

Supporting Information

for

Reversibly locked thionucleobase pairs in DNA to study base flipping enzymes

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Derivation of equation (1) and Figures S1 and S2

Derivation of equation (1)

The thermodynamic constants K_D^{unlocked} , $K_{D,\text{init}}$ and K_{flip} for the two-step binding model shown in Figure 5a are defined as follows:

$$K_D^{\text{unlocked}} = \frac{[\text{DNA}] \cdot [\text{E}]}{[\text{E} \cdot \text{DNA}]} = \frac{[\text{DNA}] \cdot [\text{E}]}{[\text{A}] + [\text{B}]} \quad (4)$$

$$K_{D,\text{init}} = \frac{[\text{DNA}] \cdot [\text{E}]}{[\text{A}]} \quad (5)$$

$$K_{\text{flip}} = \frac{[\text{B}]}{[\text{A}]} \quad (6)$$

with $[\text{DNA}]$, concentration of unbound DNA; $[\text{E}]$, concentration unbound enzyme; $[\text{E} \cdot \text{DNA}]$ concentration of all enzyme-DNA complexes; $[\text{A}]$, concentration of initial complex with innerhelical target base; $[\text{B}]$, concentration of complex with flipped target base and $[\text{E} \cdot \text{DNA}] = [\text{A}] + [\text{B}]$.

Taking the inverse of eq. (4) and separating the terms:

$$\frac{1}{K_D^{\text{unlocked}}} = \frac{[\text{A}] + [\text{B}]}{[\text{DNA}] \cdot [\text{E}]} = \frac{[\text{A}]}{[\text{DNA}] \cdot [\text{E}]} + \frac{[\text{B}]}{[\text{DNA}] \cdot [\text{E}]} \quad (7)$$

Substituting with eqs. (5) and (6), $\frac{[\text{A}]}{[\text{DNA}] \cdot [\text{E}]} = \frac{1}{K_{D,\text{init}}}$ and $\frac{[\text{B}]}{[\text{DNA}] \cdot [\text{E}]} = \frac{K_{\text{flip}}}{K_{D,\text{init}}}$, gives

$$\frac{1}{K_D^{\text{unlocked}}} = \frac{1}{K_{D,\text{init}}} + \frac{K_{\text{flip}}}{K_{D,\text{init}}} = \frac{1 + K_{\text{flip}}}{K_{D,\text{init}}} \quad (8)$$

Taking the inverse again yields equation (1):

$$K_D^{\text{unlocked}} = \frac{K_{D,\text{init}}}{1 + K_{\text{flip}}} \quad (1)$$

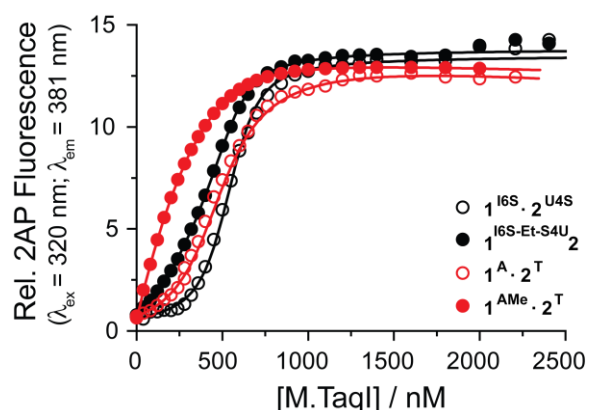


Figure S1: Direct comparison of competition binding titrations of DNA with unlocked duplex $1^{I6S} \cdot 2^{U4S}$ (black open circles), locked duplex $1^{I6S-Et-S4U}_2$ (black closed circles), A/T substrate duplex $1^A \cdot 2^T$ (red open circles) and A^{Me}/T product duplex $1^{A^{Me}} \cdot 2^T$ (red closed circles) with M.TaqI. Increasing amounts of M.TaqI were added to a mixture of the DNA of interest and a fluorescent DNA with known K_D that contains the target base analog 2-aminopurine (2AP). A more delayed increase in the 2AP fluorescence indicates tighter binding of the non-fluorescent DNA.

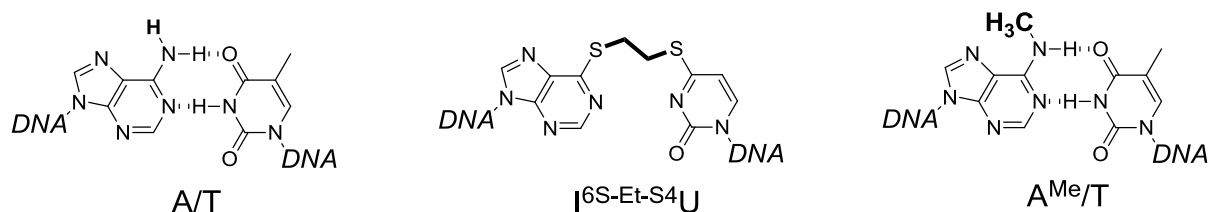


Figure S2: Comparison of the substrate A/T (left) and product A^{Me}/T (right) base pair of adenine-specific DNA MTases, with the cross-linked $I^{6S-Et-S4U}$ thionucleoside base pair (middle) used to mimic the innerhelical position of the target base in the initial M.TaqI-DNA complex before base flipping. The cross-linked base pair is missing the methyl group and therefore might be a better mimic for the A/T substrate than for the A^{Me}/T product in DNA.