

Supplementary Information

Light-guiding hydrogels for cell-based sensing and optogenetic synthesis *in vivo*

Myunghwan Choi^{1,2}, Jin Woo Choi^{1,3}, Seonghoon Kim², Sedat Nizamoglu¹,
Sei Kwang Hahn^{1,4}, and Seok Hyun Yun^{1,2*}

¹ Harvard Medical School and Wellman Center for Photomedicine, Massachusetts General Hospital, Boston, Massachusetts, USA.

² WCU Graduate School of Nanoscience and Technology, Korea Advanced Institute of Science and Technology, Daejeon, Korea.

³ Wonkwang Institute of Interfused Biomedical Science, Department of Pharmacology, School of Dentistry, Wonkwang University, Seoul, Korea.

⁴ Department of Materials Science and Engineering, Pohang University of Science and Technology, Pohang, Korea.

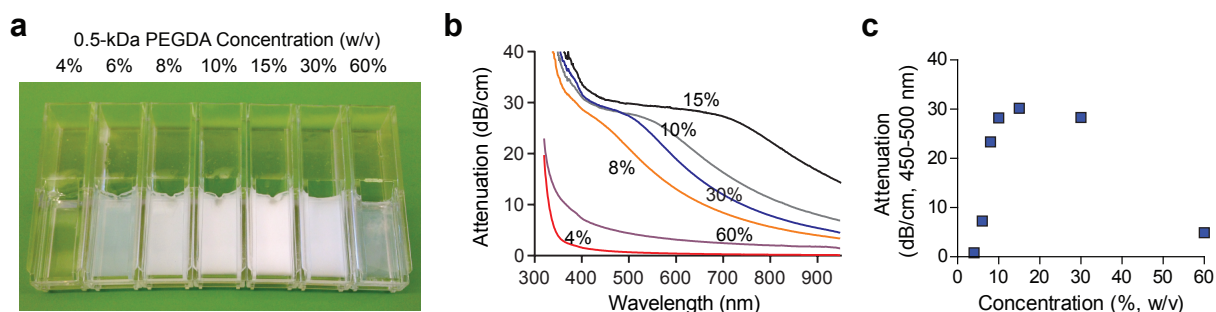
*Corresponding should be addressed to

S. H. Andy Yun, Ph.D.

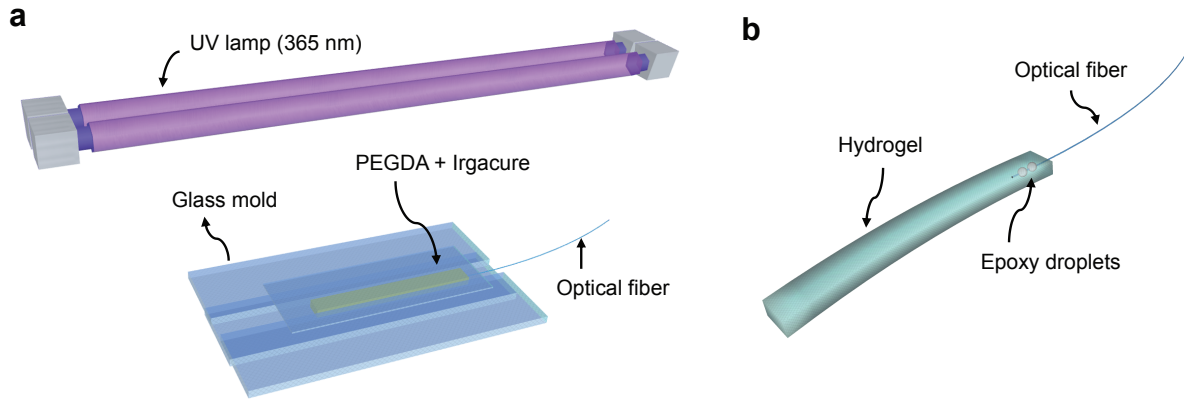
Associate Professor

65 Landsdowne St. UP-525, Cambridge, MA 02139, USA

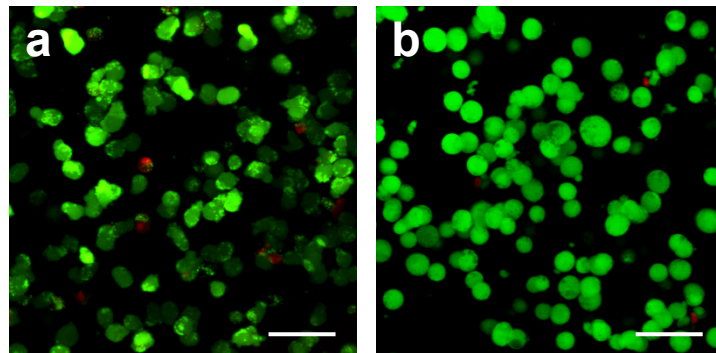
Email: syun@hms.harvard.edu



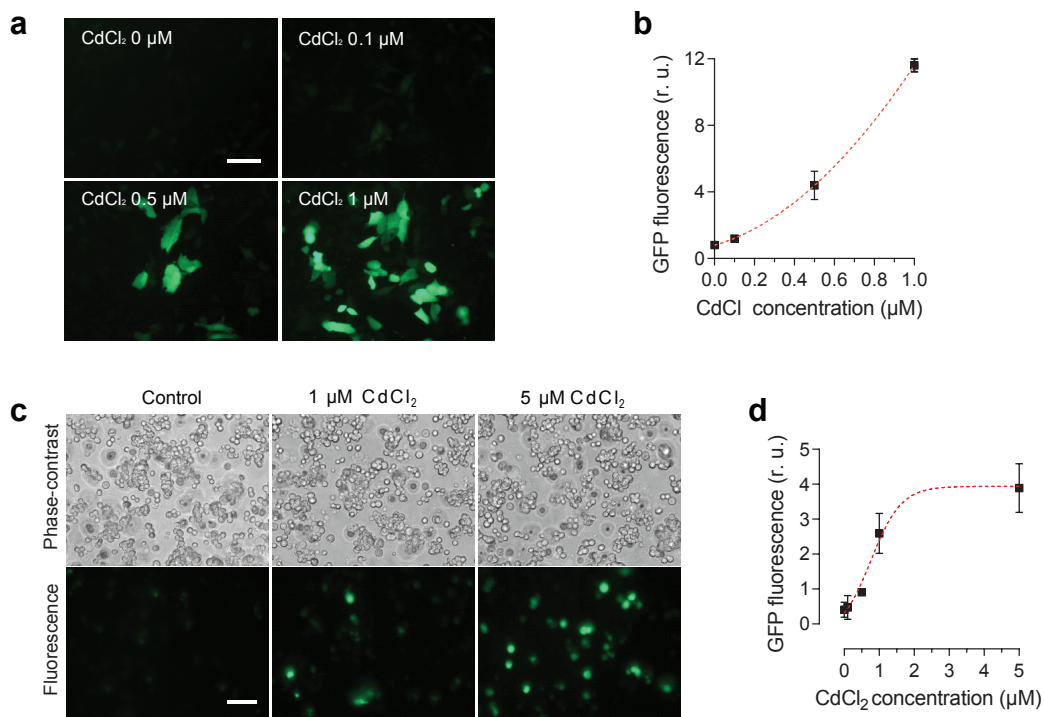
Supplementary Fig. S1 | Effect of the precursor concentration on optical transparency. **(a)** Photographs of PEG hydrogels at varying concentrations of PEGDA (0.5 kDa) in standard 1-cm-wide cuvettes. **(b)** Optical attenuation spectra. **(c)** Average attenuation coefficients averaged over a spectral range of 450-500 nm.



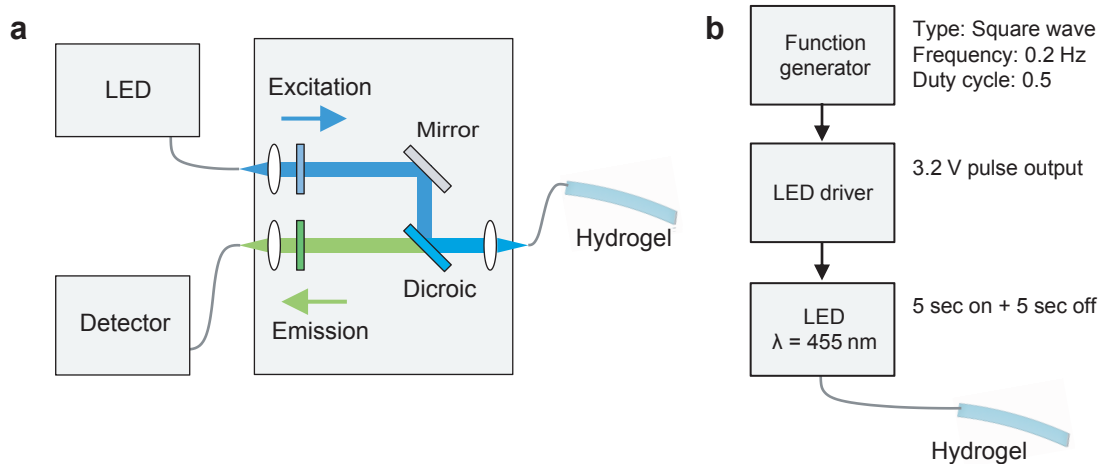
Supplementary Fig. S2 | Fabrication of a hydrogel optical waveguide. (a) Precursor solution containing PEG diacrylates (PEGDA) and photoinitiator (Irgacure) was photo-crosslinked *in situ* in a glass mold. (b) Schematic of the fabricated hydrogel optical waveguide.



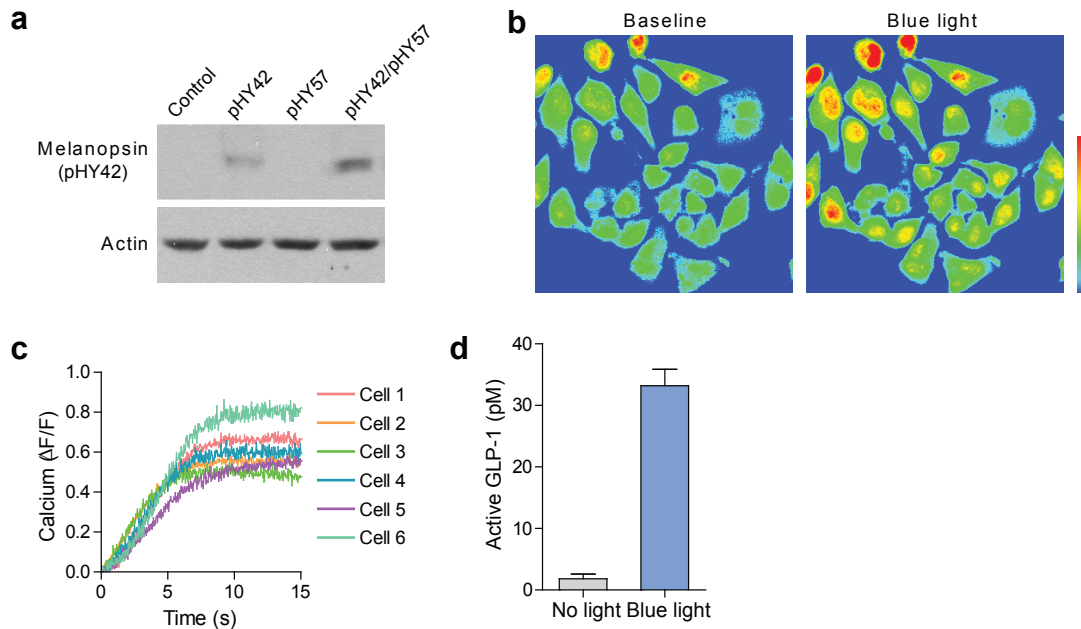
Supplementary Fig. S3 | Cell viability after hydrogel encapsulation. (a) HEK293 human embryonic kidney cell line. (b) EL4 mouse T cell line. Cells were encapsulated in PEG hydrogel through photopolymerization and cell viability was tested by staining with calcein AM (green; viable cells) and ethidium homodimer (red; dead cells). In the hydrogels, 96% of the HEK293 cells are live after encapsulation (a), and 97.5% of EL4 cells are live (b). Scale bar, 50 μm .



Supplementary Fig. S4 | Activation of heat-shock protein (hsp70) gene in response to cadmium ions. (a) Fluorescence images of the hsp-70-GFP sensing cells *in vitro*. (b) Dose-dependent activation of GFP fluorescence. (c) Phase contrast images and corresponding fluorescence images of the sensing cells in a hydrogel at 24 hours after CdCl₂ was added to the medium. (d) Dose-dependent activation of GFP signal *in vitro*.



Supplementary Fig. S5 | Schematic of the experimental setups for sending and receiving light to and from a hydrogel. (a) Setup for fluorescence sensing. A fiber-coupled blue LED ($\lambda = 455$ nm; excitation) was coupled to the hydrogel through the pigtail fiber and the fluorescence emission (500-550 nm) was collected to a photo-detector. (b) Setup for optogenetic therapy. To generate pulsed blue light, a light emitting diode (LED) was driven in a pulsed mode at 0.1 Hz.



Supplementary Fig. S6 | Stable cell line for light-induced GLP-1 secretion, produced with two plasmids named pHY42 (human melanopsin) and pHY57 (NFAT promoter driven GLP-1 expression). **(a)** Western blot analysis confirming the expression of melanopsin. **(b)** Fluorescence calcium-level images before and after illuminating blue light (10 s). The cells were preloaded with a fluorescent calcium indicator. **(c)** Time traces of the calcium signals in various cells. **(d)** The GLP-1 level in the cell media measured by ELISA before and after illuminating blue activation light.

Supplementary Video S1

A fully awake mouse with a hydrogel (4 mm x 1 mm x 40 mm) implanted in the subcutaneous pocket. Video was taken one day after implantation.