

Supplementary Information

Supplementary Table

Compound	Suggested Mechanisms	Assay	Reference
Resveratrol	Sirtuin activator, PDE inhibitor, antioxidant	Worm PolyQ neuroprotection Mouse amyloid plaque reduction	(1) (2)
Mithramycin	DNA binding agent	Worm PolyQ neuroprotection Mouse HD survival	(3) (4)
Clioquinol	Metal chelator and CLK-1 inhibitor	Worm <i>clk-1</i> partial mimic Mouse Alzheimer cognition	(5) (6)
Valproic acid	Anticonvulsant (HDAC inhibitor)	Worm lifespan increase Fly PolyQ neuroprotection	(7) (8)
Mianserin	Antidepressant (5-HT ₂ blocker)	Worm lifespan increase	(9)
C2-8	unknown	Fly PolyQ neuroprotection Mouse HD survival	(10) (11)
Rapamycin	mTOR inhibitor	Cell synuclein/PolyQ Mouse AD cognition	(12) (13)

Table S1: Compounds used in *dnj-14* lifespan screen.

Figure legends

FIG. S1: *dnj-14* gene structure, mutations and homology to human *dnajc5*.

(A) Sequence alignment of the predicted products of the human *DNAJC5* and *C. elegans dnj-14a* genes (CSP α and DNJ-14 proteins, respectively).

(B) Exon structure of the *dnj-14* gene and location of the *ok237* and *tm3223* alleles.

(C) Confirmation of the *ok237* deletion. Genomic DNA from wild type N2 and *dnj-14(ok237)* worms was amplified using primers flanking the deletion (left panel) or with one flanking primer and a primer within exon 2 of *dnj-14* (right panel).

(D) Characterisation of the *tm3223* insertion/deletion. Genomic DNA from wild type N2 and *dnj-14(tm3223)* worms was amplified using primers flanking the mutation site.

FIG. S2: Quantification of neurodegeneration in *dnj-14(ok237)* mutants.

Wild type N2 and *dnj-14(ok237)* worms were synchronised and grown on NGM plates containing vehicle control (ethanol) or 100 μ M resveratrol for at least 9 days. Animals were then immobilised for GFP imaging and scored for head neuron abnormalities ('neuron loss') based on loss of neuronal cell bodies, a reduction in the number of visible neurites, or the presence of contorted neuronal processes in the head of the worms; and the presence of large fluorescent punctae in the dorsal nerve cord ('punctae'). The number of worms analysed was 45 for N2, 35 for *dnj-14* and 20 for *dnj-14* Resveratrol.

FIG. S3: Visualisation of neurodegeneration in *dnj-14(ok237)* mutants.

Wild type N2 and *dnj-14(ok237)* worms were synchronised and grown on NGM plates for at least 9 days. Animals were then immobilised for GFP imaging. Typical images are shown for various animals. These illustrate the loss of neuronal cell bodies, reduction in the number of

visible neurites, or the presence of contorted neuronal processes in the head of the worms that was frequently seen in *dnj-14* mutants. In contrast, age-matched wild type N2 animals generally exhibited obvious neuronal cell bodies and had clearly labelled multiple neurites that extended straight to the end of the worm's head without twisting.

FIG. S4: Mechanosensation is unaffected in aged *dnj-14* mutants.

Wild type N2, *dnj-14(tm3223)* and *dnj-14(ok237)* worms were synchronised and grown on NGM plates until 6 days of age. After transfer to unseeded plates, mechanosensation was assessed by gently touching an eyelash to the side of the worm's head at a perpendicular angle. Each worm was assayed ten times and the number of times the worm either stopped or reversed its direction of movement was recorded. No significant differences between wild type and *dnj-14* mutants were seen (n=15 animals per strain).

FIG. S5: Pharyngeal pumping is unaffected in aged *dnj-14* mutants.

Wild type N2, *dnj-14(tm3223)* and *dnj-14(ok237)* worms were synchronised and grown on NGM plates until 6 days of age. After transfer to plates freshly seeded with OP50 bacteria, the number of contraction/relaxation cycles of the pharynx per thirty seconds was for each worm was recorded. No significant differences between wild type and *dnj-14* mutants were seen (n=15 animals per strain).

FIG. S6: Measurement of cAMP levels in worms treated with resveratrol and rolipram.

Wild type N2 worms were age-synchronised and grown on 60-mm NGM plates seeded with JB1669 adenylyl cyclase deficient bacteria. Approximately fifteen such plates of day 2 worms were washed in M9 buffer and then treated in M9 buffer containing vehicle control (ethanol), 100 μ M resveratrol or 100 μ M rolipram for 120 mins. The worms were then lysed,

centrifuged to pellet any debris, and the supernatant used immediately for assay. Endogenous cyclic AMP levels were measured by ELISA and normalised to total protein concentration. Data shown are mean + SEM ($n = 4$ biological replicates). No significant differences between treatments were seen.

MOVIE S1: Superficially normal locomotion in *dnj-14(ok237)* mutants

The movement of worms on NGM agar plates was recorded. Wild type N2 worms are shown first, followed by *dnj-14(ok237)* mutants.

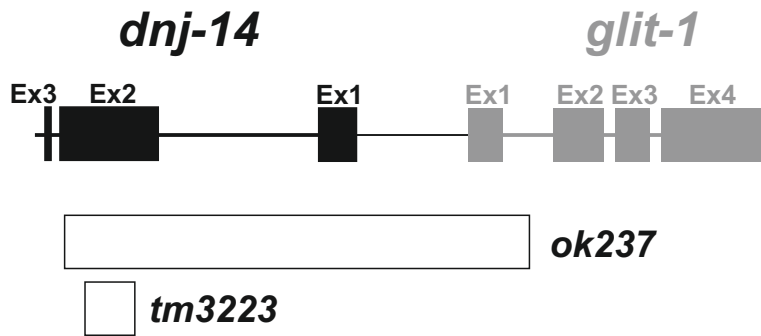
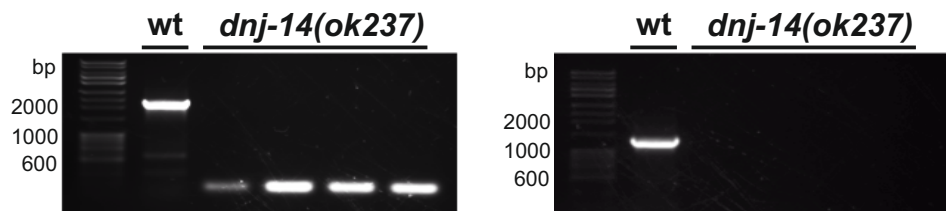
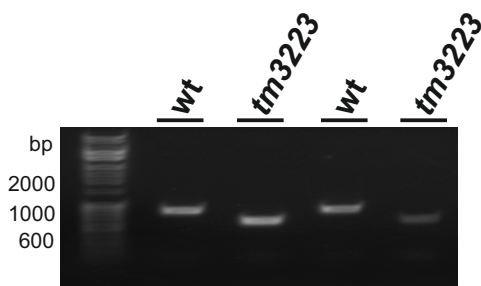
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a

Human CSP α	1	MADQRQRSLSTSG-----ES-----LYHVLGLDKNAT	27
Worm DNJ-14	1	MNSDGLREAEEGRTSGGASPREESPADHSHDPKKGLHLYNVLGIQKNAT	50
Human CSP α	28	SDDIKKSYRKLALRYHPDKNPD-NPEAADKFKKEINNAHAILTDATKRNII	76
Worm DNJ-14	51	DDEIKKAYRKLALRYHPDKNLDGDPEKTEMFKEINYANAVLSNPNKRRVY	100
Human CSP α	77	DKYGSLGLYVAEQFGEENVNTYFVLSWAKALFVFCGLLTCYCCCCCLC	126
Worm DNJ-14	101	DEMGETGLKLMQFGEDEKILQWMLKPWEKWTFFAFGLLIGFFCCCCGC	149
Human CSP α	126	-CCFNCCCGKCKPKAPEGEETEFYVSPEDLEAQLQSDEREATDTPIVIQP	175
Worm DNJ-14	150	MCCQCCCFNFCCKYKPKHDETF--ADFTSDGDVIVDQPTASEP---MPD	194
Human CSP α	176	ASATETTQLTADSHPSYHTDGFN	198
Worm DNJ-14	195	TNRRQVPIVIAMPPEPSQKD---	214

b**c****d****Fig S1 Kashyap et al**

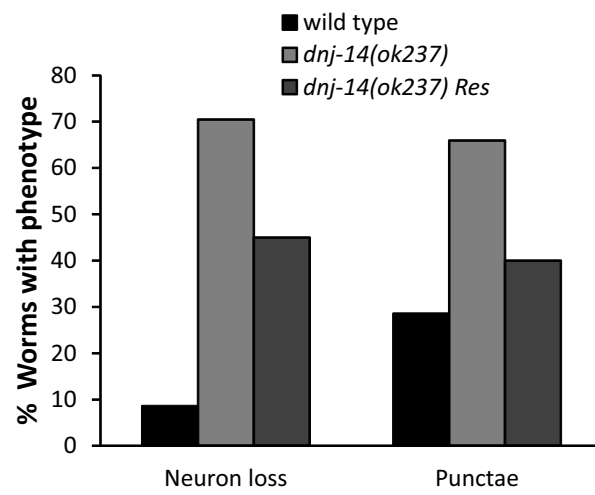


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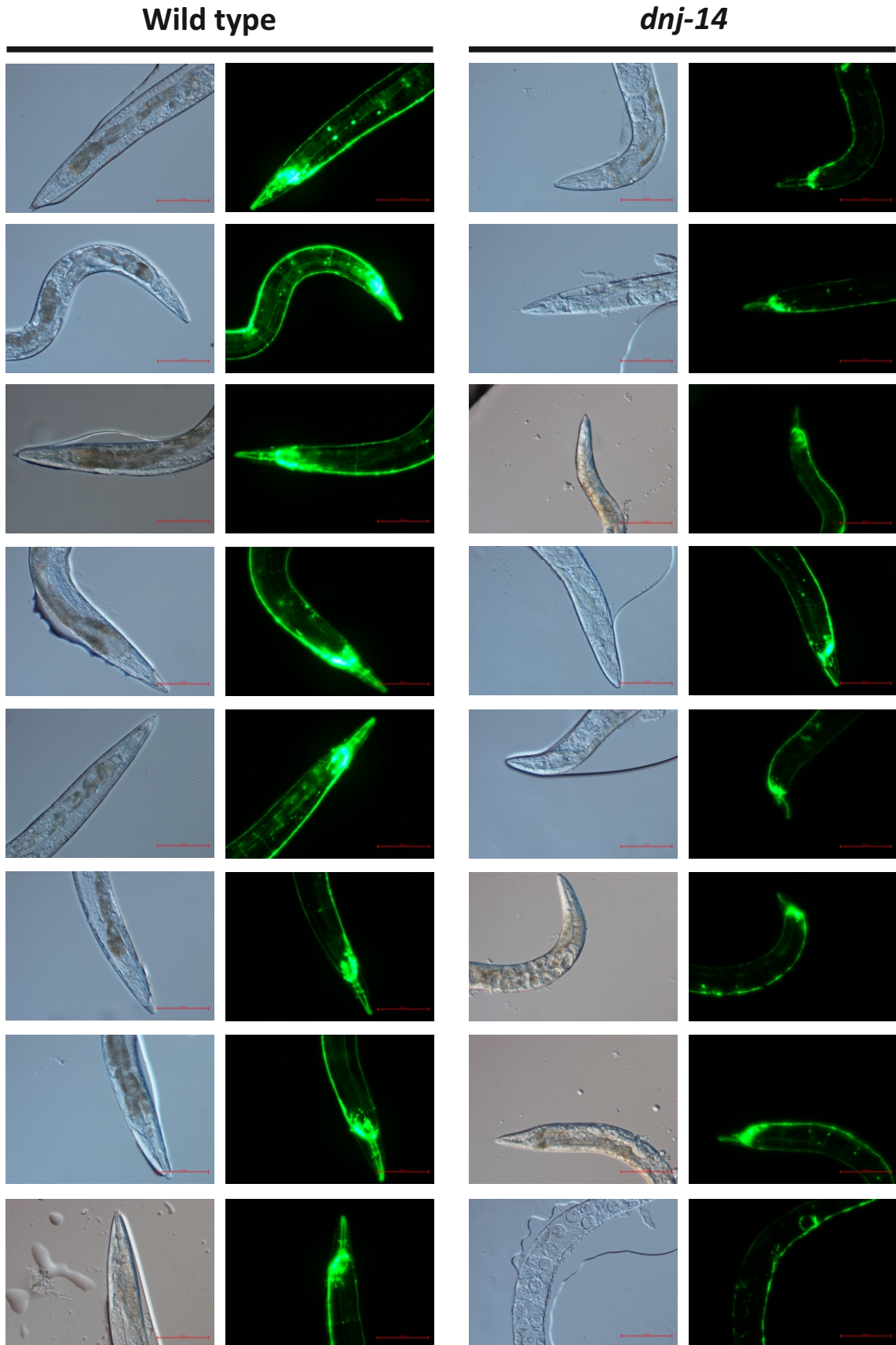


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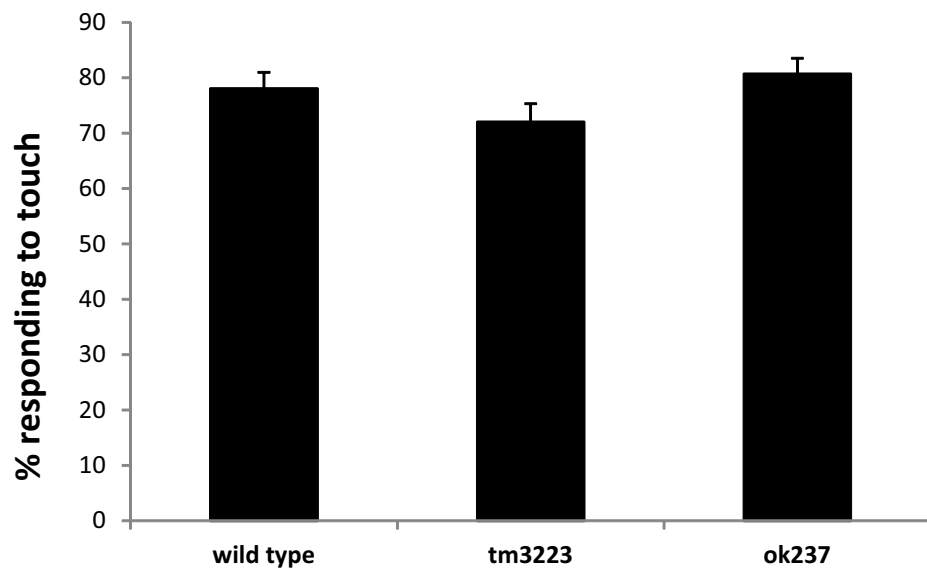


Fig S4 Kashyap et al

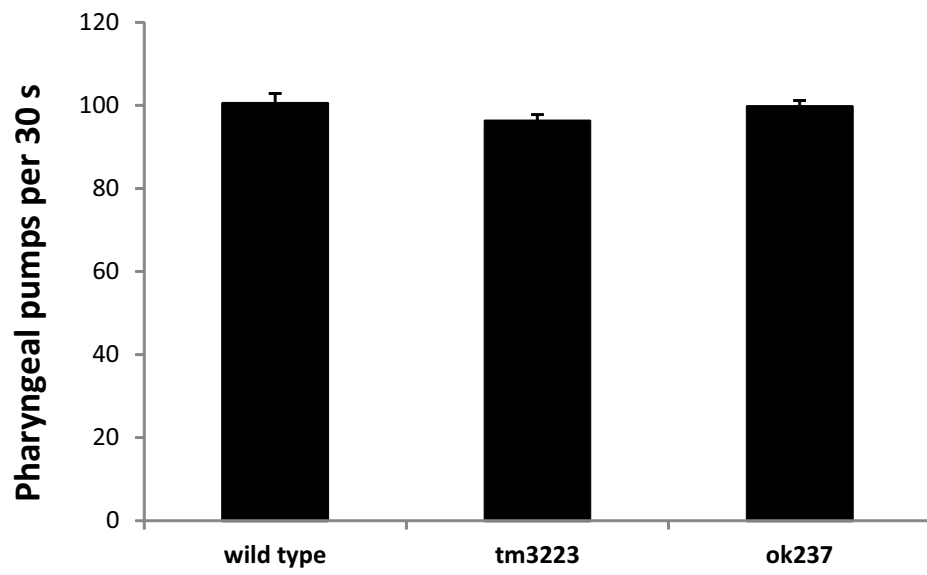


Fig S5 Kashyap et al

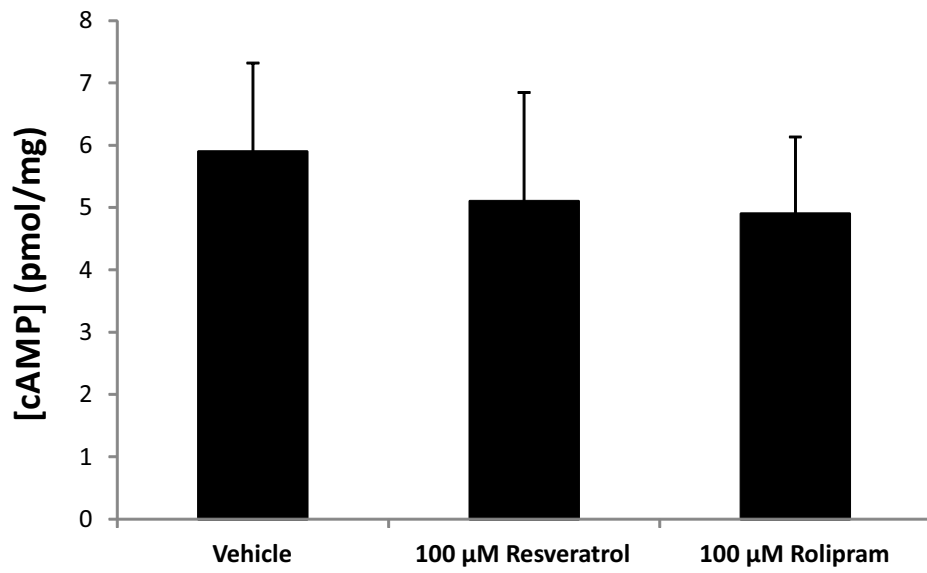


Fig S6 Kashyap et al