

Supporting Information

Bimodal-hybrid heterocyclic amine targeting oxidative pathways and copper mis-regulation in Alzheimer's disease.

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Experimental: Details of Spectrophotometric determination of K (mM⁻¹) for **cyclen** and **1**.

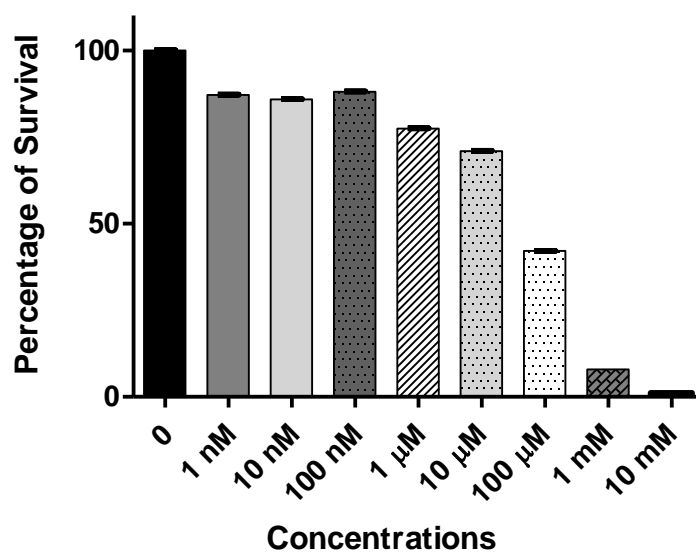


Figure S1. Screening for biocompatibility of **cyclexen** using the MTT assay, with 10 fold dilution concentrations, in HT-22 cell line.

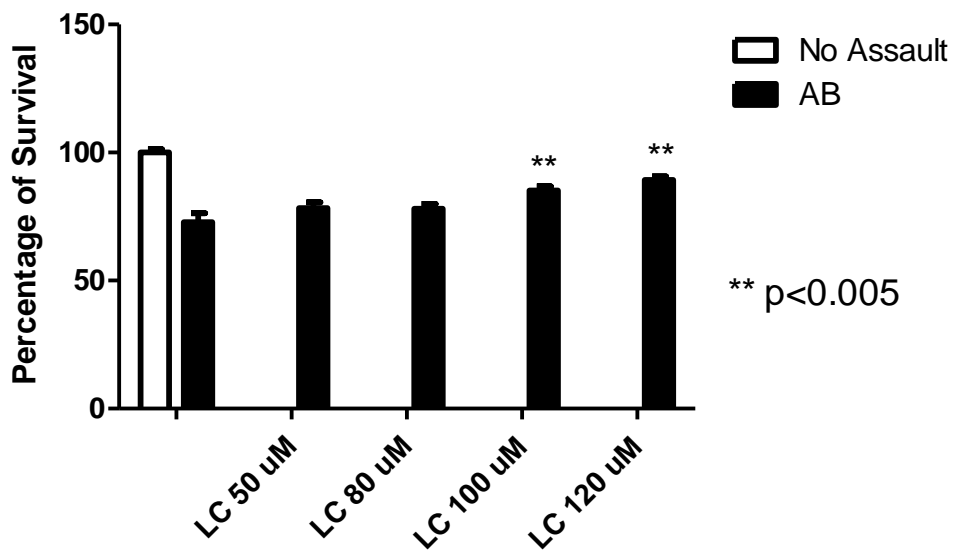


Figure S2. Protective effect of **1** against Aβ associated neurotoxicity in HT-22 cells in a 48 h. treatment using MTT assay. Aβ 15 μM; **1** 50, 80, 100, 120 μM.

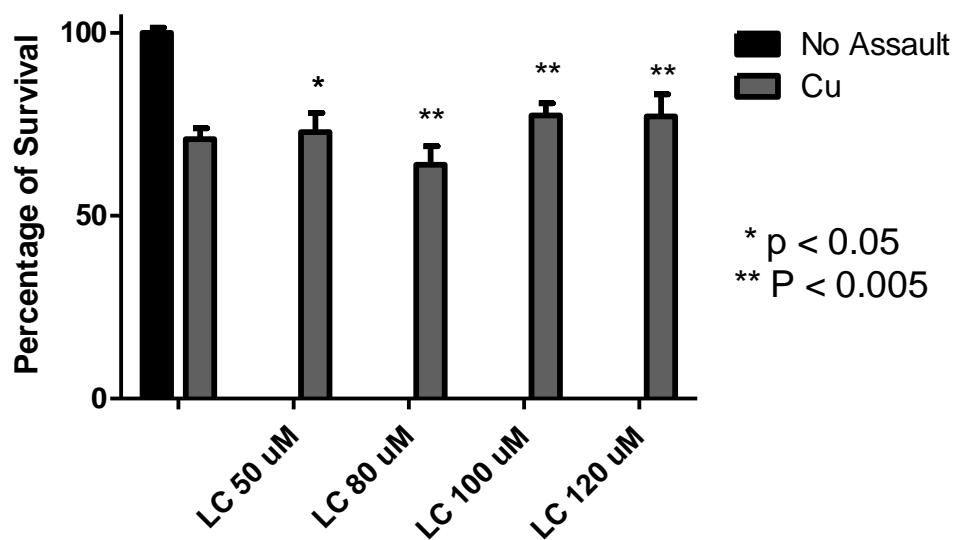
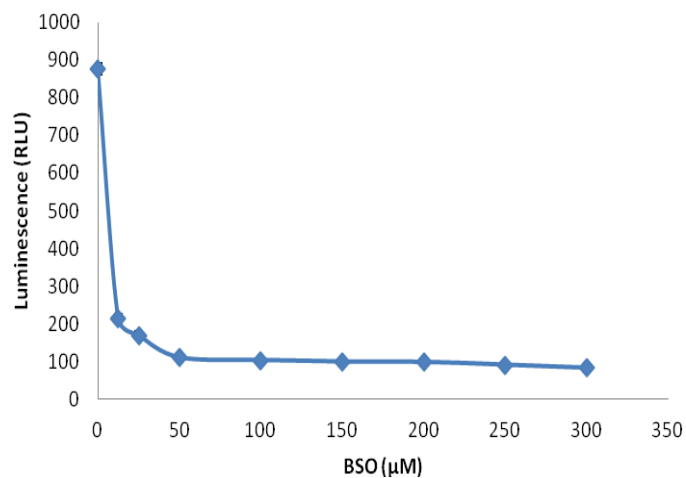
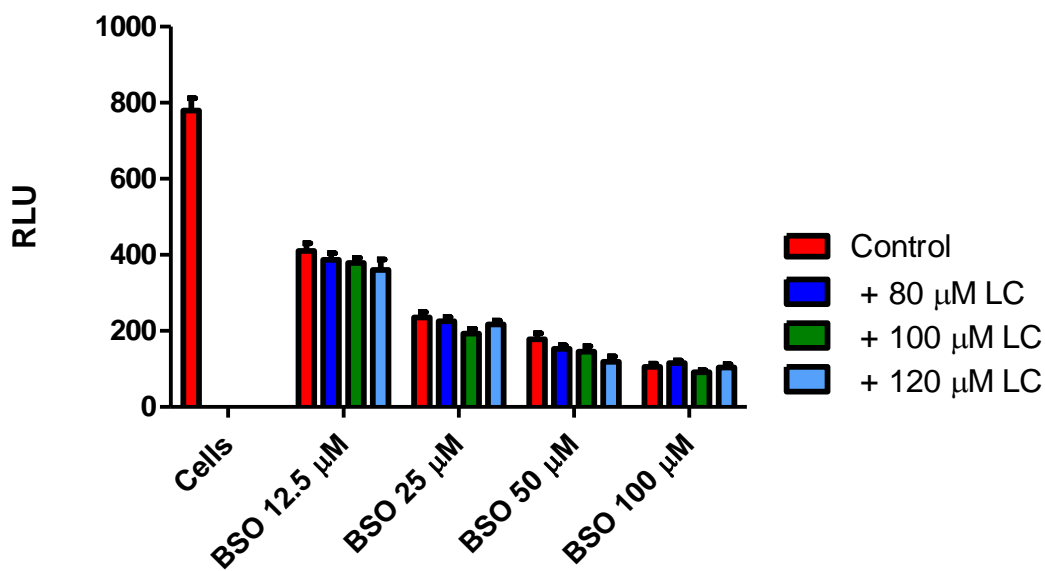


Figure S3. Protective effect of **1** against copper associated neurotoxicity in HT-22 cells in a 48 h. treatment using MTT assay. CuCl₂ 15 μM; **1** 50, 80, 100, 120 μM.



(a)



(b)

Figure S4. Evaluation of GSH levels in HT-22 Neuronal Cells measured by GSH-Glo™ Kit: (a) Treatment of HT-22 neuronal cells with BSO results in a dose dependent decrease in GSH. (b) The addition of **1** at 80, 100, 120 μM does not change GSH levels.

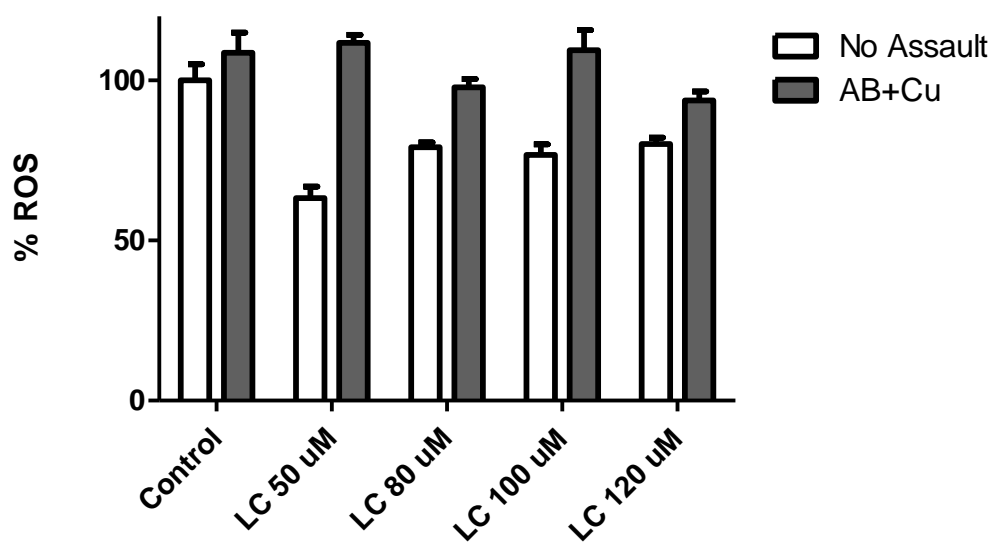


Figure S5. DCFH-DA Antioxidant assay of HT-22 neuronal cells after 12 hour exposure to A β + Cu [15 μ M each, final conc.] followed by addition of **1**. n=8 for each sample. (Note: DCFH-DA interacts with free copper ions to give unreliable readings, therefore this control was excluded.)

Log K _{ow} fragment description	Coefficient	Value obtained for 1
-CH2- [aliphatic carbon]	0.4911	3.9288
-NH- [aliphatic attach]	-1.4962	-5.9848
Equation Constant		0.2290
LogK_{ow}		-1.8270

Table S1. Calculated K_{ow} values to determine BBB permeability of **cyclen**.

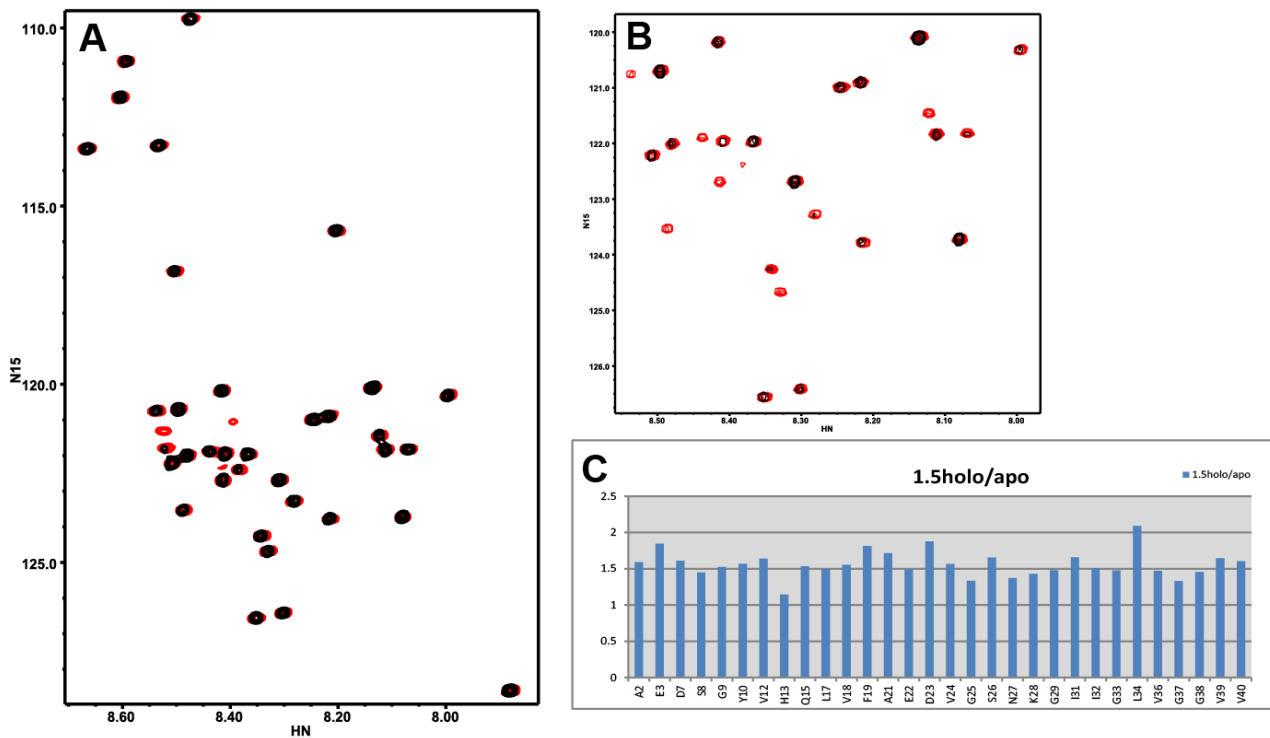


Figure S6. HSQC spectra of ^{15}N -A β_{1-40} (black) and ^{15}N -A β_{1-40} plus **1** (1.5 eq.) (red). (a) Full spectrum. (b) Expansion of the central region of the spectrum, plotted at higher contour levels than in panel (a) to emphasize the stronger intensities of the red cross-peaks compared to the corresponding black cross-peaks. (c) Changes in the HSQC cross-peak intensities of the A β_{1-40} peptide caused by addition of **1** (1.5 eq.). The ratios between the cross-peak intensities of the A β_{1-40} peptide plus 1.5 eq. of **1** (I_1) and those of the A β_{1-40} peptide alone (I_0) are plotted as a function of the residue number.

Details of Spectrophotometric determination of K (mM^{-1}) for cyclen and **1**.

Our data were examined using the one-to-one binding stoichiometry model: $\mathbf{M} + \mathbf{L} \rightleftharpoons \mathbf{ML}$, where \mathbf{M} , \mathbf{L} and \mathbf{ML} represent free copper, free ligand and copper-ligand complex, respectively. The binding constant, K , can then be expressed in terms of molar concentrations of each component:

$$K = \frac{[\mathbf{ML}]}{[\mathbf{M}][\mathbf{L}]} \quad (1)$$

where $[\mathbf{M}]$, $[\mathbf{L}]$ and $[\mathbf{ML}]$ are the corresponding molar concentrations. The total copper concentration, $C_{\mathbf{M}}$, is related to $[\mathbf{M}]$ by the mass balance $C_{\mathbf{M}} = [\mathbf{M}] + [\mathbf{ML}]$ and the total ligand concentration $C_{\mathbf{L}}$ to $[\mathbf{L}]$ by the mass balance $C_{\mathbf{L}} = [\mathbf{L}] + [\mathbf{ML}]$.

In order to express $[\mathbf{M}]$ as a function of K , $C_{\mathbf{M}}$ and $C_{\mathbf{L}}$, it is useful to start by relating the fraction of copper-ligand complex, $\left(\frac{[\mathbf{ML}]}{C_{\mathbf{L}}}\right)$, to the binding constant using equation 1 and the previous mass balances:

$$\frac{C_{\mathbf{M}} - [\mathbf{M}]}{C_{\mathbf{L}}} = \frac{K[\mathbf{M}]}{1 + K[\mathbf{M}]} \quad (2)$$

Equation 2 can be rearranged as a quadratic equation with respect to $[\mathbf{M}]$ and its positive root can be calculated:

$$[\mathbf{M}] = \frac{-(1 - KC_{\mathbf{M}} + KC_{\mathbf{L}}) + \sqrt{(1 - KC_{\mathbf{M}} + KC_{\mathbf{L}})^2 + 4KC_{\mathbf{M}}}}{2K} \quad (3)$$

At a given wavelength, the copper extinction coefficient, ε , in the presence of ligand, can be expressed as the weighted average between that of the free copper ions, $\varepsilon_{\text{free}}$, and bound copper, $\varepsilon_{\text{bound}}$, according to:

$$\varepsilon = \frac{[\mathbf{M}]}{C_{\mathbf{M}}} \varepsilon_{\text{free}} + \frac{[\mathbf{ML}]}{C_{\mathbf{M}}} \varepsilon_{\text{bound}} \quad (4)$$

In equation 4, it is useful to define $[\mathbf{M}]/C_{\mathbf{M}}$ as the fraction of free copper ions in solution, α_{free} . Therefore, we can rewrite equation 4 and express $\varepsilon/\varepsilon_{\text{free}}$ as a function of α_{free} .

$$\frac{\varepsilon}{\varepsilon_{\text{free}}} = \alpha_{\text{free}} + (1 - \alpha_{\text{free}})R \quad (3)$$

where and $R = \varepsilon_{\text{bound}}/\varepsilon_{\text{free}}$ and

$$\alpha_{\text{free}} = \frac{-(1 - KC_{\mathbf{M}} + KC_{\mathbf{L}}) + \sqrt{(1 - KC_{\mathbf{M}} + KC_{\mathbf{L}})^2 + 4KC_{\mathbf{M}}}}{2KC_{\mathbf{M}}} \quad (4)$$

The method of least squares (using KaleidaGraph software) based on equations 3 and 4 was applied to our experimental data to determine K and R (Table S3). The accuracy of the prepared **1** solutions concentration was assessed by substituting in equation (4) $C_{\mathbf{L}}$ with $fC'_{\mathbf{L}}$, where $C'_{\mathbf{L}}$ is the measured **1** concentration by weight and f is a corrective factor that takes into account that part of the total weighed material is impurity; hence, $f \leq 1$. This value of f is consistent with the actual concentration value extracted from NMR, being approximately 10% lower than that determined by sample weight.

Table S3. Fitting model parameters associated with copper-ligand binding.

	Cyclen ¹	1 ¹
K/mM^{-1}	>100	6.4±1.7
R	179±1	14.7±0.2
f	1.05±0.01	0.88±0.02

¹the uncertainties are standard deviations.