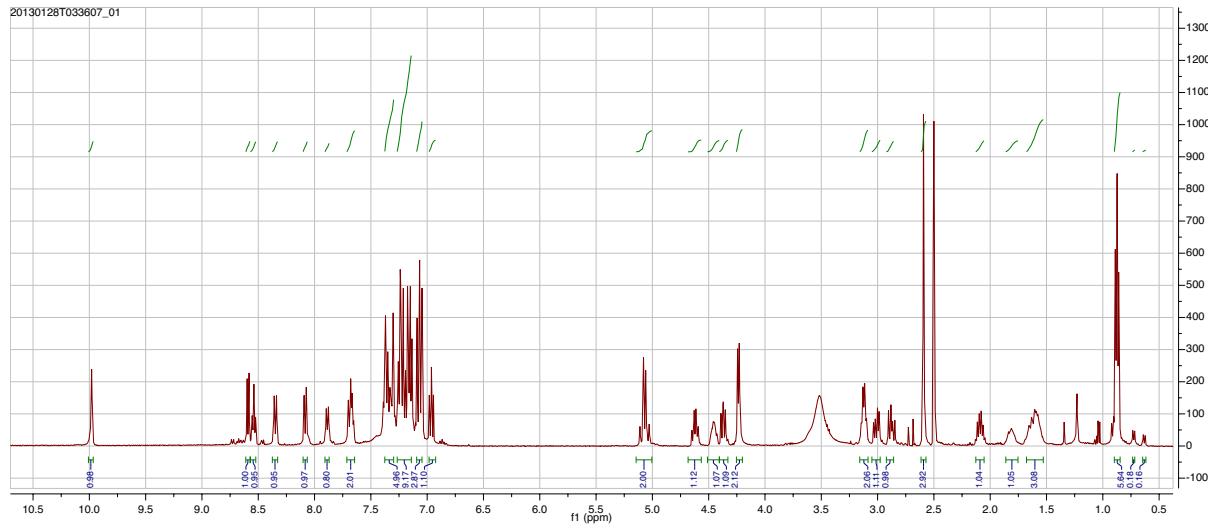


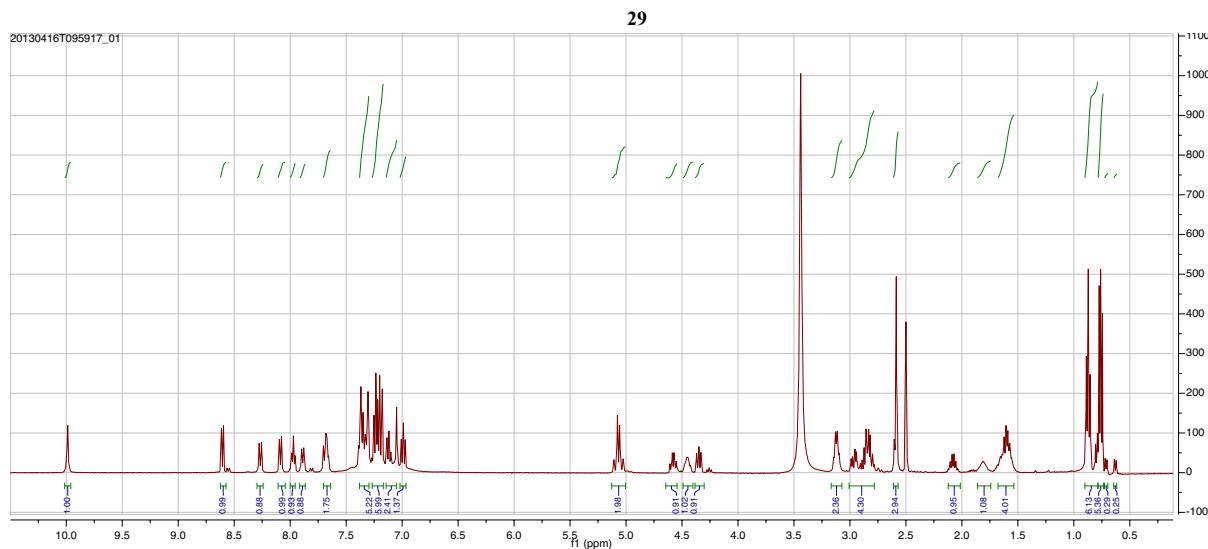
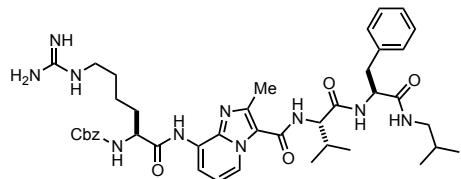
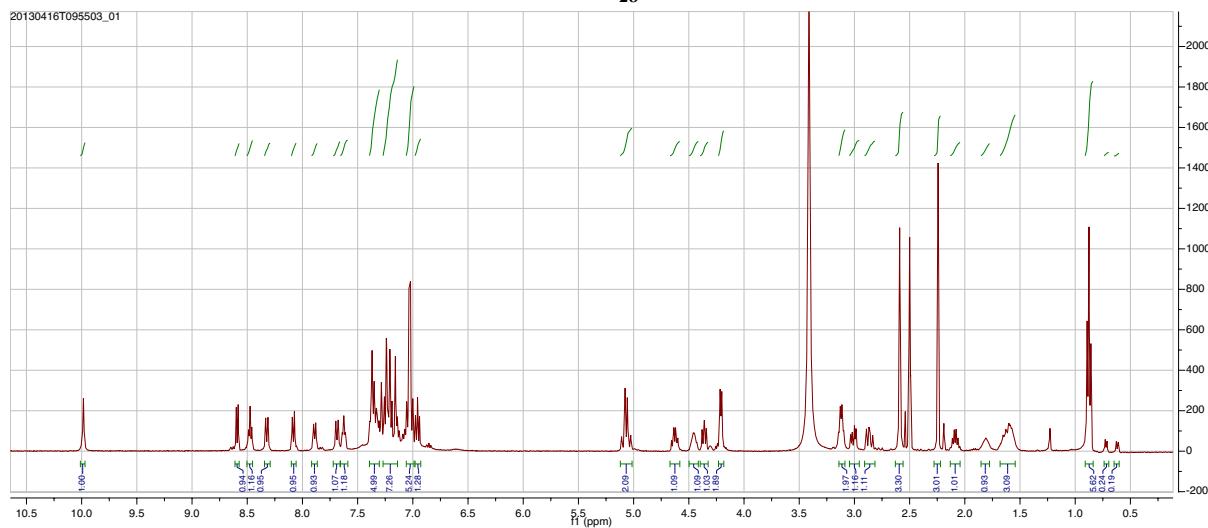
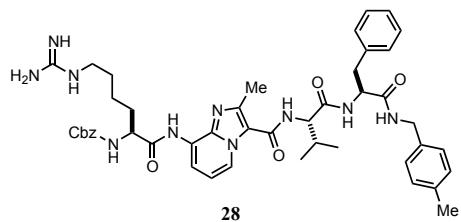


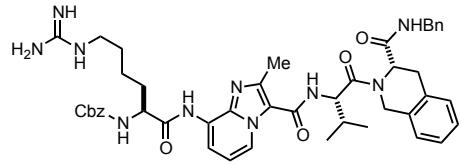
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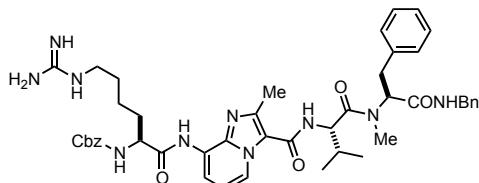
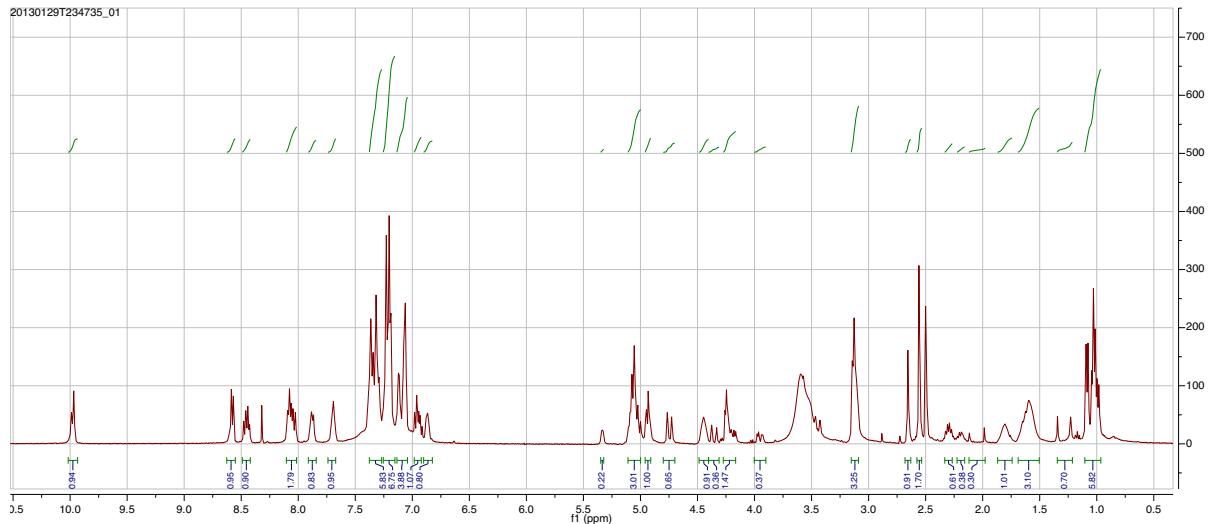
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