

Figure S1, related to Figure 1. (A) Quantification of the frequency of Foxp3⁺ Treg cells within indicated organs as in Figure 1A; (B) Quantification of frequency of Foxp3⁺Nrp1^{lo} iTreg cells or Foxp3⁺Nrp1^{hi} nTreg cells as in Figure 1B; (C) Activated WT naïve $CD4^+$ T cells were stimulated with combinations of TGF- β and recombinant galectin-9 with or without β -lactose (100mM) for 3 days. The frequency of Foxp3⁺ cells was determined by flow cytometry; Chimeric mice were generated by transferring WT or Lgals $9^{-/-}$ BM into (D-E) Lgals $9^{-/-}$ or (F-G) WT host mice. 10 weeks after reconstitution, the percentage of CD4⁺Foxp3⁺ Treg cells in mLN and LP was determined by flow and quantification; (H-I) Quantification of the frequency cvtometrv of CD45.2⁺CD4⁺Foxp3⁺ iTreg cells in PP and LP of recipient mice as in Figure 1F and Figure 1G. Data are representative of three independent experiments with $n \ge 4$ mice each group. *P < 0.05 (Student's *t*-test, error bars, SD).



Figure S2, related to Figure 2. (A) Protein expression level of galectin-9 was determined in $CD4^+$, $CD8^+$ T cells, $CD11b^+$ macrophages, and $CD19^+$ B cells by flow cytometry; (B) Quantitative real-time PCR analysis of the time course of the expression of *Lgals9* mRNA in iTreg cells; (C) Time course of galectin-9 protein expression in iTreg cells was determined by flow cytometry; (D) mRNA and (E) protein expression levels of Foxp3 and galectin-9 in naive $CD4^+$ T cells transduced with retrovirus expressing control vector (MIG) or Smad3-expressing vector and differentiated into Th0 or iTreg cells; Data

are representative of two independent experiments with $n \ge 5$ mice each group. **P* < 0.05 (Student's *t*-test, error bars, SD).



Figure S3, related to Figure 3. (A) Naïve *Foxp3^{GFP}* or *Foxp3^{GFP}Lgals9^{-/-}* T cells were differentiated into iTreg cells with TGF-β. The frequency of GFP (Foxp3⁺) cells was determined by flow cytometry; (B) Intracellular staining of IL-10 production by differentiated WT and *Lgals9^{-/-}* CD4⁺Foxp3⁺ iTreg cells was determined by flow cytometry; (C) WT and *Lgals9^{-/-}* iTreg cells were stimulated for 24 h with PMA and ionomycin, and IL-10 production in the supernatant was determined by ELISA; (D) Left: Colon lengths of *Rag2^{-/-}Lgals9^{-/-}* mice which had received the indicated cells for transfer as in **Figure 3C**, measured from the colocecal junction to the anal verge; Right: Quantification of pathological changes in the colon of mice as in **Figure 3C**; The data are representative of three independent experiments (A, B, D) or are pooled of three independent experiments (C) with n ≥ 4 mice each group. **P* < 0.05, ***P* < 0.01 (Student's *t*-test, error bars, SD).



Figure S4, related to Figure 4. (A) Left: Colon lengths of $Rag2^{-t}Lgals9^{-t}$ mice which had received the indicated cells for transfer as in Figure 4C, measured from the colocecal junction to the anal verge; Right: Quantification of pathological changes in the colon of mice as in Figure 4C 10 weeks after colitis induction; (B) Quantification of the frequency of RFP⁺ among CD4⁺ T cells in the LP of WT and $Lgals9^{-t}$ fate mapping mice; YFP expression within CD4⁺RFP⁺ T cells isolated from LP of indicated mice as in Figure 4C was determined by (C) flow cytometry and (D) quantification. Data are representative of two independent experiments with $n \ge 5$ mice each group. *P < 0.05(Student's *t*-test, error bars, SD).



Figure S5, related to Figure 5. Flow cytometry of protein expression levels of (A) Tim3 and (B) CD44 on indicated T cell subsets; (C) Immunoprecipitation (with control IgG, anti-galectin-9 or anti-CD44) of lysates of WT and $Cd44^{-/-}$ or $Lgals9^{-/-}$ iTreg cells, followed by immunoblot analysis with the indicated antibodies. Data are representative of three independent experiments with $n \ge 3$ mice each group.



Figure S6, related to Figure 6. (A) The level of phosphorylated Smad3 was assessed in WT and $Lgals9^{-/-}$ iTreg cells by immunoblot; (B) The binding of Smad3 to the *Foxp3*

CNS1 region in WT and Lgals9^{-/-} iTreg cells was determined by ChIP-PCR; (C) Foxp3 expression frequency and mean fluorescence intensity (MFI) from Foxp3^{GFP} and $Foxp3^{GFP}Lgals9^{-/-}$ iTreg cells; (D) Activated naïve CD4⁺ T cells were stimulated with TGF-β and/or recombinant galectin-9 for 2, 12 and 24 hours. *Foxp3* mRNA expression was assessed by quantitative real-time PCR analysis; (E) EL4 LAF cells were transfected with a Foxp3 promoter reporter construct containing the CNS1 enhancer and stimulated with anti-CD3 and anit-CD28 antibodies and TGF-β in the presence of increasing concentrations of galectin-9. Luciferase activity was measured 48 hours later; Chimeric mice were generated by transferring WT or $Foxp3^{\Delta CNSI}$ BM into WT or Lgals9^{-/-} host mice. 10 weeks after reconstitution, the frequency of $Foxp3^+$ Treg cells in thymus was determined by (F) flow cytometry and (G) quantification; (H) Quantification of the frequency of Foxp3⁺Nrp1^{lo} iTreg cells or Foxp3⁺Nrp1^{hi} nTreg cells in PP and LP as in Figure 6E. Data are representative of three independent experiments (A, C-D, F-H) or are pooled of three independent experiments (B, E) with $n \ge 4$ mice each group. *P < 0.05, **P < 0.01 (Student's *t*-test, error bars, SD).



Figure S7, related to Figure 7. (A) The expression of indicated co-stimulatory molecules on $Foxp3^{GFP}$ or $Foxp3^{GFP}Cd44^{-/-}$ CD4⁺GFP⁺ iTreg cells was determined by flow cytometry; (B) Left: Colon lengths of $Rag2^{-/-}$ mice which had received the indicated cells for transfer as in Figure 7E, measured from the colocecal junction to the anal verge; Right: Quantification of pathological changes in the colon of mice as in Figure 7E; (c) Hematoxylin and eosin staining of colon samples from the different groups as in Figure 7E 10 weeks after colitis induction (original magnification, ×20).

The data are representative of three independent experiments with $n \ge 5$ mice each group. *P < 0.05, (Student's *t*-test, error bars, SD).