

## SUPPORTING INFORMATION

# **Quantification of fucosylated hemopexin and complement factor H in plasma of patients with liver disease.**

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**Table S-1**

<b>Protein UNIPROT #</b>	<b>Glycosite position</b>	<b>Glycopeptide</b>
Hemopexin <i>P02790</i>	N64	CSDGWSFDATTLDDN <sup>64</sup> GTMLFFK
	N187	SWPAVGN <sup>187</sup> CSSALR
	N240/N246	N <sup>240</sup> GTGHGN <sup>246</sup> STHHGPEYMR
	N453	ALPQPQN <sup>453</sup> VTSLLGCTH
CFH <i>P08603</i>	N217	SPDVIN <sup>217</sup> GSPISQK
	N529	LN <sup>529</sup> DTLDYEC HDGYESNTGSTTGSIVCGYNGWSDLPICYER
	N718	IQVDGEWTTLPVCIVEESTCGDIPELEHGWAQLSSPPYYYGDSVEFN <sup>718</sup> CSESFTMIGHR
	N802 /N822	WDPEVN <sup>802</sup> CSMAQIQLCPPPQIPNSHN <sup>822</sup> MTTTLNYR
	N882	IPCSQPPQIEHGTIN <sup>882</sup> SSR
	N911	ISEEN <sup>911</sup> ETTCYMGK
	N1029	MDGASN <sup>1029</sup> VTCINSR
	N1095	SPYEMFGDEEVMCLNGN <sup>1095</sup> WTEPPQCK

**Table S-1. Glycopeptides obtained by tryptic digest of hemopexin and CFH**

**Table S-2**

Protein		UNIPROT position	Site occupancy (%)		
<i>UNIPROT ID</i>	Peptide		H	CIR	HCC
Hemopexin <i>P02790</i>	CSDGWSFDATTLDDNGTMLFFK	N64	96	96	97
	SWPAVGNCSSALR	N187	100	100	100
	ALPQPQNVTSLLGCTH	N453	99	99	98
CFH <i>P08603</i>	SPDVINGSPISQK	N217	87	92	91
	IPCSQPPQIEHGTINSSR	N882	76	78	80
	ISEENETTCYMGK	N911	93	96	93
	MDGASNVTCINSR	N1029	99	99	99

**Table S-2. Site occupancy.** Site occupancy, shown as percent of glycosylated peptide, was determined in pooled samples (n=2 per group, 5 patient per pool) of healthy controls (H), cirrhosis (CIR) and HCC patients for all detected singly glycosylated sites as described in Material and Methods. The site occupancy of 100% means that non-glycosylated form of peptide was not detected. The results represent average of the two pooled samples. None of the sites changes significantly ( $p < 0.05$ ) in occupancy with disease state.

**Table S-3**

Protein <i>UNIPROT #</i>	Glycosite position	Glycan structure	m/z(charge)	H (SI)	H [%]	CIR (SI)	CIR [%]	HCC (SI)	HCC [%]
Hemopexin <i>P02790</i>	N64	A2G2S1	1111.1937 (4+)	10090±1337	8.3	7523±870	6.6	9351±432	6.6
		A2G2S2	1183.9676 (4+)	101335±9745	83.4	90072±9254	79.1	112117±11511	79.1
		A2G2S2F1	1220.4821 (4+)	10115±1223	8.3	16344±1185	14.3	*20354±1585	14.4
	N187	A2G2S1	1106.7872 (3+)	8001±990	4.0	8112±135	3.9	9075±1064	3.7
		A2G2S2	1203.8190 (3+)	80533±6953	40.7	92559±3187	44.3	101139±7309	41.7
		A2G2S2F1	939.6307 (4+)	13005±1674	6.6	26131±3548	12.5	*28105±3245	11.6
		A3G3	1131.4661 (3+)	7957±563	4.0	8503±928	4.1	10202±456	4.2
		A3G3S1	1228.4979 (3+)	11316±1164	5.7	12299±1071	5.9	11319±1345	4.7
		A3G3S2	994.3992 (4+)	17563±2235	8.9	*7834±332	3.8	11891±1093	4.9
		A3G3S2F1	1030.9137 (4+)	18037±1530	9.1	25218±1544	12.1	*30159±3621	12.4
A3G3S3		1067.1731 (4+)	21269±3309	10.8	*8823±998	4.2	12707±1617	5.2	
A3G3S3F1		1103.6876 (4+)	8790±1616	4.4	8174±751	3.9	14899±2212	6.1	
A4G4	1253.1768 (3+)	11247±1201	5.7	11232±1056	5.4	13119±605	5.4		
N453	A2G2S1	913.1450 (4+)	12156±549	5.5	9513±172	2.7	9832±973	2.8	
	A2G2S1F1	949.6595 (4+)	10077±894	4.6	16144±1866	4.6	20512±2518	5.9	
	A2G2S2	985.9189 (4+)	180111±19486	82.0	270251±19091	77.3	260039±27066	74.2	
	A2G2S2F1	1022.4333 (4+)	17368±2316	7.9	*53664±7335	15.4	*60098±6294	17.1	
CFH <i>P08603</i>	N217	A2G2S2	887.3742 (4+)	20173±1864	93.5	16092±1863	83.8	21153±1992	82.4
		A2G2S2F1	923.8887 (4+)	1406±122	6.5	*3102±182	16.2	*4529±486	17.6
N882	A2G2	911.6480 (4+)	28007±2505	6.7	*16110±1860	5.3	*16998±1631	4.4	
	A2G2S1	984.4218 (4+)	10029±947	2.4	18205±2320	6.0	16005±1907	4.2	
	A2G2S2	1057.1957 (4+)	350215±34546	83.7	*220038±16664	72.7	300138±23621	78.5	
	A2G2S2F1	1093.7102 (4+)	21312±2728	5.1	36216±3892	12.0	35835±3905	9.4	
	A3G3S2	1148.4787 (4+)	2506±190	0.6	3431±294	1.1	*4112±385	1.1	
	A3G3S2F1	1184.9932 (4+)	1009±124	0.2	*3837±514	1.3	*3061±337	0.8	
	A3G3S3	1221.2526 (4+)	5546±432	1.3	5017±293	1.7	5998±309	1.6	
N911	A2G2S1	869.5878 (4+)	14141±1915	3.1	7900±925	2.0	13006±1725	3.0	
	A2G2S1F1	906.1023 (4+)	15034±1502	3.3	*30171±2637	7.6	*35208±3704	8.0	
	A2G2S2	942.3617 (4+)	420406±43788	91.9	320524±37147	80.3	351044±41205	80.0	
	A2G2S2F1	978.8762 (4+)	908±105	0.2	*32353±3951	8.1	*30213±1692	6.9	
	A3G3S2	1033.6447 (4+)	6008±893	1.3	6025±760	1.5	8121±1107	1.8	
	A3G3S3	1106.4186 (4+)	1081±25	0.2	*2107±184	0.5	#1415±130	0.3	
N1029	A2G2S1	835.3315 (4+)	18064±1954	10.4	20144±2774	12.1	16928±705	9.3	
	A2G2S1F1	871.8460 (4+)	4549±743	2.6	*10108±1380	6.1	*10975±515	6.0	
	A2G2S2	908.1054 (4+)	145238±22655	83.3	120541±15146	72.6	141714±18929	77.8	
	A2G2S2F1	944.6199 (4+)	6018±383	3.5	*12010±1366	7.2	9947±1106	5.5	
	A3G3S2	999.3884 (4+)	503±29	0.3	*3226±201	1.9	*2605±188	1.4	

**Table S-3. Relative intensities of identified site-specific glycoforms of tryptic peptides of HPX and CFH.** Glycopeptide signal intensities (SI) in pooled samples of healthy controls (H), cirrhosis (CIR) and HCC patients (2 pools per group, 5 patients per pool) are presented as peak area of precursor ion XIC normalized to the intensity of a stable non-glycosylated tryptic peptide (GGYTLVSGYPK for hemopexin and SSNLIILEEHLK for CFH). Percent value indicates the relative distribution of individual glycoforms at a particular glycosylation site. Results are shown as mean  $\pm$  SD, \* P<0.05 vs H, # P<0.05 HCC vs CIR

**Table S-4**

Protein <i>UNIPROT #</i>	Glycosite position	Glycan structure	m/z(charge)	H (SI)	H [%]	CIR (SI)	CIR [%]	HCC (SI)	HCC [%]
Hemopexin <i>P02790</i>	N64	A2G2	1038.4199 (4+)	115256±8208	91.2	1324541±30205	84.4	145095±13404	87.4
		A2G2F1	1074.9344 (4+)	7370±812	5.8	*18683±2315	11.9	14779±1859	8.9
		A3G3	1129.7029 (4+)	2561±559	2.0	848±81	0.5	1044±70	0.6
		A3G3F1	1166.2174 (4+)	1186±95	0.9	4951±1737	3.2	5077±1889	3.1
	N187	A2G2	1009.7554 (3+)	468677±54669	79.9	365518±19391	75.4	420926±76770	72.4
		A2G2F1	1058.4413 (3+)	62393±2179	10.6	63878±6919	13.2	88403±16542	15.2
		A3G3	1131.4661 (3+)	35557±17850	6.1	21607±3622	4.5	29680±2072	5.1
		A3G3F1	1180.1521 (3+)	14796±933	2.5	22069±1614	4.5	*30220±2899	5.2
		A3G3F2	1228.8380 (3+)	840±63	0.1	2406±465	0.5	*3071±335	0.5
		A4G4	940.1346 (4+)	3771±1498	0.6	6923±5556	1.4	6238±3513	1.1
		A4G4F1	976.6491 (4+)	376±134	0.1	2677±3255	0.6	2724±2575	0.5
	N453	A2G2	1120.1589 (3+)	913342±80225	96.3	886004±57384	93.7	868559±88078	92.5
A2G2F1		1168.8449 (3+)	16337±3811	1.7	34644±11919	3.7	45813±7939	4.9	
A3G3		1241.8697 (3+)	16663±7058	1.8	16466±326	1.7	15580±1794	1.7	
A3G3F1 <sup>@</sup>		1290.5556 (3+)	2288±488	0.2	*8233±894	0.9	*8745±1237	0.9	
CFH <i>P08603</i>	N217	A2G2	741.8265 (4+)	16722±560	91.6	12472±270	86.1	11888±4332	89.4
		A2G2F1	1074.9344 (4+)	1532±321	8.4	2012±241	13.9	1416±856	10.6
	N882	A2G2	911.6480 (4+)	213380±87237	85.2	167264±37896	71.1	424843±7208	70.1
		A2G2F1	948.1625 (4+)	16348±7626	6.5	38123±6225	16.2	**101879±13057	16.8
		A3G3	1002.9310 (4+)	14419±117	5.8	12298±4205	5.2	**31377±1302	5.2
		A3G3F1	1039.4455 (4+)	6278±1555	2.5	17541±7113	7.5	**48017±2769	7.9
	N911	A2G2	1062.0827 (3+)	116372±13814	83.7	102527±3449	69.1	124283±39793	66.1
		A2G2F1	1110.7687 (3+)	7297±1760	5.2	19919±4302	13.4	24900±8858	13.2
		A3G3	888.0970 (4+)	9895±1617	7.1	11834±2846	8.0	16746±938	8.9
		A3G3F1	924.6115 (4+)	5492±1478	3.9	14086±490	9.5	*22209±5271	11.8
	N1029	A2G2	762.5577 (4+)	33755±8178	90.9	31458±859	80.0	35627±20698	76.3
		A2G2F1	799.0722 (4+)	3376±513	9.1	7876±1058	20.0	11074±7923	23.7

**Table S-4. Relative intensities of site-specific glycoforms in de-sialylated pooled samples from healthy controls (H), cirrhosis (CIR) and HCC patients.** Glycopeptide signal intensities (SI) are presented as peak area of precursor ion XIC normalized to the intensity of internal control non-glycosylated peptide (GGYTLVSGYPK for hemopexin and SSNLIILEEHLK for CFH). Percent value indicates the relative distribution of individual glycoforms at a particular glycosylation site. Results are shown as mean ± SD, \* P<0.05 vs H, # P<0.05 HCC vs CIR, @ Structure out of fragmentation range confirmed by retention time and exact mass

**Table S-5**

<b>Protein</b> <i>UNIPROT ID</i>	<b>Peptide</b>	<b>UNIPROT</b> <b>position</b>	<b>Glycan</b> <b>structure</b>	<b>Precursor</b> <b>m/z (charge)</b>
Hemopexin <i>P02790</i>	CSDGWSFDATTLDDNGTMLFFK	N64	A2G2	1038.4 (4+)
			A2G2F1	1074.9 (4+)
			A3G3	1129.7 (4+)
			A3G3F1	1166.2 (4+)
	SWPAVGNCSSALR	N187	A2G2	1009.7 (3+)
			A2G2F1	1058.4 (3+)
			A3G3	1131.4 (3+)
			A3G3F1	1180.1 (3+)
			A3G3F2	1228.8 (3+)
			A4G4	940.1 (4+)
			A4G4F1	976.7 (4+)
			A4G4F2	1013.2 (4+)
	ALPQPQNVTSLLGCTH	N453	A2G2	1120.1 (3+)
A2G2F			1168.8 (3+)	
CFH <i>P08603</i>	SPDVINGSPISQK	N217	A2G2	741.8 (4+)
			A2G2F	778.3 (4+)
			A3G3	833.1 (4+)
			A3G3F	869.6 (4+)
	IPCSQPPQIEHGTINSSR	N882	A2G2	911.6 (4+)
			A2G2F	948.1 (4+)
			A3G3	1002.9 (4+)
			A3G3F	1039.4 (4+)
	ISEENETTCYMGK	N911	A2G2	1062.1 (3+)
			A2G2F	1110.7 (3+)
			A3G3	888.1 (4+)
			A3G3F	924.6 (4+)
	MDGASNVTCINSR	N1029	A2G2	762.6 (4+)
A2G2F			799.1 (4+)	

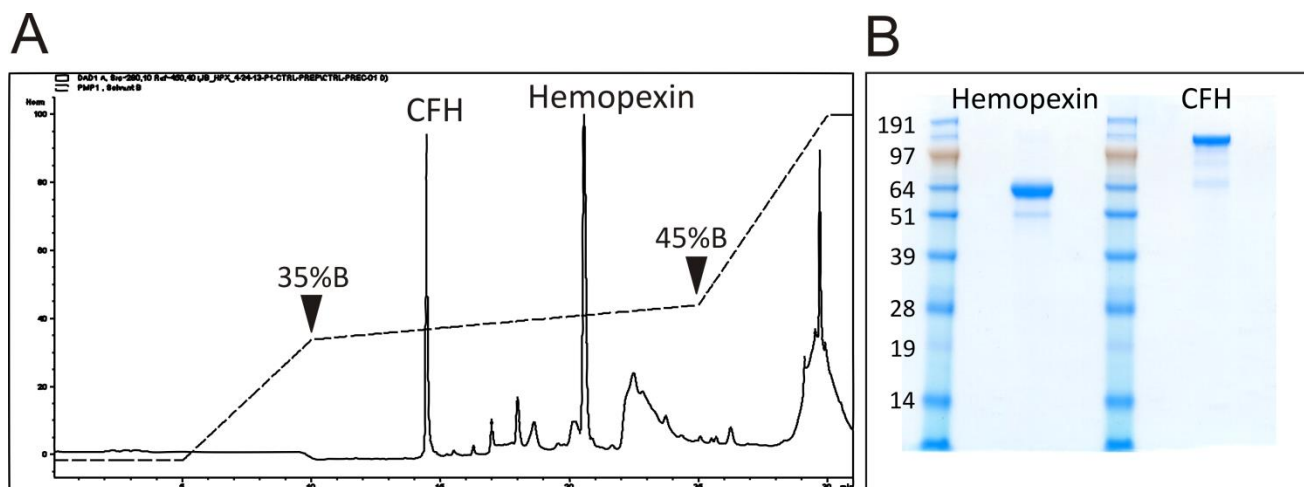
**Table S-5.** List of precursor ions selected for MRM transitions.



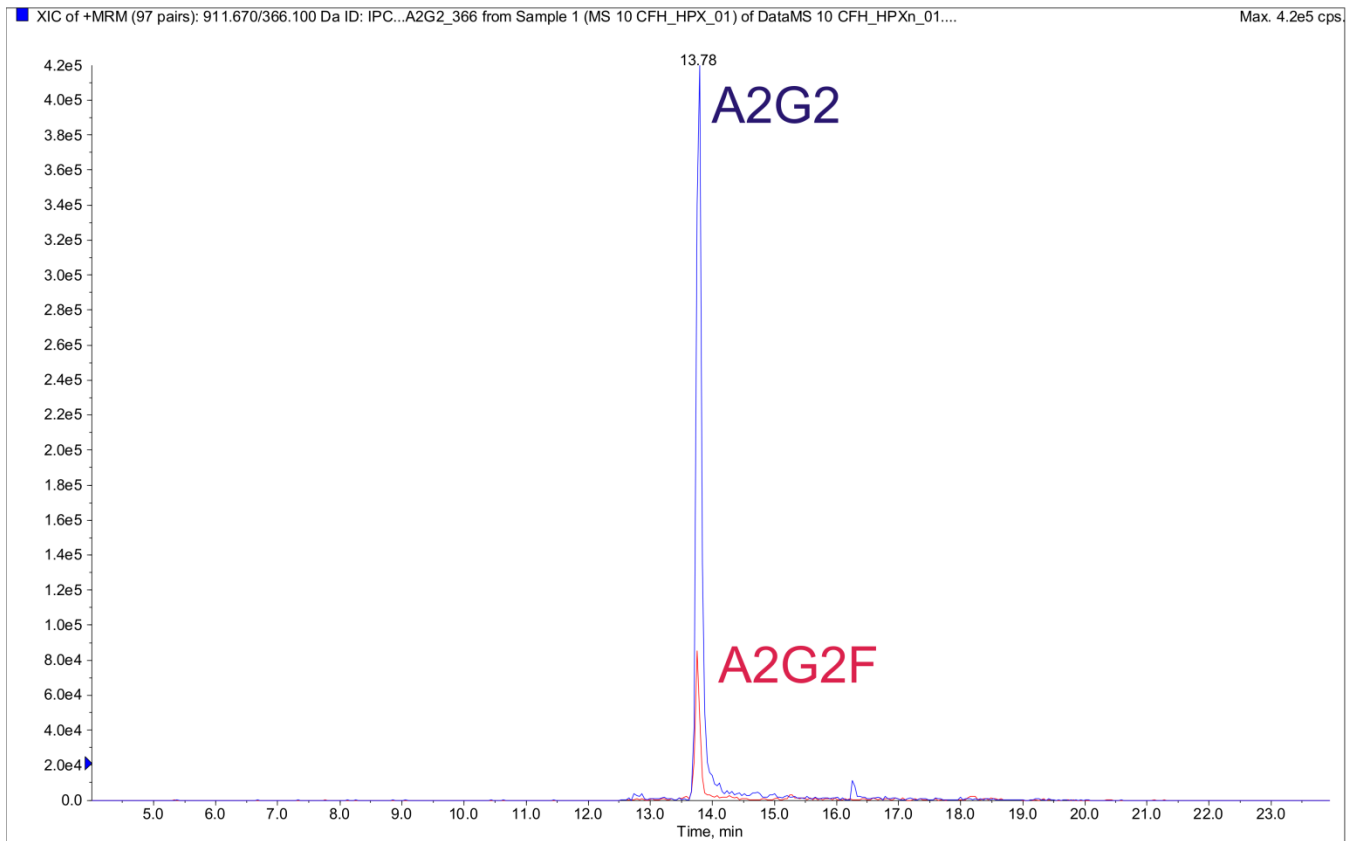
**Table S-6**

Position Structure	Hemopexin				Complement factor H						
	N64 A2G2F	N187 A2G2F	N187 A3G3F	N453 A2G2F	N217 A2G2F	N217 A3G3F	N882 A2G2F	N882 A3G3F	N911 A2G2F	N911 A3G3F	N1029 A2G2F
Healthy 1	8.5	14.1	40.9	2.5	5.9	44.8	8.3	55.9	10.3	19.2	50.2
	12.4	18.0	38.8	1.9	10.1	50.0	12.4	95.8	17.5	32.4	45.2
	14.1	10.9	30.1	1.9	4.5	18.4	10.9	83.7	32.5	49.2	35.7
	11.5	14.5	38.6	1.2	9.0	19.5	8.9	52.9	25.5	30.2	28.6
	7.7	14.8	66.5	2.9	10.7	34.6	8.7	70.9	10.6	37.1	36.5
Healthy 2	4.9	9.5	20.1	1.0	8.2	39.5	10.9	56.0	28.4	28.0	64.6
	10.6	14.9	32.7	2.4	8.2	30.2	12.3	60.3	19.1	37.4	37.6
	6.8	15.7	19.6	1.6	12.3	22.9	9.5	36.6	17.4	16.4	40.2
	8.1	12.6	18.4	1.4	9.7	18.1	8.6	27.3	12.0	8.6	27.8
	7.9	14.6	34.6	2.9	16.3	27.4	9.9	52.1	12.1	23.1	30.5
Cirrhosis 1	14.3	22.9	68.3	2.8	7.4	33.0	12.6	68.0	26.6	46.0	22.7
	14.6	8.4	59.5	5.0	13.1	92.9	31.9	168.4	35.8	121.3	54.2
	16.4	28.9	101.9	4.4	23.5	34.9	27.4	152.8	18.7	92.1	53.8
	16.3	21.3	130.5	3.5	40.7	96.9	44.4	193.5	44.4	168.4	81.0
	25.0	26.5	161.3	8.3	54.9	156.6	47.7	398.8	43.8	391.1	67.4
Cirrhosis 2	13.9	13.3	45.0	3.0	17.7	40.4	20.6	65.6	36.1	53.8	61.1
	23.1	21.1	73.3	3.3	21.2	46.4	25.8	121.5	45.2	94.2	30.9
	18.3	23.8	51.4	5.8	13.4	34.8	29.8	67.1	41.7	46.6	24.1
	61.2	35.9	107.3	12.8	47.9	177.9	72.8	222.8	84.2	307.8	88.0
	97.4	22.1	107.8	5.6	54.9	110.9	63.5	243.9	130.9	46.1	102.1
HCC 1	13.6	19.4	64.8	3.9	11.2	57.8	15.9	81.7	25.2	70.0	26.8
	17.9	22.9	152.4	8.3	14.8	102.6	25.7	169.1	27.3	132.4	35.8
	11.2	28.5	78.1	3.8	17.9	54.3	23.3	87.0	30.1	67.1	44.1
	34.1	18.2	140.6	10.1	39.8	217.6	48.8	319.2	53.5	213.8	76.5
	20.9	22.8	70.0	8.1	36.4	76.8	32.1	162.2	44.9	94.6	94.3
HCC 2	16.7	23.7	85.3	2.5	10.6	42.7	19.3	107.6	34.6	68.1	46.4
	17.8	21.5	139.5	5.7	12.9	72.1	23.4	138.2	28.8	96.0	35.2
	20.5	17.7	68.2	4.4	12.0	28.2	18.7	105.0	25.6	49.9	46.8
	59.1	27.8	130.7	10.4	62.5	199.7	65.6	384.9	85.9	289.6	78.2
	43.5	30.2	161.1	10.9	110.7	185.5	48.5	226.6	72.4	190.7	83.4

**Table S-6. Fucosylation of desialylated glycopetides in individual samples of healthy controls (H), cirrhosis (CIR) and HCC patients.** Tryptic glycopeptides of hemopexin and CFH from heme-bound fraction of individual patient samples were quantified by LC-MS MRM. Data are expressed as a relative ratio of signal intensities of fucosylated glycopeptide to its non-fucosylated counterpart monitored as the 366 transition (Hex-HexNAc) and shown as percent of the non-fucosylated form for each individual patient sample.



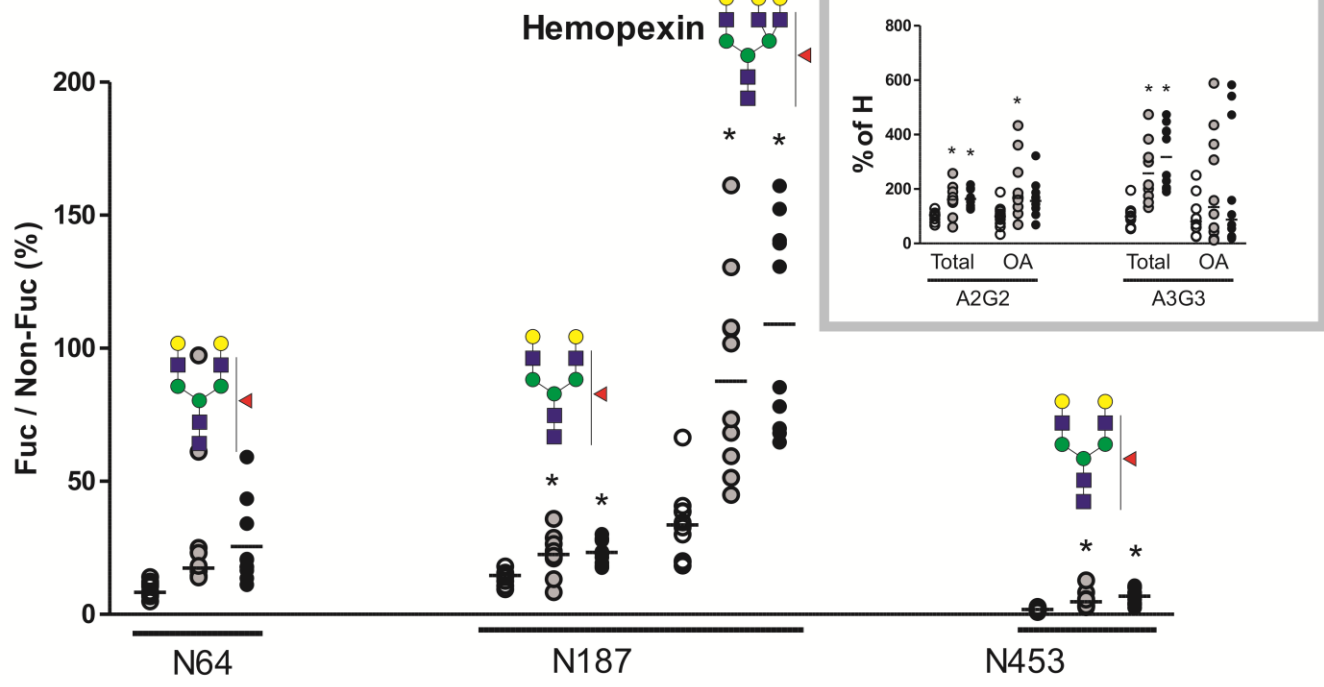
**Figure S-1. Purification of HPX and CFH** **A.** Representative chromatogram of hemin-bound fraction of plasma proteins separated on a C18 column. Two major peaks corresponding to CFH and hemopexin are labeled and the elution gradient is indicated by a dashed line. **B.** Peaks corresponding to CFH and hemopexin separated by SDS-PAGE and stained with Coomassie Blue. The two fractions were free of major contaminants.



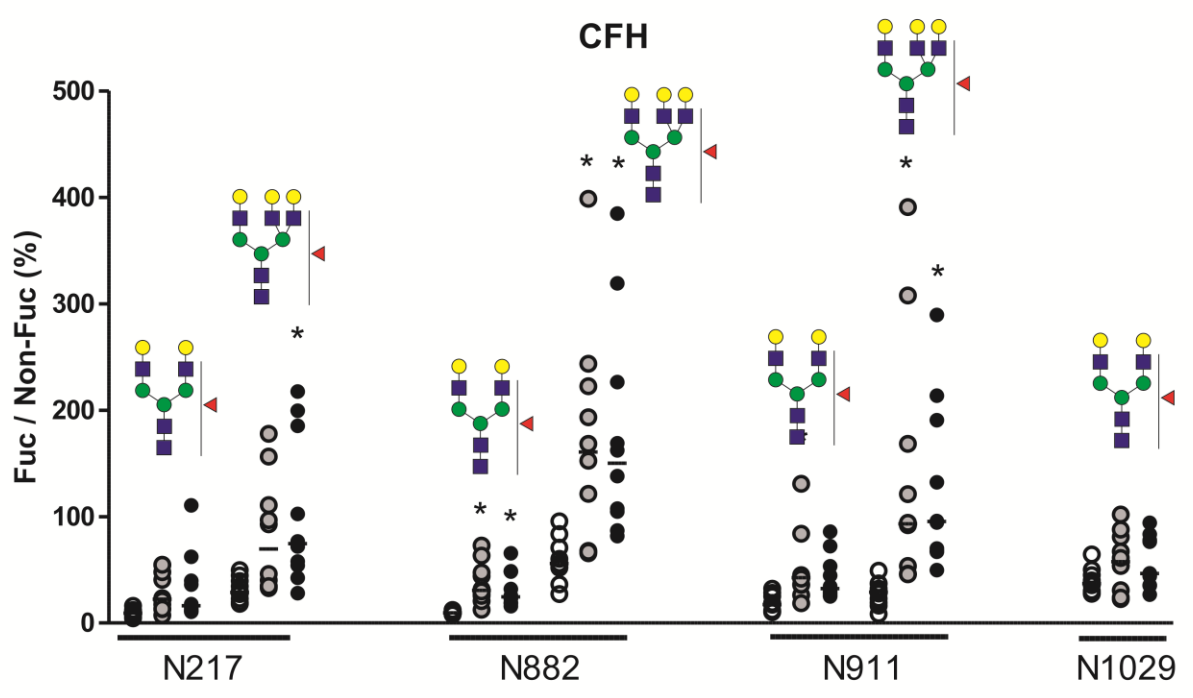
IPCSQPPQIEHG<sup>882</sup>TIN\*SSR

**Figure S-2.** Extracted ion chromatogram (XIC) of fucosylated (red) and non-fucosylated (blue) forms of bi-antennary glycans at N882 of tryptic glycopeptide IPCSQPPQIEHG<sup>882</sup>TIN\*SSR of CFH shows that retention times of both glycoforms are nearly identical.

A.

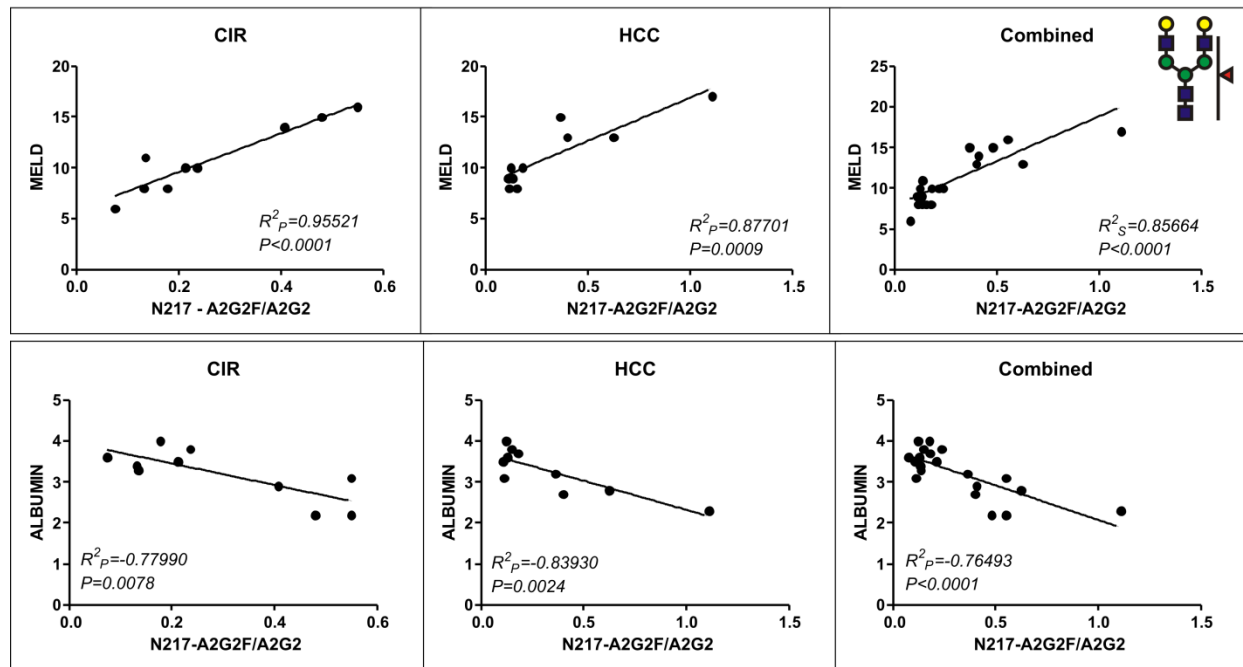


B.

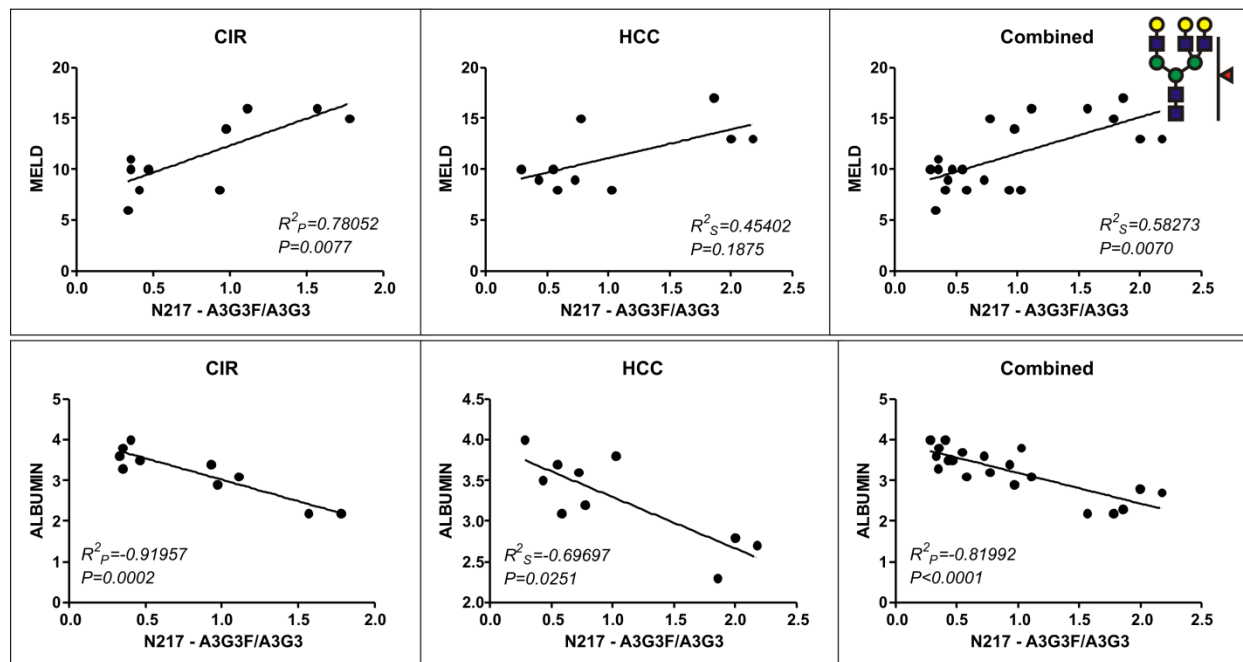


**Figure S-3. Fucosylation of desialylated glycopeptides in individual samples of healthy controls (H), cirrhosis (CIR) and HCC patients.** Tryptic glycopeptides of **A.** Hemopexin, **B.** CFH from heme-bound fraction of individual patient samples were quantified by LC-MS MRM. Data are expressed as a relative ratio of signal intensities of fucosylated glycopeptide to its non-fucosylated counterpart monitored as the 366 transition (Hex-HexNAc) and shown as percent of the non-fucosylated form. Glycan structures representing specific glycoforms are indicated above each group of corresponding bars; position of the glycosylation site in protein sequence is shown below. **Insert.** Comparison of total and outer arm fucosylation at N187 site of hemopexin. Outer arm fucosylation was quantified as 512 transition (Fuc-GlcNAc-Gal) of fucosylated precursor normalized to the 366 transition of its non-fucosylated counterpart; total fucosylation was quantified as above. The relative change in fucosylation in liver disease groups is shown as percent of H. OA; outer arm fucosylation. Distribution of individual values is shown as aligned dot plot, median is indicated by horizontal line., \* P < 0.05 vs H.

A.



B.



**Figure S-4.** Correlation between the level of fucosylation of bi- (A.) and tri- (B) antennary glycans with MELD score and serum albumin concentration, parameters of liver damage, at N217 site of CFH. The level of fucosylation is expressed as ratio of fucosylated to non-fucosylated forms quantified on individual samples by MRM protocol as described in Methods. Correlation was assessed by Pearson (P) or Spearman (S) correlation analysis depending on the sample distribution.  $R^2$  and  $P$  values are indicated in the figure.