## **Supporting Information**

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**Fig. S1.** Experimental strategies used throughout the study. (*A*) The granuloma model of nitric oxide synthase-2–deficient ( $Nos2^{-/-}$ ) mice dermally infected with *Mycobacterium tuberculosis* (*Mtb*): WT and  $Nos2^{-/-}$  mice were infected intradermally with 10<sup>4</sup> *Mtb*. At days 14 and 21 postinfection (p.i.),  $Nos2^{-/-}$  mice received 500 µg anti–IFN- $\gamma$  i.p. or remained untreated. Mice were killed at day 56 p.i., and tissues were used for further analysis. (*B*)  $Nos2^{-/-}$  and arginase-1 (*Arg1*)-*Nos2* double knockout (DKO) mice were infected intradermally with 10<sup>4</sup> *Mtb*. Mice were killed at day 56 p.i., and tissues were used to assess bacterial burden and for histological analysis. (*C*)  $Nos2^{-/-}$  and *Arg1-Nos2* DKO mice were infected intradermally with 10<sup>4</sup> *Mtb*. At days 14 and 21 p.i.  $Nos2^{-/-}$  mice received 500 µg anti–IFN- $\gamma$  i.p. or remained untreated. Mice were killed at day 56 p.i., and tissues were used to assess bacterial burden and for histological analysis. (*C*)  $Nos2^{-/-}$  and *Arg1-Nos2* DKO mice were infected intradermally with 10<sup>4</sup> *Mtb*. At days 14 and 21 p.i.  $Nos2^{-/-}$  mice received 500 µg anti–IFN- $\gamma$  i.p. or remained untreated. Mice were killed at day 56 p.i., and tissues were used for histological analysis. To increase the number of nonnecrotic and necrotic granulomas from  $Nos2^{-/-}$  mice, lung sections from  $Nos2^{-/-}$  and IFN- $\gamma$ -blocked  $Nos2^{-/-}$  mice were pooled.



**Fig. 52.** (A) Immunohistochemical (IHC) staining of representative necrotic granuloma from infected IFN-γ–blocked Nos2<sup>-/-</sup> mice at day 56 p.i. using anti-Arg1 (sc-20150; Santa Cruz) and corresponding isotype. (B) IHC staining of representative necrotic granuloma from infected Arg1-Nos2 DKO mice at day 56 p.i. using anti-Arg1 from Santa Cruz (sc-20150 and sc-18351) and from BD (610708).



**Fig. S3.** Arg1<sup>+</sup> cells surround the necrotic core of caseous granulomas where hypoxic cells are localized. At day 56 p.i., infected mice received the tissue hypoxia marker pimonidazole (PIMO) and were killed 2 h later. (Mock: one animal was left untreated.) Lung sections were stained with anti-Arg1 mAb and anti-PIMO mAb. Images show a representative necrotic tuberculosis (TB) lung granuloma from an infected IFN- $\gamma$ -blocked Nos2<sup>-/-</sup> mouse.



**Fig. 54.** Macrophages are the main cell type expressing Arg1 in lungs from infected  $Nos2^{-/-}$  mice. (A) Representative nonnecrotic and necrotic TB lung granulomas from infected  $Nos2^{-/-}$  mice at day 56 p.i. stained with anti-CD68 (red) and anti-Arg1 (blue). Staining shows colocalization of Arg1<sup>+</sup> cells and CD68<sup>+</sup> cells indicating that Arg1 is expressed in macrophages in TB lung granulomas. (B) Gating strategy used to characterize phagocytic cell populations of the lungs of infected  $Nos2^{-/-}$  mice. Populations: 1, neutrophils; 2, alveolar macrophages; 3, dendritic cells; 4, inflammatory macrophages; 5, monocytes. (C) Histograms representative of Arg1 expression in neutrophils, inflammatory macrophages, and monocytes. The red line depicts Arg1-Nos2 DKO mice, the black line represents  $Nos2^{-/-}$  mice, and the blue line represents IFN- $\gamma$ -blocked  $Nos2^{-/-}$  mice. (D) Alternative gating strategy for characterization of Arg1-expressing cell populations in the lungs of infected  $Nos2^{-/-}$  mice. Arg1<sup>+</sup> cells are gated in the leukocyte population, and then the populations expressing Arg1 are identified as in *B*.

DNA C



Fig. S5. Arg1 is expressed in human lung tissue of TB patients. Human lung samples taken from patients with TB stained with anti-Arg1 mAb. (A and B) Pericavity lung tissue. (C) Cavity wall lung tissue. (D) Distal lung tissue. N, necrotic.

A Nd

S.A



**Fig. S6.** L-arginine-derived metabolites do not affect *Mtb* growth in culture. *Mtb* H37Rv was cultured in Sauton's minimal medium supplemented with L-arginine, L-ornithine, putrescine, spermidine, and spermine. At days 1, 2, 4, and 7 bacterial growth was evaluated by measurement of  $OD_{600}$  (*A*) and by enumerating cfus (*B*). Data are representative of two independent experiments.



**Fig. S7.** Blocking of IFN- $\gamma$  in infected *Arg1-Nos2* DKO mice exacerbates *Mtb* growth and pathology in TB lung granulomas. (*A*) Experimental strategy to evaluate the effect of IFN- $\gamma$  blocking in infected *Arg1-Nos2* DKO mice: *Arg1-Nos2* DKO mice were infected intradermally with 10<sup>4</sup> Mtb. At days 14 and 21 p.i., mice received 500 µg anti–IFN- $\gamma$  i.p. or remained untreated. Mice were killed at day 56 p.i., and tissues were used for bacterial burden and histological analysis. (*B*) H&E staining of lung tissue from infected IFN- $\gamma$ -blocked *Arg1-Nos2* DKO mice at day 56 p.i. (*C*) Lung cfus from infected, nonblocked *Arg1-Nos2* DKO (*n* = 3) and IFN- $\gamma$ -blocked *Arg1-Nos2* DKO (*n* = 6) mice at day 56 p.i. Data show the median and interquartile range. Significance was assessed by the Mann–Whitney test.



**Fig. S8.** Proximity of Arg1<sup>+</sup> cells affects T-cell proliferation in microenvironments within granulomas. Arg1, Ki-67, and CD3ε immunofluorescence staining of representative nonnecrotic and necrotic TB lung granulomas from infected Nos2<sup>-/-</sup> and Arg1-Nos2 DKO mice at day 56 p.i.



**Fig. S9.** The gradients of the numbers of Ki-67<sup>+</sup> cells in nonnecrotic granulomas from infected Nos2<sup>-/-</sup> and Arg1-Nos2 DKO mice at day 56 p.i. are not different. (A) Linear regression of the gradient of number of Ki-67<sup>+</sup> cells in nonnecrotic granulomas from infected Nos2<sup>-/-</sup> and IFN- $\gamma$ -blocked Nos2<sup>-/-</sup> mice (blue lines) and from Arg1-Nos2 DKO mice (red lines) at day 56 p.i. Lines correspond to linear fits of centered Ki-67<sup>+</sup> cells values on separate granuloma regions for each of the two subgroups. Shaded areas show corresponding 95% confidence bands. (*B* and C) Counts of Ki-67<sup>+</sup> cells through granuloma regions 1–4 of individual nonnecrotic (*B*) and necrotic (*C*) granulomas from infected Nos2<sup>-/-</sup> and Arg1-Nos2 DKO mice.