

Development of α GlcN(1 \leftrightarrow 1) α Man-based Lipid A Mimetics as Novel Class of Potent Toll-like Receptor 4 (TLR4) Agonists

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Supplementary SI-Table 1

Glycosylation of lactol **8** with 2-*O*-levulinoyl-protected mannosyl donors

A coupling of the TCA donor **18** and lactol acceptor **8** using trimethylsilyl trifluoromethanesulfonate (TMSOTf) as promoter resulted in the formation of orthoester **23** as the major product (23%), along with hydrolyzed donor **16** and unreacted acceptor **8** (Scheme 4A, SI-Table 1: Entry 1). By lowering the reaction temperature and using higher amounts of promoter, the formation of the orthoester could be suppressed, although the target α,α -disaccharide **22** was obtained in only 8% yield (SI-Table 1: Entry 2) together with concurrently formed donor self-coupling product **24** (Scheme 4A). With the less reactive NPTFA donor **19** the formation of the orthoester was not observed, however the desired product **22** was isolated only in trace amounts (SI-Table 1: Entry 3).

The failure of the imidate-mediated couplings was initially linked to the diminished reactivity of the axial 1-OH group in acceptor **8** due to the H-bonding with the carbamate NH group. Therefore, an inverse glycosylation procedure¹ was employed where TMSOTf was exchanged to boron trifluoroetherate (BF₃•Et₂O) as promoter to avoid 1-*O*-silylation of the relatively unreactive acceptor **8** (SI-Table 1: Entry 4). Again, a preponderant formation of the orthoester **23** was observed. Since orthoesters can be rearranged to glycosides,² *in situ* activation of **23** with additional amount of promoter was attempted (SI-Table 1: Entry 4, 5) which did not result in the formation of the desired α,α -product **22**. Varying temperature, amount of the donor/promoter (TMSOTf) and the solvent did not improve the glycosylation outcome (SI-Table 1: Entry 5). With thioglycoside donor **14** only a trace amount of the disaccharide **22** was obtained (SI-Table 1: Entry 6).

SI-Table 1. Glycosylation outcome of the couplings between acceptor 8 and donors 18, 19 and 14 (according to Scheme 4A).

	Donor (eq)	Promotor (eq)	Solvent	T / °C	Product / Yield
1	18 (1)	TMSOTf (0.05)	CH ₂ Cl ₂	0	23 / 23%
2	18 (1)	TMSOTf (0.2)	CH ₂ Cl ₂	-15	22 / 8%
3	19 (1.5)	TMSOTf (0.05÷1.5)	CH ₂ Cl ₂	-15÷0÷25	22 / traces
4	19 (1.5)	BF ₃ •Et ₂ O (0.05)	CH ₂ Cl ₂	0	23 / <i>in situ</i> activation BF ₃ •Et ₂ O (0.05 eq.)
5	19 (1.5)	TMSOTf (0.01)	CH ₃ CN-CH ₂ Cl ₂ , 10:1	0	23 / <i>in situ</i> activation TMSOTf (0.01 eq.)
6	14 (1.2)	TfOH/NIS (0.1/1.0)	CH ₂ Cl ₂	0	22 / traces

¹H- and ¹³C- NMR-data for orthogonally protected GlcN(1↔1)Man disaccharides

Supplementary SI-Table 2

SI-Table 2: ¹H- and ¹³C-NMR shifts and the corresponding coupling constants (¹J_{C₁,H₁} and ³J_{H₁,H₂}) of the anomeric protons/carbons of GlcN(1↔1)Man disaccharides.

Compound	GlcN(1←				→)Man				
	C-1 ppm	¹ J _{C,H} Hz	H-1 ppm	³ J _{1,2} Hz	C-1' ppm	¹ J _{C,H} Hz	H-1 ppm	³ J _{1,2} Hz	H-5 ppm
22 (αα)	92.77	175	5.14	3.7	93.44	172	5.02	1.1	3.68
28 (αα)	91.95	178	5.23	3.6	92.58	174	5.11	-	3.77
35 (αα)	92.96	173	5.15	3.8	93.72	170	5.09	1.6	3.63
36 (αα)	93.07	172	5.20	3.5	93.70	173	5.12	-	3.70
37 (αα)	93.92	175	5.13	3.1	94.87	178	5.11	1.3	3.68
38 (αβ) ^[a]	97.83	173	5.15	3.6	101.35	157	4.62	-	3.28
39 (αβ)	98.20	177	5.08	3.5	101.37	155	4.61	-	3.29
40 (αβ)	97.79	175	4.94	3.2	99.86	156	4.60	-	3.26
41 (βα)	100.12	n.d.	4.74	-	100.12	n.d.	4.96	-	4.03
44 (βα)	100.20	159	4.72	8.4	99.81	172	4.94	1.3	4.04
43 (βα)	101.68	160	4.78	8.3	100.70	171	5.0	-	4.10

^[a] The first anomeric prefix (e.g. αβ) designate the configuration at the GlcN moiety as depicted in the formula αGlcN(1↔1)βMan.

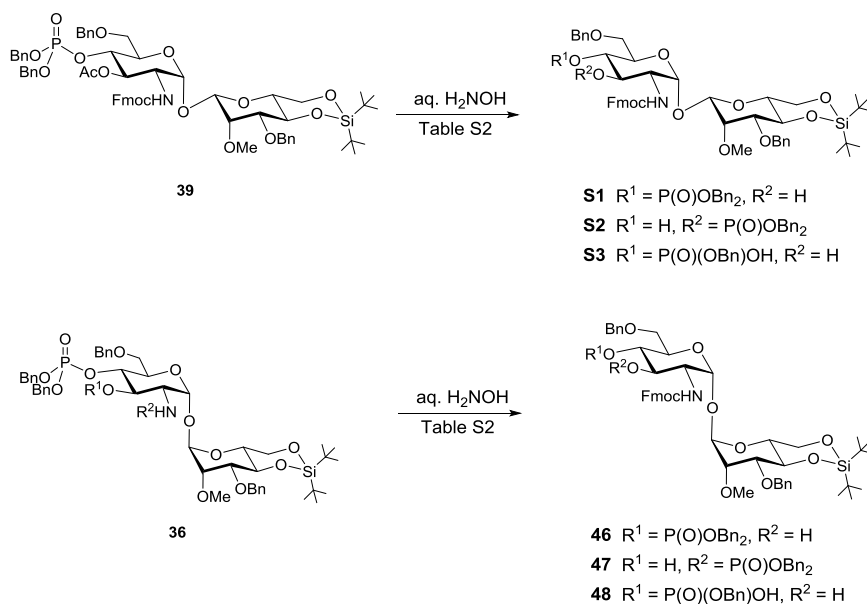
3-*O*-Deacetylation in the presence of the adjacent 2-*N*-Fmoc group.

To establish the conditions for the removal of 3-*O*-acetate protecting group in the α,α -disaccharide **36** the trial experiments with α GlcN(1 \leftrightarrow 1) β Man disaccharide **39** which possesses the same protecting group pattern as **36** but has a β -configuration at C-1 of Man were performed first. Our attempts were directed either to simultaneous removal of 3-*O*-Ac and 2-*N*-Fmoc groups or, alternatively, to the regioselective cleavage of the 3-*O*-acetate in the presence of the adjacent 2-*N*-Fmoc protection.

First, a simultaneous removal of both 3-*O*-Ac and 2-*N*-Fmoc protecting groups was attempted. If successful, the regioselective *N*-acylation of the free amino group would precede the DIC/DMAP-mediated *O*-acylation. Treatment of **39** with MeONa/MeOH or aq. Et₃N in methanol resulted in the cleavage of Fmoc group to furnish a free amine as a single product (positive ninhydrin-staining of the intermediate on TLC and appearance of a UV-positive apolar spot which was rationalized as released dibenzofulvene) whereas the acetate group persisted. Longer reaction times resulted in the removal of 3-*O*-acetate though the 4-*O*-phosphate group was cleaved. Although a successful acetate removal under Zemplén conditions in the presence of a phosphotriester group has been reported,³ the loss of the 4-*O*-dibenzylphosphate group in **39** under basic conditions could be expected.

At the same time, the procedures for the exclusive cleavage of 3-*O*-acetate were investigated. A successful application of 4 Å molecular sieves in methanol for the cleavage of acetates in the presence of *N*-Fmoc group has been reported.⁴ Upon application of these conditions for the deprotection of **39** no reaction was observed. By using 3 Å molecular sieves in methanol-CH₂Cl₂⁵ the acetate cleavage in **39** was extremely sluggish and was accompanied by decomposition. To check the feasibility of both methods in our hands, we submitted the peracetylated monosaccharide **11** to deacetylation with 3 Å or 4 Å molecular sieves in methanol. According to HPLC-MS both reactions cleanly afforded deacetylated thiomannoside **12**, demonstrating the practicability of these deprotection approaches, which, however, were not applicable to the 3-*O*-deacetylation of the (1 \leftrightarrow 1) disaccharide **39**. Application of *p*-toluenesulfonic acid⁶ for the cleavage of 3-*O*-Ac group did not bring any improvements.

Supplementary SI-Scheme 1.



Next, the possibility of using the powerful α -nucleophile hydroxylamine,^{7,8} which has been reported to serve as an effective acyl group acceptor⁹ capable of cleaving amides¹⁰ and esters¹¹ was exploited (SI-Scheme 1). Upon treatment of **39** with 50% aq. hydroxylamine in a biphasic mixture with THF the 3-*O*-deacetylated compound **S1** was isolated as the major product (55%) whereas the *N*-Fmoc group was retained. The reaction was accompanied by migration of the phosphate group from C-4 to the liberated OH group at C-3 to furnish compound **S2**. A partial loss of a benzyl protecting group at phosphorus leading to the phosphodiester **S3** was also observed. (SI-Table 3: Entry 1).

Supplementary SI-Table 3

SI-Table 3: Conditions for 3-*O*-deacetylation of **36** and **39** with aq. hydroxylamine.

	Com- pound	H ₂ NOH equiv.	Solvent	T °C	Time h	Product (Yield [%])			
1	39	725	THF	0	84	S1 (55)	S2 (7)	S3 (5)	39 (7)
2	39	160	CHCl ₃ -MeOH, 6:1	25	48	S1 (31)	n.d.	n.d.	n.d.
3	39	160	Pyr-AcOH, 2:1	25	72	decomposition			
4	39	8	DMF	0	22	decomposition			
5	36	725	THF	0	48	46 (53)	47 (2)	48 (9)	36 (15)
6	36	80	CH ₃ CN	25	24	46 (33)	47 (9)	48 (29)	36 (18)

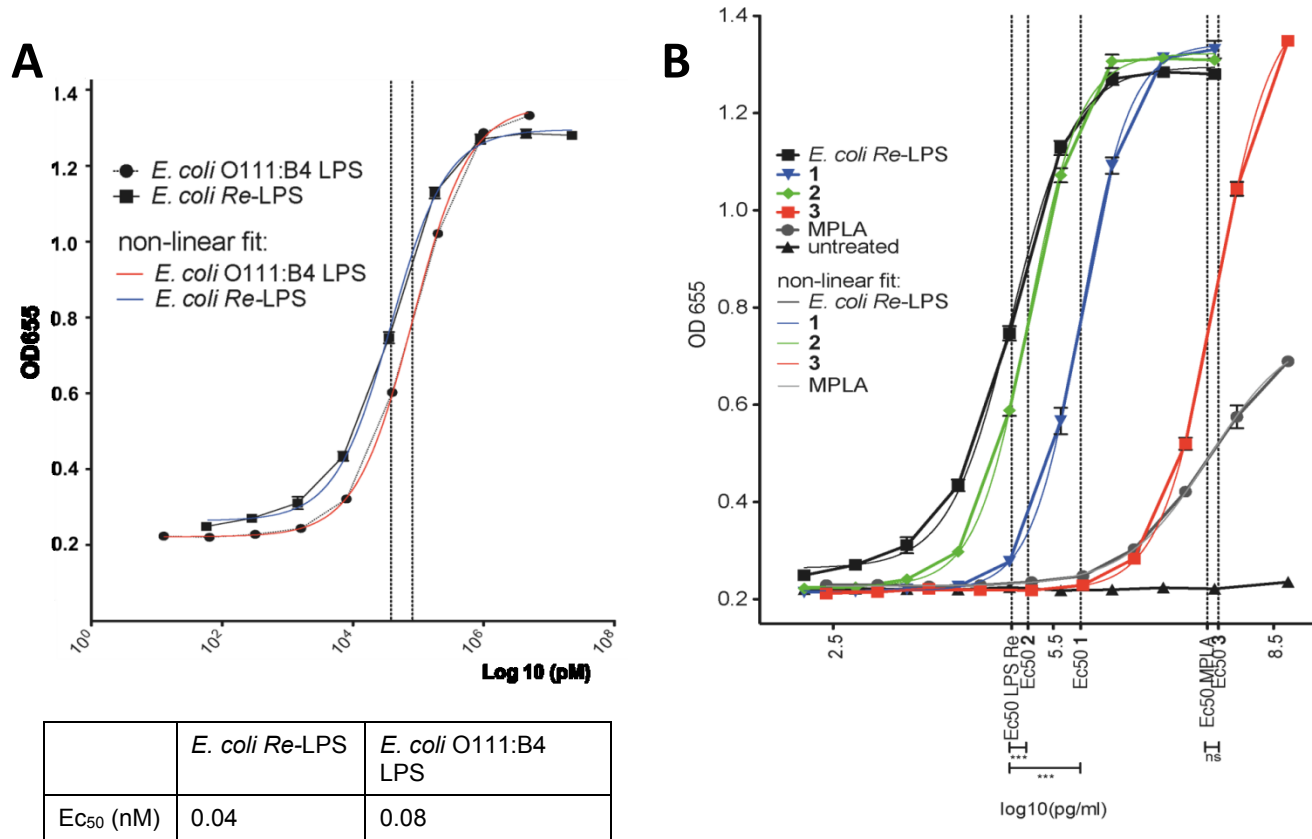
A mechanism for the partial debenzoylation of phosphotriesters upon treatment with NH₂OH was proposed by Nome.^{12,13} Accordingly, a nucleophilic attack of the OH group of hydroxylamine at the phosphorus atom results in the exchange of a benzyl ether for an hydroxyamino group. The amino group at the phosphate reacts further with another molecule of hydroxylamine and water to form monobenzylated phosphate diester and diimide, which disproportionates to N₂ and hydrazine.

Further optimization of reaction conditions by varying the amount of nucleophile (NH₂OH), reaction temperature and solvents did not result in any improvement (SI-Table 3: Entries 2-4). Thus, the conditions of Entry 1 in SI-Table 2 were applied to the deprotection of the key αGlcN(1↔1)αMan disaccharide **36** resulting in a successful isolation of the 3-*O*-deacetylated compound **46** as the main product (SI-Table 3: Entry 5). Minor amounts of concomitantly formed 3-*O*-phosphate **47** and partially debenzylated phosphodiester **48** were isolated as well. Upon application of acetonitrile as solvent the yield of the target compound **46** declined, whereas the proportion of debenzylated **48** increased (SI-Table 3: Entry 6).

Thus, the most high-yielding and reproducible conditions for the cleavage of 3-*O*-acetate in **36** entailed the stirring a 23 mM solution of **36** in THF with an equal volume of aq. hydroxylamine at 0 °C for 48 h. The reaction was stopped prior to completion, and the remaining starting material **36** was isolated and subjected to another reaction cycle. Accordingly, compound **46** was obtained in a 53% yield together with 2% of **47**, 9% of **48** and 15% of recovered **36** over three reaction cycles.

The same deprotection protocol was applied to the 3-*O*-deacetylation of the 2-*N*-Troc protected αGlcN(1↔1)αMan disaccharide **35**. The isolation of the target product was however much more tedious and complicated by the presence of the co-migrating by-products, so that this route was not pursued.

Supplementary SI-Figure 1

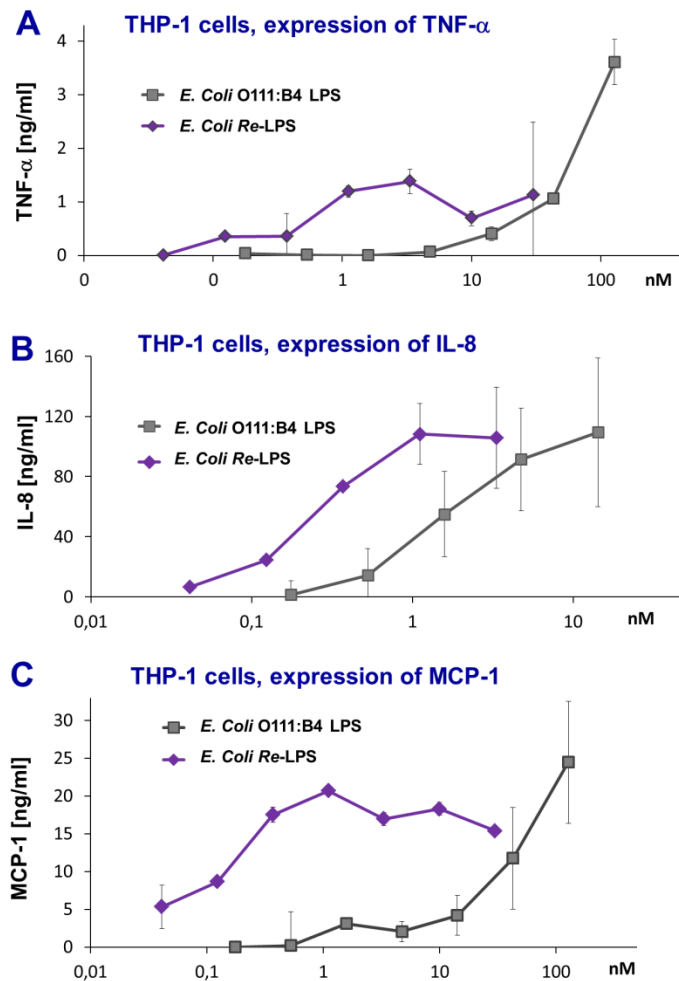


SI-Figure 1. (A) Dose-dependent activation of TLR4 signaling in hTLR4/hMD-2/hCD14 transfected HEK293 cells (HEK-Blue) by *E. coli* Re-LPS compared to *E. coli* O111:B4 LPS. [nM] concentrations were calculated from [ng/ml] concentrations based on the following molecular weights (MW): MW_{*E. coli* Re-LPS} 2339 Da, MW_{*E. coli* O111:B4 LPS} 10000 Da. **(B)** Dose-dependent activation of TLR4 signaling in hTLR4/hMD-2/hCD14 transfected HEK293 cells (HEK-Blue) by Lipid A mimetics 1-3 compared to *E. coli* Re-LPS and *S. minnesota* MPLA. Non-linear curve fit (variable slope) was calculated using GraphPad Prism and is shown for the mean curves. Dotted lines indicate corresponding EC₅₀ values. Statistical analysis was performed on non-linear curve fits on individual datasets using an un-paired two-tailed t-test (***) indicates a p value < 0.001, ns indicates non-significant change). Top, bottom, hill slope and EC₅₀ values of non-linear curve fit on individual datasets are shown.

	<i>E. coli</i> Re-LPS			1			2			3			SM-MPLA		
	n1	n2	n3	n1	n2	n3	n1	n2	n3	n1	n2	n3	n1	n2	n3
Bottom	0.262	0.276	0.253	0.212	0.225	0.215	0.216	0.232	0.236	0.217	0.221	0.216	0.217	0.236	0.229
Top	1.277	1.304	1.308	1.369	1.312	1.348	1.330	1.319	1.320	1.425	1.426	1.453	0.8932	0.7332	0.7252
HillSlope	1.115	1.082	0.8908	1.282	1.341	1.257	1.193	1.250	1.246	1.120	1.170	1.145	0.6005	0.8738	0.8395
Ec50 (ng/ml)	0.089	0.0868	0.082	0.894	0.623	0.745	0.154	0.144	0.132	51.5	47.62	57.65	111	35.04	29.9

Supplementary SI-Figure 2

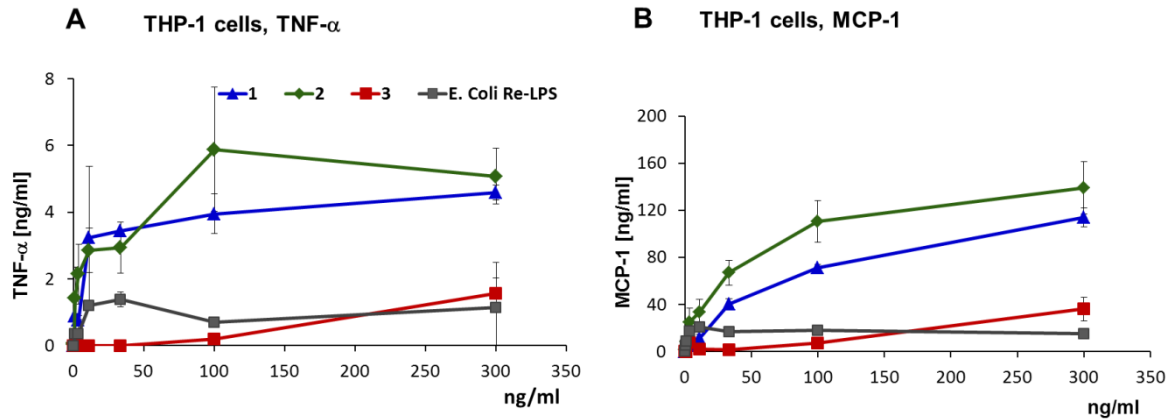
E. coli wild type O111:B4 LPS is a large heterogeneous molecule with molecular weight (MW) comprising 10-15 kDa. Since we were particularly interested in the molecular recognition mechanisms implicated in the binding of LPS by the MD-2·TLR4 complex wherein the Lipid A/Re-LPS (MW 1.8/2.2 kDa, respectively) portion of LPS is mostly involved, we have analyzed the dose-dependent THP-1 cells activation by synthetic α,α -GM-LAM 1-3 (at the concentrations 0.1 – 300 ng/ml) compared to *E. coli* Re-LPS which is more appropriate in this instance. Since the divergences in the expression of different cytokines by *E. coli* LPS vs. Re-LPS are only scarcely reported in the literature, we have performed dose-dependent activation experiments comparing expression of TNF- α , IL-8 and MCP-1 by *E. coli* O111:B4 LPS and *E. coli* Re-LPS at the concentrations 0.4 – 100 nM. Wild-type LPS (MW 10 kDa) induced the release of somewhat lower levels of pro-inflammatory cytokines at the nanomolar concentration range (0.4 – 10 nM), whereas at the concentrations above 10 nM (50 nM for MCP-1) the levels of cytokines production did not differ significantly. This might be explained by a different aggregation state of Re-LPS and wild type LPS, and, therefore, the alterations in the transfer of a single LPS molecule to TLR4·MD-2 complex by LBP and CD-14. Higher MW (> 10 kDa) of the LPS molecule applied for the calculations of the concentrations in [nM] would result in the shift of the activation profile induced by wild-type LPS. Calculation of EC₅₀ values were hardly possible, since the maximum TLR4 saturation levels were not achieved at the concentrations tested.



SI-Figure 2. Dose-dependent expression of pro-inflammatory cytokines TNF- α , IL-8 and MCP-1 in human macrophage cell line THP-1 by *E. coli* Re-LPS compared to *E. coli* wild-type O111:B4 LPS. (A) Comparative expression of TNF- α ; (B) comparative expression of IL-8; (C) comparative expression of MCP-1

Supplementary SI-Figure 3

Dose-dependent responses (THP-1 cells: α,α -GM-LAM **1-3** and *Re*-LPS) at the concentration range 0.4 – 100 ng/ml which are presented in the **Figure 5**, were extended to the higher concentration range (up to 300 ng/ml).

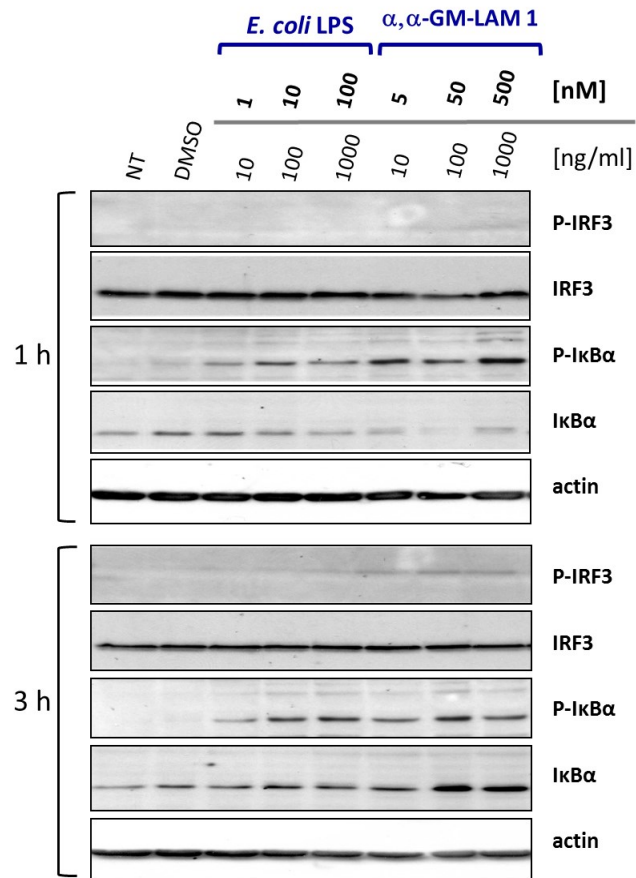


SI-Figure 3. Dose-dependent expression of cytokines induced by α,α -GM-LAM **1-3** in macrophage cell line THP-1 compared to *E. coli* *Re*-LPS at the concentration range 0.4 – 300 ng/ml. (A) Production of TNF- α ; (B) Induction of the expression of MCP-1.

Supplementary SI-Figure 4

Lipid A mimetic **1** was examined for the activation of NF- κ B (Nuclear factor kappaB) and IRF3 (Interferon regulatory factor 3). IRF3 activation (revealed by IRF-3 phosphorylation) was not noticeable after 1h stimulation, though was detectable after 3h stimulation with **1**, but not with LPS. At 1h stimulation the α,α -GM-LAM **1** was comparable with LPS in the activation NF- κ B, whereas at 3h stimulation compound **1** (at a concentration higher than 50 nM) was more potent activator of NF- κ B than LPS as revealed by stronger phosphorylation of I κ B α (P-I κ B α levels) as well as the higher levels of I κ B α (reflecting the NF- κ B dependent re-synthesis of I κ B following its degradation after 3h).

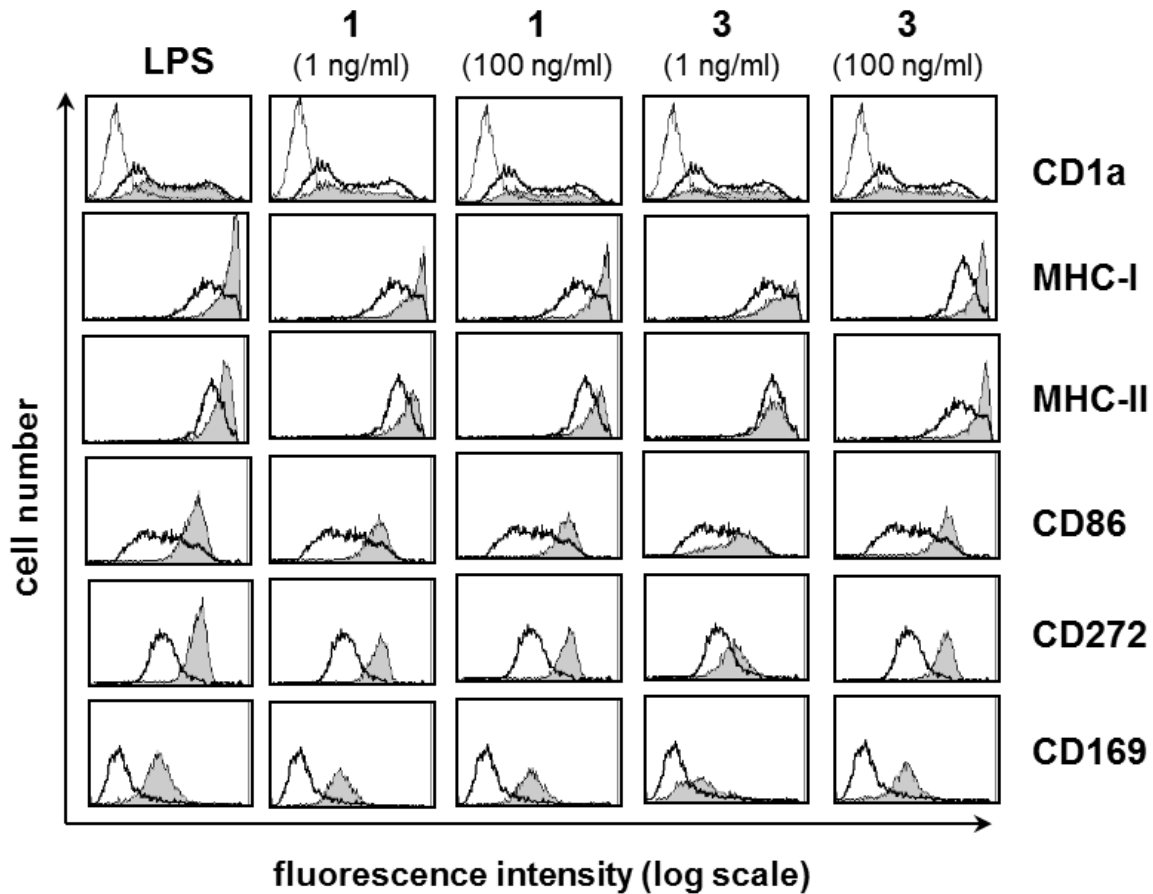
Thus, IRF3 activation and NF- κ B activation (revealed by I κ B α phosphorylation, I κ B α degradation and its NF- κ B dependent resynthesis) were more strongly induced by compound **1** compared to LPS after 3h stimulation.



SI-Figure 4. NF- κ B and IRF-3 activation by LPS and α,α -GM-LAM **1** in macrophage-like cell line THP-1. THP-1 cells were treated for 1 h (upper panel) or for 3 h (lower panel) with **1** or with *E. coli* O111:B4 LPS at the concentrations 10, 100 and 1000 ng/ml. Non treated cells (NT) or cells treated with DMSO were used as controls. Cell extracts were separated by SDS polyacrylamide gel electrophoresis. Phosphorylated or total IRF-3 and I κ B α was detected by western blotting as indicated. Actin detection was used as a control for equal loading in each line.

Supplementary SI-Figure 5

Stimulation of DCs with **1** (1 mg/ml) was as potent as by LPS in inducing DCs maturation, up-regulation of the co-stimulatory molecules CD86 as well as the antigen presenting structures MHC class I and MHC class II, which are necessary for the induction of an adaptive immune response (SI-Figure 5). Similar to LPS, α,α -GM-LAM **1** induced up-regulation of the inhibitory B7-H1 (CD272) and Sialoadhesin (CD169), which is known to be indirectly up-regulated in DCs due to release of type-I interferons. Thus, our data demonstrate that α,α -GM-LAM **1** is a potent stimulator not only of the NF- κ B- but also of the IRF-pathway. Stimulation of DCs with the monophosphoryl α,α -GM-LAM **3** resulted in a less pronounced activation marker profile.

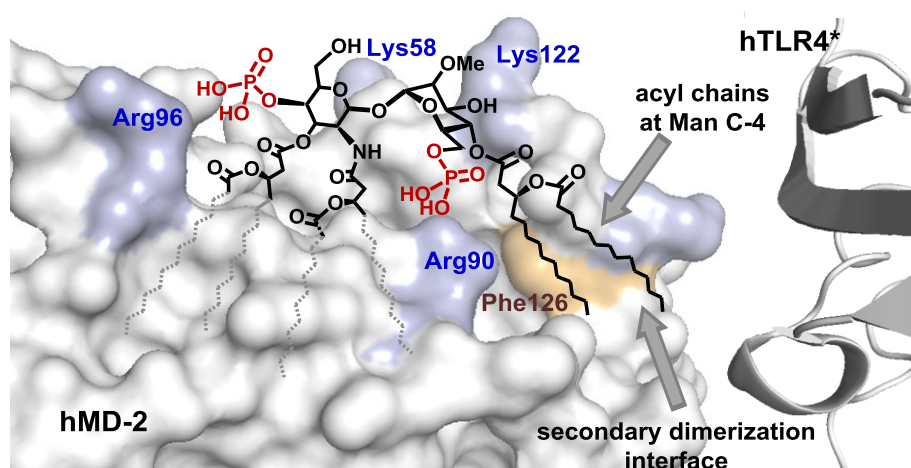


SI-Figure 5. Bisphosphoryl α,α -GM-LAM **1** and monophosphoryl α,α -GM-LAM **3** induce the expression of maturation markers in human DCs. Human peripheral blood monocytes were cultured for 6 days in GM-CSF and IL-4 to receive immature monocyte-derived DCs (*open histograms, bold line*), and were stimulated in parallel for the last 24hs of culture with **1**, **3** or LPS (10 ng/ml) (*gray histograms*). The cells were then harvested and the surface expression level of the indicated markers was measured by flow cytometry. The thin line in the CD1a histograms represents VIAP staining (isotype control). Results are representative of three (for compound **1**) and two (for compound **3**) independent experiments.

Supplementary SI-Figure 6

The relative contribution of hydrophobic (acyl chains of Lipid A) and electrostatic (phosphate groups of Lipid A) interactions at the dimerization interface of two TLR4·MD-2·LPS complexes as well as the impact of the molecular shape of MD-2-bound Lipid A on the initiation of TLR4-mediated immune response remains largely unknown. Assuming that the two acyl lipid chains at Man C-4 of α,α -GM-LAM 1 are not included into the hydrophobic cluster formed by the four long-chain acyl residues attached to the GlcN moiety, the four lipid tails linked to GlcN unit should be fully inserted into the hydrophobic binding pocket of hMD-2 whereas the two acyl chains at Man C-4 are presumed to be exposed on the surface of the protein (SI-Figure 6). Thus, α,α -GM-LAM 1-3 should be able bridging two TLR4·MD-2 complexes by exposure of two lipid chains at Man C-4 on the surface of MD-2 thereby forming an effective hydrophobic homodimerization interface. The phosphate group at Man C-6 in α,α -GM-LAMs 1 and 2 could be either involved in the interaction with the second TLR4*·MD-2*-ligand complex or, alternatively, support the binding of the ligand by MD-2 *via* establishing ionic interaction with Arg90 at the rim of the binding groove of the co-receptor protein. According to our experimental data, the presence of the phosphate group at Man C-6 in α,α -GM-LAM 1 and 2 significantly enhances the TLR-4 stimulating activity compared to the monophosphate 3. This might indicate both a stronger binding of the diphosphates 1 and 2 to hMD-2 due to the formation of salt bridges with positively charged Lys and Arg at the rim of the binding groove of hMD-2 and an involvement of the phosphate group at Man C-6 in the dimerization interface of two TLR4·MD-2· α,α -GM-LAM complexes.

Proposed interaction of Lipid A mimetic 1 with hMD-2·TLR4 complex



SI-Figure 6. Proposed orientation of Lipid A mimetic 1 within the binding pocket of hMD-2 (hMD-2 from PDB code: 3FXI). Acyl chains at Man C-4 which are exposed on the surface of hMD-2 should stabilize the inward positioning of Phe126 and support hydrophobic interactions at the dimerization interface with the second TLR4*·MD-2*· α,α -GM-LAM 1 complex. Image was generated with PyMol and ChemDraw.

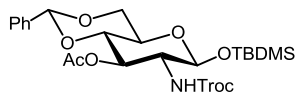
Experimental Procedures: Biological assays

Detection of IRF-3 and I κ B α phosphorylation by western blotting.

THP1 monocytic cells were seeded at (2.75 x 10⁶) cells/well in a 6-well plate in 2 ml culture medium and differentiated into macrophages by treatment with 200 nM TPA for 24 h. Next, the cells were washed twice with complete RPMI-1640 medium to remove the cells that did not adhere, refreshed with 2 ml medium and left for 1 h to recover. Cells were stimulated with 10 ng/ml, 100 ng/ml or 1000 ng/ml *E. coli* O111:B4 LPS or α,α -GM-LAM 1 which was added in 200 μ l complete RPMI-1640 medium (total volume 2.2 ml/well). The cells were stimulated for 1 h and 3 h at 37°C after which cells were washed with PBS and lysed in E1A buffer (50 mM Hepes pH 7.6, 250 mM NaCl, 5 mM EDTA and 0.5% NP-40) supplemented with protease inhibitors (2.1 mM leupeptine, 0.15 mM aprotinine, and 1 mM pefabloc) and phosphatase inhibitors (200 mM sodium orthovanadate, 10 mM sodium fluoride and 5 mg/ml β -glycerophosphate). Cell extracts were centrifuged at 14000 r.p.m. for 15 min in an Eppendorf centrifuge and 60 μ g of the soluble fraction was separated by SDS polyacrylamide gel electrophoresis, followed by western blotting. Phosphorylated and total IRF-3 or I κ B α was detected by incubating the blots with the corresponding antibodies, followed by secondary antibodies conjugated with HRP and detection by chemiluminescence. The following antibodies were used: anti-phosphoSer396-IRF3 (4D4G) and anti-phosphoSer32/36-I κ B α (5A5) (Cell Signaling Technology), anti-I κ B α (C-21; Sc-371) and anti-IRF3 (FL-425; Sc-9082) (Santa Cruz), anti-actin (MP Biomedicals). A secondary mouse or rabbit antibody conjugated with HRP was obtained from Amersham, secondary goat antibody conjugated with HRP was obtained from Santa Cruz.

Experimental Procedures: Synthesis

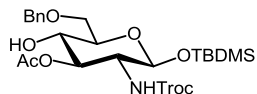
Synthesis of 5



tert-Butyldimethylsilyl 3-O-acetyl-4,6-O-benzylidene-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)-β-D-glucopyranoside (5).

To a stirred solution of **4**¹⁴ (4.50 g, 8.08 mmol) and DMAP (10 mg, 82 μmol) in pyridine (70 mL) was added acetic acid anhydride (3.00 mL, 31.9 mmol) at rt. The mixture was stirred for 3 h, then diluted with EtOAc (200 mL) and washed with aq. HCl (2 M, 500 mL), sat. aq. NaHCO₃ (250 mL), water (250 mL) and brine (250 mL). The organic layer was dried over Na₂SO₄, filtered and concentrated to afford crude **5** (4.71 g, 7.86 mmol, 97%) as white solid. For analysis 540 mg of crude **5** was purified by MPLC (toluene → toluene - EtOAc, 20 : 1) to give 400 mg of **5** as white amorphous solid. $R_f = 0.43$ (toluene - EtOAc, 10 : 1); $[\alpha]_D^{20} = -35$ (c 0.8, CHCl₃); ¹H-NMR (400 MHz, CDCl₃) δ 7.46-7.43 (m, 2H, Ph), 7.40-7.35 (m, 3H, Ph), 5.51 (s, 1H, CHPh), 5.27 (t, 1H, ³J_{3,2} = ³J_{3,4} = 9.9 Hz, H-3), 5.08 (d, 1H, ³J_{NH,2} = 9.5 Hz, NH), 4.81 (AB, 1H, ²J = 12.5 Hz, CH₂, Troc), 4.80 (d, 1H, ³J_{1,2} = 7.6 Hz, H-1), 4.61 (AB, 1H, ²J = 11.8 Hz, CH₂, Troc), 4.32 (dd, 1H, ³J_{6a,5} = 5.0 Hz, ²J_{6a,6b} = 10.5 Hz, H-6a), 3.82 (t, 1H, ³J_{6b,5} = ²J_{6b,6a} = 10.3 Hz, H-6b), 3.76-3.68 (m, 1H, H-2), 3.72 (t, 1H, ³J_{4,3} = ³J_{4,5} = 9.4 Hz, H-4), 3.52 (ddd, 1H, ³J_{5,4} = ³J_{5,6b} = 9.7 Hz, ³J_{5,6a} = 4.9 Hz, H-5), 2.07 (s, 3H, CH₃, Ac), 0.89 (s, 9H, 3×CH₃, TBDMS), 0.13 (s, 3H, CH₃, TBDMS), 0.11 (s, 3H, CH₃, TBDMS); ¹³C-NMR (101 MHz, CDCl₃) δ 170.94 (CO, Ac), 154.21 (CO, Troc), 136.93 (Cq, CHPh), 129.12, 128.23, 126.17 (5×CH, CHPh), 101.47 (CHPh), 97.13 (C-1), 78.77 (C-4), 74.63 (CH₂, Troc), 71.17 (C-3), 68.60 (C-6), 66.60 (C-5), 58.89 (C-2), 25.49 (3×CH₃, TBDMS), 20.82 (CH₃, Ac), 17.85 (Cq, TBDMS), -4.21, -5.33 (2×CH₃, TBDMS); HRMS (ESI): m/z calcd for C₂₄H₃₄Cl₃NO₈Si-H⁻: 596.1047 [M-H]⁻; found 596.1042.

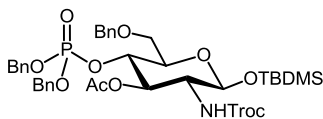
Synthesis of 6



tert-Butyldimethylsilyl 3-O-acetyl-6-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)-β-D-glucopyranoside (6).

A mixture of **5** (3.50 g, 5.84 mmol) and activated powdered 4 Å molecular sieves in dry CH₂Cl₂ (70 mL) was stirred at r.t. for 1.5 h under atmosphere of Ar. The mixture was then cooled to -78 °C and triethylsilane (1.70 mL, 10.7 mmol) and triflic acid (1.70 mL, 12.0 mmol) were added successively. After stirring for 2 h, the reaction was quenched by successive addition of MeOH (3 mL) and Et₃N (3 mL). The mixture was warmed to r.t., diluted with EtOAc (250 mL) and filtered over a pad of Celite. The filtrate was washed with aq. HCl (2 M, 100 mL), sat. aq. NaHCO₃ (200 mL), water (200 mL) and brine (200 mL). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by MPLC (toluene - EtOAc, 5 : 1) to afford compound **6** (2.40 g, 3.99 mmol, 68%) as white amorphous solid. $R_f = 0.35$ (toluene - EtOAc, 2 : 1); $[\alpha]_D^{20} = -26$ (c 0.9, CHCl₃); ¹H-NMR (400 MHz, CDCl₃): δ 7.37-7.27 (m, 5H, CH₂Ph), 5.02 (dd, 1H, ³J = 10.9 Hz, ³J = 9.1 Hz, H-3), 5.05-5.00 (m, 1H, NH), 4.77 (AB, 1H, ²J = 11.6 Hz, CH₂, Troc), 4.70 (d, 1H, ³J_{1,2} = 7.9 Hz, H-1), 4.61 (AB, 1H, CH₂, Troc), 4.61 (AB, 1H, ²J = 12.1 Hz, CH₂Ph), 4.57 (AB, 1H, ²J = 11.9 Hz, CH₂Ph), 3.82-3.73 (m, 3H, H-4, H-6a, H-6b), 3.64-3.57 (m, 1H, H-2), 3.57-3.51 (m, 1H, H-5), 2.10 (s, 3H, CH₃, Ac), 0.88 (s, 9H, 3×CH₃, TBDMS), 0.12 (s, 3H, CH₃, TBDMS), 0.09 (s, 3H, CH₃, TBDMS); ¹³C-NMR (101 MHz, CDCl₃): δ 171.82 (CO, Ac), 154.18 (CO, Troc), 137.56 (Cq, CH₂Ph), 128.48, 127.88, 127.66 (5×CH, CH₂Ph), 96.46 (C-1), 95.38 (C-1, Troc), 74.88 (C-3), 74.57 (CH₂, Troc), 73.95 (C-5), 73.80 (CH₂Ph), 71.14 (C-4), 70.55 (C-6), 57.89 (C-2), 25.53 (3×CH₃, TBDMS), 20.91 (CH₃, Ac), 17.88 (Cq, TBDMS), -4.17, -5.31 (2×CH₃, TBDMS); HRMS (ESI): m/z calcd for C₂₄H₃₆Cl₃NO₈Si-H⁻: 598.1203 [M-H]⁻; found 598.1202.

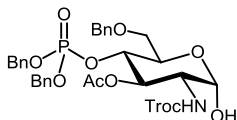
Synthesis of 7



tert-Butyldimethylsilyl 3-*O*-acetyl-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)-β-*D*-glucopyranoside (7).

To a solution of **6** (2.40 g, 3.99 mmol) in dry CH₂Cl₂ (40 mL) were added dibenzyl *N,N*-diisopropylphosphoramidite (2.00 mL, 5.36 mmol, 90%) and a solution of 1*H*-tetrazole in CH₃CN (17.7 mL, 5.99 mmol, 0.45 M in CH₃CN) at r.t. under atmosphere of Ar. After 30 min the mixture was cooled to -78 °C and *m*CPBA (2 g, 8.11 mmol, 70% in 40 ml CH₂Cl₂) was added. After stirring for 1 h, the reaction was quenched by addition of Et₃N (1.3 ml) and it was warmed to rt. The mixture was diluted with EtOAc (300 ml) and washed with sat. aq. NaHCO₃ (300 ml) and brine (300 ml). The organic layer was dried over MgSO₄, filtered and concentrated. The residue was purified by MPLC (toluene - EtOAc, 4 : 1) to afford **7** (3.10 g, 3.60 mmol, 90%) as white amorphous solid. *R*_f = 0.29 (toluene : EtOAc = 5 : 1); [α]_D²⁰ = -1 (*c* 1.0, CHCl₃); ¹H-NMR (400 MHz, CDCl₃) δ 7.36-7.22 (m, 15H, CH₂Ph), 5.26 (dd, 1H, ³*J* = 10.8 Hz, ³*J* = 9.1 Hz, H-3), 5.02-4.98 (m, 1H, 2-NH), 4.98-4.88 (m, 4H, OP(O)(OCH₂Ph)₂), 4.76 (d, 1H, ³*J*_{1,2} = 7.8 Hz, H-1), 4.74 (AB, 1H, ²*J* = 11.7 Hz, CH₂, Troc), 4.62 (AB, 1H, ²*J* = 11.7 Hz, CH₂, Troc), 4.52 (AB, 1H, ²*J* = 12.2 Hz, CH₂Ph), 4.51-4.47 (m, 1H, H-4), 4.47 (AB, 1H, ²*J* = 12.3 Hz, CH₂Ph), 3.79-3.75 (m, 1H, H-6a), 3.65-3.55 (m, 3H, H-2, H-5, H-6b), 1.89 (s, 3H, CH₃, Ac), 0.87 (s, 9H, 3×CH₃, TBDMS), 0.13 (s, 3H, CH₃, TBDMS), 0.09 (s, 3H, CH₃, TBDMS); ¹³C-NMR (101 MHz, CDCl₃) δ 171.08 (CO, Ac), 153.98 (CO, Troc), 138.11 (C_q, CH₂Ph), 135.56 (C_q, ³*J*_{C,P} = 7 Hz, OP(O)(OCH₂Ph)₂), 135.46 (C_q, ³*J*_{C,P} = 8 Hz, OP(O)(OCH₂Ph)₂), 128.61, 128.59, 128.28, 128.00, 127.95, 127.49, 127.47 (15×CH, CH₂Ph), 96.17 (C-1), 74.56 (CH₂, Troc), 74.33 (C-4 or C-5, *J*_{C,P} = 5.8 Hz), 74.05 (C-4 or C-5, *J*_{C,P} = 6.1 Hz), 73.39 (CH₂Ph), 72.39 (C-3), 69.64, 69.61, 69.58, 69.56 (2×CH₂, OP(O)(OCH₂Ph)₂), 68.66 (C-6), 58.28 (C-2), 25.53 (3×CH₃, TBDMS), 20.68 (CH₃, Ac), 17.87 (C_q, TBDMS), -4.16, -5.30 (2×CH₃, TBDMS); ³¹P-NMR (162 MHz, CDCl₃): δ -2.16; HRMS (ESI): *m/z* calcd for C₃₈H₄₉Cl₃NO₁₁PSi-H⁺: 858.1805 [M-H]⁻; found 858.1807.

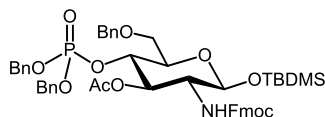
Synthesis of 8



3-*O*-Acetyl-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(2,2,2-trichloroethoxy-carbonylamino)-*D*-glucopyranose (8).

To a solution of **7** (1.50 g, 1.74 mmol) in dry THF (20 ml) in a PTFE-vial was a solution of HF·Py in Pyridine (2.5 ml, 70% HF) was added at 0 °C under atmosphere of Ar. After 10 min the mixture was warmed to rt and stirred for 12 h. The solution was diluted with EtOAc (200 ml) and washed with sat. aq. NaHCO₃ (2×30 ml), water (30 ml) and brine (30 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by MPLC (CH₂Cl₂ - MeOH, 50 : 1 → 25 : 1) to afford **8** (1.19 g, 1.59 mmol, 91%, α : β = 9 : 1) as white amorphous solid. *R*_f = 0.37 (hexane - EtOAc, 1 : 1); [α]_D²⁰ = +20 (*c* 1.0, CHCl₃); ¹H-NMR (300 MHz, CDCl₃ + 1% MeOD): δ 7.35-7.24 (m, 15H, CH₂Ph), 5.38 (dd, 1H, ³*J*_{3,2} = 10.7 Hz, ³*J*_{3,4} = 9.1 Hz, H-3), 5.21 (d, 1H, ³*J*_{1,2} = 3.5 Hz, H-1), 4.98-4.83 (m, 4H, 2×CH₂, OP(O)(OCH₂Ph)₂), 4.78 (AB, 1H, ²*J* = 12.0 Hz, CH₂, Troc), 4.62 (AB, 1H, ²*J* = 12.0 Hz, CH₂, Troc), 4.51 (AB, 1H, ²*J* = 11.8 Hz, CH₂Ph), 4.43 (AB, 1H, ²*J* = 11.7 Hz, CH₂Ph), 4.42-4.36 (m, 1H, H-4), 4.18 (ddd, 1H, ³*J*_{5,4} = 10.0 Hz, ³*J*_{5,6a} = 1.8 Hz, ³*J*_{5,6b} = 6.5 Hz, H-5), 3.94 (dd, 1H, ³*J*_{2,1} = 3.6 Hz, ³*J*_{2,3} = 10.7 Hz, H-2), 3.77 (dd, 1H, ³*J*_{6a,5} = 1.9 Hz, ²*J*_{6a,6b} = 10.6 Hz, H-6a), 3.61 (dd, 1H, ³*J*_{6b,5} = 6.5 Hz, ²*J*_{6b,6a} = 10.6 Hz, H-6b), 1.85 (s, 3H, CH₃, Ac); ¹³C-NMR (75 MHz, CDCl₃): δ 171.18 (CO, Ac), 154.14 (CO, Troc), 137.50 (C_q, CH₂Ph), 135.38 (2×C_q, OP(O)(OCH₂Ph)₂), 128.73, 128.68, 128.66, 128.60, 128.39, 128.04, 127.99, 127.90, 127.84 (15×CH, CH₂Ph), 95.33 (CCl₃, Troc), 91.47 (C-1), 74.51 (CH₂, Troc), 73.95 (C-4, ²*J*_{C4,P} = 6.2 Hz), 73.47 (CH₂, CH₂Ph), 70.96 (C-3, ³*J*_{C3,P} = 6.2 Hz), 69.65 (CH₂, ²*J*_{C,P} = 5.9 Hz, OP(O)(OCH₂Ph)₂), 69.56 (CH₂, ²*J*_{C,P} = 5.9 Hz, OP(O)(OCH₂Ph)₂), 69.31 (C-5, ³*J*_{C5,P} = 5.8 Hz), 68.57 (C-6), 54.15 (C-2), 20.69 (CH₃, Ac), ³¹P-NMR (162 MHz, CDCl₃): δ -2.21; HRMS (ESI): *m/z* calcd for C₃₂H₃₅Cl₃NO₁₁P+H⁺: 746.1089 [M+H]⁺; found 746.1086.

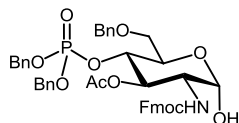
Synthesis of 9



tert-Butyldimethylsilyl 3-O-acetyl-6-O-benzyl-4-O-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)-β-D-glucopyranoside (9).

To a solution of **8** (645 mg, 749 μmol) in dry CH₂Cl₂ (40 ml), Zn dust (2.4 g) and acetic acid (18 ml) were added at 0 °C under atmosphere of Ar. The mixture was sonicated for 1.5 h, while the temperature of the reaction mixture was kept in the range of 5 °C – 20 °C. The mixture was diluted with CH₂Cl₂ (200 ml), filtered over a pad of Celite, the filtrate was washed with sat. aq. NaHCO₃ (2×200 ml) and brine (100 ml). The organic phase was dried over Na₂SO₄, filtered and concentrated to afford the intermediate amine (513 mg) as a pale yellow syrup. The crude amine was dissolved in dry CH₂Cl₂ (6 ml) and diisopropylethylamine (150 μL, 908 μmol) and a solution of 9-fluorenylmethyl chloroformate (255 mg, 986 μmol) in dry CH₂Cl₂ (1 ml) were added at 0 °C. The mixture was stirred for 2 h. at r. t., diluted with CH₂Cl₂ (80 ml) and washed with brine (100 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by MPLC (toluene – EtOAc, 5 : 1) to afford **9** (608 mg, 670 μmol, 89%, over 2 steps) as amorphous solid. *R_f* = 0.47 (toluene - EtOAc, 5 : 3); [α]_D²⁰ = -2 (c 1.1, CHCl₃); ¹H-NMR (300 MHz, DMSO-d₆): δ 7.90-7.87 (m, 2H, arom.), 7.66-7.64 (m, 2H, arom.), 7.46-7.22 (m, 19H, arom.), 5.19 (t, 1H, ³J_{2,3} = ³J_{3,4} = 9.8 Hz, H-3), 4.98-4.92 (m, 4H, OP(O)(OCH₂Ph)₂), 4.80 (d, 1H, ³J_{1,2} = 7.9 Hz, H-1), 4.47-4.28 (m, 4H, H-6, CH₂Ph, CH₂, Fmoc), 4.20-4.13 (m, 2H, CH₂, CH, Fmoc), 3.71-3.65 (m, 2H, H-5, H-6a), 3.56 (dd, 1H, *J* = 5.8 Hz, *J* = 11.6 Hz, H-6b), 3.44-3.34 (m, 2H, H-2, NH), 1.80 (s, 3H, CH₃, Ac), 0.78 (s, 9H, 3×CH₃, TBDMS), 0.06 (s, 3H, CH₃, TBDMS), 0.03 (s, 3H, CH₃, TBDMS); ¹³C-NMR (75 MHz, DMSO-d₆): δ 169.76 (CO, Ac), 155.88 (CO, Fmoc), 143.80, 143.75, 140.73, 140.71 (4×Cq, Fmoc), 138.28 (Cq, CH₂Ph), 135.88 (Cq, ³J_{C,P} = 7.0 Hz, OP(O)(OCH₂Ph)₂), 135.79 (Cq, ³J_{C,P} = 7.2 Hz, OP(O)(OCH₂Ph)₂), 128.51, 128.17, 127.93, 127.86, 127.65, 127.37, 127.23, 127.07, 125.21, 125.10, 120.15 (23×CH, arom.), 95.56 (C-1), 73.95, 73.17 (C-4, C-5, *J*_{C,P} = 5.8 Hz, *J*_{C,P} = 6.0 Hz), 72.80 (C-3), 72.26 (CH₂Ph), 68.90, 68.83 (2×CH₂, OP(O)(OCH₂Ph)₂), 68.38 (C-6), 65.60 (CH₂, Fmoc), 57.30 (C-2), 46.56 (CH, Fmoc), 25.44 (3×CH₃, TBDMS), 20.67 (Cq, TBDMS), 17.55 (CH₃, Ac), -4.34, -5.30 (2×CH₃, TBDMS); ³¹P-NMR (122 MHz, DMSO-d₆) δ -1.96; HRMS (ESI): *m/z* calcd for C₅₀H₅₈NO₁₁PSi+COOH⁺: 952.3499 [M+COOH⁺]; found 952.3501.

Synthesis of 10

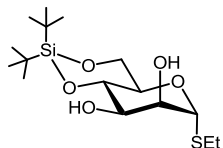


3-O-Acetyl-6-O-benzyl-4-O-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonyl)-amino)-D-glucopyranose (10).

To a solution of **9** (539 mg, 594 μmol) in dry THF (6 ml) in a PTFE-vial a solution of HF•Py (400 μL, 70% HF) was added at 0 °C under atmosphere of Ar. After 10 min the mixture was warmed to r. t. and stirred for 15 h. The solution was diluted with CH₂Cl₂ (200 ml), washed with sat. aq. NaHCO₃ (100 ml) and brine (100 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by MPLC (toluene - EtOAc, 1 : 1) to afford **10** (448 mg, 564 μmol, 94%, α/β = 9 : 1) as amorphous solid. *R_f* = 0.33 and 0.21 (toluene – EtOAc, 1 : 1); [α]_D²⁰ = +23 (c 1.4, CHCl₃); ¹H-NMR (300 MHz, CDCl₃, α-anomer): δ 7.74-7.72 (m, 2H, arom.), 7.56-7.51 (m, 2H, arom.), 7.39-7.20 (m, 19H, arom.), 5.38 (dd, 1H, ³J = 10.7 Hz, ³J = 9.2 Hz, H-3), 5.22 (t, 1H, ³J_{1,2} = ³J_{1,OH} = 3.3 Hz, H-1), 5.17 (d, 1H, ³J_{NH,2} = 9.8 Hz, NH), 4.93-4.84 (m, 4H, OP(O)(OCH₂Ph)₂), 4.54-4.35 (m, 5H, 1-OH, H-4, CH₂Ph, CH₂ Fmoc), 4.23-4.13 (m, 3H, H-5, CH₂, CH Fmoc) 3.95 (td, 1H, ³J_{1,2} = 2.6 Hz, ³J_{2,3} = ³J_{2,NH} = 10.3 Hz, H-2), 3.73-3.69 (m, 1H, H-6a), 3.60 (dd, 1H, *J* = 10.7 Hz, *J* = 5.9 Hz, H-6b), 1.83 (s, 3H, CH₃, Ac); ¹³C-NMR (75 MHz, CDCl₃, α-anomer): δ 171.29 (CO, Ac), 155.83 (CO, Fmoc), 143.73, 141.65, 141.23, 141.18 (4×Cq, Fmoc), 137.54 (Cq, CH₂Ph), 135.46 (Cq, ³J_{C,P} = 7.0 Hz, OP(O)(OCH₂Ph)₂) 135.36 (Cq, ³J_{C,P} = 7.1 Hz, OP(O)(OCH₂Ph)₂), 128.66, 128.58, 128.54, 128.32, 127.98, 127.93, 127.83, 127.73, 127.66, 127.04, 125.02, 119.93, (23×CH, arom.), 91.54 (C-1), 74.12 (C-4, ²J_{C4,P} = 6.3 Hz), 73.40 (CH₂Ph), 71.15 (C-3), 69.59 (OP(O)(OCH₂Ph)₂, ²J_{C,P} = 5.8 Hz), 69.51

(OP(O)(OCH₂Ph)₂, ²J_{C,P} = 6.1 Hz), 69.23 (C-5, ³J_{C5,P} = 6.1 Hz), 68.60 (C-6), 67.06 (CH₂, Fmoc), 54.06 (C-2), 47.00 (CH, Fmoc), 20.71 (CH₃, Ac); ³¹P-NMR (122 MHz, CDCl₃): δ -2.30; HRMS (ESI): *m/z* calcd for C₄₄H₄₄NO₁₁P+Na⁺: 816.2544 [M+Na⁺]; found 816.2543.

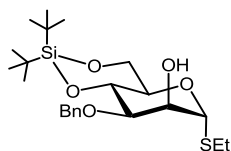
Synthesis of 12



Ethyl 4,6-*O*-di-*tert*-butylsilylene-1-thio- α -D-mannopyranoside (12).

To a solution of **11**^{15,16} (19.5 g, 49.7 mmol, α : β = 6 : 1) in MeOH (260 ml) a solution of NaOMe in MeOH (0.25 M, 4.7 ml) was added at r. t. After stirring for 2 h the pH of the solution was adjusted to 7 by addition of Dowex 50 (H⁺) cation-exchange resin. The resin was removed by filtration and the filtrate was concentrated to afford crude tetraol^[2] (10.9 g) as a syrup. A solution of the tetraol (10.9 g) and pyridine (12 ml, 148.4 mmol) in dry DMF (120 ml) was cooled to -35 °C and (*t*Bu)₂Si(OTf)₂ (17 ml, 52.47 mmol) was added dropwise over a period of 15 min under atmosphere of Ar. The mixture was stirred at -35 °C for 60 min and then the reaction was quenched by addition of MeOH (3 ml). The mixture was warmed to r. t., diluted with EtOAc (300 ml) and washed with sat. aq. NaHCO₃ (3×150 ml) and brine (150 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was co-evaporated with toluene (2×100 ml) and the crude product was purified by crystallization from Et₂O (3.5 ml×g⁻¹) to afford **12** (16.2 g, 44.4 mmol, 89% over 2 steps, α / β = 5 : 1) as white solid. *R*_f = 0.47 (toluene - EtOAc, 1 : 1); m.p. = 145 °C (Et₂O); [α]_D²⁰ = +168.2 (*c* 1.1, CHCl₃); ¹H-NMR (300 MHz, CDCl₃, α -anomer): δ 5.30 (d, 1H, ³J_{1,2} = 1.1 Hz, H-1), 4.18-3.94 (m, 5H, H-2, H-4, H-5, H-6a, H-6b), 3.81 (dd, 1H, *J* = 3.4 Hz, *J* = 8.8 Hz, H-3), 2.76-2.52 (m, 2H, CH₂, SEt), 1.29 (t, 3H, *J* = 7.4 Hz, CH₃, SEt), 1.06 (s, 9H, 3×CH₃, DTBS), 1.01 (s, 9H, 3×CH₃, DTBS). ¹³C-NMR (75 MHz, CDCl₃, α -anomer): δ 84.02 (C-1), 74.98 (C-2 or C-4), 72.28 (C-3), 71.96 (C-2 or C-4), 66.96 (C-5), 66.20 (C-6), 27.44, 27.01 (6×CH₃, DTBS), 25.17 (CH₂, SEt), 22.63, 20.00 (2×C_q, DTBS), 14.88 (CH₃, SEt); HRMS (ESI): *m/z* calcd for C₁₆H₃₂O₅ SSi+NH₄⁺: 382.2078 [M+NH₄⁺]; found 382.2083.

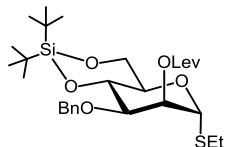
Synthesis of 13



Ethyl 3-*O*-benzyl-4,6-*O*-di-*tert*-butylsilylene-1-thio- α -D-mannopyranoside (13).

A solution of **12** (2.07 g, 5.68 mmol) and Bu₂SnO (1.65 g, 6.63 mmol) in dry toluene (100 ml) was refluxed in a Dean-Stark apparatus for 3.5 h. The mixture was cooled to r. t. and tetrabutylammonium iodide (2.31 g, 6.25 mmol), dry DMF (5 ml) and BnBr (0.75 ml, 6.3 mmol) were added. The mixture was refluxed for 3.5 h, cooled to r. t., diluted with EtOAc (200 ml) and washed with 1 M aq. HCl (3×300 ml) and NaHCO₃ (300 ml). The organic layer was filtered over a pad of Celite, the filtrate was washed with 5% aq. Na₂S₂O₃ (200 ml), water (200 ml) and brine (200 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (toluene - EtOAc, 10 : 1) to afford **13** (2.40 g, 5.28 mmol, 93%) as a pale yellow syrup. *R*_f = 0.57 (toluene - EtOAc, 3 : 1); [α]_D²⁰ = +118 (*c* 1.4, CHCl₃); ¹H-NMR (300 MHz, CDCl₃): δ 7.41-7.28 (m, 5H, CH₂Ph), 5.30 (s, 1H, H-1), 4.93 (AB, 1H, ²J = 11.9 Hz, CH₂Ph), 4.78 (AB, 1H, ²J = 11.9 Hz, CH₂Ph), 4.29 (t, 1H, ³J_{4,3} = ³J_{4,5} = 9.1 Hz, H-4), 4.14 (td, 1H, ³J_{5,6} = ³J_{5,4} = 9.8 Hz, ³J_{5,6} = 4.7 Hz, H-5), 4.10-3.96 (m, 3H, H-2, H6a, H6b), 3.63 (dd, 1H, ³J_{3,4} = 3.4 Hz, ³J_{3,4} = 8.9 Hz, H-3), 2.78 (d, 1H, *J* = 1.7 Hz, 2-OH), 2.70-2.50 (m, 2H, CH₂, SEt), 1.28 (t, 3H, *J* = 7.4 Hz, CH₃, SEt), 1.08 (s, 9H, 3×CH₃, DTBS), 1.03 (s, 9H, 3×CH₃, DTBS); ¹³C-NMR (75 MHz, CDCl₃): δ 138.24 (C_q, CH₂Ph), 128.42, 127.81, 127.71 (5×CH, CH₂Ph), 83.82 (C-1), 78.58 (C-3), 75.34 (C-4), 73.37 (CH₂, 3-OBn), 71.58 (C-2), 67.41 (C-5), 66.49 (C-6), 27.44, 27.07 (6×CH₃, DTBS), 24.99 (CH₂, SEt), 22.62, 20.02 (2×C_q, DTBS), 14.88 (CH₃, SEt); HRMS (ESI): *m/z* calcd for C₂₃H₃₈O₅ SSi+Na⁺: 477.2101 [M+Na⁺]; found 477.2101.

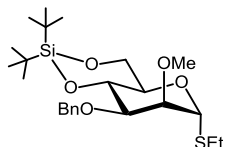
Synthesis of 14



Ethyl 3-O-benzyl-4,6-O-di-tert-butylsilylene-2-O-levulinoyl-1-thio- α -D-mannopyranoside (14).

To a stirred solution of compound **13** (604 mg, 1.33 mmol), levulinic acid (270 μ L, 2.66 mmol) and DMAP (10 mg, 82 μ mol) in dry CH_2Cl_2 (10 ml), was added N,N' -diisopropylcarbodiimide (416 μ L, 2.66 mmol) at 0 $^\circ\text{C}$ under atmosphere of Ar. After 15 min the reaction was warmed to r. t. and stirred for 4 h. The solids were removed by filtration over a pad of Celite, the filtrate was concentrated, the residue was purified by silica gel chromatography (hexane – EtOAc, 2 : 1) to afford **14** (666 mg, 1.20 mmol, 90%) as a syrup. $R_f = 0.39$ (hexane – EtOAc, 2 : 1); $[\alpha]_D^{20} = +53$ (c 1.1, CHCl_3); $^1\text{H-NMR}$ (300 MHz, CDCl_3): δ 7.32-7.16 (m, 5H, CH_2Ph), 5.28 (dd, 1H, $^3J_{2,1} = 1.4$ Hz, $^3J_{2,3} = 3.5$ Hz, H-2), 5.10 (d, 1H, $^3J_{1,2} = 1.4$ Hz, H-1), 4.64 (s, 2H, CH_2Ph), 4.12 (t, 1H, $^3J_{4,3} = ^3J_{4,5} = 8.9$ Hz, H-4), 4.06 (dd, 1H, $^3J_{6a,5} = 4.4$ Hz, $^2J_{6a,6b} = 9.7$ Hz, H-6a), 4.02-3.93 (m, 1H, H-5), 3.90 (t, 1H, $^3J_{6b,5} = ^2J_{6b,6a} = 9.7$ Hz, H-6b), 3.59 (dd, 1H, $^3J_{3,2} = 3.5$ Hz, $^3J_{3,4} = 8.9$ Hz, H-3), 2.79-2.44 (m, 6H, $3 \times \text{CH}_2$, SEt, Lev), 2.09 (s, 3H, CH_3 , Lev), 1.20 (t, 3H, $^3J = 7.4$ Hz, CH_3 , SEt), 1.02 (s, 9H, $3 \times \text{CH}_3$, DTBS), 0.94 (s, 9H, $3 \times \text{CH}_3$, DTBS); $^{13}\text{C-NMR}$ (75 MHz, CDCl_3): δ 206.36 (CO, Lev), 171.98 (CO, Lev), 138.39 (C_q , CH_2Ph), 128.25, 127.42, 127.40 ($5 \times \text{CH}$, CH_2Ph), 83.07 (C-1), 76.60 (C-3), 74.96 (C-4), 72.33 (CH_2 , CH_2Ph), 72.00 (C-2), 68.03 (C-5), 66.48 (C-6), 38.03 (CH_2 , Lev), 29.77 (CH_3 , Lev), 28.12 (CH_2 , Lev), 27.40, 27.04 ($6 \times \text{CH}_3$, DTBS), 25.58 (CH_2 , SEt), 22.67, 19.98 ($2 \times \text{C}_q$, DTBS), 14.92 (CH_3 , SEt); HRMS (ESI): m/z calcd for $\text{C}_{28}\text{H}_{44}\text{O}_7\text{SSi} + \text{Na}^+$: 575.2469 [M+Na $^+$]; found 575.2470.

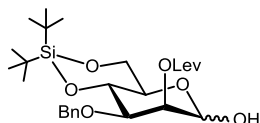
Synthesis of 15



Ethyl 3-O-benzyl-4,6-O-di-tert-butylsilylene-2-O-methyl-1-thio- α -D-mannopyranoside (15).

A slurry of **13** (1.30 g, 2.86 mmol) and NaH (210 mg, 8.75 mmol, 60% in mineral oil) in dry DMF (50 ml) was stirred at -40 $^\circ\text{C}$ for 1.5 h under atmosphere of Ar. Then CH_3I (300 μ L, 4.82 mmol) was added, the mixture was warmed to r. t. and stirred for 2.5 h. Methanol (10 ml) was added and the mixture was stirred for 10 min, diluted with CHCl_3 (100 ml) and washed with aq. HCl (1 M, 300 ml), sat. aq. NaHCO_3 (200 ml), aq. $\text{Na}_2\text{S}_2\text{O}_3$ (5%, 100 ml) and brine (100 ml). The organic layer was dried over Na_2SO_4 , filtered and concentrated. The residue was purified by silica gel chromatography (toluene – EtOAc, 30 : 1) to afford **15** (1.13 g, 2.41 mmol, 84%, $\alpha/\beta = 30 : 1$) as a pale yellow syrup. $R_f = 0.61$ (hexane – EtOAc, 3 : 1); $[\alpha]_D^{20} = +101$ (c 0.9, CHCl_3); $^1\text{H-NMR}$ (300 MHz, CDCl_3 , α -anomer): δ 7.41-7.25 (m, 5H, CH_2Ph), 5.30 (d, 1H, $^3J_{1,2} = 1.3$ Hz, H-1), 4.92 (AB, 1H, $^2J = 12.3$ Hz, CH_2Ph), 4.76 (AB, 1H, $^2J = 12.4$ Hz, CH_2Ph), 4.31 (t, 1H, $^3J_{4,3} = ^3J_{4,5} = 9.1$ Hz, H-4), 4.13-3.95 (m, 3H, H-5, H-6a, H-6b), 3.63 (dd, 1H, $^3J_{3,2} = 3.3$ Hz, $^3J_{3,4} = 9.2$ Hz, H-3), 3.59 (dd, 1H, $^3J_{2,1} = 1.4$ Hz, $^3J_{2,3} = 3.3$ Hz, H-2), 3.50 (s, 3H, CH_3 , Me), 2.70-2.52 (m, 2H, CH_2 , SEt), 1.28 (t, 3H, $^3J = 7.4$ Hz, CH_3 , SEt), 1.08 (s, 9H, $3 \times \text{CH}_3$, DTBS), 1.03 (s, 9H, $3 \times \text{CH}_3$, DTBS); $^{13}\text{C-NMR}$ (75 MHz, CDCl_3 , α -anomer): δ 138.89 (C_q , CH_2Ph), 128.28, 127.55, 127.45 ($5 \times \text{CH}$, CH_2Ph), 82.48 (C-1), 81.34 (C-2), 78.75 (C-3), 75.43 (C-4), 73.48 (CH_2Ph), 68.14 (C-5), 66.51 (C-6), 59.18 (CH_3 , Me), 27.45, 27.13 ($6 \times \text{CH}_3$, DTBS), 25.44 (CH_2 , SEt), 22.60, 20.01 ($2 \times \text{C}_q$, DTBS), 15.00 (CH_3 , SEt); HRMS (ESI): m/z calcd for $\text{C}_{24}\text{H}_{40}\text{O}_5\text{SSi} + \text{H}^+$: 469.2438 [M+H $^+$]; found 469.2431.

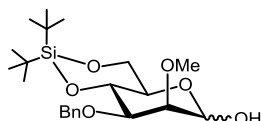
Synthesis of 16



3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-levulinoyl- α -D-mannopyranose (**16**).

Compound **14** (600 mg, 1.09 mmol) was dissolved in acetone - water (8 ml, 24 : 1) and added to a stirred solution of NBS (846 mg, 4.75 mmol) in the same solvent mixture (8 ml) at 0 °C. The mixture was stirred for 15 min and then the reaction was quenched by addition of aq. 5% Na₂S₂O₃ (5 ml). The mixture was warmed to r. t., diluted with EtOAc (50 ml) and washed with water (50 ml) and brine (50 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 2 : 1) to afford **16** (427 mg, 0.84 mmol, 77%, $\alpha/\beta = 6 : 1$) as a syrup. $R_f = 0.44$ (hexane – EtOAc, 1 : 1); $[\alpha]_D^{20} = -19$ (c 1.1, CHCl₃); ¹H NMR (300 MHz, CDCl₃, α -anomer): δ 7.40-7.23 (m, 5H, CH₂Ph), 5.30 (dd, 1H, ³ $J_{2,1} = 1.6$ Hz, ³ $J_{2,3} = 3.5$ Hz, H-2), 5.11 (dd, 1H, ³ $J_{1,2} = 1.4$ Hz, ³ $J_{1,OH} = 3.8$ Hz, H-1), 4.74 (s, 2H, CH₂Ph), 4.22-3.90 (m, 4H, H-4, H-5, H-6a, H-6b), 3.80 (dd, 1H, ³ $J_{3,2} = 3.5$ Hz, ³ $J_{3,4} = 9.4$ Hz, H-3), 3.13 (d, 1H, ³ $J_{OH,1} = 4.0$ Hz, 1-OH), 2.91-2.54 (m, 2 \times CH₂, Lev), 2.15 (s, CH₃, Lev), 1.10 (s, 3 \times CH₃, DTBS), 1.00 (s, 3 \times CH₃, DTBS); ¹³C NMR (75 MHz, CDCl₃, α -anomer): δ 206.59 (CO, Lev), 171.13 (CO, Lev), 138.61 (Cq, CH₂Ph), 128.24, 127.40, 127.35 (5 \times CH, CH₂Ph), 92.93 (C-1), 75.69 (C-3), 74.80 (C-4), 72.31 (CH₂Ph), 70.49 (C-2), 67.53 (C-5), 66.33 (C-6), 38.05 (CH₂, Lev), 29.76 (CH₃, Lev), 28.11 (CH₂, Lev), 27.43, 27.05 (6 \times CH₃, DTBS), 22.71, 19.92 (2 \times Cq, DTBS); HRMS (ESI): m/z calcd for C₂₆H₄₀NaO₈Si+Na⁺: 531.2385 [M+Na⁺]; found 531.2387.

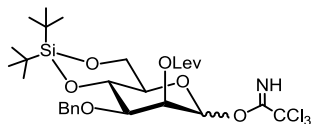
Synthesis of 17



3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- α -D-mannopyranose (**17**).

Compound **15** (1.04 g, 2.22 mmol) was dissolved in acetone - water (100 ml, 24 : 1) and added to a stirred solution of NBS (2.01 g, 11.3 mmol) in the same solvent mixture (100 ml) at 0 °C. The mixture was stirred for 15 min and then the reaction was quenched by addition of aq. Na₂S₂O₃ (5%, 50 ml). The mixture was warmed to r. t., diluted with EtOAc (300 ml) and washed with water (100 ml) and brine (100 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 2 : 1) to afford **17** (840 mg, 1.98 mmol, 89%, $\alpha/\beta = 4 : 1$) as a syrup. $R_f = 0.19$ (hexane – EtOAc, 3 : 1); $[\alpha]_D^{20} = +25$ (c 1.2, CHCl₃); ¹H-NMR (300 MHz, CDCl₃, α -anomer): δ 7.43-7.24 (m, 5H, CH₂Ph), 5.22 (dd, 1H, ³ $J_{1,2} = 1.7$ Hz, ³ $J_{1,OH} = 3.5$ Hz, H-1), 4.92 (AB, 1H, ² $J = 12.4$ Hz, CH₂Ph), 4.76 (AB, 1H, ² $J = 12.4$ Hz, CH₂Ph), 4.30 (t, 1H, ³ $J_{4,3} = 3.5$ Hz, ³ $J_{4,5} = 9.3$ Hz, H-4), 4.09-3.87 (m, 3H, H-5, H-6a, H-6b), 3.76 (dd, 1H, ³ $J_{3,2} = 3.2$ Hz, ³ $J_{3,4} = 9.5$ Hz, H-3), 3.56 (dd, 1H, ³ $J_{2,1} = 1.7$ Hz, ³ $J_{2,3} = 3.2$ Hz, H-2), 3.53 (s, 3H, CH₃, Me), 2.57 (d, 1H, ³ $J_{OH,1} = 3.5$ Hz, 1-OH), 1.08 (s, 9H, 3 \times CH₃, DTBS), 1.02 (s, 9H, 3 \times CH₃, DTBS); ¹³C-NMR (75 MHz, CDCl₃, α -anomer): δ 139.14 (Cq, CH₂Ph), 128.27, 127.46, 127.39 (5 \times CH, CH₂Ph), 93.16 (C-1), 79.81 (C-2), 78.07 (C-3), 75.32 (C-4), 73.60 (CH₂Ph), 68.00 (C-5), 66.72 (C-6), 60.01 (CH₃, Me), 27.47, 27.13 (6 \times CH₃, DTBS), 22.64, 19.95 (2 \times Cq, DTBS); HRMS (ESI): m/z calcd for C₂₂H₃₆O₆Si+Na⁺: 447.2173 [M+Na⁺]; found 447.2174.

Synthesis of 18

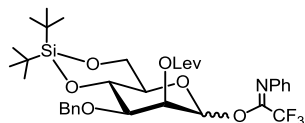


3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-levulinoyl- α -D-mannopyranose trichloroacetimidate (**18**).

To a solution of **16** (90 mg, 177 μ mol) in dry CH₂Cl₂ (2 ml) trichloroacetonitrile (75 μ L, 752 μ mol) and a solution of DBU (2 μ L, 15 μ mol) in CH₂Cl₂ (0.5 ml) were added successively at 0 °C. The mixture was stirred for 1h., then warmed to r. t., diluted with EtOAc (10 ml) and washed with sat. aq. NaHCO₃ (10 ml) and brine (10 ml). The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 3 : 1 supplemented by 1% Et₃N) to afford **18** (104 mg, 159 μ mol, 90%) as a syrup. $R_f = 0.60$ and 0.68 (hexane – EtOAc, 1 : 1 + 1% Et₃N); ¹H NMR (300 MHz, CDCl₃): δ 8.68 (s, 1H, NH), 7.39-7.24 (m, 5H, CH₂Ph), 6.13 (d, 1H, ³ $J_{1,2} = 1.9$ Hz, H-1), 5.38 (dd, 1H, ³ $J_{2,1} = 1.9$ Hz, ³ $J_{2,3} = 3.5$ Hz, H-2), 4.77 (s, 2H, -CH₂Ph), 4.25

(t, 1H, $^3J_{4,3} = ^3J_{4,5} = 9.2$ Hz, H-4), 4.15-4.11 (m, 1H, H-6a), 3.99-3.98 (m, 2H, H-5, H-6b), 3.82 (dd, 1H, $^3J_{3,2} = 3.5$ Hz, $^3J_{3,4} = 9.5$ Hz, H-3), 2.87-2.67 (m, 4H, $2\times\text{CH}_2$, Lev), 2.17 (s, 3H, CH_3 , Lev), 1.11 (s, 9H, $3\times\text{CH}_3$, DTBS), 0.97 (s, 9H, $3\times\text{CH}_3$, DTBS). ^{13}C -NMR (75 MHz, CDCl_3): δ 206.03 (CO, Lev), 171.81 (CO, Lev), 159.62 (OC(NH)CCl₃), 138.10 (C_q CH₂Ph), 128.29, 127.77, 127.58 ($5\times\text{CH}$, CH₂Ph), 94.97 (C-1), 90.66 (OC(NH)CCl₃), 75.25 (C-3), 74.23 (C-4), 72.70 (CH₂Ph), 69.90 (C-5), 68.76 (C-2), 66.35 (C-6), 37.97 (CH₂, Lev), 29.73 (CH₃, Lev), 28.01 (CH₂, Lev), 27.35, 26.92 ($6\times\text{CH}_3$, DTBS), 22.63, 19.96 ($2\times\text{C}_q$, DTBS).

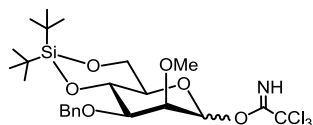
Synthesis of 19



3-O-Benzyl-4,6-O-di-tert-butylsilylene-2-O-levulinoyl-D-mannopyranose N-phenyltrifluoroacetimidate (19).

To a vigorously stirred suspension of **16** (129 mg, 254 μmol) and K_2CO_3 (75 mg, 543 μmol) in acetone (5 ml) 2,2,2-trifluoro-N-phenylacetimidoyl chloride (100 μL , 630 μmol) was added. The mixture was stirred for 1 h., diluted with acetone (10 ml), the solids were removed by filtration over a pad of Celite and the filtrate was concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 3 : 1 supplemented by 1% Et₃N) to afford **19** (172 mg, 253 μmol , 99%, α : β ~ 6 : 1) as a syrup. $R_f = 0.71$ and 0.67 (hexane – EtOAc, 1 : 1); ^1H -NMR (300 MHz, CDCl_3 , α -anomer): δ 7.44-7.23 (m, 6H, CH₂Ph), 7.18-7.08 (m, 2H, CH₂Ph), 6.83-6.80 (m, 2H, CH₂Ph), 6.07 (br. s, 1H, H-1), 5.40 (dd, 1H, $^3J_{2,1} = 1.7$ Hz, $^3J_{2,3} = 3.5$ Hz, H-2), 4.80 (AB, 1H, $^2J = 12.3$ Hz, CH₂Ph), 4.74 (AB, 1H, $^2J = 12.3$ Hz, CH₂Ph), 4.23 (t, 1H, $^3J_{4,3} = ^3J_{4,5} = 9.4$ Hz, H-4), 4.13 (dd, 1H, $^3J_{6a,5} = 4.7$ Hz, $^2J_{6a,6b} = 9.8$ Hz, H-6a), 3.96 (t, 1H, $^3J_{6b,5} = ^2J_{6b,6a} = 10.1$ Hz, H-6b), 3.88-3.84 (m, 1H, H-5), 3.78 (dd, 1H, $^3J_{3,4} = 9.4$ Hz, $^3J_{3,2} = 3.6$ Hz, H-3), 2.81-2.64 (m, 4H, $2\times\text{CH}_2$, Lev), 2.24 (s, 3H, CH_3 , Lev), 1.10 (s, 9H, $3\times\text{CH}_3$, DTBS), 1.00 (s, 9H, $3\times\text{CH}_3$, DTBS). ^{13}C -NMR (75 MHz, CDCl_3 , α -anomer): δ 205.98 (CO, Lev), 171.74 (CO, Lev), 143.01 (OC(NH)CF₃), 138.20 (C_q, CH₂Ph), 128.77, 128.29, 127.57, 124.59, 119.38 ($10\times\text{CH}$ CH₂Ph), 95.60 (CF₃), 94.14 (C-1), 75.73 (C-3), 74.13 (C-4), 72.89 (CH₂Ph), 69.75 (C-5), 68.72 (C-2), 66.31 (C-6), 37.97 (CH₂, Lev), 29.69 (CH₃, Lev), 27.98 (CH₂, Lev), 27.34, 26.91 ($6\times\text{CH}_3$, DTBS), 22.67, 19.88 ($2\times\text{C}_q$, DTBS).

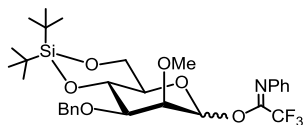
Synthesis of 20



3-O-Benzyl-4,6-O-di-tert-butylsilylene-2-O-methyl-D-mannopyranose trichloroacetimidate (20).

To a solution of **17** (113 mg, 266 μmol) in dry CH_2Cl_2 (3 ml) trichloroacetonitril (140 μL , 1.40 μmol) and a solution of DBU (3.4 μL , 23 μmol) in CH_2Cl_2 (0.5 ml) were added successively at 0 °C. The mixture was stirred for 1.5 h, then warmed to r. t., diluted with EtOAc (30 ml) and washed with sat. aq. NaHCO_3 (30 ml) and brine (30 ml). The organic layer was dried over Na_2SO_4 and concentrated. The residue was purified by silica gel column chromatography (hexane – EtOAc, 4 : 1 supplemented by 1% Et₃N) to afford **20** (143 mg, 251 μmol , 94%, $\alpha/\beta = 7$: 1) as a pale yellow syrup. $R_f = 0.61$ and 0.50 and (hexane - EtOAc, 3 : 1); ^1H -NMR (300 MHz, CDCl_3 , α -anomer): δ 8.59 (s, 1H, NH), 7.42-7.24 (m, 5H, CH₂Ph), 6.21 (d, 1H, $^3J_{1,2} = 1.8$ Hz, H-1), 4.97 (AB, 1H, $^2J = 12.3$ Hz, CH₂Ph), 4.80 (AB, 1H, $^2J = 12.3$ Hz, CH₂Ph), 4.37 (t, 1H, $^3J_{4,3} = ^3J_{4,5} = 9.3$ Hz, H-4), 4.13-4.07 (m, 1H, H-6a), 4.00-3.86 (m, 2H, H-5, H-6b), 3.77 (dd, 1H, $^3J_{3,2} = 3.3$ Hz, $^3J_{3,4} = 9.5$ Hz, H-3), 3.58 (dd, 1H, $^3J_{2,1} = 1.8$ Hz, $^3J_{2,3} = 3.4$ Hz, H-2), 3.57 (s, 3H, CH_3 , Me), 1.08 (s, 9H, $3\times\text{CH}_3$, DTBS), 0.99 (s, 9H, $3\times\text{CH}_3$, DTBS); ^{13}C -NMR (75 MHz, CDCl_3 , α -anomer): δ 160.16 (OC(NH)CCl₃), 138.64 (C_q, CH₂Ph), 128.34, 127.92, 127.61 ($5\times\text{CH}$, CH₂Ph), 95.27 (C-1), 78.15 (C-2), 77.09 (C-3), 74.65 (C-4), 73.73 (CH₂Ph), 70.33 (C-5), 66.38 (C-6), 59.84 (CH₃, Me), 27.39, 27.01 ($6\times\text{CH}_3$, DTBS), 22.57, 19.99 ($2\times\text{C}_q$, DTBS).

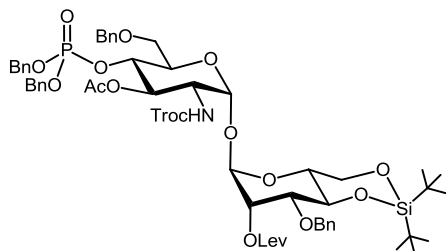
Synthesis of 21



3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- α -D-mannopyranose *N*-phenyl trifluoroacetimidate (21).

To a vigorously stirred suspension of **17** (840 mg, 1.98 μ mol) and K_2CO_3 (550 mg, 3.96 μ mol) in acetone (70 ml) 2,2,2-trifluoro-*N*-phenylacetimidoyl chloride (750 μ L, 4.97 μ mol) was added at r. t. and the mixture was stirred for 3 h. The solids were removed by filtration over a pad of Celite and the filtrate was concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 20 : 1 + 1% Et₃N \rightarrow 9 : 1 supplemented by 1% Et₃N) to afford **21** (1.1 g, 1.85 mmol, 93%, $\alpha/\beta \sim 10 : 1$) as a slightly yellow syrup. $R_f = 0.61$ and 0.56 (hexane – EtOAc, 4 : 1); ¹H-NMR (300 MHz, CDCl₃, α -anomer): δ 7.43-7.26 (m, 7H, CH₂Ph), 7.14-7.09 (m, 1H, CH₂Ph), 6.83-6.80 (m, 1H, CH₂Ph), 6.10 (br. s, 1H, H-1), 5.00 (AB, 1H, ² $J = 12.2$ Hz, CH₂Ph), 7.78 (AB, 1H, ² $J = 12.2$ Hz, CH₂Ph), 4.34 (t, 1H, ³ $J_{4,3} = ^3J_{4,5} = 9.5$ Hz, H-4), 4.11 (dd, 1H, ³ $J_{6a,5} = 4.8$ Hz, ² $J_{6a,6b} = 9.8$ Hz, H-6a), 3.96 (t, 1H, ³ $J_{6b,5} = ^2J_{6b,6a} = 10.1$ Hz, H-6b), 3.82 (td, 1H, ³ $J_{5,4} = ^3J_{5,6b} = 9.9$ Hz, ³ $J_{5,6a} = 4.8$ Hz, H-5), 3.71 (dd, 1H, ³ $J_{3,2} = 3.29$ Hz, ³ $J_{3,4} = 9.6$ Hz, H-3), 3.57 (br. s, 1H, H-2), 3.47 (s, 3H, CH₃, Me), 1.08 (s, 9H, 3 \times CH₃, DTBS), 1.02 (s, 9H, 3 \times CH₃, DTBS). ¹³C-NMR (75 MHz, CDCl₃, α -anomer): δ 143.30 (OC(NH)CF₃), 138.68 (Cq, CH₂Ph), 128.86, 128.34, 127.75, 127.63, 124.63, 119.45 (10 \times CH, arom.), 78.14 (C-2), 77.62 (C-3), 74.43 (C-4), 74.01 (CH₂Ph), 70.25 (C-5), 66.29 (C-6), 59.84 (CH₃, Me), 27.40, 26.99 (6 \times CH₃, DTBS), 22.60, 19.91 (2 \times Cq, DTBS).

Synthesis of 22

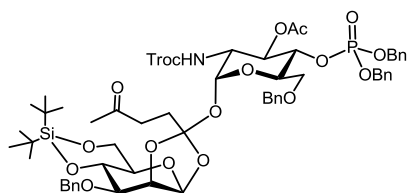


3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-levulinoyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-3-*O*-acetyl-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (22).

A solution of glycosyl donor **18** (60 mg, 92 μ mol) and glycosyl acceptor **8** (68 mg, 91 μ mol) in dry CH₂Cl₂ (1 ml) was stirred with activated powdered molecular sieves 4 Å for 1.5 h. at r. t. under atmosphere of Ar. The mixture was cooled to -15 °C and a solution of TMSOTf (3.3 μ L, 18 μ mol) in dry CH₂Cl₂ (165 μ L of a stock solution prepared from 10 μ L TMSOTf in 500 μ L CH₂Cl₂) was added. The mixture was stirred for 40 min and warmed to 0 °C. The reaction was quenched by addition of sat. aq. NaHCO₃ (500 μ L), diluted with EtOAc (20 ml), the solids were removed by filtration over a pad of Celite and the filtrate was washed with water (10 ml) and brine (10 ml). The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by HPLC (three successive columns: toluene - EtOAc, 8 : 1 \rightarrow 3 : 1; toluene – EtOAc, 10 : 1 \rightarrow 3 : 1, column B and toluene – acetone, 8 : 1) to afford compound **22** (9.4 mg, 7.6 μ mol, 8%) as a syrup. $R_f = 0.17$ (toluene – EtOAc, 3 : 1); $[\alpha]_D^{20} = +39.0$ (*c* 0.5, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.38-7.21 (m, 20H, CH₂Ph), 5.25 (t, 1H, ³ $J_{3,4} = ^3J_{3,4} = 9.7$ Hz, H-3), 5.16 (dd, 1H, ³ $J_{2',1'} = 1.4$ Hz, ³ $J_{2',3'} = 3.6$ Hz, H-2'), 5.14 (d, 1H, ³ $J_{1,2} = 3.7$ Hz, H-1), 5.10 (d, 1H, ³ $J_{NH,2} = 9.7$ Hz, NH), 5.07 (AB, 1H, ² $J = 12.2$ Hz, CH₂Ph or Troc), 5.02 (d, 1H, ³ $J_{1',2'} = 1.1$ Hz, H-1'), 4.98-4.90 (m, 4H, OP(O)(OCH₂Ph)₂), 4.86 (AB, 1H, ² $J = 11.9$ Hz, CH₂Ph or Troc), 4.77 (AB, 1H, ² $J = 11.9$ Hz, CH₂Ph or Troc), 4.67 (ddd, 1H, ³ $J_{4,5} = ^3J_{4,3} = ^3J_{4,p} = 9.2$ Hz, H-4), 4.50 (AB, 1H, ² $J = 12.1$ Hz, CH₂Ph or Troc) 4.46 (AB, 1H, ² $J = 12.1$ Hz, CH₂Ph or Troc), 4.34 (AB, 1H, ² $J = 12.1$ Hz, CH₂Ph or Troc), 4.20 (t, 1H, ³ $J_{4',3'} = ^3J_{4',5'} = 9.5$ Hz, H-4'), 4.11-4.07 (m, 1H, H-2), 4.02 (dd, 1H, ³ $J_{6a',5'} = 5.0$ Hz, ² $J_{6a',6b'} = 10.3$ Hz, H-6a'), 3.92 (t, 1H, ³ $J_{6b',5'} = ^2J_{6b',6a'} = 10.3$ Hz, H-6b'), 3.82 (dt, 1H, $J = 10.0$ Hz, $J = 2.8$ Hz, H-5), 3.77 (dd, 1H, ³ $J_{3',2'} = 3.6$ Hz, ³ $J_{3',4'} = 9.5$ Hz, H-3'), 3.70-3.65 (m, 3H, H-5', H-6a, H-6b), 2.71-2.61

(m, 4H, 2×CH₂, Lev), 2.10 (s, 3H, CH₃, Lev), 1.90 (s, 3H, CH₃, Ac), 1.10 (s, 9H, 3×CH₃, DTBS), 0.98 (s, 9H, 3×CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ 206.02 (CO, Lev), 172.00, 171.51 (2×CO, Ac, Lev), 154.19 (CO, Troc), 138.27, 138.01 (2×Cq, CH₂Ph), 135.50, 135.43 (2×Cq, OP(O)(OCH₂Ph)₂), 129.03, 128.66, 128.61, 128.54, 128.36, 128.29, 128.21, 127.99, 127.85, 127.44, 127.56, 127.52 (20×CH, CH₂Ph), 95.30 (CCl₃, Troc), 93.44 (C-1', ¹J_{C,H} = 172 Hz), 92.77 (C-1, ¹J_{C,H} = 175 Hz), 75.77 (C-3'), 74.45 (C-4'), 73.41, 73.29, (2×CH₂, Troc, CH₂Ph), 73.09 (C-4, ²J_{C₄,P} = 5.7 Hz), 71.09 (C-3), 70.44 (C-5, ³J_{C₅,P} = 6.9 Hz), 70.30 (C-2'), 69.67, 69.64, 69.60 (2×CH₂, OP(O)(OCH₂Ph)₂), 68.79 (C-5'), 67.60 (C-6), 66.28 (C-6'), 53.74 (C-2), 38.00 (CH₂, Lev), 29.67 (CH₃, Lev), 28.09 (CH₂, Lev), 27.44, 27.07 (6×CH₃, DTBS), 22.69 (Cq, DTBS), 20.70 (CH₃, Ac), 19.84 (Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃): δ -2.26; HRMS (ESI): *m/z* calcd for C₅₈H₇₃Cl₃NO₁₈PSi+H⁺: 1236.3473 [M+H]⁺; found 1236.3461.

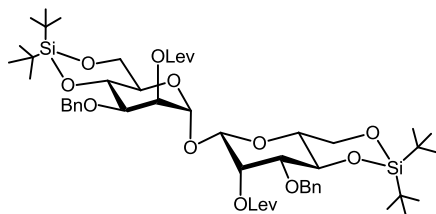
Synthesis of 23



Orthoester (23).

A solution of glycosyl donor **18** (23 mg, 35 μmol) and glycosyl acceptor **8** (27 mg, 36 μmol) in dry CH₂Cl₂ (1 ml) was stirred with activated powdered 4 Å molecular sieves for 1 h. at r. t. under atmosphere of Ar. The mixture was cooled to 0 °C and a solution of TMSOTf (0.3 μL, 2 μmol) in dry CH₂Cl₂ (30 μL of a stock solution prepared from 10 μL TMSOTf in 1 ml CH₂Cl₂) was added. The mixture was stirred for 45 min and then the reaction was quenched by addition of sat. aq. NaHCO₃ (500 μL) and diluted with EtOAc (20 ml). The solids were removed by filtration over a pad of Celite, the filtrate was washed with water (10 ml) and brine (10 ml). The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by HPLC (two successive columns: toluene - EtOAc, 4 : 1 and toluene - EtOAc, 10 : 1 → 4 : 1, column B) to afford compound **23** (11 mg, 8.9 μmol, 25%) as a syrup. *R*_f = 0.43 (toluene - EtOAc, 3 : 1); [α]_D²⁰ = +13 (*c* 0.4, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.44-7.28 (m, 15H, CH₂Ph), 7.19-7.13 (m, 3H, CH₂Ph), 7.06-7.04 (m, 2H, CH₂Ph), 5.44 (d, 1H, ³J_{NH,2} = 9.9 Hz, NH), 5.37 (d, 1H, ³J_{1,2} = 3.7 Hz, H-1), 5.22 (dd, 1H, ³J = 9.0 Hz, ³J = 10.8 Hz, H-3), 5.00-4.93 (m, 4H, OP(O)(OCH₂Ph)₂), 4.88 (AB, 1H, ²J = 12.8 Hz, CH₂Ph or Troc), 4.83 (AB, 1H, ²J = 12.9 Hz, CH₂Ph or Troc), 4.80 (AB, 1H, ²J = 12.0 Hz, CH₂Ph or Troc), 4.64 (d, 1H, ³J_{1',2'} = 2.2 Hz, H-1'), 4.64 (AB, 1H, ²J = 12.0 Hz, CH₂Ph or Troc), 4.60 (dd, 1H, ³J_{2',1'} = 2.3 Hz, ³J_{2',3'} = 4.1 Hz, H-2'), 4.34 (ddd, 1H, ³J_{4,3} = ³J_{4,5} = ³J_{4,P} = 9.4 Hz, H-4), 4.27 (AB, 1H, ²J = 10.4 Hz, CH₂Ph or Troc), 4.20 (AB, 1H, ²J = 10.5 Hz, CH₂Ph or Troc), 4.19-4.16 (m, 1H, H-5), 4.05-3.98 (m, 2H, H-2, H-6a'), 4.04 (t, 1H, ³J_{4',5'} = ³J_{4',3'} = 9.3 Hz, H-4'), 3.78-3.74 (m, 2H, H-6a, H-6b'), 3.51 (dd, 1H, *J* = 7.7 Hz, *J* = 10.4 Hz, H-6b), 3.20 (dd, 1H, ³J_{3',2'} = 4.1 Hz, ³J_{3',4'} = 9.2 Hz, H-3'), 2.79-2.70 (m, 2H, H-5', CH₂, Lev), 2.52-2.47 (m, 1H, CH₂, Lev), 2.40 (dt, 1H, *J* = 7.2 Hz, *J* = 14.3 Hz, CH₂, Lev), 2.17 (s, 3H, CH₃, Lev), 2.02-1.98 (m, 1H, CH₂, Lev), 1.85 (s, 3H, CH₃, Ac), 1.08 (s, 9H, 3×CH₃, DTBS), 1.02 (s, 9H, 3×CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ 208.04 (CO, Lev), 170.98 (CO, Lev), 154.39 (CO, Troc), 138.32, 137.86 (2×Cq, CH₂Ph), 135.54, 135.36 (2×Cq, OP(O)(OCH₂Ph)₂), 128.77, 128.72, 128.65, 128.56, 128.40, 128.34, 128.15, 128.02, 127.90, 127.89, 127.76 (20×CH, CH₂Ph), 123.87 (Cq, orthoester), 97.16 (C-1'), 95.42 (CCl₃, Troc), 90.96 (C-1), 79.09 (C-2'), 76.60 (C-3'), 74.54 (CH₂, CH₂Ph or Troc), 74.26 (C-4, ²J_{C₄,P} = 5.9 Hz), 74.14 (C-4'), 73.60, 72.91 (2×CH₂, CH₂Ph or Troc), 71.00 (C-3), 69.89 (C-5, ³J_{C₅,P} = 5.3 Hz), 69.78, 69.74, 69.71, 69.67, 69.63 (C-6, OP(O)(OCH₂Ph)₂), 68.78 (C-5'), 66.19 (C-6'), 53.81 (C-2), 37.97, 32.99 (2×CH₂, Lev), 30.20 (CH₃, Lev), 27.46, 27.03 (6×CH₃, DTBS), 22.74 (Cq, DTBS), 20.65 (CH₃, Ac), 19.89 (Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃): δ -1.92; HRMS (ESI): *m/z* calcd for C₅₈H₇₃Cl₃NO₁₈PSi+NH₄⁺: 1253.3738 [M+NH₄⁺]; found 1253.3734.

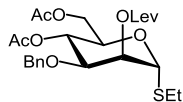
Synthesis of 24



3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-levulinoyl- β -D-mannopyranosyl-(1 \leftrightarrow 1)-3-*O*-benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-levulinoyl- α -D-mannopyranoside (24).

24 was isolated as by-product from glycosylation reactions between donors **18** or **19** and acceptor **8**. $R_f = 0.43$ (toluene – EtOAc, 3 : 1); $[\alpha]_D^{20} = -21$ (c 0.5, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.38-7.22 (m, 10H, CH₂Ph), 5.44 (dd, 1H, ³ $J_{2,1'}$ = 0.9 Hz, ³ $J_{2,3'}$ = 3.3 Hz, H-2'), 5.20 (dd, 1H, ³ $J_{2,1}$ = 1.6 Hz, ³ $J_{2,3}$ = 3.5 Hz, H-2), 4.90 (d, 1H, ³ $J_{1,2}$ = 1.3 Hz, H-1), 4.76-4.67 (m, 4H, 2 \times CH₂Ph), 4.67 (s, 1H, H-1'), 4.14-4.07 (m, 3H, H-4, H-4', H-6a'), 4.01 (dd, 1H, ³ $J_{6a,5}$ = 4.7 Hz, ² $J_{6a,6b}$ = 9.5 Hz, H-6a), 3.97-3.92 (m, 2H, H-5, H-6b'), 3.87 (t, 1H, ³ $J_{6b,5}$ = ² $J_{6b,6a}$ = 9.8 Hz, H-6b), 3.74 (dd, 1H, ³ $J_{3,2}$ = 3.6 Hz, ³ $J_{3,4}$ = 9.4 Hz, H-3), 3.41 (dd, 1H, ³ $J_{3',2'}$ = 3.4 Hz, ³ $J_{3',4'}$ = 9.3 Hz, H-3'), 3.31 (dt, 1H, J = 5.0 Hz, J = 9.8 Hz, H-5'), 2.79-2.59 (s, 8H, 4 \times CH₂, Lev), 2.14 (s, 3H, CH₃, Lev), 2.11 (s, 3H, CH₃, Lev), 1.09, 1.08, 0.99, 0.98 (4 \times s, 36H, 12 \times CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ 206.30, 205.76 (2 \times CO), 171.90, 171.86 (2 \times CO), 138.85, 138.16 (2 \times Cq, CH₂Ph), 128.36, 128.19, 127.55, 127.44, 127.33, 127.28 (10 \times CH, CH₂Ph), 98.11 (C-1), 97.78 (C-1'), 77.52 (C-3'), 76.11 (C-3), 74.37, 73.96 (C-4, C-4'), 72.42, 71.67 (2 \times CH₂Ph), 71.54 (C-5'), 69.74 (C-2'), 68.99 (C-2), 68.23 (C-5), 66.43, 66.25 (C-6, C-6'), 38.07, 38.04 (2 \times CH₂, Lev), 29.74 (2 \times CH₃, Lev), 28.11, 28.06 (2 \times CH₂, Lev), 27.43, 27.38, 27.06, 27.03 (12 \times CH₃, DTBS), 22.69, 22.64, 20.01, 19.92 (4 \times Cq, DTBS); HRMS (ESI): m/z calcd for C₅₂H₇₈O₁₅Si₂+Na⁺: 1021.4771 [M+Na⁺]; found 1021.4771.

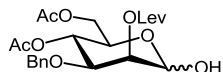
Synthesis of 25



Ethyl 4,6-di-*O*-acetyl-3-*O*-benzyl-2-*O*-levulinoyl-1-thio- α -D-mannopyranoside (25).

A solution of **14** (89 mg, 161 μ mol) in dry THF (2 ml) was stirred with solution of HF \cdot Py (200 μ L, 70%) in a PTFE-vial at r. t. for 45 min. under atmosphere of Ar. The mixture was diluted with EtOAc (20 ml) and washed with sat. aq. NaHCO₃ (2 \times 20 ml) and brine (20 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated to afford the intermediate 4,6-diol (R_f = 0.31, EtOAc). The diol (74 mg) was dissolved in pyridine (1.5 ml) and acetic acid anhydride (111 μ L, 1.18 mmol) and DMAP (catalytic amount) were successively added at 0 $^{\circ}$ C. The mixture was stirred at r. t. for 2 h, diluted with EtOAc (20 ml) and washed with aq. HCl (2 M, 10 ml), sat. aq. NaHCO₃ (20 ml) and brine (20 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 2 : 1) to afford **25** (68 mg, 137 μ mol, 85%, 2 steps) as a syrup. R_f = 0.56 (toluene – EtOAc, 1 : 2); $[\alpha]_D^{20} = +2$ (c 0.7, CHCl₃); ¹H-NMR (300 MHz, CDCl₃): δ 7.36-7.23 (m, 5H, CH₂Ph), 5.42 (dd, 1H, ³ $J_{2,1}$ = 1.7 Hz, ³ $J_{2,3}$ = 3.3 Hz, H-2), 5.30 (d, 1H, ³ $J_{1,2}$ = 1.6 Hz, H-1), 5.21 (t, 1H, ³ $J_{4,3}$ = ³ $J_{4,5}$ = 9.6 Hz, H-4), 4.60 (AB, 1H, ² J = 12.1 Hz, CH₂Ph), 4.38 (AB, 1H, ² J = 12.1 Hz, CH₂Ph), 4.29-4.22 (m, 2H, H-5, H-6a), 4.08 (dd, 1H, J = 5.0 Hz, J = 14.7 Hz, H-6b), 3.77 (dd, 1H, ³ $J_{3,2}$ = 3.3 Hz, ³ $J_{3,4}$ = 9.7 Hz, H-3), 2.79-2.53 (m, 6H, 3 \times CH₂, Lev, SET), 2.17 (s, 3H, CH₃, Lev), 2.08 (s, 3H, CH₃, Ac), 2.01 (s, 3H, CH₃, Ac), 1.29 (t, 3H, ³ J = 7.4 Hz, CH₃, SET); ¹³C-NMR (75 MHz, CDCl₃): δ 206.22 (CO, Lev), 171.83, 170.64, 169.66 (3 \times CO, Lev, Ac), 137.44 (Cq, CH₂Ph), 128.38, 127.86 (5 \times CH, CH₂Ph), 82.46 (C-1), 74.91 (C-3), 71.32 (CH₂Ph), 70.03 (C-2), 69.07 (C-5), 67.59 (C-4), 62.70 (C-6), 38.00 (CH₂, Lev), 29.74 (CH₃, Lev), 28.18 (CH₂, Lev), 25.61 (CH₂, SET), 20.80, 20.73 (2 \times CH₃, Ac), 14.84 (CH₃, SET); HRMS (ESI): m/z calcd for C₂₄H₃₂O₉S+Na⁺: 519.1659 [M+Na⁺]; found 519.1654.

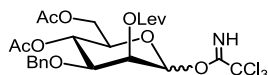
Synthesis of 26



4,6-di-O-Acetyl-3-O-benzyl-2-O-levulinoyl-D-mannopyranose (26).

Compound **25** (62 mg, 125 μ mol) was dissolved in acetone - water (2 ml, 24 : 1) and added to a solution of NBS (111 mg, 941 μ mol) in the same solvent mixture (2 ml) at 0 °C. The mixture was stirred for 10 min and then the reaction was quenched by addition of aq. 5% Na₂S₂O₃ (2 ml). The mixture was warmed to r. t., diluted with EtOAc (10 ml) and washed with sat. aq. NaHCO₃ (20 ml), water (20 ml) and brine (20 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 1 : 1) to afford **26** (50 mg, 111 μ mol, 89%, $\alpha/\beta = 7 : 1$) as a syrup. $R_f = 0.44$ (toluene – EtOAc, 1 : 5); $[\alpha]_D^{20} = 1$ (*c* 1.4, CHCl₃); ¹H-NMR (300 MHz, CDCl₃, α -anomer): δ 7.36-7.23 (m, 5H, CH₂Ph), 5.37 (dd, 1H, ³J_{2,1} = 1.9 Hz, ³J_{2,3} = 3.3 Hz, H-2), 5.25 (d, 1H, ³J_{1,2} = 1.9 Hz, H-1), 5.21 (t, 1H, ³J_{4,3} = ³J_{4,5} = 9.7 Hz, H-4), 4.63 (AB, 1H, ²J = 12.1 Hz, CH₂Ph), 4.41 (AB, 1H, ²J = 12.1 Hz, CH₂Ph), 4.21 (dd, 1H, *J* = 5.8 Hz, *J* = 12.5 Hz, H-6a), 4.15-4.07 (m, 2H, H-5, H-6b), 3.93 (dd, 1H, ³J_{3,2} = 3.4 Hz, ³J_{3,4} = 9.7 Hz, H-3), 2.90-2.58 (m, 4H, 2 \times CH₂, Lev), 2.16 (s, 3H, CH₃, Lev), 2.09 (s, 3H, CH₃, Ac), 2.01 (s, 3H, CH₃, Ac); ¹³C-NMR (75 MHz, CDCl₃, α -anomer): δ 206.47 (CO, Lev), 171.97, 170.81, 169.72 (3 \times CO, Lev, Ac), 137.71 (Cq, CH₂Ph), 128.33, 127.75, 127.72 (5 \times CH, CH₂Ph), 92.42 (C-1), 74.00 (C-3), 71.28 (CH₂Ph), 68.74, 68.67 (C-2, C-5), 67.42 (C-4), 62.86 (C-6), 38.02 (CH₂, Lev), 29.72 (CH₃, Lev), 28.16 (CH₂, Lev), 20.82, 20.79 (2 \times CH₃, Ac); HRMS (ESI): *m/z* calcd for C₂₂H₂₈O₁₀+COOH⁺: 497.1664 [M+COOH⁺]; found 497.1670.

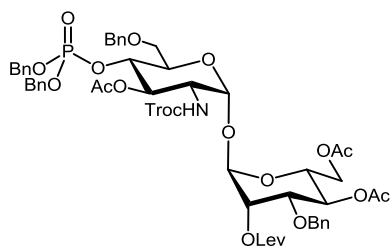
Synthesis of 27



4,6-di-O-Acetyl-3-O-benzyl-2-O-levulinoyl-D-mannopyranose trichloroacetimidate (27).

To a solution of **26** (36 mg, 80 μ mol) in dry CH₂Cl₂ (1 ml) trichloroacetonitrile (70 μ L, 718 μ mol) and DBU (2 μ L, 14 μ mol) were added successively at 0 °C. The mixture was stirred for 3.5 h at 0 °C, then warmed to r. t., diluted with EtOAc (10 ml) and washed with sat. aq. NaHCO₃ (10 ml) and brine (10 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 2 : 1 supplemented by 1% Et₃N) to afford **27** (32 mg, 54 μ mol, 67%, $\alpha/\beta = 32 : 1$) as a yellow syrup. $R_f = 0.53$ (hexane – EtOAc, 1 : 1 + 1% Et₃N); ¹H-NMR (600 MHz, CDCl₃, α -anomer): δ 8.75 (s, 1H, NH), 7.34-7.24 (m, 5H, CH₂Ph), 6.28 (d, 1H, ³J_{1,2} = 1.7 Hz, H-1), 5.48 (dd, 1H, ³J_{2,1} = 2.1 Hz, ³J_{2,3} = 3.1 Hz, H-2), 5.30 (t, 1H, ³J_{4,3} = ³J_{4,5} = 10.1 Hz, H-4), 4.64 (AB, 1H, ²J = 12.1 Hz, CH₂Ph), 4.44 (AB, 1H, ²J = 12.1 Hz, CH₂Ph), 4.21 (dd, 1H, ³J_{6a,5} = 5.3 Hz, ²J_{6a,6b} = 12.4 Hz, H-6a), 4.13 (dd, 1H, ³J_{6b,5} = 2.3 Hz, ²J_{6b,6a} = 12.4 Hz, H-6b), 4.04 (ddd, 1H, ³J_{5,4} = 10.2 Hz, ³J_{5,6a} = 5.2 Hz, ³J_{5,6b} = 2.2 Hz, H-5), 3.92 (dd, 1H, ³J_{3,2} = 3.3 Hz, ³J_{3,4} = 9.8 Hz, H-3), 2.82-2.69 (m, 4H, 2 \times CH₂, Lev), 2.18 (s, 3H, CH₃, Lev), 2.06 (s, 3H, CH₃, Ac), 2.04 (s, 3H, CH₃, Ac); ¹³C-NMR (75 MHz, CDCl₃, α -anomer): δ 206.01 (CO, Lev), 171.64, 170.58, 169.53 (3 \times CO, Lev, Ac), 159.53 (OC(NH)CCl₃), 137.11 (Cq, CH₂Ph), 128.40, 128.06, 128.00 (5 \times CH, CH₂Ph), 94.76 (C-1), 90.58 (CCl₃), 73.64 (C-3), 71.48 (CH₂, CH₂Ph), 71.30 (C-5), 66.89 (C-2), 66.58 (C-4), 62.26 (C-6), 37.93 (CH₂, Lev), 29.69 (CH₃, Lev), 28.03 (CH₂, Lev), 20.75, 20.67 (2 \times CH₃, Ac).

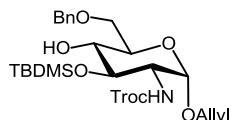
Synthesis of 28



4,6-di-*O*-Acetyl-3-*O*-benzyl-2-*O*-levulinoyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-3-*O*-acetyl-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (28).

A solution of glycosyl donor **27** (32 mg, 54 μ mol) and glycosyl acceptor **8** (33 mg, 44 μ mol) in dry CH₂Cl₂ (1 ml) was stirred with activated powdered 4 Å molecular sieves was stirred for 1.5 h. at r. t. under atmosphere of Ar. The mixture was cooled to 0 °C and a solution of TMSOTf (0.5 μ L, 3 μ mol) in dry CH₂Cl₂ (50 μ L of a stock solution prepared from 10 μ L TMSOTf in 1 ml CH₂Cl₂) was added. The mixture was stirred at 0 °C for 3 h. and then the reaction was quenched by addition of Et₃N (10 μ L). The mixture was warmed to r. t., diluted with EtOAc (10 ml), filtered over a pad of Celite and the filtrate was concentrated. The residue was purified by HPLC (two successive columns: CH₂Cl₂ – MeOH, 50 : 1 and hexane – EtOAc, 2 : 1 \rightarrow 1 : 2 \rightarrow 1 : 1, column B) to afford **28** (5 mg, 4 μ mol, 8%) as a syrup. R_f = 0.59 (hexane - EtOAc, 1 : 2); $[\alpha]_D^{20}$ = +33 (*c* 0.4, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.37-7.23 (m, 20H, CH₂Ph), 5.29 (t, 1H, ³*J*_{3,2} = ³*J*_{3,4} = 10.0 Hz, H-3), 5.23 (d, 1H, ³*J*_{1,2} = 3.6 Hz, H-1), 5.22 (br s, 1H, H-2'), 5.18 (d, 1H, ³*J*_{NH,2} = 8.8 Hz, NH), 5.15 (t, 1H, ³*J*_{4,3'} = ³*J*_{4,5'} = 10.0 Hz, H-4'), 5.11 (br s, 1H, H-1'), 5.00-4.92 (m, 4H, OP(O)(OCH₂Ph)₂), 4.71-4.65 (m, 4H, H-4, CH₂Ph or Troc), 4.54-4.46 (m, 3H, CH₂Ph or Troc), 4.14 (dd, 1H, *J* = 6.5 Hz, *J* = 12.2 Hz, H-6a'), 4.06-4.02 (m, 2H, H-2, H-6b'), 3.89 (dd, 1H, *J* = 3.1 Hz, *J* = 9.6 Hz, H-3'), 3.82 (br d, 1H, *J* = 9.6 Hz, H-5), 3.78-3.75 (m, 1H, H-5'), 3.71 (m, 2H, H-6a, H-6b), 2.73-2.63 (m, 4H, 2 \times CH₂, Lev), 2.12 (s, 3H, CH₃, Lev), 2.05 (s, 3H, CH₃, Ac), 2.02 (s, 3H, CH₃, Ac), 1.96 (s, 3H, CH₃, Ac); ¹³C-NMR (151 MHz, CDCl₃): δ 206.00 (CO, Lev), 171.87, 171.84, 170.55, 169.63 (4 \times CO, Lev, Ac), 153.93 (CO, Troc), 137.90, 137.31 (2 \times Cq, CH₂Ph), 135.47 (2 \times Cq, OP(O)(OCH₂Ph)₂), 128.71, 128.64, 128.50, 128.34, 128.06, 127.99, 127.63, 127.59 (20 \times CH, CH₂Ph), 95.21 (CCl₃, Troc), 92.58 (C-1', ¹*J*_{C,H} = 174 Hz), 91.95 (C-1, ¹*J*_{C,H} = 178 Hz), 74.74 (CH₂, CH₂Ph or Troc), 74.12 (C-3'), 73.50 (CH₂, CH₂Ph or Troc), 73.04 (C-4, ²*J*_{C4,P} = 6.2 Hz), 72.01 (CH₂, CH₂Ph or Troc), 70.71 (C-3), 70.51 (C-5, ³*J*_{C5,P} = 5.9 Hz), 69.87 (C-5'), 69.71, 69.69, 69.68, 69.66 (2 \times CH₂, OP(O)(OCH₂Ph)₂), 68.36 (C-2'), 67.54 (C-6), 67.16 (C-4'), 62.47 (C-6'), 53.81 (C-2), 37.93 (CH₂, Lev), 29.67 (CH₃, Lev), 28.09 (CH₂, Lev), 20.81, 20.79, 20.70 (3 \times CH₃, Ac); ³¹P-NMR (243 MHz, CDCl₃): δ -2.26; HRMS (ESI): *m/z* calcd for C₅₄H₆₁Cl₃NO₂₀P+Na⁺: 1202.2482 [M+Na⁺]; found 1202.2477.

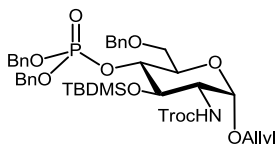
Synthesis of 30



Allyl 6-*O*-benzyl-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (30).

A solution of **29**¹⁷ (6.5 g, 10.9 mmol), trifluoroacetic anhydride (4.5 ml, 32.4 mmol), triethylsilane (8.8 ml, 54.5 mmol) and trifluoroacetic acid (4.0 ml, 54.0 mmol) in dry CH₂Cl₂ (50 ml) was stirred at 0 °C for 3 h. under atmosphere of Ar, then sat. aq. NaHCO₃ (50 ml) was added and the mixture was warmed to r. t. The mixture was diluted with EtOAc (200 ml) and washed with sat. aq. NaHCO₃ (200 ml), water (100 ml) and brine (100 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (toluene – EtOAc, 20 : 1) to afford **30** (5.9 g, 9.9 mmol, 90%) as a syrup. R_f = 0.23 (toluene–EtOAc, 20 : 1); $[\alpha]_D^{20}$ = +61 (*c* 1.2, CHCl₃); ¹H-NMR (400 MHz, CDCl₃): δ 7.38-7.28 (m, 5H, CH₂Ph), 5.90 (dddd, 1H, ³*J* = 5.3 Hz, ³*J* = 6.5 Hz, ³*J*_{cis} = 10.5 Hz, ³*J*_{trans} = 17.1 Hz, CH, Allyl), 5.28 (dq, 1H, ⁴*J* = 1.6 Hz, ³*J*_{trans} = 17.2 Hz, CH₂, Allyl), 5.21 (dq, 1H, ⁴*J* = 1.3 Hz, ³*J*_{cis} = 10.4 Hz, CH₂, Allyl), 5.05 (d, 1H, ³*J*_{NH,2} = 10.0 Hz, NH), 4.86 (d, 1H, ³*J*_{1,2} = 3.6 Hz, H-1), 4.78 (AB, ²*J* = 11.9 Hz, CH₂, Troc), 4.64 (AB, ²*J* = 12.1 Hz, CH₂Ph), 4.60 (AB, ²*J* = 11.9 Hz, CH₂, Troc), 4.56 (AB, ²*J* = 12.1 Hz, CH₂Ph), 4.17 (ddt, 1H, ⁴*J* = 1.5 Hz, ³*J* = 5.3, ²*J* = 12.9 Hz, CH₂, Allyl), 4.00 (ddt, 1H, ⁴*J* = 1.3 Hz, ³*J* = 6.4 Hz, ²*J* = 12.9 Hz, CH₂, Allyl), 3.85 (td, 1H, ³*J*_{2,1} = 3.6 Hz, ³*J*_{2,3} = ³*J*_{2,NH} = 10.1 Hz, H-2), 3.77-3.66 (m, 4H, H-3, H-5, H-6a, H-6b), 3.58 (td, 1H, ³*J*_{4,OH} = 3.1 Hz, ³*J*_{4,5} = ³*J*_{4,3} = 8.6 Hz, H-4), 2.36 (d, 1H, ³*J*_{OH,4} = 3.0 Hz, 4-OH), 0.88 (s, 9H, 3 \times CH₃, TBDMS), 0.12 (s, 3H, CH₃, TBDMS), 0.09 (s, 3H, CH₃, TBDMS); ¹³C-NMR (101 MHz, CDCl₃): δ 154.12 (CO, Troc), 137.87 (Cq, CH₂Ph), 133.57 (CH, Allyl), 128.45, 127.79, 127.71 (5 \times CH, CH₂Ph), 117.96 (CH₂, Allyl), 96.96 (C-1), 74.96 (CH₂, Troc), 73.69 (C-3), 73.65 (CH₂Ph), 72.85 (C-4), 70.20 (C-5), 69.96 (C-6), 68.31 (CH₂, Allyl), 55.60 (C-2), 25.76 (3 \times CH₃, TBDMS), 18.12 (Cq, TBDMS), -4.09, -4.62 (2 \times CH₃, TBDMS); HRMS (ESI): *m/z* calcd for C₂₅H₃₈Cl₃NO₇Si+Na⁺: 620.1375 [M+Na⁺]; found 620.1376.

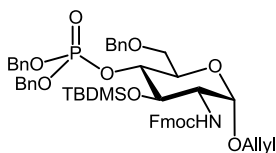
Synthesis of 31



Allyl 6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (**31**).

To a stirred solution of **30** (5.92 g, 9.88 mmol) and dibenzyl *N,N*-diisopropylphosphoramidite (6.30 ml, 16.9 mmol, 90%) in dry CH_2Cl_2 (50 ml) a solution of 1*H*-tetrazole (43.80 ml, 19.7 mmol, 0.45 M in CH_3CN) was added at r. t. under atmosphere of Ar. After 2 h the mixture was cooled to -78°C and a solution of *m*CPBA (3.60 g, 14.5 mmol, 70%) in CH_2Cl_2 (50 ml) was added. The reaction mixture was stirred for 1 h. to -78°C , then quenched by addition of Et_3N (3.20 ml) and warmed to r. t. The mixture was diluted with EtOAc (200 ml) and washed with aq. citric acid (0.25 M, 100 ml), sat. aq. NaHCO_3 (100 ml), water (100 ml) and brine (100 ml). The organic layer was dried over Na_2SO_4 , filtered and concentrated. The residue was purified by MPLC (toluene – EtOAc, 10 : 1 \rightarrow 7 : 1) to afford **31** (7.77 g, 9.04 mmol, 91%) as a syrup. $R_f = 0.45$ (hexane – EtOAc, 2 : 1); $[\alpha]_{\text{D}}^{20} = +46.0$ (c 1.1, CHCl_3); $^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 7.34-7.23 (m, 15H, CH_2Ph), 5.91 (dddd, 1H, $^3J = 5.1$ Hz, $^3J = 6.3$ Hz, $^3J_{\text{cis}} = 10.7$ Hz, $^3J_{\text{trans}} = 16.9$ Hz, CH, Allyl), 5.29 (dq, 1H, $^4J = 1.5$ Hz, $^3J_{\text{trans}} = 17.2$ Hz, CH_2 , Allyl), 5.23 (dq, 1H, $^4J = 1.3$ Hz, $^3J_{\text{cis}} = 10.5$ Hz, CH_2 , Allyl), 5.03 (d, 1H, $^3J_{\text{NH}_2} = 9.7$ Hz, NH), 4.98-4.87 (m, 5H, H-1, $\text{OP}(\text{O})(\text{OCH}_2\text{Ph})_2$), 4.80 (AB, 1H, $^2J = 11.9$ Hz, CH_2 Troc or CH_2Ph), 4.59 (m, 2H, CH_2 Troc or CH_2Ph), 4.49 (AB, $^2J = 12.0$ Hz, CH_2 Troc or CH_2Ph), 4.32 (dt, $J = 9.6$ Hz, $J = 7.9$ Hz, H-4), 4.20 (ddt, 1H, $^4J = 1.4$ Hz, $^3J = 5.2$ Hz, $^2J = 12.9$ Hz, CH_2 , Allyl), 4.03 (ddt, 1H, $^4J = 1.2$ Hz, $^3J = 6.3$ Hz, $^2J = 13.1$ Hz, CH_2 , Allyl), 3.93-3.85 (m, 4H, H-2, H-3, H-5, H-6a), 3.82 (dd, 1H, $J = 2.3$ Hz, $J = 10.7$ Hz, H-6b), 0.85 (s, 9H, $3 \times \text{CH}_3$, TBDMS), 0.13 (s, 3H, CH_3 , TBDMS), 0.10 (s, 3H, CH_3 , TBDMS); $^{13}\text{C-NMR}$ (151 MHz, CDCl_3): δ 153.99 (CO, Troc), 138.42 (Cq, CH_2Ph), 135.78, 135.72 ($2 \times \text{Cq}$, $\text{OP}(\text{O})(\text{OCH}_2\text{Ph})_2$), 133.46 (CH, Allyl), 128.50, 128.46, 128.23, 128.16, 128.02, 127.54, 127.44, 127.37 ($15 \times \text{CH}$, CH_2Ph), 118.03 (CH_2 , Allyl), 96.13 (C-1), 95.18 (CCl_3 , Troc), 76.93 (C-4, $^2J_{\text{C}_4,\text{P}} = 6.6$ Hz), 75.02, 73.00 (CH_2Ph , CH_2 Troc), 71.62 (C-3, $^3J_{\text{C}_3,\text{P}} = 4.7$ Hz), 70.65 (C-5), 69.80 ($\text{OP}(\text{O})(\text{OCH}_2\text{Ph})_2$, $^2J_{\text{C},\text{P}} = 5.2$ Hz), 69.34 ($\text{OP}(\text{O})(\text{OCH}_2\text{Ph})_2$, $^2J_{\text{C},\text{P}} = 4.7$ Hz), 68.60 (C-6), 68.37 (CH_2 , Allyl), 55.79 (C-2), 25.83 ($3 \times \text{CH}_3$, TBDMS), 17.82 (Cq, TBDMS), -4.07 ($2 \times \text{CH}_3$, TBDMS); $^{31}\text{P-NMR}$ (243 MHz, CDCl_3): δ -1.87; HRMS (ESI): m/z calcd for $\text{C}_{39}\text{H}_{51}\text{Cl}_3\text{NO}_{10}\text{PSi} + \text{Na}^+$: 880.1978 [$\text{M} + \text{Na}^+$]; found 880.1975.

Synthesis of 32

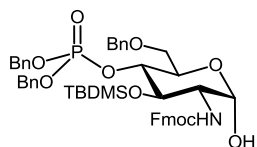


Allyl 6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)- α -D-glucopyranoside (**32**).

To a solution of **31** (5.05 g, 5.88 mmol) in dry CH_2Cl_2 (100 ml), Zn dust (16.0 g, 10 μm) and dry acetic acid (50 ml) were added at 0°C under atmosphere of Ar. The mixture was stirred for 3.5 h at 0°C , then diluted with CH_2Cl_2 (200 ml), filtered over a pad of Celite and the filtrate was washed with sat. aq. NaHCO_3 (2×200 ml) and water (200 ml). The organic layer was dried over Na_2SO_4 , filtered and concentrated to afford crude amine (3.34 g) as a syrup. $R_f = 0.21$ (hexane – EtOAc, 1 : 1). The crude amine (2.93 g, 4.28 mmol) was dissolved in dry DMF (15 ml) and diisopropylethylamine (1.12 ml, 6.41 mmol) and 9-fluorenylmethyl chloroformate (1.66 g, 6.42 mmol) were added at 0°C under atmosphere of Ar. The mixture was stirred for 2 h. at r. t., diluted with EtOAc (200 ml) and washed with water (4×100 ml) and brine (200 ml). The organic layer was dried over Na_2SO_4 , filtered and concentrated. The residue was purified by silica gel chromatography (hexane – EtOAc, 5 : 1 \rightarrow 4 : 1 \rightarrow 3 : 1) to afford **32** (3.41 g, 3.76 mmol, 87%) as a syrup. $R_f = 0.23$ (hexane - EtOAc, 3 : 1); $[\alpha]_{\text{D}}^{20} = +42$ (c 1.0, CHCl_3); $^1\text{H-NMR}$ (400 MHz, CDCl_3): δ 7.76 (d, 2H, $J = 7.5$ Hz, arom.), 7.61-7.58 (m, 2H, arom.), 7.41-7.21 (m, 19H, arom.), 5.90 ($^3J = 5.3$ Hz, $^3J = 6.4$ Hz, $^3J_{\text{cis}} = 10.5$ Hz, $^3J_{\text{trans}} = 17.1$ Hz, CH, Allyl), 5.27 (dq, $^4J = 1.6$ Hz, $^3J_{\text{trans}} = 17.2$ Hz,

CH₂, Allyl), 5.21 (dq, ⁴J = 1.3 Hz, ³J_{cis} = 10.4 Hz, CH₂, Allyl), 4.97-4.78 (m, 5H, H-1, OP(O)(OCH₂Ph)₂), 4.58 (AB, 1H, ²J = 12.0 Hz, CH₂Ph), 4.50-4.46 (m, 1H, CH₂, Fmoc), 4.47 (AB, 1H, ²J = 12.0 Hz, CH₂Ph), 4.33-4.26 (m, 2H, H-4, CH₂, Fmoc), 4.21-4.16 (m, 2H, CH₂ Allyl, CH, Fmoc), 4.03-3.96 (m, 1H, CH₂, Allyl), 3.92-3.79 (m, 5H, H-2, H-3, H-5, H-6a, H-6b), 0.80 (s, 9H, 3×CH₃, TBDMS), 0.09 (s, 3H, CH₃, TBDMS), 0.01 (s, 3H, CH₃, TBDMS); ¹³C-NMR (101 MHz, CDCl₃): δ 155.73 (CO, Fmoc), 143.87, 141.31 (4×C_q, Fmoc), 138.44 (C_q, CH₂Ph), 135.80, 135.75 (2×C_q, OP(O)(OCH₂Ph)₂), 133.61 (CH, Allyl), 128.48, 128.21, 128.14, 128.00, 127.68, 127.43, 127.34, 127.02, 126.95, 125.14, 124.92, 119.96, 119.93 (23×CH, arom.), 117.81 (CH₂, Allyl), 96.53 (C-1), 77.09 (C-4, ²J_{C4,P} = 6.6 Hz), 72.98 (CH₂Ph), 71.80 (C-5, ³J_{C5,P} = 4.7 Hz), 70.51 (C-3), 69.77 (OP(O)(OCH₂Ph)₂, ²J = 5.5 Hz), 69.30 (OP(O)(OCH₂Ph)₂, ²J = 4.8 Hz), 68.69, 68.35 (C-6, CH₂ Allyl), 66.95 (CH₂, Fmoc), 55.50 (C-2), 47.17 (CH, Fmoc), 25.78 (3×CH₃, TBDMS), 17.84 (C_q, TBDMS), -4.05, -4.11 (2×CH₃, TBDMS); ³¹P-NMR (162 MHz, CDCl₃): δ -1.89; HRMS (ESI): *m/z* calcd for C₅₁H₆₀NO₁₀PSi+Na⁺: 928.3616 [M+Na⁺]; found 928.3622.

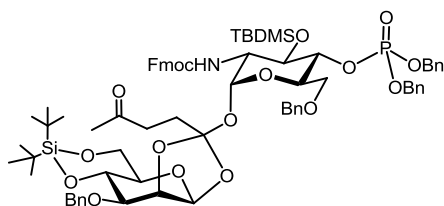
Synthesis of 33



6-*O*-Benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)-α-D-glucopyranose (33).

A degassed solution of H₂-activated (1,5-Cyclooctadiene)bis(methyldiphenylphosphine)iridium(I) hexafluorophosphate (191 mg, 226 μmol) in dry THF (30 ml) was added to a degassed solution of **32** (2.05 g, 2.26 mmol) in dry THF (100 ml) under atmosphere of Ar. The mixture was stirred for 4 h. at r. t., cooled to 0°C and a solution of I₂ (1.07 g, 4.21 mmol) in THF - H₂O (1 : 1, 16 ml) was added. The mixture was stirred for 3 h. at 0 °C, diluted with CH₂Cl₂ (200 ml) and washed with aq. 5% Na₂S₂O₃ (100 ml). The aqueous phase was extracted with CH₂Cl₂ (2×100 ml) and the combined organic phases were washed with sat. aq. NaHCO₃ (100 ml) and water (100 ml), dried over MgSO₄, filtered and concentrated. The residue was purified by silica gel chromatography (toluene – EtOAc, 3 : 1 → 1 : 1) to afford **33** (1.70 g, 1.96 mmol, 86%) as a syrup. *R*_f = 0.23 (hexane - EtOAc, 1 : 1); [α]_D²⁰ = +22.0 (*c* 1.1, CHCl₃); ¹H-NMR (400 MHz, CDCl₃): δ 7.75 (d, 2H, *J* = 7.6 Hz, arom.), 7.58 (t, 2H, *J* = 6.6 Hz, arom.), 7.38 (t, 2H, *J* = 7.4 Hz, arom.), 7.32-7.20 (m, 17H, arom.), 5.15 (t, 1H, ³J_{1,2} = ³J_{1,OH} = 3.6 Hz, H-1), 4.95-4.83 (m, 5H, NH, OP(O)(OCH₂Ph)₂), 4.55- 4.43 (m, 3H, CH₂Ph, CH₂ Fmoc), 4.32 (dd, 1H, *J* = 6.8 Hz, *J* = 10.6 Hz, CH₂, Fmoc), 4.20-4.09 (m, 3H, H-4, H-5, CH Fmoc), 3.91 (dd, 1H, *J* = 7.2 Hz, *J* = 8.9 Hz, H-3), 3.84-3.78 (m, 2H, H-2, H-6a), 3.73 (dd, 1H, *J* = 6.1 Hz, *J* = 10.4 Hz, H-6b), 3.38 (d, 1H, ³J_{OH,1} = 3.9 Hz, 1-OH), 0.80 (s, 9H, 3×CH₃, TBDMS), 0.06 (s, 3H, CH₃, TBDMS), -0.01 (s, 3H, CH₃, TBDMS); ¹³C-NMR (101 MHz, CDCl₃): δ 155.96 (CO, Fmoc), 143.88, 143.81, 141.32 (4×C_q, Fmoc), 138.03 (C_q, CH₂Ph), 135.69, 135.63 (2×C_q, OP(O)(OCH₂Ph)₂), 128.53, 128.31, 128.15, 128.03, 127.78, 127.68, 127.60, 127.07, 127.00, 125.12, 124.92, 119.95 (23×CH, arom.), 91.04 (C-1), 77.32-76.68 (C-4), 73.17 (CH₂Ph), 71.27, 70.98 (C-3, C-4), 69.76 (OP(O)(OCH₂Ph)₂, ²J = 5.4 Hz), 69.39 (OP(O)(OCH₂Ph)₂, ²J = 5.1 Hz), 68.80 (C-6), 66.92 (CH₂, Fmoc), 55.39 (C-2), 47.16 (CH, Fmoc), 25.75 (3×CH₃, TBDMS), 17.82 (C_q, TBDMS), -4.25, -4.35 (2×CH₃, TBDMS); ³¹P-NMR (243 MHz, CDCl₃): δ -1.76; HRMS (ESI): *m/z* calcd for C₄₈H₅₆NO₁₀PSi+Na⁺: 888.3303 [M+Na⁺]; found 888.3295.

Synthesis of 34

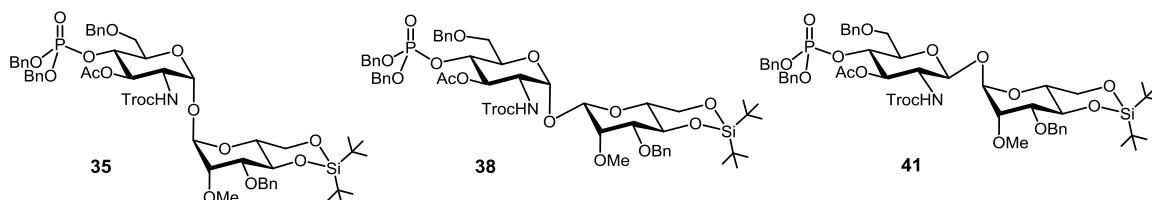


Orthoester (34).

A solution of glycosyl donor **18** (60 mg, 92 μmol) and glycosyl acceptor **33** (110 mg, 127 μmol) in dry CH₂Cl₂ (1 ml) was stirred with activated powdered 4 Å molecular sieves for 1.5 h. at r. t. under atmosphere of Ar. The

mixture was cooled to 0 °C and a solution of TMSOTf (0.83 μ L, 4.6 μ mol) in dry CH₂Cl₂ (42 μ L of a stock solution prepared from 10 μ L TMSOTf in 500 μ L CH₂Cl₂) was added. The mixture was stirred at 0 °C for 40 min and then the reaction was quenched by addition of Et₃N (10 μ L). The mixture was warmed to r. t., diluted with EtOAc (20 ml), filtered over a pad of Celite and the filtrate was washed with water (10 ml) and brine (10 ml). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by HPLC (two successive columns: toluene – EtOAc, 7 : 1 \rightarrow 1 : 1 and hexane : EtOAc, 7 : 1 \rightarrow 3 : 1, column B) to afford **34** (51 mg, 38 μ mol, 40%) as a colorless syrup. R_f = 0.17 (hexane – EtOAc, 3 : 1); $[\alpha]_D^{20}$ = +18 (*c* 1.0, DCM); ¹H-NMR (600 MHz, CDCl₃): δ = 7.74-7.66 (m, 4H, arom.), 7.44-7.27 (m, 19H, arom.), 7.16-7.09 (m, 3H, arom.), 6.99-6.98 (m, 2H, arom.) 5.33 (d, 1H, ³ $J_{1,2}$ = 3.7 Hz, H-1), 5.24 (d, 1H, ³ $J_{NH,2}$ = 10.3 Hz, NH), 5.04-4.83 (m, 6H, 3 \times CH₂, CH₂Ph, OP(O)(OCH₂Ph)₂), 4.62 (dd, 1H, ³ $J_{2',1'}$ = 2.2 Hz, ³ $J_{2',3'}$ = 3.9 Hz, H-2'), 4.50 (d, 1H, ³ $J_{1',2'}$ = 2.0 Hz, H-1'), 4.42 (dd, 1H, ³ J = 6.9 Hz, ² J = 10.4 Hz, CH₂, Fmoc), 4.29 (dd, 1H, ³ J = 7.4 Hz, ² J = 10.4 Hz, CH₂, Fmoc), 4.23 (AB, 1H, ² J = 10.3 Hz, CH₂Ph), 4.20 (t, ³ J = 7.2 Hz, CH, Fmoc), 4.17 (AB, 1H, ² J = 10.3 Hz, CH₂Ph), 4.13-4.07 (m, 2H, H-4, H-5), 4.03 (t, 1H, ³ $J_{4',3'}$ = ³ $J_{4',5'}$ = 9.3 Hz, H-4'), 3.98 (dd, 1H, ³ $J_{6a',5'}$ = 5.1 Hz, ² $J_{6a',6b'}$ = 10.4 Hz, H-6a'), 3.95-3.93 (m, 1H, H-6a), 3.90 (td, 1H, ³ $J_{2,1}$ = 3.6 Hz, ³ $J_{2,3}$ = ³ $J_{2,NH}$ = 10.2 Hz, H-2), 3.77-3.72 (m, 2H, H-3, H-6b'), 3.53 (dd, 1H, ³ $J_{6b,5}$ = 7.7 Hz, ² $J_{6b,6a}$ = 10.1 Hz, H-6b), 3.15 (dd, 1H, ³ $J_{3',2'}$ = 4.1 Hz, ³ $J_{3',4'}$ = 9.2 Hz, H-3'), 2.75 (ddd, 1H, J = 5.6 Hz, J = 8.0 Hz, J = 18.1 Hz, CH₂, Lev), 2.65 (td, 1H, ³ $J_{5',6a'}$ = 5.1 Hz, ³ $J_{5',4'}$ = ³ $J_{5',6b'}$ = 9.8 Hz, H-5'), 2.52 (dt, 1H, J = 6.2 Hz, J = 18.1 Hz, CH₂, Lev), 2.43 (ddd, 1H, J = 5.7 Hz, J = 8.2 Hz, J = 14.0 Hz, CH₂, Lev), 2.11 (s, 3H, CH₃, Lev), 2.01-1.93 (m, 1H, CH₂, Lev), 1.08 (s, 9H, 3 \times CH₃, DTBS), 1.03 (s, 9H, 3 \times CH₃, DTBS), 0.82 (s, 9H, 3 \times CH₃, TBDMS), 0.11 (s, 3H, CH₃, TBDMS), 0.07 (s, 3H, CH₃, TBDMS); ¹³C-NMR (151 MHz, CDCl₃): δ 207.86 (CO, Lev), 171.06 (CO, Lev), 155.95 (CO, Fmoc), 144.11, 143.90, 141.26, 141.22 (4 \times Cq, Fmoc), 138.34, 138.18 (2 \times Cq, CH₂Ph), 135.76, 135.72, 135.67 (2 \times Cq, OP(O)(OCH₂Ph)₂), 128.59, 128.56, 128.52, 128.45, 128.27, 128.25, 128.01, 127.83, 127.72, 127.58, 127.52, 127.07, 126.98, 125.49, 125.20 (26 \times CH, arom.), 123.61 (Cq orthoester), 119.81, 119.76 (2 \times CH, Fmoc), 97.09 (C-1', ¹ $J_{C,H}$ = 180 Hz), 91.62 (C-1, ¹ $J_{C,H}$ = 172 Hz), 78.94 (C-2'), 77.82 (C-4, ² $J_{C4,P}$ = 6.6 Hz), 76.23 (C-3'), 74.12 (C-4'), 73.41, 72.62 (2 \times CH₂, CH₂Ph), 71.65 (C-3, ³ $J_{C3,P}$ = 4.7 Hz), 70.32 (C-6), 70.24 (C-5), 69.82 ((OP(O)(OCH₂Ph)₂, ² $J_{C,P}$ = 5.3 Hz), 69.33 ((OP(O)(OCH₂Ph)₂, ² $J_{C,P}$ = 5.2 Hz), 68.63 (C-5'), 67.11 (CH₂, Fmoc), 66.24 (C-6'), 55.43 (C-2), 47.14 (CH, Fmoc), 37.95, 33.01 (2 \times CH₂, Lev), 30.15 (CH₃, Lev), 27.45, 27.01, 25.80 (9 \times CH₃, DTBS, TBDMS), 22.72, 19.86, 17.84 (3 \times Cq, DTBS, TBDMS), -4.03, -4.10 (2 \times SiCH₃, TBDMS); ³¹P-NMR (243 MHz, CDCl₃): δ -1.62; HRMS (ESI): *m/z* calcd for C₇₄H₉₄NO₁₇PSi₂+NH₄⁺: 1373.6136 [M+NH₄⁺]; found 1373.6057.

Synthesis of **35**, **38** and **41**



3-O-Benzyl-4,6-O-di-tert-butylsilylene-2-O-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-3-O-acetyl-6-O-benzyl-4-O-bis(benzyloxy)phosphoryl]-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (35**),**

3-O-benzyl-4,6-O-di-tert-butylsilylene-2-O-methyl- β -D-mannopyranosyl-(1 \leftrightarrow 1)-3-O-acetyl-6-O-benzyl-4-O-bis(benzyloxy)phosphoryl]-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (38**) and**

3-O-benzyl-4,6-O-di-tert-butylsilylene-2-O-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-3-O-acetyl-6-O-benzyl-4-O-bis(benzyloxy)phosphoryl]-2-deoxy-2-(2,2,2-trichloroethoxycarbonylamino)- β -D-glucopyranoside (41**).**

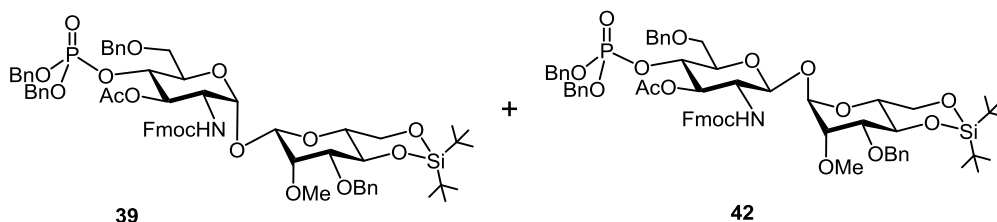
A solution of glycosyl donor **21** (72 mg, 121 μ mol) and glycosyl acceptor **8** (78 mg, 104 μ mol) in dry CH₂Cl₂ (2 ml) was stirred with powdered activated 4 Å molecular sieves for 2 h. at r. t. under atmosphere of Ar. The mixture was cooled to 0 °C and a solution of TMSOTf (1.1 μ L, 6 μ mol) in dry CH₂Cl₂ (55 μ L of a stock solution prepared from 10 μ L TMSOTf in 500 μ L CH₂Cl₂) was added. The mixture was stirred at 0 °C for 10 min and then the reaction was quenched by addition of Et₃N (10 μ L). The mixture was warmed to r. t., diluted with toluene (10 ml), filtered over a pad of Celite and the filtrate was concentrated. The residue was purified by HPLC (two successive columns: toluene – EtOAc, 4 : 1 \rightarrow 0:1 and hexane – EtOAc, 3 : 1 \rightarrow 2 : 1, column A) to afford **35** (62 mg, 54 μ mol, 51%), **38** (9 mg, 7.8 μ mol, 7 %) and **41** (4 mg, 3.5 μ mol, 3%) as syrups. **35**: R_f = 0.25 (hexane – EtOAc, 2 : 1); $[\alpha]_D^{20}$ = +64 (*c* 1.0,

CHCl₃); ¹H-NMR (600 MHz, CDCl₃) δ 7.44-7.42 (m, 2H, CH₂Ph), 7.37-7.33 (m, 9H, CH₂Ph), 7.30-7.22 (m, 9H, CH₂Ph), 5.20 (dd, 1H, ³J_{3,4} = 9.1 Hz, ³J_{3,2} = 10.9 Hz, H-3), 5.15 (d, 1H, ³J_{1,2} = 3.8 Hz, H-1), 5.09 (d, 1H, ³J_{1',2'} = 1.6 Hz, H-1'), 5.07 (AB, 1H, ²J = 12.1 Hz, CH₂Ph or Troc), 5.02 (AB, 1H, ²J = 12.0 Hz, CH₂Ph or Troc), 5.02-4.92 (m, 5H, NH, OP(O)(OCH₂Ph)₂), 4.80 (AB, 1H, ²J = 12.0 Hz, CH₂Ph or Troc), 4.50 (AB, 1H, ²J = 11.8 Hz, CH₂Ph or Troc), 4.49 (q, 1H, ³J_{4,3} = ³J_{4,5} = ³J_{4,P} = 9.2 Hz, H-4), 4.42 (AB, 1H, ²J = 11.8 Hz, CH₂Ph or Troc), 4.37 (AB, 1H, ²J = 12.1 Hz, CH₂Ph or Troc), 4.31 (t, 1H, ³J_{4',3'} = ³J_{4',5'} = 9.5 Hz, H-4'), 4.08 (ddd, 1H, ³J_{2,1} = 3.8 Hz, ³J_{2,3} = 10.9 Hz, ³J_{2,NH} = 9.5 Hz, H-2), 4.00 (dd, 1H, ³J_{6a',5'} = 4.9 Hz, ²J_{6a',6b'} = 10.2 Hz, H-6a'), 3.93 (t, 1H, ³J_{6b',5'} = ²J_{6b',6a'} = 10.4 Hz, H-6b'), 3.76-3.74 (m, 1H, H-5), 3.75 (dd, 1H, ³J_{6a,5} = 1.8 Hz, ²J_{6a,6b} = 11.3 Hz, H-6a), 3.68 (dd, 1H, ³J_{3',2'} = 3.3 Hz, ³J_{3',4'} = 9.5 Hz, H-3'), 3.65-3.62 (m, 1H, H-5'), 3.61 (dd, 1H, ³J_{6b,5} = 6.2 Hz, ²J_{6b,6a} = 11.2 Hz, H-6b), 3.38 (s, 3H, CH₃, Me), 3.37 (dd, 1H, ³J_{2',1'} = 1.7 Hz, ³J_{2',3'} = 3.2 Hz, H-2'), 1.89 (s, 3H, CH₃, Ac), 1.07 (s, 9H, 3×CH₃, DTBS), 1.01 (s, 9H, 3×CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): 171.33 (CO, Ac), 154.17 (CO, Troc), 138.72, 137.75 (2×Cq, CH₂Ph), 135.46 (Cq, ³J_{C,P} = 7.3 Hz, OP(O)(OCH₂Ph)₂), 135.37 (Cq, ³J_{C,P} = 6.9 Hz, OP(O)(OCH₂Ph)₂), 128.77, 128.76, 128.66, 128.39, 128.35, 128.05, 128.03, 127.99, 127.77, 127.71, 127.64 (20×CH, CH₂Ph), 93.72 (C-1', ¹J_{C,H} = 170 Hz), 92.96 (C-1, ¹J_{C,H} = 173 Hz), 79.62 (C-2'), 77.76 (C-3'), 74.73 (C-4'), 74.49, 74.13, 73.57 (3×CH₂, CH₂Ph, Troc), 73.46 (C-4, ²J_{C4,P} = 5.8 Hz), 71.12 (C-3), 70.65 (C-5, ³J_{C5,P} = 5.7 Hz), 69.76, 69.74, 69.72, 69.71 (2×CH₂, OP(O)(OCH₂Ph)₂), 69.30 (C-5'), 68.25 (C-6), 66.33 (C-6'), 59.69 (CH₃, Me), 53.76 (C-2), 27.45, 27.09 (6×CH₃, DTBS), 22.60 (Cq, DTBS), 20.67 (CH₃, Ac), 19.86 (Cq, DTBS); ³¹P-NMR (242 MHz, CDCl₃): δ -1.95; HRMS (ESI): *m/z* calcd for C₅₄H₆₉NO₁₆PSi+H⁺: 1152.3262 [M+H⁺]; found 1152.3281.

(38): *R_f* = 0.20 (hexane – EtOAc, 2 : 1); [α]_D²⁰ = +23 (c 0.8, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.40-7.22 (m, 20H, CH₂Ph), 5.39 (dd, 1H, ³J_{3,4} = 9.5 Hz, ³J_{3,2} = 10.7 Hz, H-4), 5.30 (d, 1H, ³J_{NH,2} = 9.1 Hz, NH), 5.15 (d, 1H, ³J_{1,2} = 3.6 Hz, H-1), 4.94-4.81 (m, 6H, OP(O)(OCH₂Ph)₂, CH₂Ph or Troc), 4.78 (AB, 1H, ²J = 12.0 Hz, CH₂Ph or Troc), 4.67 (ddd, 1H, ³J_{4,3} = ³J_{4,5} = ²J_{H,P} = 9.2 Hz, H-4), 4.63 (AB, 1H, ²J = 11.9 Hz, CH₂Ph or Troc), 4.62 (s, 1H, H-1'), 4.50 (AB, 1H, ²J = 11.9 Hz, CH₂Ph or Troc), 4.44 (AB, 1H, ²J = 11.8 Hz, CH₂Ph or Troc), 4.28 (t, 1H, ³J_{4',3'} = ³J_{4',5'} = 9.4 Hz, H-4'), 4.22-4.19 (m, 1H, H-5), 4.06 (dd, 1H, ³J_{6a',5'} = 5.0 Hz, ²J_{6a',6b'} = 10.2 Hz, H-6a'), 3.98 (t, 1H, ³J_{6b',5'} = ²J_{6b',6a'} = 10.1 Hz, H-6b'), 3.98-3.94 (m, 1H, H-2), 3.69 (s, 3H, CH₃, Me), 3.68-3.63 (m, 2H, H-6a, H-6b), 3.58 (d, 1H, ³J_{2',3'} = 2.8 Hz, H-2'), 3.35 (dd, 1H, ³J_{3',2'} = 2.9 Hz, ³J_{3',4'} = 9.2 Hz, H-3'), 3.28 (dt, 1H, ³J_{5',6a'} = 4.9 Hz, ³J_{5',6b'} = ³J_{5',4'} = 9.7 Hz, H-5'), 1.88 (s, 3H, CH₃, Ac), 1.08 (s, 9H, 3×CH₃, DTBS), 0.99 (s, 9H, 3×CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ 171.62 (CO, Ac), 154.06 (CO, Troc), 138.59, 137.87 (2×Cq, CH₂Ph), 135.56 (Cq, ³J_{C,P} = 7.5 Hz, OP(O)(OCH₂Ph)₂), 135.47 (Cq, ³J_{C,P} = 7.7 Hz, OP(O)(OCH₂Ph)₂), 128.57, 128.45, 128.32, 127.98, 127.92, 127.68, 127.61, 127.57 (20×CH, CH₂Ph), 101.35 (C-1', ¹J_{C,H} = 157 Hz), 97.83 (C-1, ¹J_{C,H} = 173 Hz), 95.45 (CCl₃, Troc), 79.89 (C-3'), 79.70 (C-2'), 74.76 (C-4'), 74.47, 73.41 (2×CH₂, CH₂Ph or Troc), 73.12 (C-4, ²J_{C4,P} = 6.0 Hz), 73.00 (CH₂, CH₂Ph or Troc), 71.79 (C-5'), 71.01 (C-3), 70.40 (C-5, ³J_{C5,P} = 5.7 Hz), 69.61 (CH₂, ²J_{C,P} = 5.8 Hz, OP(O)(OCH₂Ph)₂), 69.46 (CH₂, ²J_{C,P} = 5.3 Hz, OP(O)(OCH₂Ph)₂), 67.62 (C-6), 66.17 (C-6'), 62.34 (CH₃, Me), 54.41 (C-2), 27.44, 27.07 (6×CH₃, DTBS), 22.61 (Cq, DTBS), 20.69 (CH₃, Ac), 19.92 (Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃): δ -2.51; HRMS (ESI): *m/z* calcd for C₅₄H₆₉NO₁₆PSi+H⁺: 1152.3262 [M+H⁺]; found 1152.3268;

(41): *R_f* = 0.15 (hexane – EtOAc, 2 : 1); [α]_D²⁰ = +9 (c 0.4, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.40-7.21 (m, 20H, CH₂Ph), 5.31 (t, 1H, ³J_{2,3} = ³J_{3,4} = 10.2 Hz, H-3), 5.08 (d, 1H, ³J_{2,NH} = 9.5 Hz, NH), 4.96-4.95 (s, 1H, H-1'), 4.95 (AB, 1H, ²J = 12.1 Hz, CH₂Ph or Troc), 4.92-4.83 (m, 4H, OP(O)(OCH₂Ph)₂), 4.75-4.73 (m, 2H, H-1, AB, CH₂Ph or Troc), 4.74 (AB, 1H, ²J = 12.1 Hz, CH₂Ph or Troc), 4.69 (AB, 1H, ²J = 11.8 Hz, CH₂Ph or Troc), 4.55 (ddd, 1H, ³J_{3,4} = ³J_{4,5} = ³J_{4,P} = 9.2 Hz, H-4), 4.52 (AB, 1H, ²J = 11.9 Hz, CH₂Ph or Troc), 4.43 (AB, 1H, ²J = 11.8 Hz, CH₂, CH₂Ph or Troc), 4.27 (t, 1H, ³J_{3',4'} = ³J_{4',5'} = 9.5 Hz, H-4'), 4.07 (dd, 1H, ³J_{5',6a'} = 4.8 Hz, ²J_{6a',6b'} = 9.2 Hz, 1H), 4.05-4.01 (m, 1H, H-5'), 3.88 (t, 1H, ³J_{5',6b'} = ²J_{6a',6b'} = 9.6 Hz, H-6b'), 3.74-3.72 (m, 1H, H-6a), 3.70 (dd, 1H, ³J_{2',3'} = 2.8 Hz, ³J_{3',4'} = 9.4 Hz, H-3'), 3.67-3.58 (m, 3H, H-2, H-5, H-6b), 3.48-3.47 (m, 1H, H-2'), 3.47 (s, 3H, CH₃, OMe), 1.89 (s, 3H, CH₃, Ac), 1.06 (s, 9H, 3×CH₃, DTBS), 0.99 (s, 9H, 3×CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ 171.08 (CO, Ac), 153.90 (CO, Troc), 139.17, 137.85 (2×Cq, CH₂Ph), 128.65, 128.58, 128.31, 128.27, 127.98, 127.78, 127.63, 127.41, 127.36 (20×CH, CH₂Ph), 100.12 (C-1, C-1'), 79.16 (C-2'), 78.82 (C-3'), 75.07 (C-4'), 74.57 (C-5, ³J_{C5,P} = 6.3 Hz), 74.39, 73.86, 73.67 (3×CH₂, CH₂Ph and Troc), 73.47 (C-4, ²J_{C4,P} = 5.9 Hz), 72.19 (C-3), 69.64, 69.60, 69.58, 69.54 (OP(O)(OCH₂Ph)₂), 68.65 (C-6), 68.53 (C-5'), 66.26 (C-6'), 60.15 (CH₃, Me), 56.49 (C-2), 27.43, 27.14 (6×CH₃, DTBS), 22.59 (Cq, DTBS), 20.67 (CH₃, Ac), 19.98 (Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃): δ = -2.20; HRMS (ESI): *m/z* calcd for C₅₄H₆₉NO₁₆PSi+H⁺: 1152.3262 [M+H⁺]; found 1152.3239.

Synthesis of 39 and 42



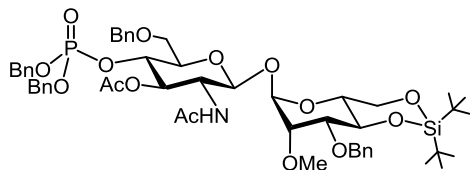
3-*O*-benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- β -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)- α -D-glucopyranoside (39) and 3-*O*-benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonyl-amino)- β -D-glucopyranoside (42)

(β , α - and α , β - by-products in the synthesis of 36)

(39): R_f = 0.25 (hexane - EtOAc, 1:1); $[\alpha]_D^{20}$ = +35 (c 1.0, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ : 7.75-7.71 (m, 2H, arom), 7.54-7.51 (m, 2H, arom), 7.40-7.20 (m, 24H, 24 \times CH, arom), 5.37 (t, 1H, ³ $J_{3,2}$ = ³ $J_{3,4}$ = 10.1 Hz, H-3), 5.08 (d, 1H, ³ $J_{1,2}$ = 3.5 Hz, H-1), 5.05 (d, 1H, ³ $J_{NH,2}$ = 9.1 Hz, NH), 4.95 (AB, 1H, ² J = 12.5 Hz, CH₂Ph), 4.93-4.83 (m, 4H, OP(O)(OCH₂Ph)₂), 4.82 (AB, 1H, ² J = 12.5 Hz, CH₂Ph), 4.65 (ddd, 1H, ³ $J_{4,3}$ = ³ $J_{4,5}$ = ³ $J_{H,P}$ = 9.2 Hz, H-4), 4.61 (s, 1H, H-1'), 4.51 (AB, 1H, ² J = 11.9 Hz, CH₂Ph), 4.43 (dd, 1H, ³ J = 6.7 Hz, ² J = 10.7 Hz, CH₂, Fmoc), 4.44 (AB, 1H, ² J = 11.1 Hz, CH₂Ph), 4.31 (dd, 1H, ³ J = 7.4 Hz, ² J = 10.6 Hz, CH₂, Fmoc), 4.29 (t, 1H, ³ $J_{4',3'}$ = ³ $J_{4',5'}$ = 9.3 Hz, H-4'), 4.22-4.19 (m, 1H, H-5), 4.16 (t, 1H, ³ J = 7.0 Hz, CH, Fmoc), 4.06 (dd, 1H, ³ $J_{6a',5'}$ = 5.0 Hz, ² $J_{6a',6b'}$ = 10.2 Hz, H-6a'), 3.98 (t, 1H, ³ $J_{6b',5'}$ = ² $J_{6b',6a'}$ = 10.2 Hz, H-6b'), 3.98-3.94 (m, 1H, H-2), 3.68-3.65 (m, 4H, H-6a, H-6b, CH₃, Me), 3.56 (d, 1H, ³ $J_{2',3'}$ = 2.8 Hz, H-2'), 3.37 (dd, 1H, ³ $J_{3',2'}$ = 2.9 Hz, ³ $J_{3',4'}$ = 9.2 Hz, H-3'), 3.29 (dt, 1H, ³ $J_{5',6a'}$ = 5.0 Hz, ³ $J_{5',4'}$ = ³ $J_{5',6b'}$ = 9.8 Hz, H-5'), 1.86 (s, 3H, CH₃, Ac), 1.08 (s, 9H, 3 \times CH₃, DTBS), 1.00 (s, 9H, 3 \times CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ : 171.63 (CO, Ac), 155.65 (CO, Fmoc), 143.78, 143.47, 141.36, 141.25 (4 \times Cq, Fmoc), 138.66, 137.89 (2 \times Cq, CH₂Ph), 135.59 (Cq, ³ $J_{C,P}$ = 7.0 Hz, OP(O)(OCH₂Ph)₂), 135.49 (Cq, ³ $J_{C,P}$ = 7.4 Hz, OP(O)(OCH₂Ph)₂), 128.55, 128.42, 128.30, 127.96, 127.91, 127.79, 127.74, 127.68, 127.65, 127.57, 127.07, 127.03, 124.99, 124.93, 120.05, 119.99 (28 \times CH, arom), 101.37 (C-1', ¹ $J_{C,H}$ = 155 Hz), 98.20 (C-1, ¹ $J_{C,H}$ = 177 Hz), 80.13 (C-3'), 79.87 (C-2'), 74.77 (C-4'), 73.40 (CH₂Ph), 73.25 (C-4, ² $J_{C4,P}$ = 6.3 Hz), 73.17 (CH₂Ph), 71.79 (C-5'), 71.13 (C-3), 70.36 (C-5, ³ $J_{C5,P}$ = 6.2 Hz), 69.57 (OP(O)(OCH₂Ph)₂, ² $J_{C,P}$ = 5.6 Hz), 69.42 (OP(O)(OCH₂Ph)₂, ² $J_{C,P}$ = 5.4 Hz), 67.70 (C-6), 67.02 (CH₂, Fmoc), 66.16 (C-6'), 62.34 (CH₃, Me), 54.12 (C-2), 47.11 (CH, Fmoc), 27.44, 27.07 (6 \times CH₃, DTBS), 22.60 (Cq, DTBS), 20.72 (CH₃, Ac), 19.92 (Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃): δ -2.53; HRMS (ESI): m/z calcd for C₆₆H₇₈NO₁₆PSi+H⁺: 1200.4900 [M+H⁺]; found 1200.4912.

Compound 42 was deprotected at C-2 and acetylated to provide compound 44 suitable for characterization by NMR.

Synthesis of 44

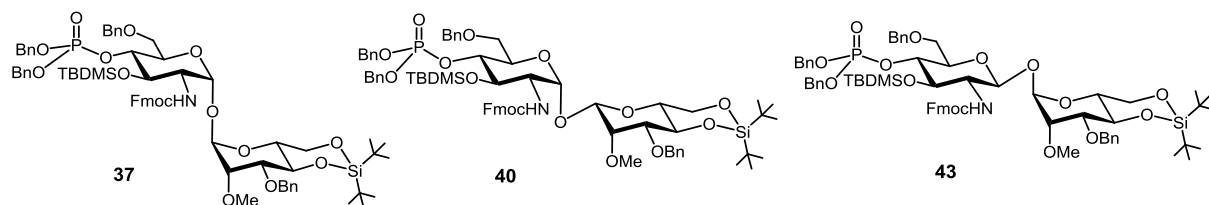


3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-2-acetamido-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy- α -D-glucopyranoside (44).

To a stirred solution of 42 (20 mg, 17 μ mol) in dry CH₂Cl₂ (1 ml) a solution of DBU (0.7 μ l, 5 μ mol) in dry CH₂Cl₂ (70 μ l of a stock solution prepared from 10 μ l DBU in 1 ml CH₂Cl₂) was added. The mixture was stirred for 1 h and successively added dropwise to a solution of acetic acid anhydride (20 μ l, 212 μ mol) in pyridine (300 μ l) at 0 $^{\circ}$ C. The stirring was continued for 20 min, the mixture was diluted with EtOAc (10 ml), washed with aq. citric acid (0.25 M, 10 ml), sat. aq. NaHCO₃ (10 ml) and brine (10 ml). The organic phase was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (toluene-EtOAc, 1:1 \rightarrow 1:2) to afford 44 (13 mg, 13 μ mol, 76%). R_f = 0.19 (toluene-EtOAc, 1:1); $[\alpha]_D^{20}$ = +11 (c 1.0, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.40-

7.39 (m, 2H, CH₂Ph), 7.34-7.20 (m, 18H, CH₂Ph), 5.50 (d, 1H, ³J_{NH,2} = 8.8 Hz, NH), 5.24 (dd, 1H, ³J = 9.0 Hz, ³J = 10.7 Hz, H-3), 4.95 (AB, 1H, ²J = 11.8 Hz, CH₂Ph), 4.94 (d, 1H, ³J_{1',2'} = 1.3 Hz, H-1'), 4.92-4.84 (m, 4H, OP(O)(OCH₂Ph)₂), 4.73 (AB, 1H, ²J = 12.0 Hz, CH₂Ph), 4.72 (d, 1H, ³J_{1,2} = 8.4 Hz, H-1), 4.54-4.49 (m, 1H, H-4), 4.52 (AB, 1H, ²J = 12.0 Hz, CH₂Ph), 4.44 (AB, 1H, ²J = 11.9 Hz, CH₂Ph), 4.26 (t, 1H, ³J_{4,3'} = ³J_{4',5'} = 9.5 Hz, H-4'), 4.08-4.00 (m, 2H, H-5', H-6a'), 3.92-3.86 (H-2, H-6b'), 3.74-3.71 (m, 2H, H-3', H-6a), 3.66-3.60 (m, 2H, H-5, H-6b), 3.49 (dd, 1H, ³J_{2',1'} = 1.6 Hz, ³J_{2',3'} = 3.1 Hz, H-2'), 3.47 (s, 3H, CH₃, Me), 1.92 (s, 3H, CH₃, Ac), 1.89 (s, 3H, CH₃, Ac), 1.06 (s, 9H, DTBS), 0.98 (s, 9H, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ 171.43, 169.98 (2×CO, Ac), 139.24, 137.92 (2×Cq, CH₂Ph), 135.52 (Cq, ³J_{C,P} = 7.1 Hz, OP(O)(OCH₂Ph)₂), 135.46 (Cq, ³J_{C,P} = 6.8 Hz, OP(O)(OCH₂Ph)₂), 128.62, 128.59, 128.57, 128.30, 128.23, 127.96, 127.94, 127.75, 127.59, 127.46, 127.31 (20×CH, CH₂Ph), 100.20 (C-1, ¹J_{C,H} = 159 Hz), 99.81 (C-1', ¹J_{C,H} = 172 Hz), 79.29 (C-2'), 78.92 (C-3'), 75.08 (C-4'), 74.66 (C-4 or C-5, ¹J_{C,P} = 6.4 Hz), 73.91, 73.68 (2×CH₂Ph), 73.57 (C-4 or C-5, ¹J_{C,P} = 6.4 Hz), 72.96 (C-3), 69.60 (CH₂, ²J_{C,P} = 5.2 Hz, OP(O)(OCH₂Ph)₂), 69.57 (CH₂, ²J_{C,P} = 5.0 Hz, OP(O)(OCH₂Ph)₂), 68.81 (C-6), 68.53 (C-5'), 66.32 (C-6'), 60.06 (CH₃, Me), 54.89 (C-2), 27.44, 27.15 (6×CH₃, DTBS), 23.27 (CH₃, Ac), 22.58 (Cq, DTBS), 20.67 (CH₃, Ac), 19.98 (Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃): δ -2.39; HRMS (ESI): *m/z* calcd for C₅₃H₇₀NO₁₅PSi⁺H⁺: 1020.4325 [M+H⁺]; found 1020.4323.

Synthesis of 37, 40 and 43



3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-3-*O*-di-*tert*-butyldimethylsilyl-2-deoxy-2-(9-fluorenylmethoxy-carbonylamino)- α -D-glucopyranoside (37),

3-*O*-benzyl-4,6-di-*O*-di-*tert*-butylsilylene-2-*O*-methyl- β -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-2-(9-fluorenylmethoxy-carbonylamino)- α -D-glucopyranoside (40) and

3-*O*-benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-2-(9-fluorenylmethoxy-carbonylamino)- β -D-glucopyranoside (43).

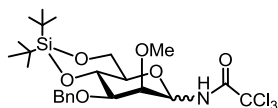
A solution of glycosyl donor **21** (57 mg, 96 μ mol) and glycosyl acceptor **33** (86 mg, 99 μ mol) in dry CH₂Cl₂ (1.5 ml) was stirred with powdered activated 4 Å molecular sieves for 2 h. at r. t. under atmosphere of Ar. The mixture was cooled to 0 °C and a solution of TMSOTf (1 μ L, 6 μ mol) in dry CH₂Cl₂ (50 μ L of a stock solution prepared from 10 μ L TMSOTf in 500 μ L CH₂Cl₂) was added. The mixture was stirred at 0 °C for 10 min and then the reaction was quenched by addition of Et₃N (10 μ L). The mixture was warmed to r. t., diluted with toluene (10 ml), filtered over a pad of Celite and the filtrate was concentrated. The residue was purified by HPLC (two successive columns: toluene - EtOAc, 4 : 1 \rightarrow 0 : 1 and hexane - EtOAc, 3 : 1 \rightarrow 2 : 1, column A) to afford **37** (63 mg, 50 μ mol, 52%), **40** (24 mg, 19 μ mol, 19%) and **43** (16 mg, 13 μ mol, 13%) as syrups. **37**: *R*_f = 0.40 (toluene - EtOAc, 3 : 1); [α]_D²⁰ = +52 (*c* 1.0, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.76-7.75 (m, 2H, arom.), 7.59-7.56 (m, 2H, arom.), 7.40-7.22 (m, 24H, arom.), 5.13 (d, 1H, ³J_{1,2} = 3.1 Hz, H-1), 5.11 (d, 1H, ³J_{1',2'} = 1.3 Hz, H-1'), 4.98-4.88 (m, 5H, OP(O)(OCH₂Ph)₂, CH₂Ph), 4.78 (AB, 1H, ²J = 12.6 Hz, CH₂Ph), 4.67 (d, 1H, ³J_{NH,2} = 9.4 Hz, NH), 4.53 (AB, 1H, ²J = 11.8 Hz, CH₂Ph), 4.44 (AB, 1H, ²J = 11.7 Hz, CH₂Ph), 4.33-4.30 (m, 3H, H-4', CH₂, Fmoc), 4.20-4.16 (m, 2H, H-4, CH, Fmoc), 4.02 (dd, 1H, ³J_{6a',5'} = 5.0 Hz, ²J_{6a',6b'} = 10.1 Hz, H-6a'), 3.96 (t, 1H, ³J_{6b',5'} = ²J_{6b',6a'} = 10.3 Hz, H-6b'), 3.88 (td, 1H, ³J_{2,1} = 3.2 Hz, ³J_{2,NH} = ³J_{2,3} = 9.4 Hz, H-2), 3.83-3.79 (m, 2H, H-5, H-6a), 3.75-3.71 (m, 2H, H-3, H-6b), 3.68 (td, ³J_{5',6a'} = 4.9 Hz, ³J_{5',4'} = ³J_{5',6b'} = 9.9 Hz, H-5'), 3.59 (dd, 1H, ³J_{3',2'} = 3.2 Hz, ³J_{3',4'} = 9.5 Hz, H-3'), 3.41 (dd, 1H, ³J_{2',1'} = 1.7 Hz, ³J_{2',3'} = 3.0 Hz, H-2'), 3.38 (s, 3H, CH₃, Me), 1.06 (s, 9H, 3×CH₃, DTBS), 0.96 (s, 9H, 3×CH₃, DTBS), 0.81 (s, 9H, 3×CH₃, TBDMS), 0.13 (s, 3H, CH₃, TBDMS), 0.03 (s, 3H, CH₃, TBDMS); ¹³C-NMR (151 MHz, CDCl₃): δ 155.79 (CO, Fmoc), 143.84, 143.77, 141.22 (4×Cq, Fmoc), 138.71, 138.08 (2×Cq,

CH₂Ph), 135.66, 135.61 (2×Cq, OP(O)(OCH₂Ph)₂), 128.65, 128.62, 128.59, 128.35, 128.31, 128.22, 128.07, 128.05, 127.74, 127.72, 127.66, 127.61, 127.09, 127.02, 125.26, 125.10, 119.98 (28×CH, arom.), 94.87 (C-1', ¹J_{C,H} = 178 Hz), 93.92 (C-1, ¹J_{C,H} = 175 Hz), 79.62 (C-2'), 77.03 (C-3'), 76.05 (C-4, ²J_{C₄P} = 6.4 Hz), 74.84 (C-4'), 73.31, 73.25 (2×CH₂Ph), 72.45 (C-5), 71.11 (C-3, ³J_{C₃P} = 4.6 Hz), 69.91 (CH₂, ²J_{C,P} = 5.3 Hz, OP(O)(OCH₂Ph)₂), 69.48 (CH₂, ²J_{C,P} = 4.8 Hz, OP(O)(OCH₂Ph)₂), 69.26 (C-5'), 68.56 (C-6), 67.81 (CH₂, Fmoc), 66.38 (C-6'), 59.57 (CH₃, Me), 54.93 (C-2), 47.07 (CH, Fmoc), 27.42, 27.08, 25.67 (9×CH₃, DTBS, TBDMS), 22.58, 19.84, 17.78 (3×Cq, DTBS, TBDMS), -4.07, -4.33 (2×CH₃, TBDMS); ³¹P-NMR (243 MHz, CDCl₃): δ -1.40ppm; HRMS (ESI) *m/z*: calcd for C₇₀H₉₀N O₁₅PSi₂+Na⁺: 1294.5479 [M+Na⁺]; found 1294.5444;

(40): *R*_f = 0.48 (toluene – EtOAc, 3 : 1); [α]_D²⁰ = +35 (*c* 1.0, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.73-7.71 (m, 2H, arom.), 7.54-7.53 (m, 2H, arom.), 7.40-7.18 (m, 24H, arom.), 4.94 (d, 1H, ³J_{1,2} = 3.2 Hz, H-1), 4.94 (AB, 1H, ²J = 12.4 Hz, CH₂Ph), 4.91-4.82 (m, 4H, OP(O)(OCH₂Ph)₂), 4.80 (AB, 1H, ²J = 12.7 Hz, CH₂Ph), 4.62-4.60 (m, 1H, NH), 4.60 (s, 1H, H-1'), 4.56 (AB, 1H, ²J = 12.2 Hz, CH₂Ph), 4.52 (dd, 1H, ³J = 6.0 Hz, ²J = 10.7 Hz, CH₂, Fmoc), 4.44 (dd, 1H, ³J = 5.8 Hz, ²J = 10.6 Hz, CH₂, Fmoc), 4.40 (AB, 1H, ²J = 12.0 Hz, CH₂Ph), 4.32 (ddd, 1H, ³J_{4,5} = ³J_{4,3} = ³J_{4,P} = 9.0 Hz, H-4), 4.26 (t, 1H, ³J_{4',5'} = ³J_{4',3'} = 9.3 Hz, H-4'), 4.15 (t, 1H, ³J = 5.8 Hz, CH, Fmoc), 4.09-4.06 (m, 1H, H-5), 4.05 (dd, 1H, ³J_{6a',5'} = 5.1 Hz, ²J_{6a',6b'} = 10.3 Hz, H-6a'), 3.97 (t, 1H, ³J_{6b',5'} = ²J_{6b',6a'} = 10.1 Hz, H-6b'), 3.87-3.80 (m, 3H, H-2, H-3, H-6a), 3.72-3.69 (m, 1H, H-6b), 3.46 (d, 1H, *J* = 2.3 Hz, H-2'), 3.44 (s, 3H, CH₃, Me), 3.35 (dd, 1H, ³J_{3',2'} = 2.9 Hz, ³J_{3',4'} = 9.3 Hz, H-3'), 3.26 (td, 1H, ³J_{5',6a'} = 4.8 Hz, ³J_{5',6b'} = ³J_{5',4'} = 9.9 Hz, H-5'), 1.07 (s, 9H, 3×CH₃, DTBS), 0.98 (s, 9H, 3×CH₃, DTBS), 0.77 (s, 9H, 3×CH₃, TBDMS), 0.03 (s, 3H, CH₃, TBDMS), -0.14 (s, 3H, CH₃, TBDMS); ¹³C-NMR (151 MHz, CDCl₃): δ 155.49 (CO, Fmoc), 143.82, 143.67, 141.47, 141.37 (4×Cq, Fmoc), 138.64, 138.32 (2×Cq, CH₂Ph), 135.78, 135.74 (2×Cq, OP(O)(OCH₂Ph)₂), 128.46, 128.44, 128.41, 128.21, 128.07, 127.94, 127.72, 127.69, 127.57, 127.50, 127.38, 127.06, 126.90, 124.72, 124.63, 119.99, 119.93 (28×CH, arom.), 99.86 (C-1', ¹J_{C,H} = 156 Hz), 97.79 (C-1, ¹J_{C,H} = 175 Hz), 80.35 (C-3'), 79.53 (C-2'), 76.57 (C-4, ²J_{C₄P} = 7.5 Hz), 74.77 (C-4'), 73.31, 72.84 (2×CH₂Ph), 71.78 (C-5'), 71.46 (C-3, ³J_{C₃P} = 4.5 Hz, 1C), 70.88 (C-5), 69.73 (OP(O)(OCH₂Ph)₂, ²J_{C,P} = 5.4 Hz), 69.25 (OP(O)(OCH₂Ph)₂, ²J_{C,P} = 4.5 Hz), 68.26 (C-6), 66.53 (CH₂, Fmoc), 66.08 (C-6'), 62.12 (CH₃, Me), 55.35 (C-2), 47.15 (CH, Fmoc), 27.43, 27.06, 25.73 (9×CH₃, DTBS, TBDMS), 22.58, 19.90, 17.77 (3×Cq, DTBS, TBDMS), -4.10, -4.26 (2×CH₃, TBDMS); ³¹P-NMR (243MHz, CDCl₃): δ -1.97; HRMS (ESI) *m/z* calcd for C₇₀H₉₀NO₁₅PSi₂+Na⁺: 1294.5479 [M+Na⁺]; found 1294.5459;

(43): *R*_f = 0.22 (toluene : EtOAc, 3 : 1); [α]_D²⁰ = +15 (*c* 1.2, CHCl₃); ¹H-NMR (600 MHz, acetone-d₆): δ 7.86 (d, 2H, *J* = 7.6 Hz, arom.), 7.69 (t, 2H, *J* = 6.0 Hz, arom.), 7.42-7.19 (m, 24H, arom.), 6.55 (d, 1H, ³J_{NH,2} = 9.7 Hz, NH), 5.04-4.96 (m, 5H, H-1', OP(O)(OCH₂Ph)₂), 4.85 (AB, 1H, ²J = 12.2 Hz, CH₂Ph), 4.78 (d, 1H, ³J_{1,2} = 8.3 Hz, H-1), 4.67 (AB, 1H, ²J = 12.1 Hz, CH₂Ph), 4.58 (AB, 1H, ²J = 11.9 Hz, CH₂Ph), 4.48-4.46 (m, 2H, CH₂, Fmoc, CH₂Ph), 4.36-4.28 (m, 2H, H-4, CH₂, Fmoc), 4.23 (t, 1H, *J* = 6.8 Hz, CH, Fmoc), 4.20 (t, 1H, ³J_{4',3'} = ³J_{4',5'} = 9.4 Hz, H-4'), 4.12-4.08 (m, 2H, H-5', H-6a'), 4.00 (t, 1H, ³J_{3,4} = ³J_{3,2} = 9.0 Hz, H-3), 3.92-3.90 (m, 1H, H-6a), 3.82 (dd, 1H, *J* = 5.2 Hz, *J* = 11.0 Hz, H-6b), 3.79-3.75 (m, 1H, H-6b'), 3.73-3.68 (m, 2H, H-3', H-5), 3.60-3.55 (m, 1H, H-2), 3.50 (br. s, 1H, H-2'), 3.23 (s, 3H, CH₃, Me), 1.06 (s, 9H, 3×CH₃, DTBS), 1.00 (s, 9H, 3×CH₃, DTBS), 0.84 (s, 9H, 3×CH₃, TBDMS), 0.12 (s, 3H, CH₃, TBDMS), 0.05 (s, 3H, CH₃, TBDMS); ¹³C-NMR (151 MHz, acetone-d₆): δ 157.06 (CO, Fmoc), 145.22, 144.96, 142.20, 140.43, 139.76, 137.34, 137.29 (6×Cq, arom.), 129.35, 129.31, 129.23, 129.19, 129.01, 128.98, 128.95, 128.60, 128.48, 128.14, 127.99, 125.99, 125.92, 120.90 (28×CH, arom.), 101.68 (C-1, ¹J_{C,H} = 160 Hz), 100.70 (C-1', ¹J_{C,H} = 171 Hz), 79.98, 79.82 (C-2', C-3'), 78.22 (C-4, ²J_{C₄P} = 6.5 Hz), 76.09 (C-4'), 75.47 (C-5), 74.96 (C-3, ³J_{C₃P} = 4.4 Hz), 73.91, 73.76 (2×CH₂Ph), 70.51 (C-6), 70.18 (OP(O)(OCH₂Ph)₂, ²J = 5.4 Hz), 69.95 (OP(O)(OCH₂Ph)₂, ²J = 4.7 Hz), 69.10 (C-5'), 67.35, 67.21 (C-6', CH₂ Fmoc), 59.85 (CH₃, Me), 59.41 (C-2), 48.10 (CH Fmoc), 27.86, 27.65, 26.53 (9×CH₃, DTBS, TBDMS), 23.19, 20.61, 18.64 (3×Cq, DTBS, TBDMS), -3.56, -3.78 (2×CH₃, TBDMS); ³¹P-NMR (243 MHz, acetone-d₆) δ -1.84; HRMS (ESI) *m/z* calcd for C₇₀H₉₀NO₁₅PSi₂+Na⁺: 1294.5479 [M+Na⁺]; found 1294.5460.

Synthesis of 45

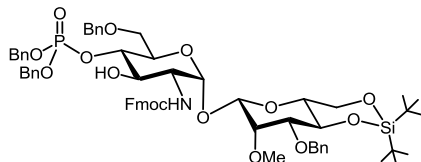


Trichloroacetyl 1-amino-3-O-benzyl-4,6-O-di-tert-butylsilylene-1-deoxy-2-O-methyl-D-mannopyranoside (45).

Glycosyl amide **45** was isolated from glycosylation reactions of acceptors **8** and **33** with donor **20**. *R*_f = 0.40 and 0.13 (hexane – EtOAc, 5 : 1); [α]_D²⁰ = +30 (*c* 0.3, CHCl₃); ¹H-NMR (300 MHz, CDCl₃, β-anomer): δ 7.55 (d, 1H, ³J_{NH,1} = 9.3 Hz, NH), 7.41-7.27 (m, 5H, arom.), 5.19 (dd, 1H, ³J_{1,2} = 1.5 Hz, ³J_{NH,1} = 9.3 Hz, H-1), 4.99 (AB, 1H, ²J =

12.3 Hz, CH_2Ph), 4.80 (AB, 1H, $^2J = 12.3$ Hz, CH_2Ph), 4.26 (t, 1H, $^3J_{4,5} = ^3J_{4,3} = 9.4$ Hz, H-4), 4.18 (dd, 1H, $^3J_{6a,5} = 5.0$ Hz, $^2J_{6a,6b} = 10.2$ Hz, H-6b), 3.93 (t, 1H, $^3J_{6b,5} = ^2J_{6b,6a} = 10.2$ Hz, H-6b), 3.71 (s, 3H, CH_3 , Me), 3.60 (dd, 1H, $^3J_{2,1} = 1.5$ Hz, $^3J_{2,3} = 3.1$ Hz, H-2), 3.53 (dd, 1H, $^3J_{3,2} = 3.0$ Hz, $^3J_{3,4} = 9.3$ Hz, H-3), 3.44 (dt, $^3J_{5,6a} = 4.9$ Hz, $^3J_{5,4} = ^3J_{5,6b} = 9.8$ Hz, H-3), 1.09 (s, 9H, $3 \times CH_3$, DTBS), 1.02 (s, 9H, $3 \times CH_3$, DTBS); ^{13}C -NMR (75 MHz, $CDCl_3$, β -anomer): δ 161.20 (CO), 138.51 (Cq, CH_2Ph), 128.47, 127.75, 127.54 ($5 \times CH$, CH_2Ph), 81.45 (C-3), 79.42, 78.99 (not assigned C-1, C-2), 74.76 (C-4), 73.83 (CH_2Ph), 73.00 (C-5), 66.10 (C-6), 62.23 (CH_3 , Me), 27.42, 27.07 ($6 \times CH_3$, DTBS), 22.64, 19.96 ($2 \times Cq$, DTBS); HRMS (ESI): m/z calcd for $C_{24}H_{35}NO_6Si$ -H: 566.1305 [M-H]⁻; found 566.1305.

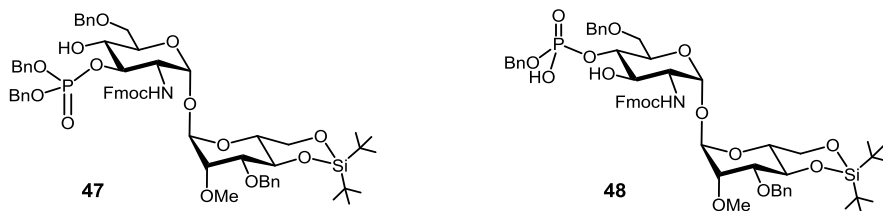
Synthesis of S1



3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- β -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)- α -D-glucopyranoside (S1).

A bi-phasic mixture of **39** (20 mg, 17 μ mol), THF (420 μ L) and aq. hydroxylamine (50%, 420 μ L) was vigorously stirred at 0 $^{\circ}C$ for 3 days. The mixture was diluted with EtOAc (10 ml) and washed with aq. citric acid (0.25 M, 15 ml), aq. sat $NaHCO_3$ (15 ml) and brine (15 ml). The organic layer was dried over $MgSO_4$, filtered and concentrated under reduced pressure. The residue was purified by MPLC (hexane - EtOAc, 3 : 1 \rightarrow 1 : 1) to afford compound **S1** (11 mg, 9.5 μ mol, 55%) as a syrup. $R_f = 0.18$ (hexane - EtOAc, 1 : 1); $[\alpha]_D^{20} = +55$ (c 1.0, $CHCl_3$); 1H -NMR (600 MHz, $CDCl_3$): δ 7.74 (t, 2H, $J = 7.5$ Hz, arom.), 7.57 (t, 2H, $J = 6.7$ Hz, arom.), 7.40-7.15 (m, 24H, arom.), 5.21 (d, 1H, $3J_{1,2} = 2.8$ Hz, H-1), 5.05 (d, 1H, $3J_{NH,2} = 7.1$ Hz, NH), 5.00-4.90 (m, 5H, CH_2Ph , $OP(O)(OCH_2Ph)_2$), 4.79 (AB, 1H, $2J = 12.5$ Hz, CH_2Ph), 4.60 (s, 1H, H-1'), 4.48-4.44 (m, 3H, CH_2Ph , CH_2 Fmoc), 4.42-4.37 (m, 2H, H-4, CH_2 Fmoc), 4.25 (t, 1H, $3J_{4',5'} = 3J_{4',3'} = 9.4$ Hz, H-4'), 4.21 (t, 1H, $J = 6.5$ Hz, CH, Fmoc), 4.10 (br. d, $J = 9.5$ Hz, H-5), 4.04 (dd, 1H, $3J_{6a',5'} = 5.0$ Hz, $2J_{6a',6b'} = 10.1$ Hz, H-6a'), 4.01-3.97 (m, 1H, H-3), 3.96 (t, 1H, $3J_{6b',6a'} = 2J_{6b',6a'} = 10.2$ Hz, H-6b'), 3.84-3.80 (m, 1H, H-2), 3.65-3.53 (m, 5H, H-6a, H-6b, CH_3 Me), 3.52 (d, 1H, $3J_{2',3'} = 2.4$ Hz, H-2'), 3.35 (dd, 1H, $3J_{3',2'} = 2.9$ Hz, $3J_{3',4'} = 9.2$ Hz, H-3'), 3.27 (td, 1H, $3J_{5',4'} = 3J_{5',6b'} = 9.8$ Hz, $3J_{5',6a'} = 4.9$ Hz, H-5'), 1.07 (s, 9H, $3 \times CH_3$, DTBS), 0.99 (s, 9H, $3 \times CH_3$, DTBS); ^{13}C -NMR (151 MHz, $CDCl_3$): δ 156.22 (CO, Fmoc), 143.95, 143.62, 141.38, 141.30 ($4 \times Cq$, Fmoc), 138.64, 137.82 ($2 \times Cq$, CH_2Ph), 135.30 (Cq, $3J_{C,P} = 7.2$ Hz, $OP(O)(OCH_2Ph)_2$), 135.25 (Cq, $3J_{C,P} = 8.0$ Hz, $OP(O)(OCH_2Ph)_2$), 128.73, 128.65, 128.59, 128.40, 128.31, 128.13, 127.87, 127.73, 127.64, 127.60, 127.55, 127.52, 127.03, 125.02, 124.93, 120.00 ($28 \times CH$, arom.), 101.34 (C-1'), 98.11 (C-1), 80.18 (C-3'), 79.80 (C-2'), 77.05 (C-4), 74.74 (C-4'), 73.45, 73.14 ($2 \times CH_2Ph$), 71.73 (C-5'), 70.97 (C-3), 70.30 (CH_2 , $^3J_{C,P} = 5.1$ Hz, $OP(O)(OCH_2Ph)_2$), 69.92 (CH_2 , $^3J_{C,P} = 5.5$ Hz, $OP(O)(OCH_2Ph)_2$), 69.74 (C-5, $3J_{C5,P} = 9.9$ Hz), 67.70 (C-6), 66.90 (CH_2 , Fmoc), 66.16 (C-6'), 62.20 (CH_3 , Me), 55.12 (C-2), 47.16 (CH, Fmoc), 27.43, 27.07 ($6 \times CH_3$, DTBS), 22.58, 19.91 ($2 \times Cq$, DTBS); ^{31}P -NMR (243 MHz, $CDCl_3$): δ 0.35; HRMS (ESI): m/z calcd for $C_{64}H_{76}NO_{15}PSi$ +H⁺: 1158.4795 [M+H]⁺; found 1158.4796.

Synthesis of 47 and 48



3-*O*-Benzyl-4,6-*O*-di-*tert*-butylsilylene-2-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-3-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)- α -D-glucopyranoside (47) and

3-O-benzyl-4,6-O-di-tert-butylsilylene-2-O-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-O-benzyl-4-O-(benzyloxyphosphoryl)-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)- α -D-glucopyranoside (48).

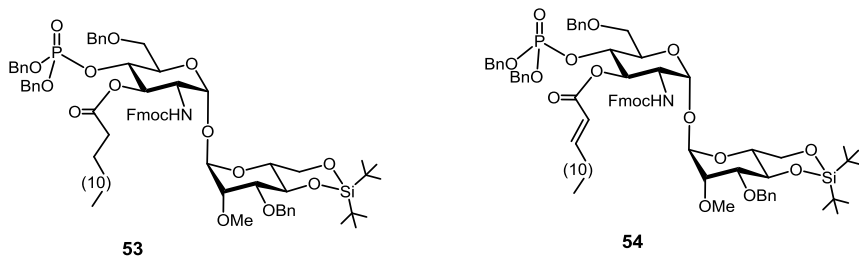
47: Phosphate-migration by-product from reaction of **36** with hydroxylamine, $R_f = 0.66$ (toluene – EtOAc, 1 : 1); $[\alpha]_D^{20} = +55$ (c 0.9, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.73-7.71 (m, 2H, arom.), 7.59-7.54 (m, 2H, arom.), 7.44-7.13 (m, 24H, arom.), 5.43 (d, 1H, ³ $J_{NH,2} = 8.5$ Hz, NH), 5.24 (d, 1H, ³ $J_{1,2} = 3.2$ Hz, H-1), 5.11 (s, 1H, H-1'), 5.09-4.95 (m, 5H, CH₂Ph, OP(O)(OCH₂Ph)₂), 4.80 (AB, 1H, ² $J = 12.0$ Hz, CH₂Ph), 4.59 (AB, 1H, ² $J = 11.8$ Hz, CH₂Ph), 4.56 (AB, 1H, ² $J = 11.9$ Hz, CH₂Ph), 4.39 (ddd, 1H, ³ $J_{3,2} = ^3J_{3,4} = ^3J_{H,P} = 8.6$ Hz, H-3), 4.33 (t, 1H, ³ $J_{4',3'} = ^3J_{4',5'} = 9.5$ Hz, H-4'), 4.30-4.27 (m, 1H, CH₂, Fmoc), 4.19-4.16 (m, 1H, CH, Fmoc), 4.13-4.11 (m, 1H, CH₂, Fmoc), 4.07 (dd, 1H, ³ $J_{6a',5'} = 4.8$ Hz, ² $J_{6a',6b'} = 10.2$ Hz, H-6a'), 3.97 (t, 1H, ³ $J_{6b',5'} = ^2J_{6b',6a'} = 10.4$ Hz, H-6b'), 3.80-3.69 (m, 6H, H-3', H-4, H-5, H-5', H-6a, H-6b), 3.43 (s, 1H, H-2'), 3.41 (s, 3H, CH₃, Me), 1.06 (s, 9H, 3 \times CH₃, DTBS), 0.95 (s, 9H, 3 \times CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃): δ 156.14 (CO, Fmoc), 143.86, 143.57, 141.21, 141.18 (4 \times Cq, Fmoc), 138.93, 137.76 (2 \times Cq, CH₂Ph), 135.26 (Cq, ³ $J_{C,P} = 7.5$ Hz, OP(O)(OCH₂Ph)₂), 135.13 (Cq, ³ $J_{C,P} = 7.5$ Hz, OP(O)(OCH₂Ph)₂), 128.69, 128.64, 128.60, 128.56, 128.42, 128.37, 127.96, 127.80, 127.75, 127.72, 127.67, 127.61, 127.15, 125.40, 125.17, 119.92, 119.88 (28 \times CH, arom.), 93.73 (C-1'), 93.53 (C-1), 79.82 (C-3), 79.62 (C-2'), 78.31 (C-3'), 74.82 (C-4'), 73.94, 73.74 (2 \times CH₂Ph), 71.17 (C-4 or C-5 or C-5'), 70.44 (C-4 or C-5 or C-5'), 70.09 (CH₂, ² $J_{C,P} = 5.8$ Hz, OP(O)(OCH₂Ph)₂), 69.90 (CH₂, ² $J_{C,P} = 5.0$ Hz, OP(O)(OCH₂Ph)₂), 69.42 (C-6), 69.13 (C-4 or C-5 or C-5'), 67.81 (CH₂, Fmoc), 66.43 (C-6'), 59.78 (CH₃, Me), 53.76 (C-2, ³ $J_{C2,P} = 4.0$ Hz), 46.93 (CH, Fmoc), 27.46, 27.08 (6 \times CH₃, DTBS), 22.60, 19.82 (2 \times Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃): δ 0.41; HRMS (ESI): m/z calcd for C₆₄H₇₆NO₁₅PSi+H⁺: 1158.4795 [M+H⁺]; found 1158.4794;

48: Debenzylation by-product from reaction of **36** with hydroxylamine, $R_f = 0.20$ (EtOAc – MeOH, 5:1); $[\alpha]_D^{20} = +36$ (c 0.9, CHCl₃); ¹H-NMR (600 MHz, CDCl₃ : MeOD = 3 : 1): δ 7.76-7.75 (m, 2H, arom.), 7.62-7.58 (m, 2H, arom.), 7.42-7.19 (m, 18H, arom.), 5.19 (br. s, 1H, H-1), 5.14 (s, 1H, H-1'), 4.99-4.89 (m, 2H, OP(O)(OCH₂Ph)OH), 4.97 (AB, 1H, ² $J = 12.0$ Hz, CH₂Ph), 4.80 (AB, 1H, ² $J = 12.1$ Hz, CH₂Ph), 4.53 (dd, 1H, ³ $J = 7.7$ Hz, ² $J = 10.2$ Hz, CH₂, Fmoc), 4.42 (AB, 1H, ² $J = 11.6$ Hz, CH₂Ph), 4.33 (AB, 1H, ² $J = 11.9$ Hz, CH₂Ph), 4.30 (t, 1H, ³ $J_{4',5'} = ^3J_{4',3'} = 9.5$ Hz, H-4'), 4.23 (t, 1H, $J = 7.3$ Hz, CH, Fmoc), 4.18-4.11 (m, 1H, CH₂, Fmoc), 4.10-4.03 (m, 1H, H-4), 4.07 (dd, 1H, ³ $J_{6a',5'} = 4.7$ Hz, ² $J_{6a',6b'} = 10.0$ Hz, H-6b'), 3.95 (t, 1H, ³ $J_{6b',5'} = ^2J_{6b',6a'} = 10.3$ Hz, H-6b'), 3.88-3.82 (m, 1H, H-2), 3.81-3.69 (m, 5H, H-3, H-3', H-5, H-5', H-6a), 3.59-3.53 (m, 1H, H-6b), 3.46 (s, 1H, H-2'), 3.33 (s, 3H, CH₃, Me), 1.07 (s, 9H, 3 \times CH₃, DTBS), 0.96 (s, 9H, 3 \times CH₃, DTBS); ¹³C-NMR (151 MHz, CDCl₃ : MeOD = 3 : 1): δ 156.80 (CO, Fmoc), 143.53, 143.38, 140.96, 140.94 (4 \times Cq, Fmoc), 138.40, 137.49 (2 \times Cq, CH₂Ph), 137.12 (Cq, OP(O)(OCH₂Ph)OH), 128.09, 128.07, 127.98, 127.64, 127.47, 127.40, 127.38, 127.31, 127.03, 126.81, 126.76, 124.85, 124.83, 119.68 (18 \times CH, arom.), 92.95, 92.89 (not assigned C-1, C-1'), 79.17 (C-2'), 77.62 (C-3'), 74.67 (C-4), 74.35 (C-4'), 73.21 (2 \times CH₂Ph), 70.90 (not assigned C-5 or C-5' or C-3), 68.86 (C-6), 68.64 (not assigned C-5 or C-5' or C-3), 67.63 (OP(O)(OCH₂Ph)OH), 67.15 (CH₂, Fmoc), 66.19 (C-6'), 58.95 (CH₃, Me), 54.40 (C-2), 46.78 (CH, Fmoc), 27.06, 26.68 (6 \times CH₃, DTBS), 22.27, 19.47 (2 \times Cq, DTBS); ³¹P-NMR (243 MHz, CDCl₃ : MeOD = 3 : 1): δ -1.87; HRMS (ESI) m/z calcd for C₅₇H₇₀NO₁₅PSi-H: 1066.4180 [M-H]; found 1066.4170.

Synthesis of 49-51

(*R*)-3-(alkanoyloxy)alkanoic acids **49-51** were prepared according to the reported procedures.^{18,19}

Synthesis of 53 and 54



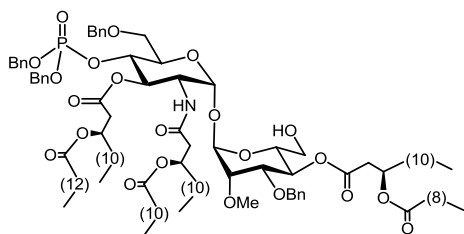
3-O-Benzyl-4,6-O-di-tert-butylsilylene-O-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-O-benzyl-4-O-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)-3-O-tetradecanoyl- α -D-glucopyranoside (53) and

3-*O*-benzyl-4,6-*O*-di-*tert*-butylsilylene-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-(9-fluorenylmethoxycarbonylamino)-3-*O*-tetradec-2-enoyl- α -D-glucopyranoside (54).

53: a by-product formed upon acylation of **46** with **49** (to afford **52** as the major product): $R_f = 0.28$ (hexane – EtOAc, 2 : 1); $[\alpha]_D^{20} = +53$ (c 1.0, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.75 (d, 2H, $J = 7.5$ Hz, arom.), 7.57-7.22 (m, 26H, arom.), 5.26 (t, 1H, $^3J_{3,4} = ^3J_{3,2} = 9.9$ Hz, H-3), 5.20 (d, 1H, $^3J_{1,2} = 3.5$ Hz, H-1), 5.12 (s, 1H, H-1'), 5.04 (AB, 1H, $^2J = 11.9$ Hz, CH₂Ph), 4.98-4.92 (m, 4H, OP(O)(OCH₂Ph)₂), 4.82 (AB, 1H, $^2J = 11.9$ Hz, CH₂Ph), 4.52 (AB, 1H, $^2J = 11.8$ Hz, CH₂Ph), 4.51 (ddd, 1H, $^3J_{4,5} = ^3J_{4,3} = ^3J_{4,P} = 9.4$ Hz, H-4), 4.43-4.40 (m, 1H, CH₂, Fmoc), 4.43 (AB, 1H, $^2J = 12.0$ Hz, CH₂Ph), 4.32 (t, 1H, $^3J_{4',3'} = 3J_{4',5'} = 9.5$ Hz, H-4'), 4.18 (t, 1H, $J = 7.5$ Hz, CH, Fmoc), 4.12-4.07 (m, 2H, H-2, CH₂, Fmoc), 4.05 (dd, 1H, $^3J_{6a',5'} = 4.7$ Hz, $^2J_{6a',6b'} = 10.2$ Hz, H-6a'), 3.96 (t, 1H, $^3J_{6b',5'} = ^2J_{6b',6a'} = 10.3$ Hz, H-6b'), 3.80-3.70 (m, 4H, H-5, H-5', H-3', H-6a), 3.64 (dd, 1H, $J = 5.9$ Hz, $J = 11.0$ Hz, H-6b), 3.42 (br. s, 1H, H-2'), 3.39 (s, 3H, CH₃, Me), 2.15 (t, 2H, $J = 7.8$ Hz, α^{Myr} -CH₂), 1.45-0.95 (m, 11 \times CH₂, 6 \times CH₃, DTBS), 0.88 (t, 3H, $J = 7.1$ Hz, ω^{Myr} -CH₃); ¹³C-NMR (151 MHz, CDCl₃): δ 174.47 (CO), 155.71 (CO, Fmoc), 143.74, 143.54, 141.21 (4 \times Cq, Fmoc), 138.81, 137.83 (2 \times Cq, CH₂Ph), 135.50 (OP(O)(OCH₂Ph)₂, $3J_{C,P} = 6.9$ Hz), 135.43 (OP(O)(OCH₂Ph)₂, $3J_{C,P} = 7.3$ Hz), 128.71, 128.64, 128.38, 128.34, 127.98, 127.95, 127.92, 127.80, 127.76, 127.68, 127.64, 127.12, 125.28, 125.13, 119.97 (28 \times CH, arom.), 93.80 (C-1'), 93.23 (C-1), 79.64 (C-2'), 78.03 (C-3'), 74.81 (C-4'), 74.10 (CH₂Ph), 73.58 (C-4), 73.55 (CH₂Ph), 71.06 (C-3), 70.67 (C-5, $3J_{C5,P} = 5.6$ Hz), 69.70, 69.67, 69.64 (OP(O)(OCH₂Ph)₂), 69.23 (C-5'), 68.36 (C-6), 67.94 (CH₂, Fmoc), 66.39 (C-6'), 59.70 (CH₃, Me), 53.76 (C-2), 46.93 (CH, Fmoc), 34.18 (α -CH₂), 31.92, 29.67, 29.62, 29.58, 29.40, 29.35, 29.20, 29.13 (9 \times CH₂), 27.44, 27.05 (6 \times CH₃, DTBS), 24.72, 22.68, 22.59, 19.81 (2 \times CH₂, 2 \times Cq, DTBS), 14.11 (ω -CH₃); ³¹P-NMR (243 MHz, CDCl₃): δ -2.04; HRMS (ESI): m/z calcd for C₇₈H₁₀₂NO₁₆PSi+H⁺ [M+H]⁺ 1368.6778, found 1368.6771.

54: a by-product formed upon acylation of **46** with **49** (to afford **52** as the major product): $R_f = 0.25$ (hexane – EtOAc, 2 : 1); $[\alpha]_D^{20} = +51$ (c 0.3, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.74-7.23 (m, 28H, arom.), 6.95 (td, 1H, $J = 6.9$ Hz, $J = 15.5$ Hz, β -CH), 5.70 (d, 1H, $J = 15.7$ Hz, α -CH), 5.36 (dd, 1H, $^3J_{3,4} = 9.7$ Hz, $^3J_{3,2} = 10.6$ Hz, H-3), 5.23 (d, 1H, $^3J_{1,2} = 3.7$ Hz, H-1), 5.13 (s, 1H, H-1'), 5.04 (AB, 1H, $^2J = 11.9$ Hz, CH₂Ph), 5.00 (d, 1H, $^3J_{NH,2} = 9.3$ Hz, NH), 4.96-4.87 (m, 4H, OP(O)(OCH₂Ph)₂), 4.83 (AB, 1H, $^2J = 11.9$ Hz, CH₂Ph), 4.63 (q, 1H, $^3J_{4,5} = ^3J_{4,3} = ^3J_{4,P} = 9.3$ Hz, H-4), 4.54-4.51 (m, 1H, CH₂, Fmoc), 4.52 (AB, 1H, $^2J = 11.8$ Hz, CH₂Ph), 4.43 (AB, 1H, $^2J = 11.8$ Hz, CH₂Ph), 4.32 (t, 1H, $^3J_{4',5'} = ^3J_{4',3'} = 9.6$ Hz, H-4'), 4.17-4.11 (m, 2H, H-2, CH₂, Fmoc), 4.04 (dd, 1H, $^3J_{6a',5'} = 4.7$ Hz, $^2J_{6a',6b'} = 9.9$ Hz, H-6a'), 3.95 (t, 1H, $^3J_{6b',6a'} = ^2J_{6b',5'} = 10.0$ Hz, 1H), 3.92 (dd, 1H, $J = 7.7$ Hz, $J = 10.3$ Hz, CH₂, Fmoc), 3.85-3.81 (m, 1H, H-5), 3.79-3.78 (m, 1H, H-6a), 3.75-3.70 (H-3', H-5'), 3.67 (dd, 1H, $J = 5.8$ Hz, $J = 10.9$ Hz, H-6b), 3.43 (br. s, 1H, H-2'), 3.40 (s, 3H, CH₃, Me), 1.87-1.82 (m, 2H, γ -CH₂), 1.35-0.94 (m, 36H, 9 \times CH₂, 6 \times CH₃, DTBS), 0.88 (t, 3H, $J = 7.1$ Hz, ω -CH₃); ¹³C-NMR (151 MHz, CDCl₃): δ 166.98 (CO), 155.78 (CO, Fmoc), 152.31 (β -CH), 143.87, 143.57, 141.22, 141.14 (4 \times Cq, Fmoc), 138.82, 137.87 (2 \times Cq, CH₂Ph), 135.57, 135.51 (2 \times Cq, OP(O)(OCH₂Ph)₂), 128.60, 128.57, 128.55, 128.39, 128.34, 127.97, 127.87, 127.77, 127.70, 127.66, 127.15, 127.06, 125.34, 125.24 (27 \times CH, arom.), 119.89, 119.77 (α -CH, CH, arom.), 93.67 (C-1'), 93.10 (C-1), 79.62 (C-2'), 78.04 (C-3'), 74.84 (C-4'), 74.12, 73.55 (2 \times CH₂Ph), 73.51 (C-4, $2J_{C4,P} = 5.3$ Hz), 71.37 (C-3), 70.77 (C-5, $3J_{C5,P} = 4.7$ Hz), 69.53, 69.49, 69.15 (OP(O)(OCH₂Ph)₂), 68.38 (C-6), 67.84 (CH₂, Fmoc), 66.39 (C-6'), 59.72 (CH₃, Me), 53.97 (C-2), 47.01 (CH, Fmoc), 32.31 (γ -CH₂), 31.92, 29.70, 29.63, 29.58, 29.43, 29.34, 29.27, 29.20, 27.64 (9 \times CH₂), 27.44, 27.05 (6 \times CH₃, DTBS), 22.69, 22.59, 19.81 (CH₂, 2 \times Cq, DTBS), 14.12 (ω -CH₃); ³¹P-NMR (243 MHz, CDCl₃): δ -2.07; HRMS (ESI) m/z calcd for C₇₈H₁₀₀NO₁₆PSi+H⁺: 1366.6622 [M+H]⁺; found 1366.6616.

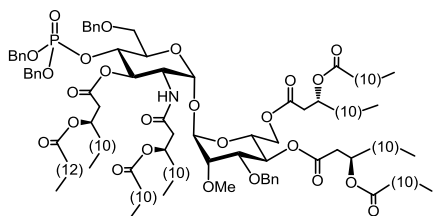
Synthesis of 58



3-*O*-Benzyl-4-*O*-[(*R*)-3-(decanoyloxy)tetradecanoyl]-2-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-[(*R*)-3-dodecanoyloxy]tetradecanoylamino]-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanyol]- α -D-glucopyranoside (58**).**

To a stirred solution of **56** (30 mg, 18 μ mol) and DMAP (0.2 mg, 1.6 μ mol) in dry CH_2Cl_2 (400 μ l) a solution of **51** (8 mg, 20 μ mol) in CH_2Cl_2 (83 μ l of a stock solution prepared from 29 mg of **51** in 300 μ l CH_2Cl_2) was added under atmosphere of Ar. The mixture was cooled to 0 $^\circ\text{C}$ and DIC (6 mg, 48 μ mol) from a 50 mg·ml⁻¹ stock solution in toluene was added dropwise over a period of 2 h. Then an additional amount of **51** (2 mg, 5.0 μ mol) and DIC (0.4 mg, 0.3 μ mol) from the indicated stock solutions was added dropwise over a period of 1 h. The mixture was diluted with EtOAc (20 ml) and washed with aq. HCl (2 M, 20 ml), sat. aq. NaHCO_3 (20 ml) and brine (20 ml). The organic layer was dried over Na_2SO_4 , filtered and concentrated. The residue was purified by HPLC (two successive columns: toluene – EtOAc, 3 : 1 \rightarrow 1 : 1 and hexane – EtOAc, 2 : 1 \rightarrow 1 : 1, column B) to afford **58** (24 mg, 12 μ mol, 65%) as a syrup. Fractions containing by-product **60** were purified HPLC (hexane – EtOAc, 4 : 1 \rightarrow 3 : 1, column B) to afford 4,6-di-*O*-acylated compound **60** (5.7 mg, 2.4 μ mol, 13%). Compound **58**: R_f = 0.40 (toluene – EtOAc, 1 : 1); $[\alpha]_D^{20}$ = +36 (1.0, CHCl_3); $^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 7.38-7.22 (m, 20H, CH_2Ph), 6.34 (d, 1H, $^3J_{\text{NH},2}$ = 8.0 Hz, NH), 5.24-5.15 (m, 5H, H-1, H-3, H-4', $2\times\beta^{\text{Myr}}\text{-CH}$), 5.12 (d, 1H, $^3J_{1',2'}$ = 2.0 Hz, H-1'), 5.07-5.03 (m, 1H, $\beta^{\text{Myr}}\text{-CH}$), 4.97 (d, 2H, J = 8.1 Hz, $\text{OP(O)(OCH}_2\text{Ph)}_2$), 4.95 (d, 2H, J = 8.2 Hz, $\text{OP(O)(OCH}_2\text{Ph)}_2$), 4.69 (AB, 1H, 2J = 12.0 Hz, CH_2Ph), 4.64 (AB, 1H, 2J = 12.1 Hz, CH_2Ph), 4.48 (AB, 1H, 2J = 11.8 Hz, CH_2Ph), 4.43 (ddd, 1H, $^3J_{4,3}$ = $^3J_{4,5}$ = $^3J_{\text{H,P}}$ = 9.3 Hz, H-4), 4.40 (AB, 1H, 2J = 11.8 Hz, CH_2Ph), 4.24 (ddd, 1H, $^3J_{2,1}$ = 3.5 Hz, $^3J_{2,\text{NH}}$ = 7.9, $^3J_{2,3}$ = 11.2 Hz, H-2), 3.93 (dd, 1H, $^3J_{3',2'}$ = 3.1 Hz, $^3J_{3',4'}$ = 9.4 Hz, H-3'), 3.75-3.72 (m, 2H, H-5, H-6a), 3.61-3.54 (m, 4H, H-5', H-6b, H-6a', H-6b'), 3.30-3.29 (m, 1H, H-2'), 3.29 (s, 3H, CH_3 , Me), 2.60 (dd, 1H, J = 7.6 Hz, J = 15.5 Hz, $\alpha^{\text{Myr}}\text{-CH}_2$), 2.54-2.47 (m, 3H, $\alpha^{\text{Myr}}\text{-CH}_2$), 2.44 (dd, J = 7.1 Hz, J = 15.1 Hz, $\alpha^{\text{Myr}}\text{-CH}_2$), 2.37 (dd, 1H, J = 5.5 Hz, J = 15.1 Hz, $\alpha^{\text{Myr}}\text{-CH}_2$), 2.32-2.26 (m, 2H, $\alpha\text{-CH}_2$), 2.24-2.19 (m, 4H, $2\times\alpha\text{-CH}_2$), 1.62-1.46 (m, 12H, $3\times\gamma^{\text{Myr}}\text{-CH}_2$, $\beta^{\text{Myr}}\text{-CH}_2$, $\beta^{\text{Lau}}\text{-CH}_2$, $\beta^{\text{Cap}}\text{-CH}_2$), 1.32 -1.20 (m, 102H, $51\times\text{CH}_2$), 0.89-0.86 (m, 18H, $4\times\omega^{\text{Myr}}\text{-CH}_3$, $\omega^{\text{Lau}}\text{-CH}_3$, $\omega^{\text{Cap}}\text{-CH}_3$); $^{13}\text{C-NMR}$ (151 MHz, CDCl_3): δ 173.63, 173.38, 173.08, 171.42, 169.91, 169.87 ($6\times\text{CO}$), 138.00, 137.87 ($2\times\text{Cq}$, CH_2Ph), 135.54, 135.50 ($2\times\text{Cq}$, $\text{OP(O)(OCH}_2\text{Ph)}_2$), 128.71, 128.64, 128.52, 128.31, 128.11, 128.10, 128.03, 127.92, 127.64, 127.59 ($20\times\text{CH}$, CH_2Ph), 94.02 (C-1'), 93.50 (C-1), 77.80 (C-2'), 75.89 (C-3'), 73.66 (C-4, $^2J_{\text{C}_4,\text{P}}$ = 5.8 Hz), 73.48 (CH_2Ph), 72.73 (C-5'), 72.51 (CH_2Ph), 71.08 (C-3), 70.83 ($\beta^{\text{Myr}}\text{-CH}$), 70.67 (C-5, $^3J_{\text{C}_5,\text{P}}$ = 5.3 Hz), 69.94 ($\beta^{\text{Myr}}\text{-CH}$), 69.87 ($\beta^{\text{Myr}}\text{-CH}$), 69.76, 69.72 ($\text{OP(O)(OCH}_2\text{Ph)}_2$), 68.35 (C-6), 68.33 (C-4'), 61.69 (C-6'), 59.09 (CH_3 , Me), 51.80 (C-2), 41.37, 39.15, 39.00 ($3\times\alpha^{\text{Myr}}\text{-CH}_2$), 34.59, 34.52, 34.43, 34.26, 34.03 ($3\times\gamma^{\text{Myr}}\text{-CH}_2$, $\alpha^{\text{Myr}}\text{-CH}_2$, $\alpha^{\text{Lau}}\text{-CH}_2$, $\alpha^{\text{Cap}}\text{-CH}_2$), 31.92, 31.86, 29.71, 29.68, 29.66, 29.61, 29.58, 29.56, 29.54, 29.50, 29.47, 29.41, 29.38, 29.36, 29.30, 29.28, 29.19, 29.18, 25.35, 25.16, 25.09, 24.99, 22.68, 22.66 ($54\times\text{CH}_2$), 14.09 ($4\times\omega^{\text{Myr}}\text{-CH}_3$, $\omega^{\text{Lau}}\text{-CH}_3$, $\omega^{\text{Cap}}\text{-CH}_3$); $^{31}\text{P-NMR}$ (243 MHz, CDCl_3): δ -1.70; HRMS (ESI): m/z calcd for $\text{C}_{119}\text{H}_{194}\text{NO}_{22}\text{P}+\text{Na}^+$: 2043.3722 [$\text{M}+\text{Na}^+$]; found 2043.3762.

Synthesis of 59

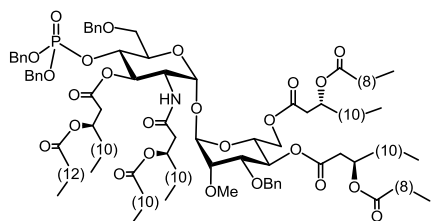


3-*O*-Benzyl-4,6-di-*O*-[(*R*)-3-(dodecanoyloxy)tetradecanoyl]-2-*O*-methyl- α -D-mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-[(*R*)-3-(dodecanoyloxy)-tetradecanoylamino]-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanyol]- α -D-glucopyranoside (59**)**

by-product from reaction of **56** with fatty acid **50** (to furnish **57** as major product), R_f = 0.44 (toluene – EtOAc, 4 : 1); $[\alpha]_D^{20}$ = +32 (c 0.7, CHCl_3); $^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 7.38-7.22 (m, 20H, CH_2Ph), 6.38 (d, 1H, $^3J_{\text{NH},2}$ = 7.7 Hz, NH), 5.27 (t, 1H, $^3J_{4',5'}$ = $^3J_{4,3'}$ = 9.8 Hz, H-4'), 5.23-5.14 (m, 5H, H-1, H-3, $3\times\beta^{\text{Myr}}\text{-CH}$), 5.12 (d, 1H, $^3J_{1',2'}$ = 1.6 Hz, H-1'), 5.08-5.04 (m, 1H, $\beta^{\text{Myr}}\text{-CH}$), 4.98-4.94 (m, 4H, $\text{OP(O)(OCH}_2\text{Ph)}_2$), 4.68 (AB, 1H, 2J = 11.9 Hz, CH_2Ph), 4.64 (AB, 1H, 2J = 12.0 Hz, CH_2Ph), 4.47 (AB, 1H, 2J = 11.9 Hz, CH_2Ph), 4.45-4.41 (m, 1H, H-4), 4.40 (AB, 1H, 2J = 11.8 Hz, CH_2Ph), 4.25 (ddd, 1H, $^3J_{2,1}$ = 3.5 Hz, $^3J_{2,3}$ = 11.2 Hz, $^3J_{2,\text{NH}}$ = 7.9 Hz, H-2), 4.16 (dd, 1H, J = 5.5 Hz, J = 12.2 Hz, H-6a'), 3.99-3.97 (m, 2H, H-3', H-6b'), 3.79-3.70 (m, 3H, H-5, H-5', H-6a), 3.58 (dd, 1H, J = 5.6 Hz, J = 10.7 Hz, H-6b), 3.28 (s, 3H, CH_3 , Me), 3.27 (br. s, 1H, H-2'), 2.61-2.18 (m, 16H, $5\times\alpha^{\text{Myr}}\text{-CH}_2$, $3\times\alpha^{\text{Lau}}$

CH₂), 1.63-1.49 (m, 16H, β^{Myr}-CH₂, 3×β^{Lau}-CH₂, 4×γ^{Myr}-CH₂), 1.31-1.22 (m, 140H, 70×CH₂), 0.89-0.87 (m, 24H, 5×ω^{Myr}-CH₃, 3×ω^{Lau}-CH₃); ¹³C NMR (151 MHz, CDCl₃): δ 173.78, 173.32, 173.00, 172.91, 171.41, 170.27, 169.68, 168.99 (8×CO), 138.02, 137.88 (2×Cq, CH₂Ph), 135.56, 135.51 (2×Cq, OP(O)(OCH₂Ph)₂), 128.70, 128.63, 128.53, 128.31, 128.11, 128.08, 128.02, 127.89, 127.63, 127.57 (20×CH, CH₂Ph), 92.86 (C-1'), 92.46 (C-1), 77.50 (C-2'), 76.14 (C-3'), 73.68 (C-4), 73.47, 72.52 (2×CH₂, CH₂Ph), 71.11 (C-3), 70.61 (C-5, ³J_{C5,P} = 5.6 Hz), 70.56 (β^{Myr}-CH), 70.15, 70.09, 69.87, 69.80 (C-5', 3×β^{Myr}-CH), 69.74 (OP(O)(OCH₂Ph)₂, ²J_{C,P} = 5.2 Hz), 69.72 (OP(O)(OCH₂Ph)₂, ²J_{C,P} = 5.4 Hz), 68.35 (C-6), 67.47 (C-4'), 62.45 (C-6'), 59.11 (CH₃, Me), 51.65 (C-2), 41.57, 38.97, 38.87, 38.77 (4×α^{Myr}-CH₂), 34.79, 34.53, 34.47, 34.43, 34.42, 34.26, 33.99, 33.94 (α^{Myr}-CH₂, 3×α^{Lau}-CH₂, 4×γ^{Myr}-CH₂), 31.94, 31.93, 29.78, 29.76, 29.74, 29.67, 29.66, 29.58, 29.57, 29.54, 29.47, 29.42, 29.40, 29.38, 29.37, 29.32, 29.22, 29.20, 25.39, 25.18, 25.15, 25.11, 25.10, 25.02, 22.69 (74×CH₂), 14.10 (5×ω^{Myr}-CH₃, 3×ω^{Lau}-CH₃); ³¹P NMR (243 MHz, CDCl₃): δ -1.71; HRMS (ESI): m/z calcd for C₁₄₇H₂₄₆NO₂₅P+Na⁺: 2479.7639 [M+Na⁺]; found 2479.7628;

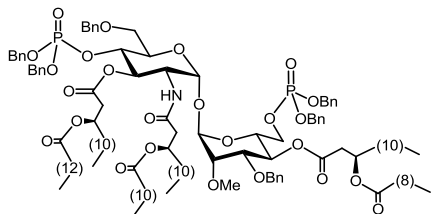
Synthesis of 60



3-*O*-Benzyl-4,6-di-*O*-[(*R*)-3-(decanoyloxy)tetradecanoyl]-2-*O*-methyl-α-*D*-mannopyranosyl-(1↔1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-[(*R*)-3-(dodecanoyloxy)-tetradecanoylamino]-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanoyl]-α-*D*-glucopyranoside (60)

by-product from reaction of **56** with fatty acid **51** (to furnish **58** as major product), *R_f* = 0.60 (toluene – EtOAc, 3 : 1); [α]_D²⁰ = +36 (*c* 0.5, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.39-7.21 (m, 20H, CH₂Ph), 6.44 (d, 1H, ³J_{NH,2} = 8.2 Hz, *NH*), 5.27 (t, 1H, *J* = 9.8 Hz, H-4'), 5.23-5.14 (m, 5H, H-1, H-3, 3×β^{Myr}-CH), 5.12 (d, 1H, ³J_{1,2'} = 1.9 Hz, H-1'), 5.08-5.04 (m, 1H, β^{Myr}CH), 4.97 (d, 2H, *J* = 8.2 Hz, OP(O)(OCH₂Ph)₂), 4.94 (d, 2H, *J* = 8.2 Hz, OP(O)(OCH₂Ph)₂), 4.68 (AB, 1H, ²J = 12.1 Hz, CH₂Ph), 4.64 (AB, 1H, ²J = 12.0 Hz, CH₂Ph), 4.47 (AB, 1H, ²J = 11.9 Hz, CH₂Ph), 4.45-4.41 (m, 1H, H-4), 4.40 (AB, 1H, ²J = 11.9 Hz, CH₂Ph), 4.27-4.23 (m, 1H, H-2), 4.16 (dd, 1H, *J* = 5.5 Hz, *J* = 12.2 Hz, H-6a'), 4.00-3.96 (m, 2H, H-3', H-6b'), 3.79-3.76 (m, 1H, H-5'), 3.75-3.70 (m, 2H, H-5, H-6a), 3.58 (dd, 1H, *J* = 5.6 Hz, *J* = 10.7 Hz, H-6b), 3.28 (s, 3H, CH₃, Me), 3.26-3.25 (m, 1H, H-2), 2.61-2.36 (m, 8H, 4×α^{Myr}-CH₂), 2.36-2.17 (m, 8H, α^{Myr}-CH₂, α^{Lau}-CH₂, 2×α^{Cap}-CH₂), 1.63-1.46 (m, 16H, 4×γ^{Myr}-CH₂, β^{Myr}-CH₂, β^{Lau}-CH₂, 2×β^{Cap}-CH₂), 1.33-1.20 (m, 132H, 66×CH₂), 0.89-0.86 (m, 24H, 5×ω^{Myr}-CH₃, ω^{Lau}-CH₃, 2×ω^{Cap}-CH₃); ¹³C-NMR (151 MHz, CDCl₃): δ 173.77, 173.33, 173.01, 172.94, 171.41, 170.28, 169.70, 168.99 (8×CO), 138.02, 137.88 (2×Cq, CH₂Ph), 135.55, 135.51 (2×Cq, OP(O)(OCH₂Ph)₂), 128.70, 128.63, 128.53, 128.31, 128.12, 128.09, 128.02, 127.90, 127.63, 127.57 (20×CH, CH₂Ph), 92.87 (C-1'), 92.49 (C-1), 77.50 (C-2'), 76.11 (C-3'), 73.68 (C-4, ²J_{C4,P} = 5.5 Hz), 73.47, 72.51 (2×CH₂Ph), 71.10 (C-3, ³J_{C3,P} = 2.0 Hz), 70.64, 70.60, 70.58 (C-5, β^{Myr}-CH), 70.15, 70.08, 69.89, 69.81 (C-5', 3×β^{Myr}-CH), 69.76, 69.74, 69.73 (OP(O)(OCH₂Ph)₂), 68.35 (C-6), 67.47 (C-4'), 62.44 (C-6'), 59.11 (CH₃, Me), 51.63 (C-2), 41.55, 38.99, 38.88, 38.77 (4×α^{Myr}-CH₂), 34.78, 34.54, 34.47, 34.43, 34.42, 34.26, 34.00, 33.94 (4×γ^{Myr}-CH₂, α^{Myr}-CH₂, α^{Lau}-CH₂, 2×α^{Cap}-CH₂), 31.93, 31.88, 29.78, 29.76, 29.74, 29.71, 29.69, 29.67, 29.66, 29.64, 29.57, 29.54, 29.53, 29.51, 29.47, 29.42, 29.40, 29.37, 29.35, 29.32, 29.31, 29.20, 29.19, 25.39, 25.17, 25.15, 25.11, 25.09, 25.01, 22.69 (70×CH₂), 14.09 (5×ω^{Myr}-CH₃, ω^{Lau}-CH₃, 2×ω^{Cap}-CH₃); ³¹P-NMR (243 MHz, CDCl₃): δ -1.71; HRMS (ESI): m/z calcd for C₁₄₃H₂₃₈NO₂₅P+Na⁺: 2423.7013[M+Na⁺]; found 2423.6967;

Synthesis of 62



3-*O*-Benzyl-6-*O*-[bis(benzyloxy)phosphoryl]-4-*O*-[(*R*)-3-(decanoyloxy)tetradecanoyl]-2-*O*-methyl- α -mannopyranosyl-(1 \leftrightarrow 1)-6-*O*-benzyl-4-*O*-[bis(benzyloxy)phosphoryl]-2-deoxy-2-[(*R*)-3-(dodecanoyloxy)tetradecanoylamino]-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanoyl]- α -D-glucopyranoside (62).

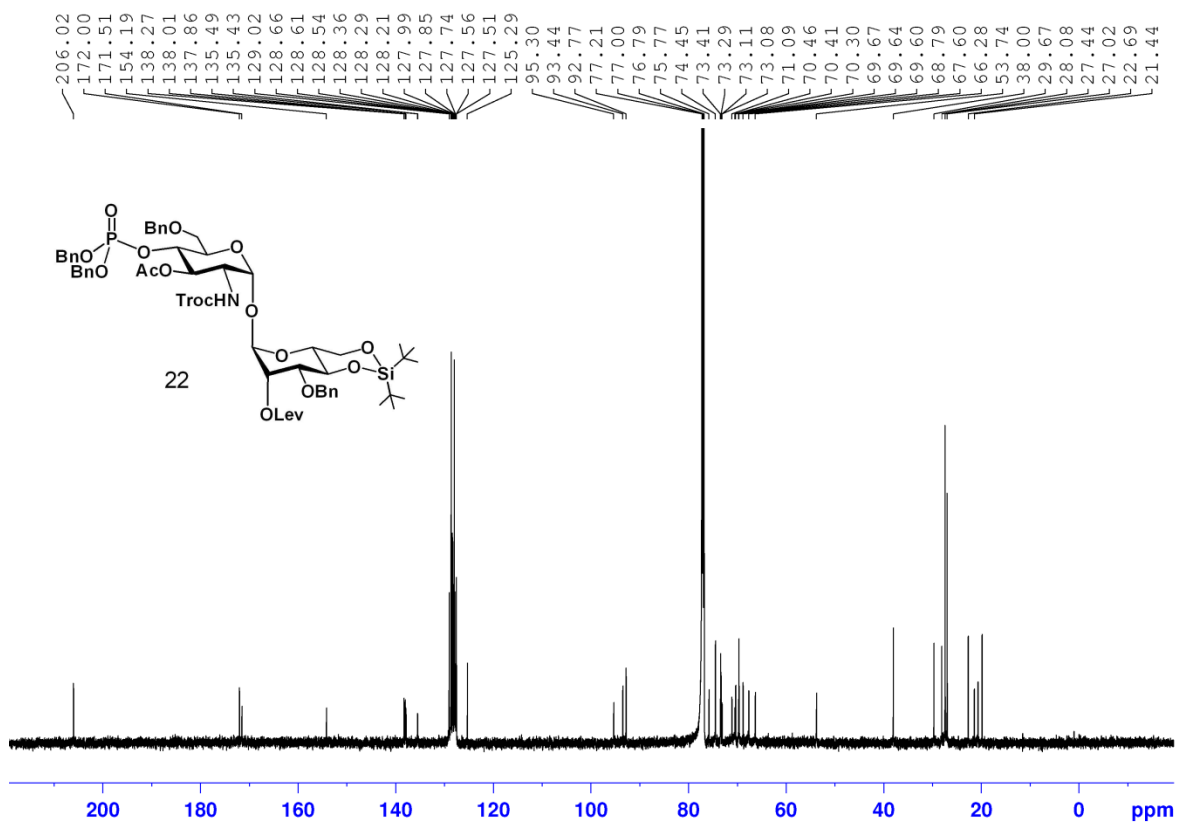
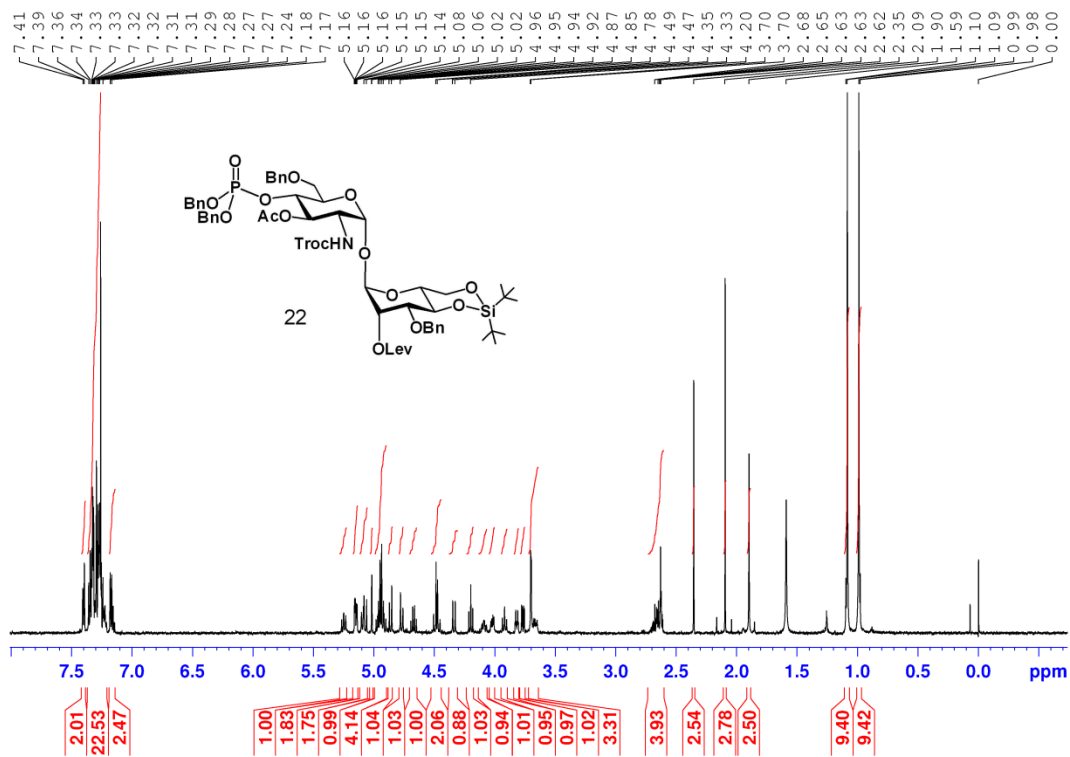
To a stirred solution of **58** (18 mg, 8.9 μ mol) and dibenzyl *N,N*-diisopropylphosphoramidite (10 μ L, 27 μ mol, 90%) in dry CH_2Cl_2 (400 μ L) a solution of 1*H*-tetrazole (100 μ L, 45 μ mol, 0.45 M in CH_3CN) was added under atmosphere of Ar. The mixture was stirred for 1 h, cooled to -78° and a solution of *m*CPBA (10 mg, 41 μ mol, 70%) in CH_2Cl_2 (150 μ L of a stock solution prepared from 20 mg in 300 μ L CH_2Cl_2) was added. After stirring for 1 h, the reaction was quenched by addition of Et_3N (15 μ L), the mixture was warmed to r. t., diluted with EtOAc (20 ml) and washed with aq. citric acid (0.25 M, 20 ml), sat. aq. NaHCO_3 (20 ml) and brine (20 ml). The organic layer was dried over Na_2SO_4 , filtered and concentrated. The residue was purified by HPLC (two successive columns: toluene – EtOAc, 10 : 1 \rightarrow 5 : 1 and hexane : EtOAc, 2 : 1 \rightarrow 1 : 1, column B) to afford **62** (18 mg, 7.9 μ mol, 88%) as a syrup. R_f = 0.53 (toluene – EtOAc, 1 : 1), $[\alpha]_D^{20}$ = +32 (*c* 1.0, CHCl_3); $^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 7.38-7.20 (m, 30H, CH_2Ph), 6.46 (d, 1H, $^3J_{\text{NH},2}$ = 8.5 Hz, NH), 5.30 (t, 1H, $^3J_{4',3'} = ^3J_{4',5'} = 9.7$ Hz, H-4'), 5.23-5.16 (m, 3H, H-3, $2 \times \beta^{\text{Myr}}\text{-CH}$), 5.15 (d, 1H, $^3J_{1',2'} = 3.7$ Hz, H-1), 5.10-5.06 (m, 1H, $\beta^{\text{Myr}}\text{-CH}$), 5.10 (d, 1H, $^3J_{1',2'} = 1.9$ Hz, H-1'), 5.03-4.93 (m, 8H, $\text{OP(O)(OCH}_2\text{Ph)}_2$), 4.68 (AB, 1H, $^2J = 12.1$ Hz, CH_2Ph), 4.63 (AB, 1H, $^2J = 12.1$ Hz, CH_2Ph), 4.45 (AB, 1H, $^2J = 11.8$ Hz, CH_2Ph), 4.43-4.39 (m, 1H, H-4), 4.38 (AB, 1H, $^2J = 11.9$ Hz, CH_2Ph), 4.28 (ddd, 1H, $^3J_{2,1} = 3.5$ Hz, $^3J_{2,\text{NH}} = 8.3$ Hz, $^3J_{2,3} = 11.1$ Hz, H-2), 4.09-4.03 (m, 2H, H-6a', H-6b'), 4.01 (dd, 1H, $^3J_{3',2'} = 3.0$ Hz, $^3J_{3',4'} = 9.5$ Hz, H-3'), 3.89-3.86 (m, 1H, H-5'), 3.76-3.69 (m, 2H, H-5, H-6a), 3.56 (dd, 1H, $J = 5.9$ Hz, $J = 10.7$ Hz, H-6b), 3.28 (t, 1H, $J = 2.5$ Hz, H-2'), 3.23 (s, 3H, CH_3 , Me), 2.58-2.44 (m, 5H, $\alpha^{\text{Myr}}\text{-CH}_2$), 2.40 (dd, 1H, $J = 4.7$ Hz, $J = 15.6$ Hz, $\alpha^{\text{Myr}}\text{-CH}_2$), 2.34-2.25 (m, 2H, $\alpha\text{-CH}_2$), 2.20-2.16 (m, 4H, $2 \times \alpha\text{-CH}_2$), 1.61-1.49 (m, 12H, $3 \times \gamma^{\text{Myr}}\text{-CH}_2$, $\beta^{\text{Myr}}\text{-CH}_2$, $\beta^{\text{Lau}}\text{-CH}_2$, $\beta^{\text{Cap}}\text{-CH}_2$), 1.32-1.20 (m, 102H, $51 \times \text{CH}_2$), 0.89-0.86 (m, 18H, $4 \times \omega^{\text{Myr}}\text{-CH}_3$, $\omega^{\text{Lau}}\text{-CH}_3$, $\omega^{\text{Cap}}\text{-CH}_3$); $^{13}\text{C-NMR}$ (151 MHz, CDCl_3): δ 173.89, 173.23, 172.92, 171.19, 169.75, 169.07 ($6 \times \text{CO}$), 137.98, 137.85 ($2 \times \text{Cq}$, CH_2Ph), 135.98, 135.94, 135.56, 135.51 ($4 \times \text{Cq}$, $\text{OP(O)(OCH}_2\text{Ph)}_2$), 128.68, 128.62, 128.53, 128.45, 128.43, 128.29, 128.28, 128.27, 128.13, 128.01, 127.94, 127.91, 127.86, 127.63, 127.59 ($30 \times \text{CH}$, CH_2Ph), 93.35 (C-1'), 93.16 (C-1), 77.54 (C-2'), 75.95 (C-3'), 73.67 (C-4, $^2J_{\text{C,P}} = 5.9$ Hz), 73.48, 72.45 ($2 \times \text{CH}_2\text{Ph}$), 71.24 (C-3), 70.93 (C-5', $^3J_{\text{C,P}} = 7.3$ Hz), 70.63 (C-5, $^3J = 5.4$ Hz), 70.58, 69.79, 69.74 ($3 \times \beta^{\text{Myr}}\text{-CH}$), 69.76, 69.72, 69.68 ($\text{OP(O)(OCH}_2\text{Ph)}_2$), 69.28, 69.26 ($\text{OP(O)(OCH}_2\text{Ph)}_2$), 68.46 (C-6), 67.50 (C-4'), 66.09 (C-6', $^2J = 5.0$ Hz), 58.99 (CH_3 , Me), 51.43 (C-2), 41.53, 38.92, 38.75 ($3 \times \alpha^{\text{Myr}}\text{-CH}_2$), 34.75, 34.52, 34.42, 43.39, 34.19, 33.98 ($3 \times \gamma^{\text{Myr}}\text{-CH}_2$, $\alpha^{\text{Myr}}\text{-CH}_2$, $\alpha^{\text{Lau}}\text{-CH}_2$, $\alpha^{\text{Cap}}\text{-CH}_2$), 31.94, 31.92, 31.87, 29.76, 29.74, 29.72, 29.70, 29.69, 29.67, 29.64, 29.62, 29.60, 29.55, 29.54, 29.50, 29.47, 29.46, 29.40, 29.38, 29.36, 29.34, 29.32, 29.30, 29.28, 29.19, 29.17, 25.35, 25.17, 25.14, 25.11, 25.00, 22.68 ($54 \times \text{CH}_2$), 14.09 ($4 \times \omega^{\text{Myr}}\text{-CH}_3$, $\omega^{\text{Lau}}\text{-CH}_3$, $\omega^{\text{Cap}}\text{-CH}_3$); $^{31}\text{P-NMR}$ (243 MHz, CDCl_3): δ -1.61, -1.69; HRMS (ESI): m/z calcd for $\text{C}_{133}\text{H}_{207}\text{NO}_{25}\text{P} + \text{Na}^+$: 2303.4325 [M+Na $^+$]; found 2303.4282.

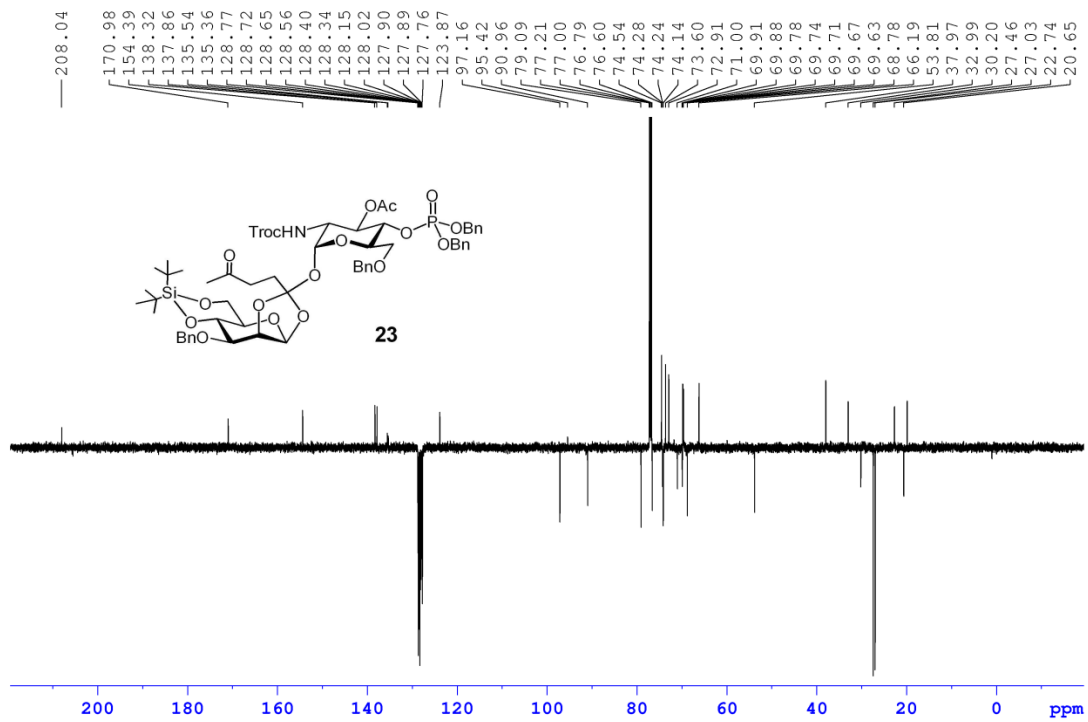
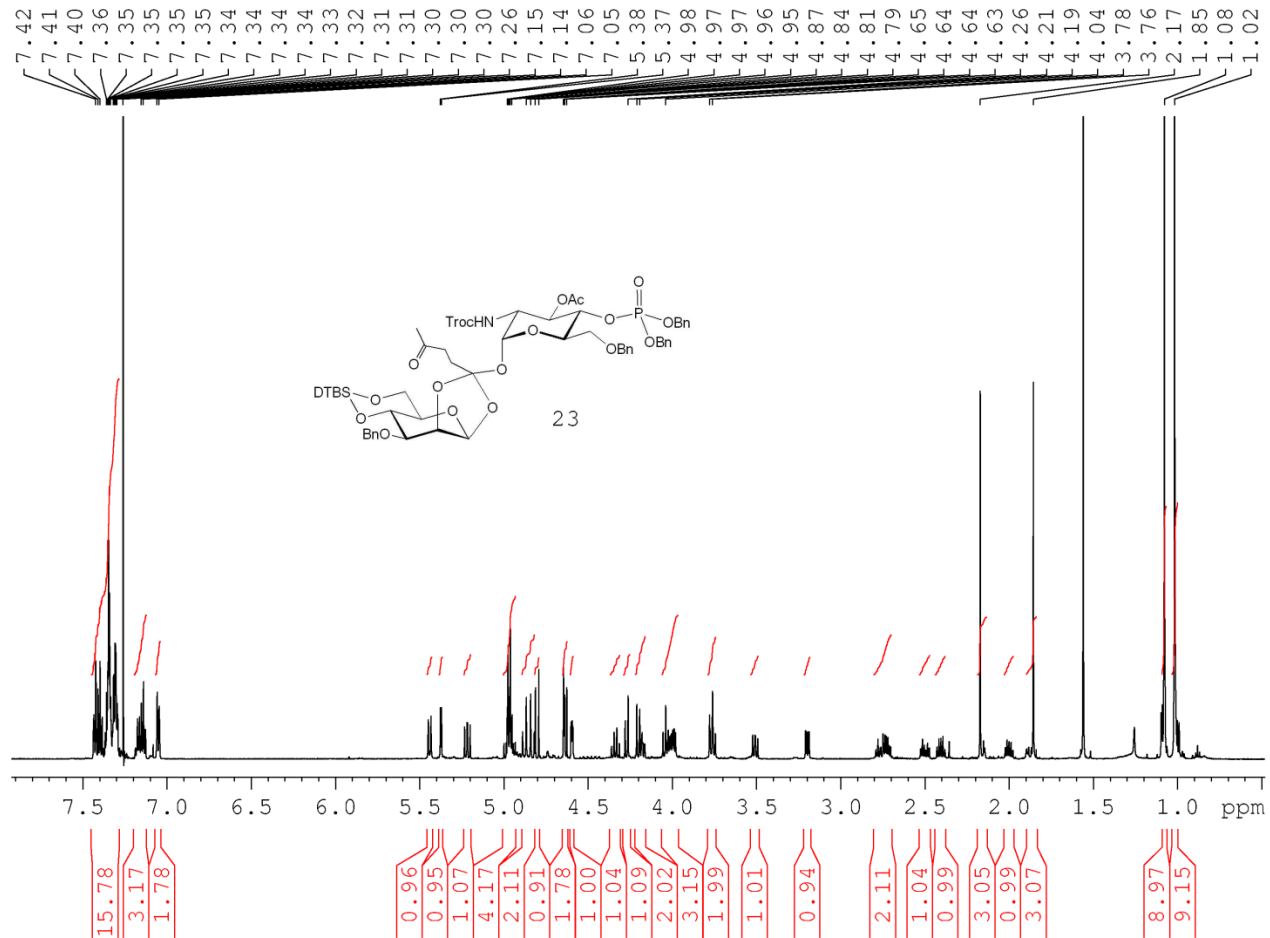
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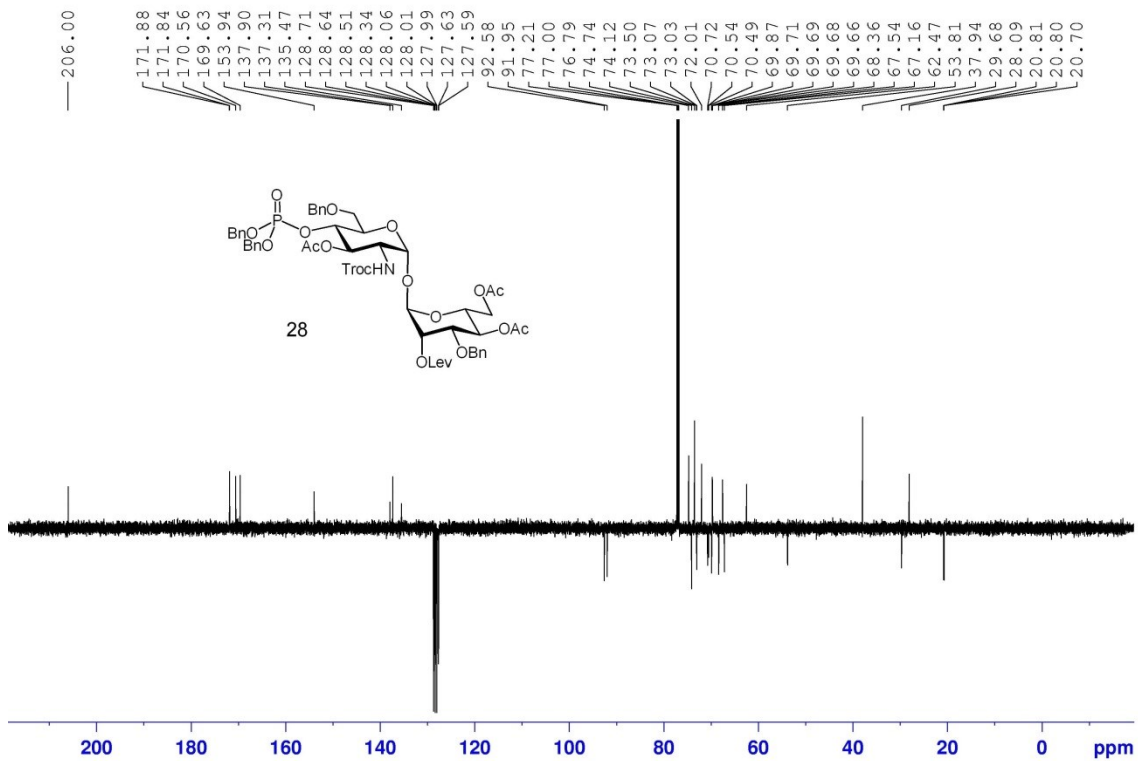
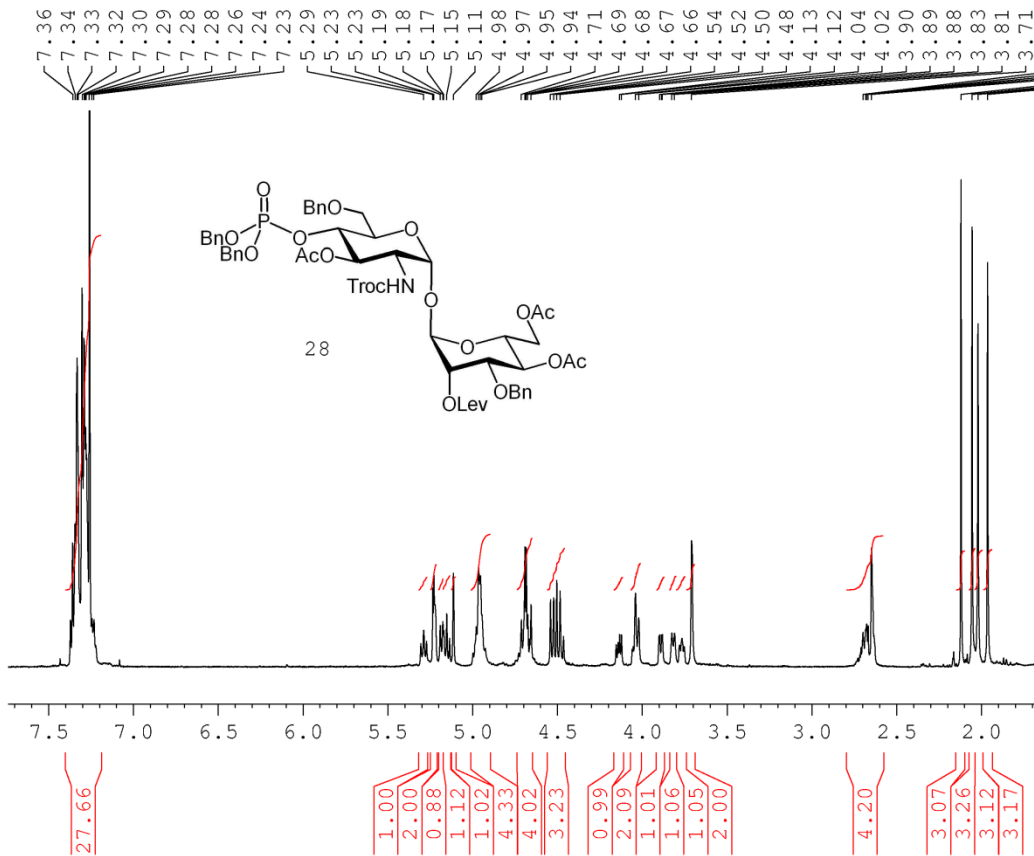
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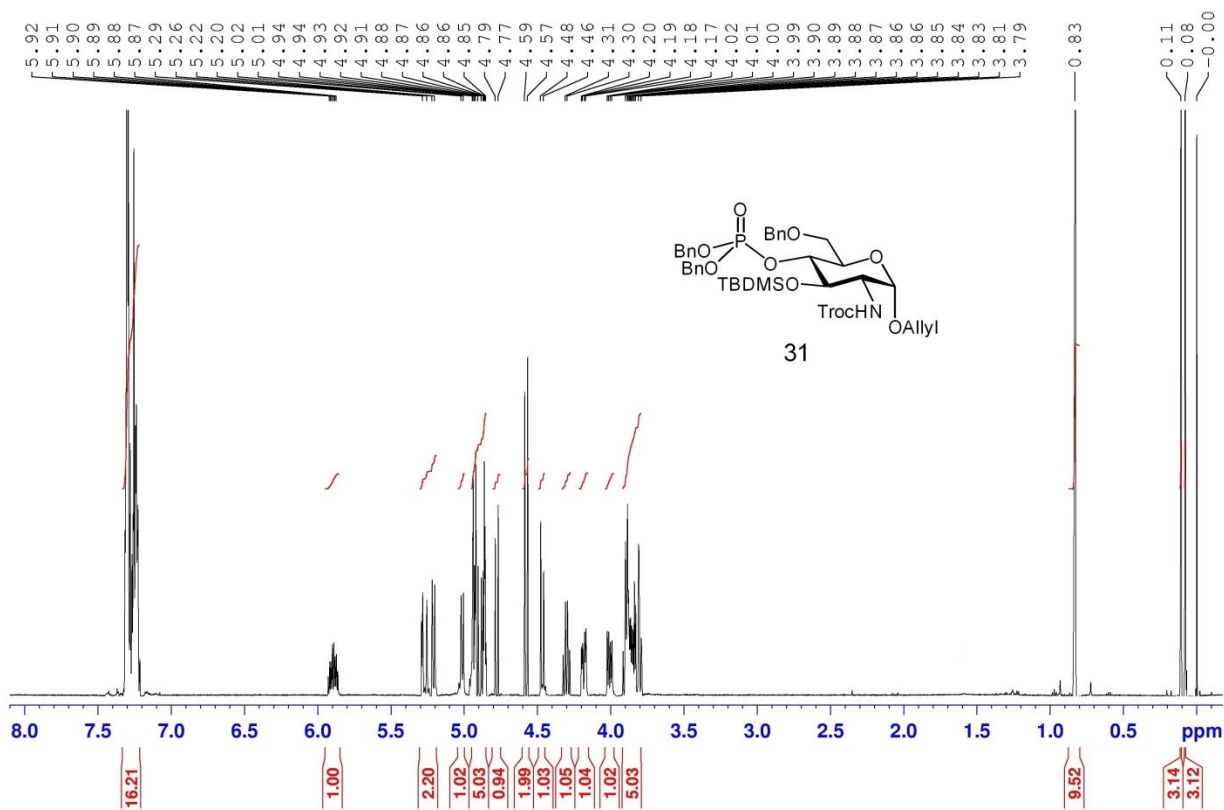
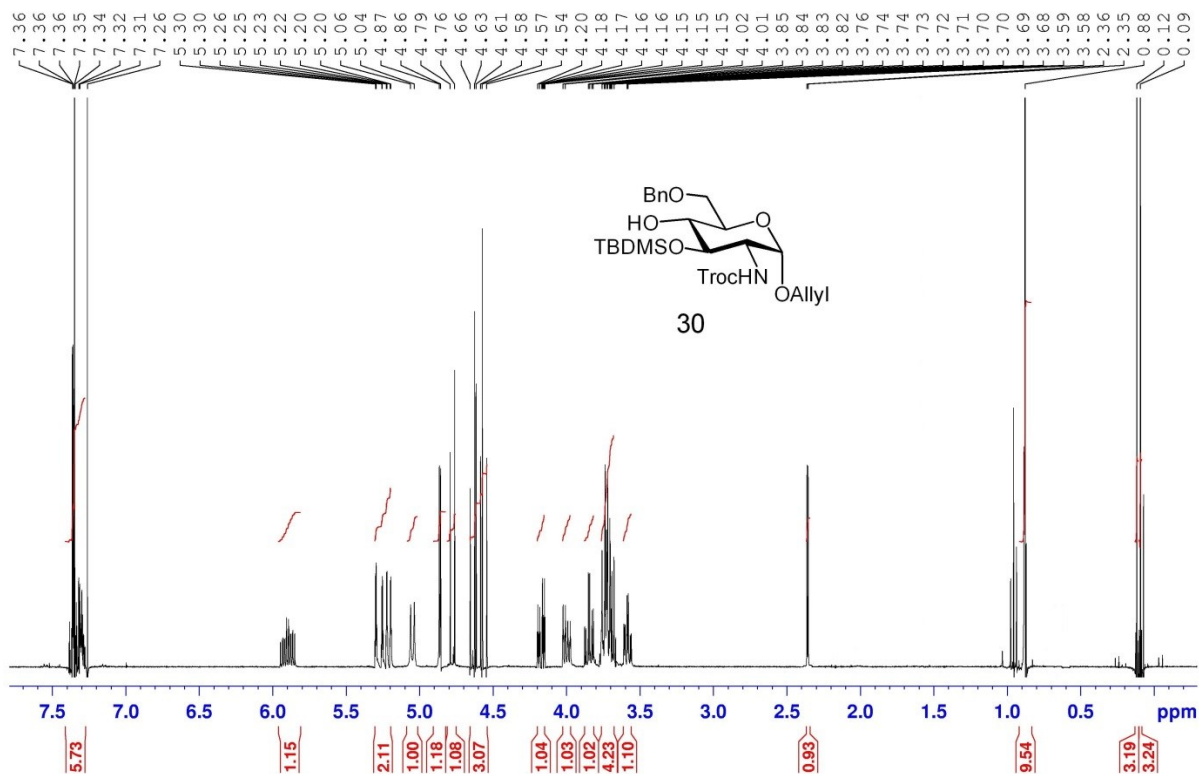
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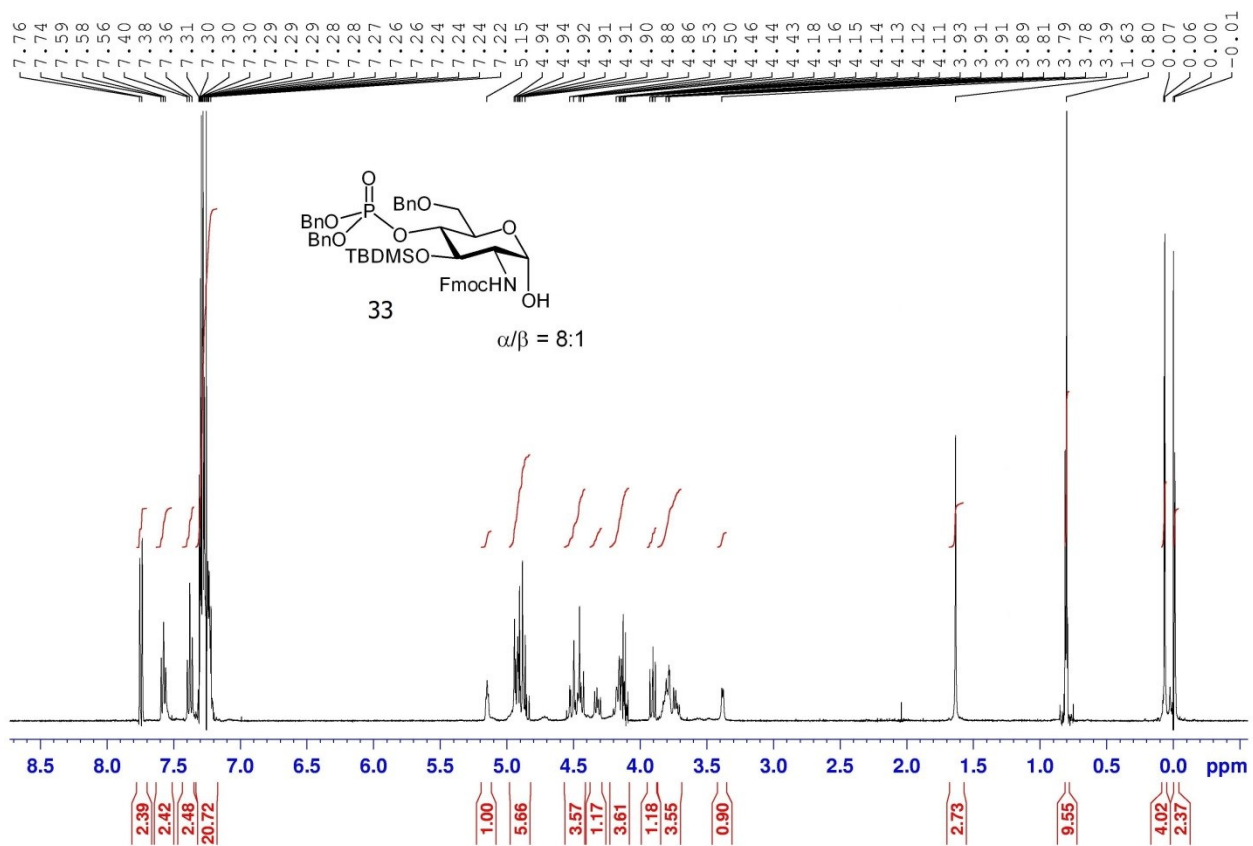
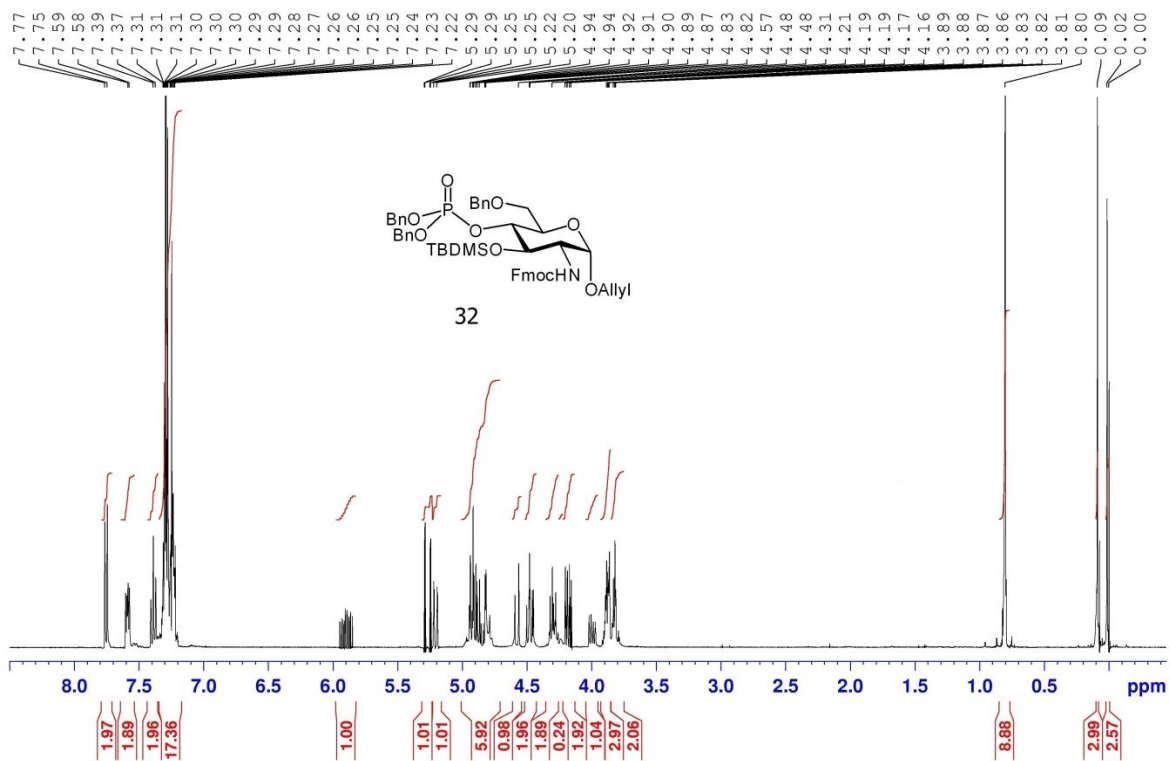
NMR and MALDI-TOF spectra

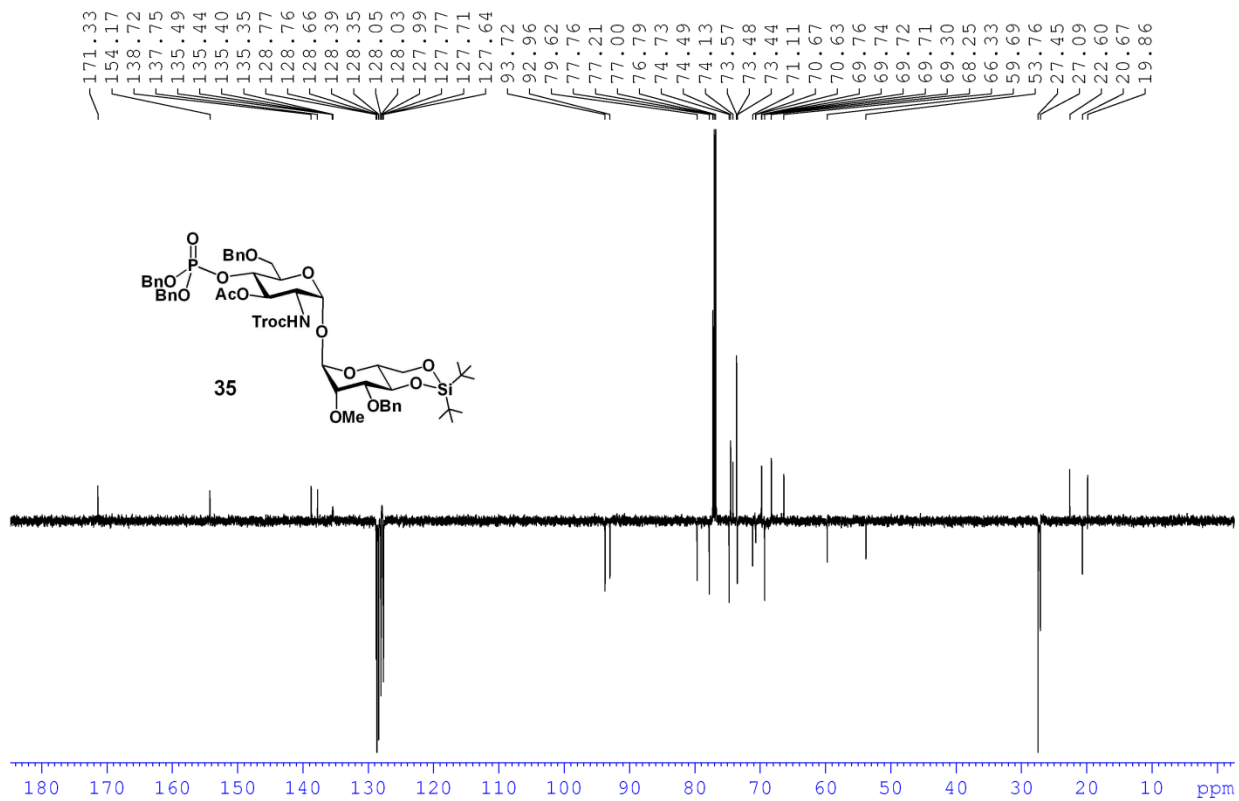
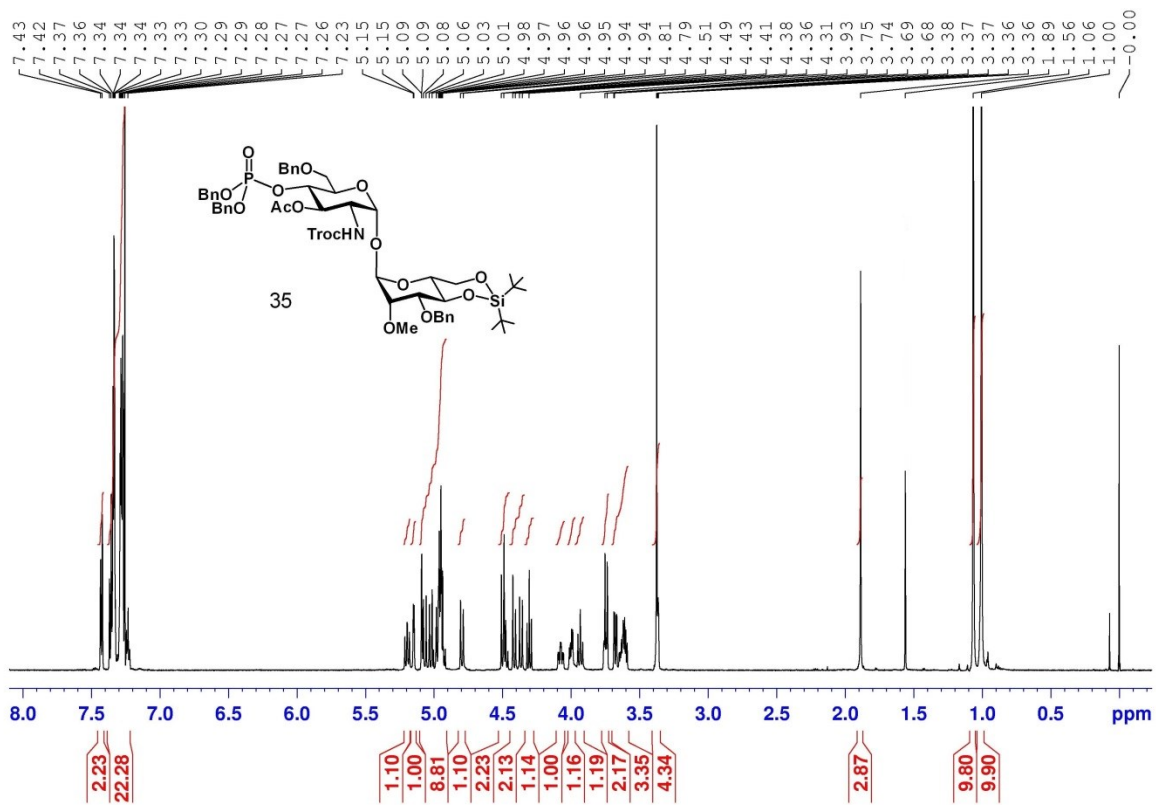


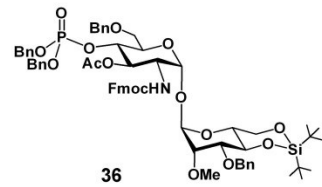
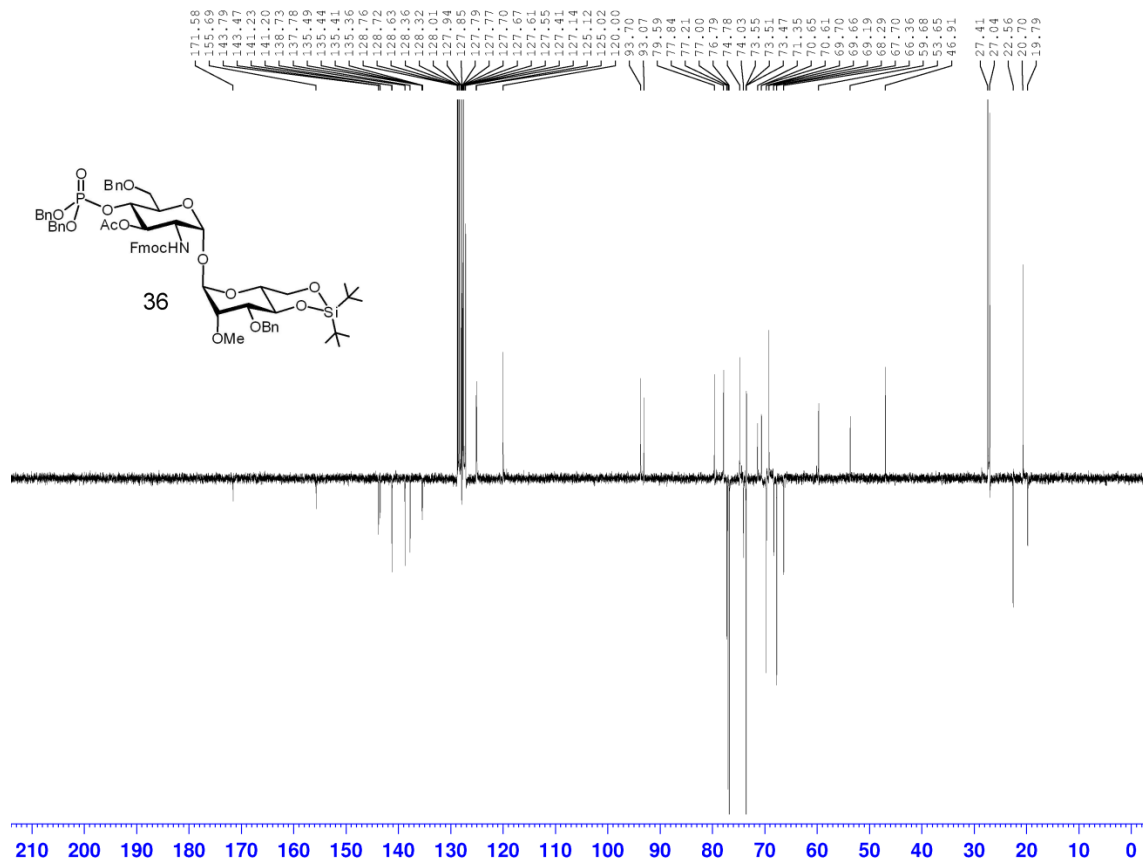
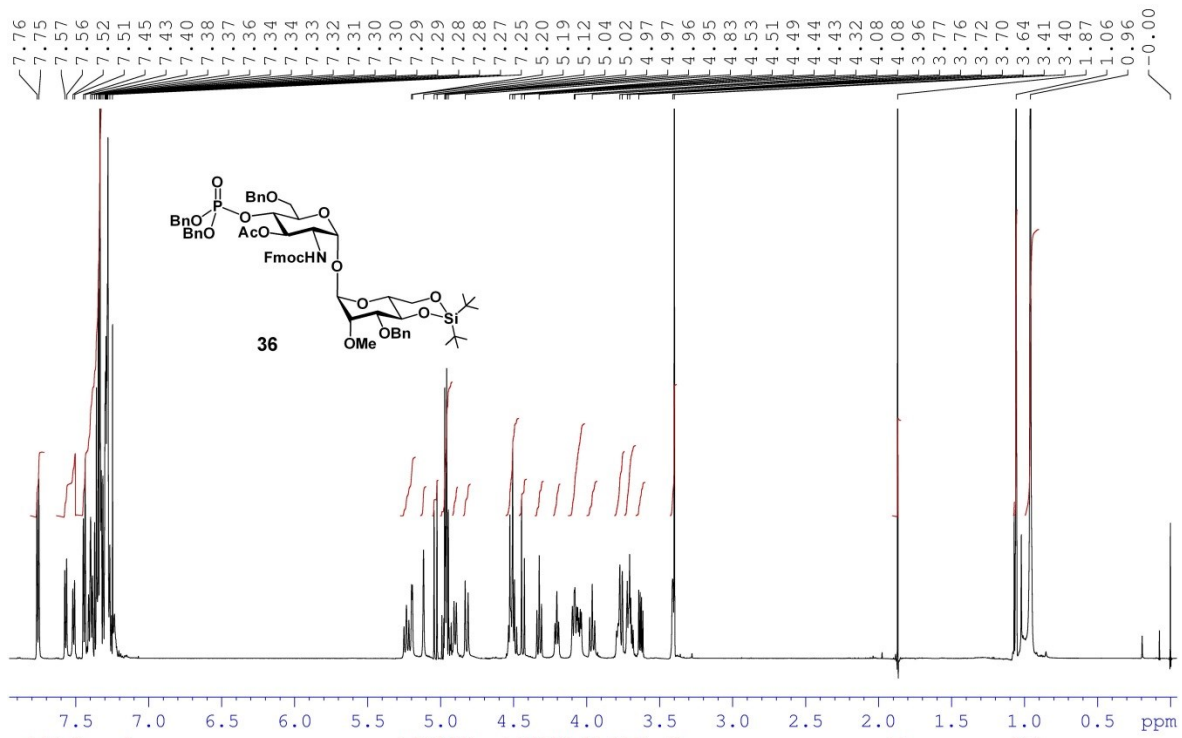


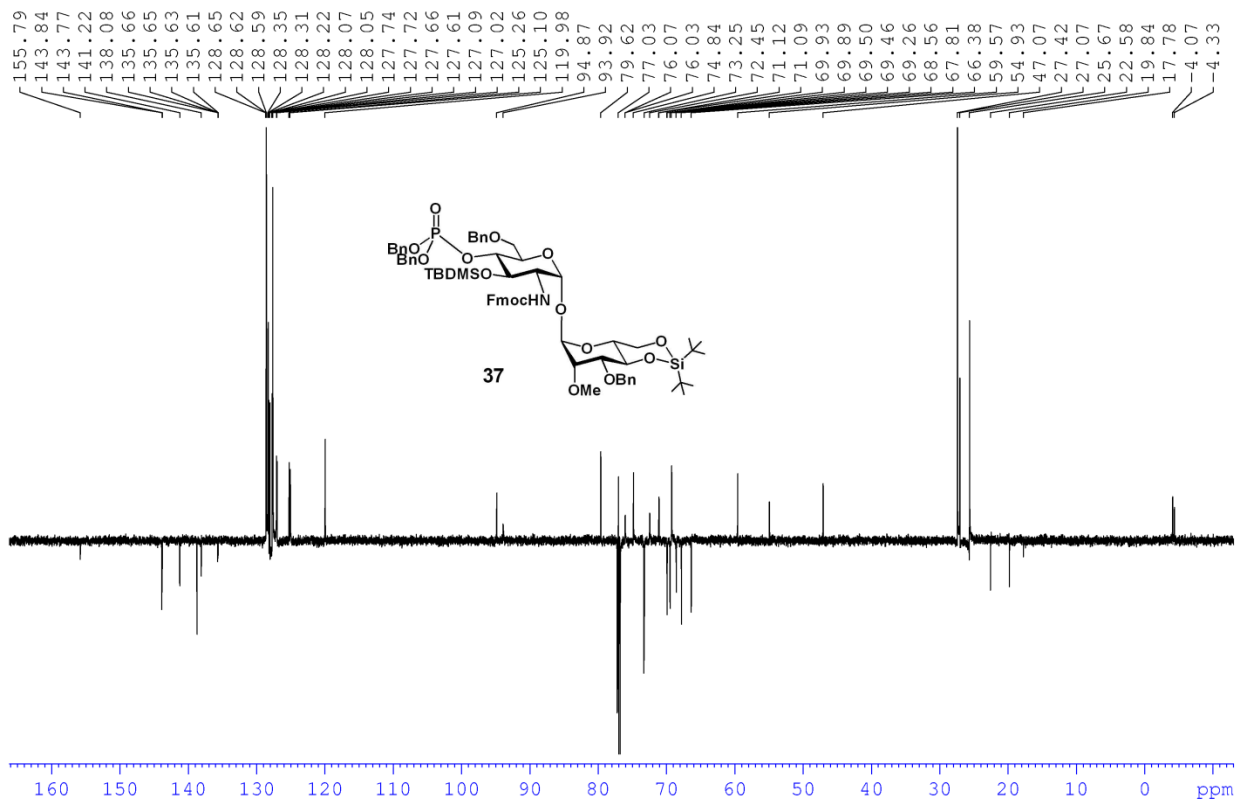
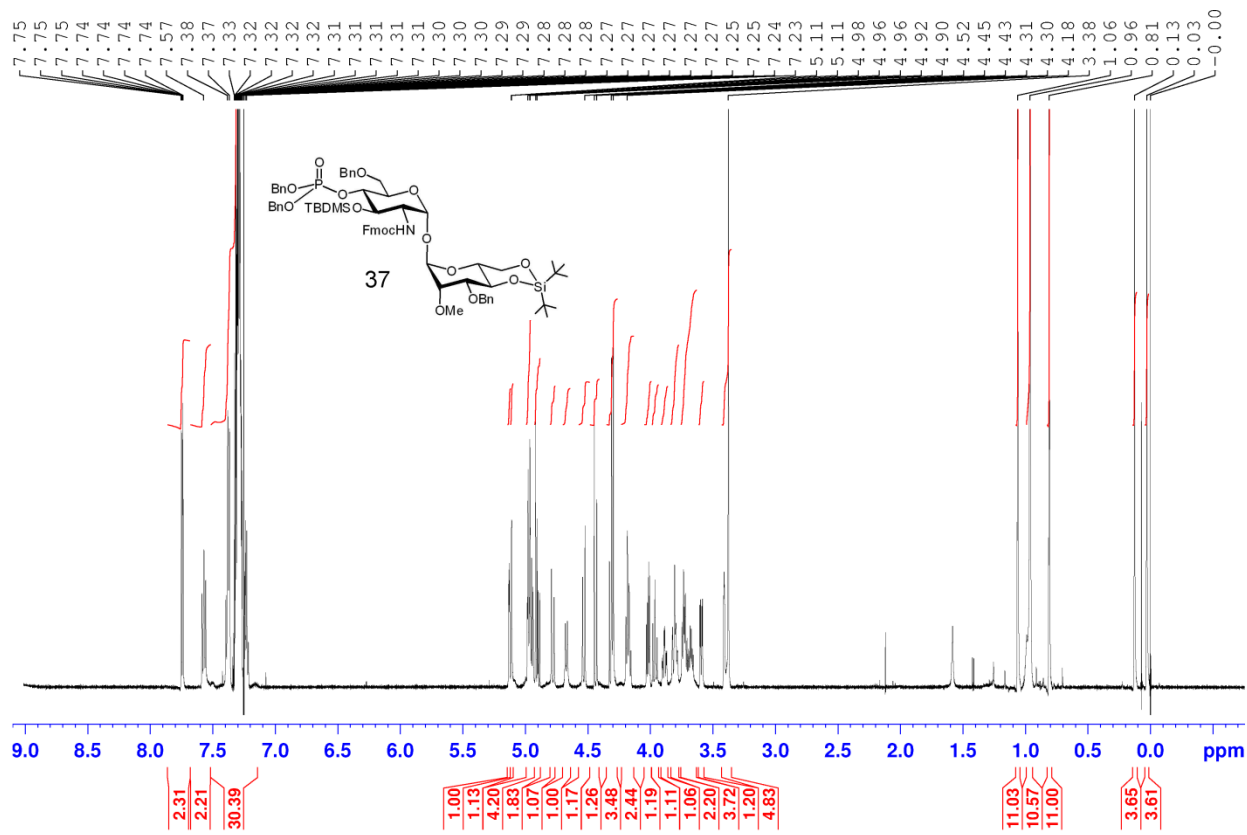


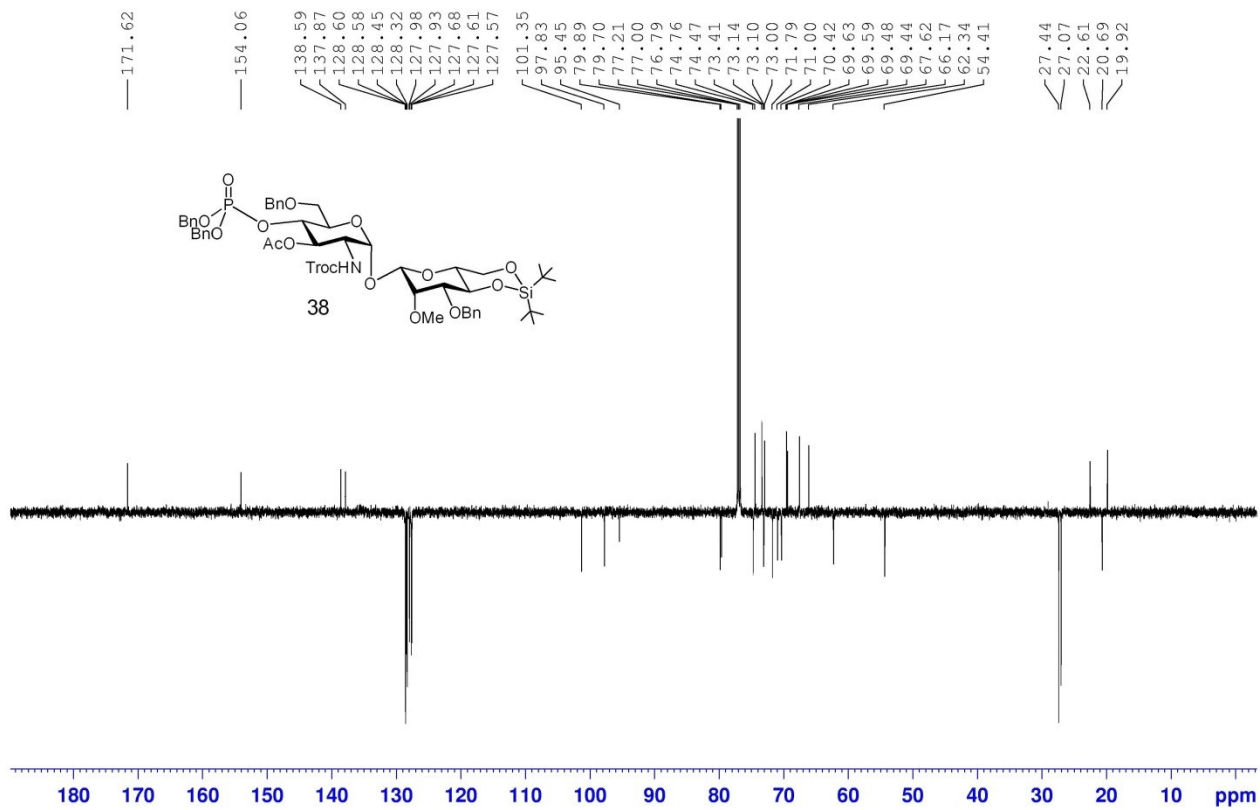
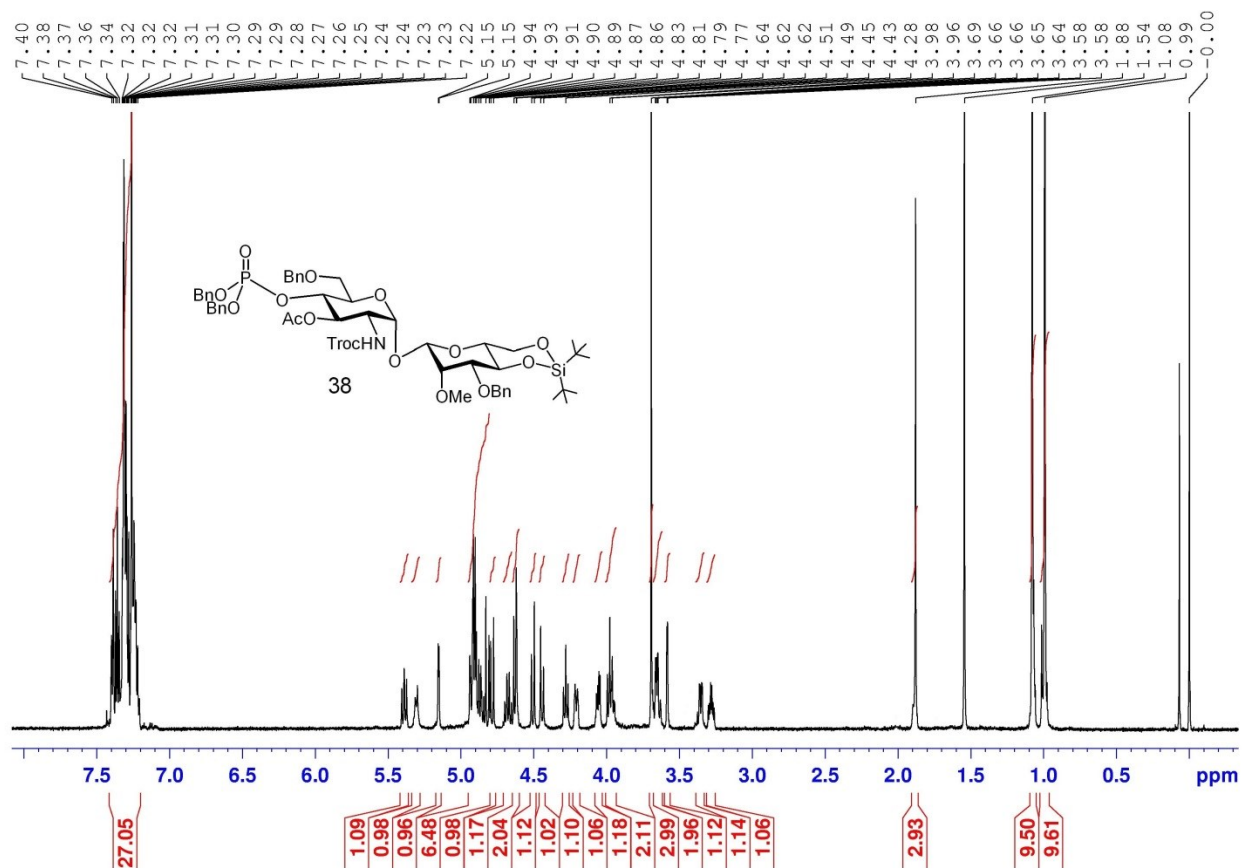


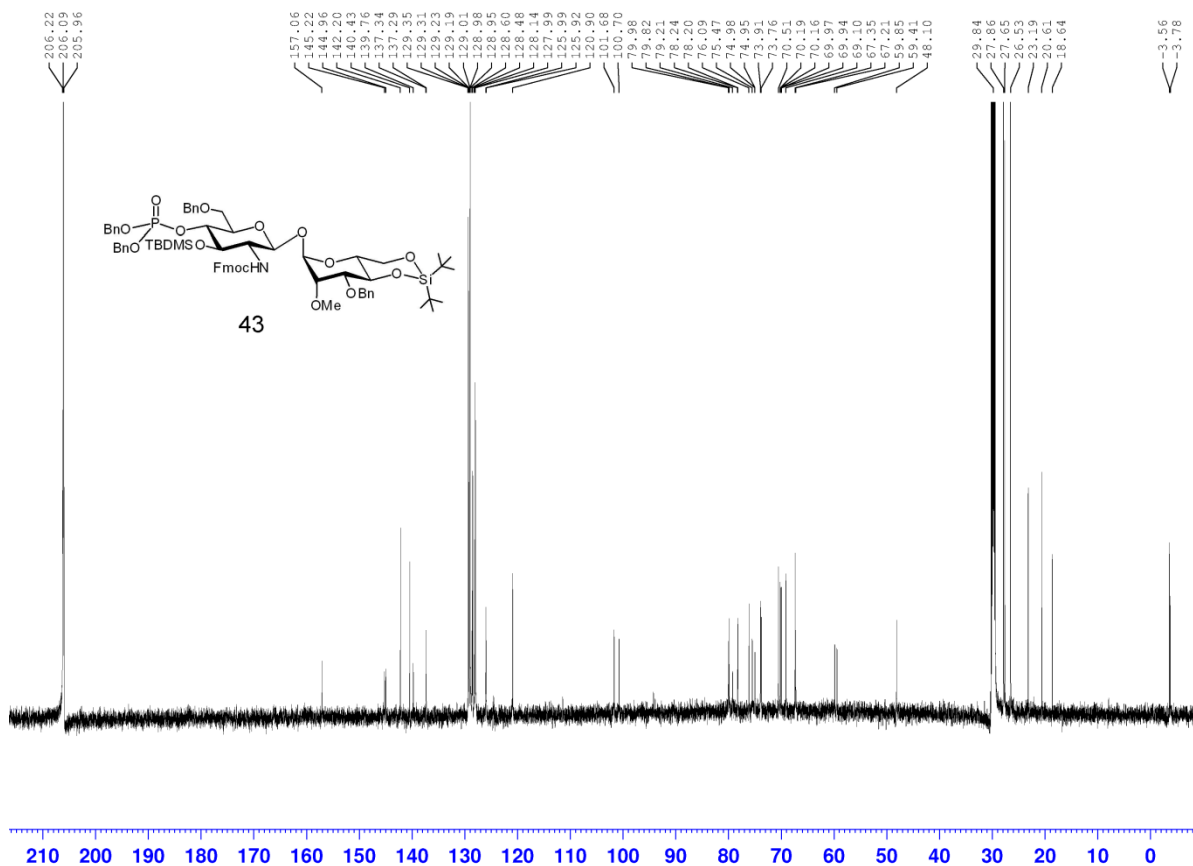
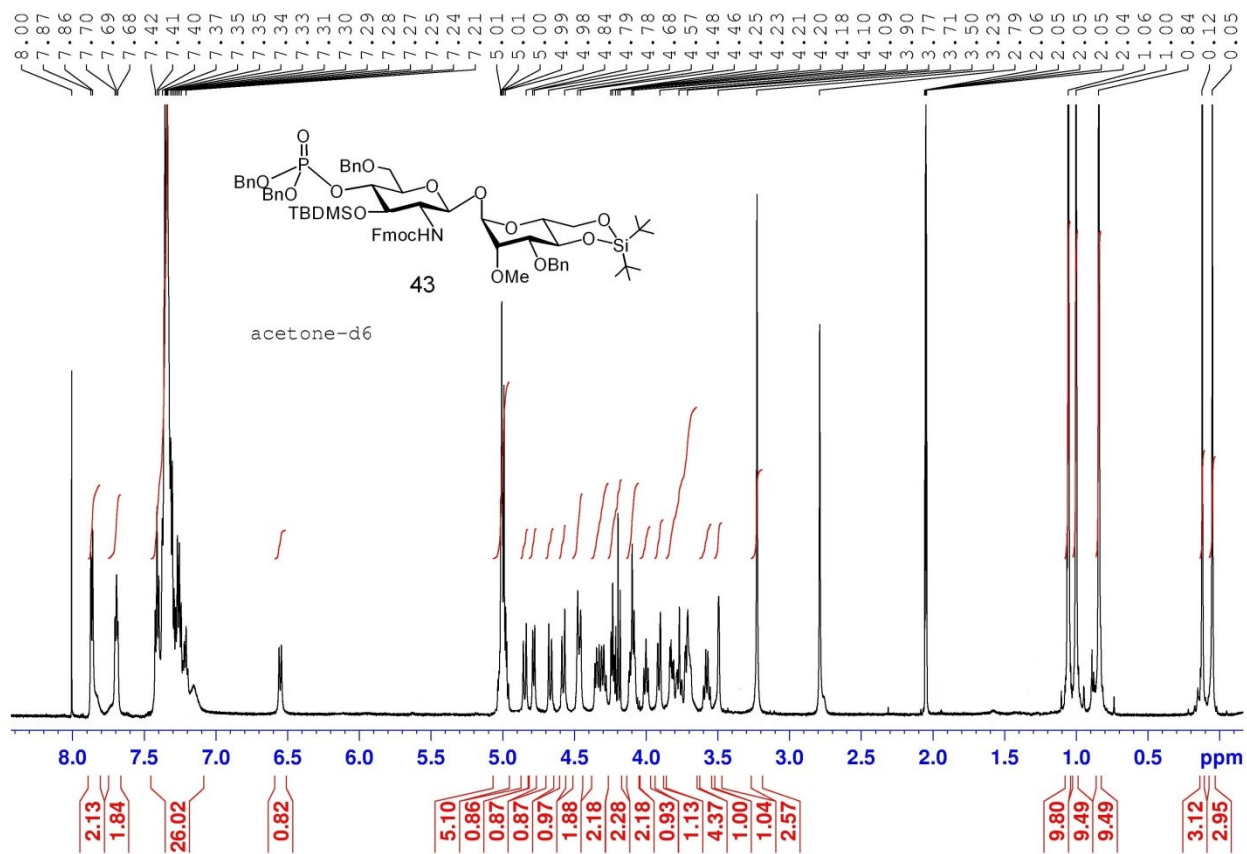


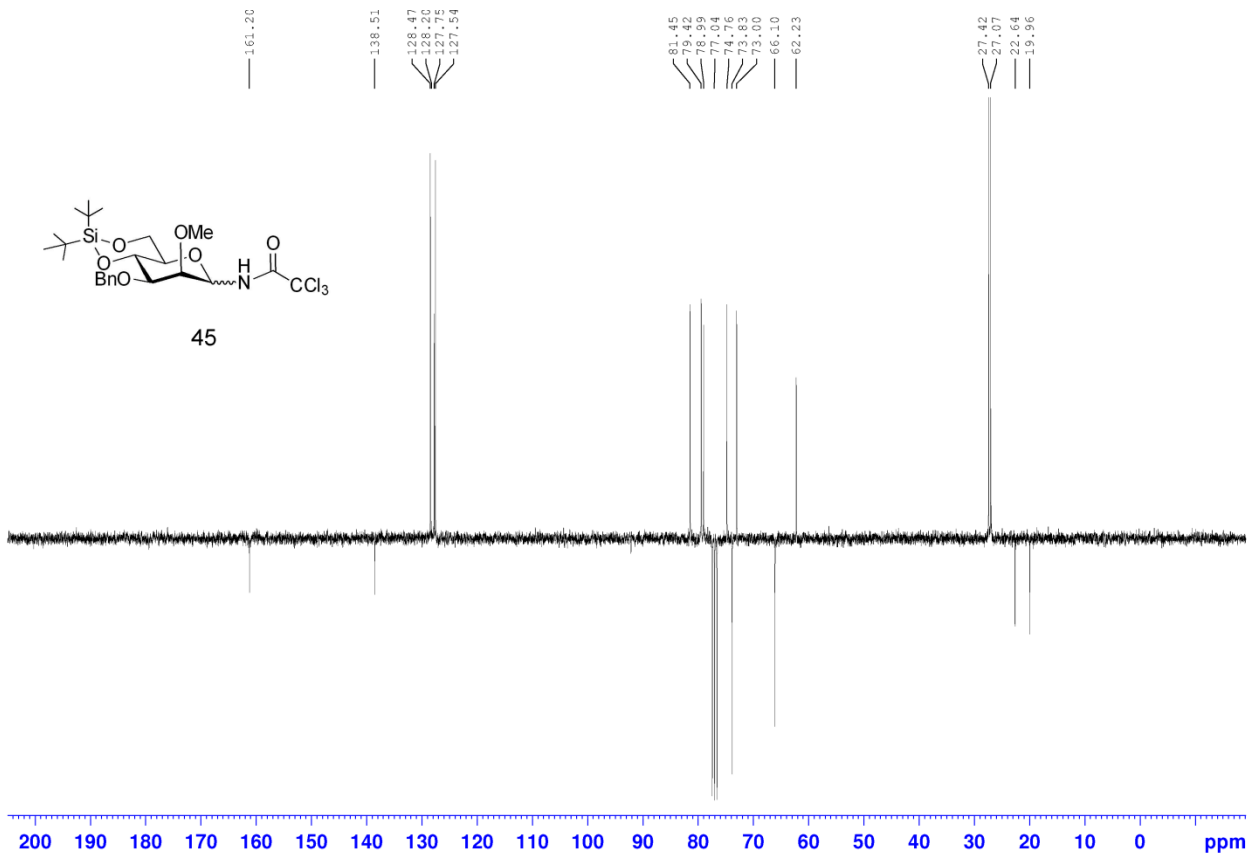
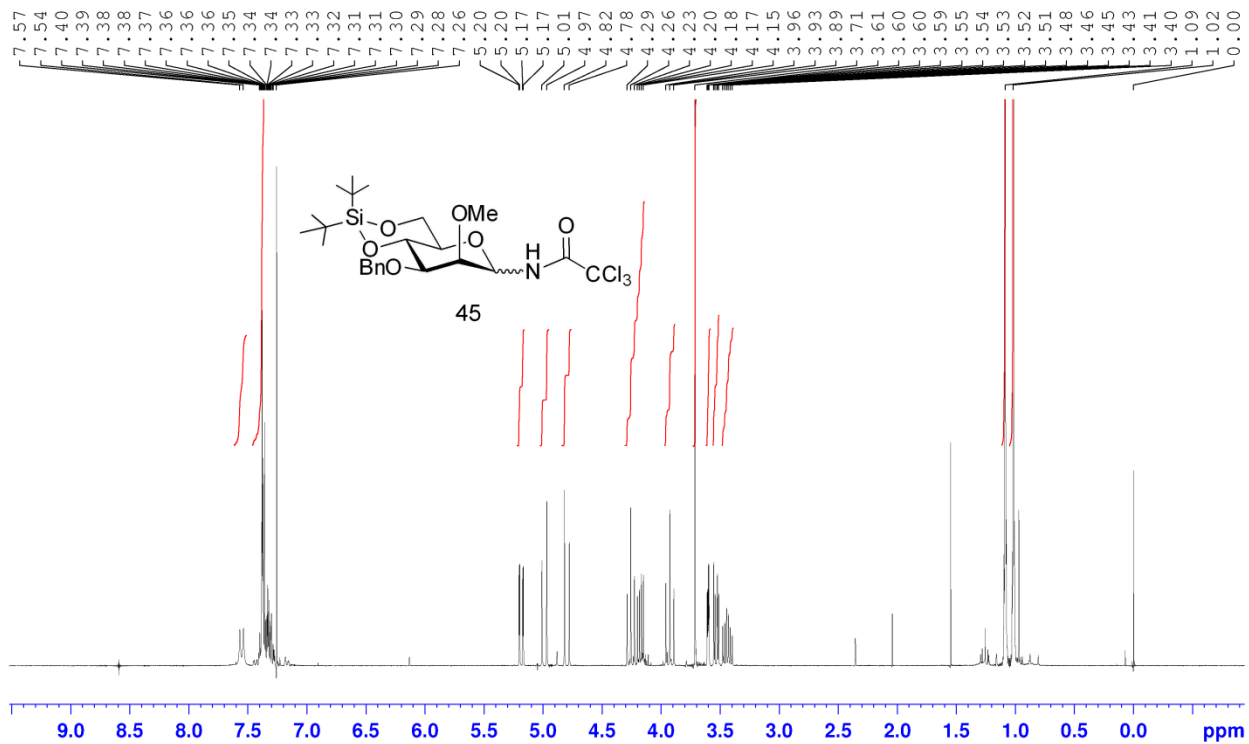


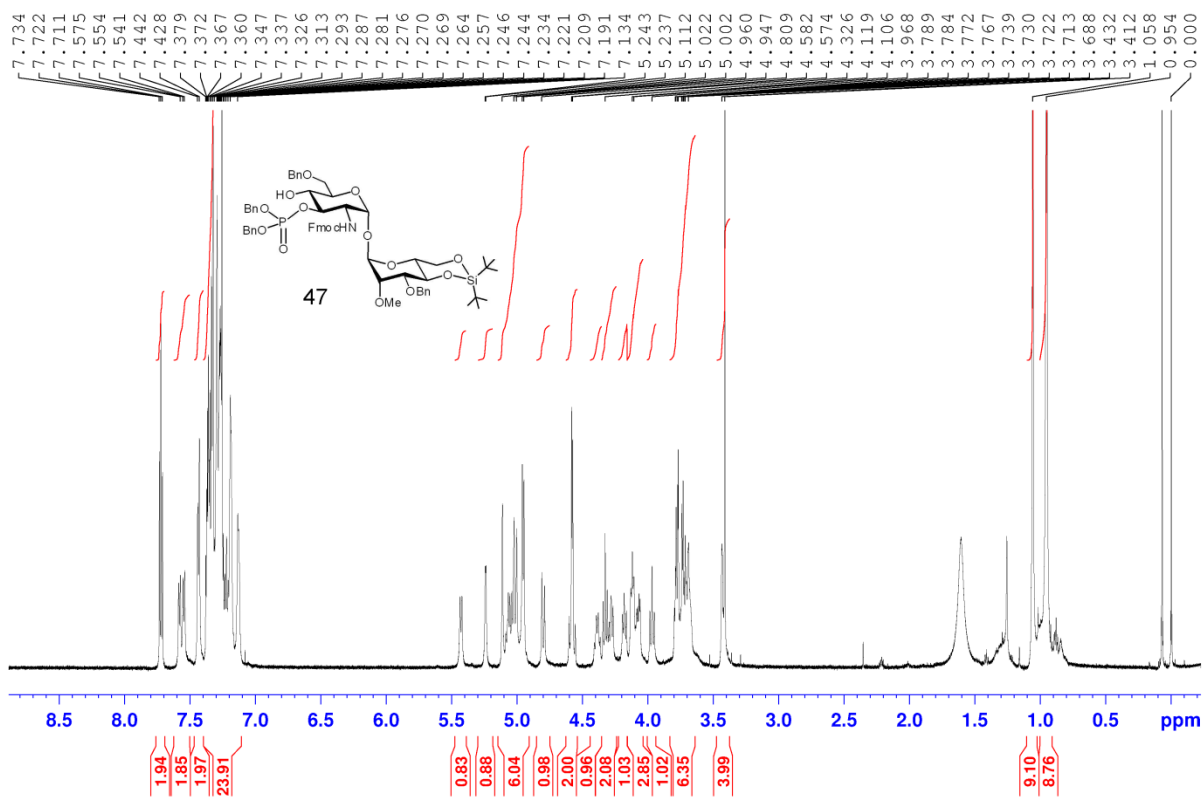
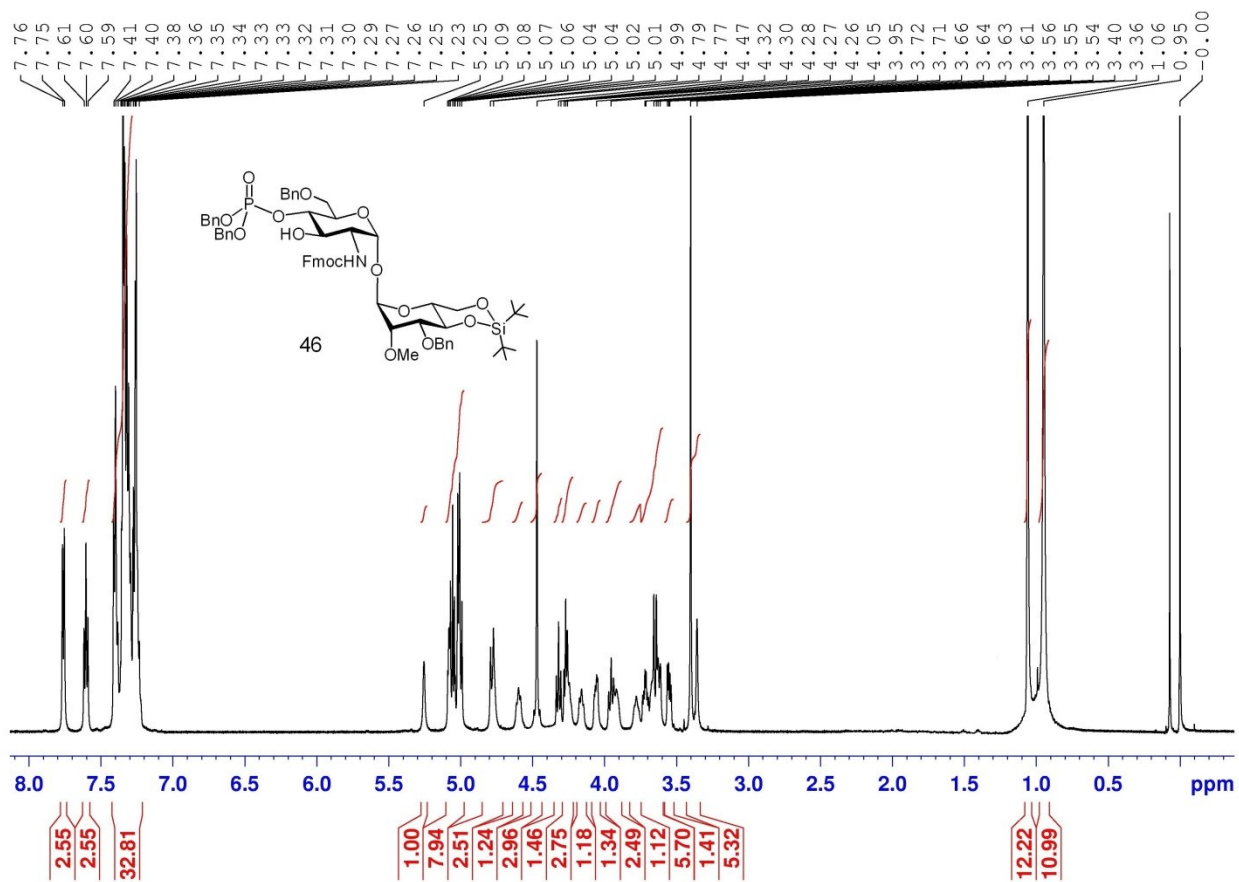


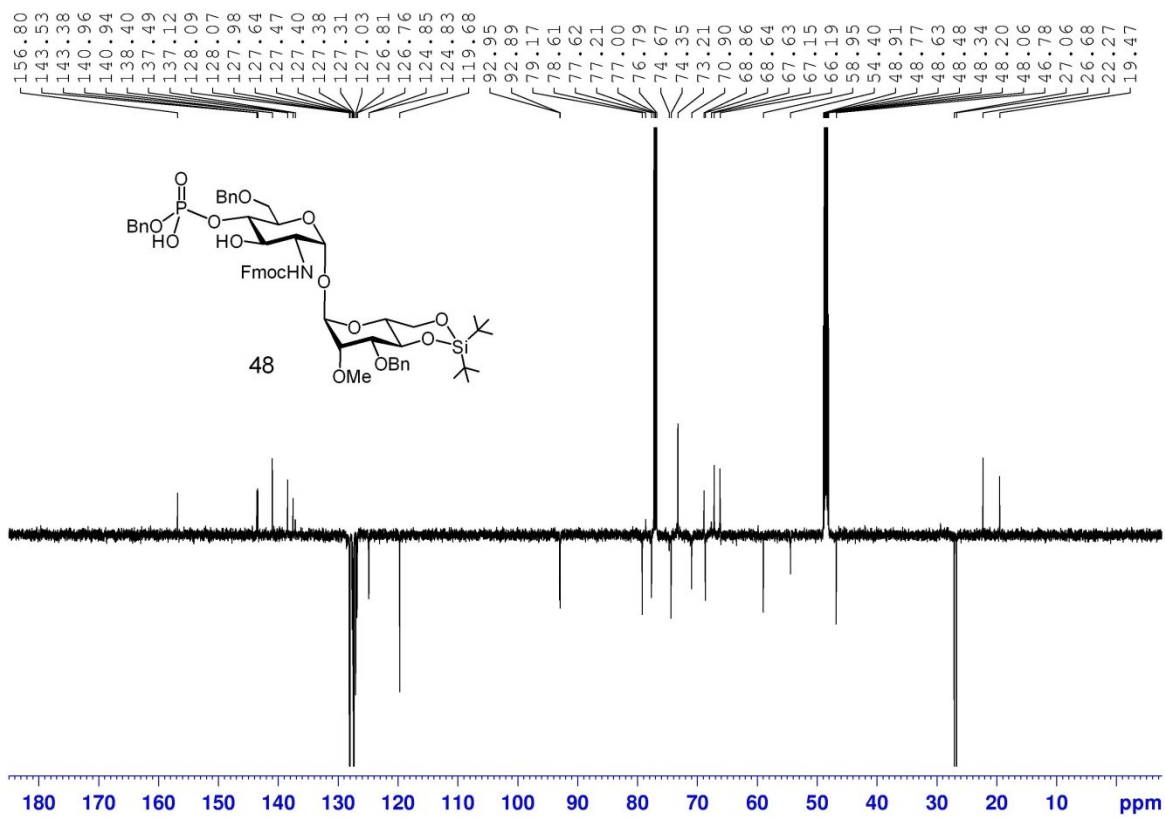
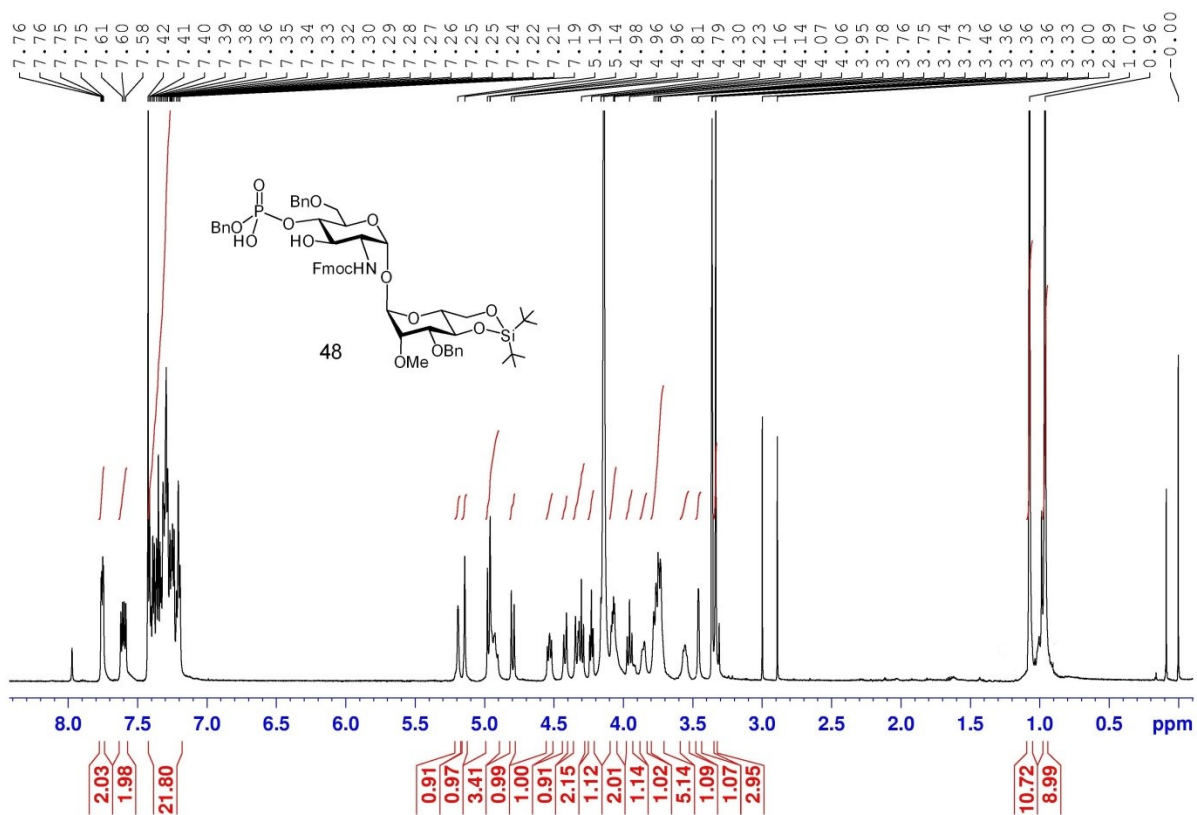


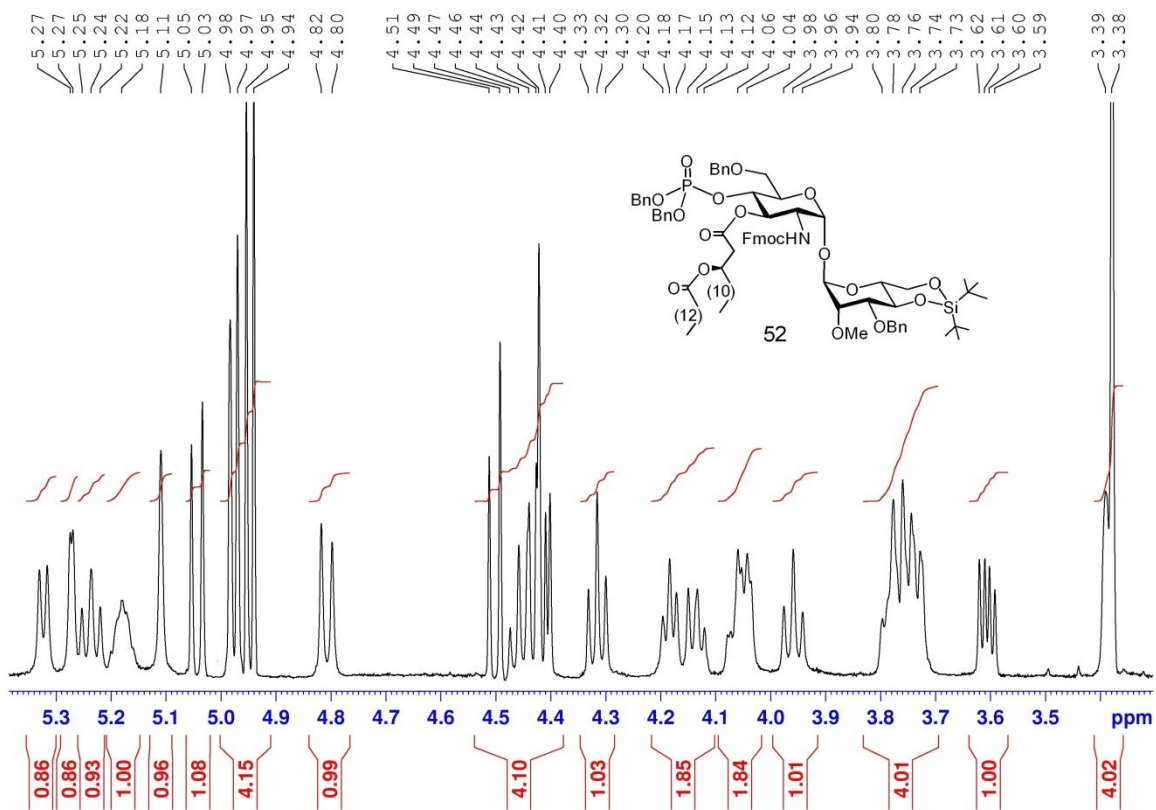
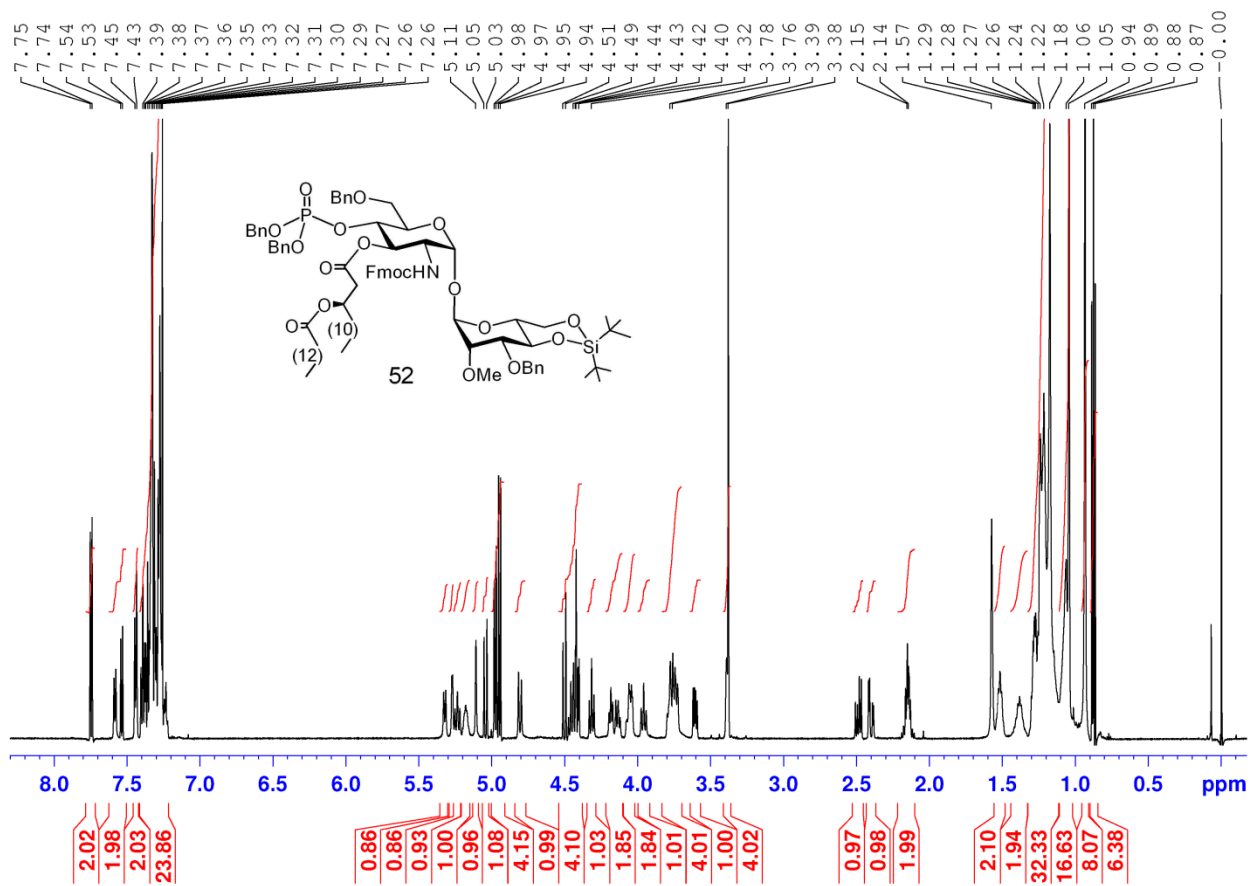


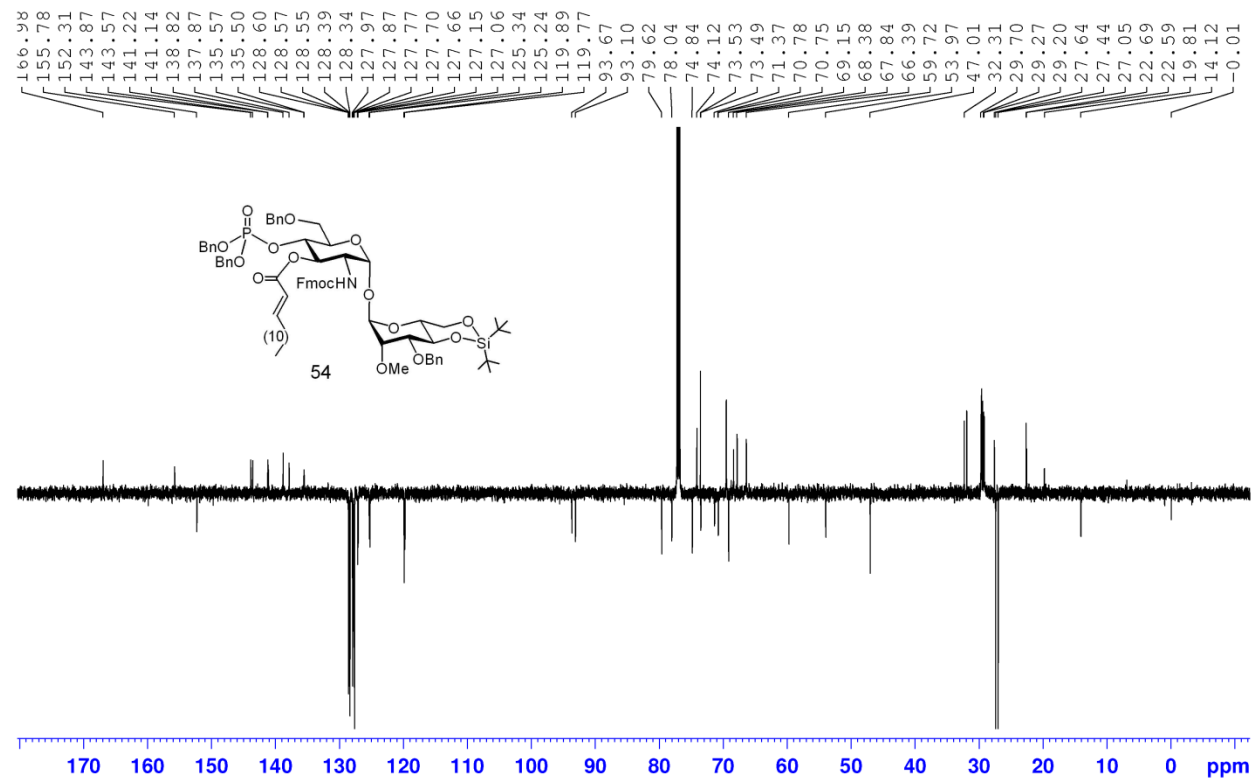
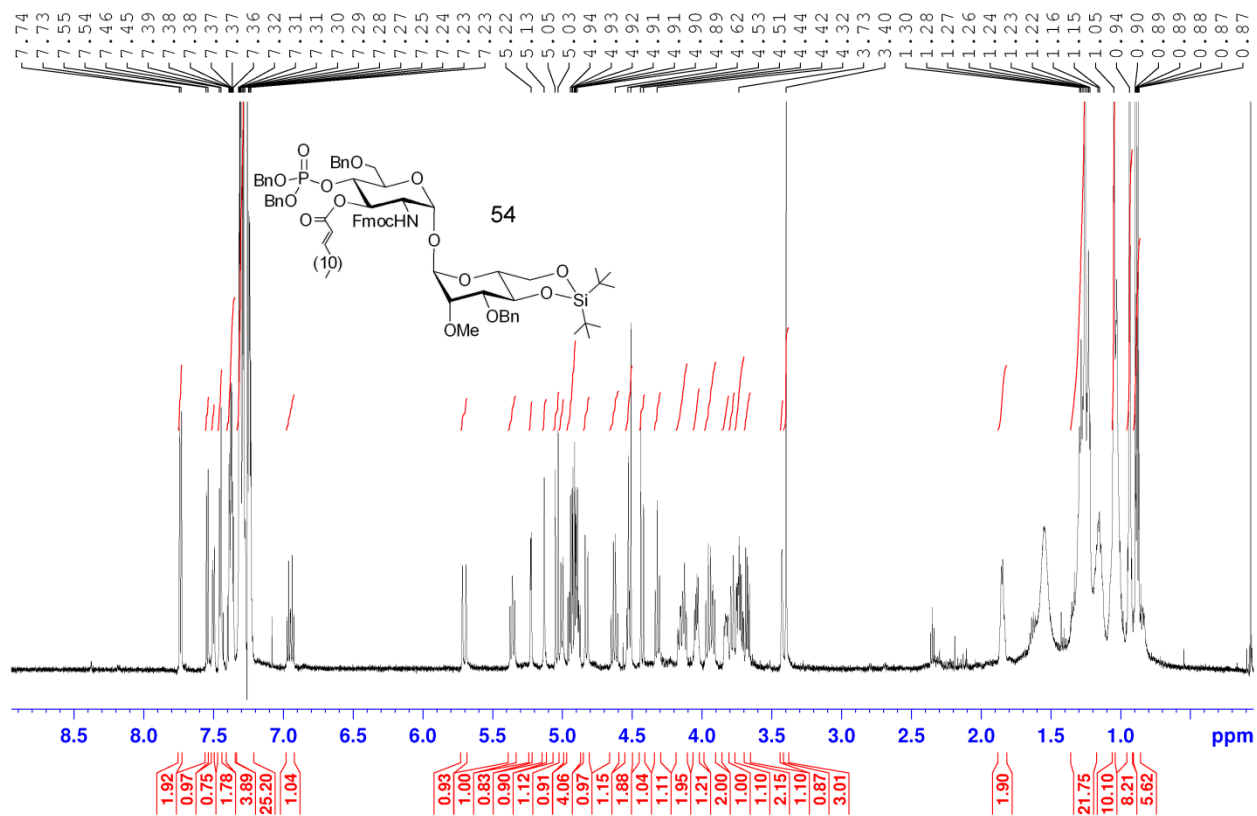


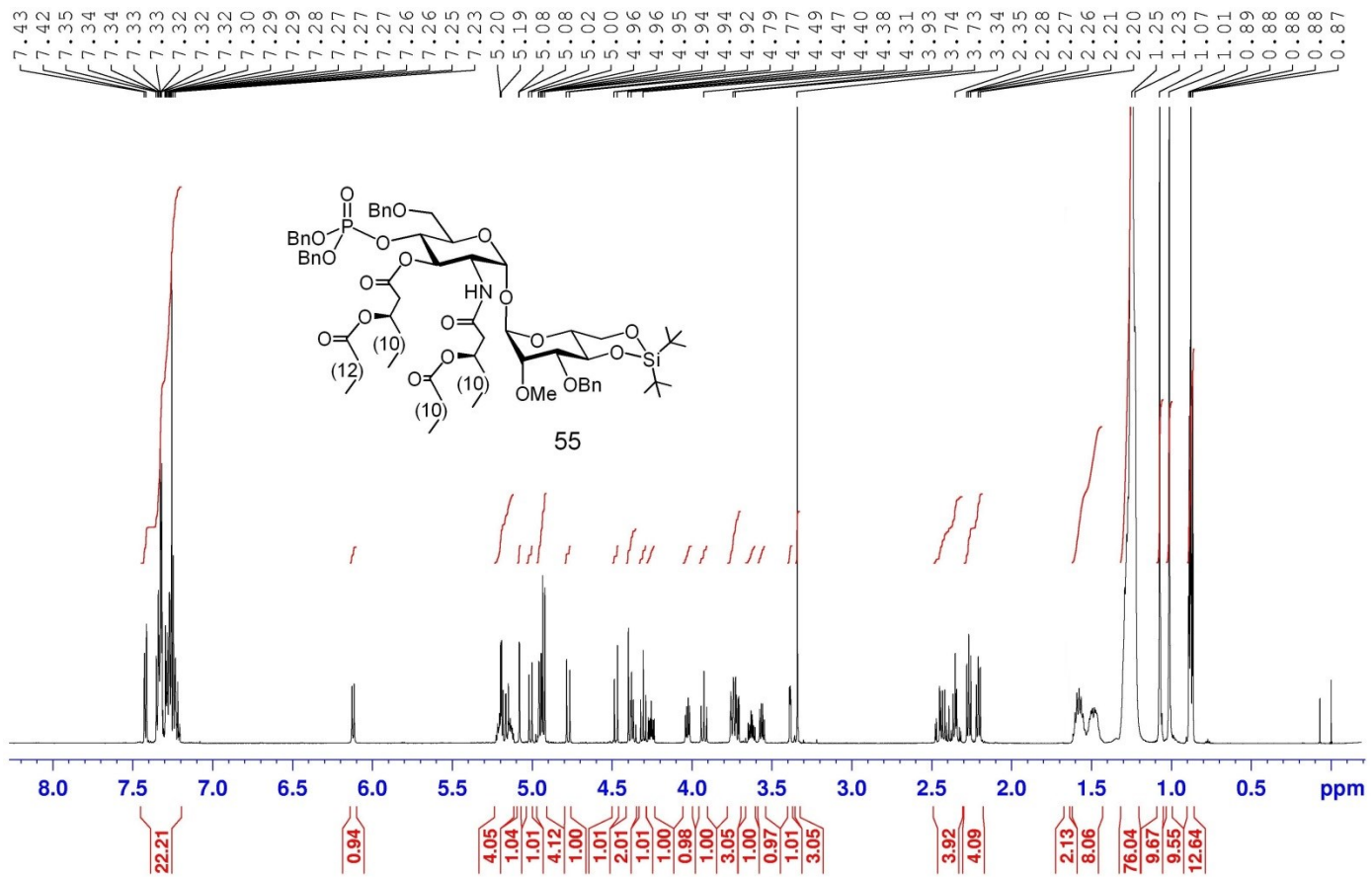


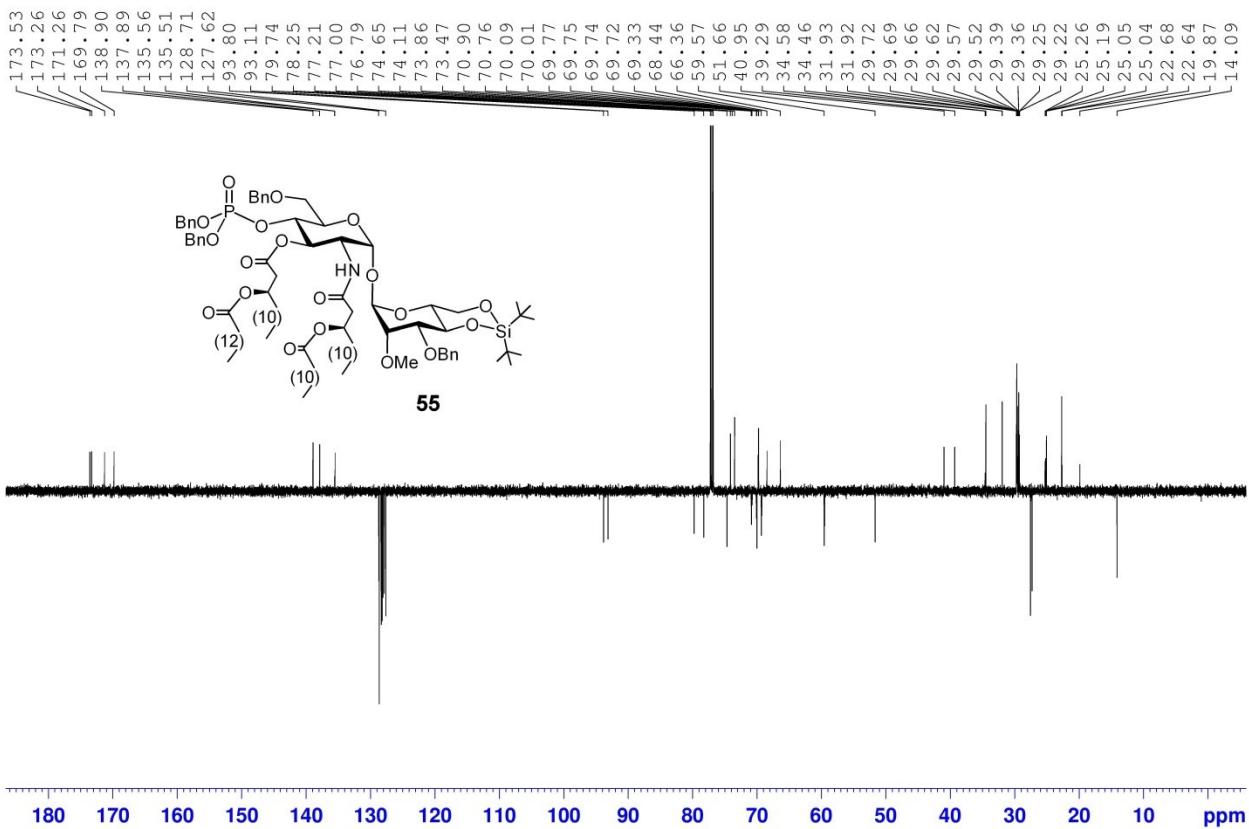
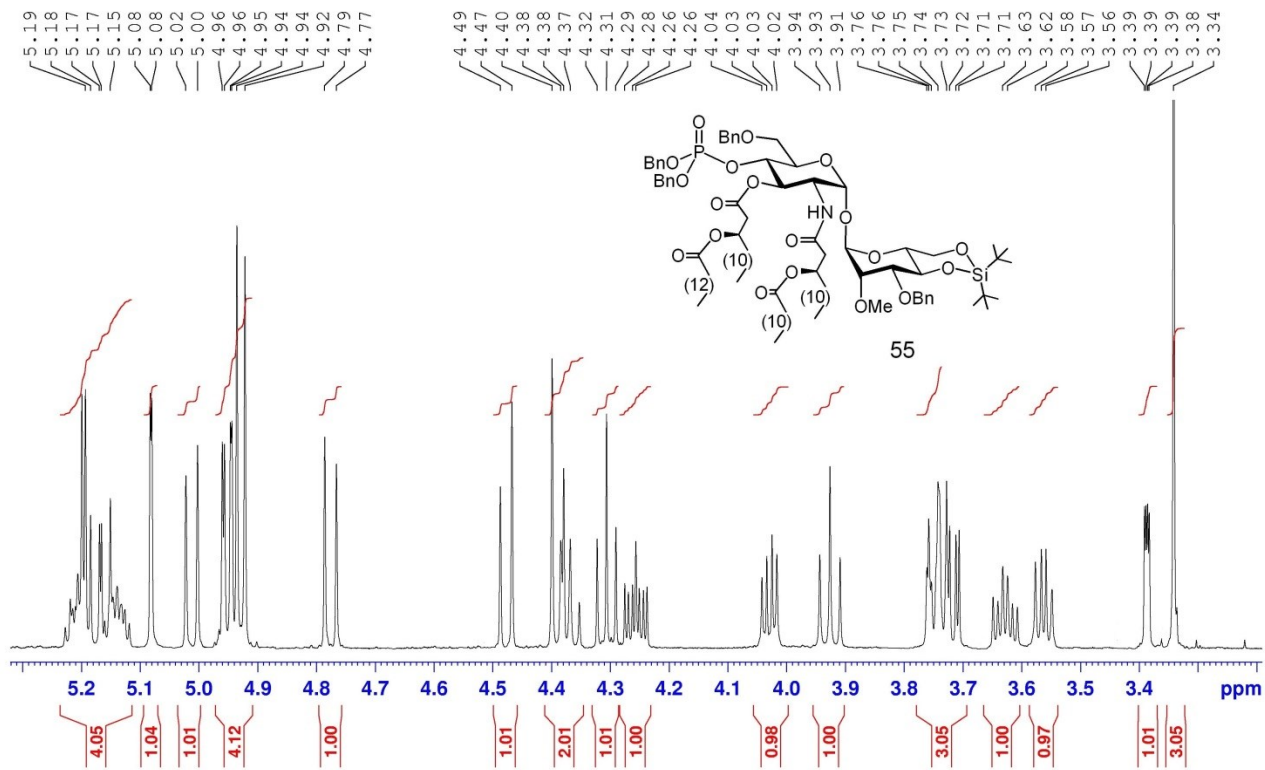


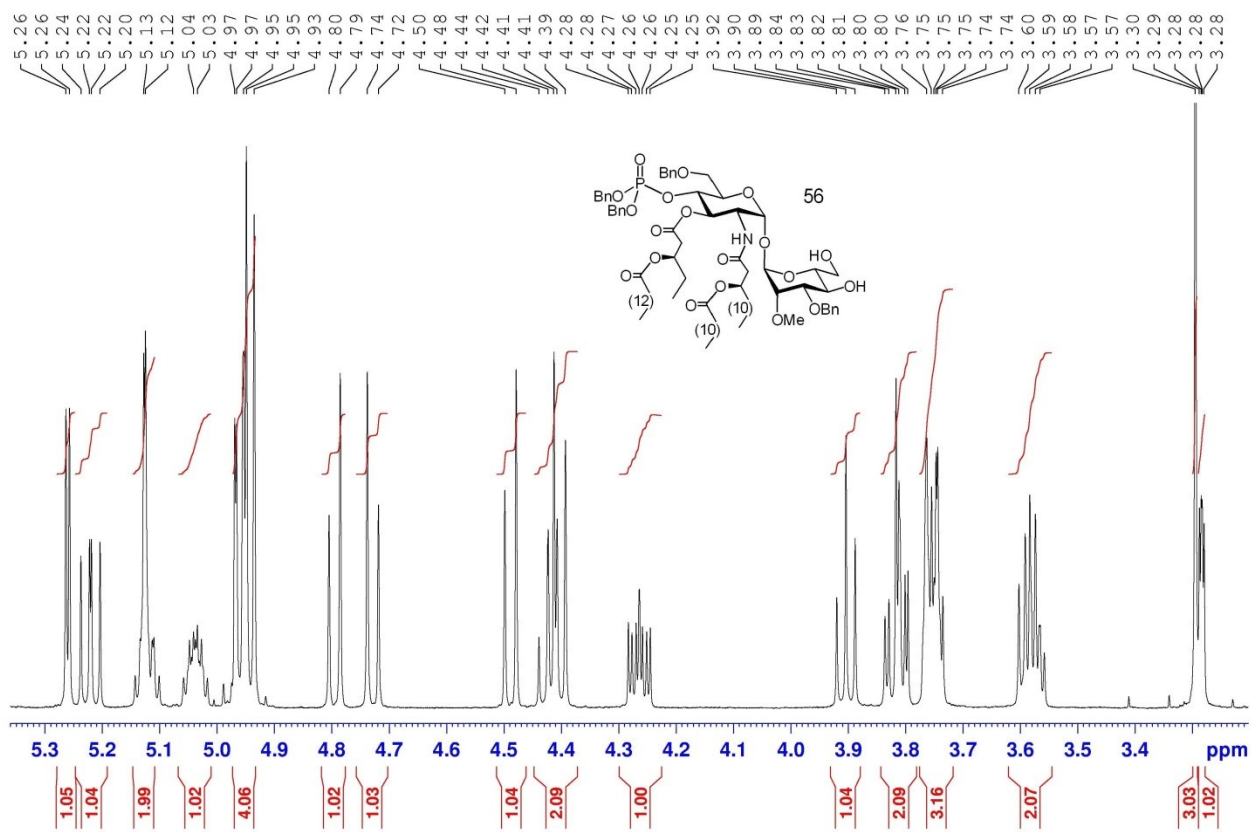
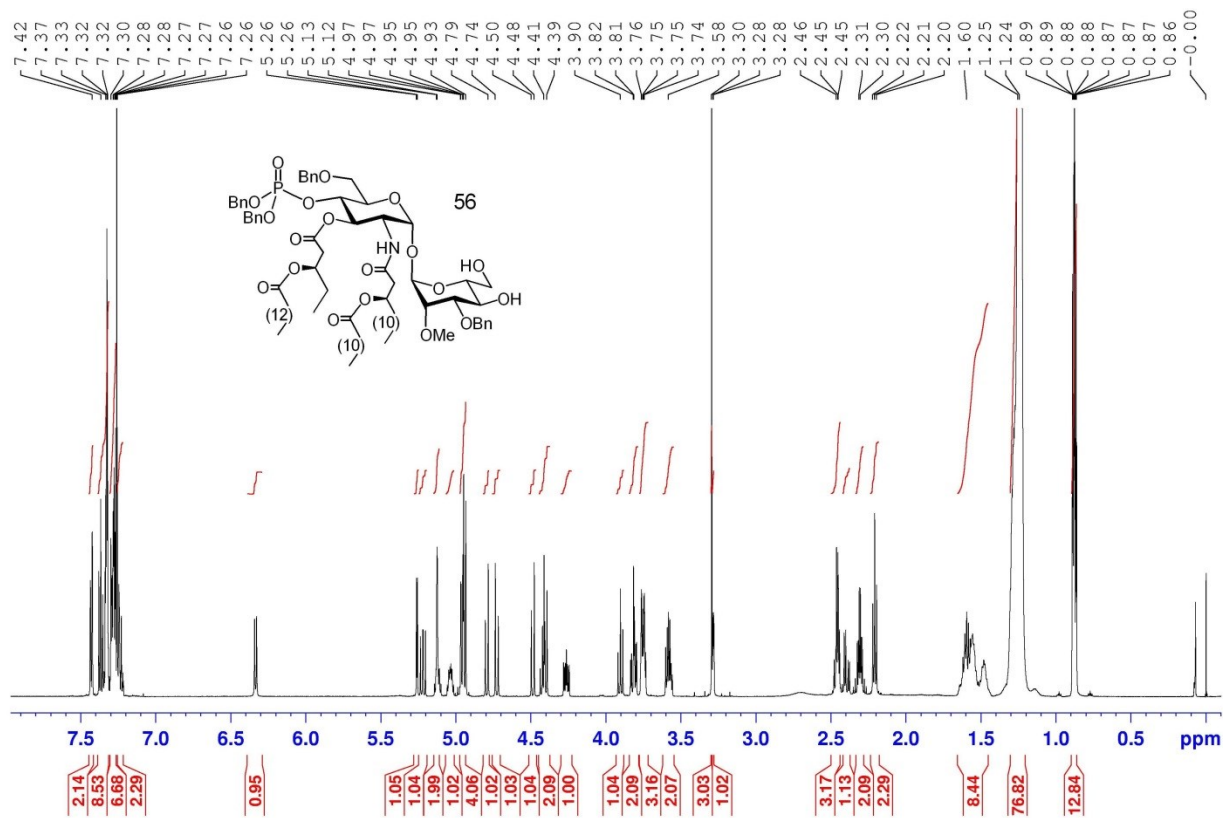


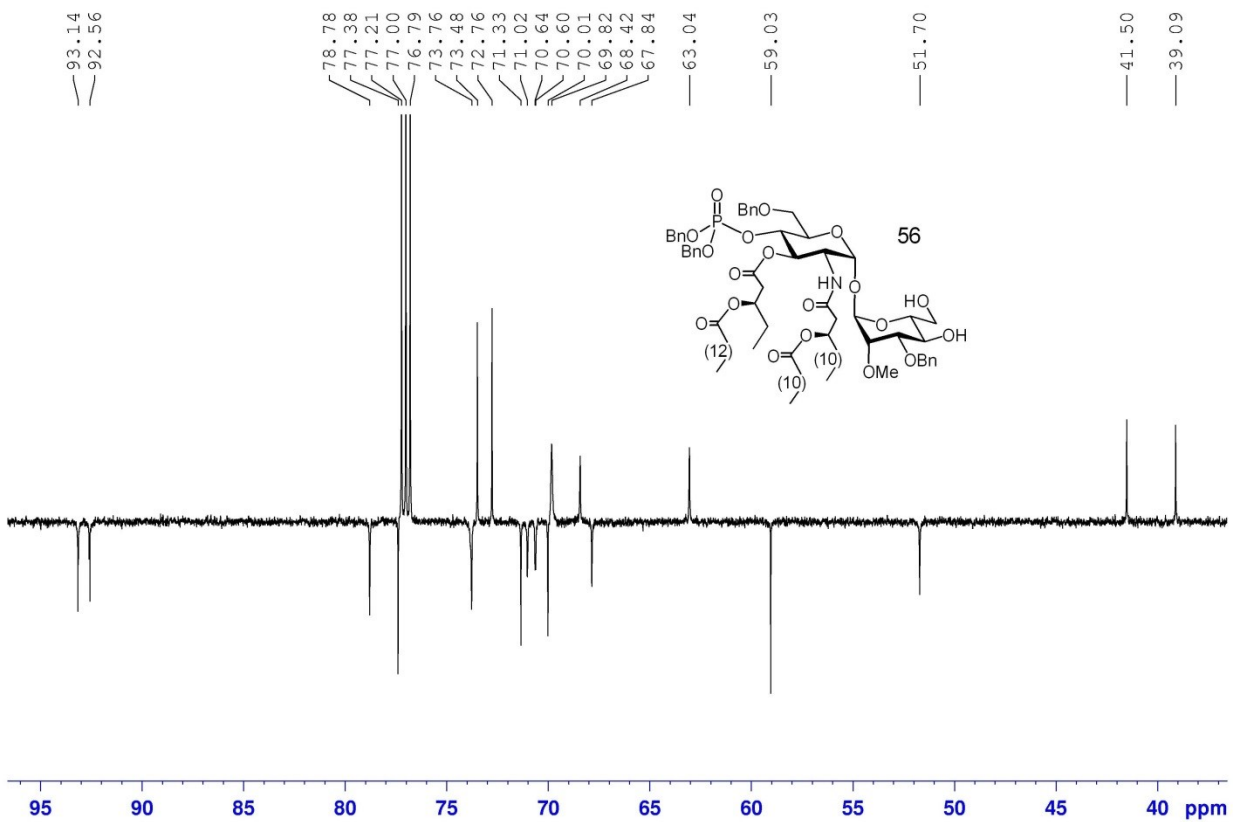
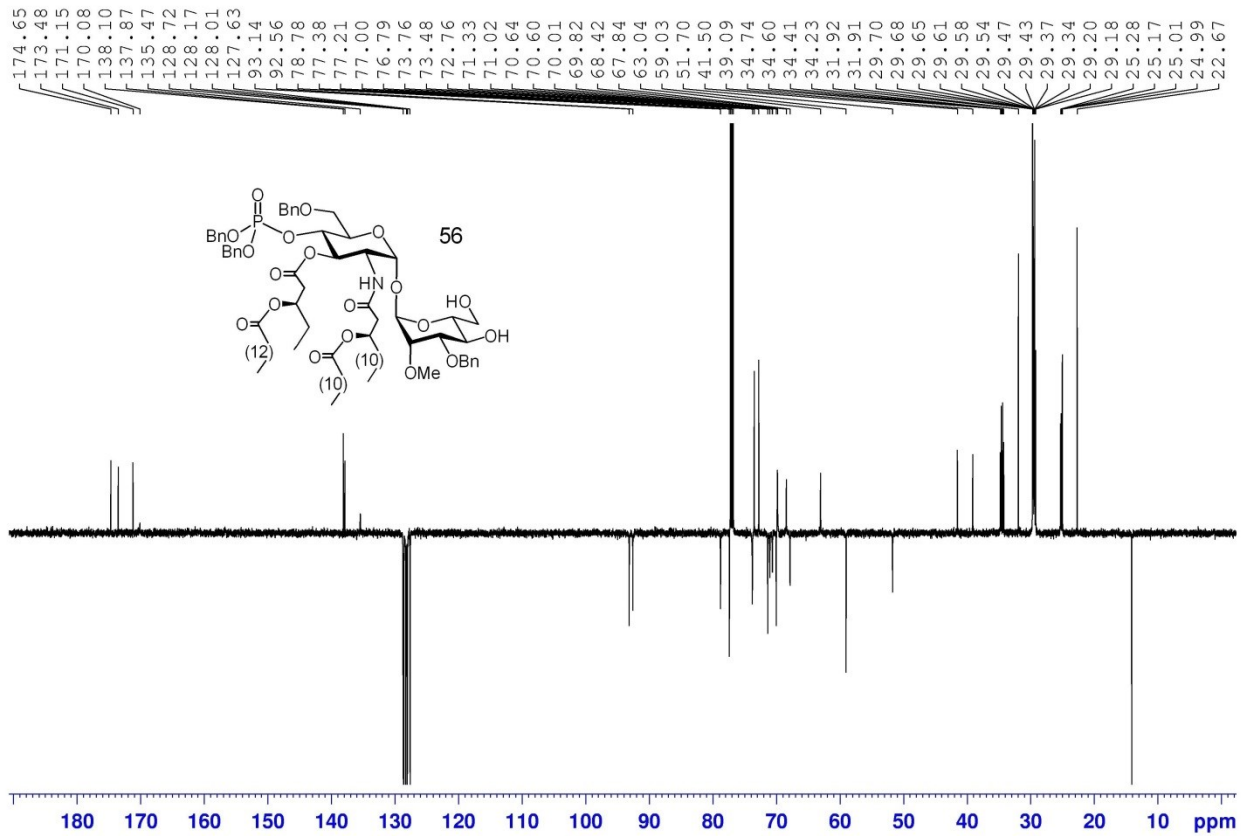


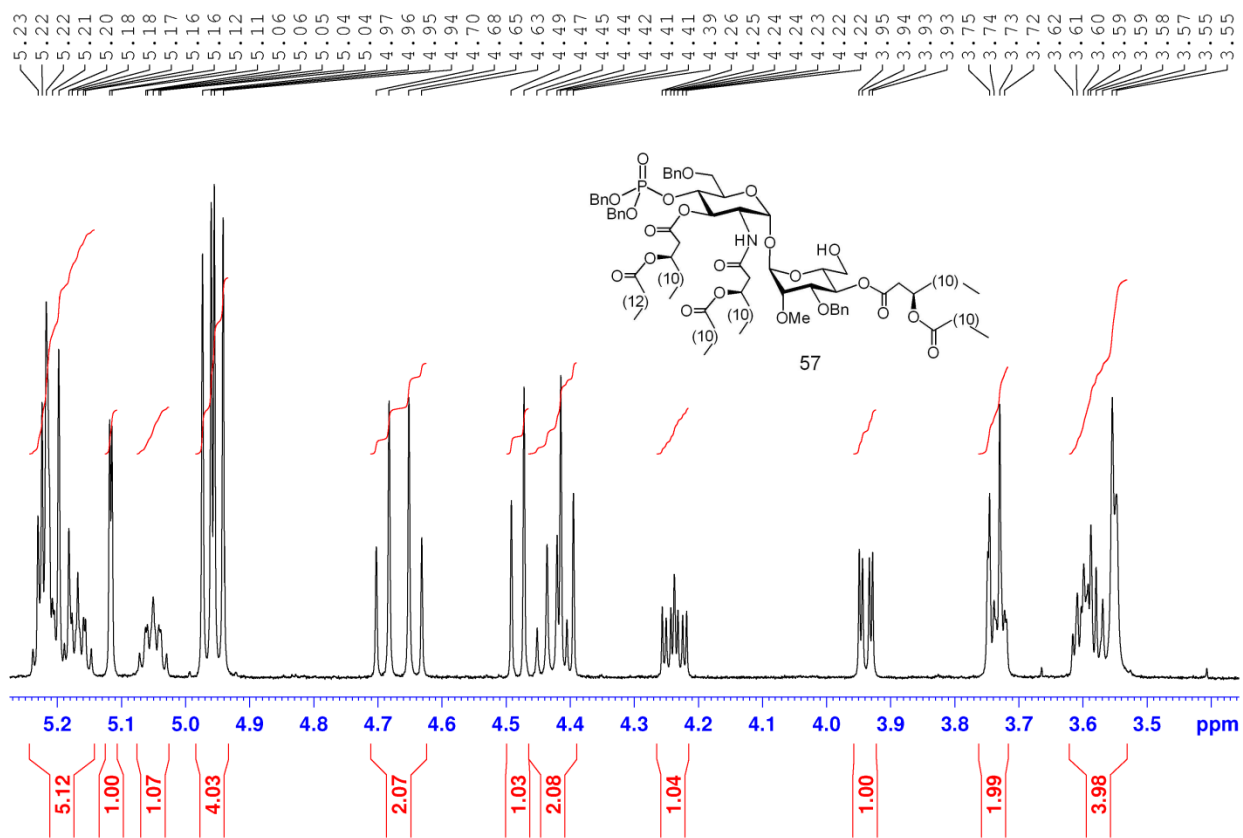
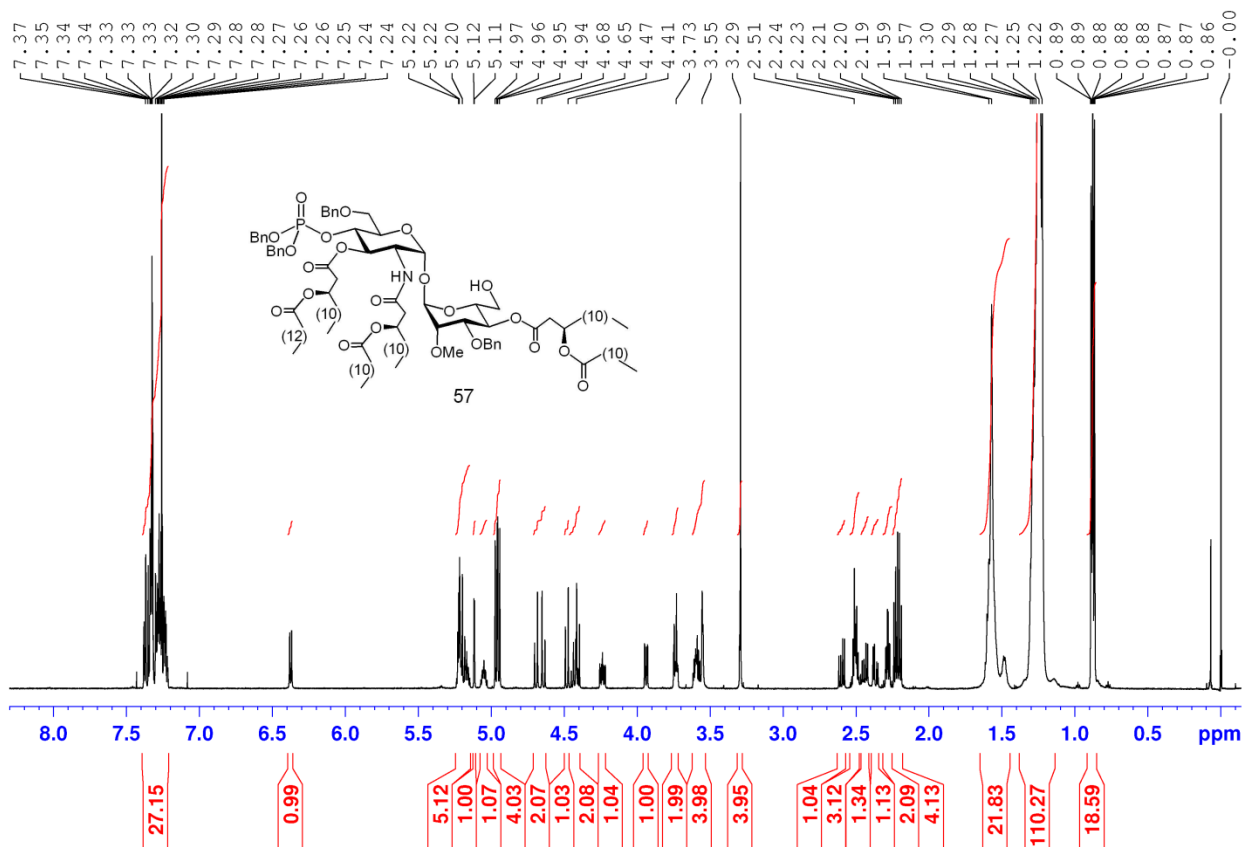


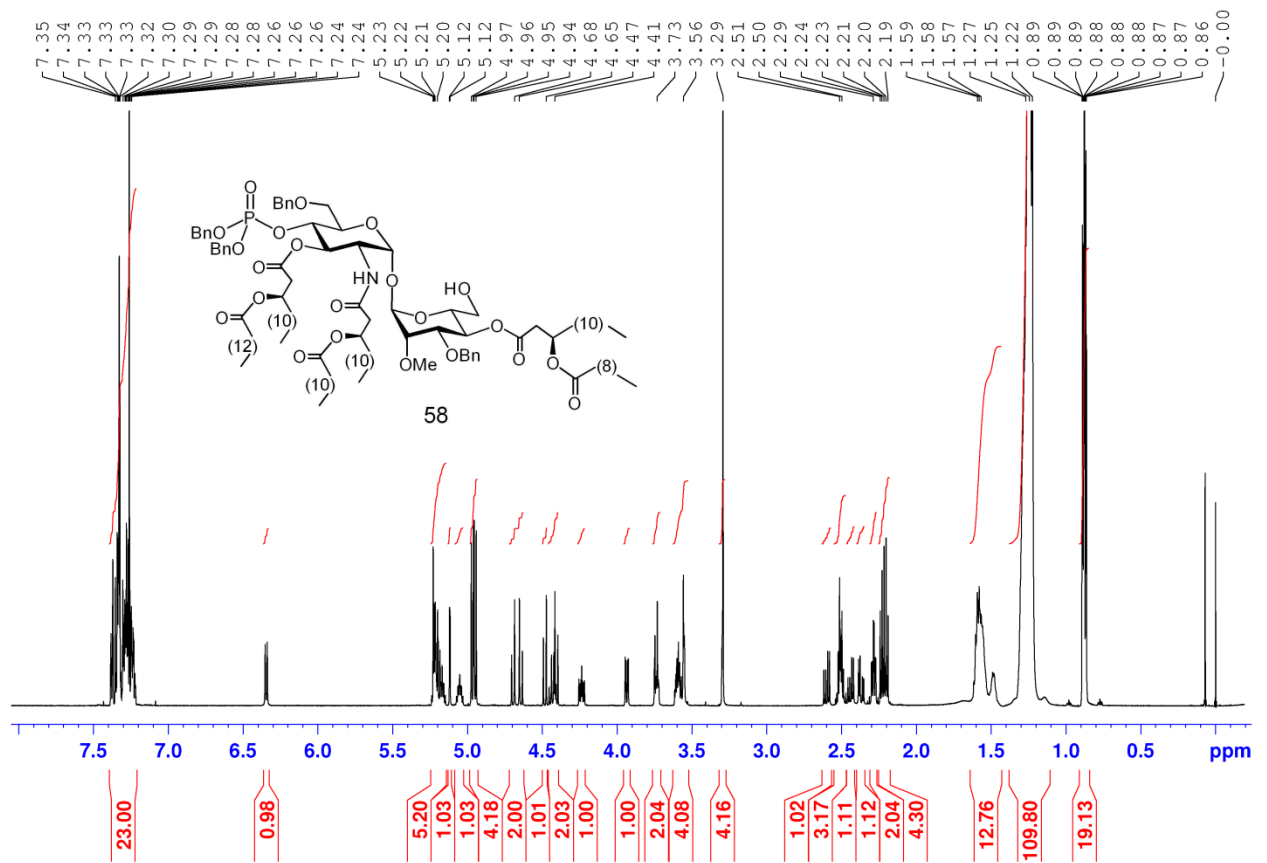
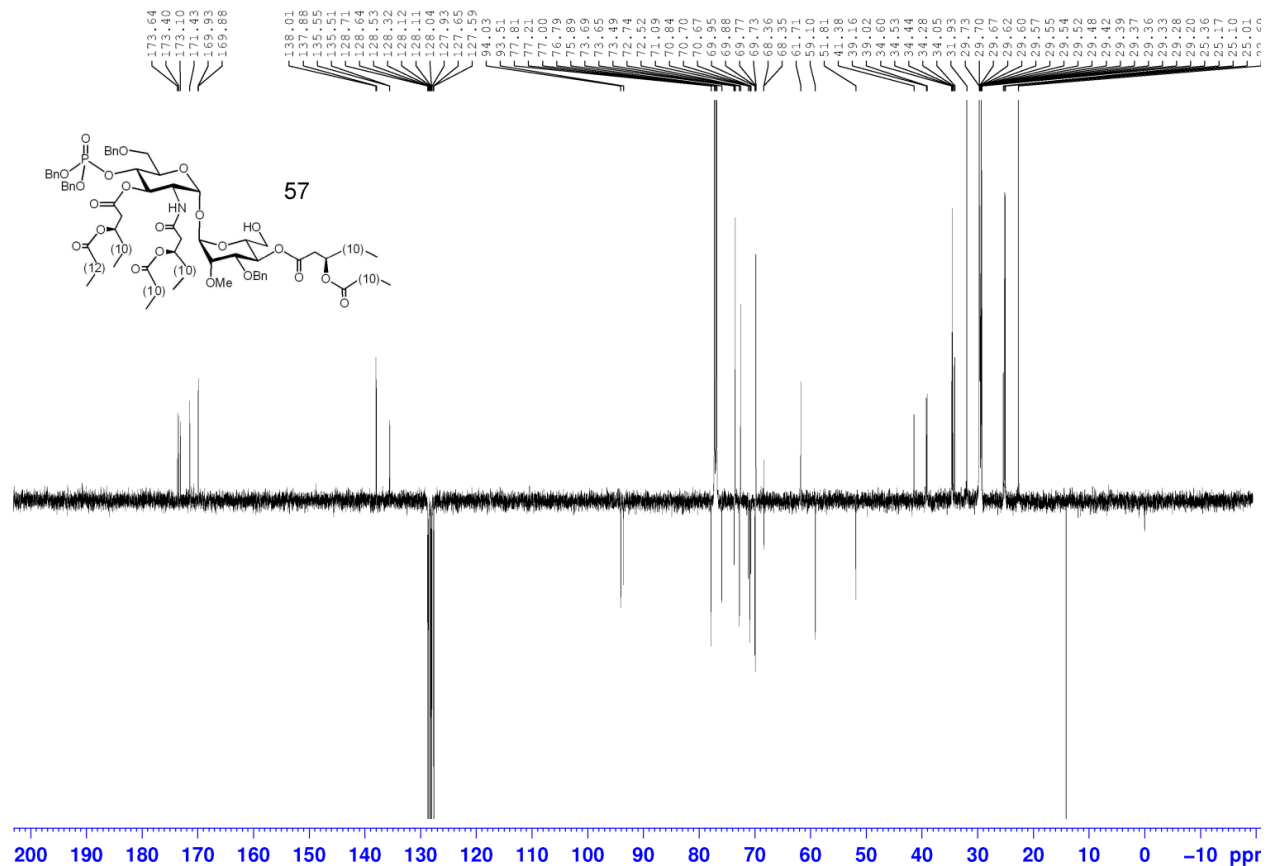


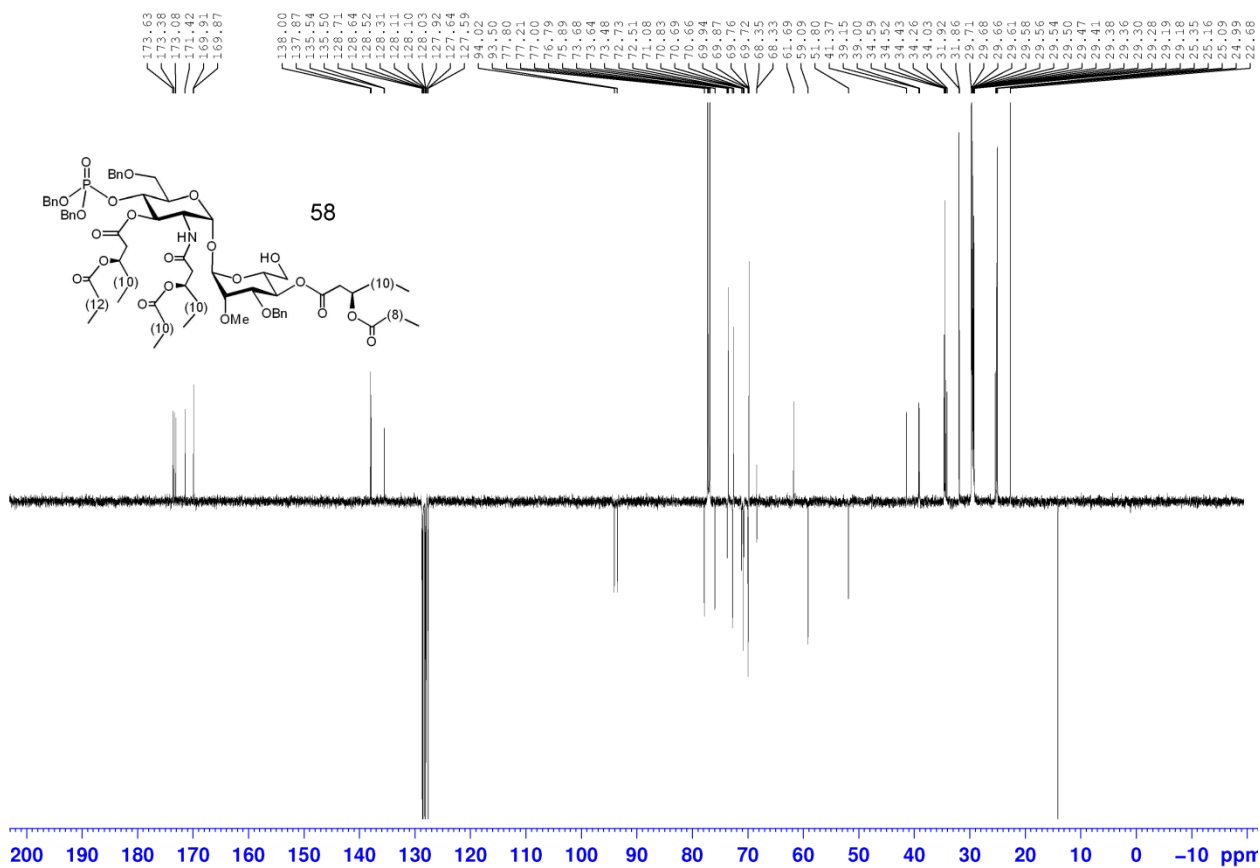
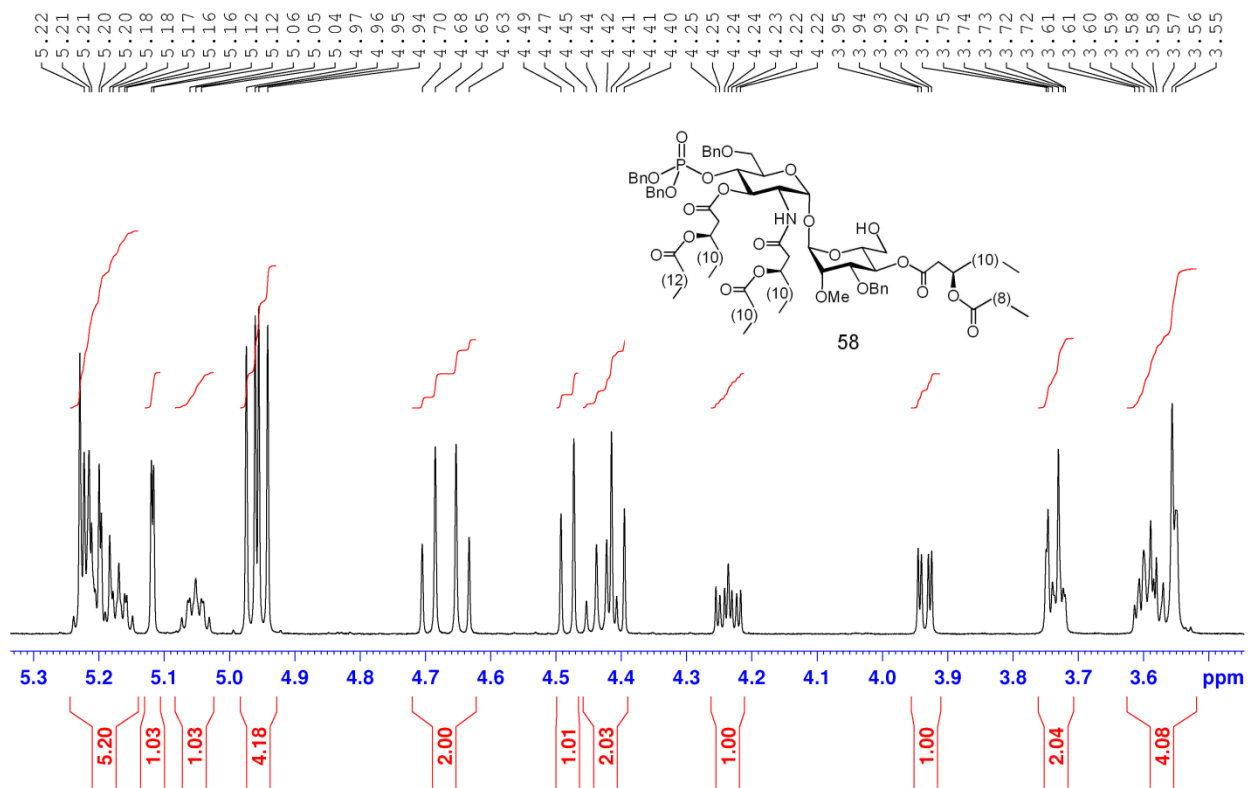


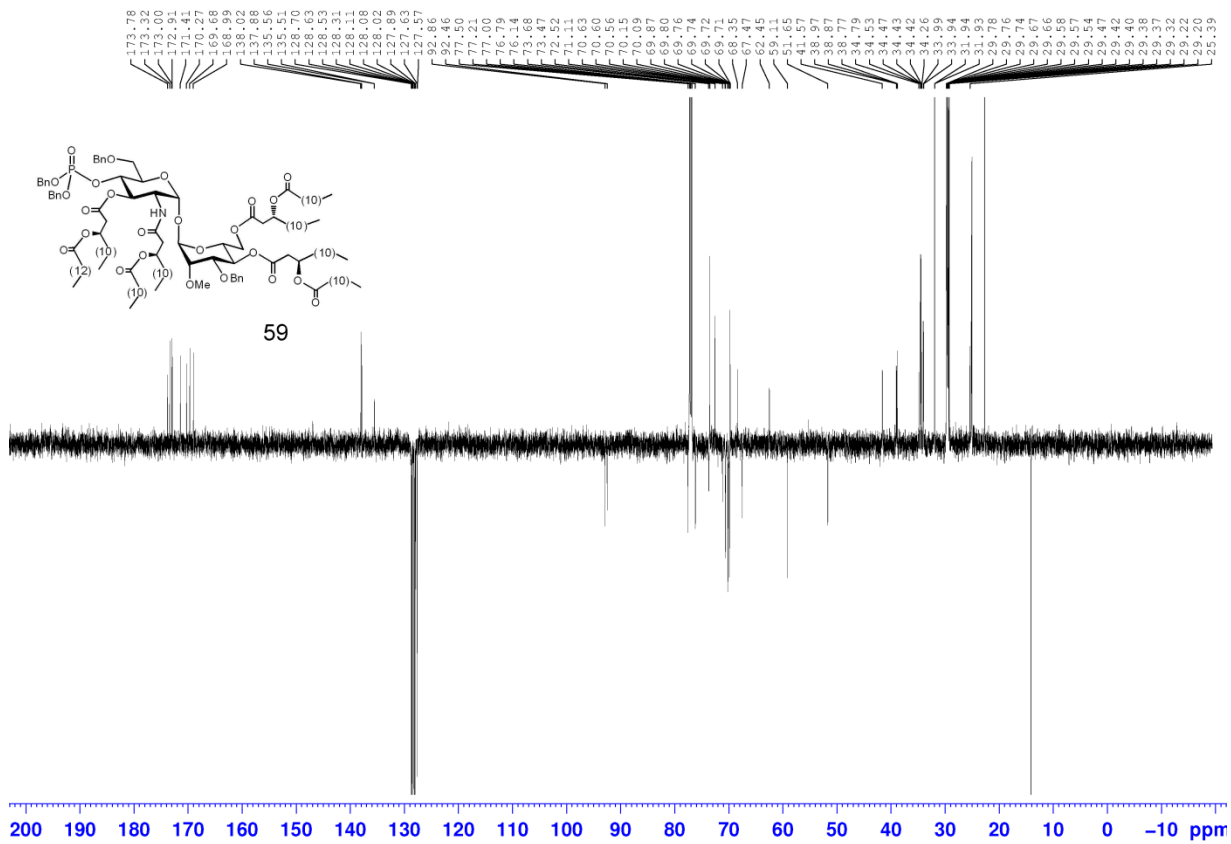
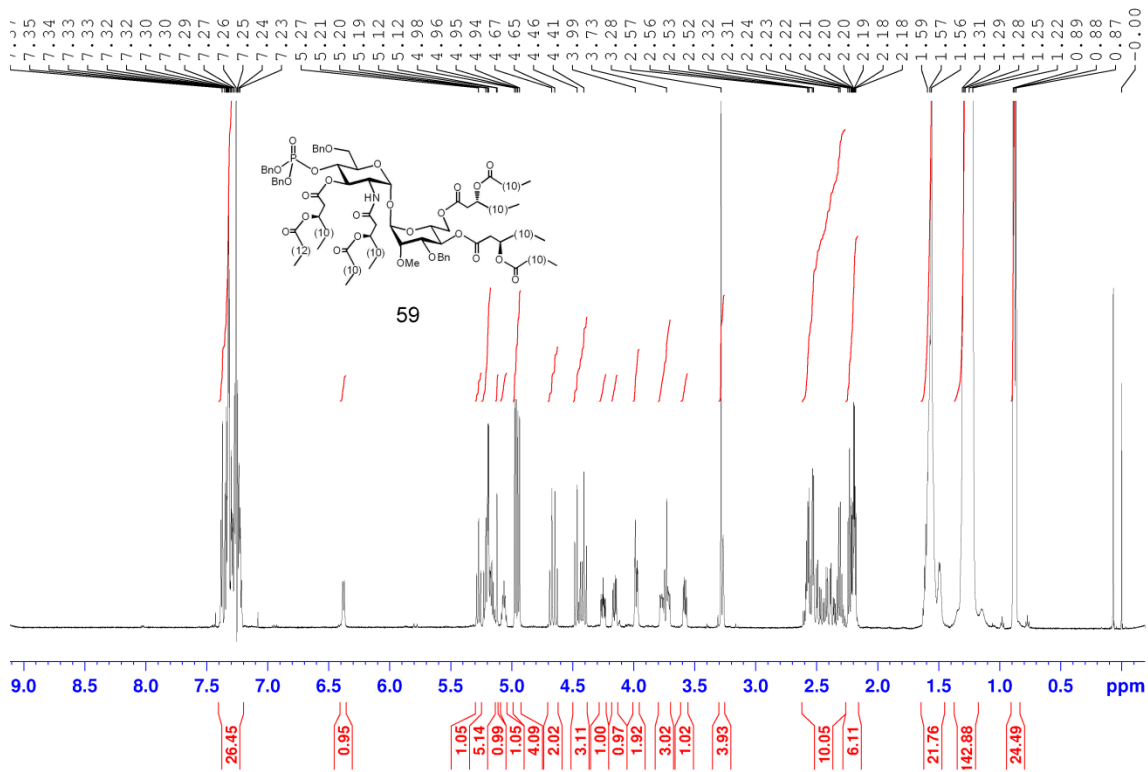


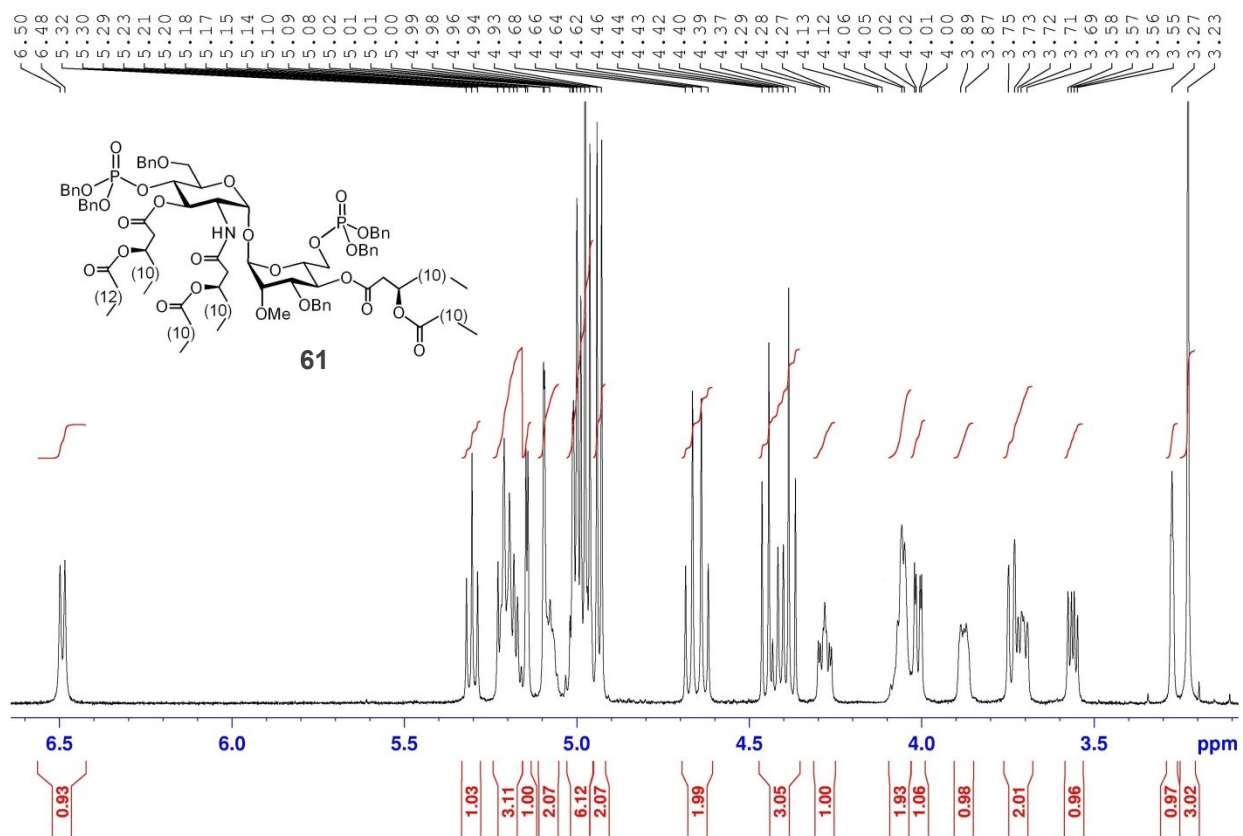
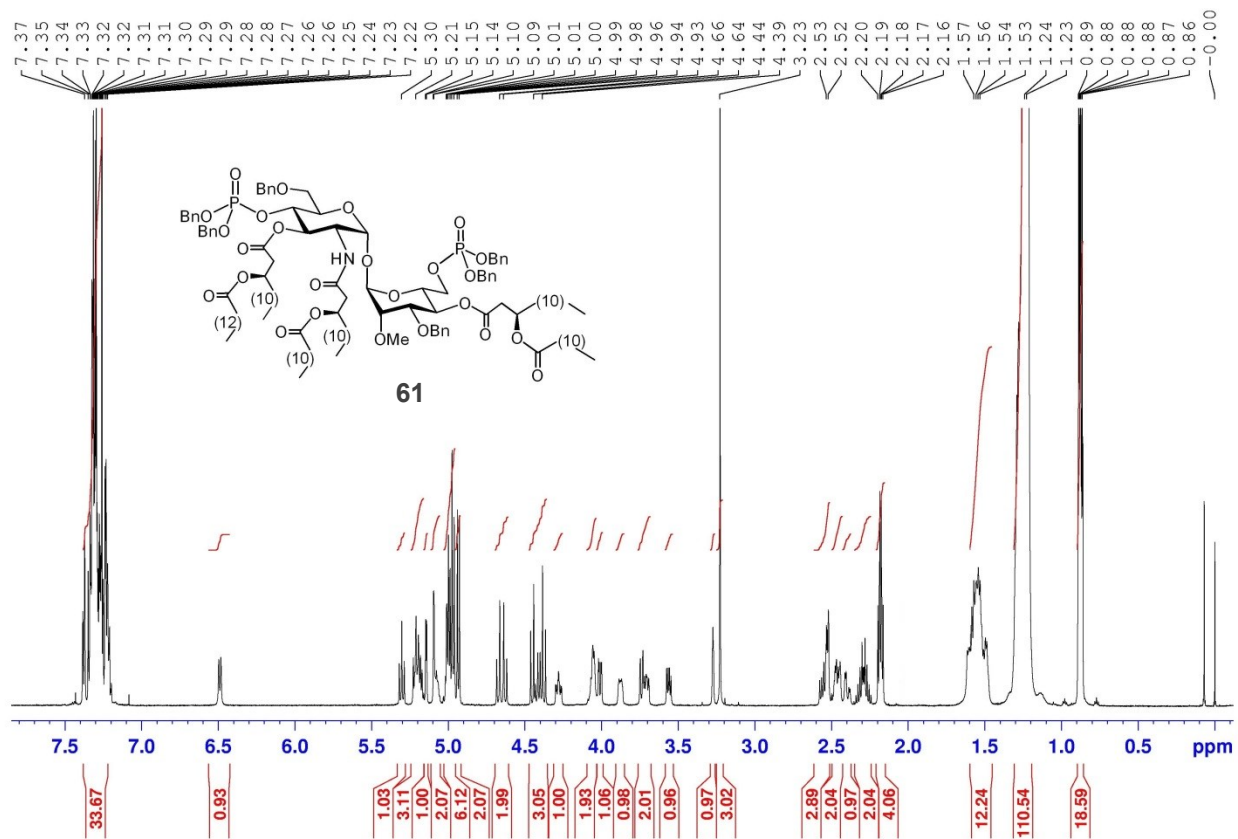


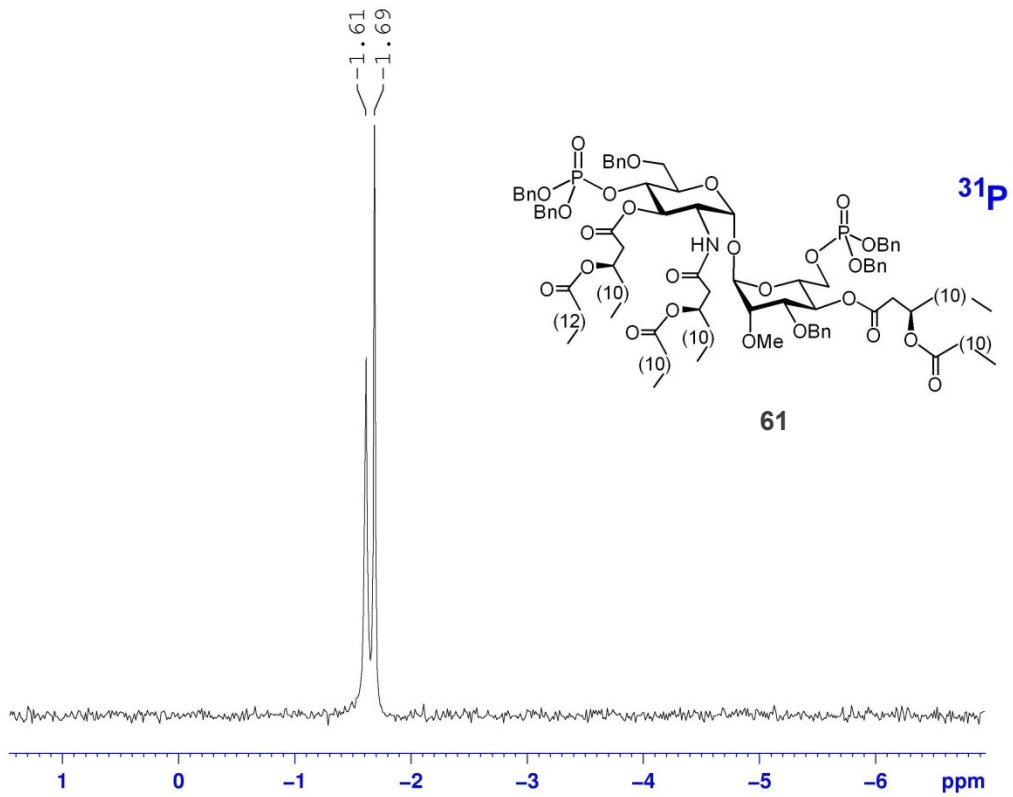
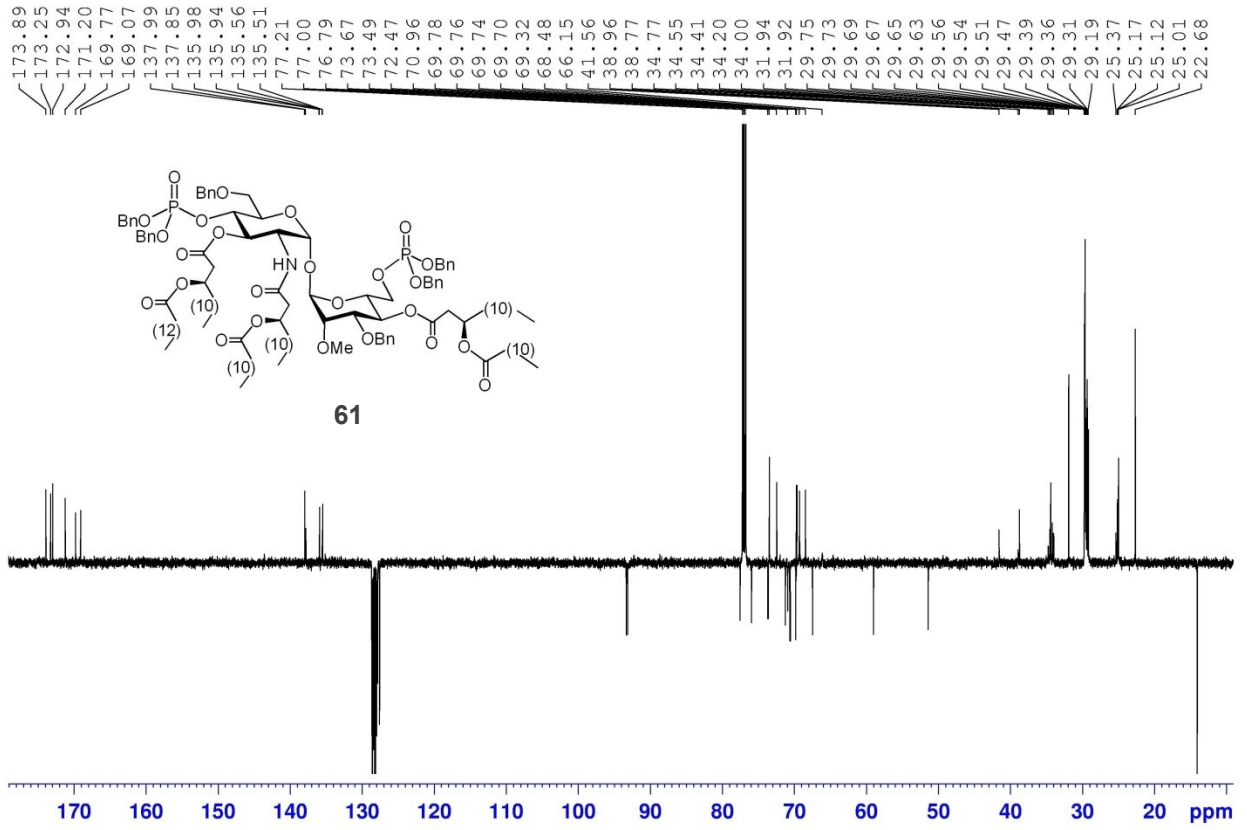


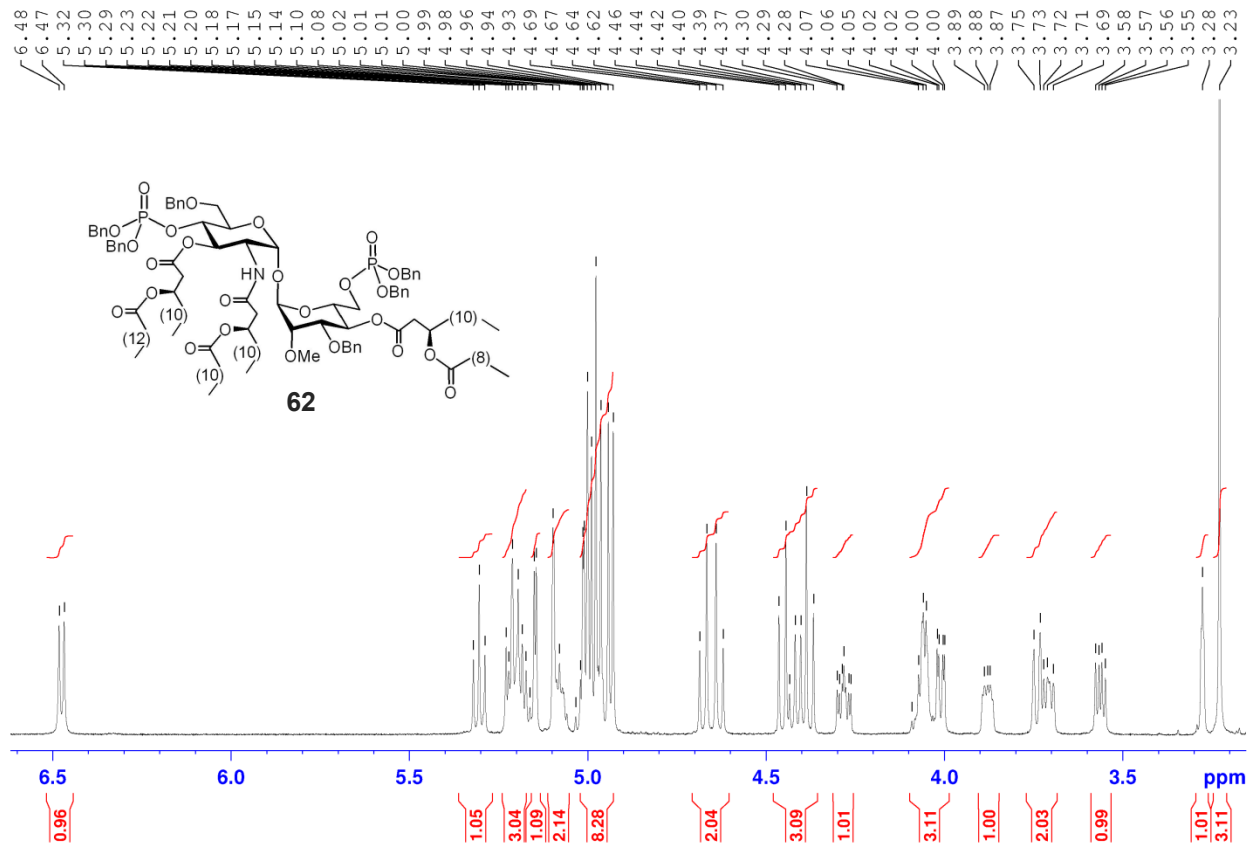
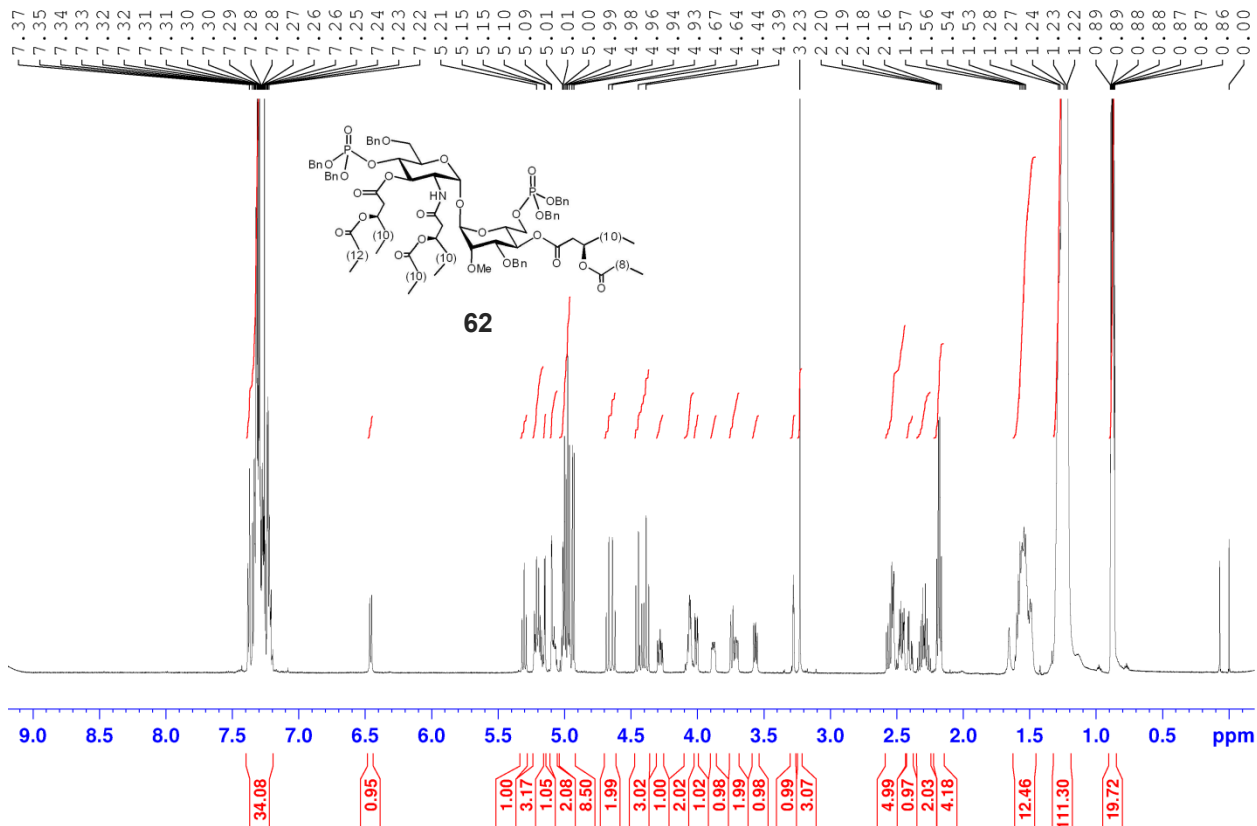


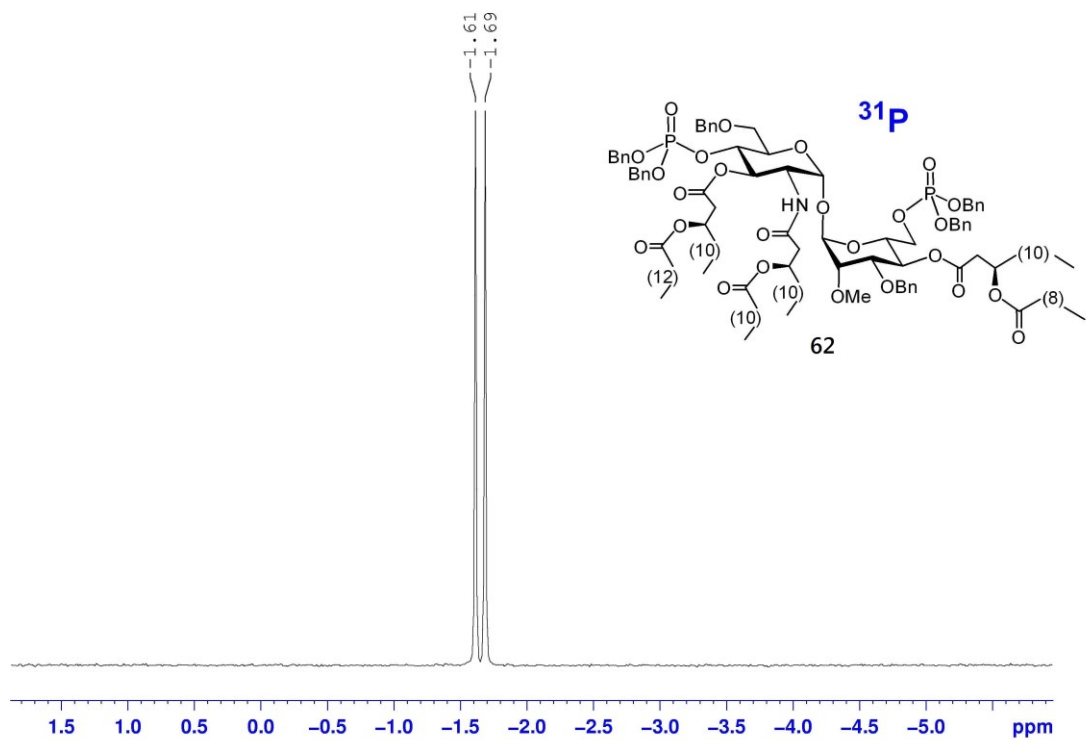
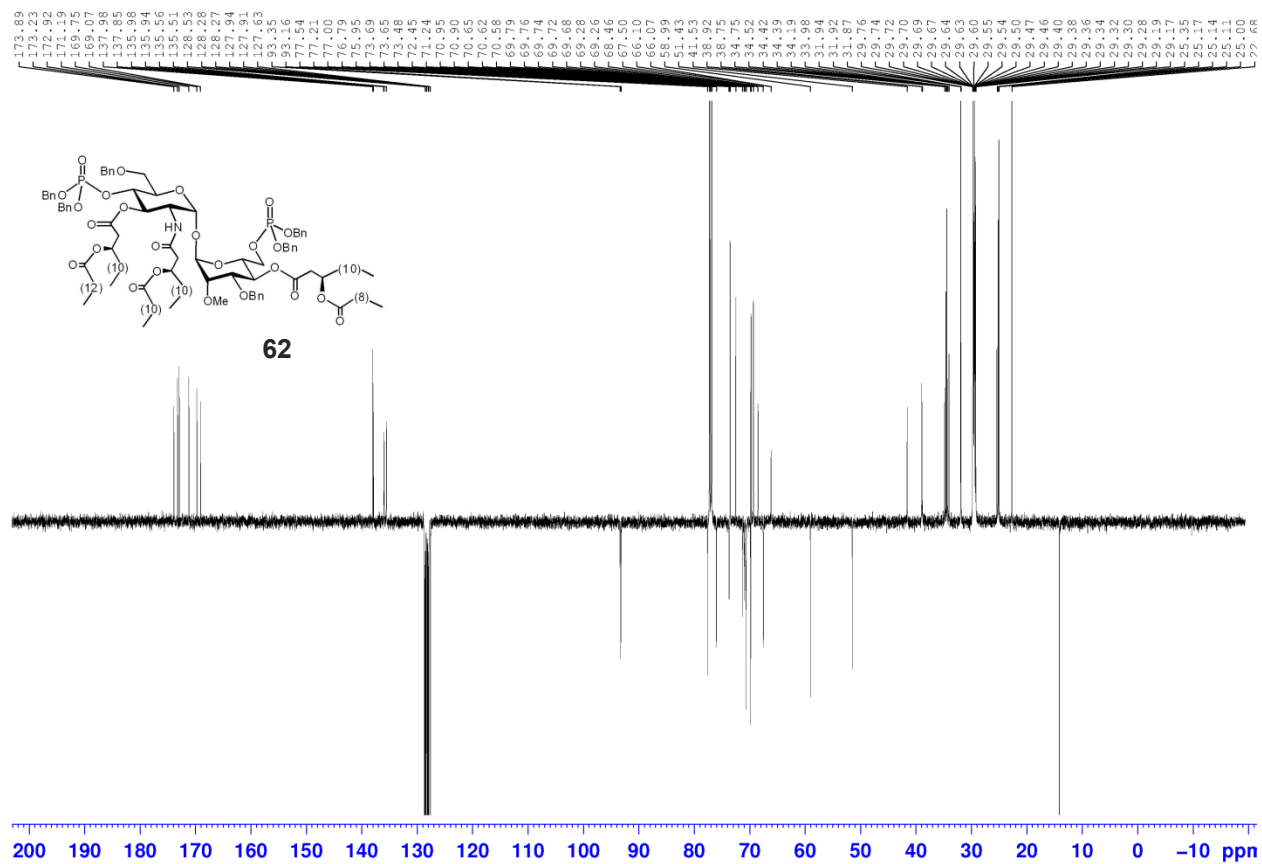


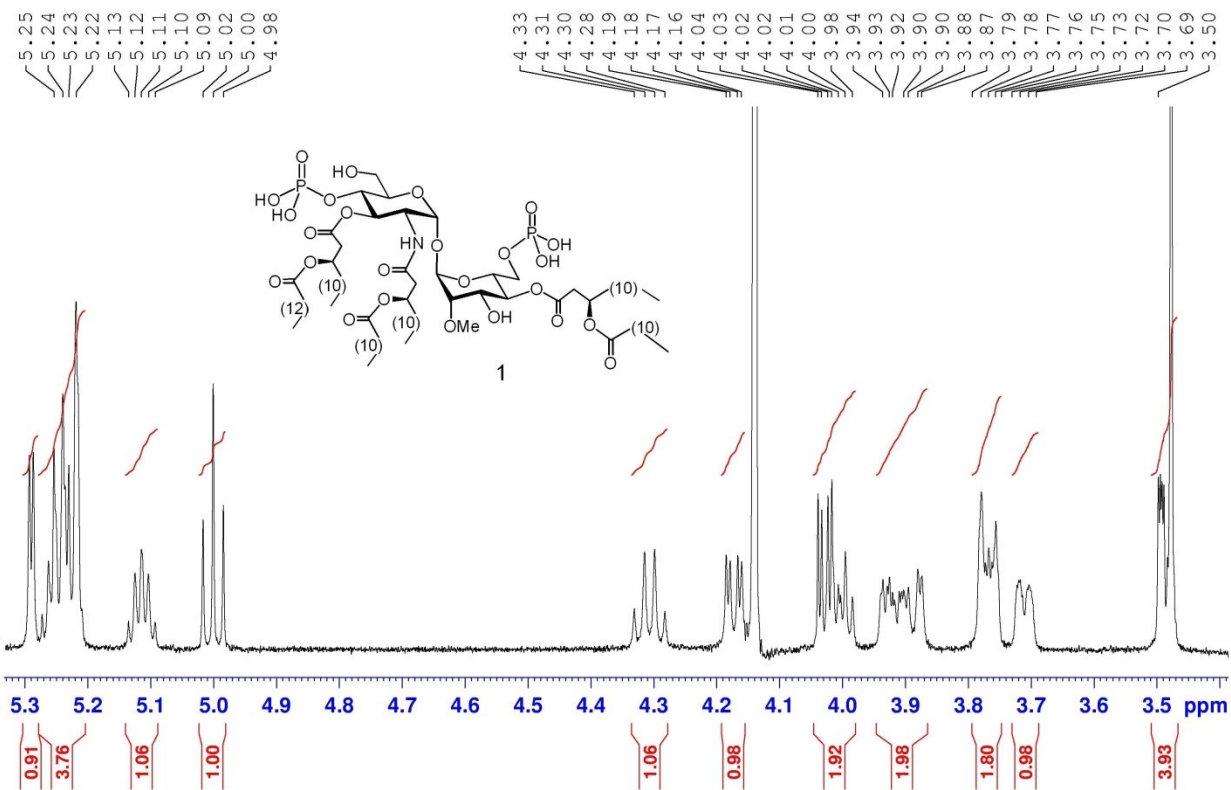
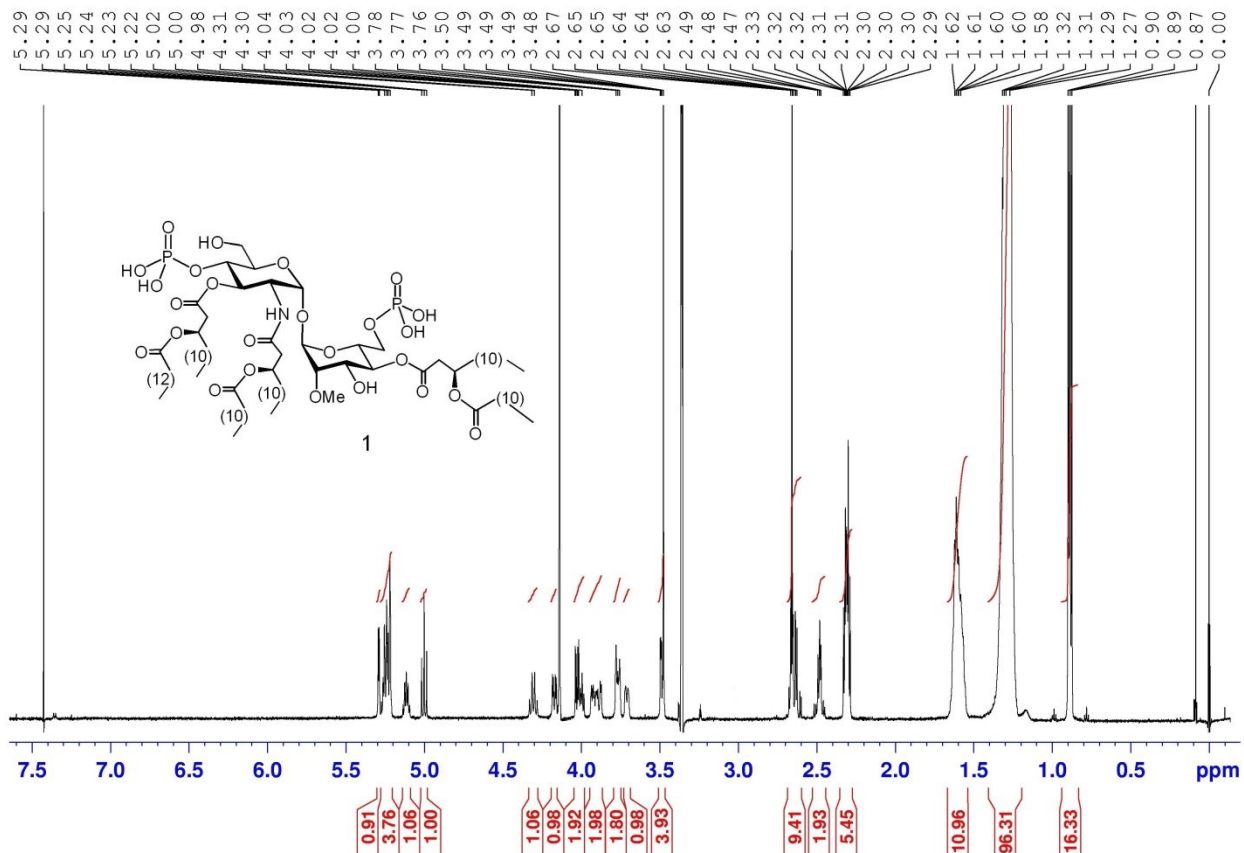




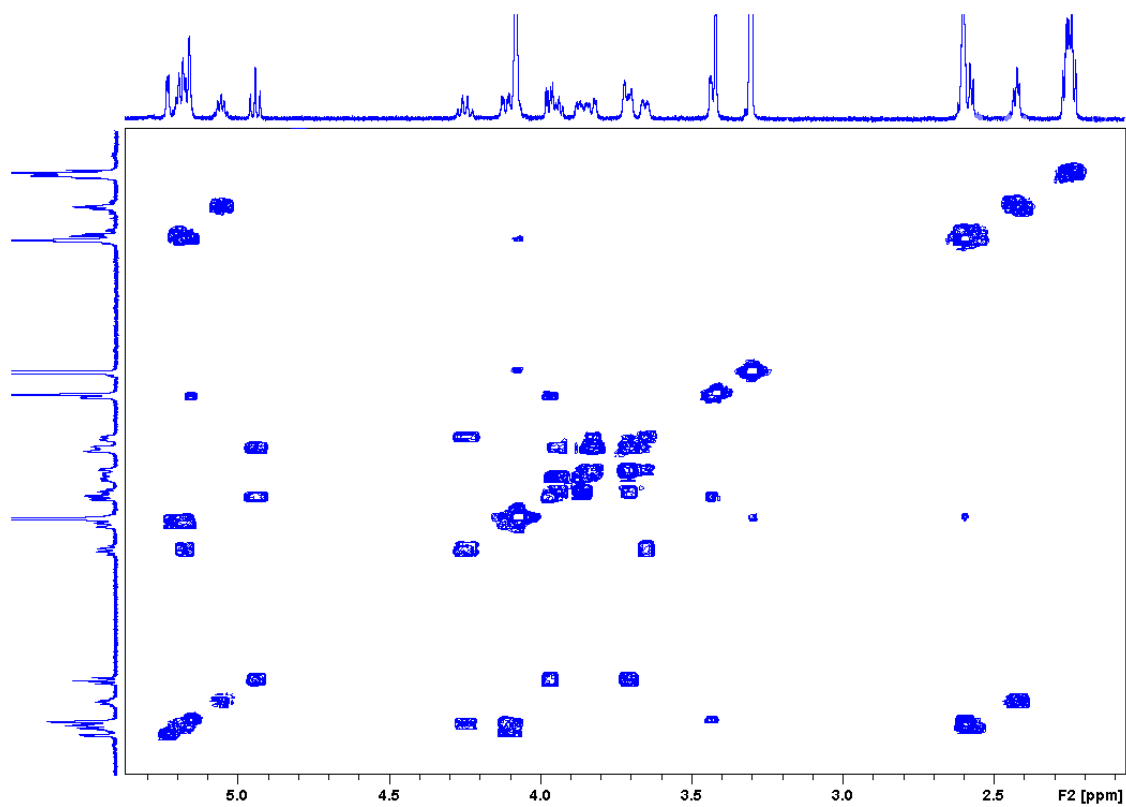




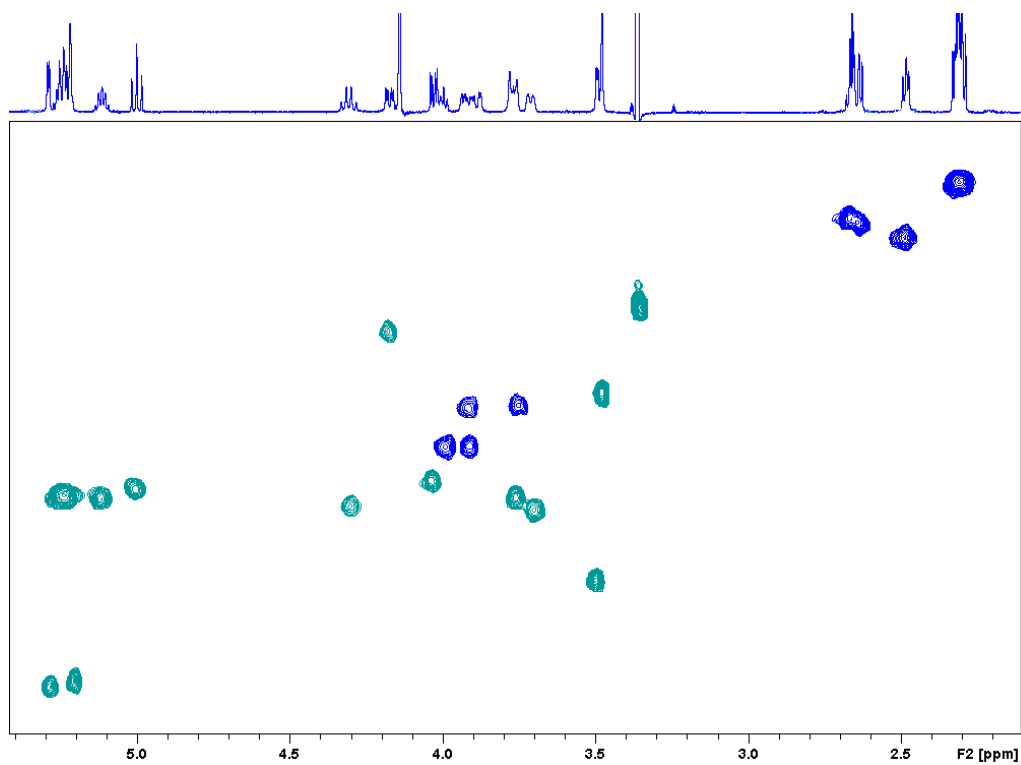




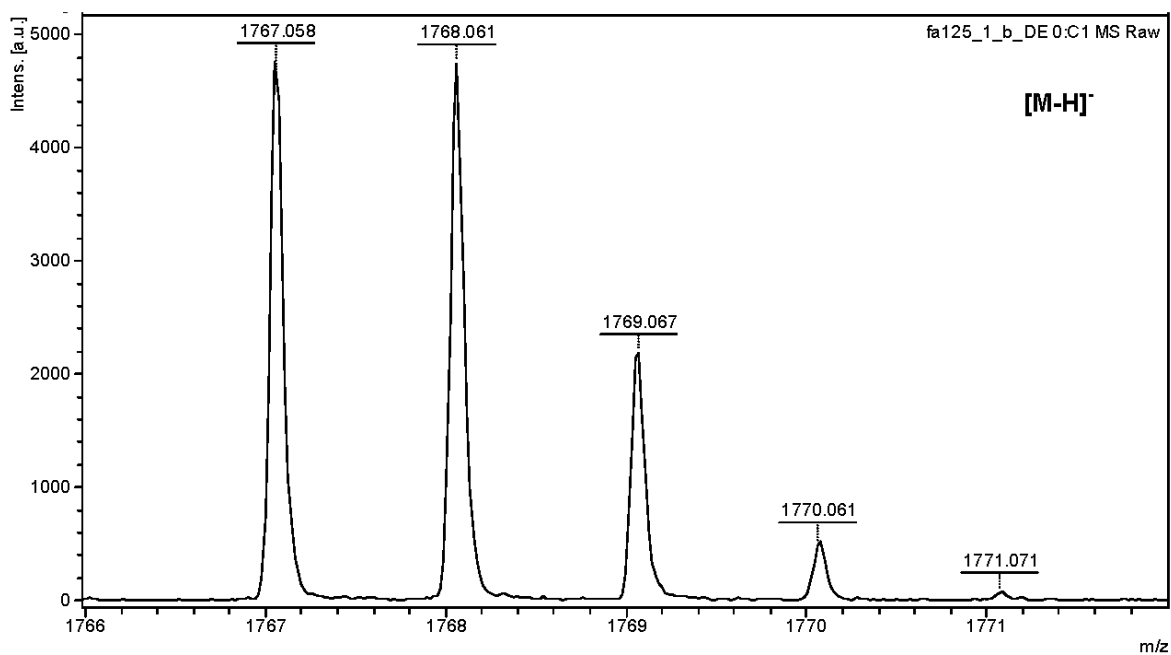
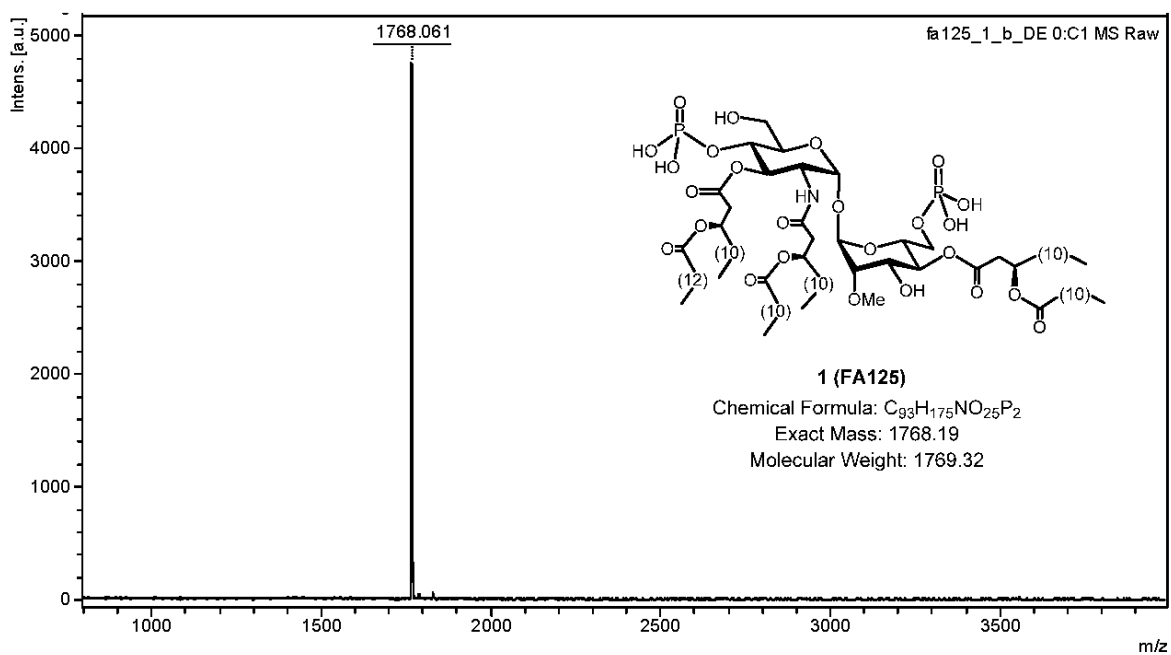
α,α -GM-LAM **1**, COSY, 600 MHz, CDCl₃-CD₃OD, 3:1

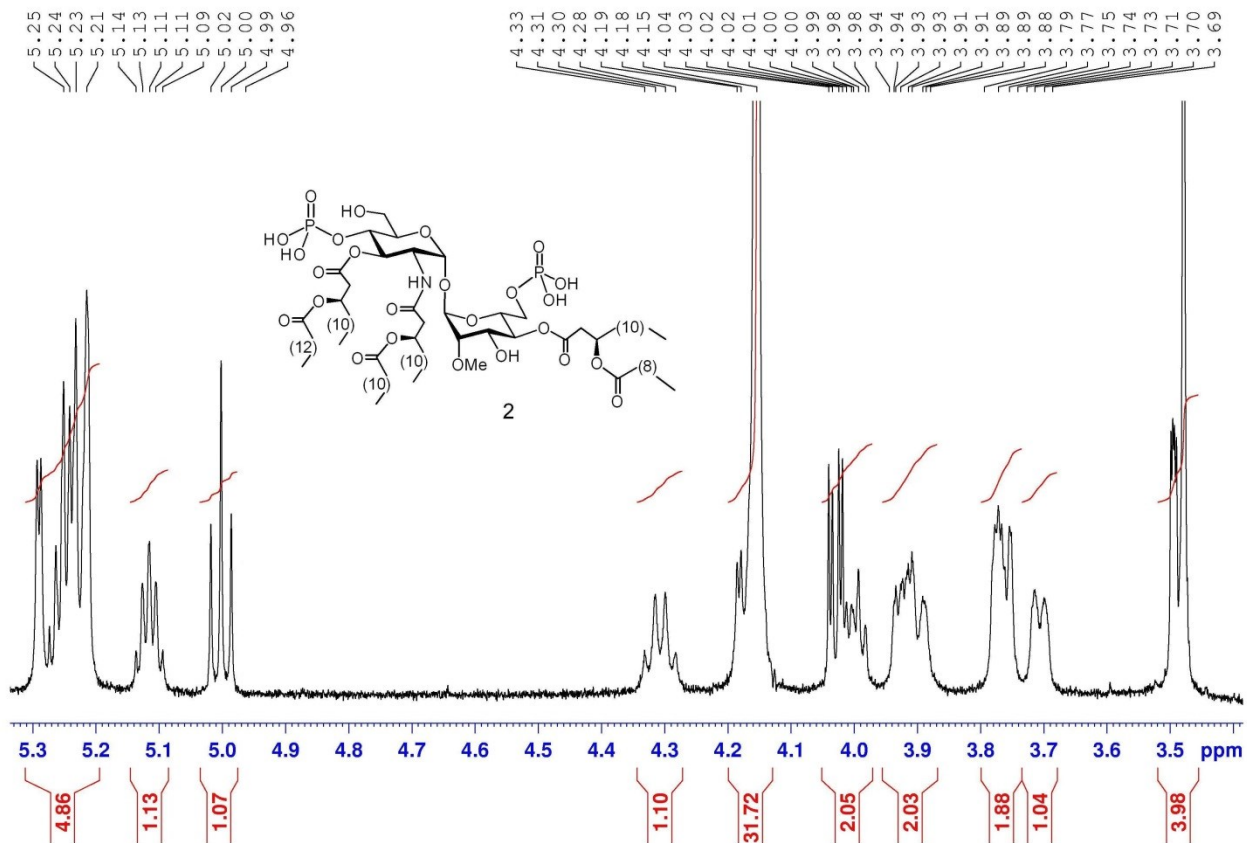
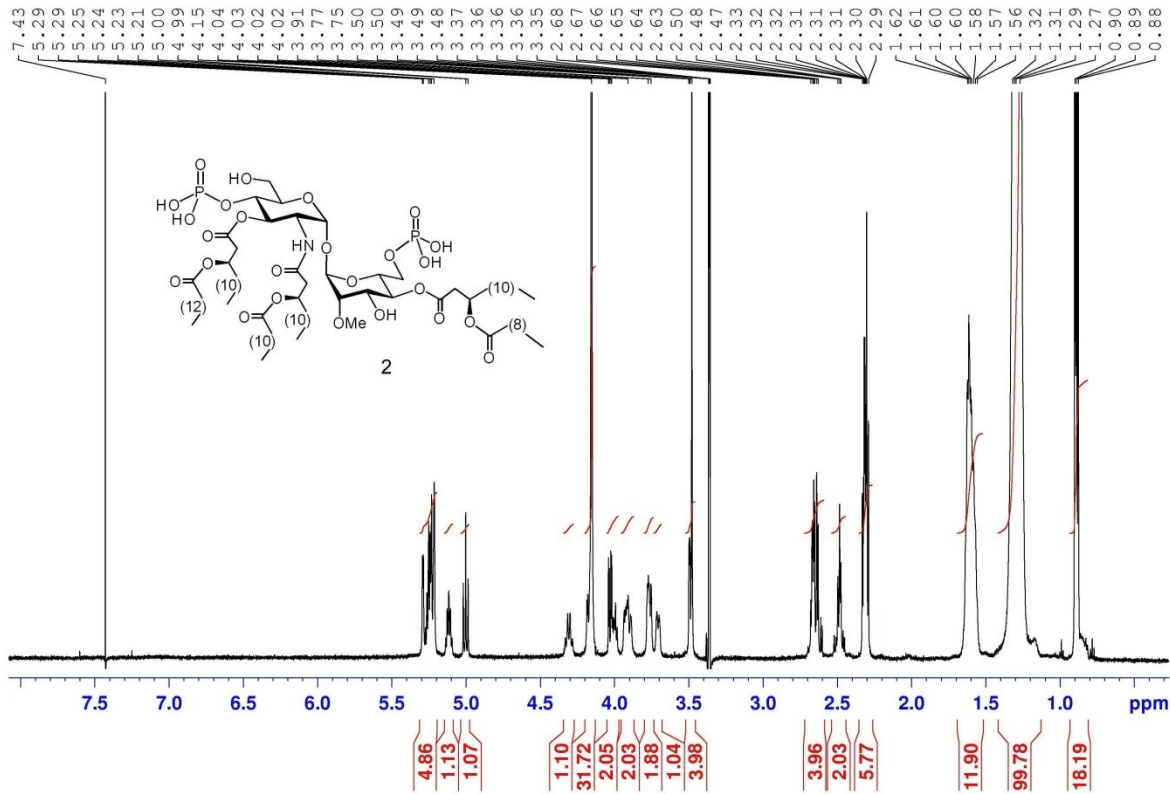


α,α -GM-LAM **1**, HSQC, CDCl₃-CD₃OD, 3:1

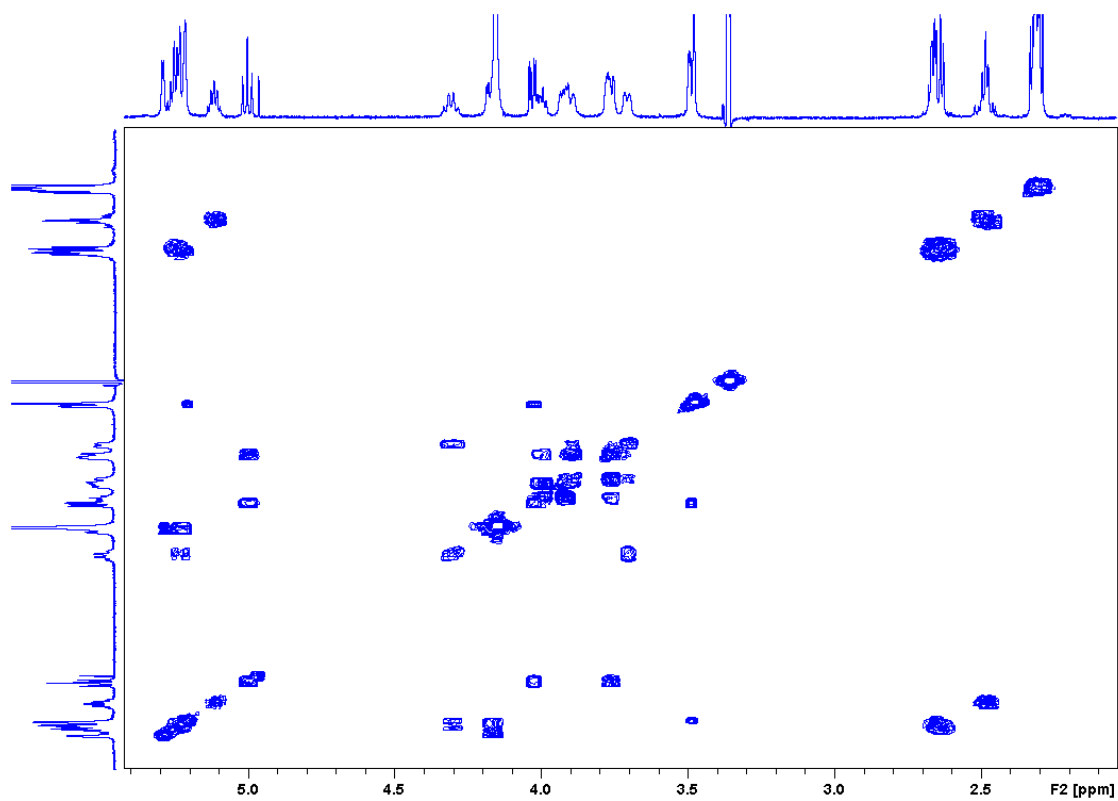


MALDI-TOF (neg. mode) spectrum of **1**

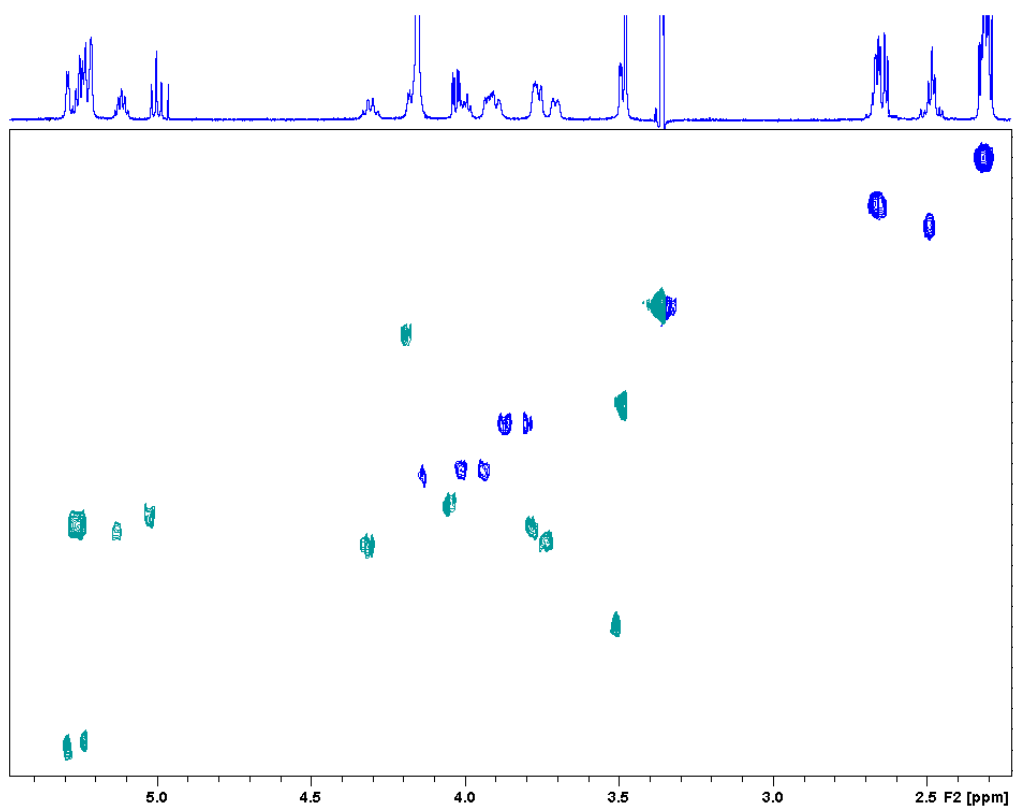




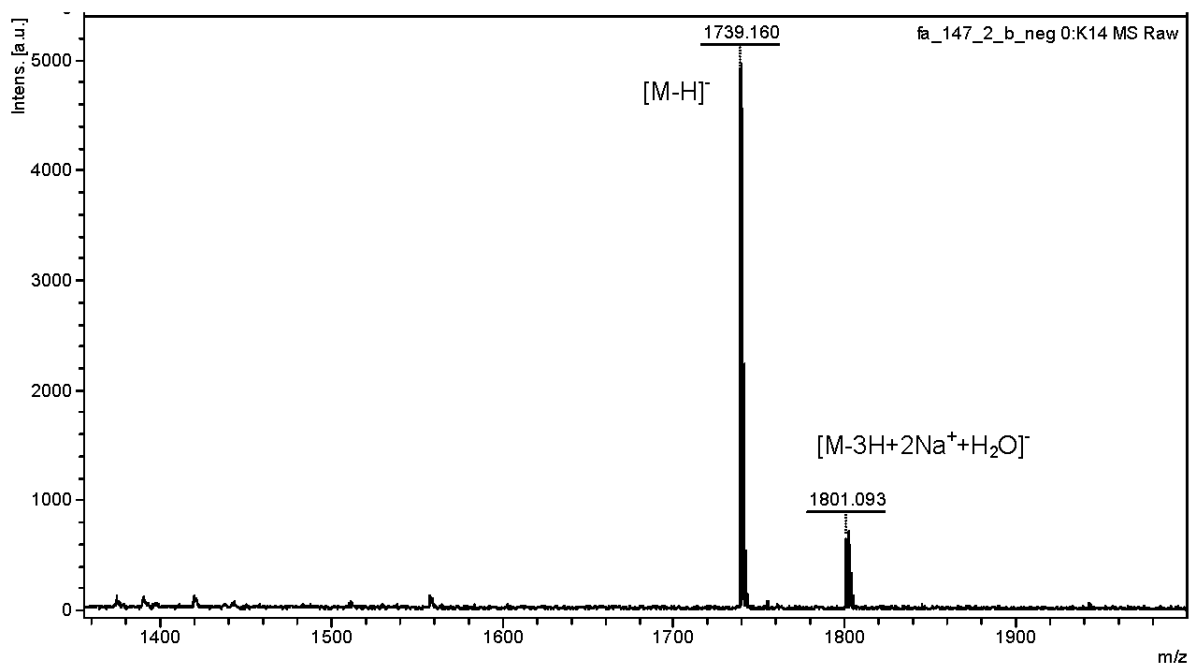
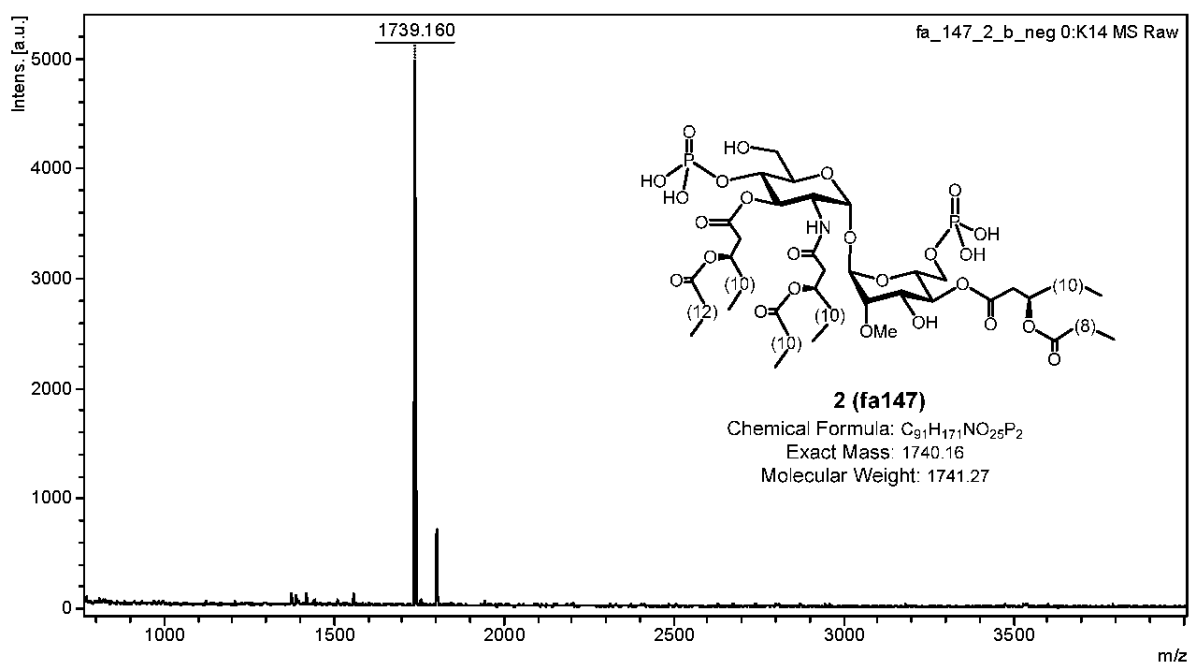
α,α -GM-LAM **2**, COSY, 600 MHz, CDCl₃-CD₃OD, 3:1

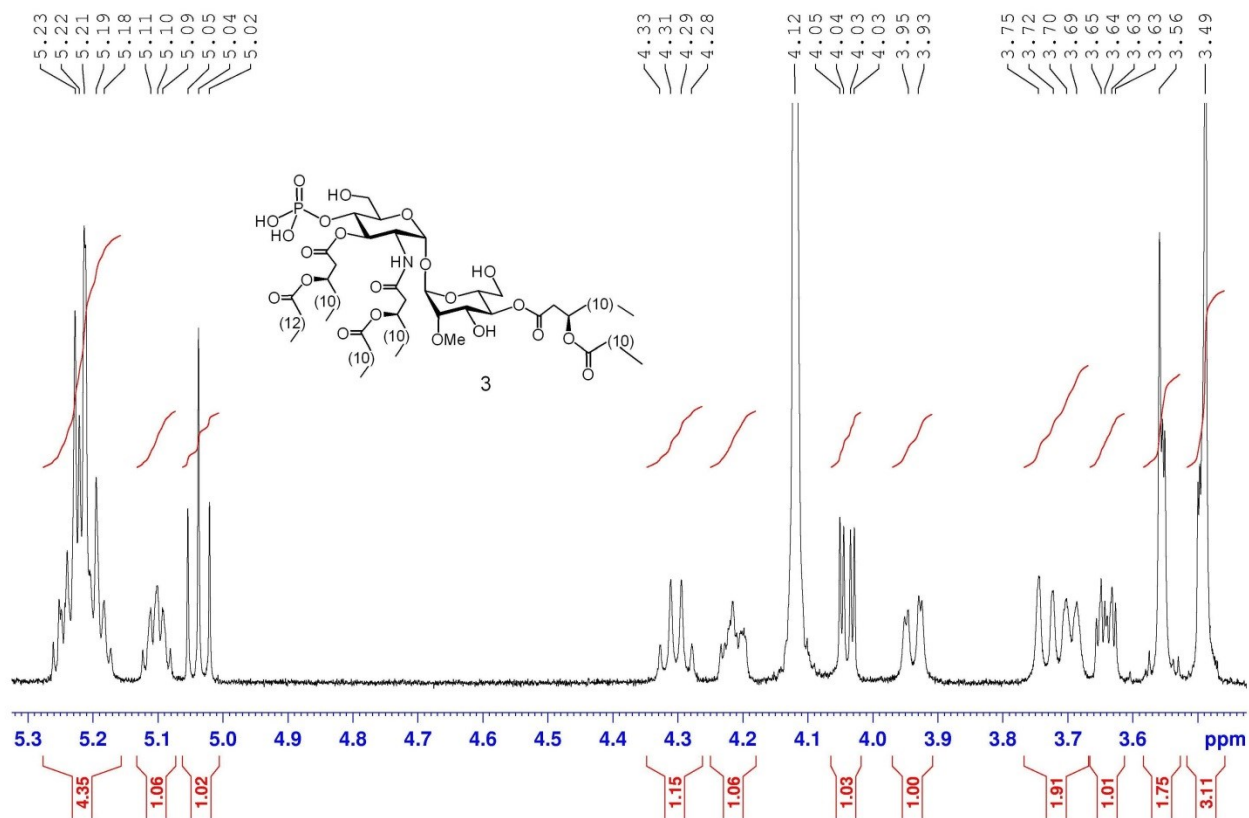
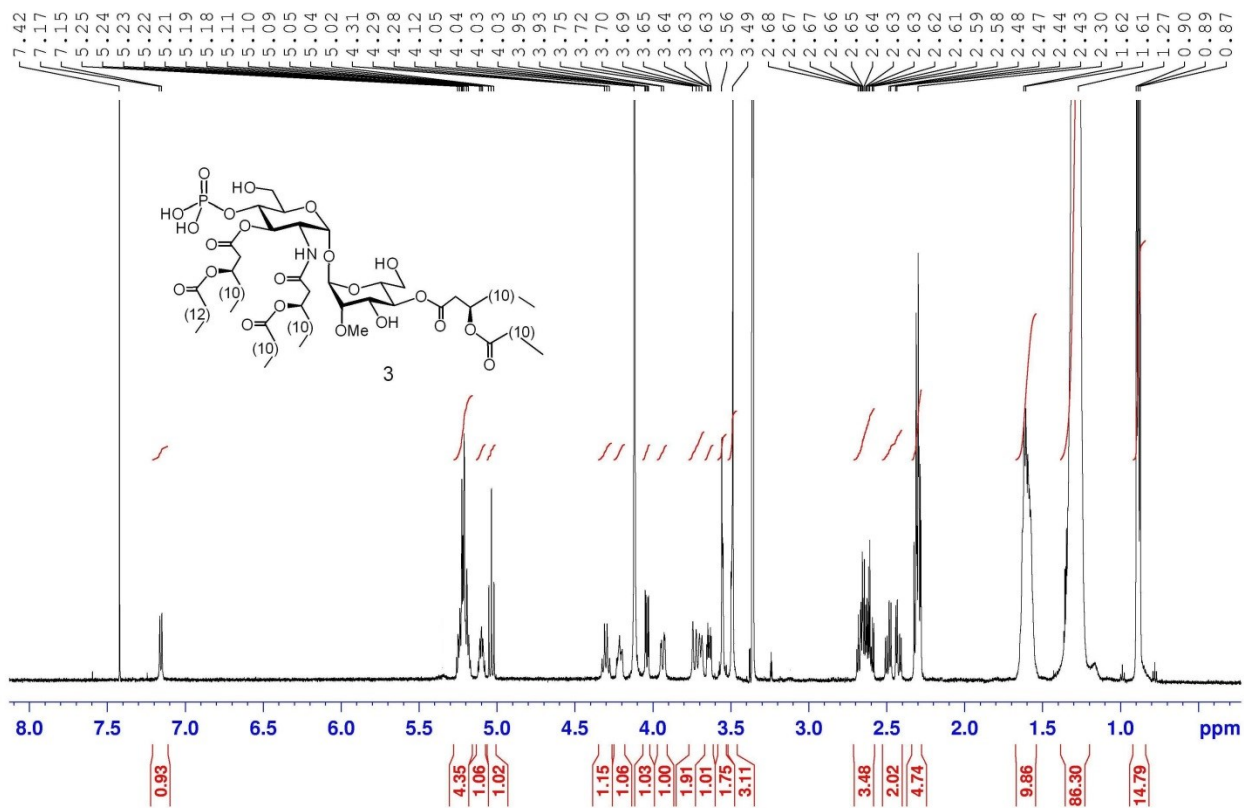


α,α -GM-LAM **2**, HSQC, CDCl₃-CD₃OD, 3:1

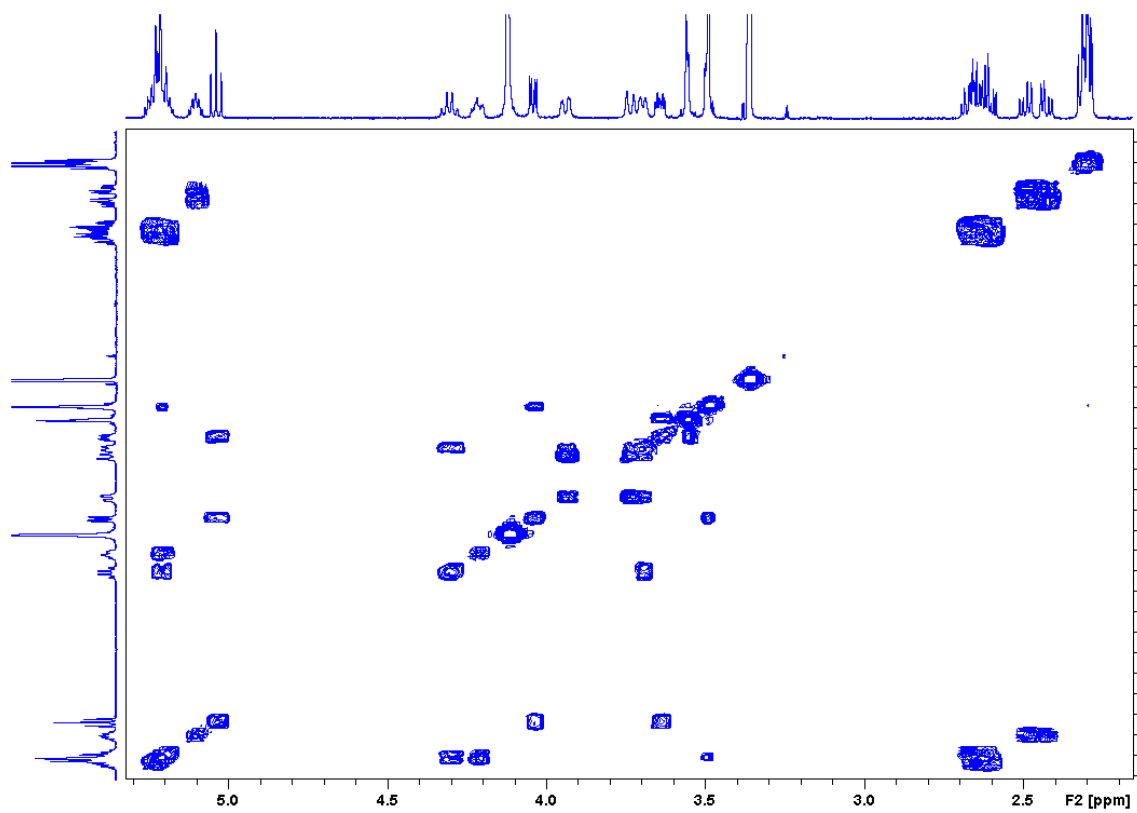


MALDI-TOF (neg. mode) of 2

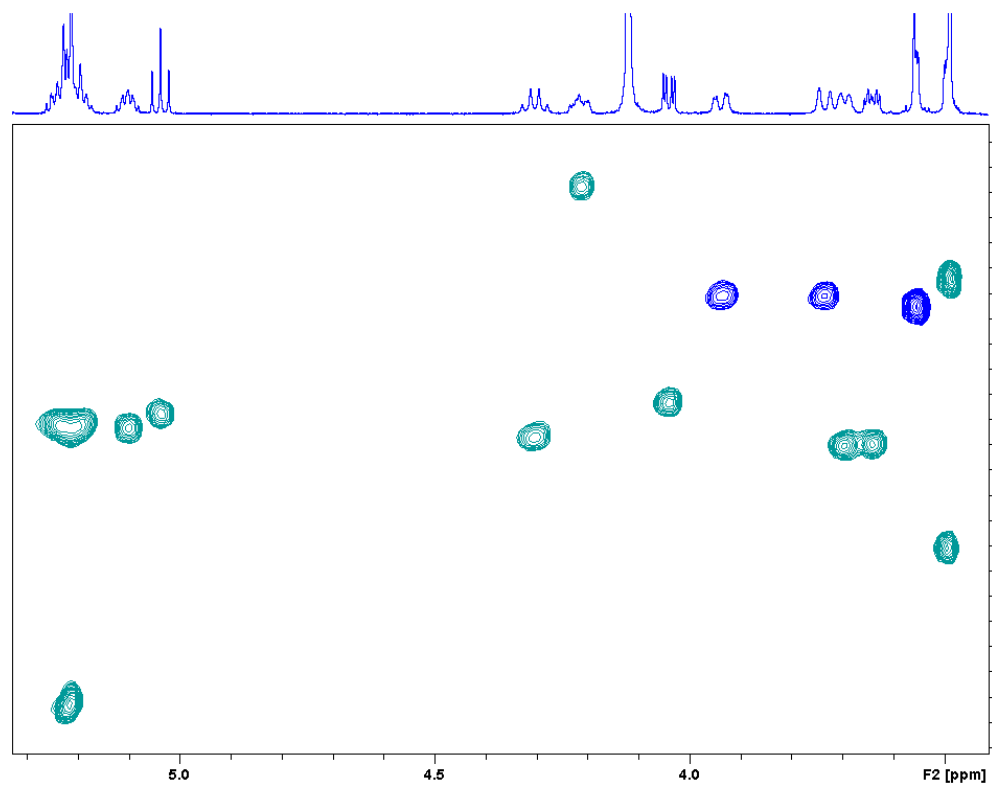




α,α -GM-LAM **3**, COSY, 600 MHz, CDCl₃-CD₃OD, 3:1



α,α -GM-LAM **3**, HSQC, CDCl₃-CD₃OD, 3:1



ESI-TOF (neg. mode) spectrum of 3

