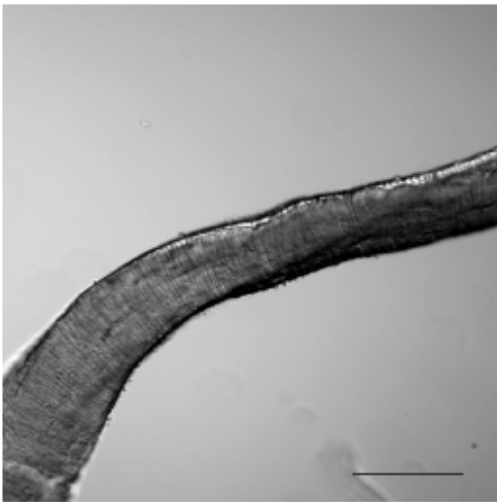


## SUPPLEMENTARY MATERIAL

### Human satellite cells as a manageable tool for gene therapy

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#### Supplemental Figure 1



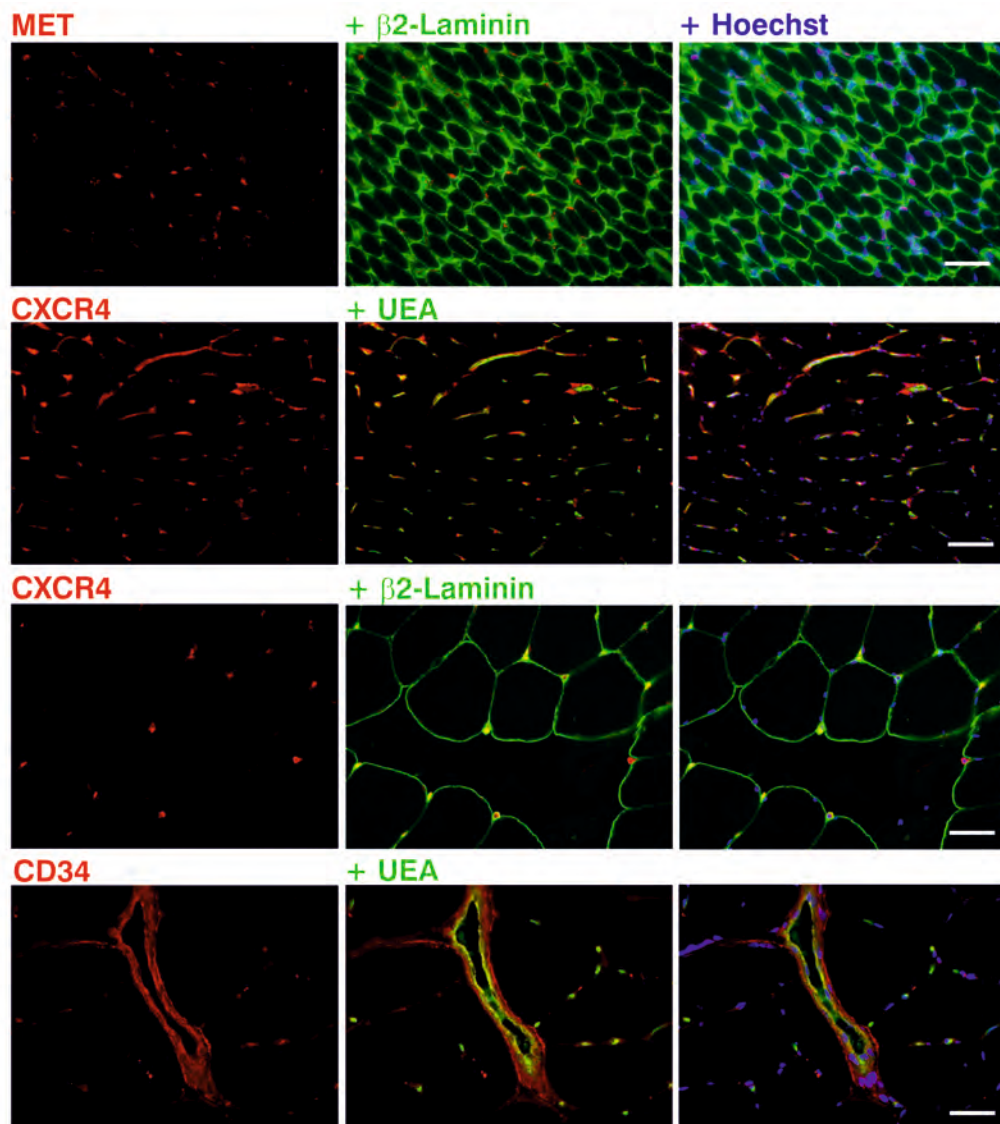
Single human muscle fiber fragment after manual dissection. Bar 200  $\mu\text{m}$

**Supplementary Table 1. Summary of experiments**

<b>Number of muscle biopsy specimens</b>	<b>Age and gender of donors</b>	<b>Procedure</b>	<b>Number of experiments</b>
69	20-80 years	HMFF <sup>1</sup> characterization	580 HMFF
	34 male, 35 female	Characterization of colonies that grew onto culture dish	304 HMFF
		Characterization of colonies after hypothermic treatment (4°C)	272 HMFF
6	44-64 years 3 male, 3 female	Transplantation	33 transplantations in TA muscle

<sup>1</sup> Human muscle fiber fragment

## Supplementary Figure 2

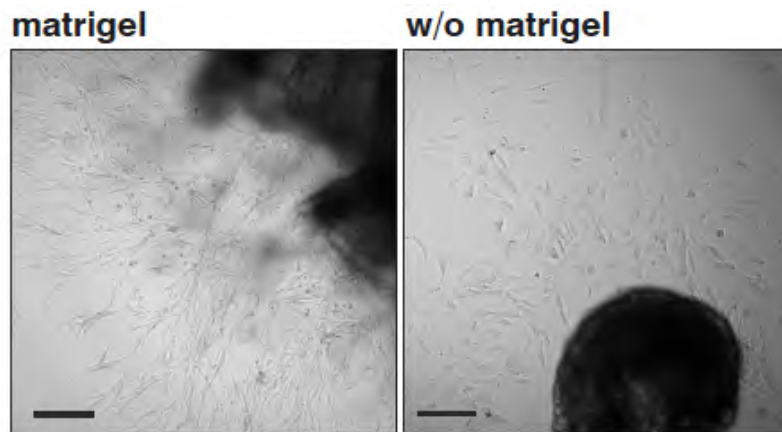


Surface marker profile of human satellite cells. Upper panel: Skeletal muscle cryosections from a three months old infant. MET positive cells (red) are frequent in the satellite cell niche. In muscle section from adult probands MET<sup>+</sup> cells could not be detected. Second and third panel: CXCR4 (red) is expressed in endothelial cells of skeletal muscle tissue; Ulex European Agglutinin (UEA), green. Some CXCR4 positive cells are located in the interstitial space outside of the basal lamina ( $\beta$ 2 laminin, green). Lower panel: Endothelial cells of skeletal muscle tissue express high levels of CD34 (CD34: red, UEA: green). No cells in the satellite cell niche are positive for CD34. Bars: 50/100/50/50  $\mu$ m.

**Supplementary Video 1** Viable satellite cells in single human fiber fragment.

Satellite cells migrating in the satellite cell niche of human a single fiber. The fiber has been cultured for 13 days prior to recording for 17 h. Images were taken every 15 min. Note the sliding cell inside of the surrounding membrane. Bar: 75  $\mu$ m.

**Supplementary Figure 3**

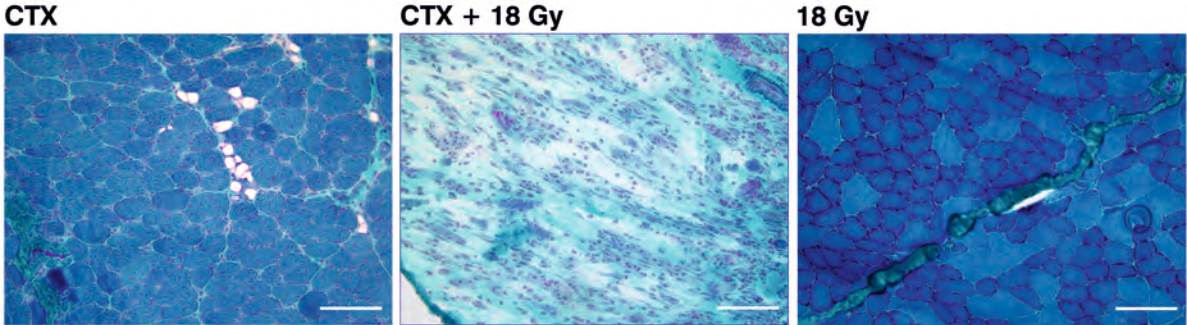


Colony formation of cells outgrowing the HMFF could be observed 10 days after initiating the HMFF irrespective whether the fiber was placed on Matrigel® (left) or on plastic (right). Bar: 200  $\mu$ m.

**Supplemental Table 2: Antibodies used for immunohistochemistry**

<b>Antibody</b>	<b>Species</b>	<b>Working concentration</b>	<b>Catalog number, Company</b>
Anti-BrdU	rat	10 µg/ml	ab 6326, Abcam, Cambridge, UK
Anti-CD34	rabbit	2 µg/ml	sc-9095, Santa Cruz Biotechnology, Santa Cruz, CA, USA
Anti-CXCR4	rabbit	2.5 µg/ml	ab 2074, Abcam
Anti-c-met	rabbit	2 µg/ml	sc-161, Santa Cruz Biotechnology
Anti-desmin	mouse	1:500 (cells) 1:50 (histology)	M 0760, Dako, Glostrup, Denmark
Anti-desmin	rabbit	4 µg/ml	ab 8592, Abcam
Anti-hu Lamin A/C	rabbit	1:2000	ab 108595, Epitomics, Cambridge, UK
Anti-β2 laminin	mouse	0.1 µg/ml	Novus Biologicals, Littleton, CO, USA
Anti-Myf5	rabbit	0.2 µg/ml	sc-302, Santa Cruz Biotechnology
Anti-MyoD	mouse	1 µg/ml	sc-32758, Santa Cruz Biotechnology
Anti-NCAM (CD56)	mouse	2 µg/ml	130-090-955, Miltenyi Biotech, Bergisch Gladbach, Germany
Anti-Pax7	mouse	Supernatant, undiluted	DSHB, Iowa City, Iowa, USA

**Supplemental Figure 4**



Effect of irradiation (18 Gy) on mouse tibialis anterior (TA) muscle. Cardiotoxin (CTX) was injected into TA muscle without irradiation (left) or 24 h after irradiation (middle). Irradiation completely abolished muscle regeneration, but had no effect on muscle morphology in the absence of CTX (right). Histology was obtained 9 days after irradiation Bar, 100  $\mu$ m.