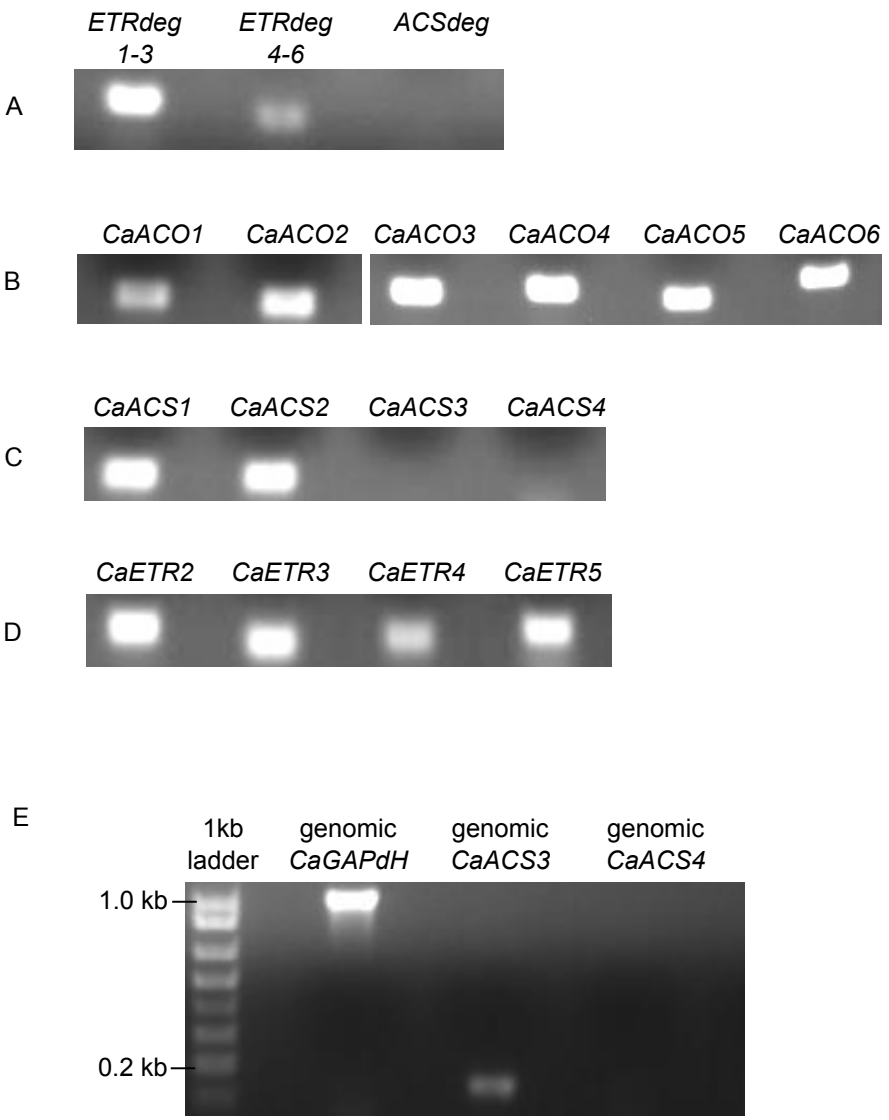

Additional file 3: Figure S2. End-point PCR for isolating capsicum *ACO*, *ACS* and *ETR* isoforms. Degenerate primers of *ETR* Type I (*ETRdeg1-3*) and Type II (*ETRdeg4-6*) as well as *ACS* isoforms (*ACSdeg*) were run in the end-point PCR (using cDNA pooled from six stages of ripening) to isolate other possible isoforms due to the lack of information in the databases (A). The primers for six *CaACO* isoforms (B), four *CaACS* isoforms (C) and four *CaETR* isoforms (D) were run in the end-point PCR using the pooled cDNA template. All bands are less than 0.26 kb using primers listed in Additional file 1: Table S1. The differences between band intensity (especially *CaACO1* and *CaETR4*) could be due to loading. The absence of *CaACS3* and *CaACS4* in the cDNA mix was also confirmed in another independent RT-PCR and qPCR experiments (data not shown). (E) Genomic PCR using DNA template (annealing temperature 60°C) was also performed but only the *CaACS3* (and positive control *CaGAPdH* plus introns) produced specific products and not *CaACS4*. Experiments were repeated in two other annealing temperatures (55°C and 65°C), again without any amplification of *CaACS4* (data not shown). *CaACS4* primers were obtained from a previous qPCR study in capsicum and tomato [14] and no other information regarding its sequence was available in either the NCBI or capsicum EST databases.