

**Supplementary Figure 1:** Uninjured optic nerves have few  $GFP^+Iba1^+$  microglia and little arginase-1, but arginase-1 staining is greatly increased after injury. (**a**) Immunohistochemical staining of arginase-1 (green) and CD68 (red) in the uninjured and injured optic nerve (scale bar = 100 µm). (**b**) Immunohistochemical staining of an optic nerve in a C57Bl/6 mouse that was lethally irradiated and transplanted with GFP bone marrow showing sparse transplanted GFP<sup>+</sup> cells (green) among plentiful resident Iba1<sup>+</sup> microglia (red). Image is from uninjured nerve 6 weeks post-transplantation (scale bar = 100 µm).



**Supplementary Figure 2:** *Characterization of 40 \mug/kg DTx. treatment.* (**a**) Flow cytometry on the day of injury of CD4 and FoxP3 in peripheral blood of DEREG mice treated with vehicle or 40  $\mu$ g/kg DTx. (**b**) Flow cytometry in the skin-draining lymph nodes of DEREG or wild type littermates treated with DTx two days before injury and on

the day of injury, showing percent of  $CD25^{+}Foxp3^{-}T_{eff}$  cells, graphed as a percentage of TCR $\beta^+$ CD4<sup>+</sup> cells (n = 12 wild type and 9 DEREG treated mice; \*, p < 0.05, Student's ttest; representative of three experiments). (c) Representative images of H&E staining in eyes of DEREG mice treated with 40 µg/kg DTx. Abnormal architecture or immune cell infiltration was found in 0/17 C57Bl/6 and 0/21 B6AF1 mice. (d) Retinal ganglion cell counts from DEREG and wild type mice injected with 40 ug/kg DTx two days before injury and on the day of injury (n = 19 wild type and 25 DEREG; Student's t-test; representative of three experiments). (e) No difference was observed in RGC survival of C57Bl/6 mice treated with 40 µg/kg DTx or saline 2 days before injury and on the day of injury (n = 10 mice per group; Student's t-test; representative of 2 experiments). (f) Flow cytometry in the skin-draining lymph nodes of wild type mice treated with 250µg anti-CD25 or IgG 8 days before injury, showing percent of  $CD25^{+}Foxp3^{+}T_{reg}$  cells, graphed as a percentage of TCR $\beta^+$ CD4<sup>+</sup> cells (n = 4 per group; \*, p < 0.05, Student's t-test; representative of three experiments). (g) RGC survival of C57Bl/6 mice treated with 250 µg anti-CD25 or IgG 8 days before injury (n = 6 IgG and 7 anti-CD25; \*, p < 0.05, Student's t-test; representative of 2 experiments).



**Supplementary Figure 3:** *There is no change in*  $CD68^+$  *area after injury with*  $T_{reg}$  *manipulation.* (a) Quantification of  $CD68^+$  area in immunohistochemical staining of injured optic nerves of C57Bl/6 treated with DTx or DEREG mice treated with DTx (n = 3 C57Bl/6 treated with DTx and 9 DEREG treated with DTx Student's t-test; representative of two experiments). (b) Quantification of CD68<sup>+</sup> area in immunohistochemical staining of injured optic nerves of C57Bl/6 mice injected with  $1x10^6$  T<sub>eff</sub> or T<sub>reg</sub> cells 2 days before injury and on the day of injury (n = 9 T<sub>eff</sub> injected and 6 T<sub>reg</sub> injected; Student's t-test; representative of two experiments).



**Supplementary Figure 4:** *ATRA treatment leads to decreased*  $T_{eff}$  *proportion in the skindraining lymph nodes and to increased*  $T_{reg}$  *proportion in vitro.* (**a**, **b**) Flow cytometry in the skin-draining lymph nodes of wild type treated with vehicle or ATRA showing percent of CD25<sup>+</sup>Foxp3<sup>+</sup>  $T_{reg}$  cells (**a**) and of CD25<sup>+</sup>Foxp3<sup>-</sup>  $T_{eff}$  cells (**b**), graphed as a percentage of TCR $\beta^+$ CD4<sup>+</sup> cells (n = 7 vehicle treated and n = 9 ATRA treated; \*, p <

0.05, Student's t-test; representative of two experiments). (c) Representative flow cytometry plots of viable cells in lymph node cultures that have been treated with 10 nM ATRA, 5 ng/mL TGF $\beta$ , 250 U/mL IL-2, 1 µg/mL anti-CD3, and 1 µg/mL anti-CD28 (T<sub>reg</sub>) or only 1 µg/mL anti-CD3, and 1 µg/mL anti-CD28 (T<sub>eff</sub>) for 5 days. The boxes denote the CD4<sup>+</sup>Foxp3<sup>+</sup> T<sub>reg</sub> population, and numbers represent percent of viable cells that are CD4<sup>+</sup>Foxp3<sup>+</sup>. (d) Representative flow cytometry plots of viable cells in the deep cervical lymph node cultures after injection of 1 X 10<sup>6</sup> Thy1.1 T<sub>reg</sub> cells or vehicle. The gates on the top plots denote the CD4<sup>+</sup>Foxp3<sup>+</sup> T<sub>reg</sub> cells.