

## SUPPLEMENTARY ONLINE DATA



## Elucidation of the preferred routes of C8-vinyl reduction in chlorophyll and bacteriochlorophyll biosynthesis

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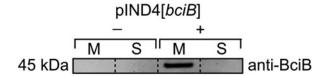


Figure S1 Expression of recombinant *bciB* in *R. sphaeroides* detected by Western blotting

Separated membrane (M) and soluble (S) fractions of  $\triangle bchCXF/\triangle bciA$  lacking ( - ) and containing ( + ) plND4[bciB] grown under expression-inducing conditions were separated by SDS/PAGE and transferred to a membrane that was probed with an anti-BciB antibody.

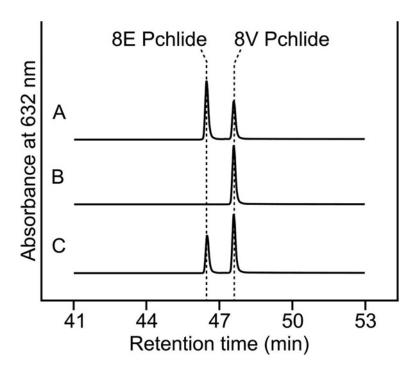


Figure S2 HPLC elution profiles of pigments extracted from *Synechocystis* strains lacking *chlB* 

HPLC elution profiles of extracts from pellets of (A)  $\Delta ch/B$ , (B)  $\Delta bciB/\Delta ch/B$  and (C)  $\Delta ch/B/\Delta bciB$ :: $bciA^{Rs}$  strains after 16 h growth without illumination. Traces are normalized to major peak height for clarity.

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Table S1 List of strains and plasmids described in the present study

Strain/plasmid	Properties	Source
E. coli		
JM109	Cloning strain for pK18 <i>mobsacB</i> and pIND4 constructs	Promega
S17-1	Conjugative strain for pK18 <i>mobsacB</i> and pIND4 constructs	[1]
R. sphaeroides		
WT	2.4.1	S. Kaplan*
V3	Unmapped mutant in a DPOR subunit-encoding gene	[2]
V3::bciB <sup>Syn</sup>	V3 harbouring pIND4[ <i>bciB</i> ]	The present stud
V3/∆bciA V3/∆bciA::bciB <sup>Syn</sup>	Unmarked deletion mutant of rsp_3070 in V3 V3/ $\Delta bciA$ harbouring pIND4[ $bciB$ ]	The present stud
$\Delta bchCXF$	Unmarked deletion mutant of bchC, bchX and bchF in WT	The present stud
$\Delta bchCXF/\Delta bciA$	Unmarked deletion mutant of rsp_3070 in △bchCXF	The present stud
∆bchCXF::bciB <sup>Syn</sup>	$\triangle bchCXF$ harbouring pIND4[ $bciB$ ]	The present stud
$\Delta bchCXF/\Delta bciA::bciB^{Syn}$	$\Delta bchCXF/\Delta bciA$ harbouring pIND4[ $bciB$ ]	The present stud
Synechocystis		
WT	sp. PCC6803	R. Sobotka†
∆bciB	Em <sup>R</sup> replacement of central portion of slr1923 in WT	[3]
∆bciB::bciA <sup>Rs</sup>	rsp_3070 and $Kan^R$ replacement of $psbAll$ in $\triangle bciB$	[3]
∆chlB	Zeo <sup>R</sup> replacement of central portion of slr0772 in WT	The present stud
$\Delta bciB/\Delta chlB$	$Zeo^R$ replacement of central portion of slr0772 in $\triangle bciB$	The present stud
$\Delta bciB/\Delta chlB::bciA^{Rs}$	$Zeo^R$ replacement of central portion of slr0772 in $\triangle bciB$ :: $bciA$	The present stud
Plasmid		
pK18mobsacB	Allelic exchange vector, <i>Km</i> <sup>R</sup>	J. Armitage‡
pIND4	IPTG-inducible expression vector for <i>R. sphaeroides</i> , <i>Km<sup>R</sup></i>	[4]
pIND4[bciB]	slr1923 cloned into the BamHI/HindIII sites of pIND4	The present stud
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## Table S2 List of primers used in the present study

Restriction enzyme cleavage sites used for cloning are underlined in the primer sequence.

Primer	Sequence $(5' \rightarrow 3')$	Cleavage site
rsp_3070UpF	CCGGAATTCGGACATCCTGACCGGTTTCCTGTCC	EcoRI
rsp_3070UpR	GCTCTAGAGGACATGGCGGAACTCCTCGGG	Xbal
rsp_3070DownF	GCTCTAGACGTTGACATTGGTGCCGGTCGG	Xbal
rsp_3070DownR	CCCAAGCTTCGAAGGCGATGCGCGCGAGGC	HindIII
rsp_3070CheckF	GACGACGAGAAGCTGGCCTACGG	
rsp_3070CheckR	GGCAGGTACCGGAGAGCGGTTAGG	
bchCXUpF	CCGGAATTCGCTCCTGCACCGGGTGCGCG	EcoRI
bchCXUpR	GCGCTCTAGAAGCGTTTTCCCCCGCGCTCTTC	Xbal
bchCXDownF	GCGCTCTAGATCTCGATACCCTCGCGCGGC	Xbal
bchCXDownR	CCCCAAGCTTGTCCTCGAACAGCTTGCCCGTG	HindIII
bchCXCheckF	GACGATCCACTGCCGCTCGG	
bchCXCheckR	CAGCGGCACGCCCGAGGCG	
bchFUpF	CCGGAATTCCCCCGCCCTGTCTCTCTGCAAGCC	EcoRI
bchFUpR	CGGGGGCGAAGGTCAAGGCTCATCTTGAGGTTCGCCTTCCGAGGAGGGCCC	
bchFDownF	CCTACGTCATCAACGCCGGGCAGTTCC	
bchFDownR	CCC <u>AAGCTT</u> GCGCCGAGAGCCGCTCGGCCCCGG	HindIII
bchFCheckF	GACGGCCACCGGGCCCTC	
bchFCheckR	GAGGCGAGGCATCCTC	
1923INDF	CGC <u>GGATCC</u> ACCGTTCCTGCCCCCCACC	BamHI
1923INDR	CCC <u>AAGCTT</u> TTATTGCTGGGGAAGTTTATACTGC	HindIII
chlBUpF	GCATCGCTTATTGTTCTCAACG	
chIBUpR	ACATTAATTGCGTTGCGCTCACTGCCGTGCATAATGGCGTGGACG	
chlBDownF	CAACTTAATCGCCTTGCAGCACATGCAGAATTGAATAAAGTGCCAGGG	
chlBDownR	CCTTCAAAGGCCATCACCC	
zeo <sup>R</sup> F	TGACCAGCGCCGTTCCGGTG	
zeo <sup>R</sup> R	CGGGTCGCGCAGGGCGAAC	

## **REFERENCES**

- 1 Simon, R., Priefer, U. and Pühler, A. (1983) A broad host range mobilization system for in vivo genetic engineering: transposon mutagenesis in Gram negative bacteria. Nat. Biotechnol. 1, 784–791 CrossRef
- 2 Nasrulhaq-Boyce, A., Griffiths, W. T. and Jones, O. T. G. (1987) The use of continuous assays to characterize the oxidative cyclase that synthesizes the chlorophyll isocycic ring. Biochem. J. 243, 23–29 PubMed
- 3 Canniffe, D. P., Jackson, P. J., Hollingshead, S., Dickman, M. J. and Hunter, C. N. (2013) Identification of an 8-vinyl reductase involved in bacteriochlorophyll biosynthesis in *Rhodobacter sphaeroides* and evidence for the existence of a third distinct class of the enzyme. Biochem. J. **450**, 397–405 <u>CrossRef PubMed</u>
- 4 Ind, A. C., Porter, S. L., Brown, M. T., Byles, E. D., de Beyer, J. A., Godfrey, S. A. and Armitage, J. P. (2009) Inducible-expression plasmid for *Rhodobacter sphaeroides* and *Paracoccus denitrificans*. Appl. Environ. Microbiol. **75**, 6613–6615 <u>CrossRef PubMed</u>

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