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# **Abstract**

**Objectives:** We aimed to investigate the suitable condition of 8-hydroxy-2' -deoxyguanosine (8-OHdG) and micronucleus(MN) at different levels of occupational chromate exposure.

**Design**: A cross-sectional study was used.

**Participants:** 84 workers exposing chromate at least one year were chosen as exposed group, while 30 non-exposed individuals were used as controls.

**Main outcome measures:** Environmental and biological exposure of chromate was assessed respectively by measuring the concentration of chromate in the air (CrA) and blood (CrB) by ICP-MS in all subjects. CBMN was conducted, in which many indexes including MNCC, MNC, NPB and NBUD were calculated, while the urinary 8-OHdG was measured by ELISA method and normalized by the concentration of Cre.

**ants:** 84 workers exposing chromate at least one year were c<br>group, while 30 non-exposed individuals were used as controls.<br>**Ecome measures:** Environmental and biological exposure of chroma<br>respectively by measuring the **Results:** Compared with the control group, the levels of CrA, CrB, MNCC, MNC and 8-OHdG in chromate exposed group were all significantly higher (*P*<0.05). There was a positive correlation between log(8-OHdG) and LnMNCC or LnMNC (*r*=0.377 and *r*=0.362 ). The levels of LnMNCC , LnMNC and Log (8-OHdG) did not have the simple linear relationship with the concentration of CrB. There was a significantly positive correlation between Log (8-OHdG) and CrB when CrB level was below 10.50 ug/L (*r*=0.355). While a positive correlation was found between LnMNCC or LnMNC with CrB when CrB level was lower than  $9.10 \text{ ug/L}(r=0.365 \text{ and } r=0.269$ respectively).

**Conclusions:** The MN and 8-OHdG can be used as the biomarker for the genetic damage in chromate exposed group, but only when CrB levels were lower than 9.10ug/L and 10.50ug/L respectively, they can accurately reflect the degree of genetic damage.

**Key terms:** chromate; Genetic damage; 8-OHdG; Micronucleus, Concentration of chromate in the blood

# **Abbreviation:**

Cytokinesis-block micronucleus test-----CBMN

Micronuclei cell count ------MNCC

Micronucleus count -------MNC

Nuclear bridge------NPB

Nuclear bud --------NBUD

Creatinine------Cre

# *Strengths and limitations of this study(Article summary)*

**PAGE** 

 Strengths: All our results had provided new insight that only when the concentration of CrB was lower than 9.10ug/L and 10.45ug/L respectively, the MN and 8-OHdG can be used as the effective biomarkers to show the degree of genetic damage for the chromate occupational exposure, otherwise, above these levels, the cytotoxic effects might play an important role and the fate of cells with serious genetic damages may turn into apoptosis or necrosis, consequently which could lead to the false appearance of lower degree of MN and 8-OHdG at higher chromate exposed level.

Limitations: the size of this study population was not very large, especially the group of control individuals, so we chose some references to give the normal value of MN and 8-OHdG in other controls to reduce the mitigation of bias. In this research, we got the recommended condition of 8-OHdG and MN as genetic damage biomarker in chromate exposed group. It is necessary that more occupational epidemiological survey in different chromate producing factories should be chosen to verify this conclusion.

#### *Introduction*

Chromate is a widely used chemical in industrial and agricultural production in china, which could generate many pollutants including the waste water, waste gas and waste residue in chromate production, usage, transportation and storage. Long-term chromate exposure in occupational workplace can affect the health status of workers even cancer, so it has been declared as a well-known environmental and occupational hazards.

Example 10 and the minimizary and computer of the periadion of the term and the expansion of the c(Cr-VI) is the most harmful one. Cr-VI can enter human body n during occupational activities. When entering into respiratory There are many different valences of chromate, in which the hexavalent chromate (Cr-VI) is the most harmful one. Cr-VI can enter human body mainly by inhalation during occupational activities. When entering into respiratory system (nose, bronchial and lung), some Cr-VI could be converted to Cr-V, Cr-IV and Cr-III, and accumulated in bronco-alveolar lining fluid, mucosa and pulmonary tissues<sup>1,2</sup>. This transformation also can consequently form many reactive intermediates with oxidative stress<sup>3,4</sup>. Both Cr-III and ROS could contribute to interact with various proteins, DNA and other biological macromolecule to cause damage on DNA and chromosome, which can initiate carcinogenesis if accumulated to some degree<sup>5, 6, 7</sup>.

Based on the evidence above, 8-OHdG as a product of oxidative DNA damage, can be used to assess the oxidative damage and DNA mutations that are induced by the ROS in clinical<sup>8</sup>, environmental<sup>9,10</sup> and occupational setting in vitro<sup>11,12</sup>or in cell cultures<sup>13</sup>. MN in peripheral blood originates from chromosome fragments that are attacked by certain physical and chemical factors such as chromium or whole chromosomes that lag behind at anaphase during nuclear division. When the excision-repairable DNA lesions induced in G0/G1 phase, they can be converted to MN by using inhibitors of the gap filling step of excision repairmen, so that unfilled gaps are converted to double strand breaks after S phase<sup>14</sup>, so the frequency of MN can be used to reflect the genetic damage<sup>15</sup>. The CBMN test is a common method to detect the MN frequencies including many indexes such as MNCC, MNC, NPB and NBUD. In these indexes, MNCC and MNC were commonly used to detect the DNA  $\text{damage}^{14}$ .

 Previous studies have chose the relationship between CrB and urinary 8-OHdG or MN to discuss the feasibility of 8-OHdG and MN as the biomarker for chromium exposure, however, the conclusion about this connection have yielded conflicting results: Kuo found the linear correlation between urinary 8-OHdG and  $CrB<sup>16</sup>$ , but Gao, Kim and Zhang didn't confirm it subsequently<sup>17,18,19</sup>, many studies had been identified there was some association between CrB and MN frequencies<sup>20,21,22</sup>, but the suitable conditions and limitations for 8-OHdG and MN as biomarkers for occupational chromate exposure have still been unclear.

So our researches aimed to observe the effect of chromate exposure on genetic damage in occupational workers, especially urinary 8-OHdG for the oxidative DNA damage and MN for chromosome damage. Then it was discussed whether MN frequency and 8-OHdG can be used as the effective genotoxic biomarkers at different levels of chromate exposure.

# *Materials and methods*

### *Study design and population*

Is and limitations for 8-OHdG and MN as biomarkers for occ<br>
exposure have still been unclear.<br>
In occupational workers, especially urinary 8-OHdG for the oxidat<br>
and MN for chromosome damage. Then it was discussed whe<br>
y a A cross-sectional survey was designed for this research. The factory was chosen as the work place in Henan province in china because (1) the product--potassium dichromate was relatively simple, most of which was water-soluble hexavalent chromate. (2) Annual health check-ups were offered in this factory for workers, which allowed us to easily collect specimens from workers to minimize the interference with normal work schedules.

In this research, 84 workers exposed to chromate in the factory were chosen as exposed group, while 30 non-exposed individuals working in the administration office were as control group. The criteria of subjects included: (1) workers in exposed group were at least one year employment and 3 months working in the same work position. (2) aged between 25-50. (3) no medical history of allergy, asthma or allergic rhinitis, (4) all subjects with skin infections, fever or other clinical diseases should be excluded during the sampling period. (5) Pregnant and nursing women were not

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enrolled. .

All subjects were in the same factory with similar education and social backgrounds. They all were requested to complete a questionnaire and had a clinical examination. The questionnaire included a lot of information including occupational history, personal medical history, medication used in 4 weeks before the study, body weight and height, hair dye, house decoration, radiation exposure, individual protection, smoking status and alcohol intake.

#### *Ethical consideration*

This study was approved by Medical Ethics Committee of Peking University, Health Science Center (HSC), Beijing, China. Written informed consent from each study subject was also obtained.

#### *Air and Biological exposure assessment*

m, smoking status and alcohol intake.<br> **Formulation**<br> **Fo** Six air sampling sections were respectively chosen in the workplaces of two groups, 10 sampling points were chosen in each section. The sampling process was used pumping at 1L/min for 8 hours (Sp730, TSI Corporation, USA), the membranes used in this study were MCE mixed cellulose ester filters (Φ37mm, pall, America).The average concentration of all sampling points on the membranes in the same group was measured by atomic absorption spectrometry and then calculated to evaluate the CrA during the whole production process, the detection limit for the CrA was 0.001 µg/L.

2ml anticoagulant (EDTA and heparin) blood in peripheral was drew from each subject after finishing the questionnaire and then assigned to two tubes on average. They were respectively used to measure the CrB and CBMN. The concentration of CrB was measured by inductively coupled plasma mass spectrometer  $(ICP-MS)^{26}$ . the detection limits was 0.0012 µg/L. The tubes used above had been detected their background value before research to ensure less elements and heavy metals contamination. .

 At the end of the work-shift, 30mL urine sample of each subject was collected into a 50 mL metal-free polypropylene centrifuge tube (Falcon, BD Biosciences) and

#### *Determination of urinary 8-OHdG*

According to the manufacturer's instructions, urine samples were firstly centrifuged at 1500 rpm for 7 min, secondly the supernatant was chosen to determine the content of urinary 8-OHdG using ELISA kit (USA Cayman chem, USA Cayman chem, 8-OHdG EIA kit) by Multiskan MK3 (Thermo, USA) and Cre by alkaline picric acid assay with a commercial kit (Ausbio Laboratories Co., Ltd. China) using a Hitachi 7170A automatic analyzer (Hitachi Corp, Japan). The results of 8-OHdG were regulated by Cre to avoid the potential interference of different urine density among the subjects.

### *CBMN test*

**For the Amplitude Community (Fig. 1997)** and EV by das a commercial kit (Ausbio Laboratories Co., Ltd. China<br>170A automatic analyzer (Hitachi Corp, Japan). The results of 8-OH<br>by Cre to avoid the potential interference of Peripheral venous blood which was designed in heparinized tubes with individual numbers was taken to measure the MN frequency. The indexes (MNCC, MNC, NPB and NBUD) were counted in 1000 binuclear lymphocytes of each individual according to Fenech's protocol<sup>23</sup>. All scoring was carried out by two independent researchers through double - blind method. If the result from these two researchers was different, another researcher should be asked to verify the scores.

#### *Statistical analysis*

 Epidata 3.0 software was used to entry the questionnaire and experimental data into computer. The whole process was utilized double-entry and logistical error check to ensure the accuracy.

 All analysis was performed with SPSS16.0. Normality was assessed by Kolmogorov-Smirnov (*K-S*) test, the variables including 8-OHdG, MNCC and MNC did not meet the normality, so Log or Ln transformation was made for normality approximation. Continuous and categorical parameters between chromate exposed group and control group were tested using the two sample *t* test (or Mann-Whitney U nonparametric test) and  $\chi^2$  test. Curving correlation and linear regression were performed. Statistical significance was for two-sided. The *P* values were defined as *α*<0.05.

#### *Results*

#### *General information analysis*

 A total of 114 subjects including 84 chromate exposed workers (mainly in form of  $K_2Cr_2O_7$ ) and 30 controls were recruited in this study. The working age of chromate exposed group was  $(7.82 \pm 5.51)$  years. Furthermore, the personal protection (gloves and masks) of workers was above 90%. The demographic characteristics of all subjects in this research were presented in Table 1, which showed that there were no significant differences in the distribution of gender, age, smoking and alcohol consumption between the two groups.

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consumption between the two groups.												
Table 1 General information of chromate exposed group and control group												
	Group	Exposed group	Control group	T	$\chi^2$	$\boldsymbol{P}$						
		$(n=84)$	$(n=30)$									
Age	$\overline{X} \pm S$	$35.73 \pm 7.85$	$34.83 \pm 8.83$	0.432		0.666						
$\leq$ 35	$n$ (%)	40 (47.62)	17(56.67)		0.187	0.493						
$>35$		44 (52.38)	13(43.33)									
Gender	$n$ (%)											
Male		62(73.81)	19(63.33)		0.869	0.351						
Female		22 (26.19)	11(36.67)									
Smoke	$n$ (%)											
Yes		30(35.71)	8(26.67)		0.610	0.435						
N <sub>0</sub>		54 (64.29)	22(73.33)									
Alcohol	$n$ (%)											
Yes		29 (34.52)	16(53.33)		2.530	0.092						
No		55 (65.48)	9(46.67)									
8-OHdG												
$(ug/g$ Cre)	$\overline{X} \pm S$	43.76±34.89	$27.21 \pm 13.76$	3.354		< 0.001						
$MNCC$ $(\%_0)$	M(Q)	6.00(4.00)	3.20(2.10)	2.420		0.004						
MNC $(\%_0)$	M(Q)	7.40(4.47)	3.74(2.94)	3.401		0.001						
NBUD $(\%_0)$	M(Q)	1.11(1.20)	1.16(1.17)	0.163		0.871						
$NPB$ $(\%$ <sup>0</sup>	M(Q)	1.28(1.15)	1.42(1.34)	0.476		0.635						

**Table 1 General information of chromate exposed group and control group** 

 Note: Smoking referred to suck at least one cigar per day and last one year or more, smoking quit but ess than one year was also included. Alcohol was definitive by weekly drinking no less than three times

#### *Concentration of CrA and CrB*

As was shown in Figure 1, the concentration of CrA in chromate exposed group

[ $(15.45 \pm 19.00)$  µg/m<sup>3</sup>] was much higher than that in control group [ $(0.23 \pm 0.38)$ ]  $\mu$ g/m<sup>3</sup>](*P*<0.001), but still under the exposure limitation of chromate [ $(50\mu g/m^3)$  (2012, ACGIH)]. The levels of CrB in chromate exposed group [ $(9.45 \pm 1)$ 9.47)  $\mu$ g/L] were also significantly higher than that in control group [(4.05  $\pm$ 1.87)  $\mu$ g/L] (*P*<0.001).

#### *Levels of urinary 8-OHdG, and serum CBMN indexes*

As for the data distribution of indexes such as 8-OHdG, MNCC, MNC, NBUD and NPB (Table 1), there were three kinds of indexes (8-OHdG, MNCC and MNC) . They were higher in chromate exposed group than that in control group  $(P<0.05)$ , which showed that 8-OHdG, MNCC and MNC could be used as the genetic damage biomarkers feasibly caused by chromate exposure.

#### *Correlation*

 As two biomarkers for genetic damage, there was a positive correlation between log (urinary 8-OHdG) and LnMNCC and LnMNC (*r*=0.377 and *r*=0.362 respectively) *P*<0.05)(Figure 2).

**EXECUTE:** The were three kinds of indexes such as 8-OHdG, MNCC, MNC, N<br>ble 1), there were three kinds of indexes (8-OHdG, MNCC and MN<br>her in chromate exposed group than that in control group (*P*<0.05<br>that 8-OHdG, MNCC a Correlation was analyzed between the concentration of urinary 8-OHdG and CrB. As was recorded in Figure 3, there was no linear correlation but a curve fitting between CrB and Log (8-OHdG), we found the value of 8-OHdG was decreased when the concentration of CrB was more than 10.50µg/L. Based on the results above, the concentration of CrB was stratified into two groups: the high exposed group (CrB  $\geq 10.50\mu g/L$ ) and the low exposed group (CrB <10.50 $\mu g/L$ ). A positive correlation was shown between CrB and log (8-OHdG) when the CrB Level was lower than10.50µg/L (r=0.355, *P*<0.05), while there was a negative correlation between CrB and log (8-OHdG) when the CrB Level was higher than10.50µg/L.

The relationship between the concentration of MNCC or MNC and CrB was also analyzed in this research (Figure 4). there were no linear correlation but a curve fitting between CrB and LnMNCC or LnMNC. We found the value of MNCC or MNC was decreased when the concentration of CrB was more than 9.10µg/L. Based on the results above, the concentration of CrB was stratified into two groups: the high

exposed group (CrB  $\geq 9.10 \mu g/L$ ) and the low exposed group (CrB  $\leq 9.10 \mu g/L$ ). A positive correlation was shown betweenCrB and LnMNCC or LnMNC in higher chromate exposed group  $(r=0.365$  and  $r=0.269$  respectively,  $P<0.05$ ), while a significantly negative relationship was found between CrB and LnMNCC or LnMNC in the lower chromate exposed group. $(r=-0.279$  and  $r=-0.261$  respectively,  $P<0.05$ ).

# *Discussion*

**For peer review only** Long-term and low level chromate exposure can not only increase the body's internal load but also cause a variety of harmful effects on workers' health even increase the incidence of human cancer  $24, 25$ . In occupational activity, chromate could enter into workers' body mainly by respiratory system, then be metabolism and excreted by urine, so our previous researches have proved the concentration of chromium in whole blood and urine can be used as the indicators to assess chromate biological exposure<sup>6, 26</sup>. In this study, it was found that the concentrations of CrA and CrB in exposed group were all significantly higher than that in control group (*P*<0.05) ,while the CrB level in chromate exposed group was nine times more than that in the general population of our country  $(1.19 \mu g/L)^{27}$ , which showed that the conclusion was credible by contrasting the genetic damage indexes between chromate exposed group and control group.

As we all known when Cr-VI enters human blood, it will be converted into other valence chromate compounds such as Cr-III, which could not only produce a large amount of ROS that can cause the redox system imbalance, but also directly or indirectly coordinate with DNA or protein. The transformation above can affect genetic stability including oxidative DNA lesion, DNA cross-links, single and double strand breaks and so  $\text{on}^{28,29,30}$ . Besides, long-term chromate exposure can also cause cyto-toxicity to lead cells apoptosis $3<sup>1</sup>$ .

 Urinary 8-OHdG has been demonstrated as a biomarker for oxidative DNA damage in chromate exposure not only by animal experiment but also by many

redict the degree of DNA damage. There are some reasons for th<br>umner E.R has proved that oxidative DNA damage by chromate<br>trget on specifically certain glycol tic enzymes on 8-OHdG<sup>34</sup>, whice<br>the product of CrB was produce epidemiological researches<sup>32,33</sup>, because it is the site that ROS often attacks, but the dose-response relationship with occupational exposure indicators was not depicted clearly. Overall, in this research, the relationship between urinary 8-OHdG and CrB was two-way changes. The concentration of urinary 8-OHdG was not significantly increasing when the level of CrB is more than  $10.50\mu g/L$ , which showed that when the CrB was higher than some degree, the concentration of urinary 8-OHdG would fail to predict the degree of DNA damage. There are some reasons for this result, firstly Sumner E.R has proved that oxidative DNA damage by chromate exposure mainly target on specifically certain glycol tic enzymes on -OHd $G<sup>34</sup>$ , which means once higher burden of CrB was produced, it could oxidize and change the structure of glycolytic enzymes to reduce the production of free 8-OHdG. secondly, some researchers have proved that 8-OHdG was not the final product of redox reaction, so when the level of CrB is higher, 8-OHdG could be converted to these further oxidation products such as Spiroiminodihydantoin  $(Sp)^{35}$ , so the concentration of free 8-OHdG was reduced. Thirdly, as we all know, the kidney can be damaged by chronic chromate exposure. As the level of CrB increased, the kidney was at greater health risk, which ultimately affect the exertion of 8-OHdG in urine<sup>36</sup>.

Micronuclei is another biomarker which commonly used as the genetic damage of chromate exposure<sup>37,38</sup>. Many researches showed that chromate exposure could cause the increasing of MN frequency in cell research, animals experiment, and human being<sup>39,40</sup>. In this research, the similar conclusion was proved that the MN frequency was significantly higher in chromate exposed group than that in control group and the general population<sup>41</sup>( $P$ <0.05). The frequency of MNCC and MNC (as the sensitive indexes of MN) has statistical differences between these two groups. However, the dose-response relationship with CrB still needs many further investigations, so this research was meaningful to discuss the suitable condition of MN as genetic damage. In this study, no linear correlation was shown between CrB and LnMNCC or LnMNC, the MN frequency did not increase as the CrB elevated. When all subjects were divided into two subgroups by the concentration of CrB, a positive correlation was shown between CrB and LnMNCC and LnMNC in high CrB

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group( $CrB\geq 9.10\mu g/L$ ) ( $r=0.365$  and  $r=0.269$  respectively), while a significantly inverse relationship was found between CrB with LnMNCC and LnMNC in low CrB group( $CrB < 9.10 \mu g/L$ ) ( $r = -0.279$  and  $r = -0.261$  respectively). The reason for these results may be that the occupational chromate exposure can increase the frequency of the MN in some range. As the increasing of CrB, more serious genotoxic damage should be caused such as the apoptosis or necrosis of lymphocyte, which contrarily decrease the frequency of MN in this situation<sup>42,43</sup>.

 Both MN and urinary 8-OHdG can be used to predict different types of DNA damage. A positive correlation between urinary 8-OHdG and MN was found, which suggested that these two indicators as genetic damage biomarker can be verified each other.

# *Conclusions*

the frequency of MN in this situation<sup>42,43</sup>.<br>
MN and urinary 8-OHdG can be used to predict different types<br>
A positive correlation between urinary 8-OHdG and MN was found<br>
that these two indicators as genetic damage bioma All these above had provided new insight that both MN and urinary 8-OHdG can be used as the genetic damage biomarkers caused by occupational chromate exposure at some leve, the combination of these indicators can improve the credibility of the results. It is not the simple linear relationship between their concentration and the level of chromate exposure. Only when the level of CrB is below 9.10µg/L and 10.50µg/L, the MN frequency and urinary 8-OHdG can show the degree of genetic damage respectively produced by chromate exposure quantitatively.

# *Contributor ship statement*

All the authors included in the paper grant the criteria of the authorship. PL together with YG conceived and executed this investigation, analyzed of the data and described this manuscript. YL and JY supported this investigation from the epidemiological aspect. GJ is responsible for the whole conduct of this study and all

content of this. SFY contribute to the organization and arrangement for the scene investigation. All authors commented critically on the manuscript and agreed with this submitting.

### *Competing interests*

There are not competing interests.

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#### *Data sharing*

There is not data sharing.

### **Reference**

- 1. Monika Gube, Joachim Ebel, Peter Brand, et al. Biological effect markers in exhaled breath condensate and biomonitoring in welders: impact of smoking and protection equipment[J]. Int Arch Occup Environ Health, 2010, 83 (7): 803–811.
- 2. Andrea Caglieri, Matteo Goldoni, Olga Acampa, et al. The effect of inhaled chromium on different exhaled breath condensate biomarkers among chrome-plating workers[J]. Environmental Health Perspectives, 2006, 114 (4): 542-546.
- 3. Shrivastava Richa, Upreti RK, Seth PK, et al. Effects of chromium on the immune system[J]. Immunology and Medical Microbiology, 2002, 34 (4): 1-7.
- 4. Shashwat Katiyar, Awasthi SK, Srivastava JK. Effect of chromium on the level of IL-12 and IFN- $\gamma$  in occupationally exposed workers[J]. Science of the total environment, 2009, 407 (6): 1868-1874.
- 5. Mohammad BH, Marie Vahter, Gabriela Concha, et al. Low level environmental cadmium exposure is associated with DNA hypomethylation in argentinean women[J]. Environmental Health Perspectives, 2012, 120 (6): 879–884.
- 6.Xiaohua L, Yanshuang S, Li W, Yuhui L, Ji Z, Yanhui M, Yun W, Wenjun M, Lei Y, Guang J. Evaluation of the correction between genetic damage and occupational chromate exposure through BMMN frequenciess[J]. J Occup Environ Med 2012, 54(2):166-170.
- 7. De Flora S, Bagnasco M, Serra D, et al. Genotoxicity of chromium compounds: A review[J]. Mutation Research/Reviews in Genetic Toxicology, 1990, 238 (5): 99-172.
- 8. Wu LL, Chiou C-C, Chang P-Y, et al. Urinary 8-OHdG: a marker of oxidative stress to DNA

and a risk factor for cancer, atherosclerosis and diabetics[J]. Clinica Chimica Acta, 2004, 339  $(1-2)$ : 1-9.

- 9. Ren C, Fang S, Wright RO, et al. Urinary 8-hydroxy-2-deoxyguanosine as a biomarker of oxidative DNA damage induced by ambient pollution in the Normative Aging Study[J]. Occupational and environmental medicine, 2011, 68 (2): 562-569.
- 10. Lin CY, Lee HL, Chen YC, et al.Positive association between urinary levels of 8-hydroxydeoxyguanosine and the acrylamide metabolite N-acetyl-S-(propionamide)-cysteine in adolescents and young adults[J]. J Hazard Mater, 2013, 261: 372-377.
- 11. Kim JY, Mukherjee S, Ngo LC, et al. Urinary 8-hydroxy-2'-deoxyguanosine as a biomarker of oxidative DNA damage in workers exposed to fine particulates[J]. Environmental health perspectives, 2004, 112 (6): 666-671.
- 12. Valavanidis A, Loridas S, Vlahogianni T, etal.Influence of ozone on traffic-related particulate matter on the generation of hydroxyl radicals through a heterogeneous synergistic effect[J].J Hazard Mater. 2009, 162 (2-3): 886-892.
- 13. Valavanidis A, Vlachogianni T, Fiotakis C. 8-hydroxy-2-deoxyguanosine (8-OHdG): a critical biomarker of oxidative stress and carcinogenesis[J]. Journal of environmental science and health Part C, 2009, 27 (2): 120-139.
- Formation of hytomation in eta. Thumatente to location of the control of the characterial position of the generation of hydroxyl radicals through a heterogeneous synergistic eff Mater. 2009, 162 (2-3): 886-892.<br>
For peer 14. M. Fenech, M. Kirsch-Volders1, A. T. Natarajan, etal. Molecular mechanisms of micronucleus, nucleoplasmic bridge and nuclear bud formation in mammalian and human cells. Mutagenesis vol, 2011, 26: 125 - 132.
- 15. Sobol Z, Schiestl RH. Intracellular and extracellular factors influencing Cr (VI) and Cr (III) genotoxicity[J]. Environmental and Molecular Mutagenesis, 2012. 53 (2): 94-100.
- 16. Kuo H, Chang S, Wu K, et al. Chromium (VI) induced oxidative damage to DNA: increase of urinary 8-hydroxydeoxyguanosine concentrations (8-OHdG) among electroplating workers[J]. Occupational and environmental medicine, 2003, 60 (8): 590-594.
- 17.Kim H, Cho S-H, Chung M-H. Exposure to hexavalent chromium does not increase 8-hydroxydeoxyguanosine levels in Korean chromate pigment workers[J]. Industrial health, 1999, 37 (3): 335-341.
- 18. Gao M, Levy LS, Faux SP, et al. Use of molecular epidemiological techniques in a pilot study on workers exposed to chromium[J]. Occupational and environmental medicine, 1994, 51 (10): 663-668.
- 19. Zhang XH, Zhang X, Wang XC, et al. Chronic occupational exposure to hexavalent chromium causes DNA damage in electroplating workers.[J] BMC Public Health, 2011, 11 (3): 224.
- 20. Xiaohua L, Yanshuang S, Li W, et al. Evaluation of the correction between genetic damage and occupational chromate exposure through BMMN frequencies. J Occup Environ Med 2012, 54 (2): 166-170.
- 21. Danadevi K, Rozati R, Banu BS, et al. Genotoxic evaluation of welders occupationally exposed to chromium and nickel using the Comet and micronucleus assays[J]. Mutagenesis, 2004, 19 (1): 35-41.
- 22. Thompson CM, Fedorov Y, Brown DD, et al. Assessment of Cr (VI)-induced cytotoxicity and genotoxicity using high content analysis[J]. PloS one, 2012, 7 (8): 742-720.
- 23. Fenech M. Cytokinesis-block micronucleus cytome assay[J]. Nature protocols, 2007, 2 (1): 1084-1104.
- 24. Clancy HA, Sun H, Passantino L, et al. Gene expression changes in human lung cells exposed to arsenic, chromium, nickel or vanadium indicate the first steps in cancer[J]. Metallomics. 2012, 4(8):784-793.
- 25. Chhabra D, Oda K, Jagannath P, etal. Chronic heavy metal exposure and glbladder cancer risk in India, a comparative study with Japan[J]. Asian Pacific Journal of Cancer Prevention. 2012, 13(1): 187-190.
- 26.Yan L, Jia G, Zhang J, He PP, Wang TC, Zhang N, Liu LZ, Li GR, Wang X, Wang L. The correlation between personal occupational exposure to soluble chromate and urinary chromium content. Zhonghua yu Fang yi xue za zhi (Chinese Journal of Preventive Medicine) 2006, 40(6): 386-389 (Article in Chinese).
- 27. Ding CG, Pan Yj, Zhang AH, et al. Distribution of chromium in whole blood and urine among general population in China between year 2009 and 2010. Zhonghua yu Fang yi xue za zhi (Chinese Journal of Preventive Medicine) 2012, 46(8): 679-682. ( Article in Chinese).

- 28. Zhitkovich, A. Importance of chromium-DNA adducts in mutagenicity and toxicity of chromium(VI)[J]. Chem. Res. Toxicol., 2005, 18 (1): 3-11.
- 29. Wang TC, Song YS, Wang H, etal. Oxidative DNA damage and global DNA hypomethylation are related to folate deficiency in chromate manufacturing workers[J]. Journal of Hazardous Materials,2012, 213.214:440-446.
- 30. Brien, T.J.; Ceryak, S.; Patierno, S.R. Complexities of chromium carcinogenesis: role of cellular response, repair and recovery mechanisms[J]. Mutat. Res. Fundam. Mol. Mech. Mutagen., 2003, 533 (14): 3-36.
- 31. Martini CN, Brandani JN, Gabrielli M, etal. Effect of hexavalent chromium on proliferation and differentiation to adipocytes of 3T3-L1 fibroblasts[J]. Toxicol In Vitro, 2014 28(4):700-706.
- 32. Goulart M, Batoreu M, Rodrigues A, et al. Lipoperoxidation products and thiol antioxidants in chromium exposed workers[J]. Mutagenesis, 2005, 20 (5): 311-315.
- **FM**, Batoreu M, Rodrigues A, et al. Lippeproxidation products and thiol anto M, Batoreu M, Rodrigues A, et al. Lippeproxidation products and thiol antio and DNA damage in chromium exposed population products and thin expo 33. Khan FH, Ambreen K, Fatima G, et al. Assessment of health risks with reference to oxidative stress and DNA damage in chromium exposed population[J]. Science of the Total Environment, 2012, 430 (2): 68-74.
- 34. Sumner ER, Shanmuganathan A, Sideri TC, et al. Oxidative protein damage causes chromium toxicity in yeast[J]. Microbiology, 2005, 151: 1939-1948.
- 35. Steenken S, Jovanovic SV, Bietti M, et al. The trap depth (in DNA) of 8-oxo-7, 8-dihydro-2' deoxyguanosine as derived from electron-transfer equilibria in aqueous solution[J]. Journal of the American Chemical Society, 2000, 122 (10): 2373-2374.
- 36. Wang T, Jia G , Zhang J, etal. Renal Impairment Caused by Chronic Occupational Chromate Exposure[J]. Int Arch Occ Env Hea, 2011, 84 (4):393–401.
- 37. Urbano AM, Ferreira LM, Alpoim MC. Molecular and cellular mechanisms of hexavalent chromium-induced lung cancer: an updated perspective[J]. Curr Drug Metab, 2012, 13 (3): 284-305.
- 38. de Lemos CT, Rödel PM, Terra NR, et al. Evaluation of basal micronucleus frequency and hexavalent chromium effects in fish erythrocytes[J]. Environmental toxicology and chemistry, 2001, 20 (6): 1320-1324.
- 39. Reynolds M, Armknecht S, Johnston T, et al. Undetectable role of oxidative DNA damage in cell cycle, cytotoxic and clastogenic effects of Cr(VI) in human lung cells with restored ascorbate levels[J]. Mutagenesis, 2012, 27 (4): 437-443.
- 40. Bagchi D, Stohs SJ, Downs BW, et al. Cytotoxicity and oxidative mechanisms of different forms of chromium[J]. Toxicology, 2002, 180 (1): 5-22.
- 41. The HUMN and HUMNxL international collaboration projects on human micronucleus assays in lymphocytes and buccal cells—past, present and future. Mutagenesis, 2011, 10 (1): 239–245.
- 42. Vasylkiv OY, Kubrak OI, Storey KB, et al. Cytotoxicity of chromium ions may be connected with induction of oxidative stress[J]. Chemosphere, 2010, 80 (9): 1044-1049.
- 43. Arakawa H, Tang MS. Recognition and incision of Cr(III) ligand-conjugated DNA adducts by the nucleotide excision repair proteins UvrABC: importance of the Cr(III)-purine moiety in the enzymatic reaction[J]. Chem Res Toxicol, 2008, 21 (6): 1284-1289.



 **Figure1 The concentration of CrA and CrB in chromate exposed group and control group** 



 **Figure 2 Correlation between MNCC and MNC with the concentration of 8-OHdG in urine in exposed group**



**Figure3 The concentration between urinary 8-OHdG and CrB, a means the Curve fitting between urinary 8-OHdG and CrB in chromate exposed group, b means the concentration between urinary 8-OHdG and CrB in lower chromate exposed group(CrB<10.50), c means the concentration between urinary 8-OHdG and CrB in higher chromate exposed group(CrB≥10.50).** 



**Figure4 The concentration between LnMNC or LnMNCC and CrB, a means the Curve fitting between LnMNC and CrB in chromate exposed group, b means the Curve fitting between LnMNCC and CrB in chromate exposed group, c means the concentration between LnMNC or LnMNCC and CrB in lower chromate exposed group(CrB<9.10), d means the concentration between LnMNC or LnMNCC and CrB in higher chromate exposed group(CrB≥9.10).**

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# **A cross-sectional study: the suitable condition of 8-OHdG and micronucleus as genotoxic biomarkers in chromate exposed workers**

![](_page_19_Picture_108.jpeg)

# **A cross-sectional study: the suitable condition of 8-OHdG and micronucleus as genotoxic biomarkers in chromate exposed workers**

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**Key terms:** Chromate; Genotoxic; 8-hydroxy-2'-deoxyguanosine; Micronucleus; Concentration of chromate in the blood.

# **Abstract**

**Objectives:** We aimed to investigate the suitable condition of 8-hydroxy-2'-deoxyguanosine (8-OHdG) and micronucleus (MN) as genetoxic biomarkers at different levels of occupational chromate exposure.

**Design**: A cross-sectional study was used.

Participants: 84 workers who exposed chromate at least one year were chosen as exposed group, while 30 non-exposed individuals were used as controls.

**Main outcome measures:** Environmental and biological exposure of chromate was assessed respectively by measuring the concentration of chromate in the air (CrA) and blood (CrB) by ICP-MS in all participants. CBMN including MNCC, MNC, NPB and NBUD were calculated, while the urinary 8-OHdG was measured by ELISA method and normalized by the concentration of Cre.

**ants:** 84 workers who exposed chromate at least one year were c<br>group, while 30 non-exposed individuals were used as controls.<br>**Ecome measures:** Environmental and biological exposure of chroma<br>respectively by measuring th **Results:** Compared with the control group, the levels of CrA, CrB, MNCC, MNC and 8-OHdG in chromate exposed group were all significantly higher (*P*<0.05). There was a positive correlation between log(8-OHdG) and LnMNCC or LnMNC (*r*=0.377 and  $r=0.362$ ). The levels of LnMNCC, LnMNC and Log  $(8\text{-}OHdG)$  all have parabola correlation with the concentration of CrB. But there was a significantly positive correlation between Log (8-OHdG) and CrB when CrB level was below 10.50 ug/L (r=0.355), while a positive correlation was also found between LnMNCC or LnMNC with CrB when CrB level was lower than  $9.10\mu\text{g/L}$  (r=0.365 and r=0.269) respectively).

**Conclusions:** The MN and 8-OHdG can be used as the genotoxic biomarkers in chromate exposed group, but only when CrB levels were lower than 9.10ug/L and 10.50ug/L respectively, they can accurately reflect the degree of genetic damage.

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# *Strengths and limitations of this study (Article summary)*

 Strengths: All our results had provided new insight that only when the concentration of CrB was lower than 9.10ug/L and 10.45ug/L respectively, the MN and 8-OHdG can be used as the effective biomarkers to show the degree of genetic damage in chromate exposed group. Otherwise, when the concentration of CrB was above these levels, the cytotoxic effect might play an important role and the cells with serious genetic damage may turn into apoptosis or necrosis, consequently which could lead to the false appearance of lower degree of MN and 8-OHdG.

**Frameworth Constrainer School Sc**  Limitations: the sample size of this study was not very large, especially the control group, so we chose some references to give the value of MN and 8-OHdG in normal population to reduce the generation of bias. And it is necessary that more sample size epidemiological surveys in different chromate producing factories should be chosen to verify our conclusion.

# **Abbreviation:**

Cytokinesis-block micronucleus test-----CBMN Micronuclei cell count ------MNCC Micronucleus count -------MNC Nuclear bridge------NPB Nuclear bud --------NBUD Creatinine------Cre

# *Introduction*

Chromate is a widely used chemical in industrial and agricultural production in china, which could generate many pollutants including the waste water, gas and residue in its production, usage, transportation and storage. Previous studies had proved that long-term chromate exposure in occupational workplace could affect the health status of workers even cancer, so it has been declared as a well-known environmental and occupational hazards<sup>1</sup>.

at long term embridate exposite in occupational incripate codia a<br>tatus of workers even cancer, so it has been declared as a we<br>ental and occupational hazards<sup>1</sup>.<br>For are many different valences of chromate, in which the h There are many different valences of chromate, in which the hexavalent chromate (Cr-VI) is the most harmful one. Cr-VI can enter human body mainly by inhalation during occupational activities. When entering into respiratory system (nose, bronchial and lung), some Cr-VI could accumulate in bronco-alveolar lining fluid, mucosa and pulmonary tissues<sup>1,2</sup> and then cross the cell membrane through non-specific phosphate/ sulfate anionic transporters to the blood. This transformation also can consequently form many reactive intermediates (Cr-V, Cr-IV and Cr-III) and reactive oxygen species (ROS) with oxidative stress<sup>3,4</sup>. Both Cr-III and ROS could contribute to interact with various biological macromolecule such as Cr-DNA adducts, Cr-protein adducts, and protein-Cr-DNA adducts. This can cause damage on DNA and chromosome including base modification, single-strand breaks and double-strand breaks. These changes can result in genetic damage and ultimate carcinogenesis if accumulated to some degree<sup>5, 6, 7</sup>.

Based on the evidence above, many studies have proved that 8-OHdG can be used as a biomarker to assess the oxidative damage and DNA mutations that are induced by the ROS in clinical<sup>8</sup>, environmental<sup>9,10</sup> and occupational setting in vitro $11,12$ or in cell cultures<sup>13</sup>. MN in peripheral blood was another biomarker to show the genetic damage. It originates from chromosome fragments that are attacked by certain physical and chemical factors such as chromate or whole chromosomes that lag behind at anaphase during nuclear division. When the excision repairable DNA lesions induced in G0/G1 phase, they can be converted to MN by using inhibitors of

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the gap filling step of excision repairmen, so that unfilled gaps are converted to double strand breaks after S phase<sup>14</sup>, so the frequency of MN can be used to reflect the genetic damage<sup>15</sup>. The CBMN test is a common method to detect the MN frequencies including many indexes such as MNCC, MNC, NPB and NBUD. In these indexes, MNCC and MNC were commonly used to detect the DNA damage $^{14}$ .

Previous studies have discussed the relationship between CrB and urinary 8-OHdG or MN in chromate exposed group to investigate the feasibility of 8-OHdG and MN as genetic damage biomarkers. However, the conclusion about this connection have yielded conflicting results: Kuo found the linear correlation between urinary 8-OHdG and  $CrB<sup>16</sup>$ , but Gao, Kim and Zhang all didn't confirm it subsequently<sup>17,18,19</sup>, some research had been identified there was some association between CrB and MN frequencies<sup>20,21,22</sup>, but the suitable conditions and limitations of 8-OHdG and MN as genotoxic biomarkers for occupational chromate exposure have still been unclear So our researches aimed to observe the effect of chromate exposure on genetic damage in occupational workers, especially urinary 8-OHdG for the oxidative DNA damage and MN for chromosome damage. Then we discussed whether MN frequency and 8-OHdG can be used as the effective genotoxic biomarkers at different levels of chromate exposure.

# *Materials and methods*

#### *Study design and population*

A cross-sectional survey was designed for this research. The factory was chosen as the work place in Henan province in china because (1) the product--potassium dichromate was relatively simple, most of which was water-soluble hexavalent chromate. (2) Annual health check-ups were offered in this factory for workers, which allowed us to easily collect specimens to minimize the interference with normal work schedules.

the was relatively simple, most of which was water-soluble he interval and the check-ups were offered in this factory for worke as to easily collect specimens to minimize the interference with non s.<br>
So review only collec In this research, 84 workers exposed to chromate in the factory were chosen as exposed group, while 30 non-exposed individuals working in the administration office were as control group. The criteria of subjects included: (1) workers in exposed group were at least one year employment and 3 months working in the same work position. (2) aged between 25-50. (3) no medical history of allergy, asthma or allergic rhinitis, (4) all subjects with skin infections, fever or other clinical diseases should be excluded during the sampling period. (5) Pregnant and nursing women were not enrolled. .

All subjects were in the same factory with similar education and social backgrounds. They all were requested to complete a questionnaire and had a clinical examination before sample collection. The questionnaire included a lot of information such as occupational history, personal medical history, medication used in 4 weeks before the study, body weight and height, hair dye, house decoration, radiation exposure, individual protection, smoking status and alcohol intake.

#### *Ethical consideration*

This research was approved by Medical Ethics Committee of Peking University, Health Science Center (HSC), Beijing, China. Written informed consent from each study subject was also obtained.

#### *Air and Biological exposure assessment*

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According to the sampling criterion in monitoring of hazardous substances in the air  $(GBZ \ 159-2004)^{23}$ , six air sampling sections were respectively chosen in the workplaces of two groups, 10 sampling points were chosen in each section. The sampling process was used pumping at 1L/min for 8 hours (Sp730, TSI Corporation, USA), the membranes used in this study were MCE mixed cellulose ester filters (Φ37mm, pall, America). The average concentration of all sampling points on the membranes in the same group was measured by atomic absorption spectrometry<sup>24</sup> and then calculated to evaluate the CrA during the whole production process; the detection limit for the CrA was  $0.001 \mu g/L$ .

ies in the same group was measured by atomic absorption spectroms<br>ulated to evaluate the CrA during the whole production process; the<br>the CrA was 0.001 µg/L.<br>anticoagulant (EDTA and heparin) blood in peripheral was drew fi 4ml anticoagulant (EDTA and heparin) blood in peripheral was drew from each subject after finishing the questionnaire and then assigned to these two tubes on average. They were respectively used to measure the CrB and CBMN. The concentration of CrB was measured by inductively coupled plasma mass spectrometer  $(ICP-MS)^{24}$ , the detection limits was 0.0012  $\mu$ g/L.

At the end of the work-shift, 30mL urine sample of each subject was collected into a 50 mL metal-free polypropylene centrifuge tube (Falcon, BD Biosciences) and stored at -80℃ until used

The tubes used above had been detected their background value before research to ensure less elements and heavy metals contamination.

#### *Determination of urinary 8-OHdG*

According to the manufacturer's instructions, urine samples were firstly centrifuged at 1500 rpm for 7 min, secondly the supernatant was chosen to determine the content of urinary 8-OHdG using ELISA kit (USA Cayman chem, USA Cayman chem, 8-OHdG EIA kit) by Multiskan MK3 (Thermo, USA).The concentration of Cre in urine was determinate by alkaline picric acid assay with a commercial kit (Ausbio Laboratories Co., Ltd. China) using a Hitachi 7170A automatic analyzer (Hitachi Corp, Japan). The results of 8-OHdG were regulated by Cre to avoid the potential interference of different urine density among the subjects.

### *CBMN test*

 Peripheral venous blood which was designed in heparin tubes was taken to measure the MN frequency. The indexes (MNCC, MNC, NPB and NBUD) were counted in 1000 binuclear lymphocytes of each individual according to Fenech's  $\text{protocol}^{25}$ . All scoring was carried out by two independent researchers through double - blind method. If the scoring difference from these two researchers was less than 20 percents, the average was calculated as the final result. If the scoring difference from these two researchers was more than 20 percents, another researcher should be asked to verify the scores, then the average were used after removing the most different value.

### *Statistical analysis*

 Epidata 3.0 software was used to entry the questionnaire and experimental data into computer. The whole process was utilized double-entry and logistical error check to ensure the accuracy.

**For performal and 20** percents, another researcher should b<br>the scores, then the average were used after removing the most different<br>and 3.0 software was used to entry the questionnaire and experiment<br>that 3.0 software w All analysis was performed with SPSS16.0. Normality was assessed by Kolmogorov-Smirnov (*K-S*) test, the variables including 8-OHdG, MNCC and MNC did not meet the normality, so Log or Ln transformation was made for normality approximation. Continuous and categorical parameters between chromate exposed group and control group were tested using the two sample *t* test (or Mann-Whitney U nonparametric test) and  $\chi^2$  test. Curving correlation and linear regression were performed. Statistical significance was for two-sided. The *P* values were defined as *α*<0.05.

# *Results*

#### *General information analysis*

 A total of 114 subjects including 84 chromate exposed workers (mainly in form of  $K_2Cr_2O_7$ ) and 30 controls were recruited in this study. The working age of chromate exposed group was  $(7.82 \pm 5.51)$  years. Furthermore, the personal protection (gloves and masks) of workers was above 90%. The demographic characteristics of all subjects in this research were presented in Table 1, which showed that there were no significant differences in the distribution of gender, age, smoking and alcohol consumption between these two groups.

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Table 1 General information of chromate exposed group and control group											
Indexes	Group	<b>Exposed</b> group $(n=84)$	Control group $(n=30)$	$\overline{T}$	$\chi^2$	$\cal P$					
Age	$\overline{X} \pm S$	$35.73 \pm 7.85$	$34.83 \pm 8.83$	0.432		0.666					
$\leq$ 35	$n$ (%)	40 (47.62)	17(56.67)		0.187	0.493					
$>35$		44 (52.38)	13 (43.33)								
Gender	$n$ (%)										
Male		62(73.81)	19(63.33)		0.869	0.351					
Female		22(26.19)	11(36.67)								
Smoke	$n$ (%)										
Yes		30 (35.71)	8(26.67)			0.435					
N <sub>o</sub>		54 (64.29)	22(73.33)		0.610						
Alcohol	$n$ (%)										
Yes		29 (34.52)	16(53.33)								
No		55 (65.48)	9(46.67)		2.530	0.092					
CrA	$\overline{X} \pm S$	$15.45 \pm 19.00$	$0.23 \pm 0.38$	6.963		< 0.001					
CrB	$\overline{X} \pm S$	$9.45 \pm 9.47$	$4.05 \pm 1.87$	3.215		0.018					
8-OHdG											
	$\overline{X} \pm \mathbf{S}$	43.76±34.89	$27.21 \pm 13.76$	3.354		< 0.001					
$(ug/g$ Cre)											
MNCC $(\%_0)$	M(Q)	6.00(4.00)	3.20(2.10)	2.420		0.004					
MNC $(\%_0)$	M(Q)	7.40 (4.47)	3.74(2.94)	3.401		0.001					
NBUD $(\%_0)$	M(Q)	1.11(1.20)	1.16(1.17)	0.163		0.871					
$NPB$ $(\%_0)$	M(Q)	1.28(1.15)	1.42(1.34)	0.476		0.635					

**Table 1 General information of chromate exposed group and control group** 

 Note: Smoking referred to suck at least one cigar per day and last one year or more, smoking quit but ess than one year was also included. Alcohol was definitive by weekly drinking no less than three times

#### *Concentration of CrA and CrB*

As was shown in Table 1, the concentration of CrA in chromate exposed group [ $(15.45 \pm 19.00)$  µg/m<sup>3</sup>] was much higher than that in control group [ $(0.23 \pm 0.38)$ ]  $\mu$ g/m<sup>3</sup>](*P*<0.001), but still under the exposure limitation of chromate [ $(50\mu g/m^3)$  (2012, ACGIH)]. The levels of CrB in chromate exposed group [ $(9.45 \pm 1)$ 9.47)  $\mu$ g/L] were also significantly higher than that in control group [(4.05  $\pm$  1.87)  $\mu$ g/L].

#### *Levels of urinary 8-OHdG, and serum CBMN indexes*

The data distribution of indexes such as 8-OHdG, MNCC, MNC, NBUD and NPB was shown in Table 1, the levels of 8-OHdG, MNCC and MNC were all significantly higher in chromate exposed group than that in control group (*P*<0.05), which showed that 8-OHdG, MNCC and MNC could be used as the genetic damage biomarkers caused by chromate exposure.

#### *Correlation*

 As two biomarkers for genetic damage, there was a positive correlation between log (urinary 8-OHdG) and LnMNCC or LnMNC (*r*=0.377 and *r*=0.362 respectively) *P*<0.05)(Figure 1).

**For also Equilibrium CEMP** in the ends of CEM in anomate exposed group [(4.0<br> **EVALUATE: FOR and serum CBMN indexes**<br>
and distribution of indexes such as 8-OHdG, MNCC, MNC, NBUD<br>
or in Table 1, the levels of 8-OHdG, MN Correlation was also analyzed between the concentration of urinary 8-OHdG and CrB. As was recorded in Figure 2 and 3, there was no linear correlation but a curve fitting between CrB and Log (8-OHdG), the value of 8-OHdG was decreased when the concentration of CrB was more than 10.50µg/L. Based on the results above, the concentration of CrB was stratified into two groups: the high exposed group (CrB  $\geq 10.50\mu g/L$ ) and the low exposed group (CrB <10.50 $\mu g/L$ ). A positive correlation was shown between CrB and log (8-OHdG) when the CrB Level was lower than10.50 $\mu$ g/L (r=0.355, *P*<0.05), while there was a negative correlation between CrB and log (8-OHdG) when the CrB Level was higher than10.50µg/L.

The relationship between the concentration of MNCC or MNC and CrB was also analyzed in this research (Figure 4 and 5). There were no linear correlation but a

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curve fitting between CrB and LnMNCC or LnMNC. The value of MNCC or MNC was decreased when the concentration of CrB was more than 9.10µg/L. Based on the results above, the concentration of CrB was stratified into two groups: the high exposed group (CrB  $\geq 9.10 \mu g/L$ ) and the low exposed group (CrB  $\leq 9.10 \mu g/L$ ). A positive correlation was shown betweenCrB and LnMNCC or LnMNC in higher chromate exposed group  $(r=0.365$  and  $r=0.269$  respectively,  $P<0.05$ ), while a significantly negative relationship was found between CrB and LnMNCC or LnMNC in the lower chromate exposed group. $(r=-0.279$  and  $r=-0.261$  respectively,  $P<0.05$ .

# *Discussion*

ntly negative relationship was found between CrB and LnMNCC or<br>ver chromate exposed group ( $r=0.279$  and  $r=0.261$  respectively,  $P_5$ <br>**For permision** and low level chromate exposure can not only increase the<br>load but als Long-term and low level chromate exposure can not only increase the body's internal load but also cause a variety of harmful effects on workers' health even increasing the incidence of human cancer  $^{26, 27}$ . In occupational activity, chromate could enter into workers' body mainly by respiratory system, then be metabolism and excreted by urine, so our previous researches have proved the concentration of chromate in whole blood and urine can be used as the indicators to assess chromate biological exposure<sup>6, 28</sup>. In this study, it was found that the concentrations of CrA and CrB in exposed group were all significantly higher than that in control group (*P*<0.05) ,while the CrB level in chromate exposed group was nine times more than that in the general population of our country  $(1.19 \mu g/L)^{29}$ , which showed that the conclusion was credible by contrasting the genetic damage indexes between chromate exposed group and control group.

As we all known when Cr-VI enters human blood, it will be converted into other valence chromate compounds such as Cr-III, which could not only produce a large amount of ROS that can cause the redox system imbalance, but also directly or indirectly coordinate with DNA or protein. The transformation above can affect genetic stability including oxidative DNA lesion, DNA cross-links, single and double strand breaks and so  $\text{on}^{30,31,32}$ . Besides, long-term chromate exposure can also cause

cyto-toxicity to lead cells apoptosis.

way changes. The concentration of urinary 8-OHdG was not signtly when the level of CrB was more than 10.50µg/L, which showed tion of urinary 8-OHdG would fail to predict the degree of DNA CrB was higher than some degree. T Urinary 8-OHdG has been demonstrated as a biomarker for oxidative DNA damage in chromate exposure not only by animal experiment but also by many epidemiological researches<sup>34, 35</sup>, because it is the site that ROS often attacks, but the dose-response relationship with occupational exposure indicators was not depicted clearly. Overall, in this research, the relationship between urinary 8-OHdG and CrB was two-way changes. The concentration of urinary 8-OHdG was not significantly increasing when the level of CrB was more than  $10.50\mu g/L$ , which showed that the concentration of urinary 8-OHdG would fail to predict the degree of DNA damage when the CrB was higher than some degree. There are some reasons for this result, firstly Sumner E.R has proved that oxidative DNA damage by chromate exposure mainly targets on specifically certain glycol tic enzymes on -OH $dG<sup>36</sup>$ , which means once higher burden of CrB was produced, it could oxidize and change the structure of glycolytic enzymes to reduce the production of free 8-OHdG. secondly, some researchers have proved that 8-OHdG was not the final product of redox reaction, so when the level of CrB was higher, 8-OHdG could be converted to these further oxidation products such as Spiroiminodihydantoin  $(Sp)^{37}$ , so the concentration of free 8-OHdG was reduced. Thirdly, as we all know, the kidney can be damaged by chronic chromate exposure. As the level of CrB increased, the kidney was at greater health risk, which ultimately affect the exertion of 8-OHdG in urine<sup>38</sup>.

Micronuclei is another biomarker which commonly used as the genetic damage of chromate exposure $39.40$ . Many researches showed that chromate exposure could cause the increasing of MN frequency in cell research, animals experiment, and human being<sup>41,42</sup>. In this research, the similar conclusion was proved that the MN frequency was significantly higher in chromate exposed group than that in control group and the general population<sup>43, 44</sup> $(P< 0.05)$ . The frequencies of MNCC and MNC (as the sensitive indexes of MN) have statistical differences between these two groups. However, the dose-response relationship with CrB still need many further investigations, so this research was meaningful to discuss the suitable condition of MN as genetic damage. In this study, no linear correlation was shown between CrB

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and LnMNCC or LnMNC, the MN frequency did not increase as the CrB elevated. When all subjects were divided into two subgroups by the concentration of CrB, a positive correlation was shown between CrB and LnMNCC and LnMNC in high CrB group( $CrB\geq 9.10\mu g/L$ ) ( $r=0.365$  and  $r=0.269$  respectively), while a significantly inverse relationship was found between CrB with LnMNCC and LnMNC in low CrB group( $CrB < 9.10 \mu g/L$ ) ( $r = -0.279$  and  $r = -0.261$  respectively). The reason for these results may be that the occupational chromate exposure can increase the frequency of the MN in some range. As the increasing of CrB, the cytotoxic effect might play an important role and the cells with serious genetic damage may turn into apoptosis or  $MN^{45,46}$ .

Both MN and urinary 8-OHdG can be used to predict different types of DNA damage in some range. A positive correlation between urinary 8-OHdG and MN was found, which suggested that these two indicators as genetic damage biomarker can be verified each other.

ay be that the occupational chromate exposure can increase the free<br>in some range. As the increasing of CrB, the cytotoxic effect migh<br>t role and the cells with serious genetic damage may turn into apc<br>MN and urinary 8-OHd There was still some limitation in this study: the sample size of this study was not very large, especially the control group, so we chose some references<sup>38,43,44</sup> to give the value of 8-OHdG and MN in normal population to reduce the generation of bias. And it is necessary that more sample size epidemiological surveys in different chromate producing factories should be chosen to verify our conclusion.

# *Conclusions*

All these above had provided new insights that both MN and urinary 8-OHdG can be used as the genetic damage biomarkers caused by occupational chromate exposure at some levels. The combination of these indicators can improve the credibility of the results. It is not the simple linear relationship between their concentration and the level of chromate exposure. Only when the level of CrB is below 9.10µg/L and 10.50µg/L, the MN frequency and urinary 8-OHdG can respectively show the degree of genetic damage quantitatively.

# *FOOTNOTES:*

# *Contributor ship statement*

 All the authors included in the paper granted the criteria of the authorship. PL together with YG conceived and executed this investigation, analyzed of the data and described this manuscript. YL and JLY supported this investigation from the epidemiological aspect. GJ is responsible for the whole conduct of this study and all content of this. SFY contribute to the organization and arrangement for the scene investigation. All authors commented critically on the manuscript and agreed with this submitting.

# *Competing interests*

There are none competing interests.

# *Funding*

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### *Data sharing*

No additional data available

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# **Reference**

- 1. Monika G, Joachim E, Peter B, et al. Biological effect markers in exhaled breath condensate and biomonitoring in welders: impact of smoking and protection equipment [J]. Int Arch Occup Environ Health, 2010, 83 (7): 803–811.
- 2. Andrea C, Matteo G, Olga A, et al. The effect of inhaled chromium on different exhaled breath condensate biomarkers among chrome-plating workers [J]. Environmental Health Perspectives, 2006, 114 (4): 542-546.
- 3. Shrivastava R, Upreti RK, Seth PK, et al. Effects of chromium on the immune system [J]. Immunology and Medical Microbiology, 2002, 34 (4): 1-7.
- 4. Shashwat K, Awasthi SK, Srivastava JK. Effect of chromium on the level of IL-12 and IFN-γ in occupationally exposed workers [J]. Science of the total environment, 2009, 407 (6): 1868-1874.
- and the persists and the control of the entired of the persists of the persist of the same of the persist of the A Seth PK, et al. Effects of chromium on the immune system at K, Awasthi SK, Stivastava JK. Effect of chromiu 5. Mohammad BH, Marie V, Gabriela C, et al. Low level environmental cadmium exposure is associated with DNA hypomethylation in argentinean women [J]. Environmental Health Perspectives, 2012, 120 (6): 879–884.
- 6.Xiaohua L, Yanshuang S, Li W, et al. Evaluation of the correction between genetic damage and occupational chromate exposure through BMMN frequenciess[J]. J Occup Environ Med, 2012, 54(2):166-170.
- 7. De Flora S, Bagnasco M, Serra D, et al. Genotoxicity of chromium compounds: A review[J]. Mutation Research/Reviews in Genetic Toxicology, 1990, 238 (5): 99-172.
- 8. Wu LL, Chiou CC, Chang PY, et al. Urinary 8-OHdG: a marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetics[J]. Clinica Chimica Acta, 2004, 339 (1-2 ): 1-9.
- 9. Ren C, Fang S, Wright RO, et al. Urinary 8-hydroxy-2-deoxyguanosine as a biomarker of oxidative DNA damage induced by ambient pollution in the Normative Aging Study[J]. Occupational and environmental medicine, 2011, 68 (2): 562-569.
- 10. Lin CY, Lee HL, Chen YC, et al. Positive association between urinary levels of 8-hydroxydeoxyguanosine and the acrylamide metabolite N-acetyl-S-(propionamide)-cysteine in adolescents and young adults[J]. J Hazard Mater, 2013, 261: 372-377.
- 11. Kim JY, Mukherjee S, Ngo LC, et al. Urinary 8-hydroxy-2'-deoxyguanosine as a biomarker of oxidative DNA damage in workers exposed to fine particulates[J]. Environmental health perspectives, 2004, 112 (6): 666-671.
- 12. Valavanidis A, Loridas S, Vlahogianni T, et al. Influence of ozone on traffic-related particulate matter on the generation of hydroxyl radicals through a heterogeneous synergistic effect [J]. Hazard Mater. 2009, 162 (2-3): 886-892.
- 13. Valavanidis A, Vlachogianni T, Fiotakis C. 8-hydroxy-2-deoxyguanosine (8-OHdG): a critical biomarker of oxidative stress and carcinogenesis[J]. Journal of environmental science and health Part C, 2009, 27 (2): 120-139.
- 14. Fenech M, Kirsch-Volders1 M, Natarajan AT, et al. Molecular mechanisms of micronucleus, nucleoplasmic bridge and nuclear bud formation in mammalian and human cells. Mutagenesis, 2011, 26: 125–132.
- 15. Sobol Z, Schiestl RH. Intracellular and extracellular factors influencing Cr (VI) and Cr (III) genotoxicity[J]. Environmental and Molecular Mutagenesis, 2012, 53 (2): 94-100.
- 16. Kuo H, Chang S, Wu K, et al. Chromium (VI) induced oxidative damage to DNA: increase of urinary 8-hydroxydeoxyguanosine concentrations (8-OHdG) among electroplating workers[J]. Occupational and environmental medicine, 2003, 60 (8): 590-594.
- 17.Kim H, Cho S-H, Chung M-H. Exposure to hexavalent chromium does not increase 8-hydroxydeoxyguanosine levels in Korean chromate pigment workers[J]. Industrial health, 1999, 37 (3): 335-341.
- 18. Gao M, Levy LS, Faux SP, et al. Use of molecular epidemiological techniques in a pilot study

 on workers exposed to chromium[J]. Occupational and environmental medicine, 1994, 51  $(10): 663-668.$ 

- 19. Zhang XH, Zhang X, Wang XC, et al. Chronic occupational exposure to hexavalent chromium causes DNA damage in electroplating workers.[J] BMC Public Health, 2011, 11 (3): 224.
- 20. Xiaohua L, Yanshuang S, Li W, et al. Evaluation of the correction between genetic damage and occupational chromate exposure through BMMN frequencies. J Occup Environ Med 2012, 54 (2): 166-170.
- 21. Danadevi K, Rozati R, Banu BS, et al. Genotoxic evaluation of welders occupationally exposed to chromium and nickel using the Comet and micronucleus assays[J]. Mutagenesis, 2004, 19 (1): 35-41.
- 22. Thompson CM, Fedorov Y, Brown DD, et al. Assessment of Cr (VI)-induced cytotoxicity and genotoxicity using high content analysis[J]. PloS one, 2012, 7 (8): 742-720.
- 23. GBZ 159-2004. Specifications of air sampling for hazardous substances monitoring in the workplace. China.
- 24. Yanshuang Song, Ji Zhang, Shanfa Yu, et al. Effects of chronic chromium (VI) exposure on blood element homeostasis: An epidemiological study. Metallomics, 2012, 4: 463–472
- 25. Fenech M. Cytokinesis-block micronucleus cytome assay[J]. Nature protocols, 2007, 2 (1): 1084-1104.
- 26. Clancy HA, Sun H, Passantino L, et al. Gene expression changes in human lung cells exposed to arsenic, chromium, nickel or vanadium indicate the first steps in cancer[J]. Metallomics. 2012, 4(8):784-793.
- 27.Chhabra D, Oda K, Jagannath P, et al. Chronic heavy metal exposure and glbladder cancer risk in India, a comparative study with Japan[J]. Asian Pacific Journal of Cancer Prevention. 2012, 13(1): 187-190.
- 28. Yan L, Jia G, Zhang J, et al. The correlation between personal occupational exposure to soluble chromate and urinary chromium content. Zhonghua yu Fang yi xue za zhi (Chinese Journal of Preventive Medicine) 2006, 40(6): 386-389 (Article in Chinese).
- 29. Ding CG, Pan Yj, Zhang AH, et al. Distribution of chromium in whole blood and urine among general population in China between year 2009 and 2010. Zhonghua yu Fang yi xue za zhi (Chinese Journal of Preventive Medicine) 2012, 46(8): 679-682. ( Article in Chinese).
- 30.Zhitkovich, A. Importance of chromium-DNA adducts in mutagenicity and toxicity of chromium(VI)[J]. Chem. Res. Toxicol., 2005, 18 (1): 3-11.
- Solvicy, Frotow P, Jowin DJ, et al. Nassessment of CT (*P* Junitate Vyotox<br>Solvicy using high content analysis[J]. PloS one, 2012, 7 (8): 742-720,<br>69-2004. Specifications of air sampling for hazardous substances monitoring 31. Wang TC, Song YS, Wang H, et al. Oxidative DNA damage and global DNA hypomethylation are related to folate deficiency in chromate manufacturing workers[J]. Journal of Hazardous Materials,2012, 213.214:440-446.
- 32. Brien, T.J.; Ceryak, S.; Patierno, S.R. Complexities of chromium carcinogenesis: role of cellular response, repair and recovery mechanisms[J]. Mutat. Res. Fundam. Mol. Mech. Mutagen., 2003, 533 (14): 3-36.
- 33. Martini CN, Brandani JN, Gabrielli M, et al. Effect of hexavalent chromium on proliferation and differentiation to adipocytes of 3T3-L1 fibroblasts[J]. Toxicol In Vitro, 2014 28(4):700-706.
- 34. Goulart M, Batoreu M, Rodrigues A, et al. Lipoperoxidation products and thiol antioxidants in chromium exposed workers[J]. Mutagenesis, 2005, 20 (5): 311-315.
- 35. Khan FH, Ambreen K, Fatima G, et al. Assessment of health risks with reference to oxidative stress and DNA damage in chromium exposed population[J]. Science of the Total Environment, 2012, 430 (2): 68-74.
- 36. Sumner ER, Shanmuganathan A, Sideri TC, et al. Oxidative protein damage causes chromium toxicity in yeast[J]. Microbiology, 2005, 151: 1939-1948.
- 37. Steenken S, Jovanovic SV, Bietti M, et al. The trap depth (in DNA) of 8-oxo-7, 8-dihydro-2' deoxyguanosine as derived from electron-transfer equilibria in aqueous solution[J]. Journal of the American Chemical Society, 2000, 122 (10): 2373-2374.
- 38. Wang T, Jia G , Zhang J, etal. Renal Impairment Caused by Chronic Occupational Chromate Exposure[J]. Int Arch Occ Env Hea, 2011, 84 (4):393–401.
- 39. Urbano AM, Ferreira LM, Alpoim MC. Molecular and cellular mechanisms of hexavalent chromium-induced lung cancer: an updated perspective[J]. Curr Drug Metab, 2012, 13 (3): 284-305.
- 40. de Lemos CT, Rödel PM, Terra NR, et al. Evaluation of basal micronucleus frequency and hexavalent chromium effects in fish erythrocytes[J]. Environmental toxicology and chemistry, 2001, 20 (6): 1320-1324.
- 41. Reynolds M, Armknecht S, Johnston T, et al. Undetectable role of oxidative DNA damage in cell cycle, cytotoxic and clastogenic effects of Cr(VI) in human lung cells with restored ascorbate levels[J]. Mutagenesis, 2012, 27 (4): 437-443.
- 42. Bagchi D, Stohs SJ, Downs BW, et al. Cytotoxicity and oxidative mechanisms of different forms of chromium[J]. Toxicology, 2002, 180 (1): 5-22.
- 43. The HUMN and HUMNxL international collaboration projects on human micronucleus assays in lymphocytes and buccal cells—past, present and future. Mutagenesis, 2011, 10 (1): 239–245.
- 44. Bonassi S, Fenech M, Lando C, et al. Human MicroNucleus Project: International database Comparison for results with the cytokinesis-block micronucleus assay in human lymphocytes: effect of laboratory protocol, scoring criteria, and host factors on the frequency of micronuclei. Environ Mol Mutagen.2001;37:31–45
- 45. Vasylkiv OY, Kubrak OI, Storey KB, et al. Cytotoxicity of chromium ions may be connected with induction of oxidative stress[J]. Chemosphere, 2010, 80 (9): 1044-1049.
- 46. Arakawa H, Tang MS. Recognition and incision of Cr(III) ligand-conjugated DNA adducts by the nucleotide excision repair proteins UvrABC: importance of the Cr(III)-purine moiety in the enzymatic reaction[J]. Chem Res Toxicol, 2008, 21 (6): 1284-1289.

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micronucleus as genotoxic-

 **damage biomarkers in occupational chromate exposed workers** 

**A cross-sectional study: The suitable condition of 8-OHdG and** 

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#### **Abstract**

**Objectives:** We aimed to investigate the suitable condition of 8-hydroxy-2'

-deoxyguanosine (8-OHdG) and micronucleus (MN) **as genetoxic biomarkers** at different levels of occupational chromate exposure.

**Design**: A cross-sectional study was used.

Participants: 84 workers who exposed chromate exposing chromate at least one year were chosen as exposed group, while 30 non-exposed individuals were used as controls.

**Main outcome measures:** Environmental and biological exposure of chromate was assessed respectively by measuring the concentration of chromate in the air (CrA) and blood (CrB) by ICP-MS in all **participants** subjects. CBMN was conducted, in which many indexes including MNCC, MNC, NPB and NBUD were calculated, while the urinary 8-OHdG was measured by ELISA method and normalized by the concentration of Cre.

med to investigate the suitable condition of 8-hydroxy-2'<br>
8-OHdG) and micronucleus\_(MN) <u>as genetoxic biomarkers</u> at<br>
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sexposed group, whi **Results:** Compared with the control group, the levels of CrA, CrB, MNCC, MNC and 8-OHdG in chromate exposed group were all significantly higher (*P*<0.05). There was a positive correlation between log(8-OHdG) and LnMNCC or LnMNC (*r*=0.377 and *r*=0.362 ). The levels of LnMNCC , LnMNC and Log (8-OHdG) **all** did not have the **parabola correlation** simple linear relationship with the concentration of CrB. **But There there** was a significantly positive correlation between Log (8-OHdG) and CrB when CrB level was below 10.50 ug/L  $(r=0.355)$ . W<sub>r</sub> while a positive correlation was **also** found between LnMNCC or LnMNC with CrB when CrB level was lower than 9.10 ug/ $L(r=0.365$  and  $r=0.269$  respectively).

**Conclusions:** The MN and 8-OHdG can be used as the **genotoxic** biomarkers for the genetic damage in chromate exposed group, but only when CrB levels were lower

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than 9.10ug/L and 10.50ug/L respectively, they can accurately reflect the degree of genetic damage.

Key terms: **chromateChromate**; Genotoxicetic damage; 8-hydroxy-2'**deoxyguanosine8-OHdG**; Micronucleus, Concentration of chromate in the blood

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**Abbreviation:** 

Cytokinesis-block micronucleus test-----CBMN Micronuclei cell count ------MNCC Micronucleus count -------MNC Nuclear bridge------NPB Nuclear bud --------NBUD Creatinine------Cre

#### *Strengths and limitations of this study(Article summary)*

**For peer review only** Strengths: All our results had provided new insights that only when the concentration of CrB was lower than 9.10ug/L and 10.45ug/L respectively, the MN and 8-OHdG can be used as the effective biomarkers to show the degree of genetic damage for the in chromate **occupational exposed groupure**, o. Otherwise, **when the concentration of CrB was** above these levels, the cytotoxic effects-might play an important role and the **fate of** cells with serious genetic damages may turn into apoptosis or necrosis, consequently which could lead to the false appearance of lower degree of MN and 8-OHdG-at higher chromate exposed level.

Limitations: the **sample** size of this study population was not very large, especially the **group of control individuals group**, so we chose some references to give the **normal** value of MN and 8-OHdG **in normal population in other controls**  to reduce the **mitigation generation** of bias. **In this research**, **recommended condition of 8-OHdG and MN as genetic damage biomarker in** 

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**chromate exposed group. And It it** is necessary that more **sample sizeoccupational** epidemiological surveys in different chromate producing factories should be chosen to verify **this our** conclusion. **Formatted:** Font: Bold **Formatted:** Font: Bold

### *Introduction*

**For all the set of the** Chromate is a widely used chemical in industrial and agricultural production in china, which could generate many pollutants including the waste water, waste-gas and waste residue in chromate its production, usage, transportation and storage. Previous studies had proved that Longlong-term chromate exposure in occupational workplace ean-could affect the health status of workers even cancer, so it has been declared as a well-known environmental and occupational hazards  $\frac{1}{2}$ .

There are many different valences of chromate, in which the hexavalent chromate (Cr-VI) is the most harmful one. Cr-VI can enter human body mainly by inhalation during occupational activities. When entering into respiratory system (nose, bronchial and lung), some Cr-VI could accumulate in bronco-alveolar lining fluid, mucosa and pulmonary tissues<sup>1,2</sup> and then cross the cell membrane through non-specific phosphate/ sulfate anionic transporters to the blood. be converted to Cr-V, Cr-IV and Cr-III, and accumulated in bronco-alveolar lining fluid, mucosa and pulmonary tissues<sup>1,2</sup>. This transformation also can consequently form many reactive intermediates (Cr-V, Cr-IV and Cr-III) and reactive oxygen species (ROS) with oxidative stress<sup>3,4</sup>. Both Cr-III and ROS could contribute to interact with various biological macromolecule such as Cr-DNA adducts, Cr-protein adducts, and

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protein-Cr-DNA adducts proteins, DNA. This can and other biological macromolecule to cause damage on DNA and chromosome, including base modification, single-strand breaks and double-strand breaks. These damages can result in genetic damage and ultimate carcinogenesis which can initiate carcinogenesis if accumulated to some degree<sup>5, 6, 7</sup>.

evidence above, <u>many studies have proved that</u> -8-OHdG can be<br> **Extra** Formarker of exidative DNA damage, can be used-to assess the<br>
and DNA mutations that are induced by the ROS in clinical<sup>8</sup>,<br>
and occupational setting Based on the evidence above, many studies have proved that -8-OHdG can be asused as a product-biomarker of oxidative DNA damage, can be used to assess the oxidative damage and DNA mutations that are induced by the ROS in clinical<sup>8</sup>, environmental<sup>9,10</sup> and occupational setting in vitro<sup>11,12</sup>or in cell cultures<sup>13</sup>. MN in peripheral blood was another biomarker to show the genetic damage.  $\frac{1}{\text{in}}$  peripheral blood It originates from chromosome fragments that are attacked by certain physical and chemical factors such as chromium chromate or whole chromosomes that lag behind at anaphase during nuclear division. When the excision-repairable DNA lesions induced in G0/G1 phase, they can be converted to MN by using inhibitors of the gap filling step of excision repairmen, so that unfilled gaps are converted to double strand breaks after S phase<sup>14</sup>, so the frequency of MN can be used to reflect the genetic damage<sup>15</sup>. The CBMN test is a common method to detect the MN frequencies including many indexes such as MNCC, MNC, NPB and NBUD. In these indexes, MNCC and MNC were commonly used to detect the DNA damage $^{14}$ .

Previous studies have chose discussed the relationship between CrB and urinary<sup>\*</sup> 8-OHdG or MN in to discuss the feasibility of 8-OHdG and MN as the biomarker for chromium chromate exposureexposed group to investigate the feasibility of 8-OHdG and MN as genetic damage biomarkers., howeverHowever, the conclusion about this connection have yielded conflicting results: Kuo found the linear correlation between urinary 8-OHdG and  $CrB<sup>16</sup>$ , but Gao, Kim and Zhang all didn't confirm it subsequently<sup>17,18,19</sup>, many some research studies had been identified there was some association between CrB and MN frequencies<sup>20,21,22</sup>, but the suitable conditions and limitations for of 8-OHdG and MN as genotoxic biomarkers for occupational chromate exposure have still been unclear. So our researches aimed to observe the effect of chromate exposure on genetic damage in occupational workers, especially

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urinary 8-OHdG for the oxidative DNA damage and MN for chromosome damage. Then it was we discussed whether MN frequency and 8-OHdG can be used as the effective genotoxic biomarkers at different levels of chromate exposure.

#### *Materials and methods*

#### *Study design and population*

**Formally any of the Control C** A cross-sectional survey was designed for this research. The factory was chosen as the work place in Henan province in china because (1) the product--potassium dichromate was relatively simple, most of which was water-soluble hexavalent chromate. (2) Annual health check-ups were offered in this factory for workers, which allowed us to easily collect specimens from workers to minimize the interference with normal work schedules.

In this research, 84 workers exposed to chromate in the factory were chosen as exposed group, while 30 non-exposed individuals working in the administration office were as control group. The criteria of subjects included: (1) workers in exposed group were at least one year employment and 3 months working in the same work position. (2) aged between 25-50. (3) no medical history of allergy, asthma or allergic rhinitis, (4) all subjects with skin infections, fever or other clinical diseases should be

excluded during the sampling period. (5) Pregnant and nursing women were not enrolled. .

All subjects were in the same factory with similar education and social backgrounds. They all were requested to complete a questionnaire and had a clinical examination before sample collection. The questionnaire included a lot of information including such as occupational history, personal medical history, medication used in 4 weeks before the study, body weight and height, hair dye, house decoration, radiation exposure, individual protection, smoking status and alcohol intake.

#### *Ethical consideration*

This study research was approved by Medical Ethics Committee of Peking University, Health Science Center (HSC), Beijing, China. Written informed consent from each study subject was also obtained.

#### *Air and Biological exposure assessment*

**Example 12** Theory, personal medical history, medication used in 4<br>tudy, body weight and height, hair dye, house decoration, radiation<br>all protection, smoking status and alcohol intake.<br>**For performance of Peking**<br>Scienc According to the sampling criterion in monitoring of hazardous substances in the air  $(GBZ 159-2004)<sup>23</sup>$ , Six-six air sampling sections were respectively chosen in the workplaces of two groups, 10 sampling points were chosen in each section. The sampling process was used pumping at 1L/min for 8 hours (Sp730, TSI Corporation, USA), the membranes used in this study were MCE mixed cellulose ester filters (Φ37mm, pall, America).The average concentration of all sampling points on the membranes in the same group was measured by atomic absorption spectrometry $\frac{24}{1}$  and then calculated to evaluate the CrA during the whole production process, the detection limit for the CrA was 0.001 µg/L.

2ml 4ml anticoagulant (EDTA and heparin) blood in peripheral was drew from each subject after finishing the questionnaire and then assigned to these two tubes on average. They were respectively used to measure the CrB and CBMN. The concentration of CrB was measured by inductively coupled plasma mass spectrometer  $(ICP-MS)^{\frac{2624}{}}$ . the detection limits was 0.0012  $\mu$ g/L.

The tubes used above had been detected their background value before research to ensure less elements and heavy metals contamination...

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 At the end of the work-shift, 30mL urine sample of each subject was collected into a 50 mL metal-free polypropylene centrifuge tube (Falcon, BD Biosciences) and stored at -80℃ until used to measure the contents of 8 OHdG and Cre.

The tubes used above had been detected their background value before research to ensure less elements and heavy metals contamination.

#### *Determination of urinary 8-OHdG*

According to the manufacturer's instructions, urine samples were firstly centrifuged at 1500 rpm for 7 min, secondly the supernatant was chosen to determine the content of urinary 8-OHdG using ELISA kit (USA Cayman chem, USA Cayman chem, 8-OHdG EIA kit) by Multiskan MK3 (Thermo, USA) and The concentration of Cre in urine was determinate by alkaline picric acid assay with a commercial kit (Ausbio Laboratories Co., Ltd. China) using a Hitachi 7170A automatic analyzer (Hitachi Corp, Japan). The results of 8-OHdG were regulated by Cre to avoid the potential interference of different urine density among the subjects.

#### *CBMN test*

**Example 8-OHdG** the manufacturer's instructions, urine samples were firstly of the manufacturer's instructions, urine samples were firstly of the manufacturer's instructions, urine samples were firstly and  $\lambda$  For  $\lambda$  Peripheral venous blood which was designed in heparin ized tubes with individual numbers was taken to measure the MN frequency. The indexes (MNCC, MNC, NPB and NBUD) were counted in 1000 binuclear lymphocytes of each individual according to Fenech's protocol<sup>23</sup>. All scoring was carried out by two independent researchers through double - blind method. If the scoring difference from these two researchers was less than 20 percents, the average was calculated as the final result. If the scoring difference from these two researchers was more than 20 percents, another researcher should be asked to verify the scores, then the average were used after removing the most different value.

If the result from these two researchers was different, another researcher should be asked to verify the scores.

#### *Statistical analysis*

 Epidata 3.0 software was used to entry the questionnaire and experimental data into computer. The whole process was utilized double-entry and logistical error check

to ensure the accuracy.

**For peer review only** All analysis was performed with SPSS16.0. Normality was assessed by Kolmogorov-Smirnov (*K-S*) test, the variables including 8-OHdG, MNCC and MNC did not meet the normality, so Log or Ln transformation was made for normality approximation. Continuous and categorical parameters between chromate exposed group and control group were tested using the two sample *t* test (or Mann-Whitney U nonparametric test) and  $\chi^2$  test. Curving correlation and linear regression were performed. Statistical significance was for two-sided. The *P* values were defined as *α*<0.05.

#### *Results*

#### *General information analysis*

 A total of 114 subjects including 84 chromate exposed workers (mainly in form of  $K_2Cr_2O_7$ ) and 30 controls were recruited in this study. The working age of chromate exposed group was  $(7.82 \pm 5.51)$  years. Furthermore, the personal protection (gloves and masks) of workers was above 90%. The demographic characteristics of all subjects in this research were presented in Table 1, which showed that there were no significant differences in the distribution of gender, age, smoking and alcohol consumption between these two groups.

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 Note: Smoking referred to suck at least one cigar per day and last one year or more, smoking quit but ess than one year was also included. Alcohol was definitive by weekly drinking no less than three times

#### *Concentration of CrA and CrB*

As was shown in FigureTable 1, the concentration of CrA in chromate exposed group [(15.45  $\pm$  19.00)  $\mu$ g/m<sup>3</sup>] was much higher than that in control group [(0.23  $\pm$ 0.38)  $\mu$ g/m<sup>3</sup>]( $P$ <0.001), but—still under the exposure limitation of chromate

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[ $(50\mu g/m^3)$  (2012, ACGIH)]. The levels of CrB in chromate exposed group [ $(9.45 \pm 1)$ 9.47)  $\mu$ g/L] were also significantly higher than that in control group [(4.05  $\pm$ 1.87)  $\mu$ g/L]  $\rightarrow$  (*P*<0.001).

#### *Levels of urinary 8-OHdG, and serum CBMN indexes*

TAs for the data distribution of indexes such as 8-OHdG, MNCC, MNC, NBUD and NPB was shown in Table 1<del>(Table 1)</del>, the levels of re were three kinds of indexes (8-OHdG, MNCC and MNC) . They were all significantly higher in chromate exposed group than that in control group (*P*<0.05), which showed that 8-OHdG, MNCC and MNC could be used as the genetic damage biomarkers feasibly caused by chromate exposure.

#### *Correlation*

 As two biomarkers for genetic damage, there was a positive correlation between log (urinary 8-OHdG) and LnMNCC  $\frac{and}{end}$  or LnMNC ( $r=0.377$  and  $r=0.362$ respectively)  $P<0.05$ )(Figure  $\frac{21}{21}$ ).

**For all standard Consumeration** of masses such as 8-Ornox, MNCC, MNC, NBOD<br> **For in Table 1-(Table 1)**, the <u>levels of</u> ree were three kinds of indexessed and MNC)—They were <u>all significantly</u> higher in chromate exposed Correlation was also analyzed between the concentration of urinary 8-OHdG and<sup>4</sup> CrB. As was recorded in Figure 2 and 3, there was no linear correlation but a curve fitting between CrB and Log (8-OHdG), we found the value of 8-OHdG was decreased when the concentration of CrB was more than 10.50µg/L. Based on the results above, the concentration of CrB was stratified into two groups: the high exposed group (CrB  $\geq 10.50\mu g/L$ ) and the low exposed group (CrB <10.50 $\mu g/L$ ). A positive correlation was shown between CrB and log (8-OHdG) when the CrB Level was lower than10.50 $\mu$ g/L ( $r=0.355$ ,  $P<0.05$ ), while there was a negative correlation between CrB and log (8-OHdG) when the CrB Level was higher than10.50µg/L.

The relationship between the concentration of MNCC or MNC and CrB was also analyzed in this research (Figure 4 and 5). there There were no linear correlation but a curve fitting between CrB and LnMNCC or LnMNC. We found tThe value of MNCC or MNC was decreased when the concentration of CrB was more than 9.10µg/L. Based on the results above, the concentration of CrB was stratified into two groups: the high exposed group (CrB  $\geq$ 9.10µg/L) and the low exposed group (CrB **Formatted:** Indent: First line: 1.5 ch

<9.10µg/L). A positive correlation was shown betweenCrB and LnMNCC or LnMNC in higher chromate exposed group  $(r=0.365$  and  $r=0.269$  respectively,  $P<0.05$ ), while a significantly negative relationship was found between CrB and LnMNCC or LnMNC in the lower chromate exposed group. $(r=0.279$  and  $r=0.261$  respectively, *P*<0.05).

# *Discussion*

Long-term and low level chromate exposure can not only increase the body's internal load but also cause a variety of harmful effects on workers' health even increase increasing the incidence of human cancer  $^{26, 27}$ . In occupational activity, chromate could enter into workers' body mainly by respiratory system, then be metabolism and excreted by urine, so our previous researches have proved the concentration of chromium chromate in whole blood and urine can be used as the indicators to assess chromate biological exposure<sup>6, 28</sup>. In this study, it was found that the concentrations of CrA and CrB in exposed group were all significantly higher than that in control group  $(P<0.05)$ , while the CrB level in chromate exposed group was nine times more than that in the general population of our country  $(1.19 \mu g/L)^{29}$ , which showed that the conclusion was credible by contrasting the genetic damage indexes between chromate exposed group and control group.

As we all known when Cr-VI enters human blood, it will be converted into other valence chromate compounds such as Cr-III, which could not only produce a large amount of ROS that can cause the redox system imbalance, but also directly or indirectly coordinate with DNA or protein. The transformation above can affect

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genetic stability including oxidative DNA lesion, DNA cross-links, single and double strand breaks and so  $\frac{a^{28}}{9} \frac{\text{m}^{30,31,32,29,30}}{9}$ . Besides, long-term chromate exposure can also cause cyto-toxicity to lead cells **apoptosis**<sup>31</sup> apoptosis<sup>33</sup>.

**Exercularies**<sup>34</sup>**:Essearches**<sup>34</sup>**:**<sup>33135</sup>, because it is the site that ROS often se-response relationship with occupational exposure indicators was tryly. Overall, in this research, the relationship between urinary w Urinary 8-OHdG has been demonstrated as a biomarker for oxidative DNA damage in chromate exposure not only by animal experiment but also by many epidemiological researches<sup>32</sup>researches<sup>34,3335</sup>, because it is the site that ROS often attacks, but the dose-response relationship with occupational exposure indicators was not depicted clearly. Overall, in this research, the relationship between urinary 8-OHdG and CrB was two-way changes. The concentration of urinary 8-OHdG was not significantly increasing when the level of CrB  $\frac{1}{15}$  was more than 10.50 $\mu$ g/L, which showed that when the CrB was higher than some degree, the concentration of urinary 8-OHdG would fail to predict the degree of DNA damage when the CrB was higher than some degree. There are some reasons for this result, firstly Sumner E.R has proved that oxidative DNA damage by chromate exposure mainly targets on specifically certain glycol tic enzymes on  $8-\theta H dG^{34}OH dG^{36}$ , which means once higher burden of CrB was produced, it could oxidize and change the structure of glycolytic enzymes to reduce the production of free 8-OHdG. secondly, some researchers have proved that 8-OHdG was not the final product of redox reaction, so when the level of CrB is was higher, 8-OHdG could be converted to these further oxidation products such as Spiroiminodihydantoin  $(Sp)^{\frac{3537}{25}}$ , so the concentration of free 8-OHdG was reduced. Thirdly, as we all know, the kidney can be damaged by chronic chromate exposure. As the level of CrB increased, the kidney was at greater health risk, which ultimately affect the exertion of 8-OHdG in urine<sup>36</sup>urine<sup>38</sup>.

Micronuclei is another biomarker which commonly used as the genetic damage of chromate exposure  $\frac{39,4039.40}{9}$ . Many researches showed that chromate exposure could cause the increasing of MN frequency in cell research, animals experiment, and human being $44,4241,42$ . In this research, the similar conclusion was proved that the MN frequency was significantly higher in chromate exposed group than that in control group and the general  $\frac{1}{2}$  population<sup>41</sup> population<sup>43, 44</sup>(*P*<0.05). The frequency frequencies of MNCC and MNC (as the sensitive indexes of MN) has have statistical

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> on of CrB, a positive correlation was shown between CrB and<br>MNC in high CrB group(CrB29.10µg/L) ( $r=0.365$  and  $r=0.269$ <br>e a significantly inverse relationship was found between CrB with<br>MNC in low CrB group(CrB<9.10µg/L) differences between these two groups. However, the dose-response relationship with CrB still needs many further investigations, so this research was meaningful to discuss the suitable condition of MN as genetic damage. In this study, no linear correlation was shown between CrB and LnMNCC or LnMNC, the MN frequency did not increase as the CrB elevated. When all subjects were divided into two subgroups by the concentration of CrB, a positive correlation was shown between CrB and LnMNCC and LnMNC in high CrB group(CrB≥9.10 $\mu$ g/L) ( $r=0.365$  and  $r=0.269$ respectively), while a significantly inverse relationship was found between CrB with LnMNCC and LnMNC in low CrB group(CrB $\leq$ 9.10µg/L) ( $r$ =-0.279 and  $r$ =-0.261 respectively). The reason for these results may be that the occupational chromate exposure can increase the frequency of the MN in some range. As the increasing of CrB, more serious genotoxic damage should be caused the cytotoxic effect might play an important role and the cells with serious genetic damage may turn into apoptosis or necrosis, consequently which could lead to the false appearance of lower degree of MN such as the apoptosis or necrosis of lymphocyte, which contrarily decrease the frequency of MN in this situation<sup>425,4346</sup>.

 Both MN and urinary 8-OHdG can be used to predict different types of DNA damage in some range. A positive correlation between urinary 8-OHdG and MN was found, which suggested that these two indicators as genetic damage biomarker can be verified each other.

There was still some limitation in this study: the sample size of this study was not<sup>\*</sup> very large, especially the control group, so we chose some references<sup>38,43,44</sup> to give the value of 8-OHdG and MN in normal population to reduce the generation of bias. And it is necessary that more sample size epidemiological surveys in different chromate producing factories should be chosen to verify our conclusion.

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All these above had provided new insights that both MN and urinary 8-OHdG can be used as the genetic damage biomarkers caused by occupational chromate exposure at some levels—  $\frac{f}{g}$ . the The combination of these indicators can improve the credibility of the results. It is not the simple linear relationship between their concentration and the level of chromate exposure. Only when the level of CrB is below 9.10µg/L and 10.50µg/L, the MN frequency and urinary 8-OHdG can respectively show the degree of genetic damage respectively produced by chromate exposure quantitatively.

#### *Contributor ship statement*

**Conclusions**<br>
All these above had provided new insights that both MN and urinary 8-OHdG can<br>
be used as the genetic damage biomarkers caused by occupational chromate exposure<br>
at some levels<sup>-</sup><sub>-the-Ihe</sub> combination of t All the authors included in the paper granted the criteria of the authorship. PL together with YG conceived and executed this investigation, analyzed of the data and described this manuscript. YL and  $JLY$  supported this investigation from the epidemiological aspect. GJ is responsible for the whole conduct of this study and all content of this. SFY contribute to the organization and arrangement for the scene investigation. All authors commented critically on the manuscript and agreed with this submitting.

#### *Competing interests*

There are not competing interests.

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# *Data sharing*

There is not data sharing.

# **Reference**

- 1. Monika Gube, Joachim Ebel, Peter Brand, et al. Biological effect markers in exhaled breath condensate and biomonitoring in welders: impact of smoking and protection equipment [J]. Int Arch Occup Environ Health, 2010, 83 (7): 803–811.
- 2. Andrea Caglieri, Matteo Goldoni, Olga Aeampa, et al. The effect of inhaled chromium on different exhaled breath condensate biomarkers among chrome-plating workers [J]. Environmental Health Perspectives, 2006, 114 (4): 542-546.
- 3. Shrivastava Richa, Upreti RK, Seth PK, et al. Effects of chromium on the immune system [J]. Immunology and Medical Microbiology, 2002, 34 (4): 1-7.
- 4. Shashwat Katiyar, Awasthi SK, Srivastava JK. Effect of chromium on the level of IL-12 and IFN- $\gamma$  in occupationally exposed workers[J]. Science of the total environment, 2009, 407 (6): 1868-1874.
- 5. Mohammad BH, Marie Vahter, Gabriela Concha, et al. Low level environmental cadmium exposure is associated with DNA hypomethylation in argentinean women [J]. Environmental Health Perspectives, 2012, 120 (6): 879–884.
- 6. Xiaohua L, Yanshuang S, Li W, <del>Yuhui L, Ji Z, Yanhui M, Yun W, Wenjun M, Lei Y, Gua</del> Evaluation of the correction between genetic damage and occupational chromate exposure through BMMN frequenciess[J]. J Occup Environ Med, 2012, 54(2):166-170.
	- 7. De Flora S, Bagnasco M, Serra D, et al. Genotoxicity of chromium compounds: A review[J]. Mutation Research/Reviews in Genetic Toxicology, 1990, 238 (5): 99-172.
- 8. Wu LL, Chiou C-C, Chang P-Y, et al. Urinary 8-OHdG: a marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetics[J]. Clinica Chimica Acta, 2004, 339  $(1-2)$ :  $1-9$ .
	- 9. Ren C, Fang S, Wright RO, et al. Urinary 8-hydroxy-2-deoxyguanosine as a biomarker of oxidative DNA damage induced by ambient pollution in the Normative Aging Study[J]. Occupational and environmental medicine, 2011, 68 (2): 562-569.
	- 10. Lin CY, Lee HL, Chen YC, et al.Positive association between urinary levels of 8-hydroxydeoxyguanosine and the acrylamide metabolite N-acetyl-S-(propionamide)-cysteine in adolescents and young adults[J]. J Hazard Mater, 2013, 261: 372-377.
	- 11. Kim JY, Mukherjee S, Ngo LC, et al. Urinary 8-hydroxy-2'-deoxyguanosine as a biomarker of oxidative DNA damage in workers exposed to fine particulates[J]. Environmental health perspectives, 2004, 112 (6): 666-671.
- m Health, 2010, 83 (7): 803 811.<br>
Hattee Goldeni, Olga Asampa, et al. The effect of inhaled chromium on<br>
reach condense. Some sects borandsers among chrome-plating workers [J].<br>
Upred RK, Seth PK, et al. Frfects of chrom 12. Valavanidis A, Loridas S, Vlahogianni T, et al.Influence of ozone on traffic-related particulate matter on the generation of hydroxyl radicals through a heterogeneous synergistic effect[J].J Hazard Mater. 2009, 162 (2-3): 886-892.
	- 13. Valavanidis A, Vlachogianni T, Fiotakis C. 8-hydroxy-2-deoxyguanosine (8-OHdG): a critical biomarker of oxidative stress and carcinogenesis[J]. Journal of environmental science and health Part C, 2009, 27 (2): 120-139.
- 14. M. Fenech M., M. Kirsch-Volders1, M.  $-A$ . T. Natarajan AT, etal. Molecular mechanisms of micronucleus, nucleoplasmic bridge and nuclear bud formation in mammalian and human cells. Mutagenesis, 2011, 26: 125–132.
	- 15. Sobol Z, Schiestl RH. Intracellular and extracellular factors influencing Cr (VI) and Cr (III) genotoxicity[J]. Environmental and Molecular Mutagenesis, 2012, 53 (2): 94-100.
	- 16. Kuo H, Chang S, Wu K, et al. Chromium (VI) induced oxidative damage to DNA: increase of urinary 8-hydroxydeoxyguanosine concentrations (8-OHdG) among electroplating workers[J]. Occupational and environmental medicine, 2003, 60 (8): 590-594.
	- 17.Kim H, Cho S-H, Chung M-H. Exposure to hexavalent chromium does not increase 8-hydroxydeoxyguanosine levels in Korean chromate pigment workers[J]. Industrial health, 1999, 37 (3): 335-341.
	- 18. Gao M, Levy LS, Faux SP, et al. Use of molecular epidemiological techniques in a pilot study

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![](_page_56_Figure_2.jpeg)

![](_page_56_Figure_3.jpeg)

Figure1 Correlation between LnMNCC or LnMNC with Lg (concentration of 8-OHdG in urine) in chromate exposed group 173x140mm (300 x 300 DPI)

![](_page_57_Figure_3.jpeg)

Figure 2 Correlation between urinary 8-OHdG and CrB in chromate exposed group 173x140mm (300 x 300 DPI)

![](_page_58_Figure_3.jpeg)

Figure 3 a showed the correlation between urinary 8-OHdG and CrB in higher chromate exposed group (CrB≥10.50). b showed the the linear relationship between urinary 8-OHdG and CrB in lower chromate exposed group(CrB<10.50) 173x230mm (300 x 300 DPI)

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![](_page_59_Figure_2.jpeg)

![](_page_59_Figure_3.jpeg)

Figure 4 Correlation between LnMNC or LnMNCC and CrB in chromate exposed group 173x230mm (300 x 300 DPI)

![](_page_60_Figure_3.jpeg)

Figure 5 a showed the correlation between LnMNC or LnMNCC and CrB in higher chromate exposed group (CrB≥10.50). b showed the the linear relationship between LnMNC or LnMNCC and CrB in lower chromate exposed group(CrB<10.50) 173x230mm (300 x 300 DPI)