

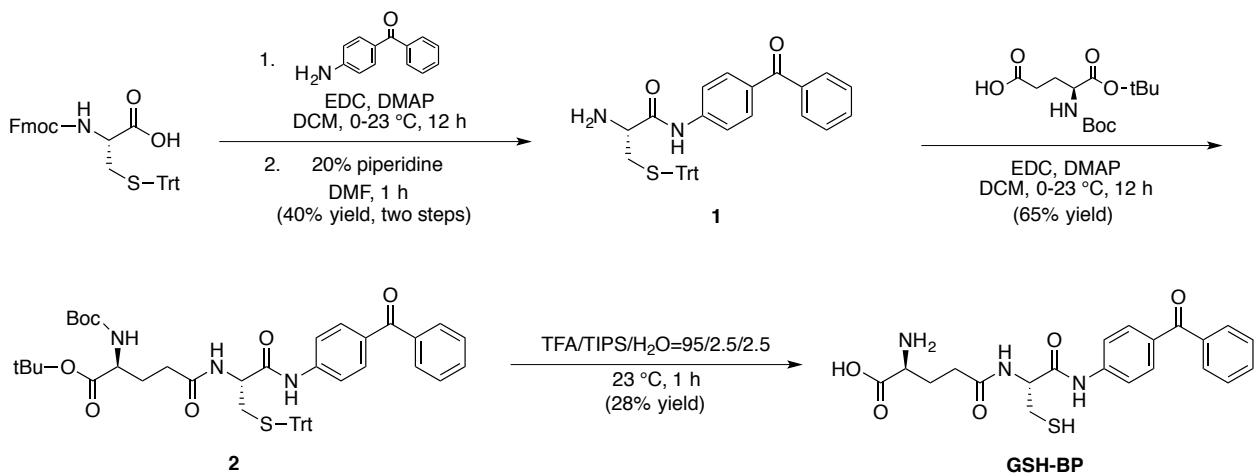
Protein-Polymer Conjugation *via* Ligand Affinity and Photoactivation of Glutathione *S*-Transferase

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Supporting Information

Scheme S1. Synthesis of GSH-BP.



Synthesis of 1. In a round bottom flask over ice, Fmoc-Cys(Trt)-OH (1.63 g, 2.79 mmol) was dissolved in anhydrous DCM (5 mL) and stirred to dissolve. 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC·HCl, 0.97 g, 5.07 mmol) was then added to the reaction mixture. In a separate flask, 4-aminobenzophenone (4-ABP, 0.51 g, 2.58 mmol) was dissolved in anhydrous DCM (5 mL). After keeping in an ice bath for 20 min, the 4-ABP solution was added dropwise to the reaction flask. 4-Dimethylaminopyridine (DMAP, 61.4 mg, 0.5 mmol) was then added, and the reaction was stirred from 0 °C to 23 °C for 12 h. The completion of the reaction was confirmed by thin layer chromatography (TLC, EtOAc:Hexanes=1:2, stained with ninhydrin). The crude mixture was washed with saturated NaHCO₃ three times. The organic layer was then dried over MgSO₄ and concentrated *in vacuo*. 20% piperidine in DMF (11 mL) was added to the crude solid, and allowed to stir at 23 °C for 1 h. The solvent was removed *in vacuo* at 40 °C. The crude product was purified via silica gel flash chromatography with EtOAc : Hexane = 1 : 2, then changed to DCM : MeOH = 95 : 5 to obtain **1** with 40% yield containing impurities, and was brought forward to the next step. ¹H NMR (500

MHz in MeOD) δ : 7.81-7.14 (m, 24H), 3.44-3.41 (t, J = 6.15 Hz, 1H), 2.67-2.46 (m, 2H). ^{13}C NMR (500 MHz in MeOD) δ : 196.25, 172.80, 144.60, 142.37, 137.77, 132.63, 132.17, 131.10, 129.48, 129.35, 128.09, 127.61, 126.52, 118.84, 118.15, 116.98 66.44, 54.71, 36.85. DART-MS (\pm 1.0) observed (predicted): H^+ 543.2064 (543.2062).

Synthesis of Protected GSH-BP 2. In a round bottom flask over ice, Boc-Glu-OtBu (93.9 mg, 0.31 mmol) was dissolved in anhydrous DCM (2 mL) and stirred to dissolve. EDC·HCl (98.9 mg, 0.52 mmol) was then added to the reaction mixture. In a separate flask, **1** (140 mg, 0.26 mmol) was dissolved in anhydrous DCM (2 mL). After keeping over ice for 20 min, the solution of **1** was added dropwise to the reaction flask. DMAP (5.8 mg, 0.05 mmol) was then added, and the reaction was stirred from 0 °C to 23 °C for 24 h. The crude mixture was washed with saturated NaHCO_3 three times. The organic layer was then dried over MgSO_4 and concentrated *in vacuo*. The crude product was purified using silica gel flash chromatography with EtOAc : Hexane = 1 : 2 to obtain 140 mg white solid with 65% yield containing slight amount of impurities. ^1H NMR (500 MHz in MeOD) δ : 7.81-7.16 (m, 24H), 4.49-4.46 (m, 1H), 4.03-3.84 (m, 1H), 2.71-2.49 (m, 2H), 2.41-2.26 (m, 2H), 2.16-1.77 (m, 2H), 1.52-1.36 (m, 18H). ^{13}C NMR (500 MHz in MeOD) δ : 197.51, 174.77, 173.14, 170.96, 158.09, 145.82, 143.80, 139.90, 134.29, 134.05, 133.53, 132.50, 132.39, 130.82, 130.68, 130.31, 129.47, 129.22, 129.03, 127.94, 120.47, 120.38, 114.07, 82.75, 80.54, 68.00, 55.34, 55.25, 54.64, 54.52, 34.89, 32.86, 28.73, 28.58, 28.24. IR: ν = 3292, 2972, 1703, 1652, 1595, 1522, 1445, 1366, 1310, 1278, 1251, 1149, 1047, 923, 845, 741. ESI-MS (\pm 1.0) observed (predicted): H^+ 828.3785 (828.3638).

Synthesis of GSH-BP. Protected GSH-BP **2** (41.5 mg, 0.05 mmol) was weighed in a glass vial. Under argon, trifluoroacetic acid (TFA, 1.33 mL), triisopropylsilane (TIPS, 35 μ L, 0.17 mmol), and H₂O (35 μ L) were added to the reaction with a final ratio of TFA/TIPS/H₂O = 95/2.5/2.5. Upon the addition of TFA, the solution immediately turned bright yellow. After the addition of TIPS, the color quickly turned pale yellow with the formation of a white precipitate. The reaction was allowed to stir for 1 h at 23 °C and then dried *in vacuo* to remove TFA. The solid was dissolved in 30% acetonitrile (ACN) and filtered for HPLC purification. HPLC chromatography was carried out with a gradient elution from 30% to 70% ACN in 30 min with a flow rate of 10 mL/min. The collected fractions were lyophilized to obtain 6.1 mg white solid with 28% yield. ¹H NMR (500 MHz in MeOD) δ : 7.79-7.51 (m, 9H), 4.67-4.58 (m, 1H), 3.70-3.58 (m, 1H), 3.04-2.83 (m, 2H), 2.56-2.53 (t, J = 7.13 Hz, 2H), 2.22-2.07 (m, 2H). ¹³C NMR (500 MHz in MeOD) δ : 196.16, 173.88, 172.48, 169.69, 142.55, 137.73, 132.71, 132.17, 130.99, 129.41, 128.10, 119.06, 56.77, 54.20, 31.31, 26.36, 25.33. IR: ν = 3293, 2977, 1645, 1595, 1517, 1446, 1407, 1365, 1311, 1279, 1252, 1174, 1149, 1047, 938, 924, 851, 793, 741. ESI-MS (\pm 1.0) observed (predicted): H⁺ 430.1450 (430.1392). UV-Vis (MeOH) λ_{max} = 295 nm.

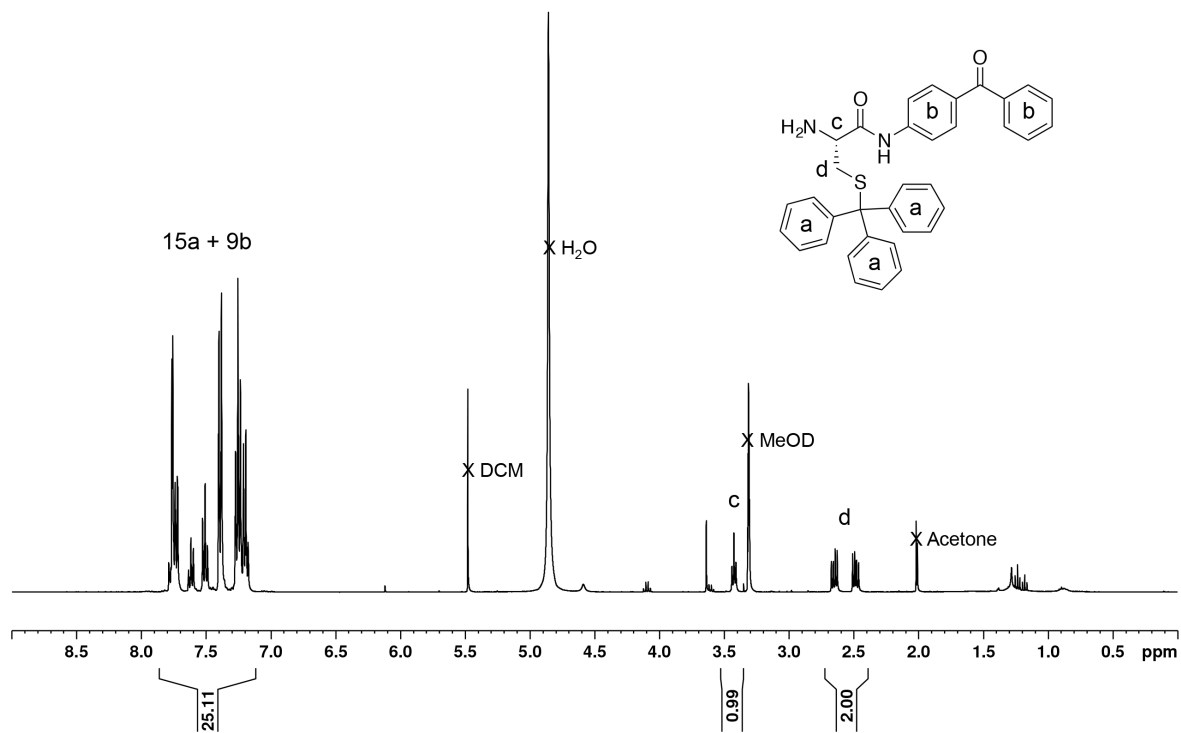


Figure S1. ^1H NMR spectrum of **1** (in MeOD).

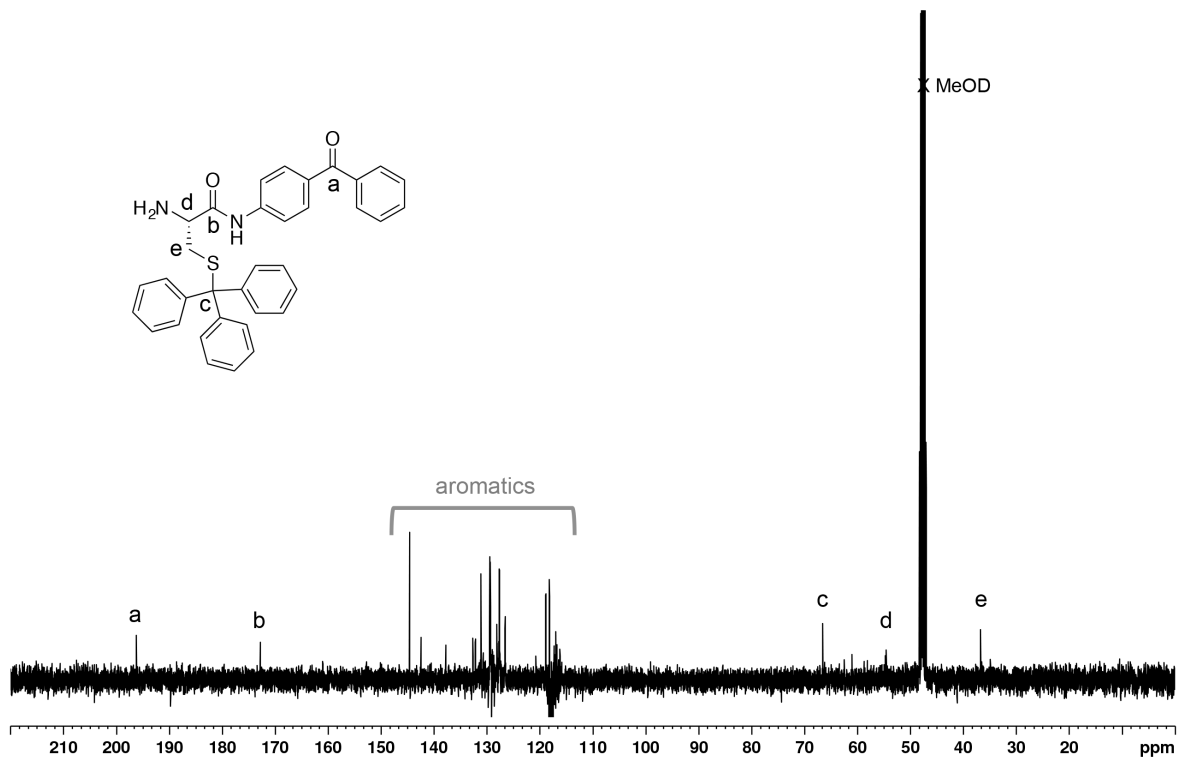


Figure S2. ^{13}C NMR spectrum of 1 (in MeOD).

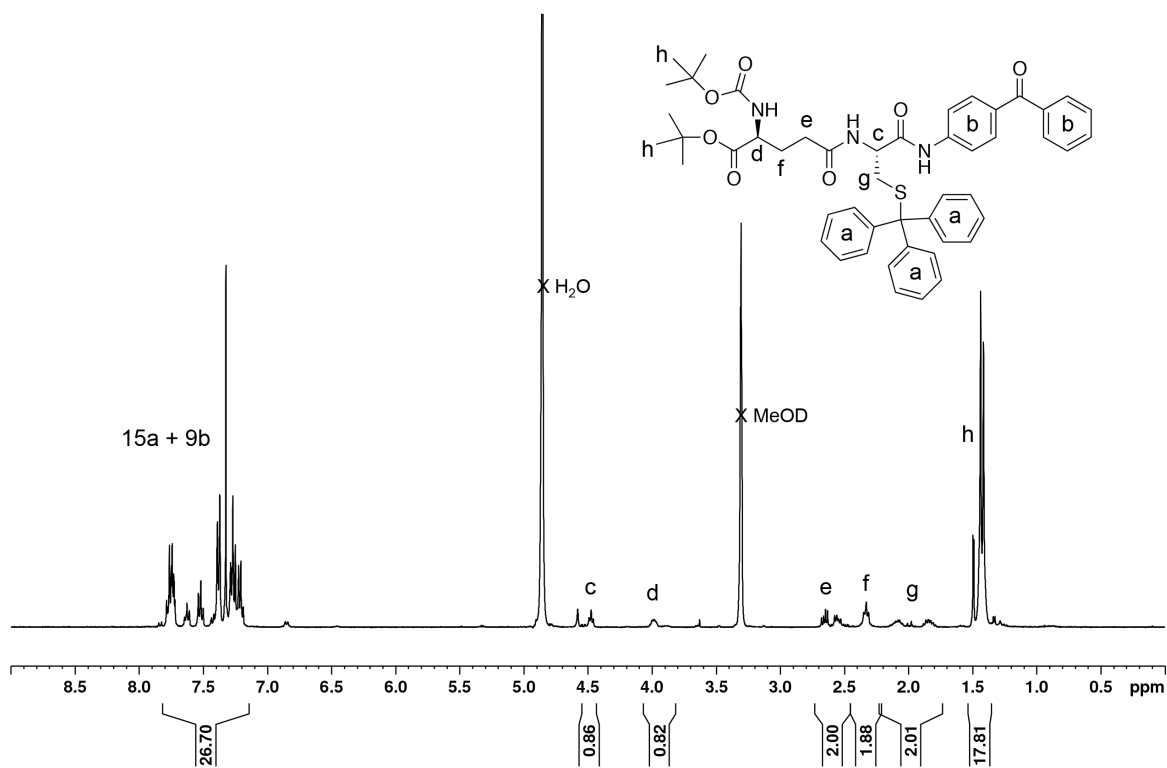


Figure S3. ^1H NMR spectrum of **2** (in MeOD).

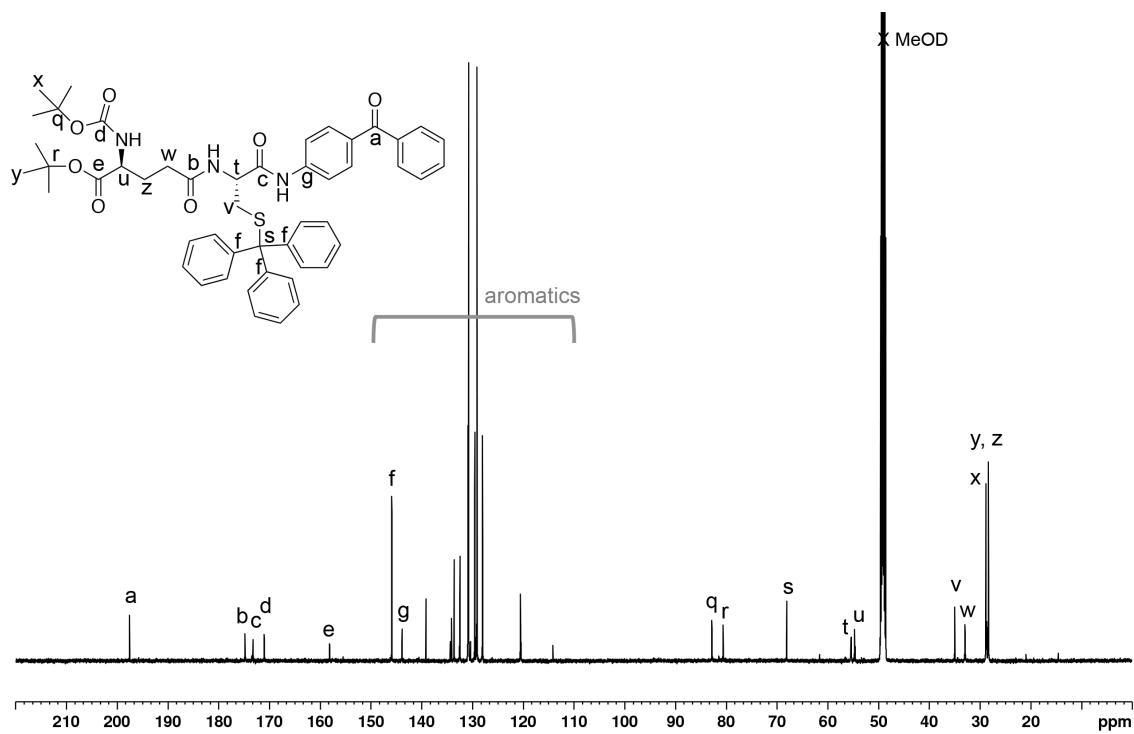


Figure S4. ^{13}C NMR spectrum of **2** (in MeOD).

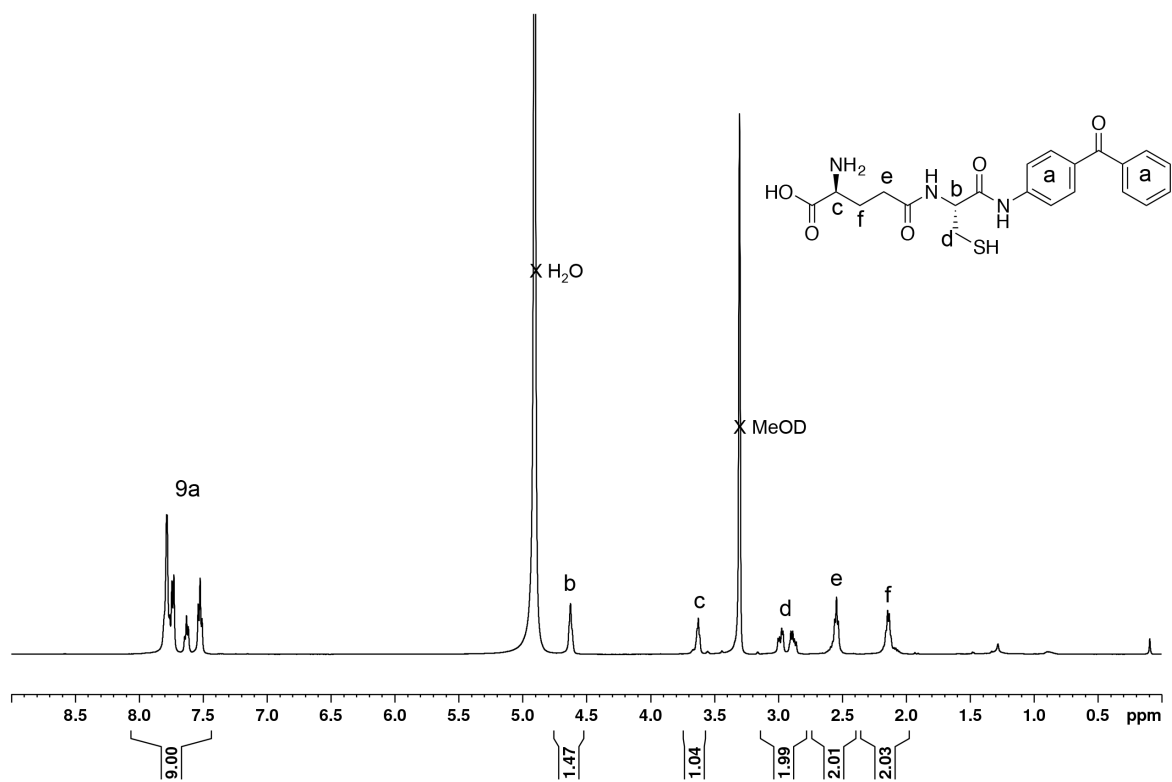


Figure S5. ¹H NMR spectrum of **3** (in MeOD).

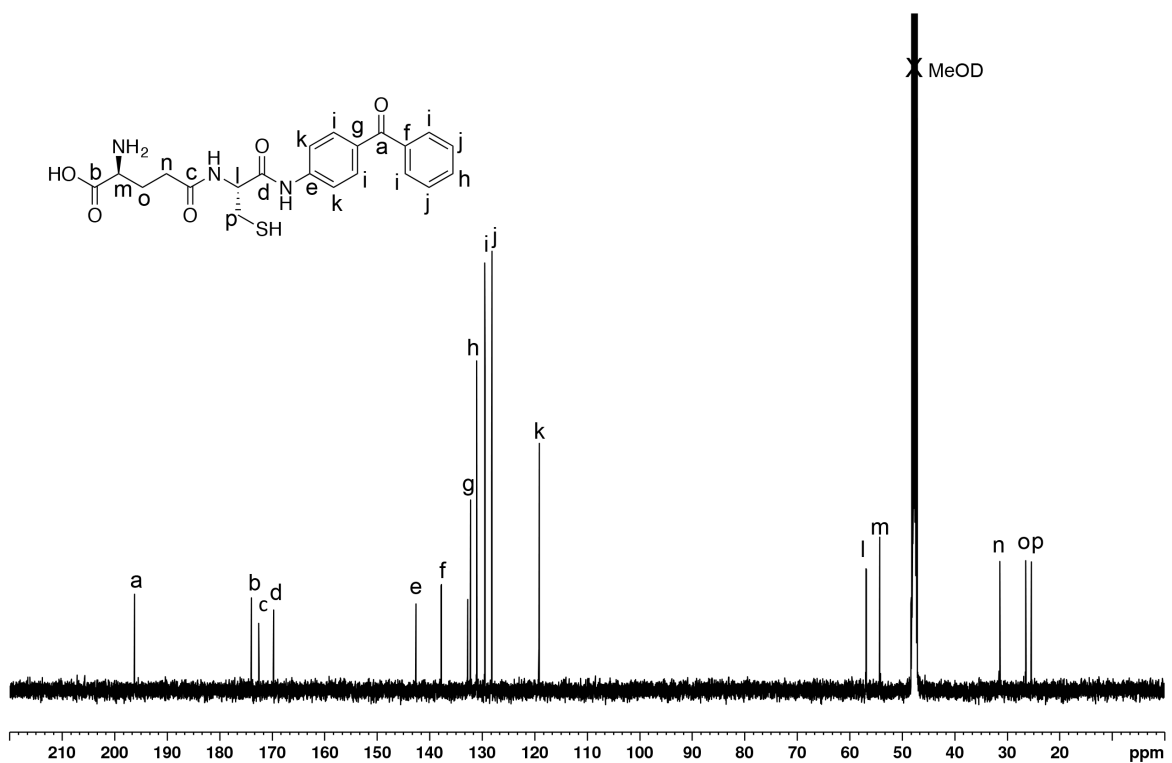
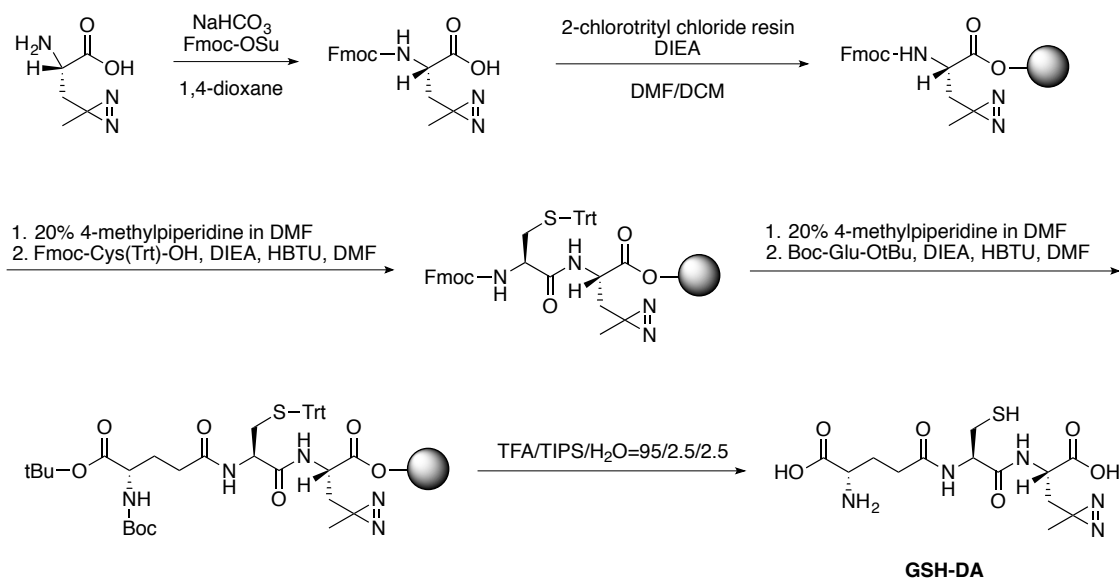


Figure S6. ^{13}C NMR spectrum of **3** (in MeOD).

Scheme S2. Synthesis of GSH-DA.



Synthesis of Fmoc-PhotoLeu. Fmoc-PhotoLeu was synthesized using a previously reported procedure.¹ Fmoc-OSu (118 mg, 0.35 mmol) was dissolved in 1,4-dioxane (2 mL). L-Photo-Leucine (50 mg, 0.35 mmol) and NaHCO₃ (47 mg, 0.56 mmol) were dissolved in H₂O (1.5 mL). 1,4-dioxane (2.5 mL) was added to this solution before adding Fmoc-OSu solution over 15 minutes. The reaction was then stirred for 26 hours at 23°C. HCl was added to the solution to acidify to pH 3 before extracting 3 times into EtOAc. Organic layers were combined and dried over MgSO₄ and concentrated *in vacuo*. Silica gel chromatography (5% MeOH in CH₂Cl₂ + 0.1% AcOH) yielded 107 mg (85%) as a beige solid. All steps were carried out in the dark. ¹H NMR (500 MHz in MeOD) δ: 7.81-7.63 (d, *J*= 7.5 Hz, 2H), 7.74-7.68 (dd, *J*= 4.6, 7.0 Hz, 2H), 7.41-7.34 (t, *J*= 7.5 Hz, 2H), 7.33-7.27 (dt, *J*= 1.05, 7.5 Hz, 2H), 4.42-4.30 (m, 2H), 4.28-4.22 (t, *J*= 7.15 Hz, 1H), 4.15-4.07 (dd, *J*= 4.3, 10.5 Hz, 1H), 2.03-1.95 (dd, *J*= 4.4, 14.95 Hz, 1H), 1.67-1.59 (dd, *J*= 10.6, 14.95 Hz, 1H), 1.06-1.00 and 0.93-0.89 (s, rotamer, 3H). ¹³C NMR (500 MHz in MeOD) δ: 173.39, 157.04, 143.81, 141.18, 127.38, 126.76, 124.94, 119.50, 66.70, 50.10, 36.19, 23.60, 18.48.

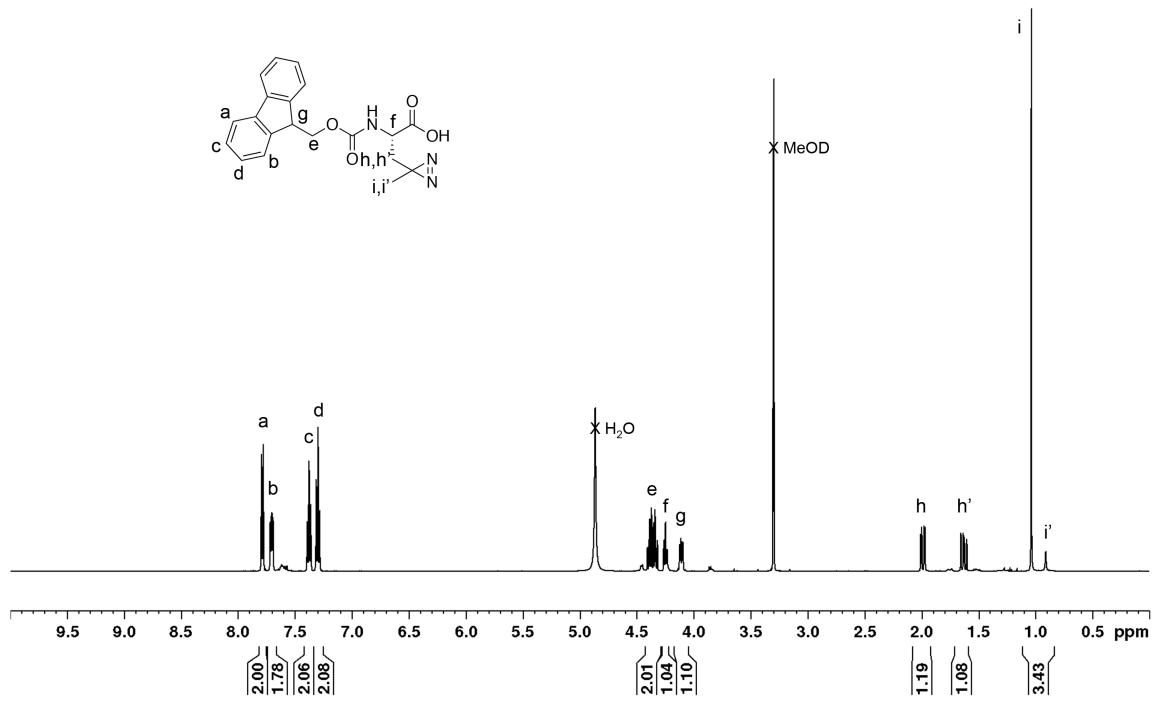


Figure S7. ^1H NMR spectrum of Fmoc-photo-Leucine (in MeOD).

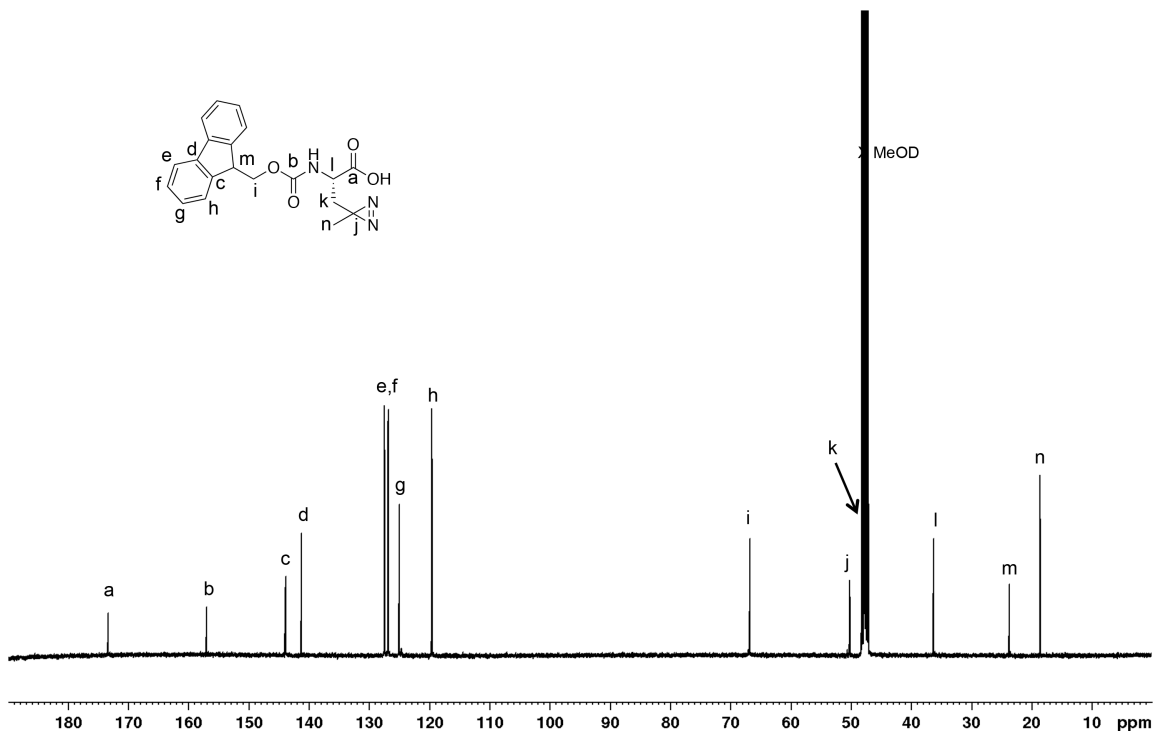


Figure S8. ^{13}C NMR spectrum of Fmoc-photo-Leucine (in MeOD). Note: It is anticipated that the carbon peak k is under the MeOD peak.

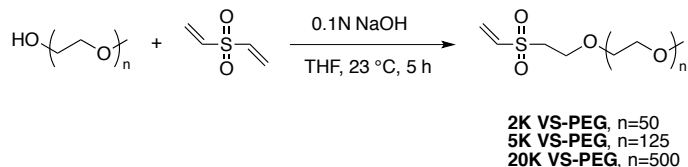
Synthesis of GSH-DA:

Resin Loading.² Fmoc-PhotoLeu (106 mg, 0.29 mmol) and DIEA (202 μL , 1.2 mmol) were dissolved in anhydrous DCM (2 mL). A small amount of DMF (200 μL) was added to dissolve the acid. The solution was added to the 2-chlorotrityl chloride resin (219 mg, .29 mmol) and mixed for 45 minutes in a peptide synthesis flask. The solution was drained and resin was rinsed with DCM/MeOH/DIEA (17:2:1), DCM, DMF, DCM, MeOH, DCM, DMF. The loaded resin was stored under argon until further use.

Amino Acid Coupling. 20% 4-methyl piperidine in DMF (10 mL) was added to resin in synthesis flask for 20 minutes to remove the Fmoc group. Solution was drained and resin was washed with DMF/DCM/MeOH/DCM/DMF. Fmoc-Cys(Trt)-OH (250 mg, 0.38 mmol) and HBTU (161 mg, 0.42 mmol) were dissolved in DMF (10 mL) and DIEA (148 μ L, 1.5 mmol) was added before adding solution to resin and mixing for 3 hours at 23°C. Solution was drained and resin was washed again. The Fmoc group was removed as described previously. Boc-Glu-OtBu (129 mg, 0.41 mmol) and HBTU (161 mg, 0.42mmol) were dissolved in DMF (10 mL) before adding DIEA (148 μ L, 1.5 mmol). This solution was added to resin and mixed at 23°C for 2 hours before draining solution and washing the resin.

Peptide Cleavage. The peptide was cleaved from the chlorotriyl resin using 95% TFA, 2.5% TIPS and 2.5% H₂O. The resin was mixed in cleavage cocktail for 45 minutes before draining the solution and rinsing the resin with DCM twice. The filtrate was concentrated *in vacuo* before precipitating in cold diethyl ether. The crude peptide was purified using HPLC with a C18 column with a gradient elution from 5% to 95% ACN in 30 min with a flow rate of 10 mL/min. The trityl group was subsequently deprotected using a solution of 95% TFA, 2.5% TIPS and 2.5% H₂O to yield 16.7 mg of pure GSH-DA as a white solid after precipitation in cold diethyl ether. ESI-MS (\pm 1.0) observed (predicted): H⁺ 376.1289 (376.1291).

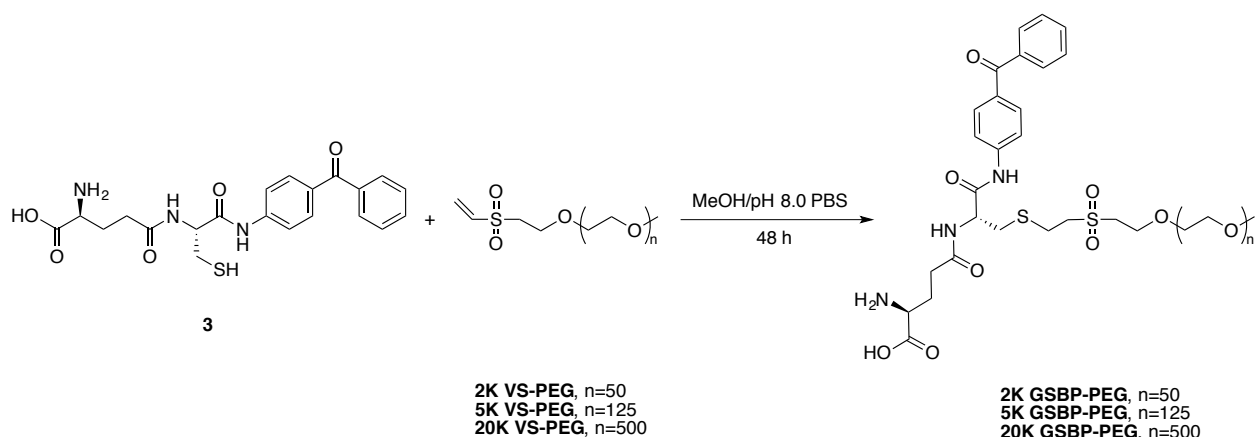
Scheme S3. Synthesis of VS-PEG.



Synthesis of VS-PEG. Here, we describe the synthesis of 2K VS-PEG. 5K and 20K VS-PEG were prepared according to this same method modified from a previous report.³ To poly(ethylene glycol) methyl ether of average $M_n \sim 2,000$ (1.32 g, 0.66 mmol), THF (35 mL) was added and warmed to 30 °C. After the polymer had dissolved, the solution was cooled down to room temperature. Divinyl sulfone (0.2 mL, 1.98 mmol) was added, followed by addition of 0.1 N aqueous NaOH solution (2.4 mL). The reaction mixture was stirred at 23 °C for 5 h, then neutralized with 0.1 N HCl solution (2.4 mL) and filtered. The filtrate was concentrated *in vacuo* to remove most of the THF. The residual solution was diluted in approximately 10 mL of H₂O, washed with diethyl ether (x3 times), and extracted with chloroform (x3 times). The organic layer was then washed with H₂O, dried over MgSO₄, and concentrated *in vacuo*. The white solid was then dissolved in minimum amount of DCM, and precipitated into cold ether (50 mL x3). The precipitant was dried *in vacuo* to obtain 1.1 g of white powder with 79% yield. ¹H NMR of 2K VS-PEG (500 MHz in CDCl₃) (conversion: 89%) δ : 6.86-6.79 (dd, $J= 9.95, 16.60$ Hz, 1H), 6.42-6.36 (d, $J= 16.65$ Hz, 1H), 6.11-6.06 (d, $J= 9.90$ Hz, 1H), 4.00-3.22 (m, 200H), 3.40-3.37 (s, 3H). ¹H NMR of 5K VS-PEG (500 MHz in CDCl₃) (conversion: 85%) δ : 6.84-6.76 (dd, $J= 9.90, 16.65$ Hz, 1H), 6.40-6.34 (d, $J= 16.65$ Hz, 1H), 6.09-6.04 (d, $J= 9.90$ Hz, 1H), 3.96-3.20 (m, 500H), 3.37-3.34 (s, 3H). ¹H NMR of 20K VS-PEG (500 MHz in CDCl₃) (conversion: 90%) δ : 6.83-6.75 (dd, $J= 10.00, 16.65$ Hz, 1H), 6.38-

6.32 (d, $J= 16.75$ Hz, 1H), 6.07-6.03 (d, $J= 10.00$ Hz, 1H), 3.95-3.19 (m, 2000H), 3.36-3.33 (s, 3H).

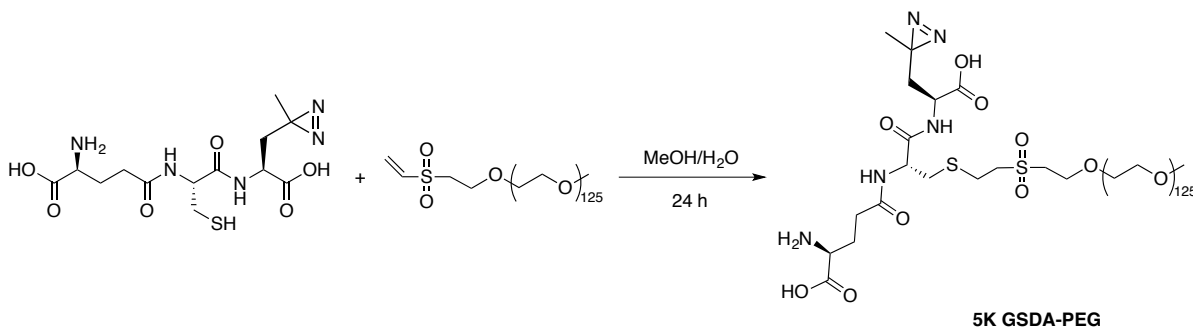
Scheme S4. Synthesis of GSBP-PEG.



Synthesis of GSBP-PEG. Here, we describe the synthesis of 2K GSBP-PEG. 5K and 20K GSBP-PEG were prepared according to the same method. Compound **3** (5.6 mg, 0.013 mmol) was dissolved in MeOH (0.2 mL). In a separate vial, VS-PEG (2K, 8.7 mg, 4.35×10^{-3} mmol) was dissolved in pH 8.0 phosphate buffer saline (0.2 mL). The two solutions were combined, and stirred under argon at 23 °C for 48 h. A small aliquot was taken for crude ^1H NMR in D_2O to confirm the disappearance of the vinyl peaks. The crude mixture was purified by HPLC with elution gradient of 60%-90% MeOH over 30 min. Alternatively, the crude mixture was purified by dialysis (MWCO 1,000) against H_2O for two days, followed by filtration to remove insoluble solids. The purified solution was then lyophilized to obtain 5.6 mg of white powder with 64% yield. ^1H NMR of 2K GSBP-PEG (500 MHz in D_2O) (conversion: 94%) δ : 7.88-7.49 (m, 9H), 4.76-4.69 (dd,

$J = 6.20, 8.00$ Hz, 1H), 3.96-3.37 (m, 200H), 3.36-3.34 (s, 3H), 3.20-3.15 (dd, $J = 6.20, 13.75$ Hz, 1H), 3.05-2.98 (m, 2H), 2.96-2.90 (dd, $J = 8.20, 13.75$ Hz, 1H), 2.58-2.52 (m, 2H), 2.18-2.10 (m, 2H). ^1H NMR of 5K GSBP-PEG (500 MHz in D_2O) (conversion: 100%) δ : 7.86-7.48 (m, 9H), 4.75-4.69 (dd, $J = 6.20, 8.05$ Hz, 1H), 3.96-3.40 (m, 500H), 3.36-3.34 (s, 3H), 3.21-3.14 (dd, $J = 6.10, 13.70$ Hz, 1H), 3.05-2.98 (m, 2H), 2.96-2.90 (dd, $J = 8.20, 13.75$ Hz, 1H), 2.58-2.52 (m, 2H), 2.17-2.09 (m, 2H). ^1H NMR of 20K GSBP-PEG (500 MHz in D_2O) (conversion: 95%) δ : 7.80-7.37 (m, 9H), 4.60-4.56 (m, 1H), 3.98-3.29 (m, 2000H), 3.27-3.25 (s, 3H), 3.08-2.86 (m, 4H), 2.48-2.33 (m, 2H), 2.08-2.96 (m, 2H).

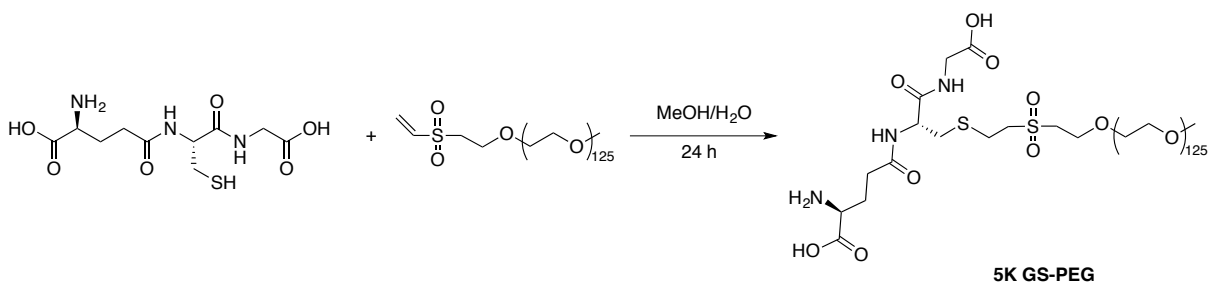
Scheme S5. Synthesis of 5K GSDA-PEG.



Synthesis of GSDA-PEG. GSH-DA (16.6 mg, 4.4×10^{-2} mmol) was dissolved in pH 8.0 phosphate buffer (1 mL). The solution was then added to VS-PEG (5K, 20.4 mg, 4.1×10^{-3} mmol), and the reaction mixture was stirred under Argon at 23 °C for 14 h. A small aliquot for crude ^1H NMR was taken in D_2O to confirm the disappearance of the vinyl peaks. The crude solution was concentrated via centrifugal filtration (0.5 mL, MWCO 3,000) and washed with H_2O for 5 times. The solution was then collected and

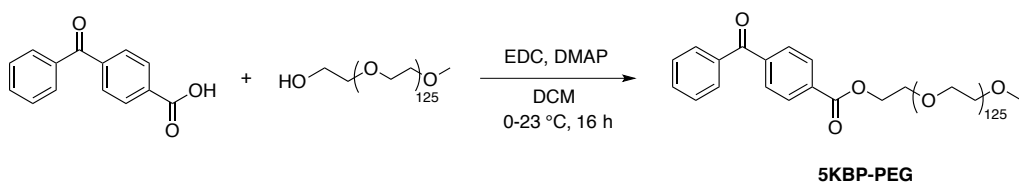
lyophilized. ^1H NMR (500 MHz in D_2O) (conversion: 75%) δ : 4.55-4.50 (dd, $J= 5.05$, 9.20 Hz, 1H), 4.08-4.02 (dd, $J= 4.20$, 9.50 Hz, 1H), 3.96-3.36 (m, 500H), 3.28-3.25 (s, 3H), 3.09-3.02 (dd, $J= 5.00$, 14.05 Hz, 1H), 2.97-2.89 (m, 2H), 2.88-2.81 (dd, $J= 9.25$, 14.05 Hz, 1H), 2.47-2.41 (m, 2H), 2.09-2.01 (m, 2H), 1.98-1.91 (dd, $J= 4.20$, 15.15 Hz, 1H), 1.54-1.46 (dd, $J= 9.55$, 15.15 Hz, 1H), 0.95-0.90 (s, 3H).

Scheme S6. Synthesis of 5K GS-PEG.



Synthesis of GS-PEG. Glutathione (153.7 mg, 0.5 mmol) was dissolved in pH 8.0, D-PBS (2 mL). The pH, which was around 3.0, was adjusted to 8.0 with 1 N NaOH and the total volume was diluted to 2 mL with additional phosphate buffer. The glutathione solution was added to VS-PEG (5K, 250 mg, 0.05 mmol), and the reaction mixture was stirred under argon at 23 °C for 24 h. A small aliquot for crude ^1H NMR was taken in D_2O to confirm the disappearance of the vinyl peaks. The crude was purified by dialysis (MWCO 1,000) against H_2O for two days, and then lyophilized. ^1H NMR of 5K GS-PEG (500 MHz in D_2O) (conversion: 86%) δ : 4.52-4.47 (dd, $J= 5.10$, 8.50 Hz, 1H), 3.90-3.39 (m, 500H), 3.27-3.23 (s, 3H), 3.05-2.97 (dd, $J= 4.95$, 14.10 Hz, 1H), 2.93-2.86 (m, 2H), 2.86-2.78 (dd, $J= 8.65$, 14.10 Hz, 1H), 2.47-2.35 (m, 2H), 2.08-1.99 (m, 2H).

Scheme S7. Synthesis of 5K BP-PEG.



Synthesis of BP-PEG. In a round bottom flask over ice, 4-benzoylbenzoic acid (181 mg, 0.8 mmol) was dissolved in DCM (4 mL). EDC·HCl (230.1 mg, 1.2 mmol), poly(ethylene glycol) methyl ether of average $M_n \sim 5,000$ (200 mg, 0.04 mmol), and DMAP (24.4 mg, 0.2 mmol) were then added to the reaction mixture. The reaction mixture was allowed to stir from 0 °C to 23 °C for 23 h. The crude mixture was washed once with H₂O, twice with saturated NaHCO₃, and twice with brine. The organic layer was then dried over MgSO₄ and concentrated *in vacuo*. The solid was purified by dialysis (MWCO 1,000) against H₂O for two days, followed by 0.22 μm PTFE syringe filtration to remove insoluble solids. The purified solution was then lyophilized. ¹H NMR (500 MHz in D₂O) (conversion: 100%) δ: 8.12-7.46 (m, 9H), 4.47-4.42 (m, 2H), 3.85-3.40 (m, 500H), 3.29-3.24 (s, 3H).

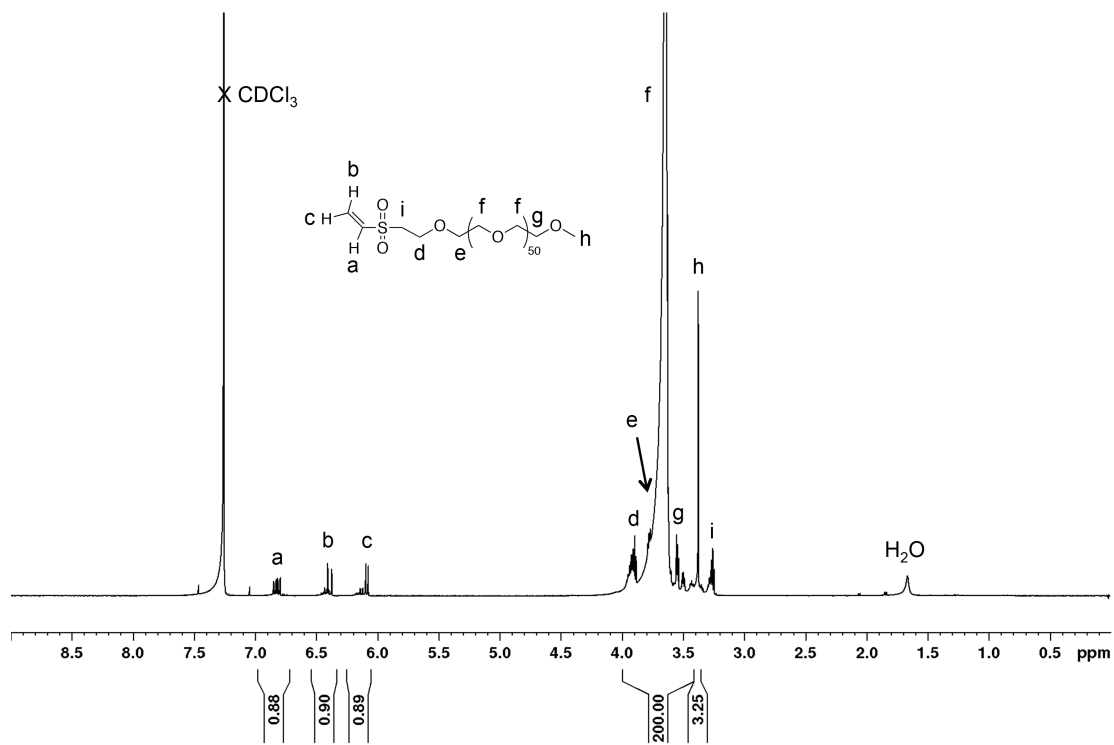


Figure S9. ^1H NMR spectrum of 2K VS-PEG (in CDCl_3).

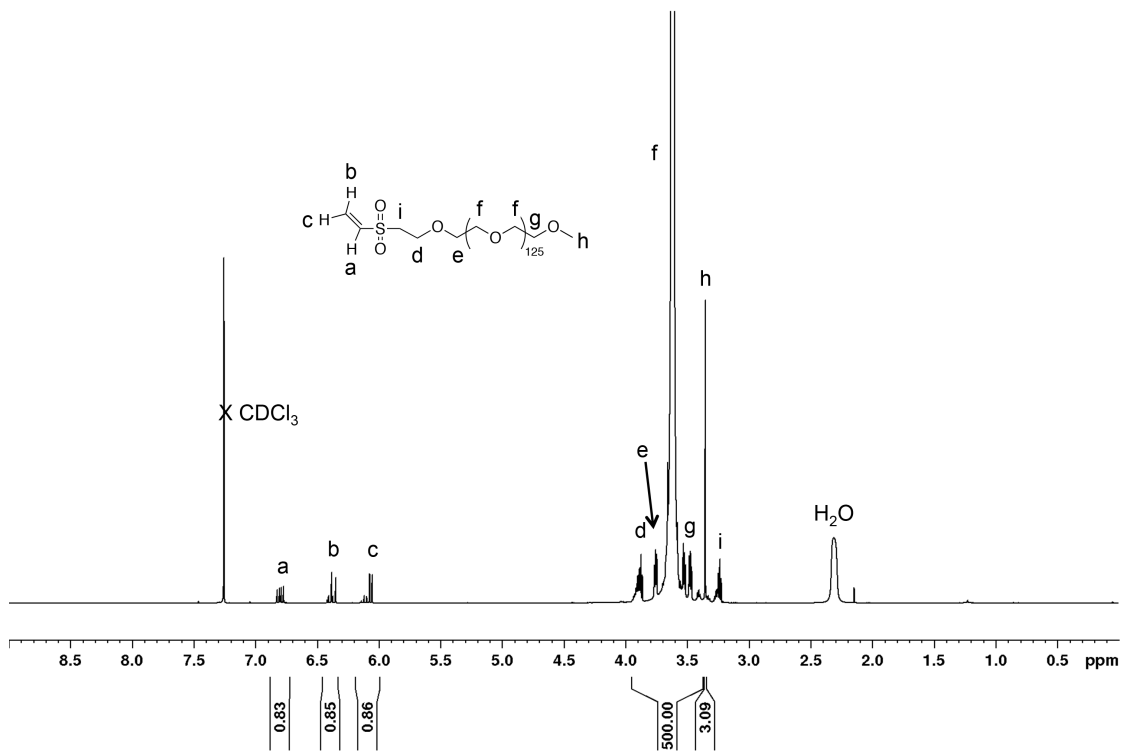


Figure S10. ^1H NMR spectrum of 5K VS-PEG (in CDCl_3).

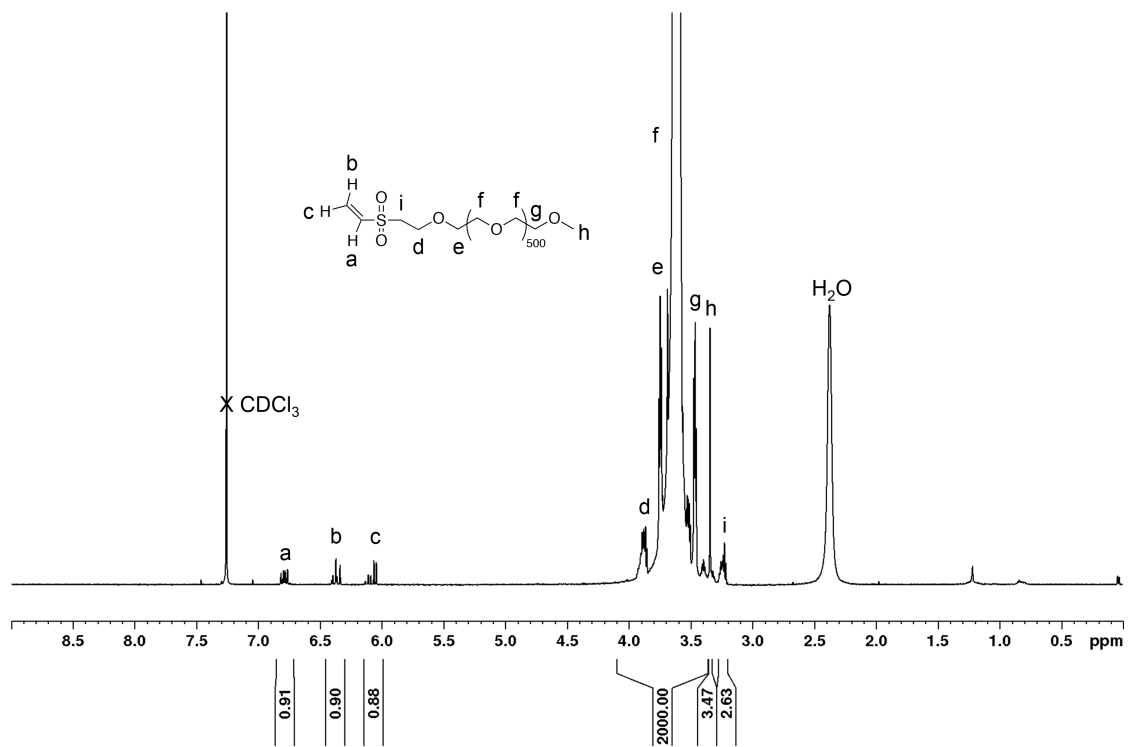


Figure S11. ^1H NMR spectrum of 20K VS-PEG (in CDCl_3).

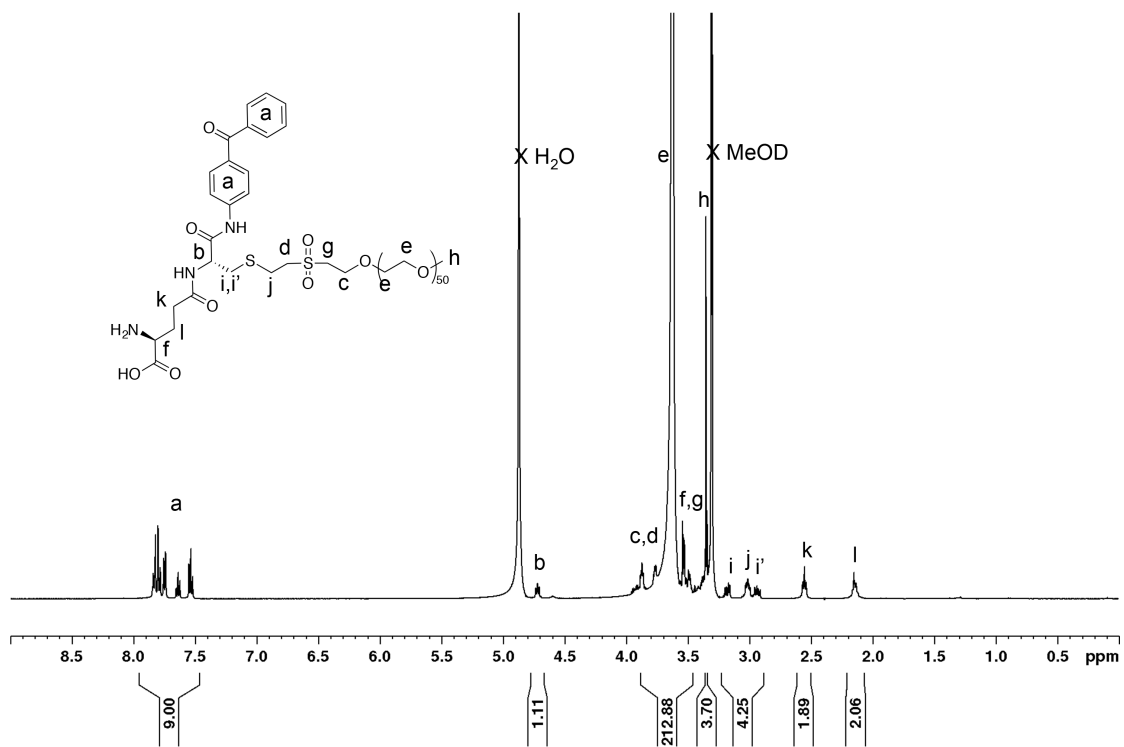


Figure S12. ^1H NMR spectrum of 2K GSBP-PEG (in MeOD).

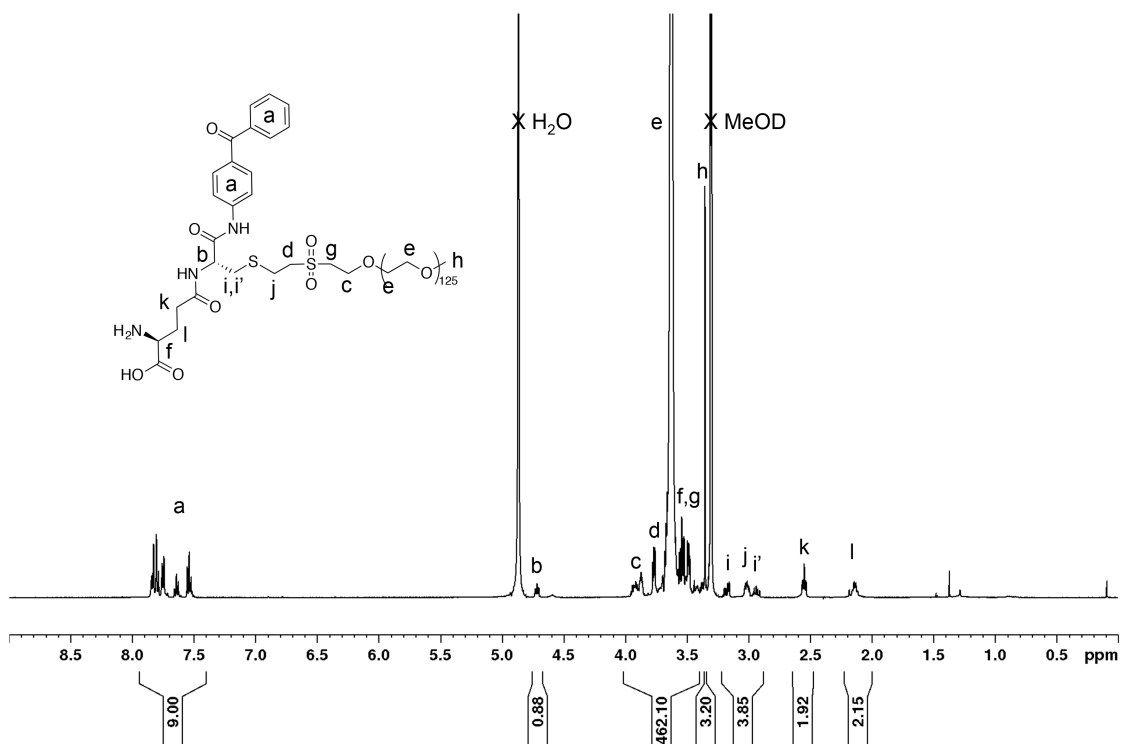


Figure S13. ^1H NMR spectrum of 5K GSBP-PEG (in CDCl_3).

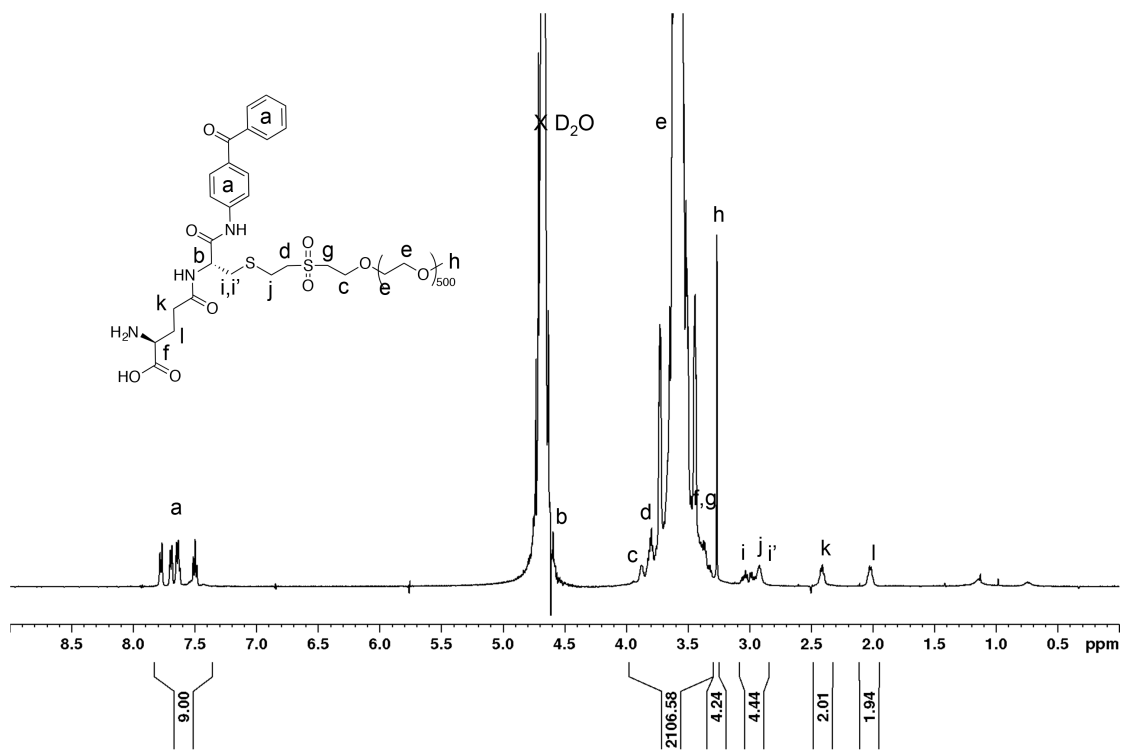


Figure S14. ¹H NMR spectrum of 20K GSBP-PEG (in MeOD).

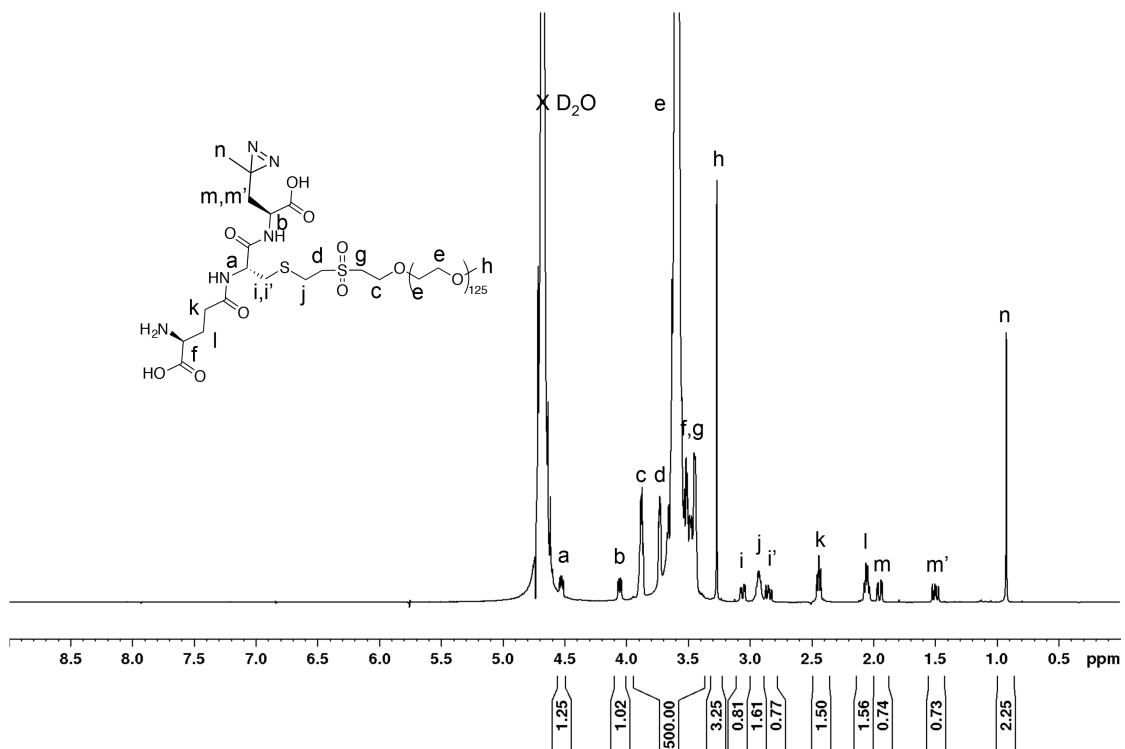


Figure S15. ¹H NMR spectrum of 5K GSDA-PEG (in D₂O).

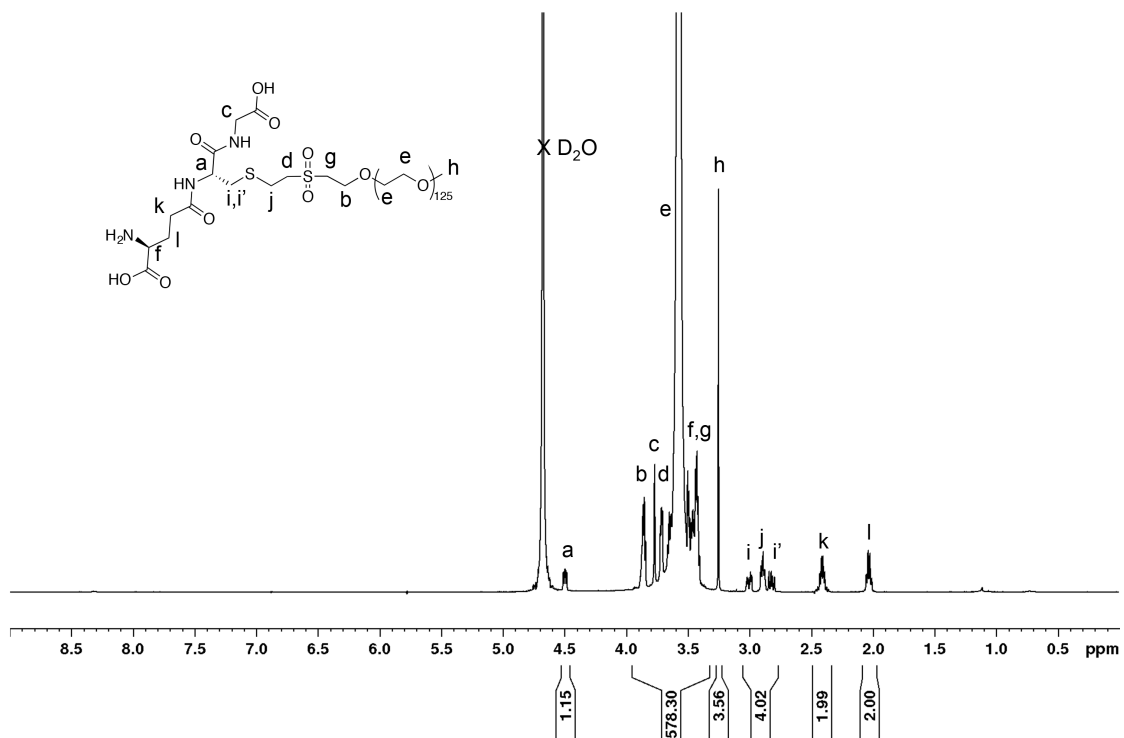


Figure S16. ¹H NMR spectrum of 5K GS-PEG (in D₂O).

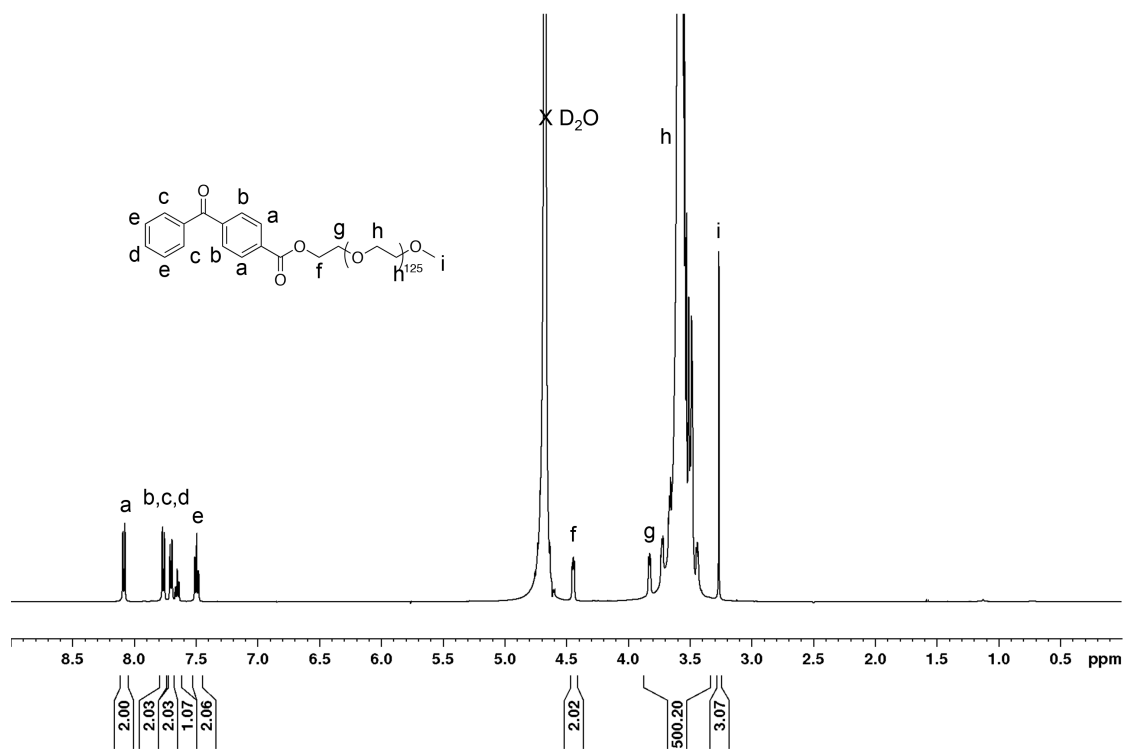


Figure S17. ^1H NMR spectrum of 5K BP-PEG (in D_2O).

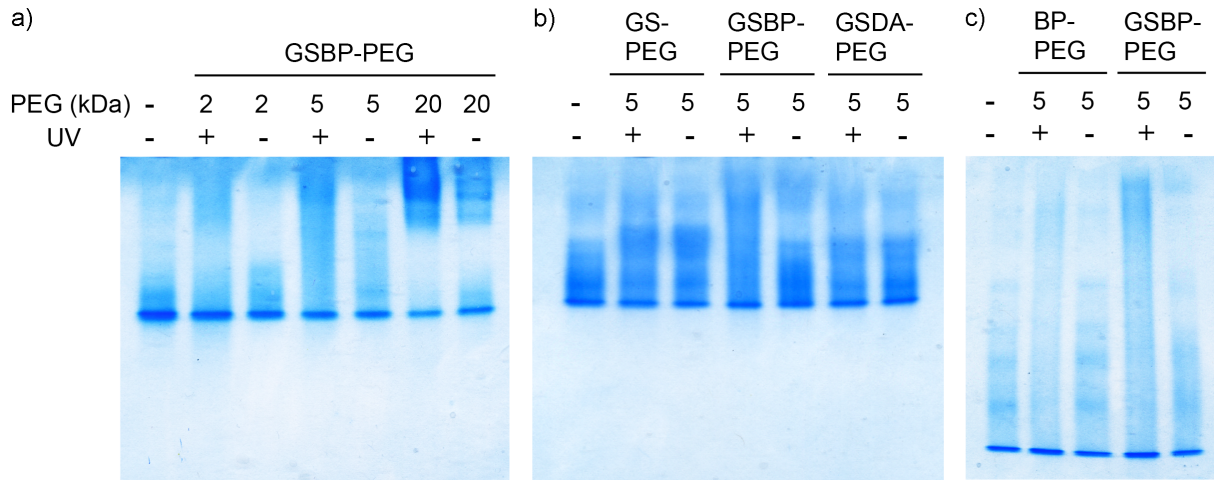


Figure S18. Native PAGE of a) GSBP-PEG conjugation to GST; b) comparison between GS-PEG, 5K GSBP-PEG, and GSDA-PEG; c) comparison between BP-PEG and 5K GSBP-PEG.

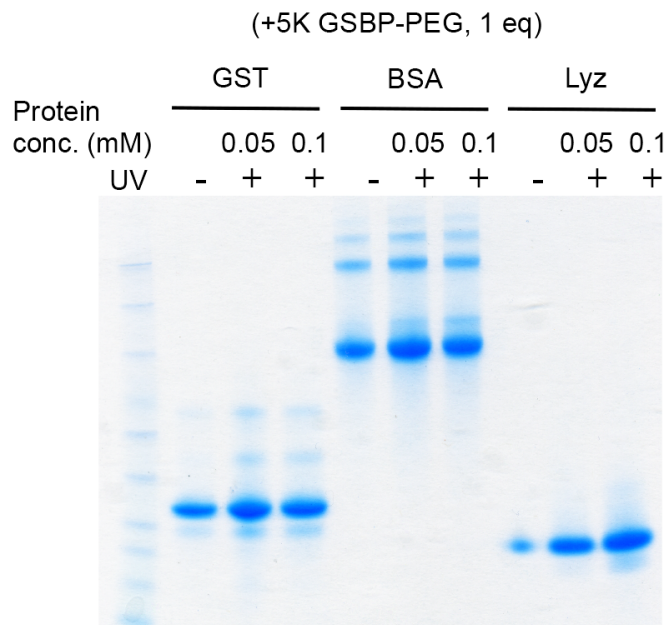


Figure S19. SDS-PAGE of the conjugation of 5K GSBP-PEG to GST, BSA, and Lyz at low concentrations. No conjugation is observed for Lyz.

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