

Supporting Information

Abu-Odeh et al. 10.1073/pnas.1409252111

SI Materials and Methods

List of Antibodies. Antibodies are listed: polyclonal anti-WWOX antibody (1:500; gift from Kay Huebner, Ohio State University, Columbus, Ohio); p-Histone H2A.X (S139; 20E3) rabbit mAb (9718S; Cell Signaling); P-Chk2 (T68) rabbit (2661S; Cell Signaling); ATM (pS1981) rabbit mAb (2152-1; Cell Signaling); Gout anti-ATM (A300-136A; Cell Signaling); P-Chk1 (S296) rabbit (2349S; Cell Signaling); Gout anti-Chk1 (A300-162A; Cell Signaling); Rb x-p-KAP-1 (S824; A300-767A; Bethyl); rabbit anti-KAP1 (A300-274A; Bethyl); P-Histone H3 (S10; D2C8; Alexa647) XP L rabbit mAb (Cell Signaling); anti-Ub, clone FK2 (that recognize both mono- and polyubiquitin; 04-263; Millipore); anti-Ub, clone FK1 (that mostly recognize polyubiquitin chains; 04-262; Millipore); anti-Ub, Lys63-specific, clone HWA4C4, mouse mAb (05-1313; Millipore); rabbit anti-P73 (A300-126A; Bethyl); anti-GAPDH mouse mAb (CB 100 1; CALBIOCHEM); anti-Lamin A/C (N-18; Santa Cruz Biotechnology); and anti-MYC HRB(9E10) (SC-40; Santa Cruz Biotechnology).

List of Primers Used for Quantitative RT-PCR.

Gene	Direction	Primer sequence
<i>MAF</i>	Forward	CCG TCC TCT CCC GAG TTT TTC
<i>MAF</i>	Reverse	ACA CTG GTA AGT ACA CGA TGC T
<i>WWOX-7-8</i>	Forward	TCC TCA GAG TCC CAT CGA TTT
<i>WWOX-8-9</i>	Reverse	CGG CAG TTG AAG TA
<i>GAPDH</i>	Forward	CAT GAG AAG TAT GAC AAC AGC CT
<i>GAPDH</i>	Reverse	AGT CCT TCC ACG ATA CCA AAG T
<i>UBC</i>	Forward	ATT TGG GTC GCG GTT CTT G
<i>UBC</i>	Reverse	TGC CTT GAC ATT CTC GAT GGT

Sequences of shRNAs Against the Human *WWOX* Gene. shRNA lentiviral plasmids (pLKO.1-puro; clone ID: NM_016373.1-720s1c1; Sigma) were used to deplete *WWOX* expression in MCF7 cells according to standard protocol.

Sequence: CCGGGCCAAGAATGTGCCTCTTCATCTCGAG-ATGAAGAGGCACATTCTTGGCTTTTGTG.

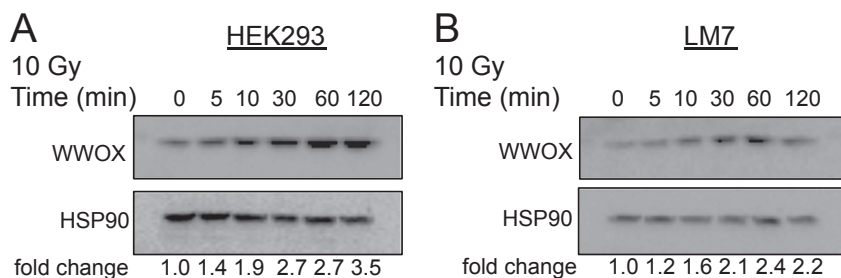


Fig. S1. Induction of *WWOX* expression early after DNA damage stimuli. Immunoblot analysis of *WWOX* levels in (A) HEK293 and (B) osteosarcoma LM7 cells after treatment with 10 Gy.

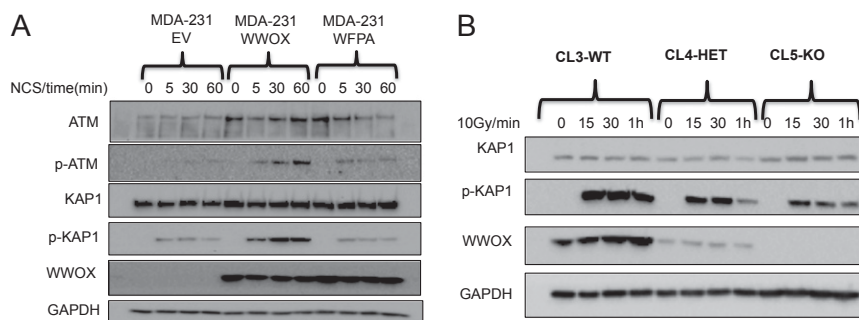


Fig. S2. Checkpoint activation in *WWOX*-manipulated cells after DSBs. (A) MDA-MB231 cells were transduced with Lenti-EV, Lenti-*WWOX*, or Lenti-*WWOX*-WFPA. Cells were selected, and clones were treated with NCS (200 ng/mL) for the indicated time points. Whole-cell lysates were blotted with antibodies against ATM, p-ATM, KAP1, p-KAP1, *WWOX*, and GAPDH. (B) Early-passage MEFs isolated from embryonic day 13.5 embryos were γ -irradiated for the indicated time points. Cell lysates were then probed with antibodies against KAP1, p-KAP1, *WWOX*, and GAPDH.

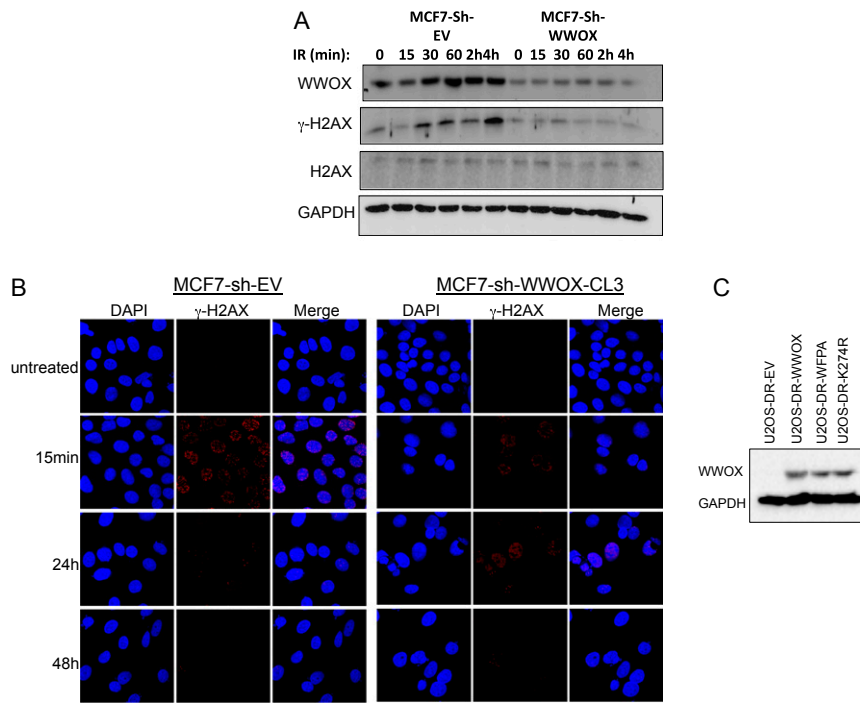


Fig. S3. (A and B) Impaired phosphorylation of H2AX in WWOX-depleted MCF7 cells. (A) Immunoblot analysis of control and WWOX-depleted MCF7 cells after ionizing radiation treatment for the indicated times. Whole-cell lysates were propped with antibodies against WWOX, Ser139-H2AX, total H2AX, and GAPDH. (B) Representative confocal microscopy images of γ -H2AX foci (red) in control MCF7-Sh-EV and WWOX-depleted (MCF7-Sh-WWOX) cells after treatment of ionizing radiation at different time points. DAPI staining (blue) was used to stain nuclei. Foci were quantified and are presented in Fig. 3C. (C) Overexpression of WWOX in U2OS-DR-GFP cells. Cells were transduced with the indicated Lenti-viral constructs (EV, WWOX, WWOX-WFPA, or WWOX-K274R), and stable clones were established. Immunoblot analysis showing WWOX expression in the different clones. GAPDH was used as loading control. IR, ionizing radiation.

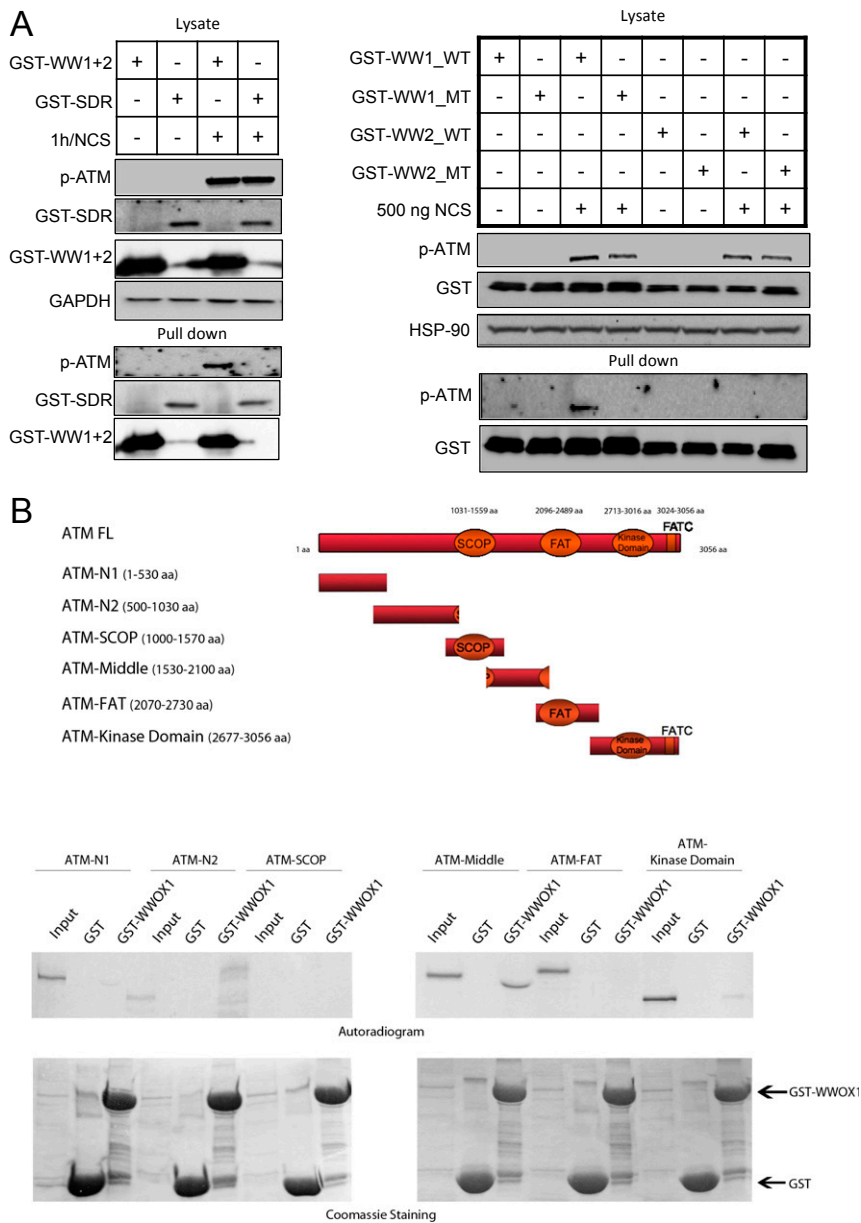


Fig. S4. (A) WW1 domain of WWOX physically associates with ATM. HEK293 cells were transfected with (Left) GST-WW1+2 or GST-SDR or (Right) GST-WW1, mutant GST-WW1-WFPA (WW1_MT), GST-WW2, or GST-WW2-YFPA (GST-WW2_MT). At 24 h, cells were treated with NCS (200 ng/mL) for an additional 1 h. GST pull down was performed for 2 h, and complexes were analyzed by immunoblotting using antibodies against GST, p-ATM, and GAPDH or HSP90. (B) GST-WWOX interacts with ATM (amino acids 1,530–2,100) *in vitro*. (Top) Schematic representation of ATM deletion constructs used in GST pull down as *in vitro*-transcribed, translated, and 35S-labeled fragments. The 35S-labeled fragments were incubated with bacterial extracts expressing GST-WWOX followed by washing and electrophoresis. GST alone was used as the negative control. Middle shows the autoradiogram of 35S-labeled ATM fragments pulled down with GST alone or GST-WWOX. Bottom shows the Coomassie-stained loading control of inputs and pulled fragments.

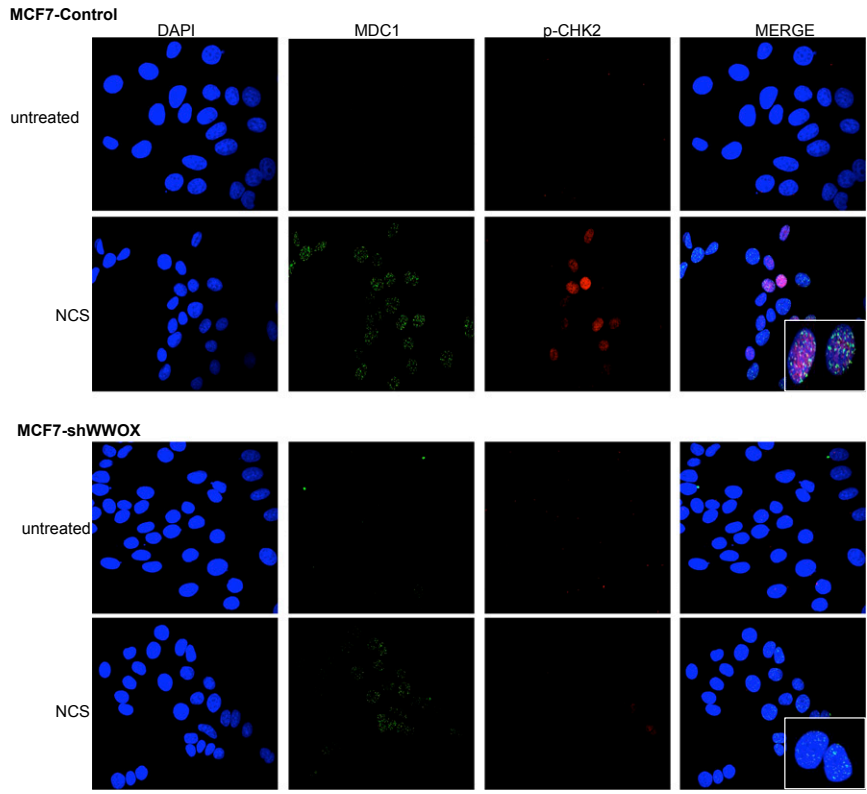


Fig. 55. Impaired activation and recruitment of MDC1 and p-CHK-2 in WWOX-depleted MCF7 cells. MCF7 and MCF7-ShWWOX cells were treated or untreated with NCS (200 ng/mL) for 30 min. Cells were then fixed and stained with anti-MDC1 (green) and anti-p-CHK2 (red). Nuclei were counterstained with DAPI (blue). Cells were visualized with confocal microscopy.

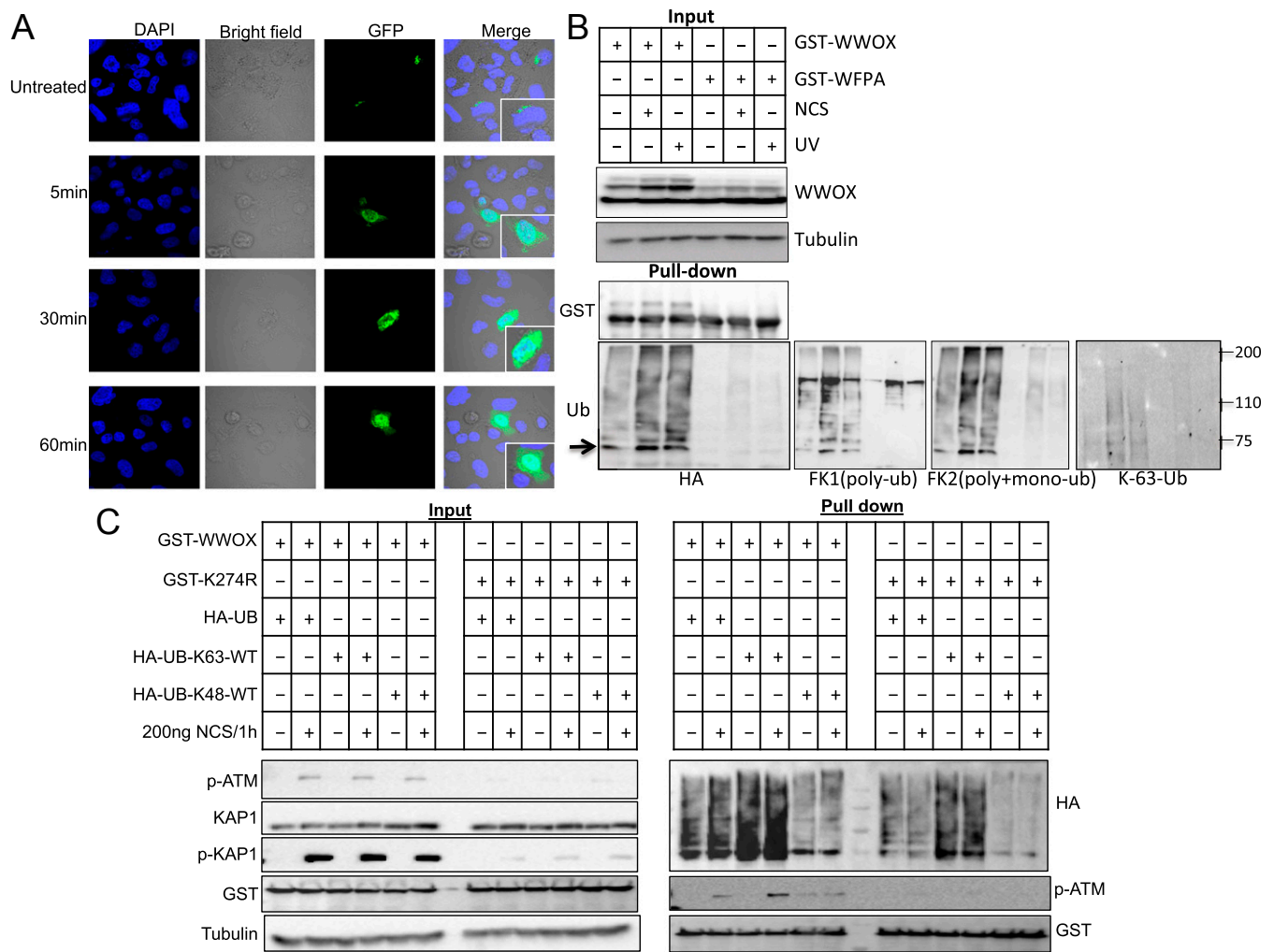


Fig. S6. (A) Nuclear localization of WWOX after NCS treatment. MCF7 or HeLa cells were transfected with GFP-WWOX. Twenty-four hours later, cells were untreated or treated with NCS (200 ng/mL). At the indicated times after NCS treatment, the cells were fixed and visualized with confocal microscopy using 60x objective lens. Nuclei were counterstained with DAPI (blue); bright field is shown in gray. Nuclear GFP-WWOX was observed in 50–70% of cells. (B and C) WWOX ubiquitination after DNA damage. (B) HEK293 cells transfected with HA-Ub and *pEBG-WWOX* or *pEBG-WWOX-WFPA* plasmids. At 24 h, cells were treated with UV (100 J/m²) or NCS (200 ng/mL) for an additional 1 h. Cells were then subjected to GST pull down, and lysates were blotted against GST (WWOX) and tubulin. Pulled down complexes were blotted with anti-GST (WWOX), anti-HA (Ub), anti-FK1 (recognizing polyubiquitin), anti-FK2 (recognizing both mono- and polyubiquitin), and anti-K63 polyubiquitin antibodies. (C) HEK293 cells were transfected with vectors encoding GST-WWOX or GST-K274R and HA-Ub, HA-Ub-K63-WT, or HA-Ub-K48-WT (ubiquitin constructs are either K63 only or K48 only, whereas all other lysines are mutated to arginines). At 24 h, cells were treated with NCS for an additional 1 h; (Right) lysates were then subjected to GST pull down, and precipitates were blotted as indicated.

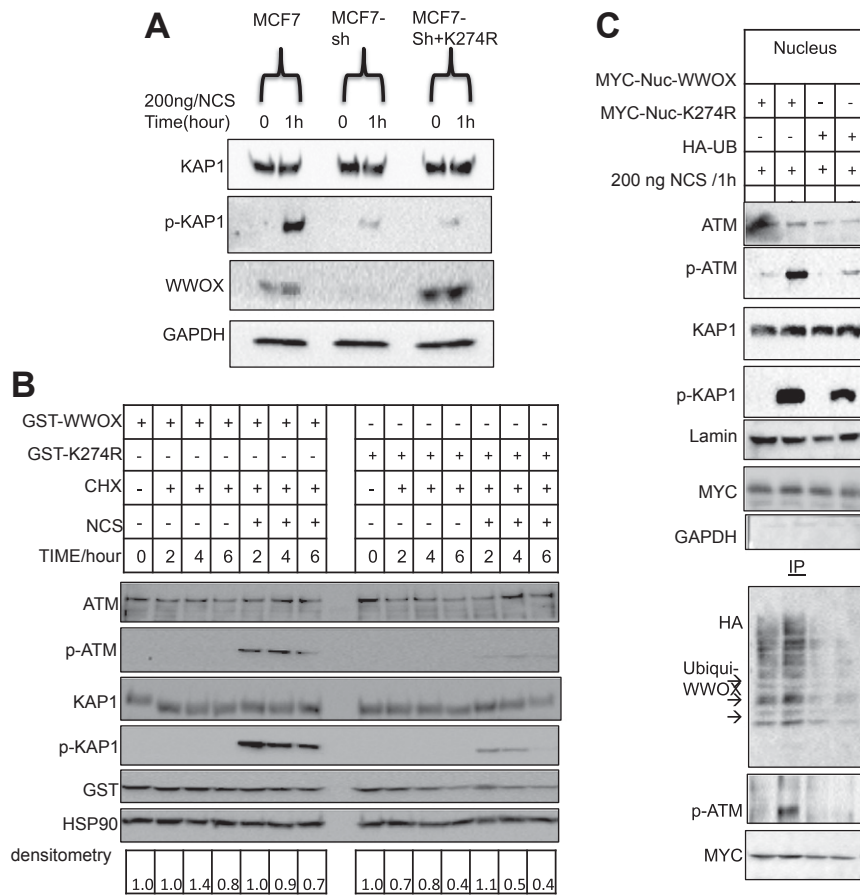


Fig. S7. (A) WWOX-depleted MCF7 cells were transduced with lentiviral vectors expressing EV or WWOX-K274R mutant and selected with neomycin. The different cells were untreated or treated with NCS (200 ng/mL). Whole-cell lysates were prepared at the indicated time points, and immunoblot analysis was performed using specific antibodies against KAP1, p-KAP1, WWOX, and GAPDH. (B) HEK293 cells were transfected with vectors encoding GST-WWOX or GST-K274R. At 24 h posttransfection, cells were treated with cyclohexamide (CHX; 100 μg/mL) in the presence or absence of NCS (200 ng/mL) for 0, 2, 4, or 6 h. Whole-cell lysates were blotted with antibodies as indicated. Densitometry shows levels of GST-WWOX or GST-WWOXK274R relative to HSP90 levels. (C) HEK293 cells were transfected with Myc-Nuc-WWOX or Myc-Nuc-WWOX-K274R. At 24 h, cells were treated with NCS for an additional 1 h. Cells were then lysed and immunoprecipitated with anti-Myc antibody. (Lower) Precipitates were blotted with anti-HA, anti-p-ATM, and anti-Myc. (Upper) Input lysates were blotted as indicated. IP, immunoprecipitation.