

At PEX2

Glycine max

Oryza sativa

Brachypodium

Selaginella

Physcomitrella

Gallus gallus

Danio rerio

Homo sapiens PEX2

C. elegans PRX-2

S. cerevisiae Pex2p

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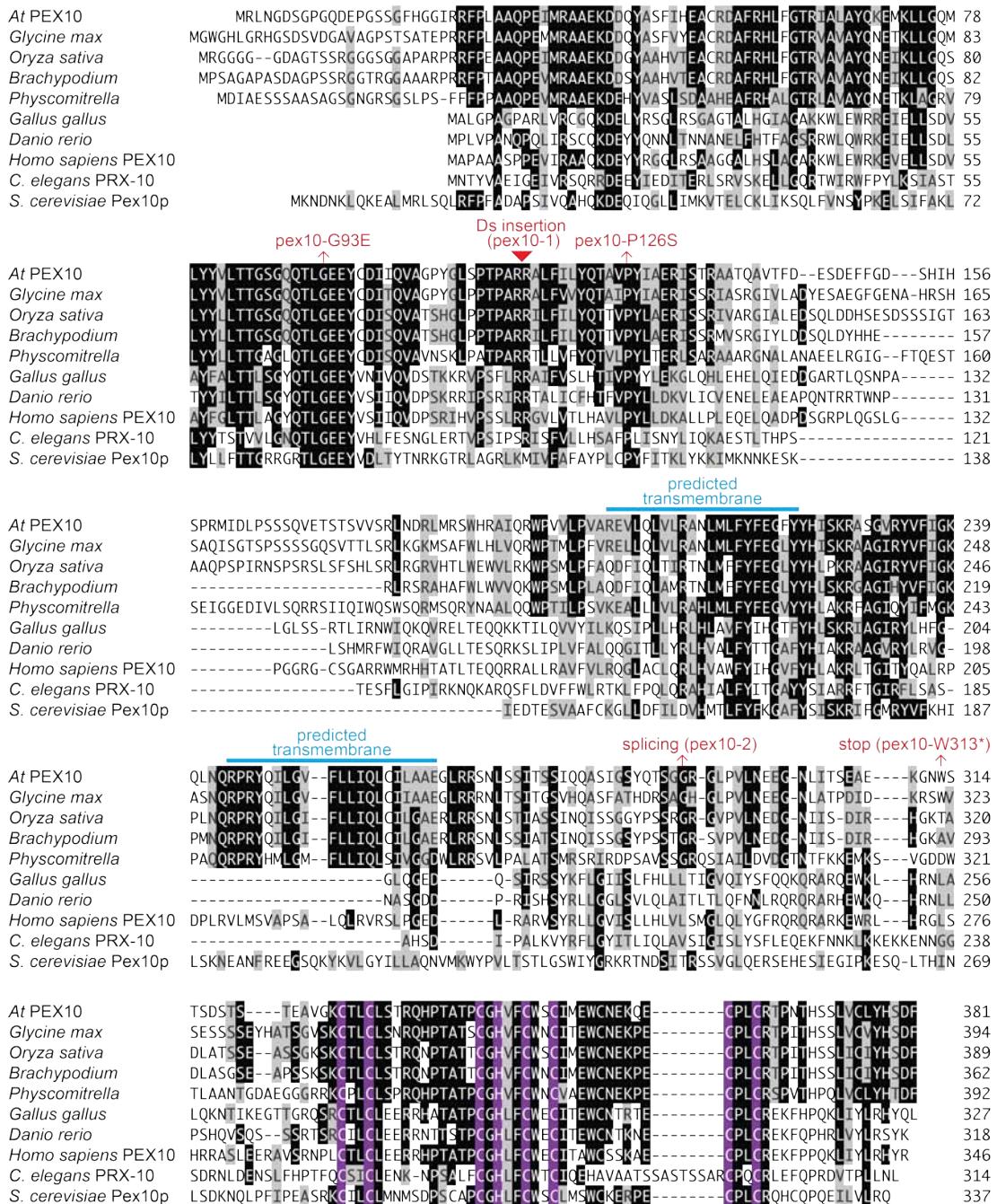
S. cerevisiae Pex2p

M (ted3)

Salk_033081

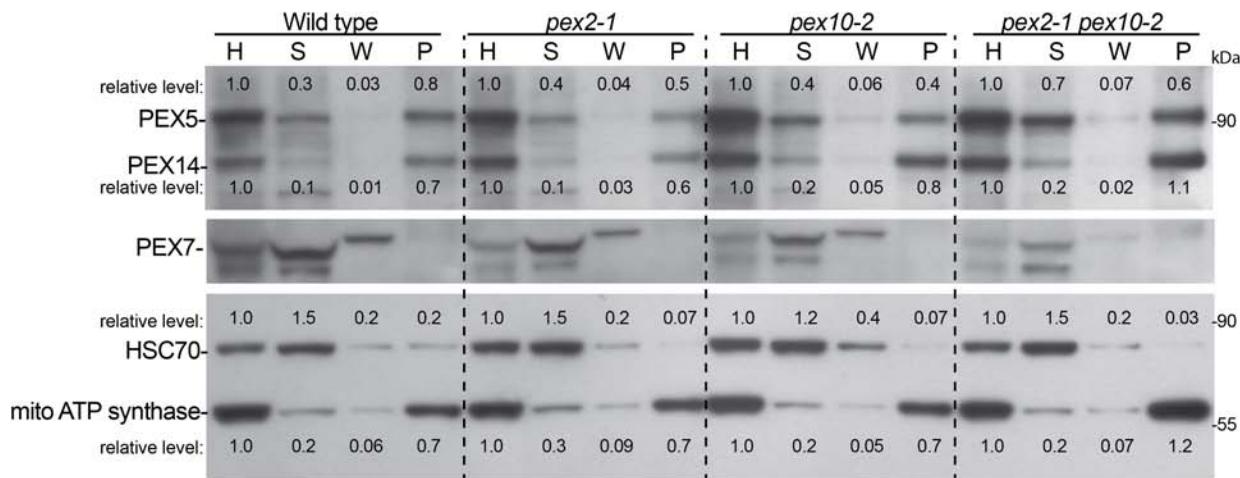
Supplemental Figure S1. Alignment of PEX2 proteins from various organisms.

Arabidopsis (*At*) PEX2 (At1g79810) was aligned with homologs from *Glycine max* (XP_003530138.1), *Oryza sativa* (Os05g0275700), *Brachypodium distachyon* (XP_003573500.1), *Selaginella moellendorffii* (XP_002976539.1), *Physcomitrella patens* (XP_001781834.1), *Gallus gallus* (NP_001008454.1), *Danio rerio* (XP_684073.2), *Homo sapiens* (NP_000309.1), *Caenorhabditis elegans* (CAA92640.2), and *Saccharomyces cerevisiae* (NP_012325.1). The alignment was generated using Lasergene MegAlign (DNASTAR, Madison, WI) with the Clustal W default settings and the Gonnet series protein weight matrix. Residues identical in at least six sequences are shaded in black or purple; chemically similar residues are shaded in gray. Transmembrane domains predicted using the ARAMEMNON database (Schwacke et al., 2003) are indicated in blue, metal-coordinating residues of the RING-finger domain are highlighted in purple, and positions of *Arabidopsis* pex2 mutants are indicated in red.



Supplemental Figure S2. Alignment of PEX10 proteins from various organisms.

Arabidopsis (At) PEX10 (At2g26350) was aligned with homologs from *Glycine max* (XP_003544551.1), *Oryza sativa* (Os07g0608800), *Brachypodium distachyon* (XP_003559973.1), *Physcomitrella patens* (XP_001777562.1), *Gallus gallus* (NP_001185583.1), *Danio rerio* (NP_001005994.1), *Homo sapiens* (NP_722540.1), *Caenorhabditis elegans* (NP_001021200.2), and *Saccharomyces cerevisiae* (NP_010551.1). The alignment was generated using Lasergene MegAlign (DNASTAR, Madison, WI) with the Clustal W default settings and the Gonnet series protein weight matrix. Residues identical in at least five sequences are shaded in black or purple; chemically similar residues are shaded in gray. Transmembrane domains predicted using the ARAMEMENON plant membrane protein database (Schwacke et al., 2003) are indicated in blue, metal-coordinating residues of the RING-finger domain are highlighted in purple, and positions of *Arabidopsis* pex10 mutants are indicated in red.



Supplemental Figure S3. Membrane association of peroxisome matrix protein receptors in *pex2-1*, *pex10-2*, and *pex2-1 pex10-2*.

Light-grown 8-d-old seedlings were homogenized and the resultant extracts were centrifuged to separate soluble proteins from proteins associated with organellar membranes. Homogenate (H), soluble (S), wash (W) and pellet (P) fractions were processed for immunoblotting and serially probed with the indicated antibodies. HSC70 and mitochondrial membrane complex V (mito ATP synthase) were used as cytosolic and organellar controls, respectively. Numbers above or below the bands indicate the relative level of the indicated protein in the S, W, or P fraction relative to the amount in the H fraction. This experiment was repeated three times with similar results.

Supplemental **Table 1.** *Markers used in recombination mapping of pex2-1 and pex10-2*

Marker	Nearest gene	Restriction enzyme	Fragment size (bp)		Primer sequences
			Col-0	Ler	
LCS1127	<i>At1g32130</i>	<i>TaqI</i>	206	138, 68	CAAAATCCTGAAGTGGCTCTGTAG ATTTCGATTCGTTTTGTG
T14L22	<i>At1g51913</i>	<i>HinfI</i>	~700	~450, ~250	GAAAAAATTAAAGCCCCAACCC CACCCGCAAACACAACAAACC
SNP10592	<i>At1g69250</i>	<i>RsaI</i>	115, 29	144	AGCGACGGCAACCTAAAAAGTAAAG AGGTTCAAGCACATTGATGATACTCTGTA
SEC238	<i>At2g18950</i>	<i>DdeI</i>	145, 69	214	GAGTTCTGCCTTGATTGAGTAAAGTTG GAAAACCTGTAAAACGCATCTAACGAGTC
SEC241	<i>At2g20770</i>	<i>PvuII</i>	177, 27	204	CAACTCAAACCTATTACCATCAAACAAACAA TATCTCTCCTTGACATGGTCGACCCAGCT
SEC244	<i>At2g25095</i>	<i>AvaII</i>	197, 28	225	TAAAAAATAGTAAACTTGTCCCGATAAA GTCGATAGATTACGGACGTATTAGAAAGG
SEC245	<i>At2g28360</i>	<i>RsaI</i>	116, 28	144	CTGCAGCTCGTTCTAGGATATTGGTCAT CTATAAGAATTGAAGAAAGGCGATGAAGTA
SEC246	<i>At2g32340</i>	<i>RsaI</i>	114, 27	141	TTTACCTGTTCCGCCAATCACTCCTC GAACCTGCGTCGCTATCTGCTTCCTCCGTA

Supplemental Table 2. Primers used for amplification and sequencing of *PEX2* and *PEX10*

Gene/Accession #	Primer names	Primer sequences
<i>PEX2/At1g798100</i>	PEX2-12 and	CATGGATGAATCTCTGTGGGTGTA
	PEX2-13	GAAAGAAAAAGAAGAGTAGCGGAAGTGAGAA
	PEX2-14 and	TCGCTATGGTGATTCTTTCTCCTT
	PEX2-15	GCTGCTGCCTGGCTTTACACTTC
<i>PEX10/At2g26350</i>	PEX10-3 and	GACCACCTAACATCTCCAACCTTTT
	PEX10-4	ATGGCCACTCGACTGATCCCGCTCTAC
	PEX10-5 and	AAGGAGTGGCCGTTGGGTGCTGAC
	PEX10-6	TTCTGGGGTTTCCTTAATC
	PEX10-7 and	CCTATAGAACGCCTGCTGAATGGAG
	PEX10-8	TCAGGGCAACAAACTTAGGAGAGGAAT
	PEX10-9 and	TGGAACTGCGGTCTGGTACAATATGAACAAAG
	PEX10-10	CTTGAAATTAAAAAAATATCAAAGGC

Supplemental Table 3. PCR-based markers for determining mutant genotypes

Mutant	Primer names	Primer sequences	Restriction enzyme	Product size (bp)	
				Wt	Mutant
<i>pfl36/pex2-1</i>	PEX2-18	TGC GTT GCCTCCGGTGGTGGCAG	<i>Dpn</i> II	65, 18	93
	PEX2-DpnII ¹	CATA CAG ACCT GCT CAG AAT CACCC <u>GAT</u>			
<i>pex2-2</i>	PEX2-20	GCT TATTATATGTGGTCTTGATGGCAGTC	<i>Nde</i> I	134	102
	PEX2-NdeI ¹	GAGTGA AAAAC CCTTAACGAGCTGTTCC <u>TTCCA</u>			
<i>ted3</i>	TED3-1	ATG CTTT GTT GCTT CTCCC GCT GCTC	<i>Tsp</i> 45I	341, 99	440
	TED3-2	TCTAAGACACCGGAACGATGCTGCTG			
<i>pfl81/pex10-2</i>	PEX10-11	CGTTGAAGTTGAATCGGAGGTAGAC	<i>Pst</i> I	92, 28	120
	PEX10-PstI ¹	AATATAGTTTGGTATTGTT <u>CCTGCA</u>			
<i>pex10-P126S</i> ²	PEX10-8	TCAGGGCAACAAACTT TAGGAGAGGAAT	-	229	229
	PEX10-TaqI	ACGAAACCTAATTCTCTGCGATATATC			

¹dCAPS oligonucleotide (Michaels and Amasino, 1998; Neff et al., 1998); the underlined nucleotide differs from wild-type sequence to create a restriction site in either the mutant or wild-type PCR amplicon.

²*pex10-P126S* was amplified with the listed primers and sequenced with primer PEX10-8 to confirm genotype.

Supplemental literature cited:

- Michaels SD, Amasino RM** (1998) A robust method for detecting single-nucleotide changes as polymorphic markers by PCR. *Plant J* **14**: 381-385
- Neff MM, Neff JD, Chory J, Pepper AE** (1998) dCAPS, a simple technique for the genetic analysis of single nucleotide polymorphisms: experimental applications in *Arabidopsis thaliana* genetics. *Plant J* **14**: 387-392
- Schwacke R, Schneider A, van der Graaff E, Fischer K, Catoni E, Desimone M, Frommer WB, Flugge UI, Kunze R** (2003) ARAMEMNON, a novel database for Arabidopsis integral membrane proteins. *Plant Physiol* **131**: 16-26