

Nonhuman Primate Model for the Study of Respiratory *Klebsiella pneumoniae* Infection

R. F. BERENDT,* G. L. KNUTSEN, AND M. C. POWANDA†

U.S. Army Medical Research Institute of Infectious Diseases, Fort Detrick, Frederick, Maryland 21701

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Squirrel monkeys were inoculated by the intratracheal inoculation of 700 *Klebsiella pneumoniae* organisms and developed lobar pneumonia in about 24 h. Characteristic clinical findings were fever, anorexia, and coughing. Laboratory findings included leukocytosis or leukopenia (with the latter more prominent in ultimately fatal infections), bacteremia, and shedding of bacteria into the pharynx. Infected monkeys showed increased plasma lysozyme activity as well as increased plasma ceruloplasmin, haptoglobin and α_1 -antitrypsin. The mortality rate was 60%, and the mean time of death was 50.5 h. Pathologically, the disease spread by means of Kohn's pores and other pathways that generally did not involve airways as a means of dissemination until about 30 h. Squirrel monkeys seem to be better models for human respiratory *K. pneumoniae* infection than rats or mice.

Mice and rats are the experimental animals most commonly employed for the study of respiratory *Klebsiella pneumoniae* infection (2, 3, 10). Clinical and biochemical studies are difficult to carry out on these animals, however, because of their small size. Recently, we have found that squirrel monkeys (*Saimiri sciureus*) developed pneumonia after intratracheal instillation of *Streptococcus pneumoniae* (4), and also were susceptible to *K. pneumoniae* administered by this route (1). Therefore, in anticipation of studies of aerosol therapy, we have carried out experiments designed to characterize the squirrel monkey model in greater detail. This report includes clinical, histopathological, and selected biochemical observations.

MATERIALS AND METHODS

Test organism. Techniques for growing, storing, and enhancing the virulence of the A-D strain of type 1 *K. pneumoniae* have been described previously (2). Inocula (1.5 ml each) for the infection of monkeys contained 600 to 800 viable organisms each and were prepared as previously described (1).

Test animals. Healthy, juvenile, male squirrel monkeys weighing from 0.5 to 1.0 kg were used. They were housed individually in wire-bar cages and allowed free access to commercial monkey chow and water. Their diet was supplemented with fresh fruit several times weekly. During experiments, fruit was eliminated, and each monkey was limited to six biscuits daily to facilitate estimation of food consumption.

Intratracheal inoculation. Intratracheal inoculation as a method of infecting monkeys has been de-

scribed previously (1). The dose administered (700 organisms) was determined to be approximately one 50% lethal dose (1).

Preparation of samples for histological examination and estimation of bacteria concentration in tissue. In a separate experiment to determine histopathology, a blood sample was obtained, monkeys were killed 6, 24, 30, or 48 h after infection by intravenous injection of pentobarbital, and the abdominal and thoracic cavities were opened aseptically. For estimation of bacterial concentration, samples of selected organs were removed aseptically, weighed, homogenized in Trypticase soy broth (TSB), and brought to a final volume of 4.0 ml. Appropriate dilutions were made in TSB and plated on Trypticase soy agar. After incubation, colonies were counted and concentrations calculated. The remaining portion of the organs was fixed in 10% neutral buffered Formalin, cut in 6- μ m sections, and stained by hematoxylin and eosin or Brown and Haup stains. Sections were then examined by light microscopy.

Clinical determinations. Once daily, 0.75 ml of heparin-treated blood was obtained from the saphenous vein for determination of total and differential leukocyte concentration, hematocrit, and bacteremia. Bacteremia was determined by spreading 0.1 ml of blood on the surface of a Trypticase soy agar plate, incubating for 18 h at 37°C, and counting the colonies produced. Additional daily determinations or observations included rectal temperature, respiratory rate, culturing pharyngeal swabs for *Klebsiella*, weight, activity, sneezing or coughing, dyspnea, and food consumption. Three base-line determinations were made on each of the above parameters, and testing continued for 8 days after infection. The volume of the daily blood sample was increased to 1.5 ml on 2 separate days before infection as well as on days 1, 2, 3, 6, and 8 for the determination of plasma lysozyme, α_1 -antitrypsin, ceruloplasmin, and haptoglobin. Prior experi-

† Present address: Biochemistry Branch, U.S.A. Institute of Surgical Research, Brooke Army Medical Center, Fort Sam Houston, TX 78234.

mentation indicated that this bleeding schedule would cause a 20% drop in hematocrit value and no other changes. Lysozyme activity was measured by the method of Osserman and Lawlor (8). The concentration of three other serum proteins was estimated by radial immunodiffusion procedures, using kits obtained from Behring Diagnostics (American Hoechst Corp., Somerville, N.J.). Although these kits were designed for determination of human proteins, preliminary investigation showed sufficient cross-reactivity with serum proteins of rhesus, cynomolgus, and squirrel monkeys to permit assay of proteins in these species.

Experimental design. Sixteen monkeys were infected and serially killed in groups of four at 6, 24, 30, and 48 h for histopathological examination and estimation of tissue bacterial concentrations. Three replicate groups of five monkeys each were infected after three base-line determinations and then were assessed for 8 days to evaluate clinical course of illness and mortality. Simultaneously, three groups of five monkeys each were sham inoculated with sterile Trypticase soy broth. Approximately 10 days elapsed between the end of one replicate and the beginning of the next. The results of replicate experiments were not different for each of the parameters under investigation, so the data from the three groups were combined.

RESULTS

Histological and bacteriological studies.

The concentration of *K. pneumoniae* in selected tissues at various times after infection is presented in Fig. 1. The 6-h period was arbitrarily chosen as a period early in infection; at 24 h, clinical signs were just becoming apparent, and at 30 h illness was clearly visible. The 48-h time was chosen because we anticipated that the first deaths would occur shortly thereafter. One of the four monkeys reserved for the 48-h period died 2 to 3 h early. The data from liver and kidney are not shown because concentrations in these organs were almost identical to those in the spleen at all time periods. The data indicated that bacteria multiplied in the lungs and were carried in small numbers by the blood to other tissues. Of interest was the relatively small number of bacteria isolated from blood even at 48 h. At 24 h, a few hundred bacteria were isolated from the brain and stomach of each of the four monkeys (not shown).

Histological examination of the four monkeys 6 h after inoculation with *K. pneumoniae* revealed minimal to mild lobular alveolitis that was restricted, in every instance, to a single diaphragmatic lobe. This lesion was characterized by an infiltration of alveolar spaces by a few neutrophils and extravasated red blood cells (Fig. 2A).

Upon gross examination 24 h after inoculation, the four monkeys examined had patchy grayish-

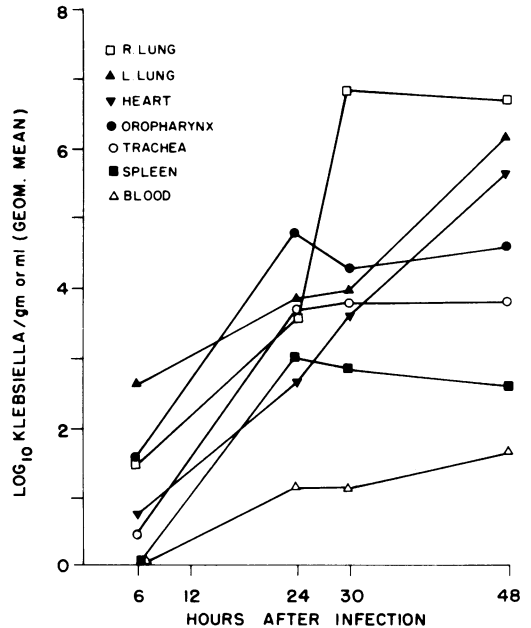


FIG. 1. Concentration of *K. pneumoniae* in selected tissues at selected times after intratracheal instillation. Values presented indicate the geometric mean of results from four monkeys examined at each time.

red areas of consolidation confined to the diaphragmatic lobe on one side. Histologically, the inflammation had progressed to an early moderate lobar pneumonia confined to a single lobe. Alveolar spaces, ducts, and some respiratory bronchioles in affected areas were filled with fluid, fibrin, and a cellular infiltrate composed primarily of neutrophils and a few macrophages (Fig. 2B). Numerous encapsulated gram-negative pleomorphic bacilli that were consistent with *K. pneumoniae* were present in the edematous exudate (inset). Interlobular septa were moderately distended with fibrinous fluid and were infiltrated with neutrophils. Lymphatic channels within the septa were moderately dilated.

Monkeys at 30 h after inoculation had total consolidation of one diaphragmatic lobe and patchy areas of involvement in the middle lobes of the same side. Microscopically, these monkeys had a lobar pneumonia that involved the entire diaphragmatic lobe and portions of the middle lobe. The lesion at this time was characterized by an intense outpouring of neutrophils, some macrophages, and a fibrinous edema that gave the lung parenchyma a solid appearance. The lesion was accompanied by foci of alveolar septal necrosis. Interlobular septa were markedly distended by an inflammatory exudate and con-

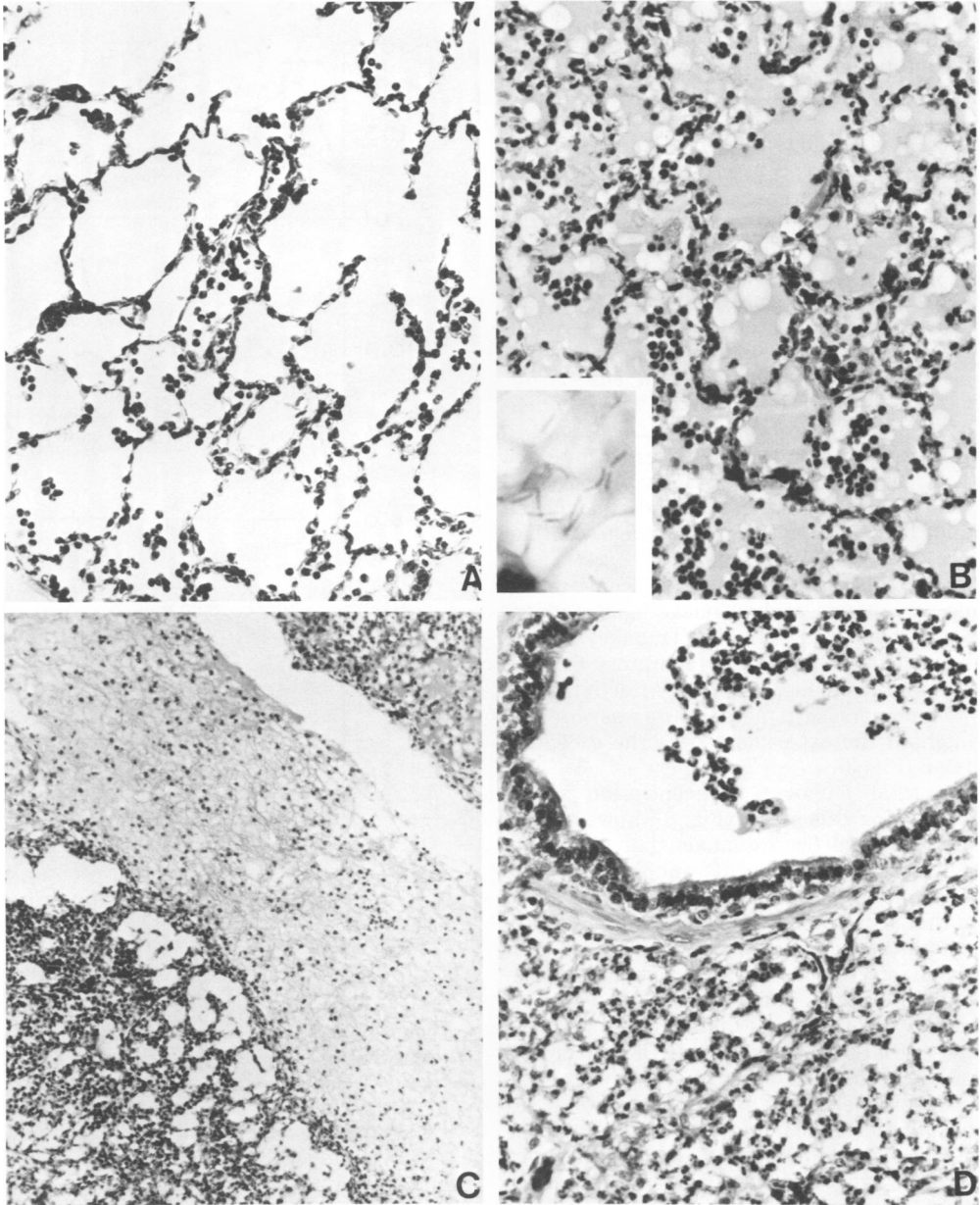


FIG. 2. (A) Infiltration of alveolar spaces by a few neutrophils and extravasated red blood cells (6 h postinfection). $\times 133$. (B) Alveolar spaces filled with edema fluid, fibrin, and a cellular infiltrate composed primarily of neutrophils and a few macrophages (24 h). $\times 133$. Inset: Gram-negative bacterial rods within alveolar exudate. $\times 665$. (C) Interlobular septa markedly distended by fibrinous edema and a neutrophilic infiltrate (24 h). $\times 133$. (D) Neutrophilic exudate within alveolar spaces and the lumen of a bronchiole without involvement of the wall of the airway (30 h). $\times 133$.

tained severely dilated lymphatics (Fig. 2C). The exudate continued onto the pleural surface of the affected lobes. In addition to the lobar pneumonia, one monkey in this group had an intense neutrophilic infiltrate within the lumen and

walls of bronchioles. The airways in other animals contained variable amounts of exudate, but the walls of these airways were generally uninvolved (Fig. 2D).

The inflammatory process at 48 h after inoc-

ulation was qualitatively similar to that seen at 30 h, but more extensive. A lobar pneumonia involving diaphragmatic and middle lobes with patchy areas of inflammation extending into the apical lobes was seen in two of the monkeys.

There was a mild to moderate splenitis in 5 of 12 monkeys observed 24 h or more after infection. Similar splenic changes are reported in rats experimentally infected with *K. pneumoniae* (2).

A patchy interstitial pneumonia was present in 13 of the 16 monkeys that were necropsied. This lesion was subacute to chronic in nature and was apparently unrelated to the experimental infection. In several monkeys, the interstitial involvement was associated with adult *Filaroides* spp. In only one monkey was *K. pneumoniae* found in association with this lesion.

Clinical and laboratory observations. All of the 15 noninfected control monkeys survived without overt illness, whereas 9 of the 15 infected animals (60%) died with a mean time to death of 50.5 h.

Illness, first observed at 24 h, consisted of lethargy, anorexia, and dyspnea. Signs of illness were pronounced in virtually all monkeys within 30 h. Principal clinical and laboratory findings are presented in Fig. 3 and 4. After infection, significant increase in temperature was observed throughout the experiment with the exception of day 5 (Fig. 3).

The total leukocyte concentration varied widely during this study (Fig. 3); however, the counts obtained from animals that ultimately died were significantly different than those of infected monkeys that survived (Table 1). Monkeys that survived the infection had leukocytosis; those that ultimately died, leukopenia. Virtually all infected monkeys showed marked neutrophilia on day 1 (not shown). This value returned to within normal limits in 2 to 3 days in monkeys that survived, but relative neutrophilia persisted in those that died.

Klebsiella cells were isolated from the pharynx of all inoculated monkeys by the second day (Fig. 3). The frequency of isolation declined thereafter, and pharyngeal cultures were negative for *K. pneumoniae* after day 5. Bacteria were isolated from peripheral blood samples from more than 80% of the monkeys 24 h after inoculation and from all monkeys on the third and fourth days. Bacteremia was not detected after day 6. A greater number of bacteria were detected in the blood of monkeys that died than in those that survived (Table 2).

Increased respiratory rates were also characteristic of this infection (Fig. 4), reaching peak values on days 2 to 4 and then slowly declining. All infected monkeys had inspiratory and expiratory dyspnea throughout the test period.

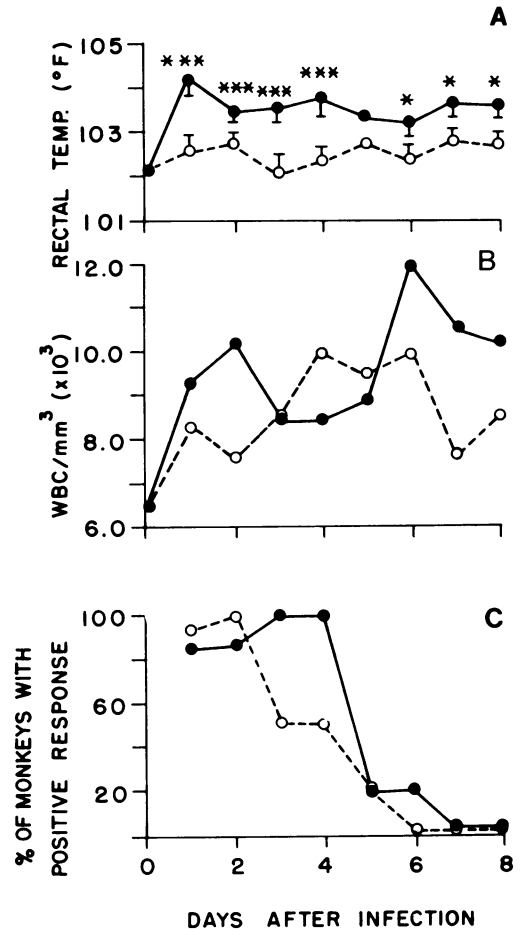


FIG. 3. (A) Rectal temperature of infected (●) and control (○) monkeys. Vertical bars indicate standard error of the mean and are shown only where significant differences exist. Symbols: (*) $P < 0.05$, (***) $P < 0.005$. (B) Total leukocyte (WBC) counts for infected (●) and control (○) monkeys. (C) Isolation of *Klebsiella* from blood (●) and throat (○).

Food consumption and body weight data are also shown in Fig. 4. The loss of weight was more pronounced than the anorexia, possibly indicating that some of the loss of weight was catabolic in origin.

Biochemical reactions. Plots of changes in four plasma proteins are presented in Fig. 5. Unfortunately, the variation between monkeys was very great, and statistical analysis often failed to discern differences. When the data were normalized to preinfection base lines, however, ceruloplasmin seemed to be the protein most affected by infection. Concentrations were significantly higher ($P < 0.05$) on the first day after infection and were still significantly elevated ($P < 0.005$) on day 8. The pattern for α_1 -antitrypsin

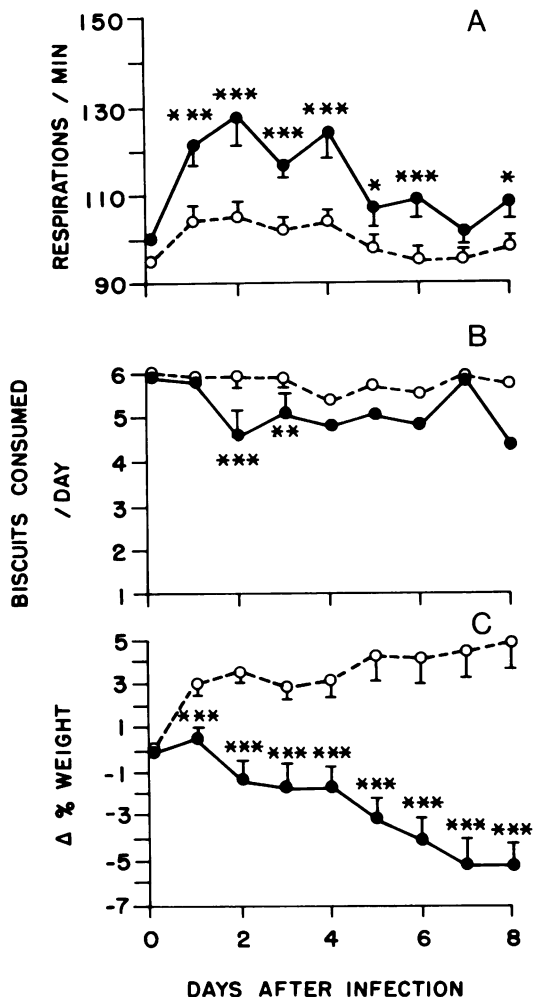


FIG. 4. (A) Respiratory rate in infected (●) and control (○) monkeys. Vertical bars indicate standard error of the mean. (B) Appetite as measured by biscuit consumption. (C) Body weight during infection. Symbols: (*) $P < 0.05$, (**) $P < 0.025$, (***) $P < 0.005$.

seems to be about the same as that of ceruloplasmin, but infected animals differed significantly from controls only on days 2, 3, and 6. The values for haptoglobin and lysozyme activity also were quite variable, but differed from controls on days 2 and 3. When the plasma protein values of surviving and dying monkeys were compared, only lysozyme exhibited a significant difference (Table 3), and this difference was seen only at one time period.

DISCUSSION

Intratracheally instilled *K. pneumoniae* (700 cells) caused an acute lobar pneumonia in squirrel monkeys. Clinically, the disease was charac-

terized by fever, anorexia, coughing, leukocytosis or leukopenia, bacteremia, and shedding of bacteria into the pharynx. The monkeys also showed increased concentrations of plasma α_1 -antitrypsin, ceruloplasmin, and haptoglobin, as well as increased plasma lysozyme activity. Mortality occurred in 60% of infected animals, with a mean time to death of about 50 h.

Pathologically, the disease was characterized by initial multifocal involvement of the alveoli (6 h) in the diaphragmatic lobe that spread to involve major portions of the same lobe by 24 h. The lesion at 24 h was still confined to peripheral tissues with involvement of alveolar spaces, ducts, and a few respiratory bronchioles. This observation suggests that initially the disease spread by way of Kohn's pores and other alveolar duct-bronchiolar pathways, including Lemberg's canals, rather than by the airways. Inflammatory exudate was not seen in larger bronchioles and bronchi until 30 h. Airways themselves were not involved. It is assumed that the spread of pneumonia to adjacent lobes occurred when the inflammatory exudate was either expelled up airways during respiration or moved by mucociliary action. Usually, involvement was unilateral, with the disease originating in the

TABLE 1. Total leukocyte concentration in dying and surviving monkeys during *K. pneumoniae* infection

Day after infection	Leukocytes/mm ³ ($\times 10^3$) ^a		P^b
	Surviving monkeys	Dying monkeys	
Base line	6,275 (6)	6,955 (9)	
1	12,633	6,522 (9)	<0.01
2	12,750	2,233 (3)	<0.01
3	8,117	2,900 (1)	
4	7,000	6,500 (1)	

^a Values in parentheses indicate number of monkeys tested.

^b Calculated by *t* test.

TABLE 2. Bacteremia in *K. pneumoniae*-infected monkeys

Day after infection	No. of bacteria/ml of blood		P^b
	Surviving monkeys	Dying monkeys	
1	3.9×10^1 (6)	5.6×10^2 (9)	<0.05
2	7.4×10^1 (6)	1.5×10^3 (3)	<0.05
3	2.3×10^2 (6)	5.4×10^2 (1)	
4	6.3×10^0 (6)	5.0×10^3 (1)	
5	0 (6)	No survivors	

^a Values represent geometric mean. Values in parentheses indicate number of monkeys tested.

^b Calculated by *t* test, computed on log transformation.

diaphragmatic lobes and extending in time to the apical lobe on the same side.

The most important question concerning this work is how well the disease in the model simulates that in humans. Clinically, the lobar form of the disease in humans as described by Julianelle (6) is characterized by abrupt onset, variable fever (frequency low), cough, extensive rusty sputum production, neutrophilia, and bacteremia in about 60% of patients. Extension of infection to other tissues is infrequent. Generally, our clinical and laboratory findings are consistent with these observations. We have not seen sputum production, but this lack may be due to difficulties in observation. Conversely, the observation that 100% of monkeys have bacteremia in contrast to 60% of human patients may be the result of more frequent blood sampling in the monkeys. The association of leukopenia and persistent bacteremia with mortality, as seen in monkeys, has also been reported in humans (6, 7).

When pathological lesions are considered, the similarity between human and simian models is even more striking. In both species, the lesions may be described as lobar pneumonia with severe edema of interlobular septa and distended lymphatic channels. Airways are relatively free of involvement early in the disease in humans (5, 11). The principal difference between the two species seems to be the low frequency of abscess formation in the squirrel monkey. Although the necrosis of alveolar septa, believed to be a pre-

TABLE 3. Lysozyme concentration in *K. pneumoniae*-infected monkeys

Day after infection	Plasma lysozyme concn ($\mu\text{g/ml}$) ^a		P ^b
	Surviving monkeys	Dying monkeys	
Base line	1.6 (6)	1.8 (9)	
1	2.2 (6)	2.4 (9)	
2	3.4 (6)	5.5 (3)	<0.05
3	2.6 (6)	4.6 (1)	
6	2.8 (6)	No survivors	
8	3.4 (6)		

^a Values in parentheses indicate number of monkeys tested.

^b Calculated by *t* test.

requisite for abscess formation in humans (5, 11), also occurs in monkeys, abscess formation was seen very rarely in our studies, even in monkeys in which the infection had caused death. The absence of abscess formation, however, may be due to the rapidity with which death occurred in monkeys.

Specific plasma proteins were measured in the expectation of using alterations in concentration as prognostic indices, as we have done earlier with rats (2). Too much variation was encountered among monkeys, however, to permit use of concentration estimates for this purpose, except with lysozyme. The changes that were observed, however, are consistent with the presence of inflammatory disease; the possible significance of these metabolic changes is discussed in detail elsewhere (9).

Although squirrel monkeys appear to be good models for acute pneumonia due to *K. pneumoniae*, the question of their utility compared to mice and rats arises. The low price and small size of small rodents is particularly appealing because more animals can be employed and, thus, statistical analysis can be more readily utilized. However, intranasal injection of organisms into mice and rats results in bronchopneumonia (2, 4) rather than in lobar pneumonia. Intratracheal instillation has also been reported to produce lobar pneumonia in rodents (10). However, the use of gastric mucin as an adjuvant in the rodent study (10) may have had profound effects. Squirrel monkeys, in contrast, mimic humans in the form that the *Klebsiella* pneumonia takes and, in addition, more readily allow the sequential measurement of conventional clinical signs such as fever, appetite, respiratory rate, weight change, blood and throat cultures, and hematology in the same animal than do rodents. Also, the cross-reactivity between certain human and monkey plasma proteins not only allows measurement of changes in specific acute-phase reactants, as opposed to fractions

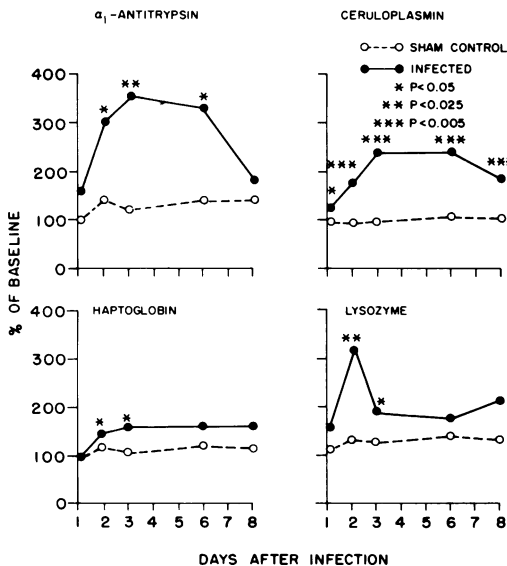


FIG. 5. Values of four selected plasma proteins during infection. Standard error bars omitted for clarity.

thereof, but may, assuming that the right protein or combination of proteins can be found, allow for additional quantitation of the severity of disease and the success of therapy.

With subhuman primate models for lobar pneumonia, we can now more thoroughly study the pathogenesis of such pneumonia, evaluate the effective uses of chemotherapy, and devise new approaches to therapy, both antimicrobial and supportive, with more confidence that our findings will be relevant to humans.

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