

## Supporting Information

### Regio- and Stereoselective Mono-Epoxidation of Dienes using Methyltrioxorhenium: Synthesis of Allylic Epoxides

Saroj Ranjan De, Ganesh Kumar, Jawahar L. Jat, Saritha Birudaraju, Biao Lu, Rajkumar Manne, Narender Puli, Adeniyi Michael Adebesein, and John R. Falck\*

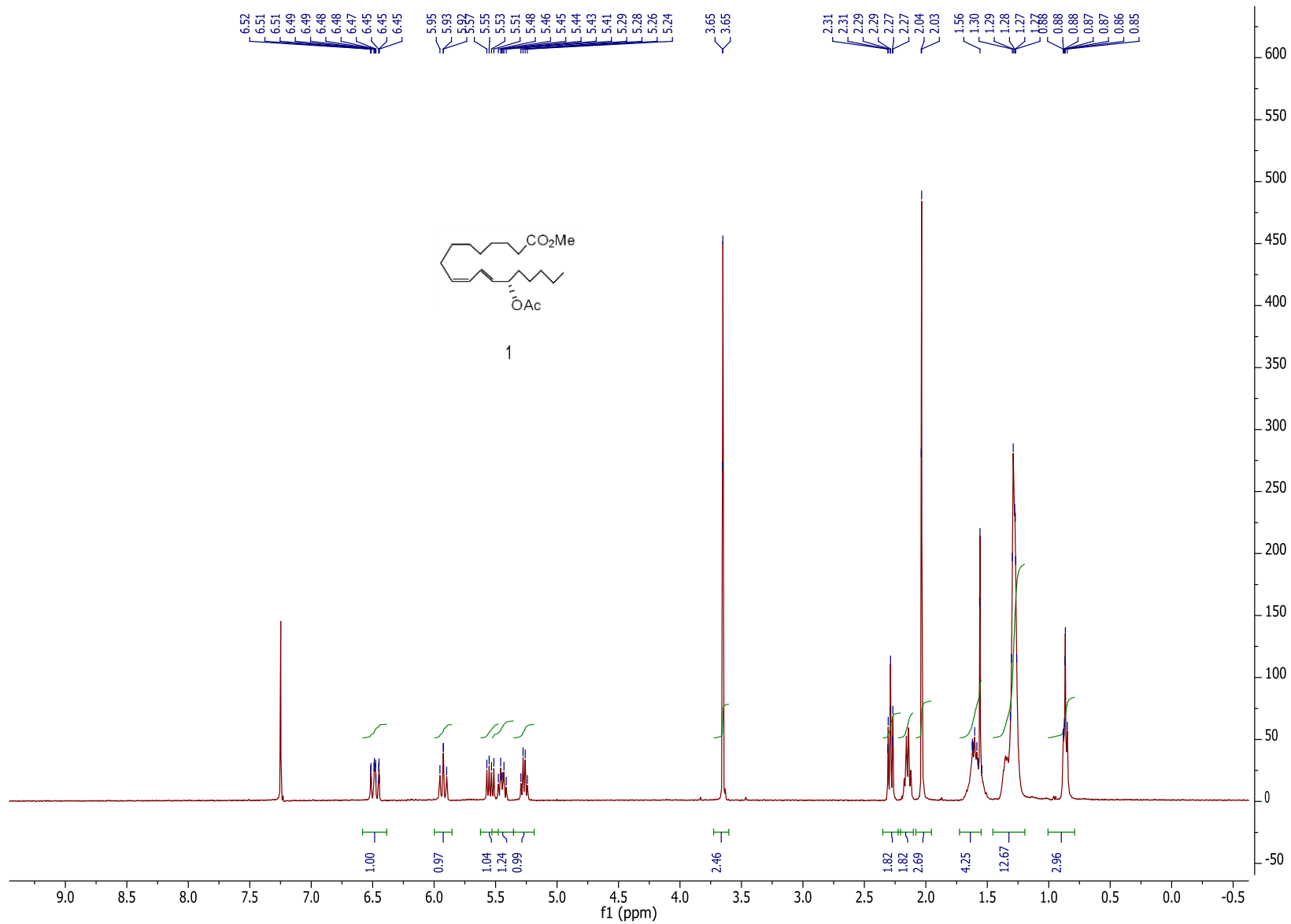
*Division of Chemistry, Department of Biochemistry, University of Texas Southwestern Medical Center, Dallas, Texas 75390*

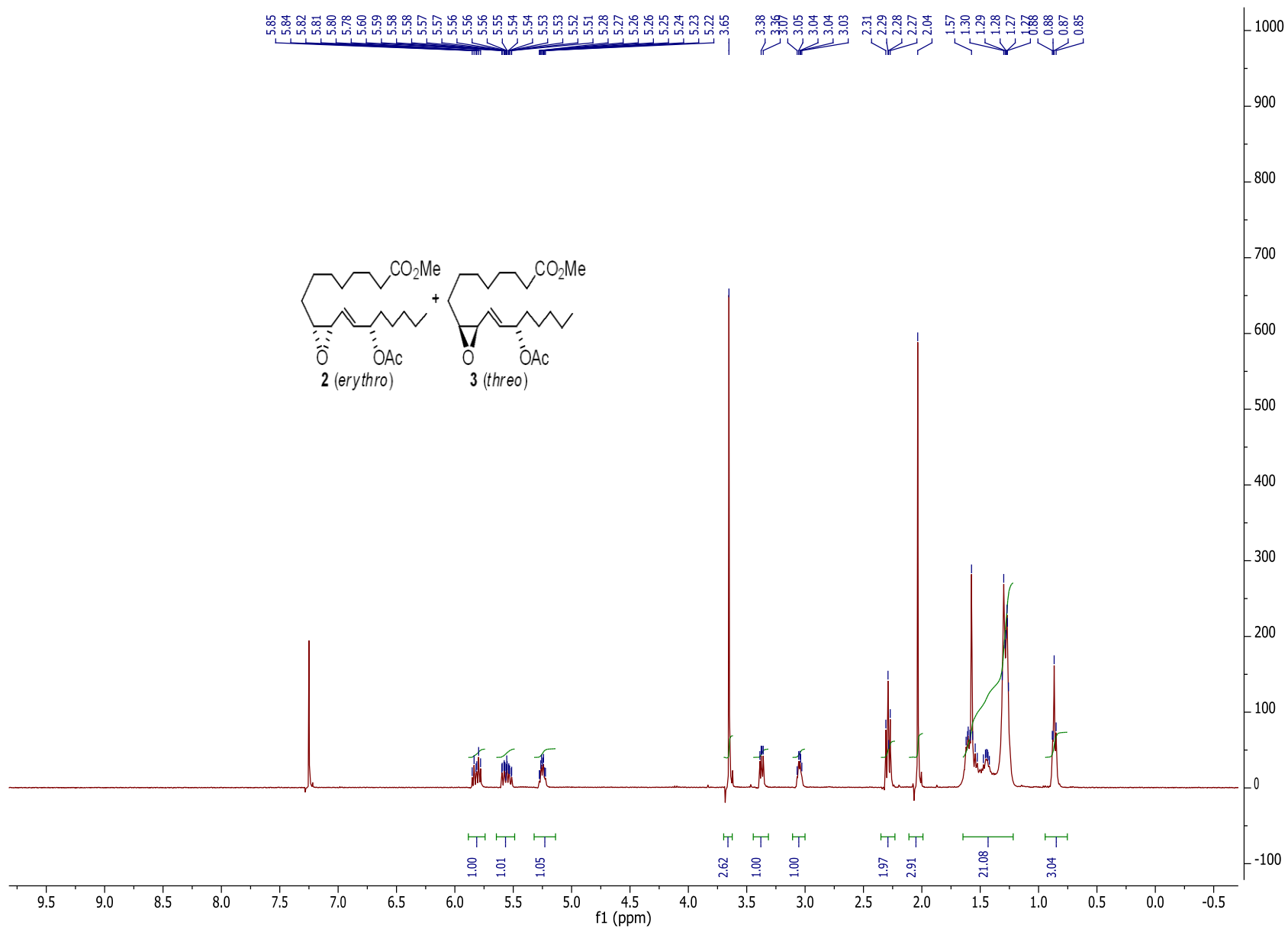
#### Table of Contents

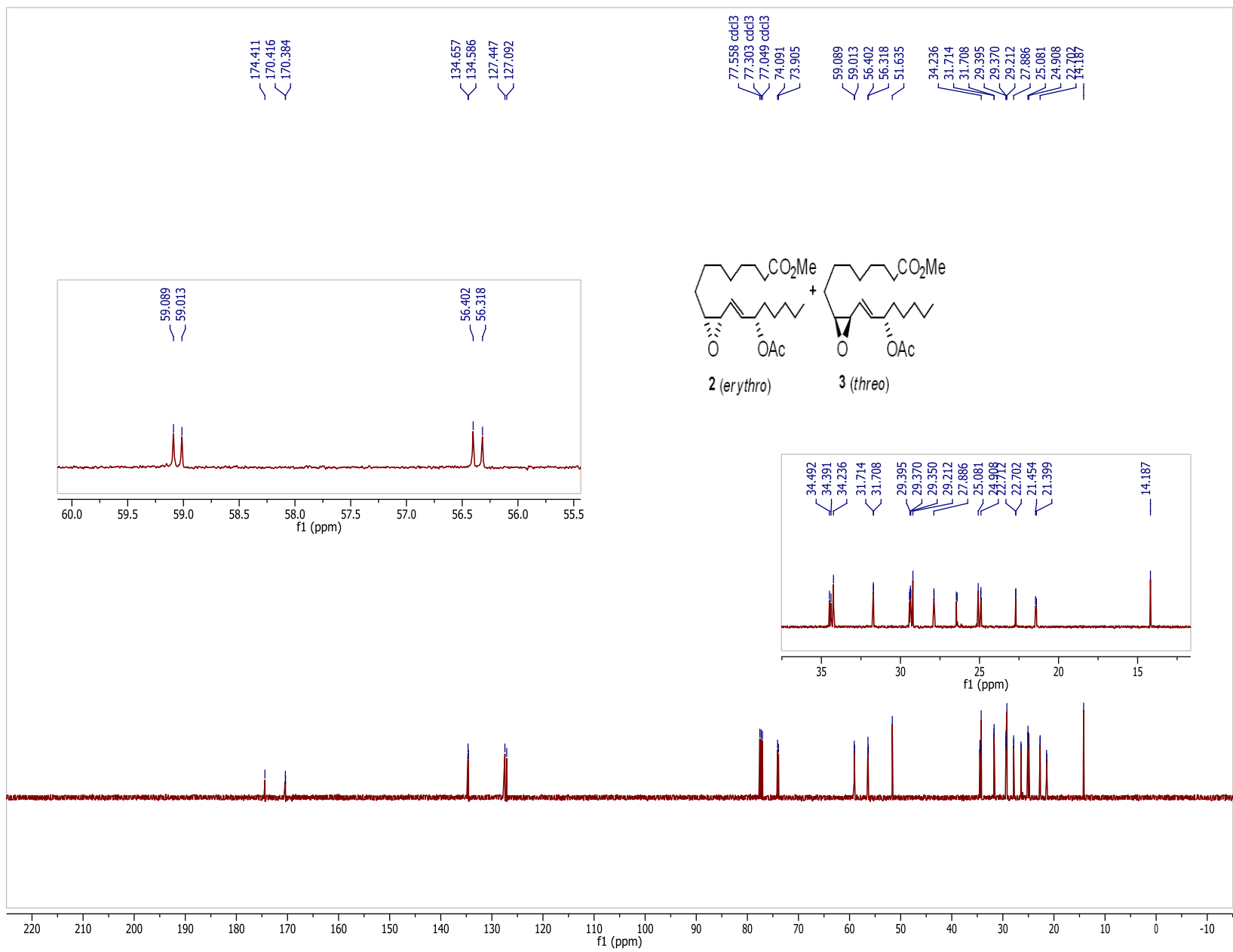
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>1</b>	S4
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>2/3</b>	S5-S6
HPLC chromatogram of <b>2/3</b>	S7
HPLC of diastereo-enriched <b>2</b>	S8
<sup>1</sup> H NMR spectra of <b>4a</b>	S9
<sup>1</sup> H NMR spectra of <b>5a/6a</b>	S10
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>4b</b>	S11-S12
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>5b/6b</b>	S13-S14
<sup>1</sup> H NMR spectra of <b>4c</b>	S15
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>5c/6c</b>	S16-S17
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>4d</b>	S18-S19
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>5d/6d</b>	S20-S21
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>4e</b>	S22-S23
<sup>1</sup> H/ <sup>13</sup> C NMR spectra of <b>5e/6e</b>	S24-S25

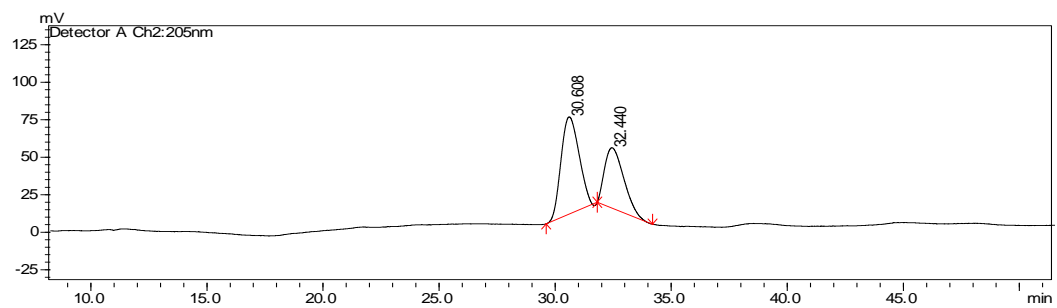
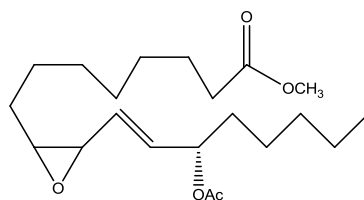
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>5f/6f</b>	S26-S27
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>7</b>	S28-S29
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>8</b>	S30-S31
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>9</b>	S32-S33
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>10</b>	S34-S35
$^1\text{H}$ NMR spectra of <b>12/13</b>	S36
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>15</b>	S37-S38
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>17</b>	S39-S40
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>19</b>	S41-S42
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>20</b>	S43-S44
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>21</b>	S45-S46
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>22</b>	S49-S50
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>23</b>	S51-S52
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>24</b>	S55-S56
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>25</b>	S57-S58
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>26</b>	S59-S60
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>27</b>	S61-S62
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>28</b>	S63-S64
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>29</b>	S65-S66
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>30</b>	S67-S68
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>31</b>	S69-S70
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>32</b>	S71-S72

$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>33</b>	S73-S74
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>34</b>	S75-S76
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>35</b>	S77-S78
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>36</b>	S79-S80
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>38</b>	S81-S82
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>40</b>	S83-S84
$^1\text{H}/^{13}\text{C}$ NMR spectra of <b>42</b>	S85-S86
HPLC chromatogram of standard chiral epoxide.	S87
HPLC chromatogram of chiral epoxide from chiral salan ligand and Ti(IV)	S88
HPLC chromatogram of 1:1 mixture of chiral Ti(salan)-generated epoxide + epoxide standard	S89
HPLC chromatogram of epoxide 5a/6a from MTO-pyridine epoxidation	S90
HPLC chromatogram of epoxide 5a/6a from MTO-pyridine epoxidation + epoxide standard	S91



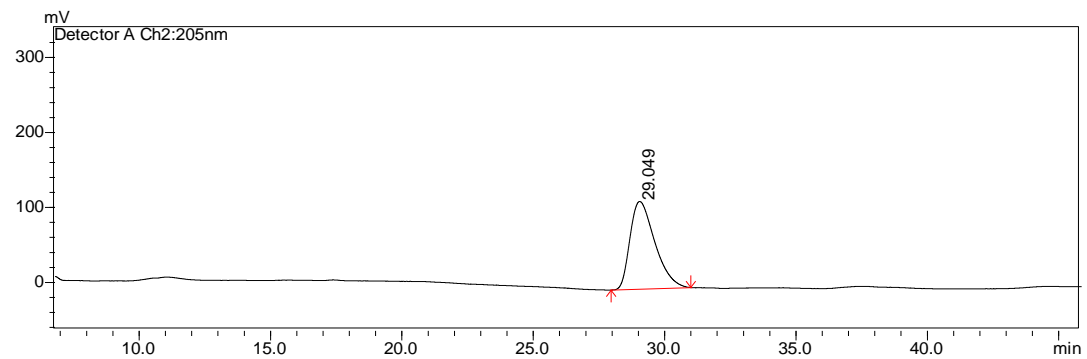
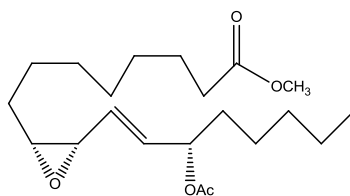






Peak	Ret. Time	Area	Height	Area %
1.	30.608	3568595	65420	58.429
2.	32.440	2539018	42270	41.571

CHIRALCEL OJ-H, 15 CM, 4.6MM, 2.7 MICRON, Hexane:IPA (99.8:0.2), 0.8 mL/min, 205.

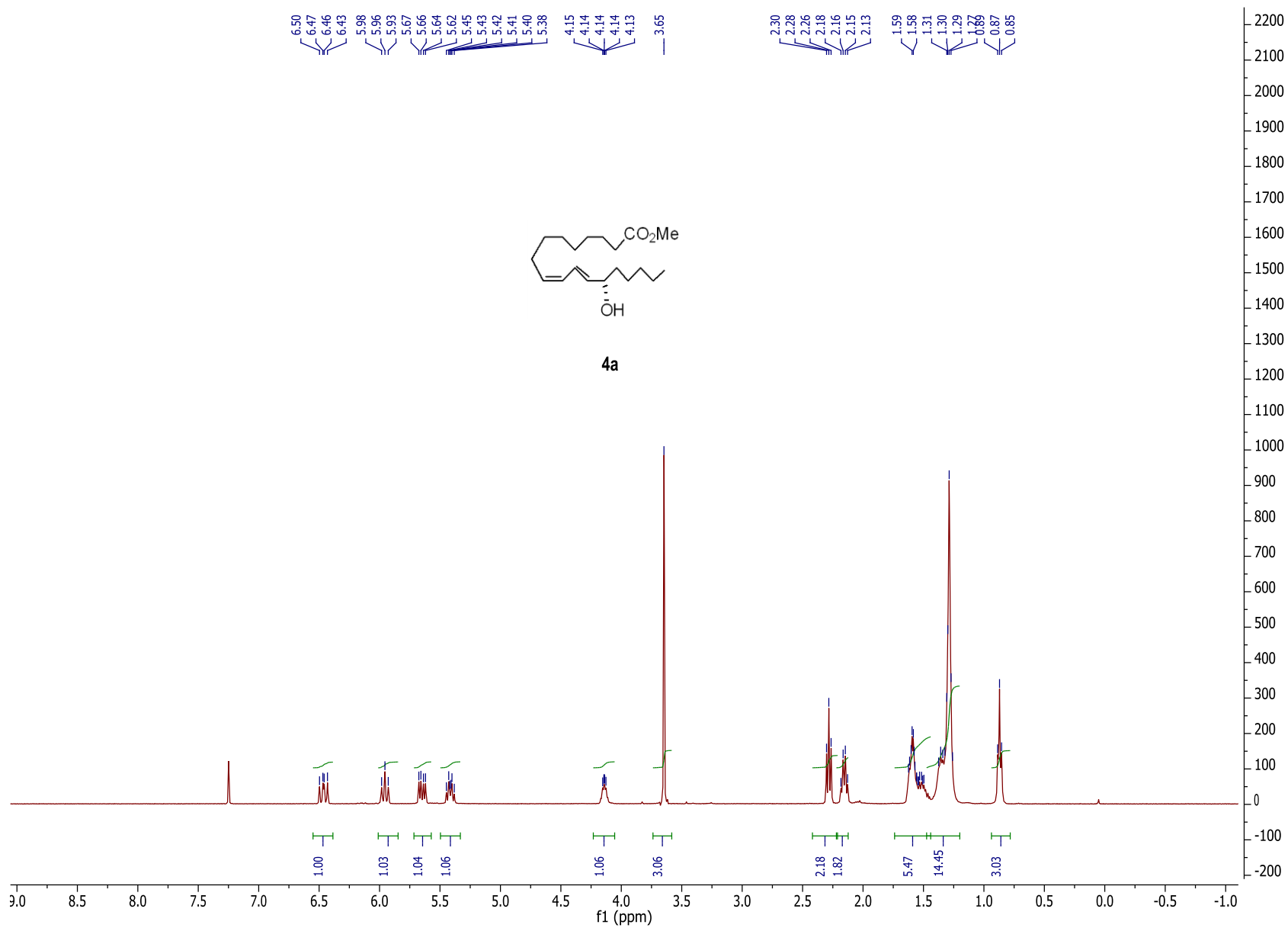


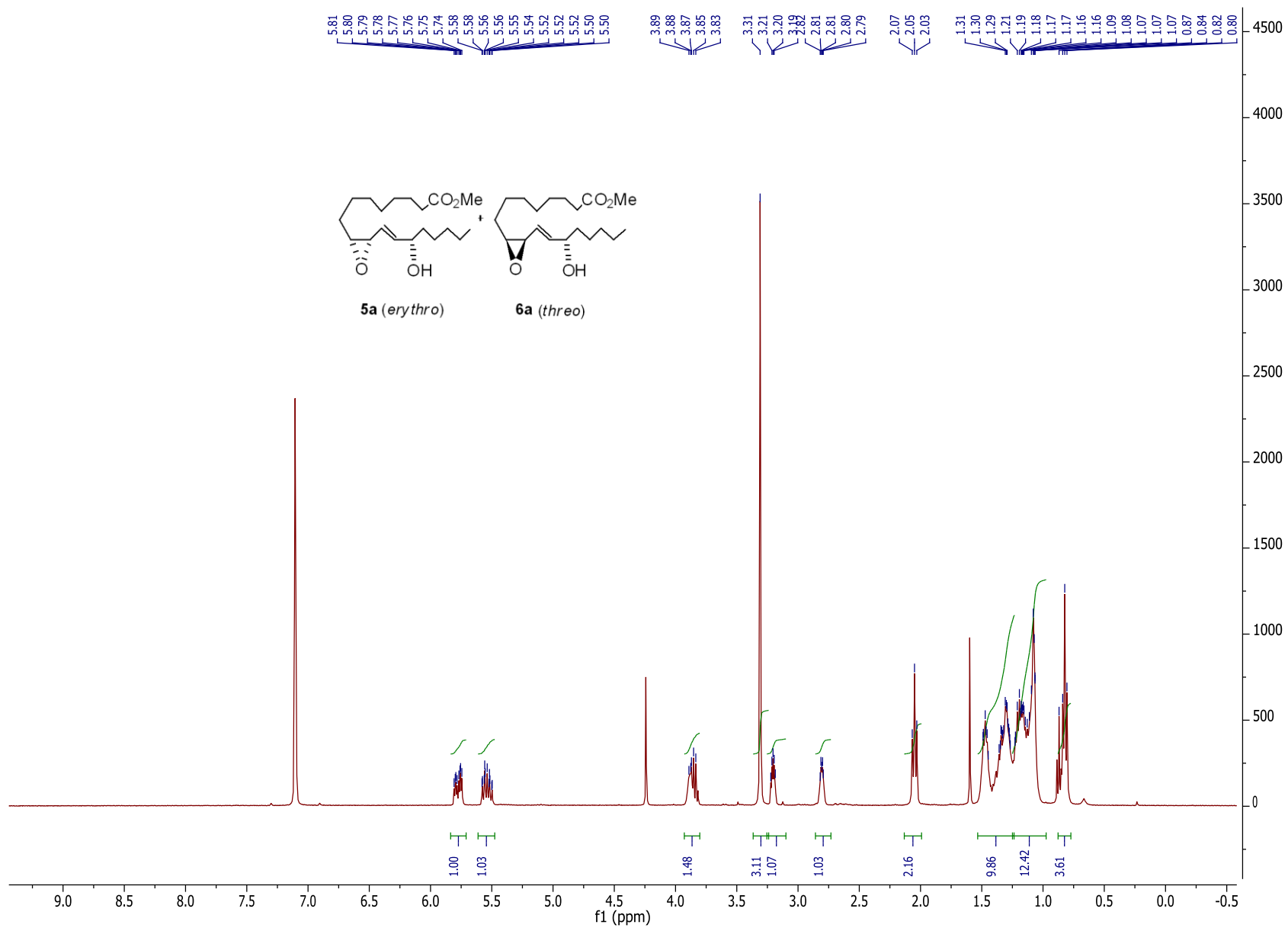
Peak	Ret. Time	Area	Height	Area %
1.	29.049	7725264	116942	100

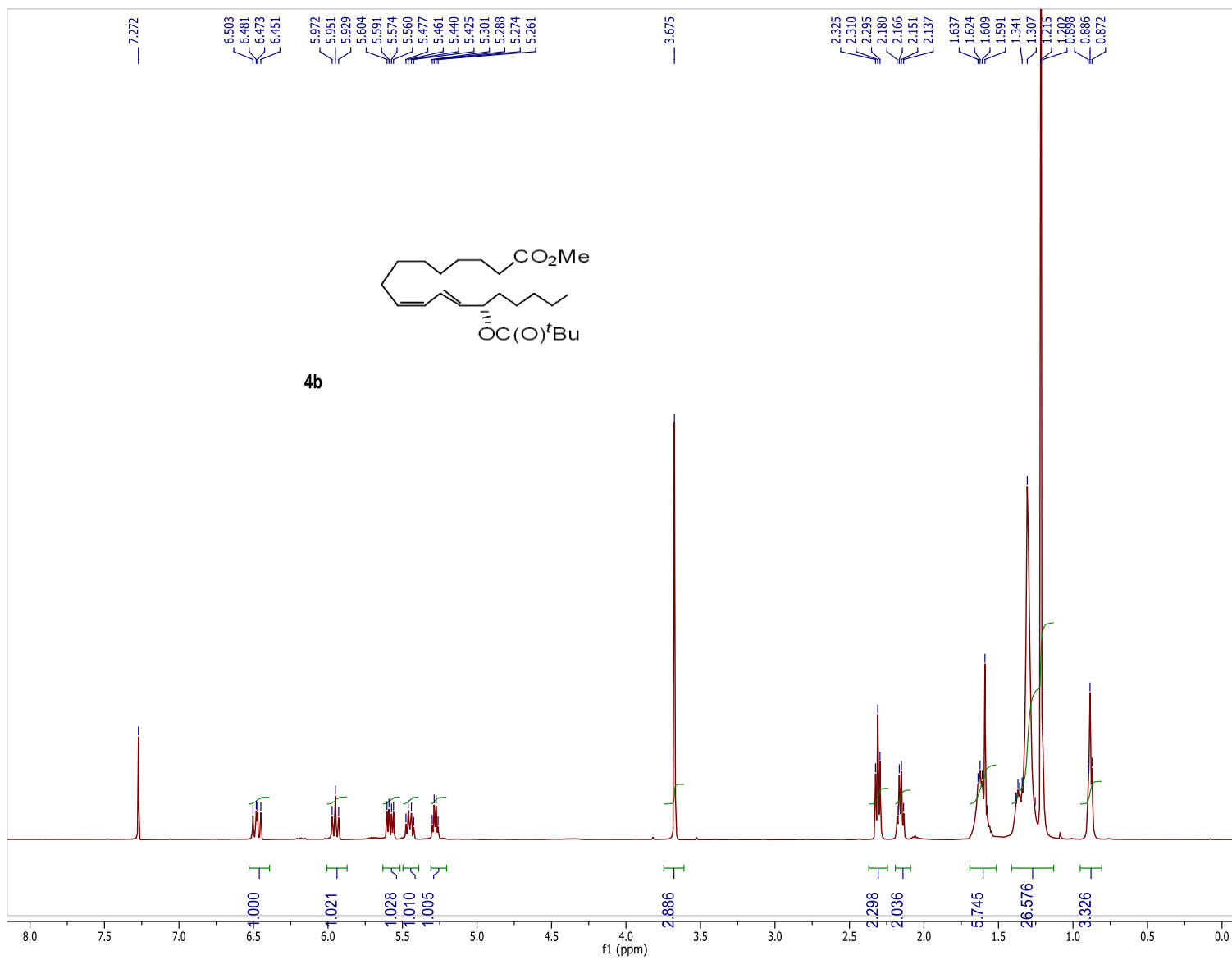
CHIRALCEL OJ-H, 15 CM, 4.6MM, 2.7 MICRON, Hexane:IPA (99.8:0.2), 0.8 mL/min, 205.

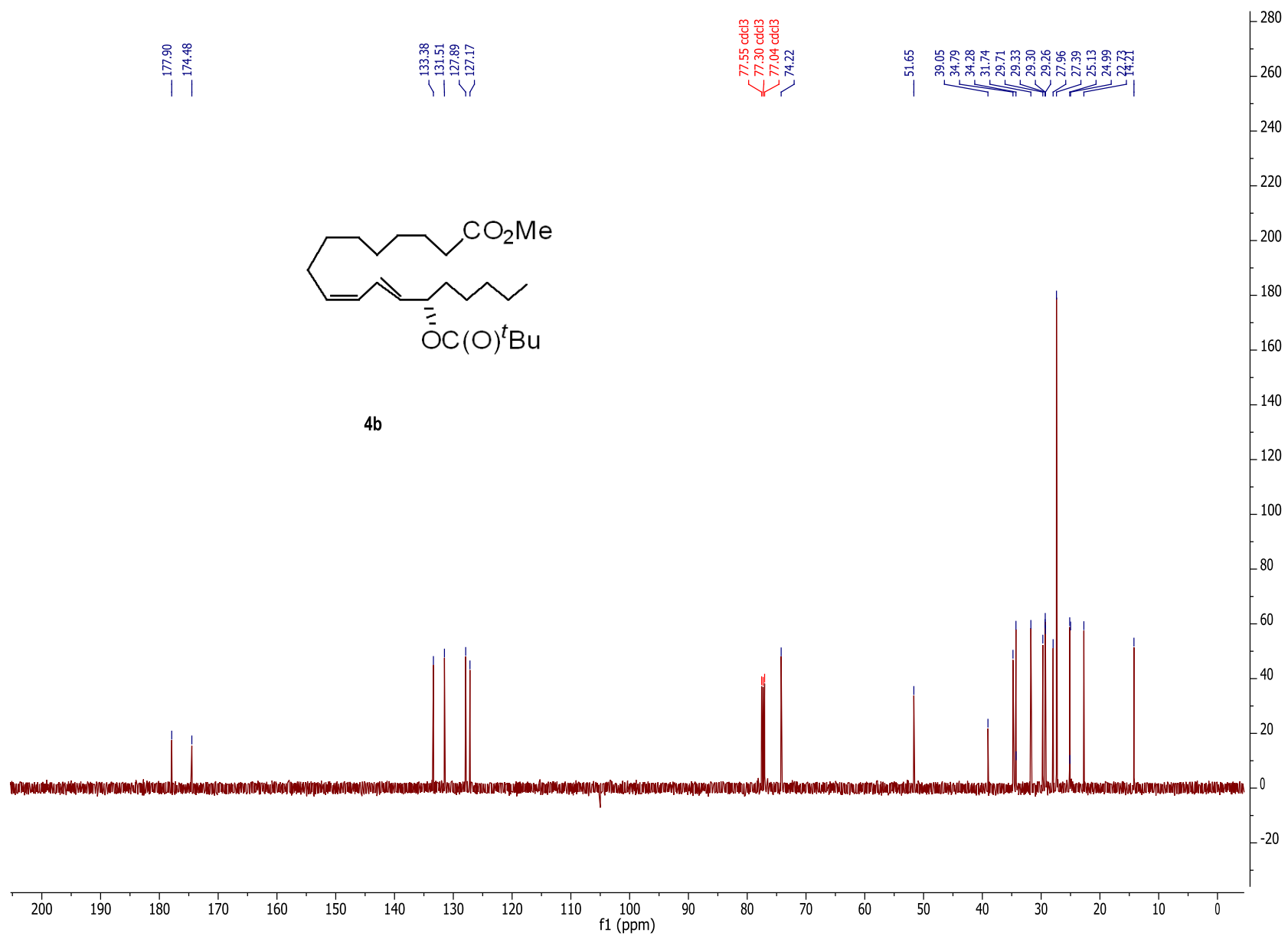
Note: Compound obtained from chiral epoxidation as described in reference

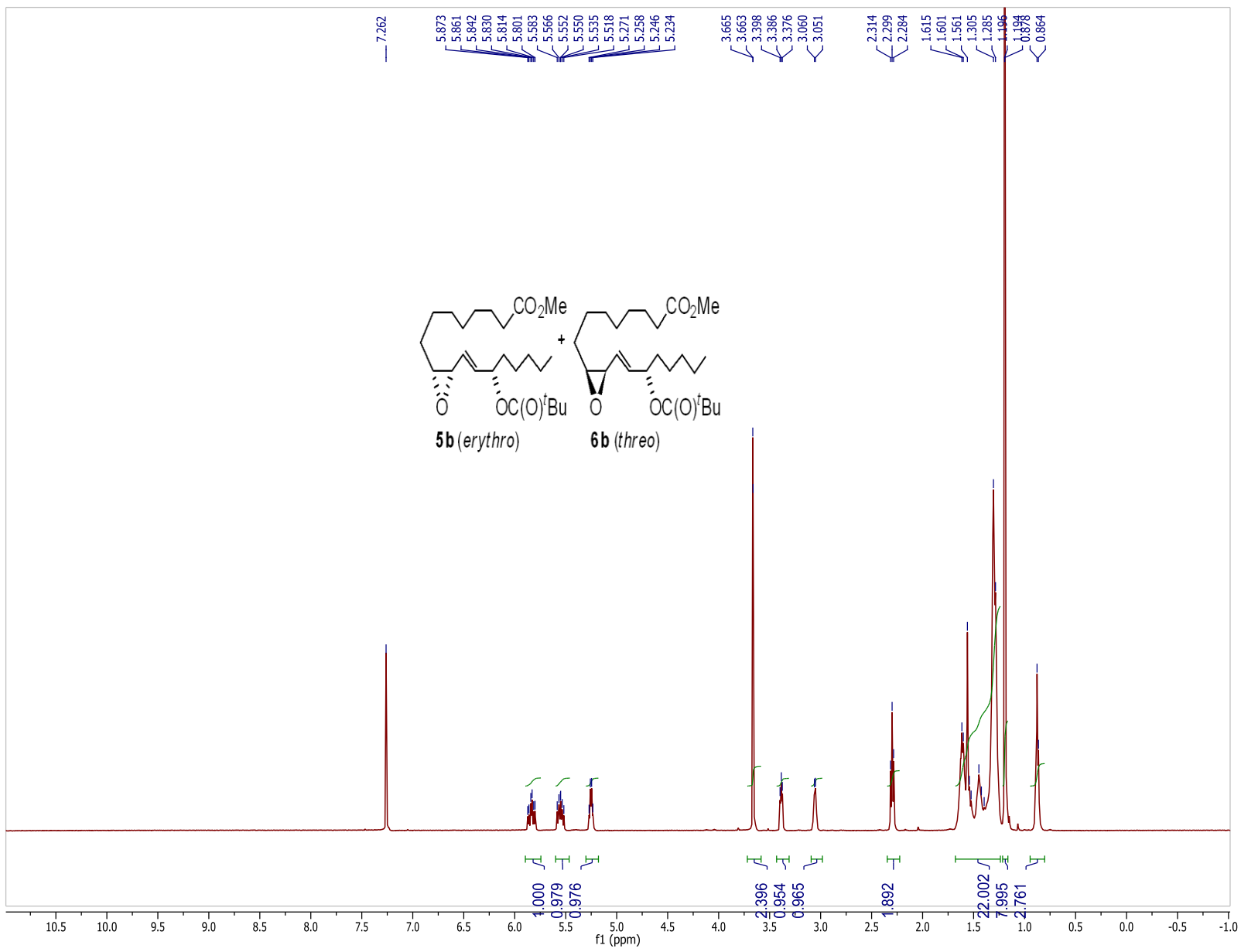








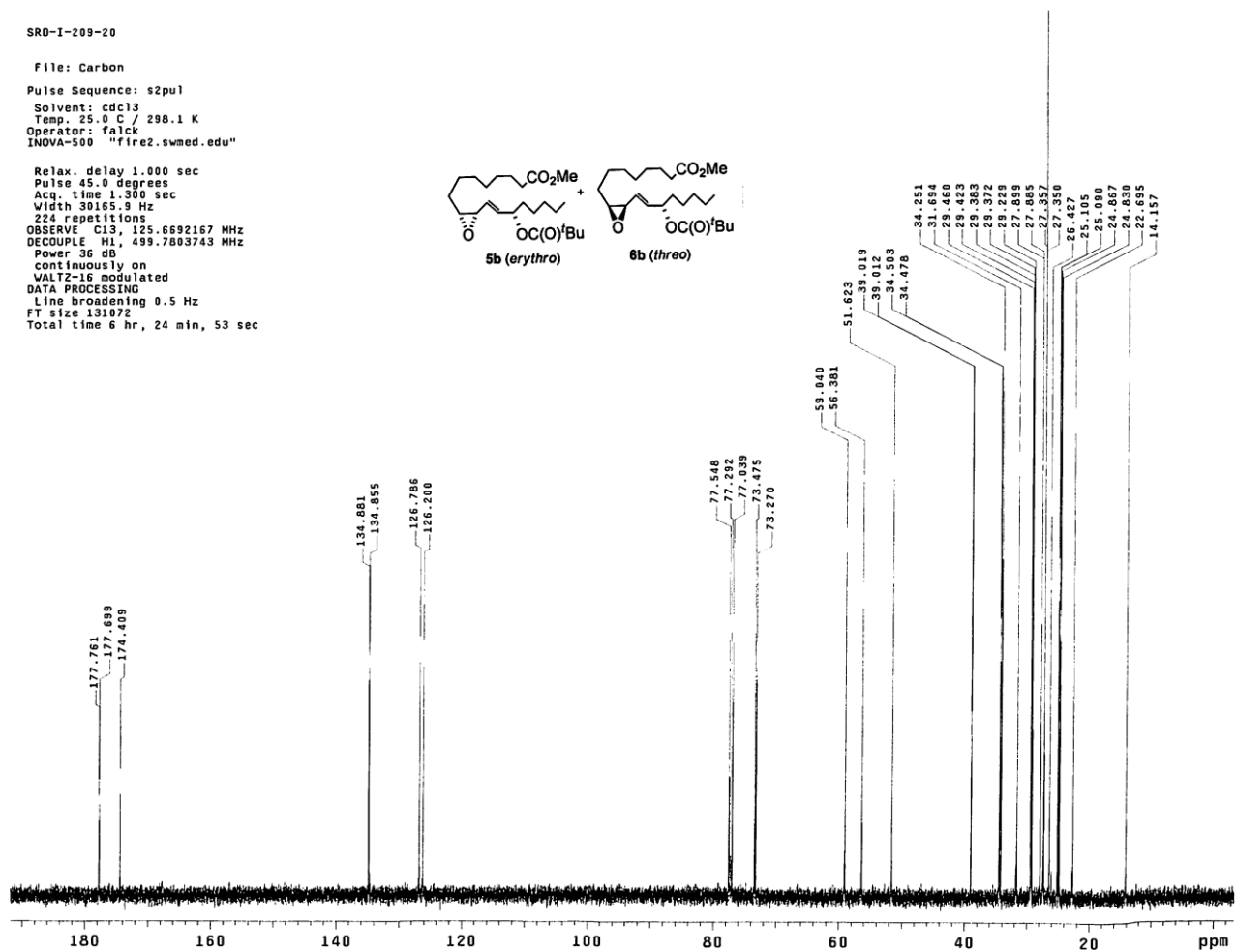
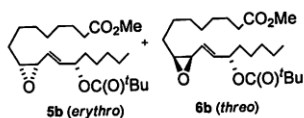


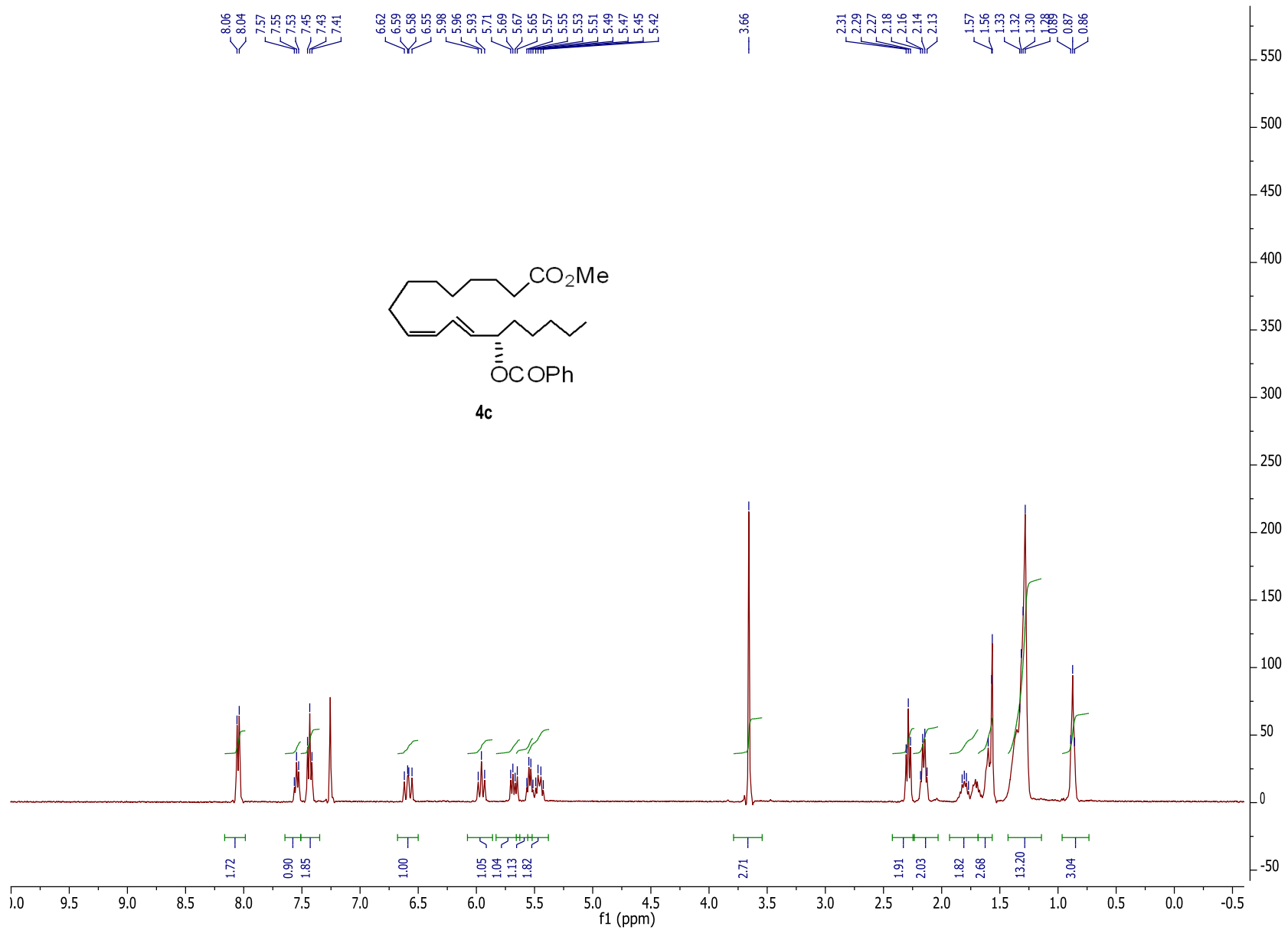


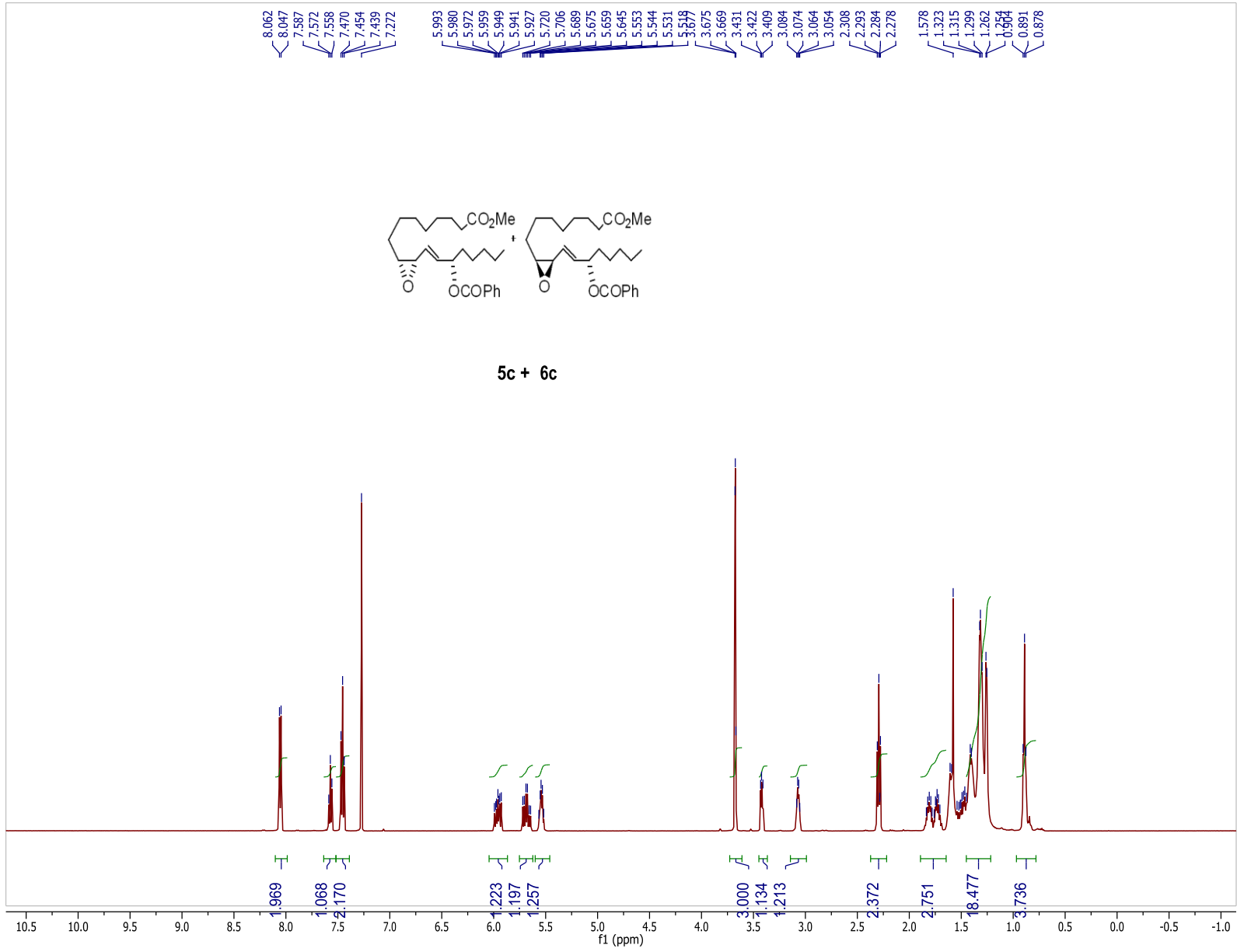
SR0-I-209-20

File: Carbon  
Pulse Sequence: s2pu1  
Solvent: cdCl3  
Temp. 25.0 C / 298.1 K  
Operator: falck  
INOVA-500 "fire2.swmed.edu"

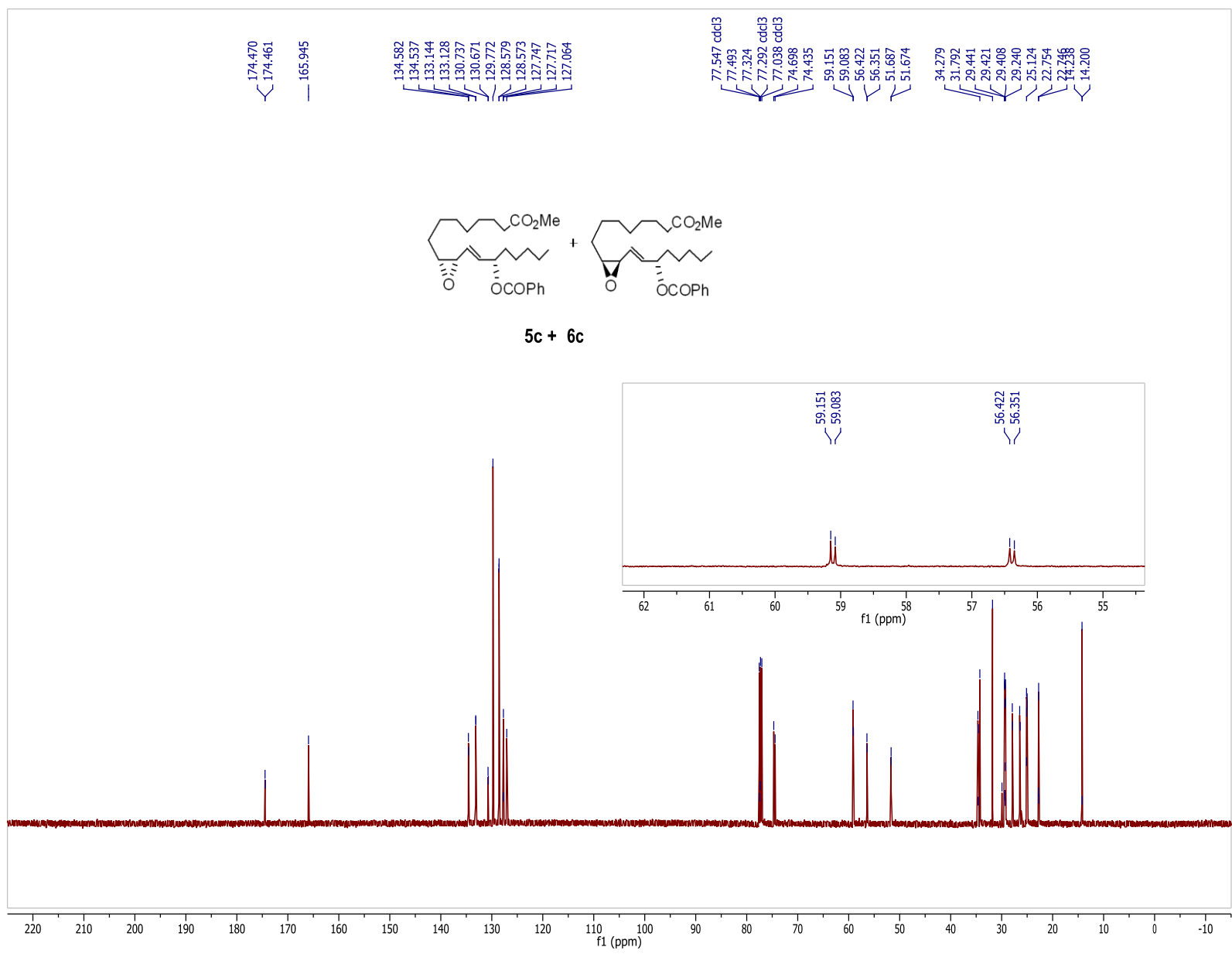
Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.300 sec  
Width 30165.9 Hz  
224 repetitions  
OBSERVE C13, 125.6692167 MHz  
DECOUPLE H1, 499.7803743 MHz  
Power 36 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.5 Hz  
FT size 131072  
Total time 6 hr, 24 min, 53 sec

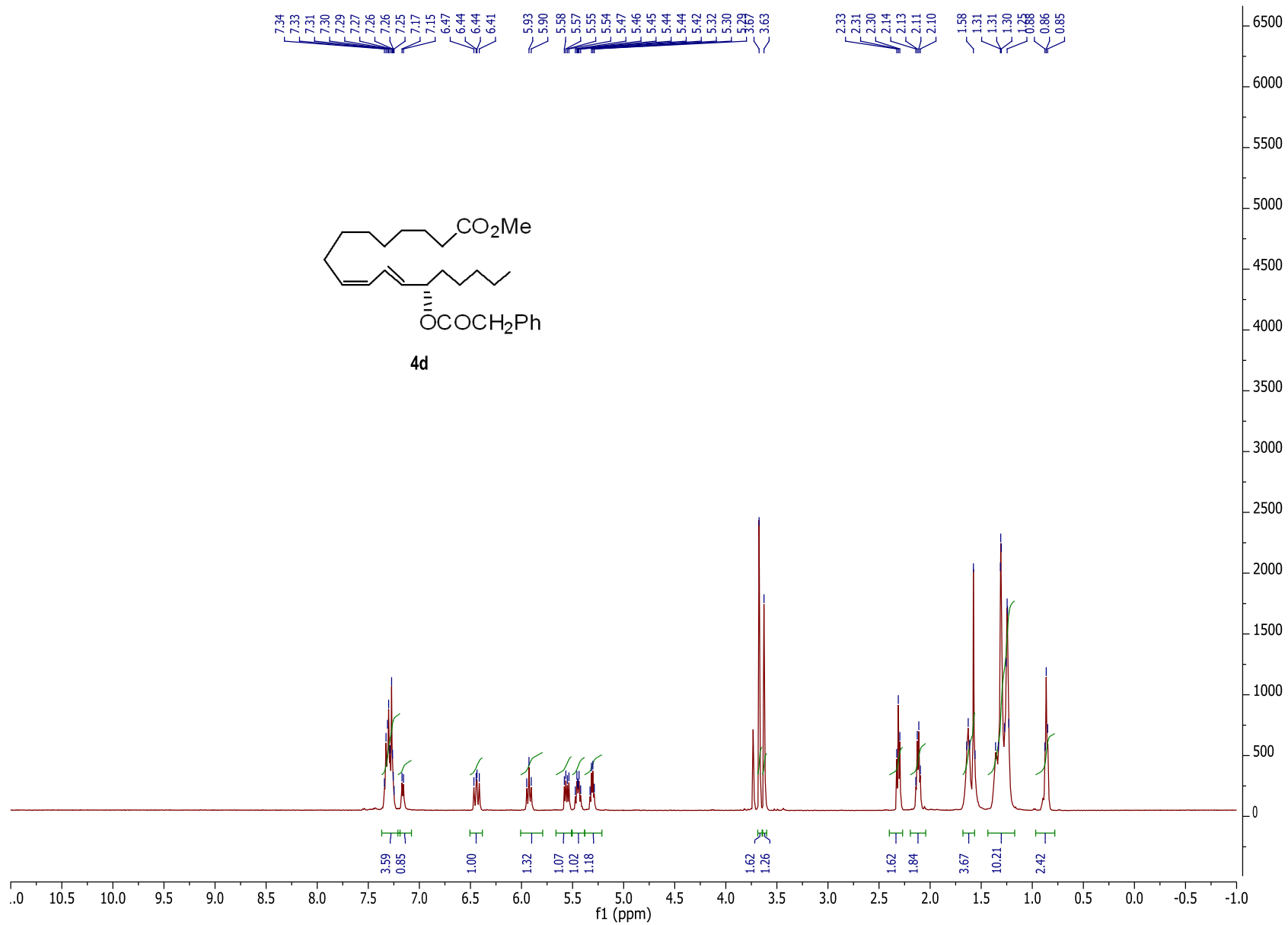


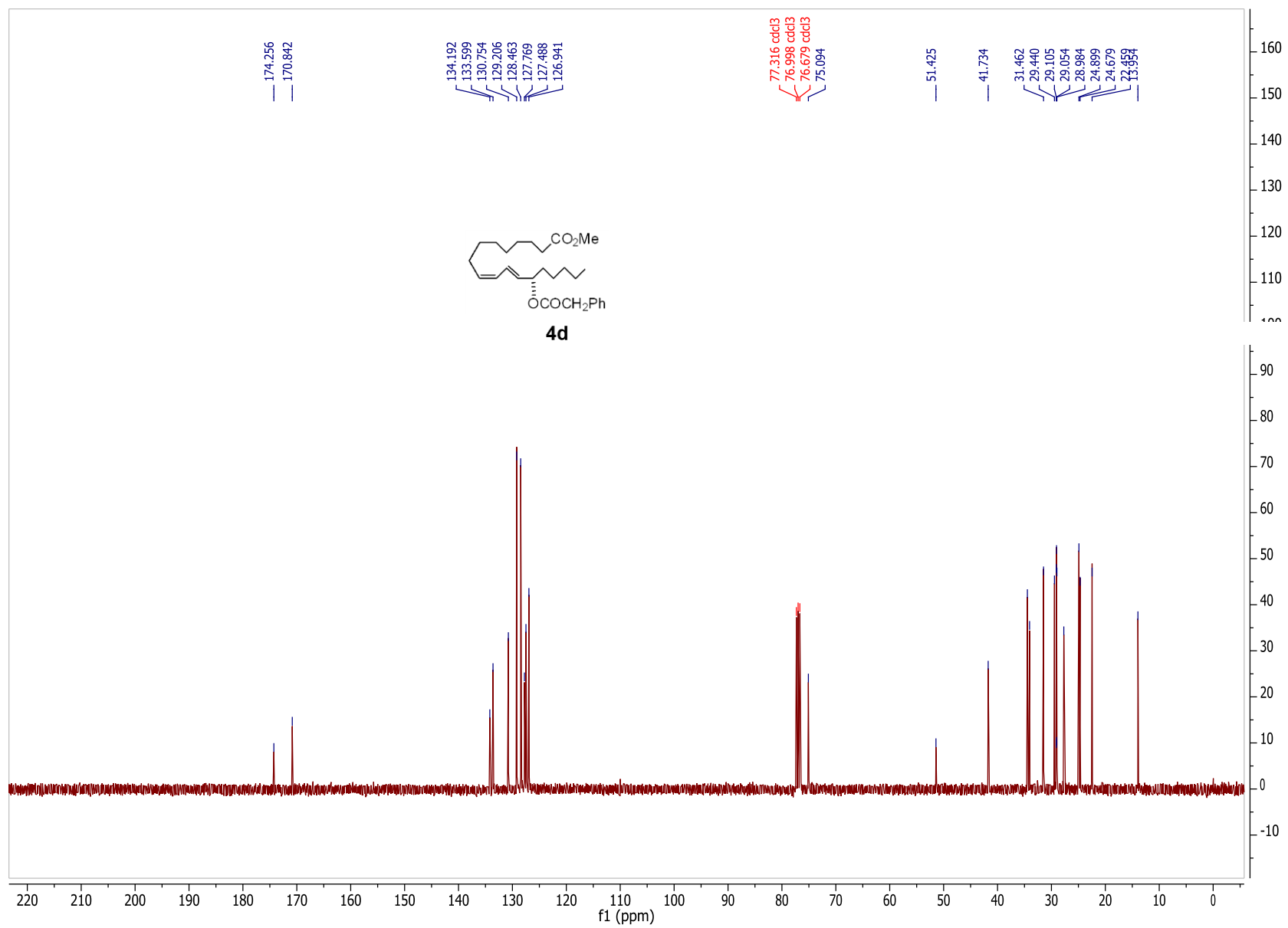


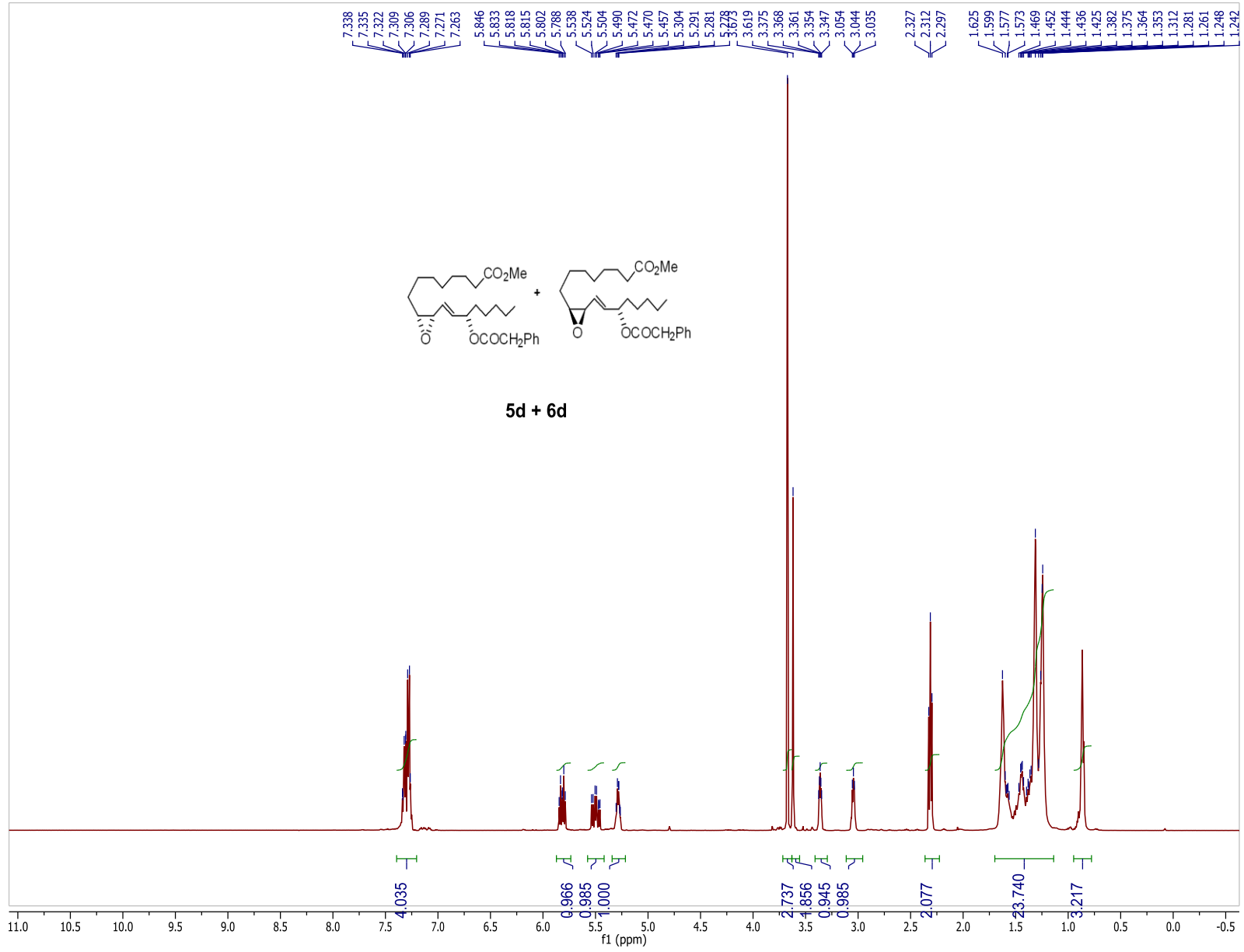


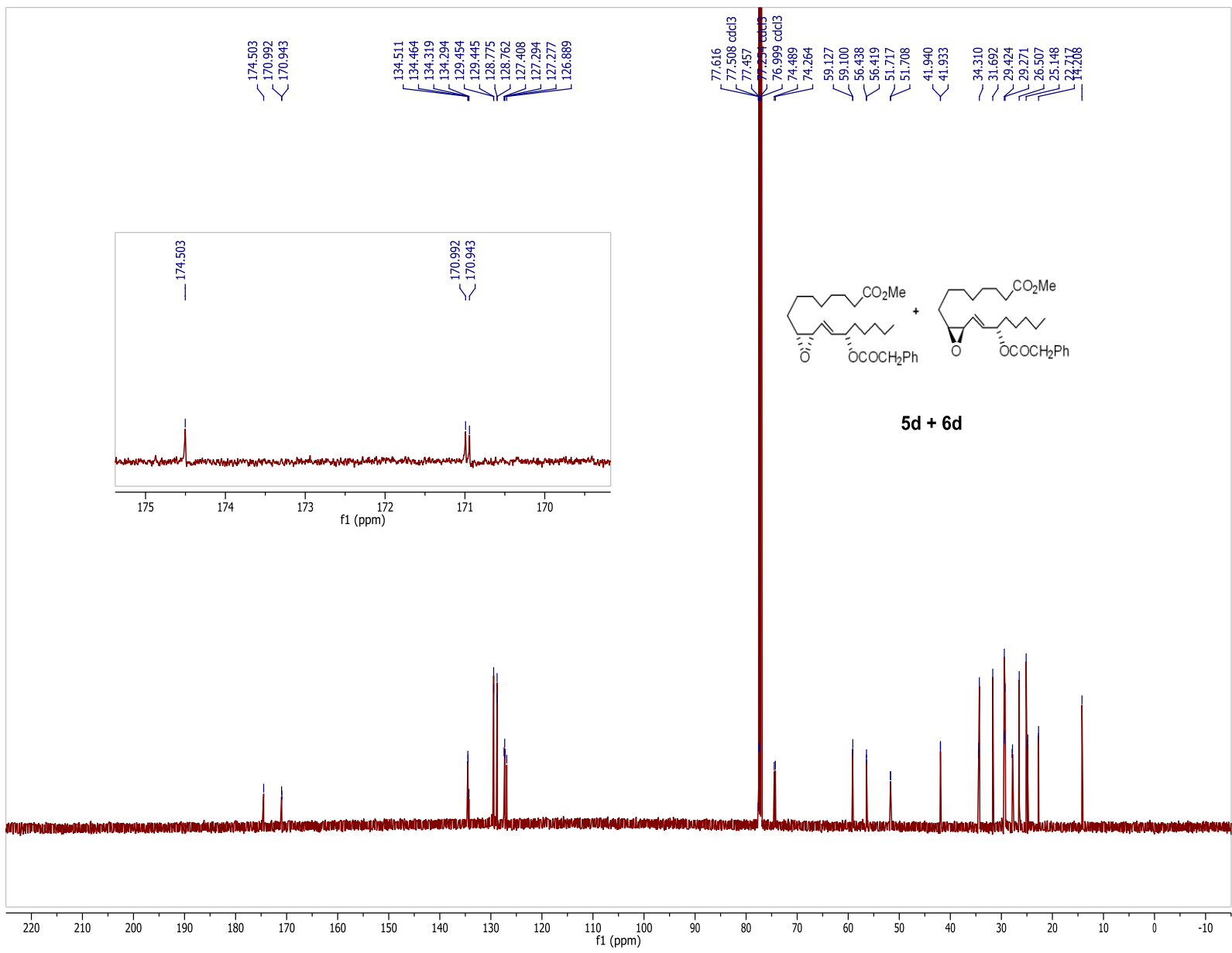


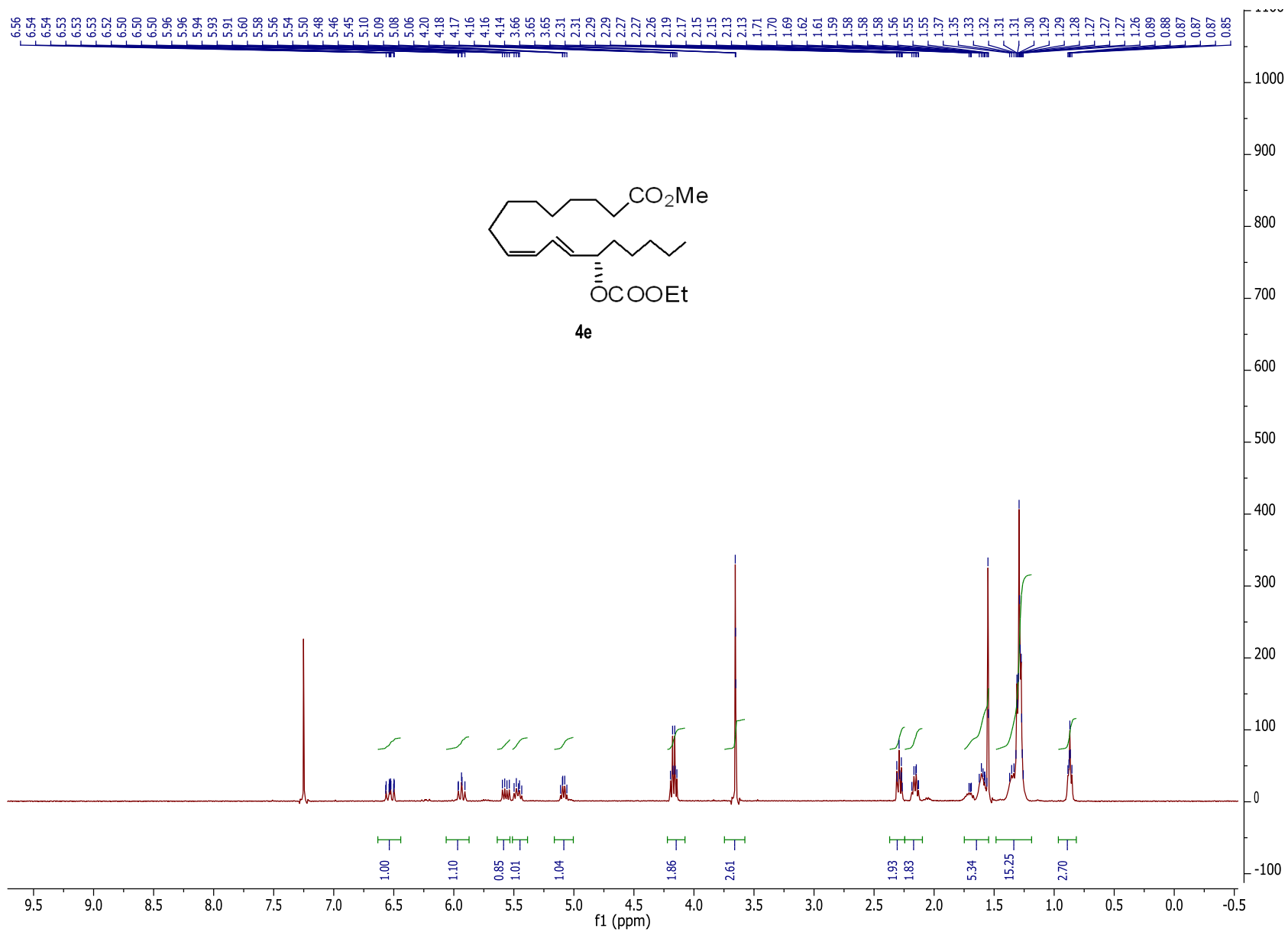


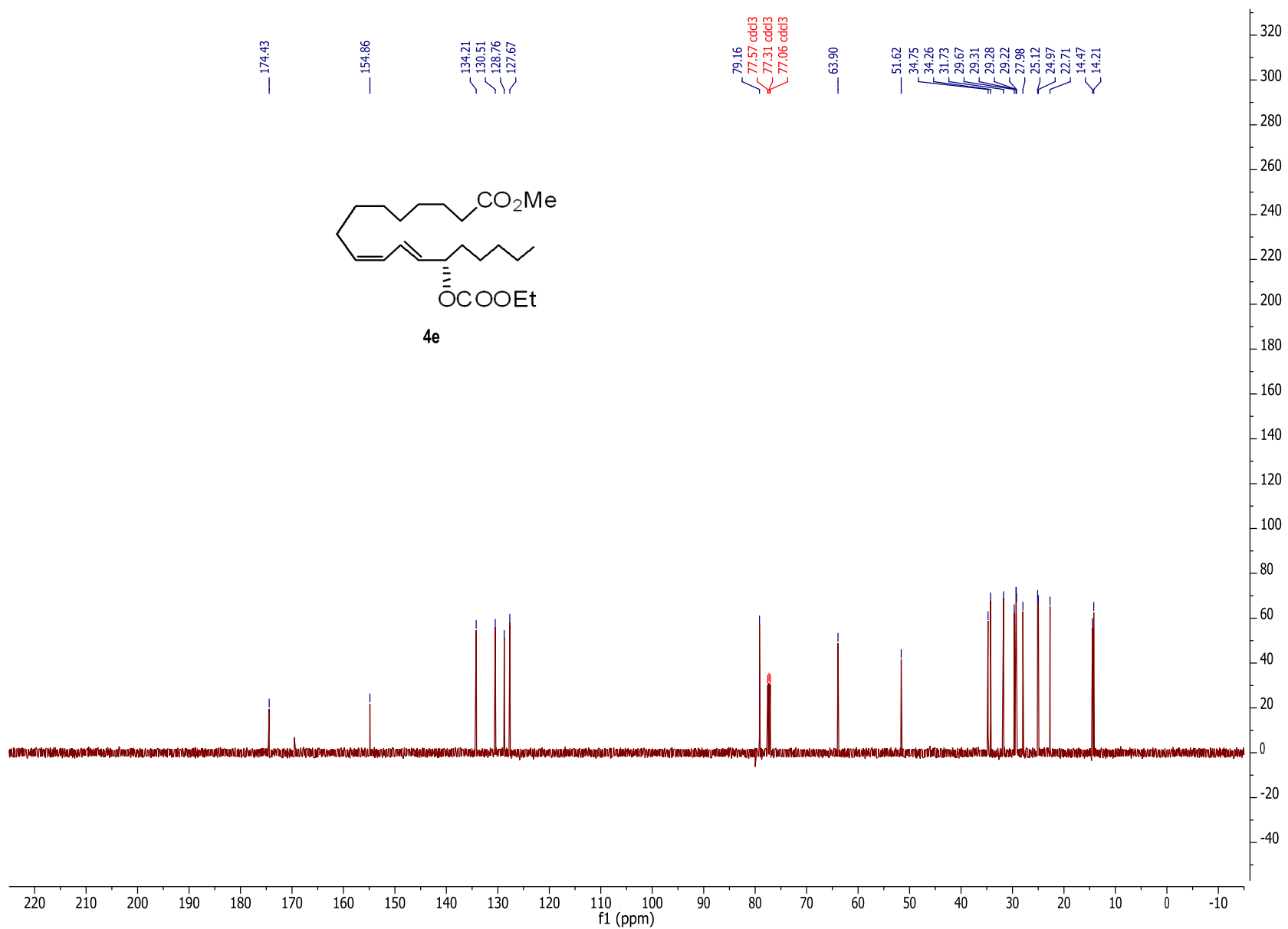


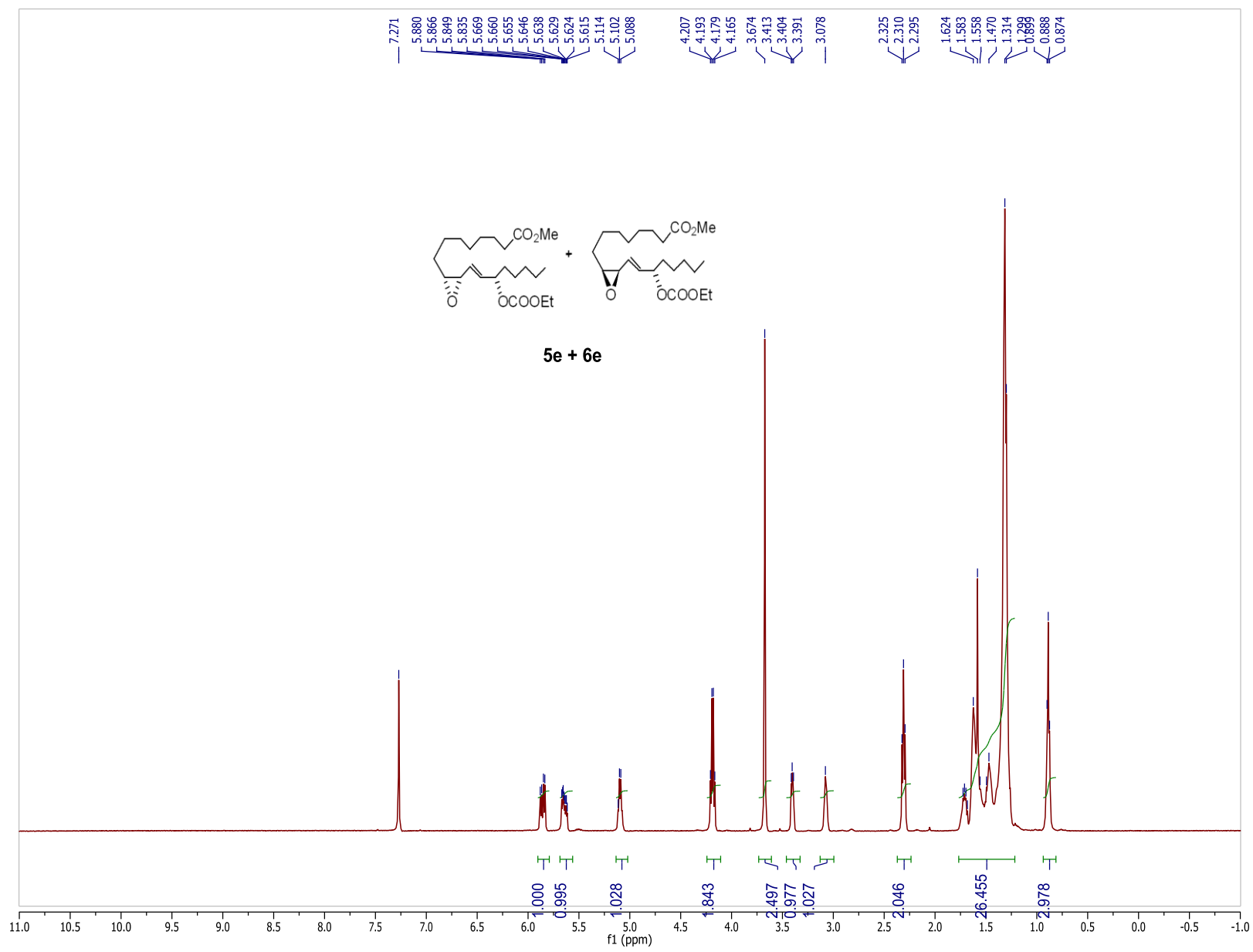




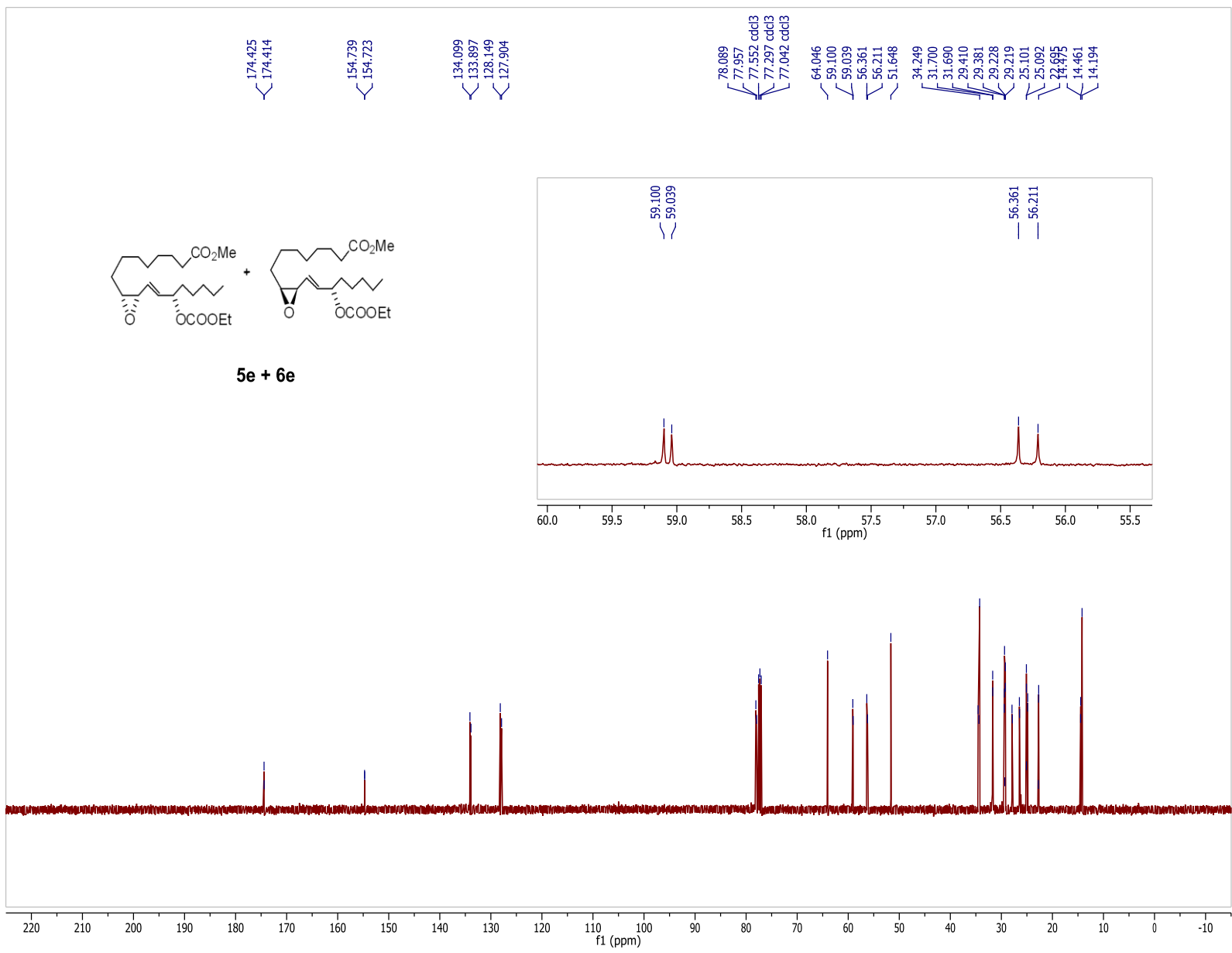


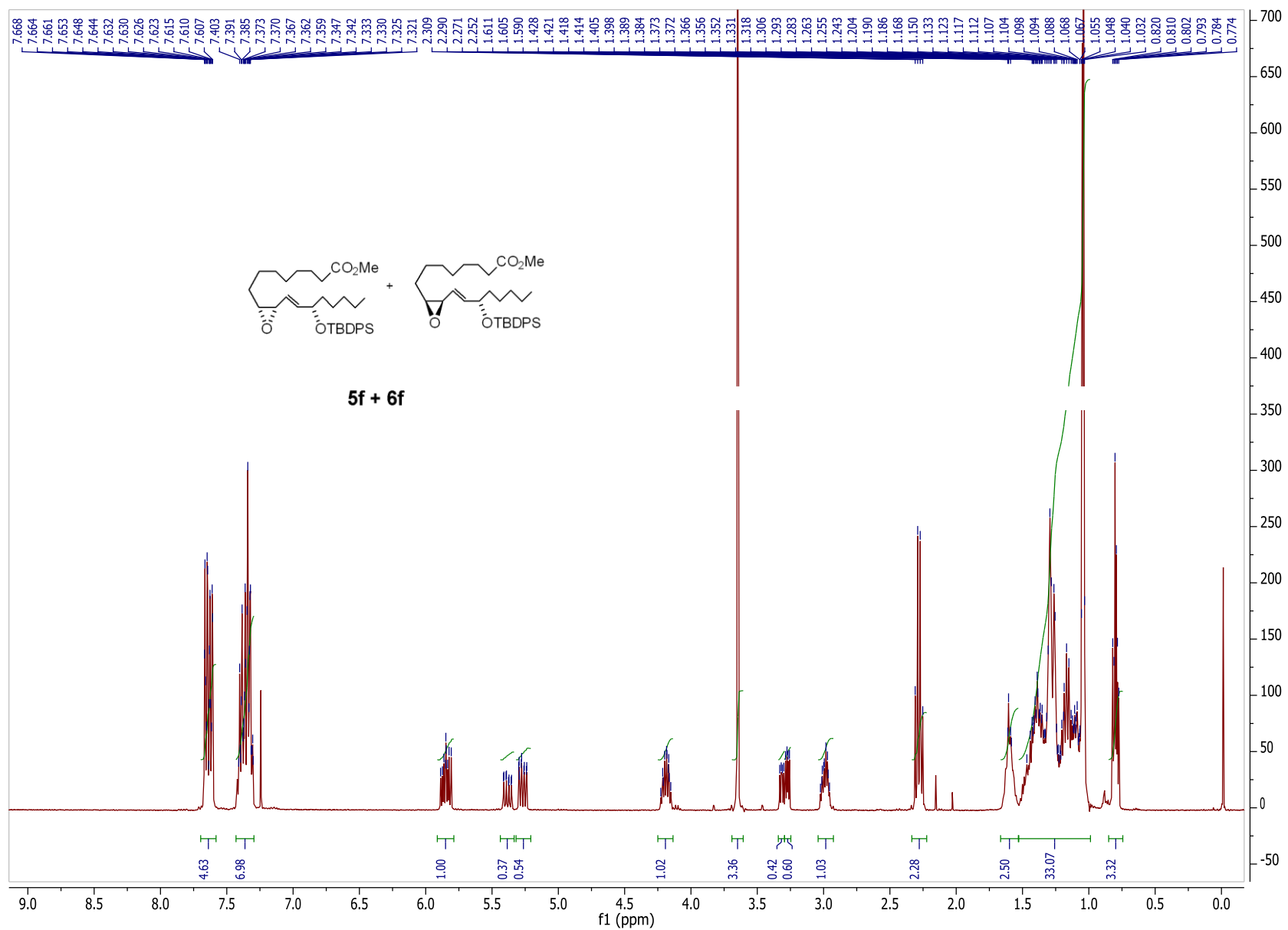


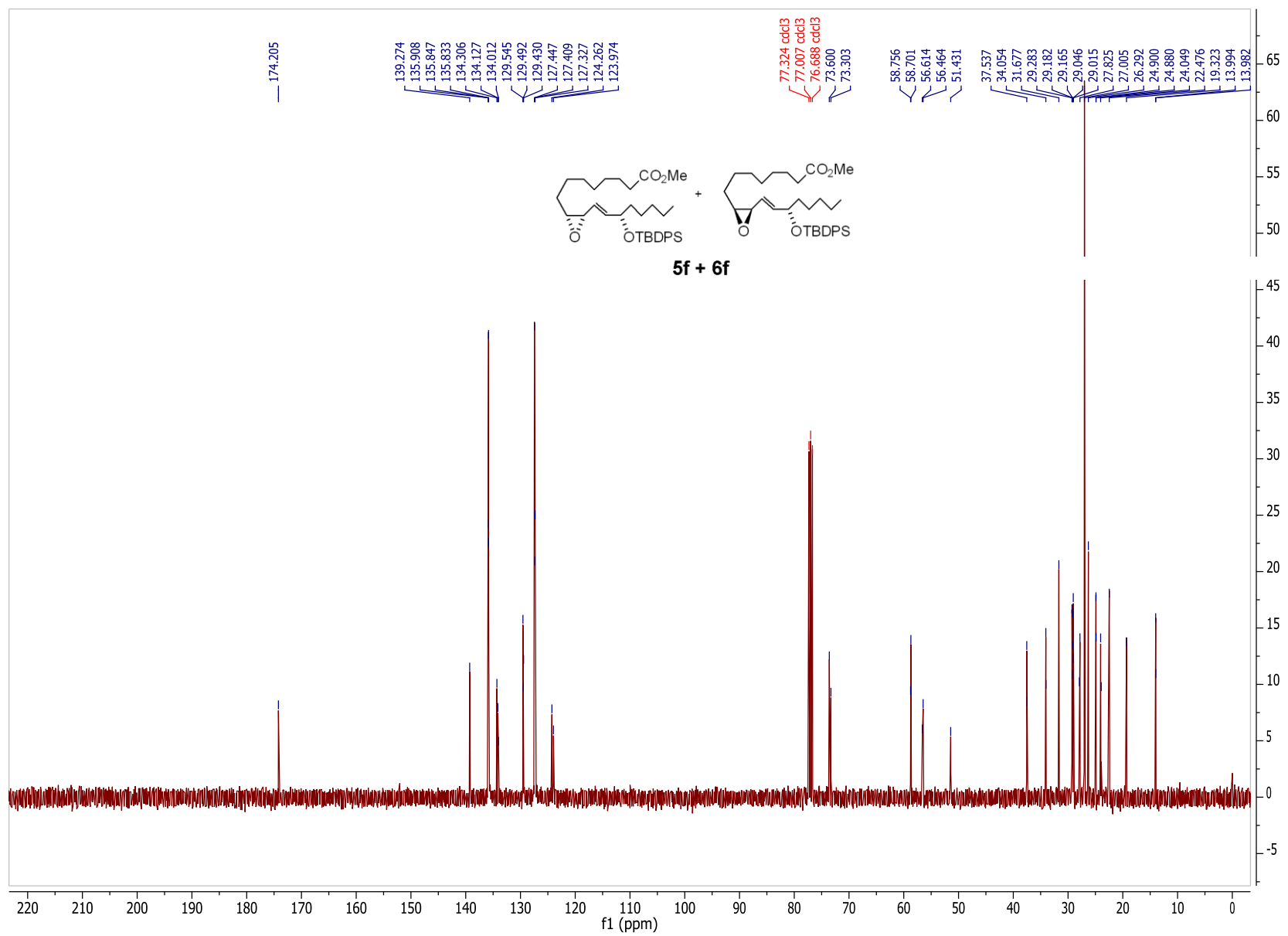


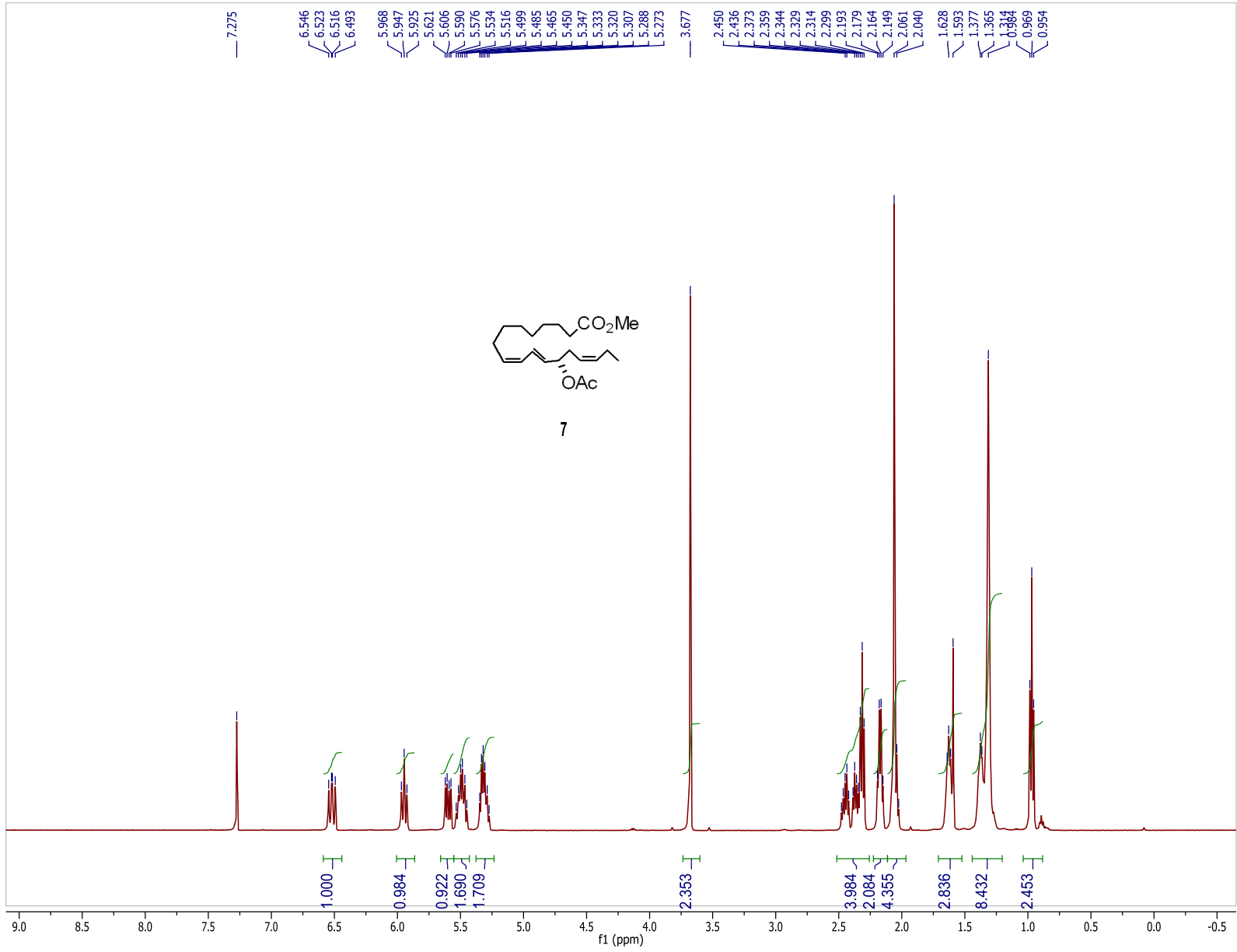


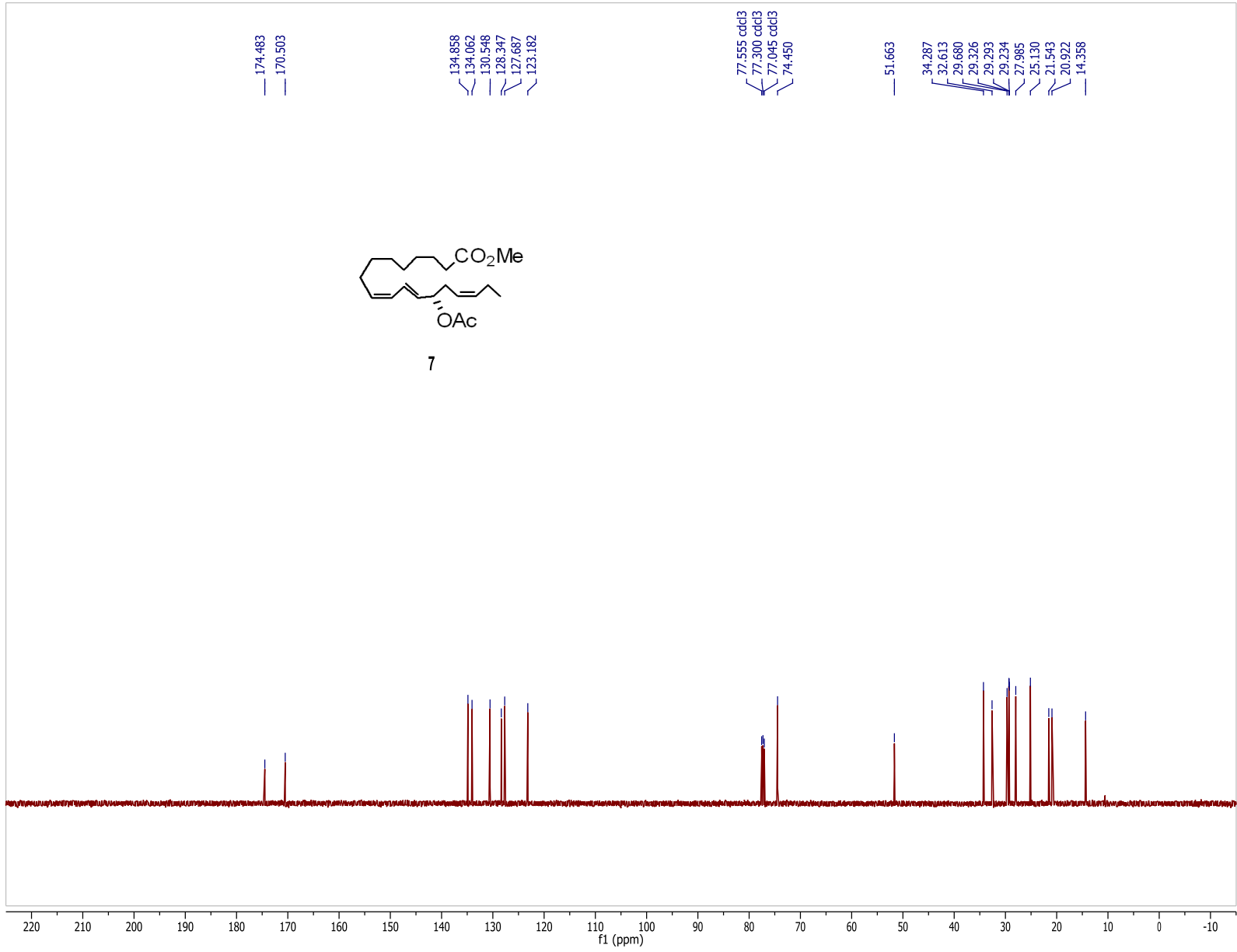


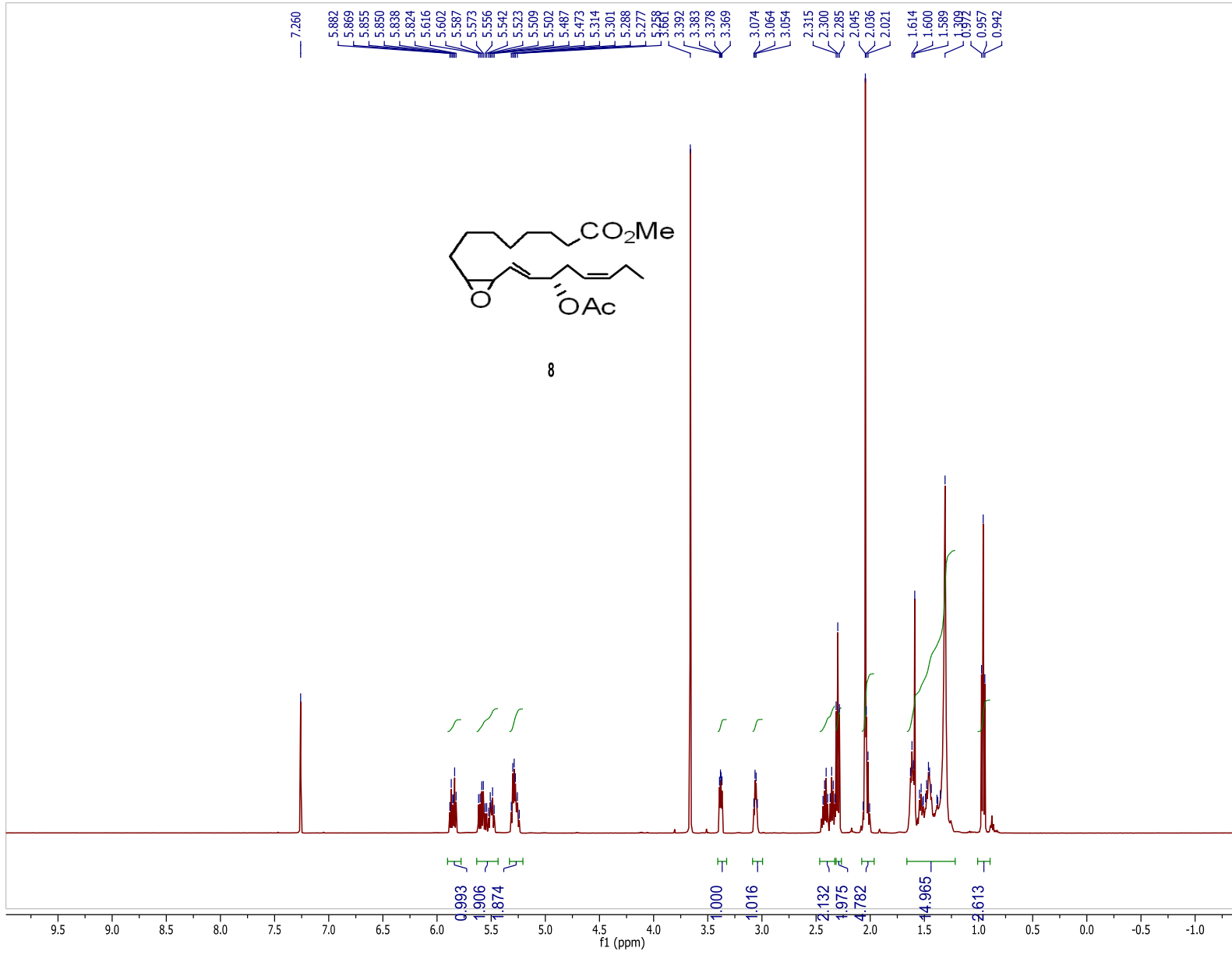


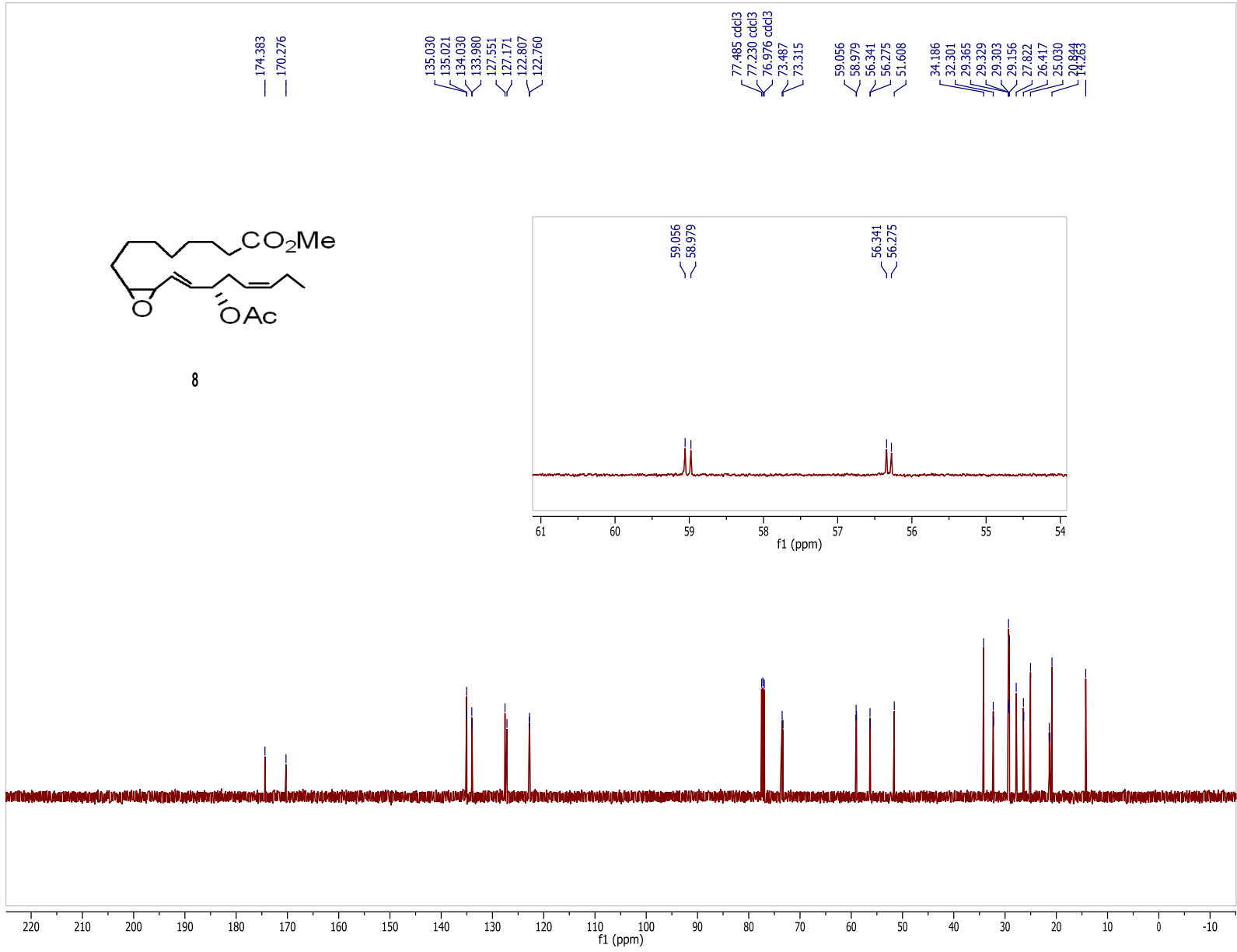


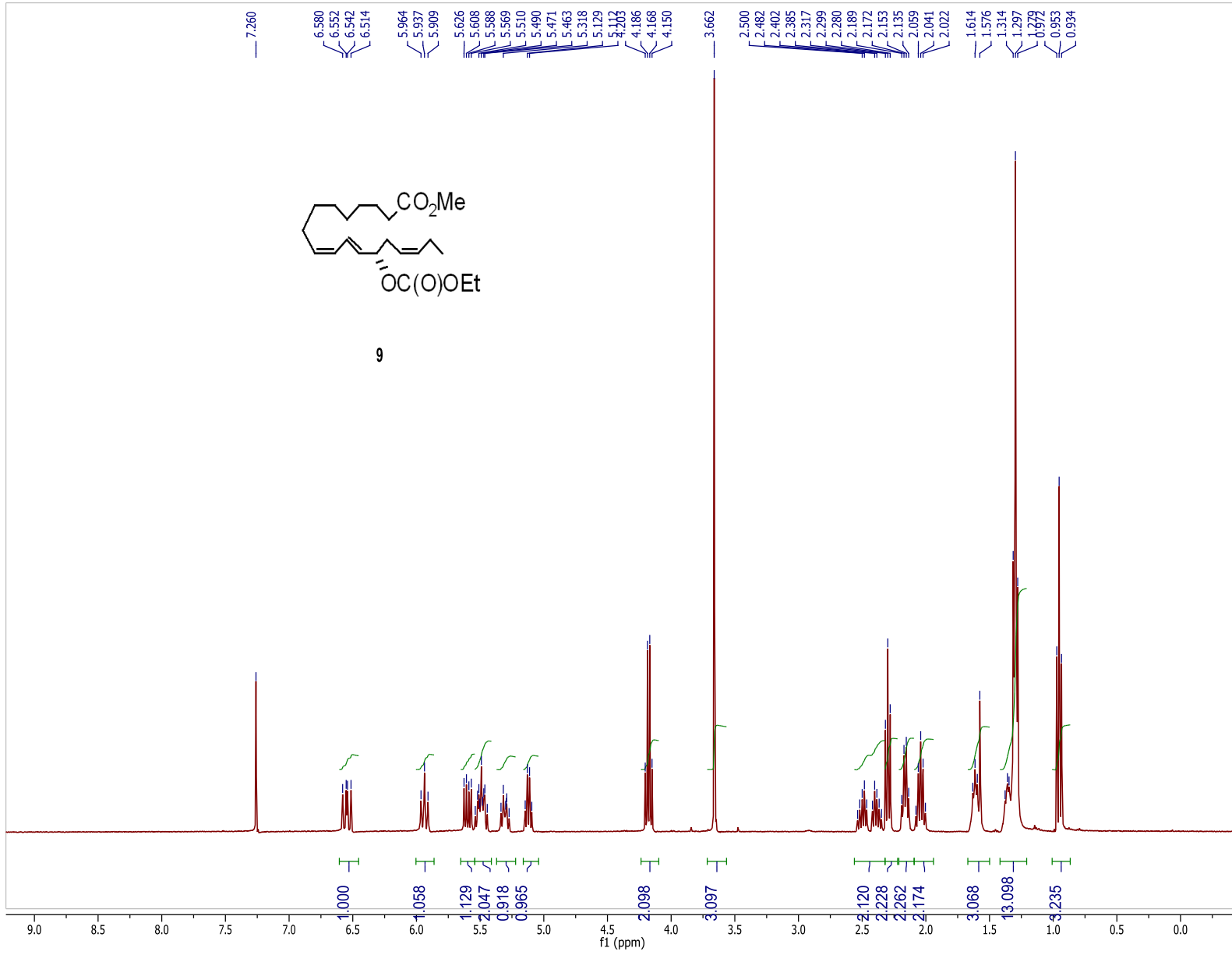




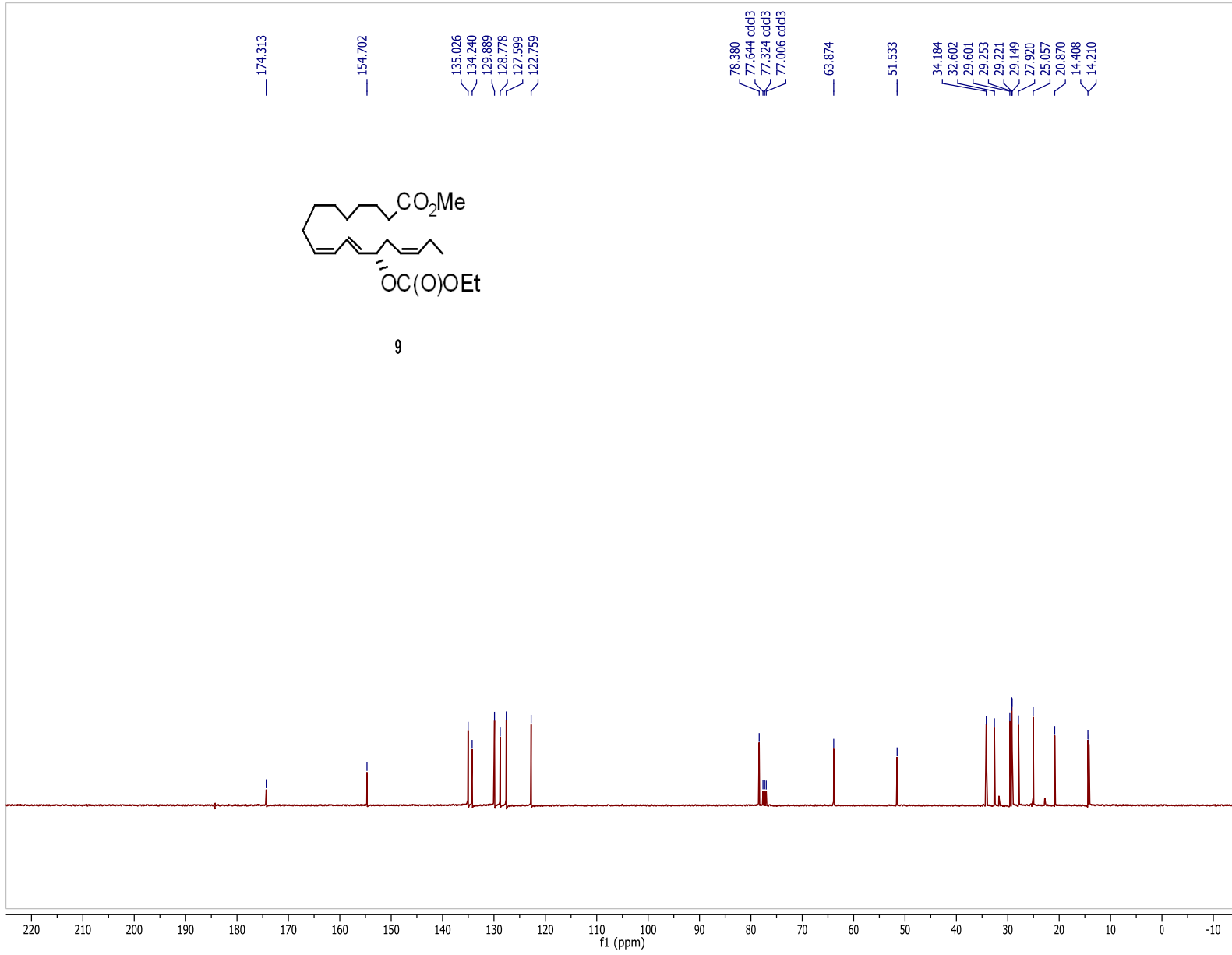


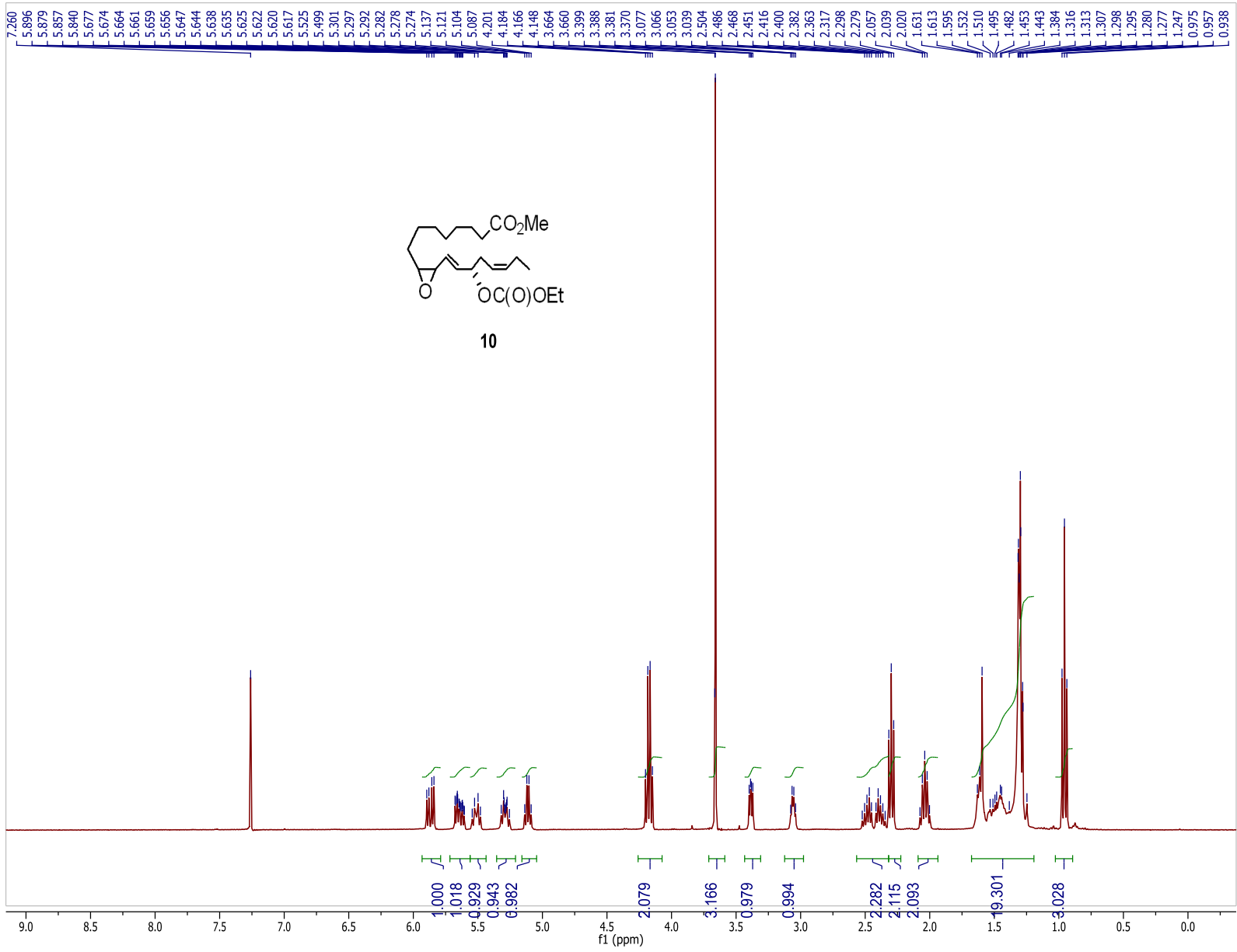


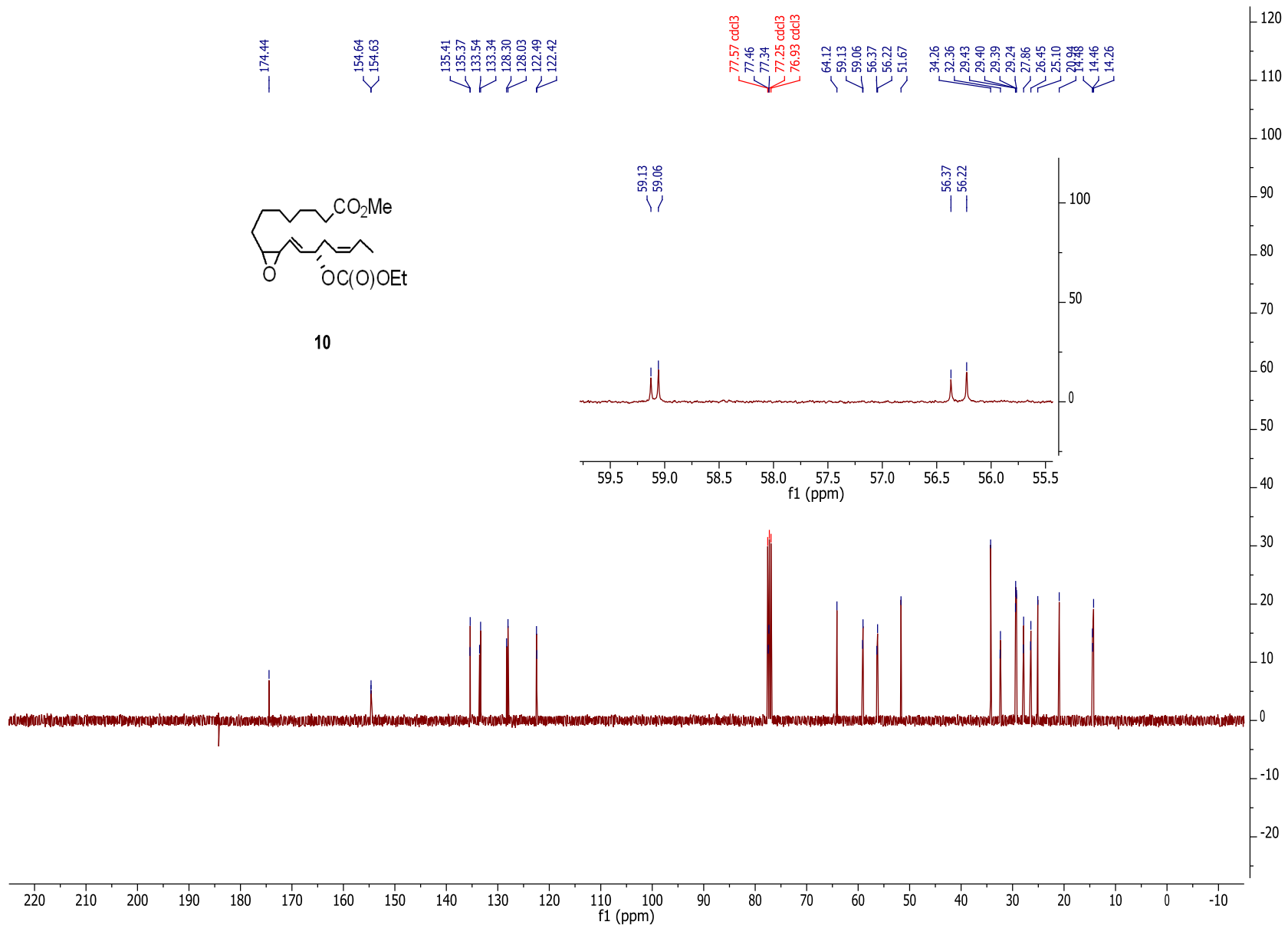


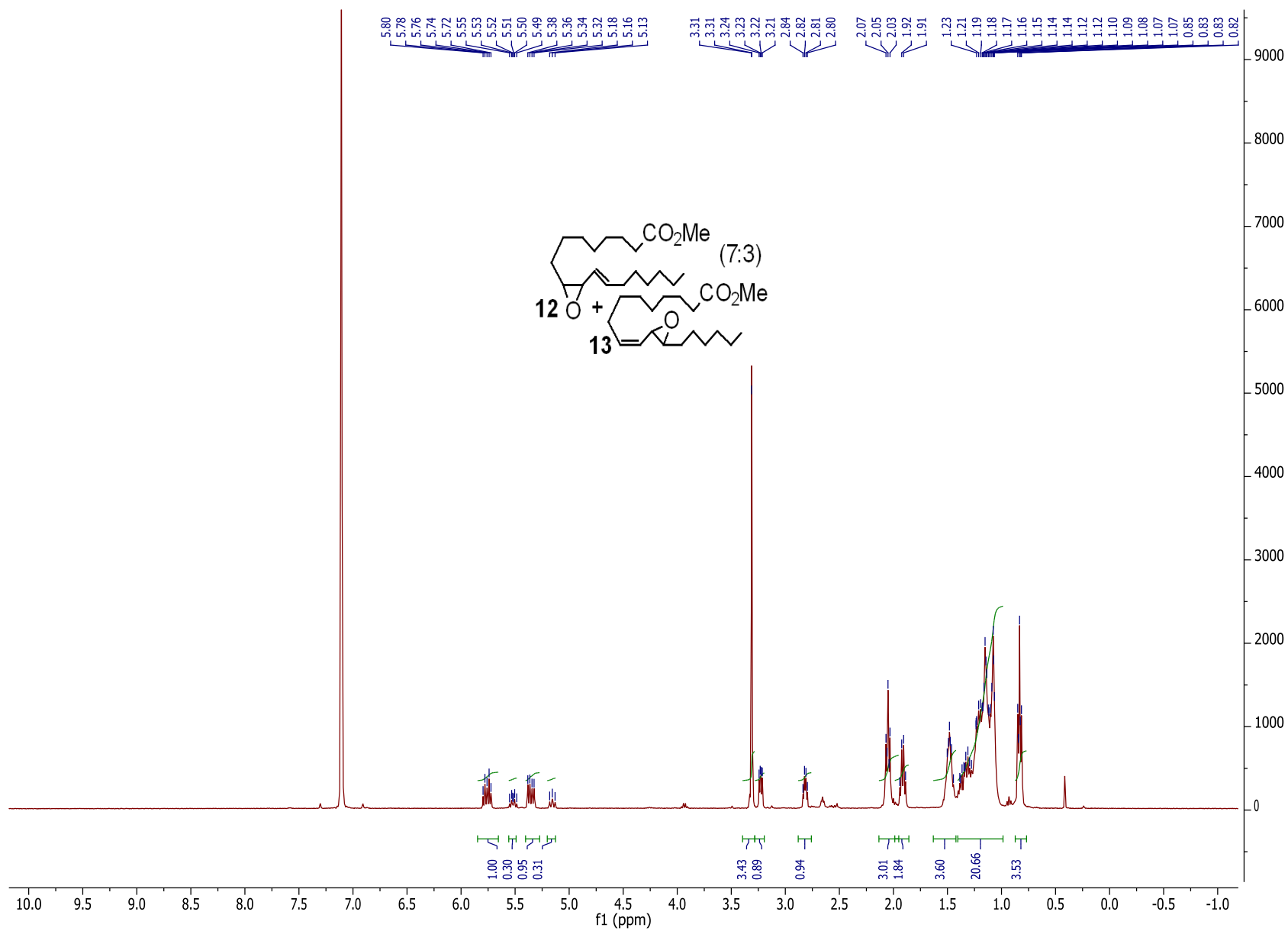


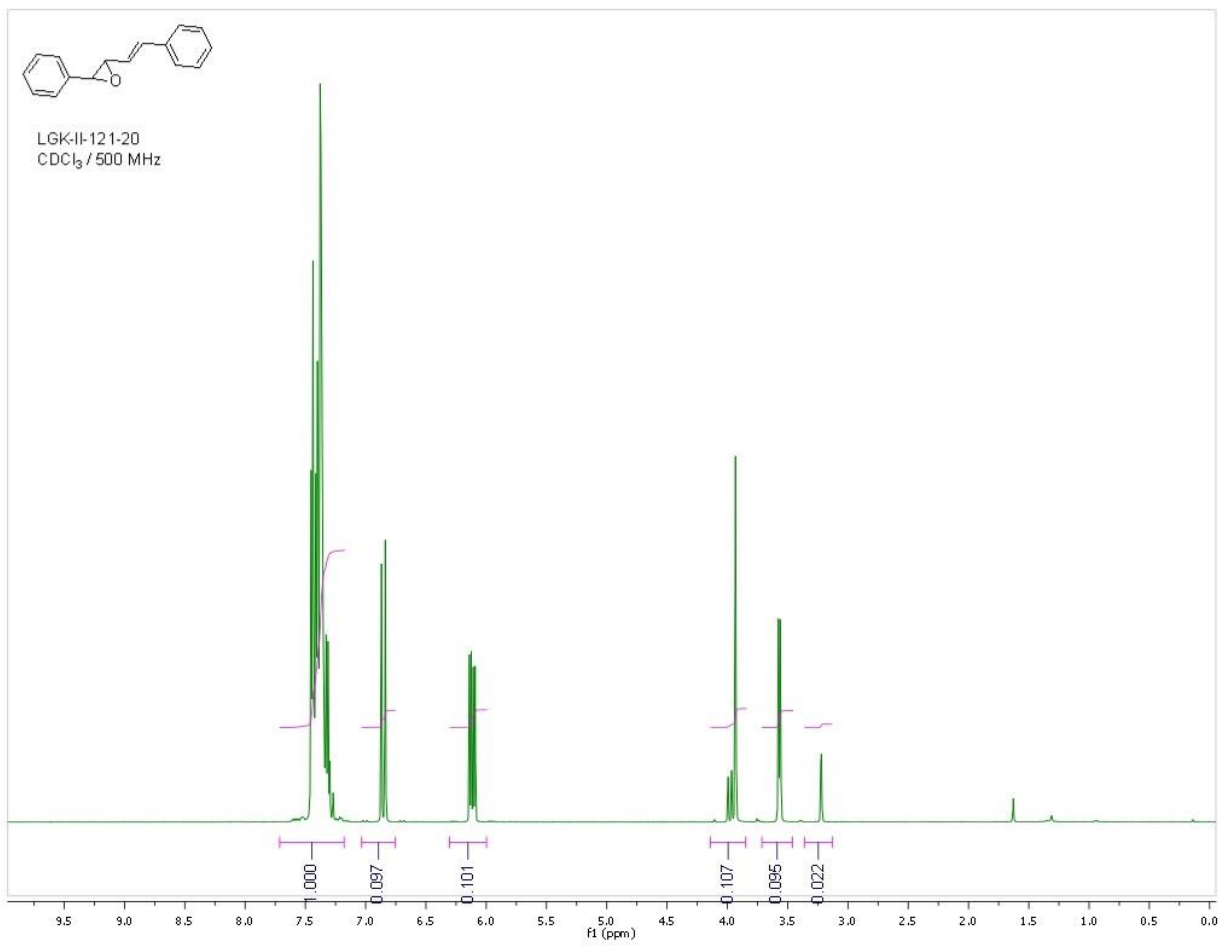


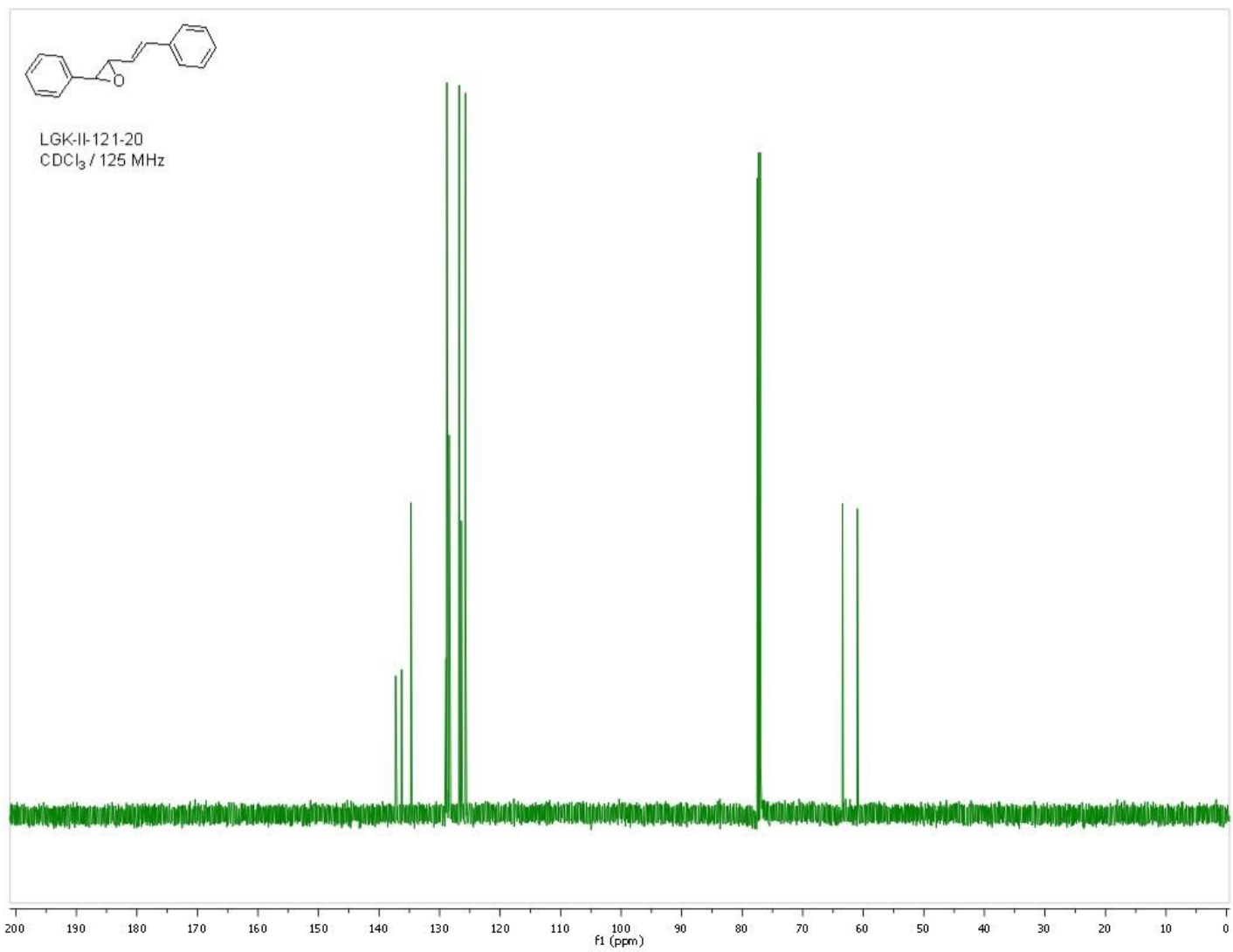


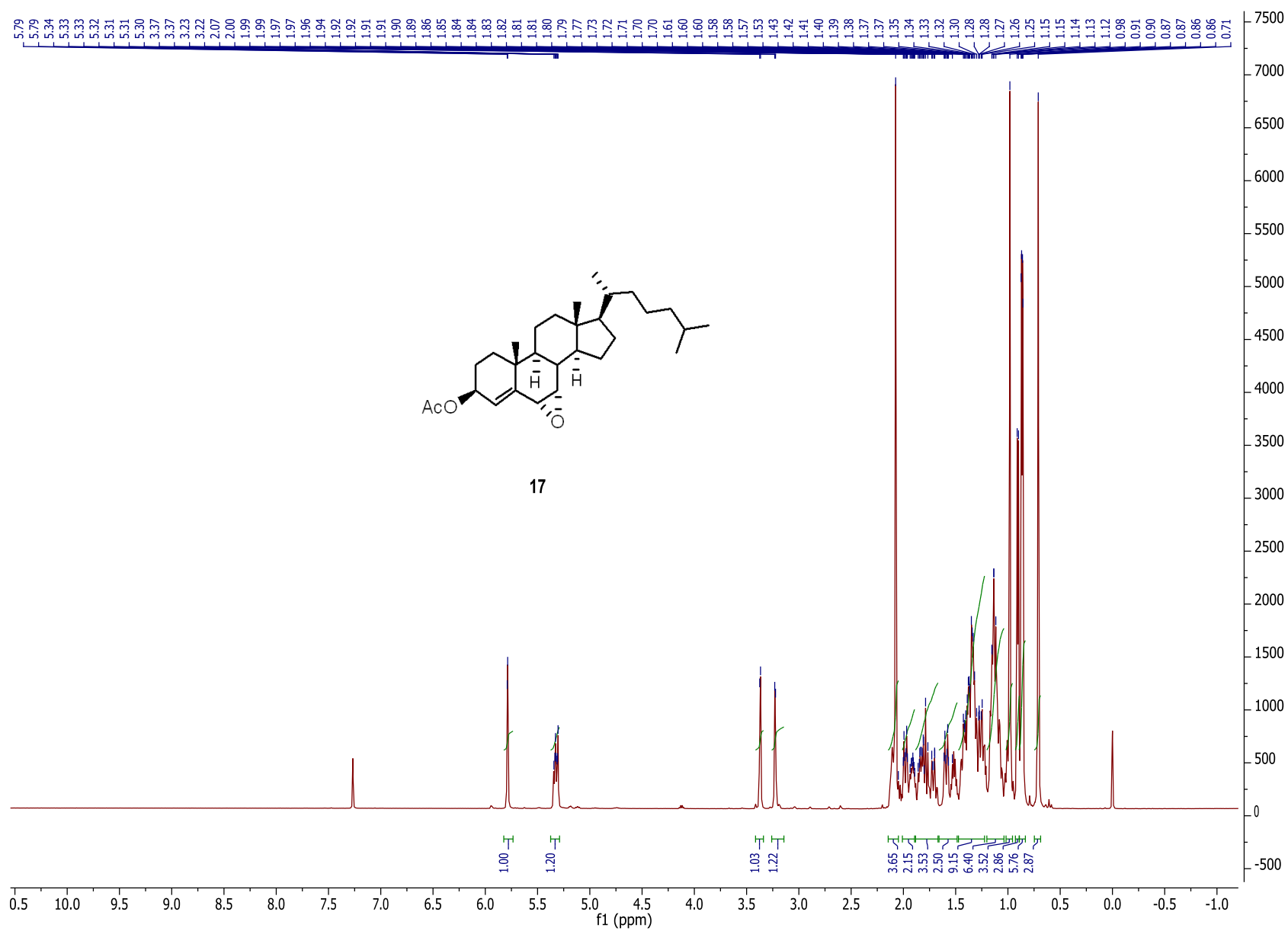


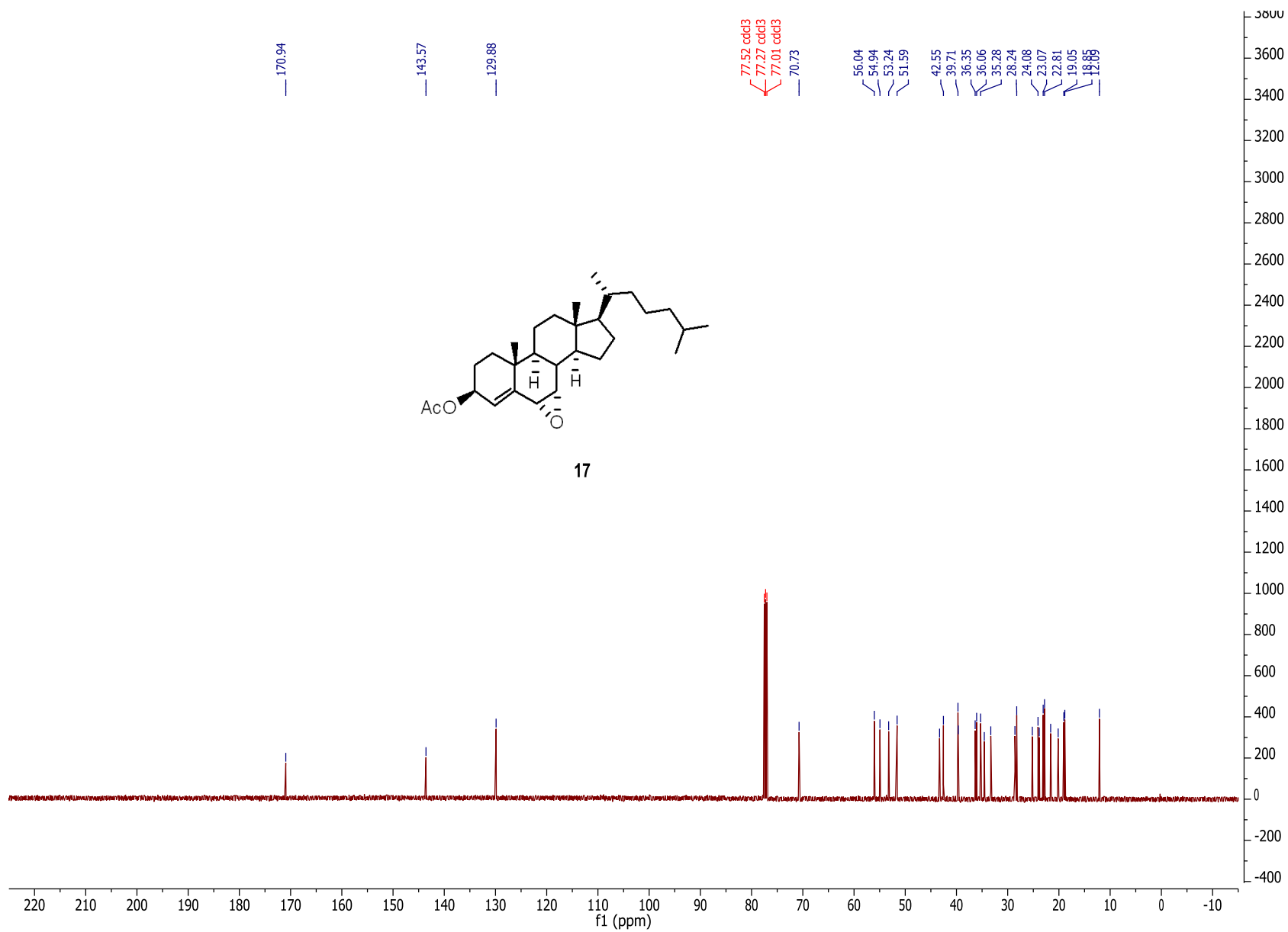




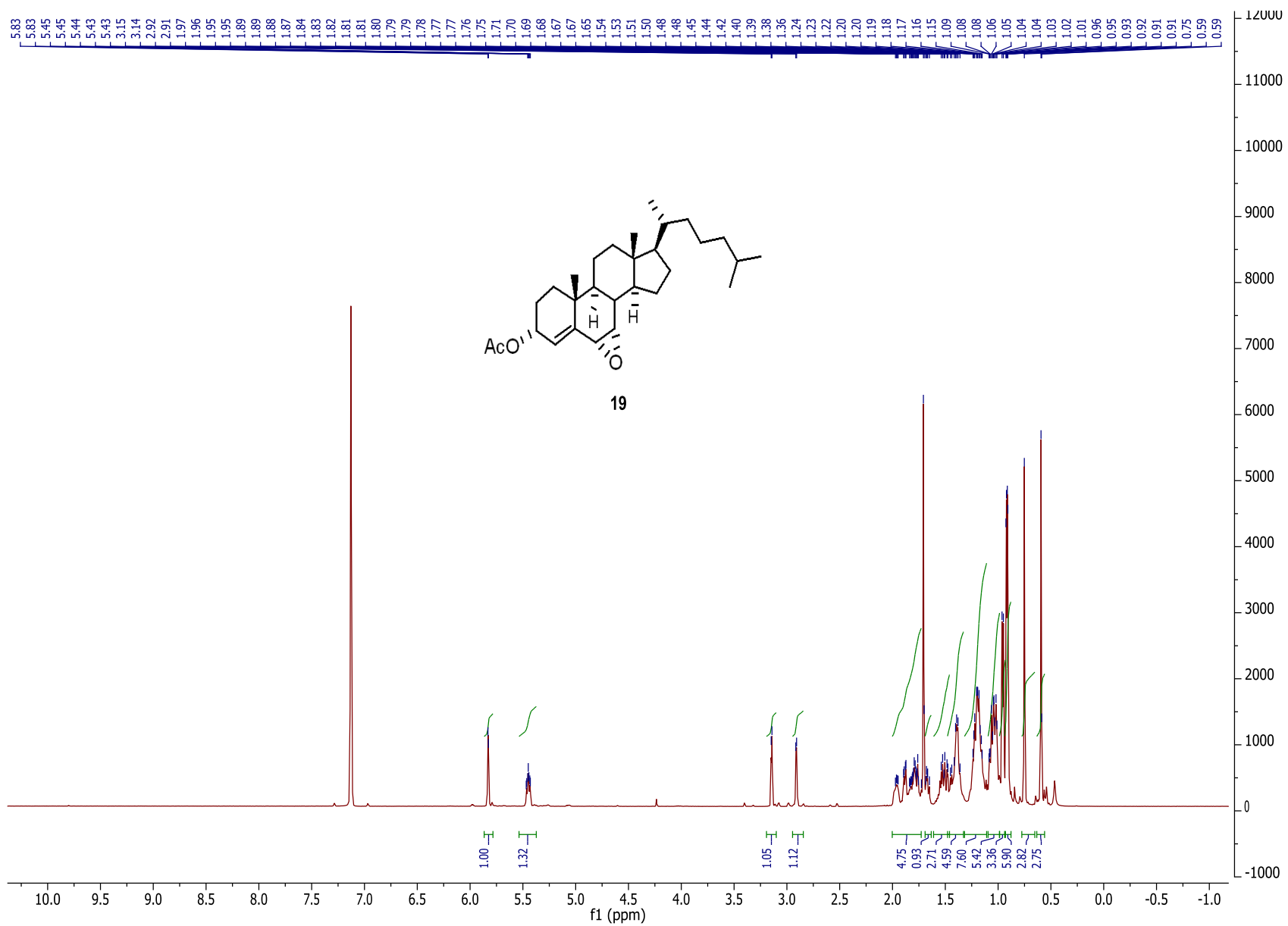


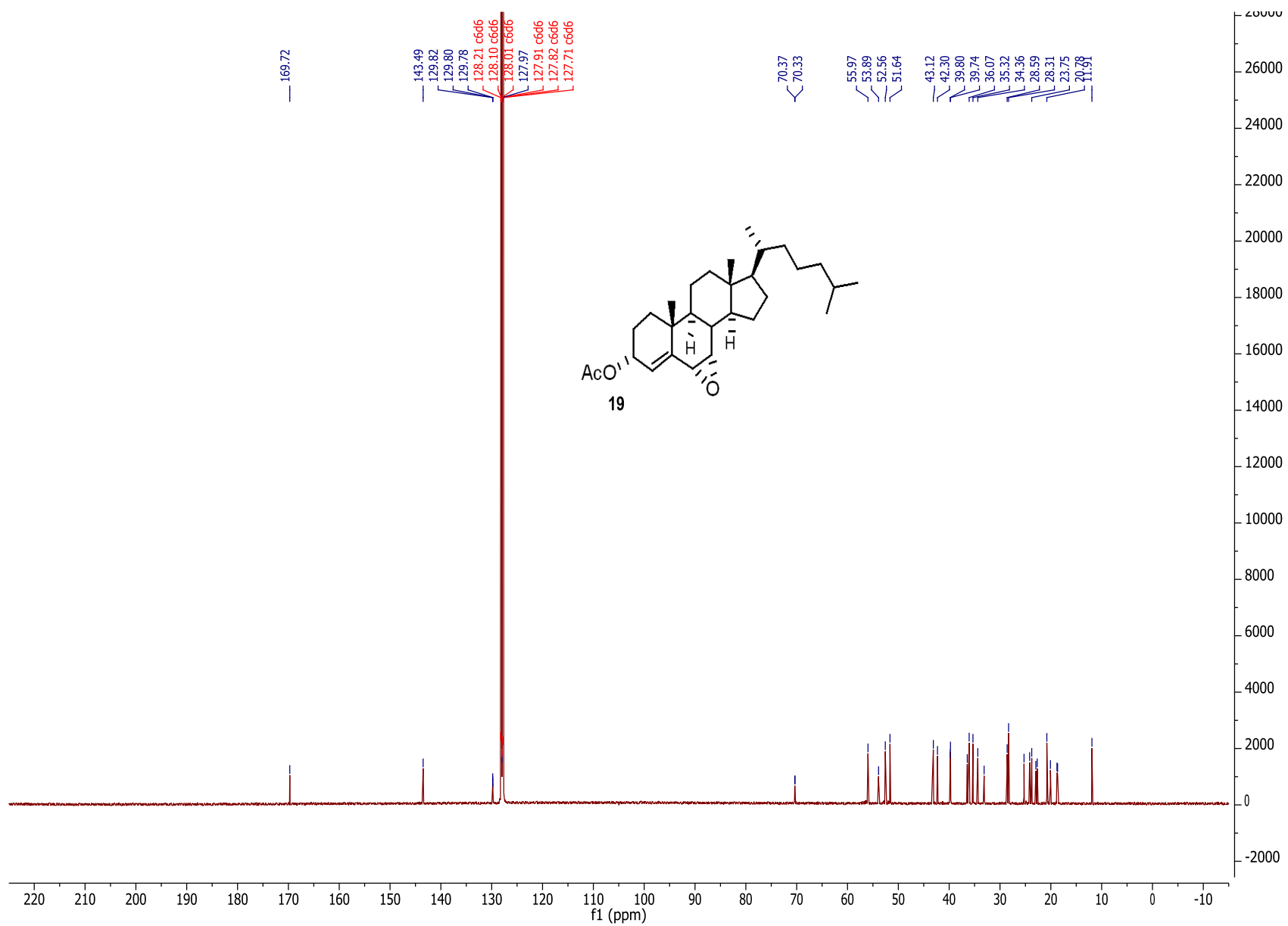


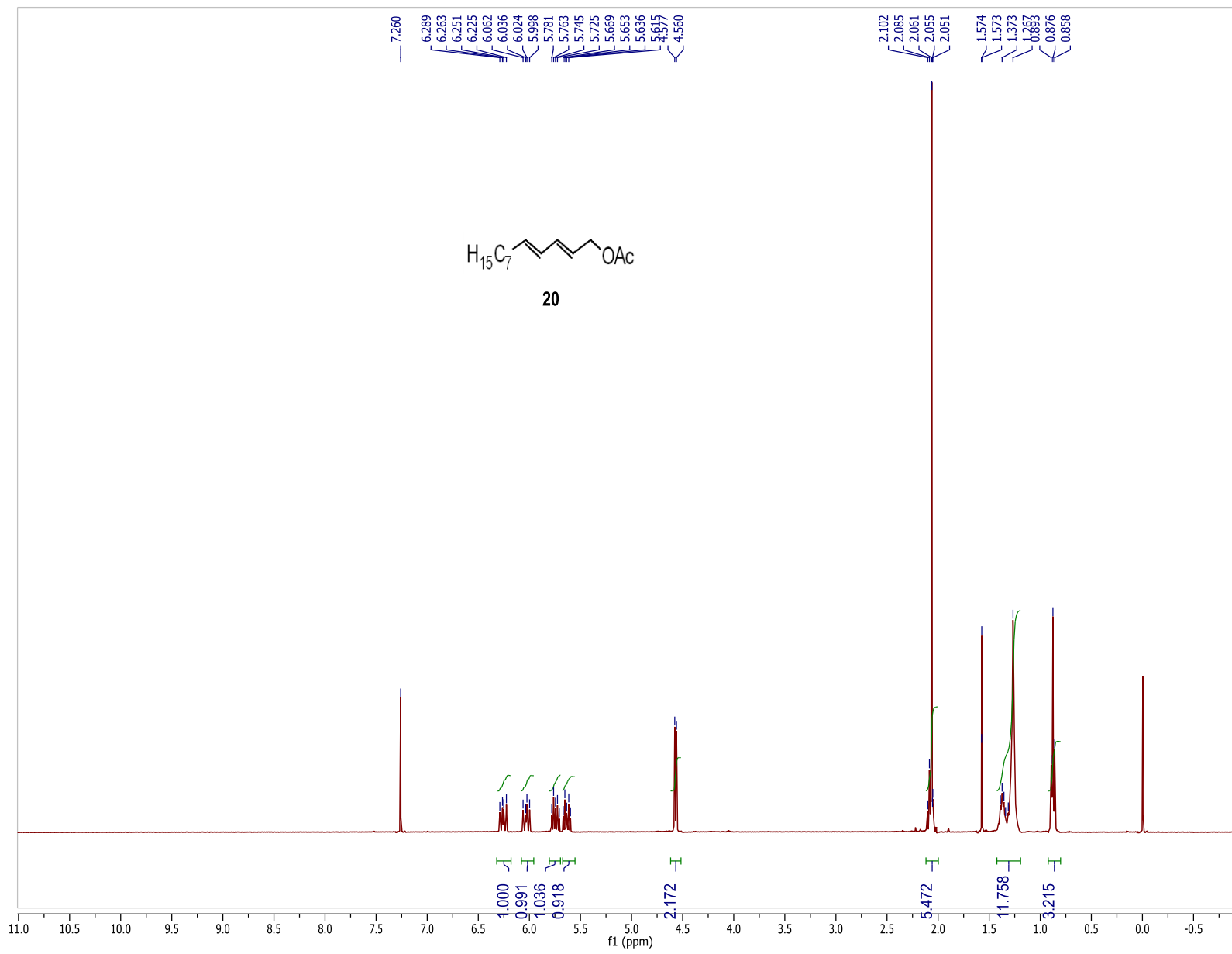








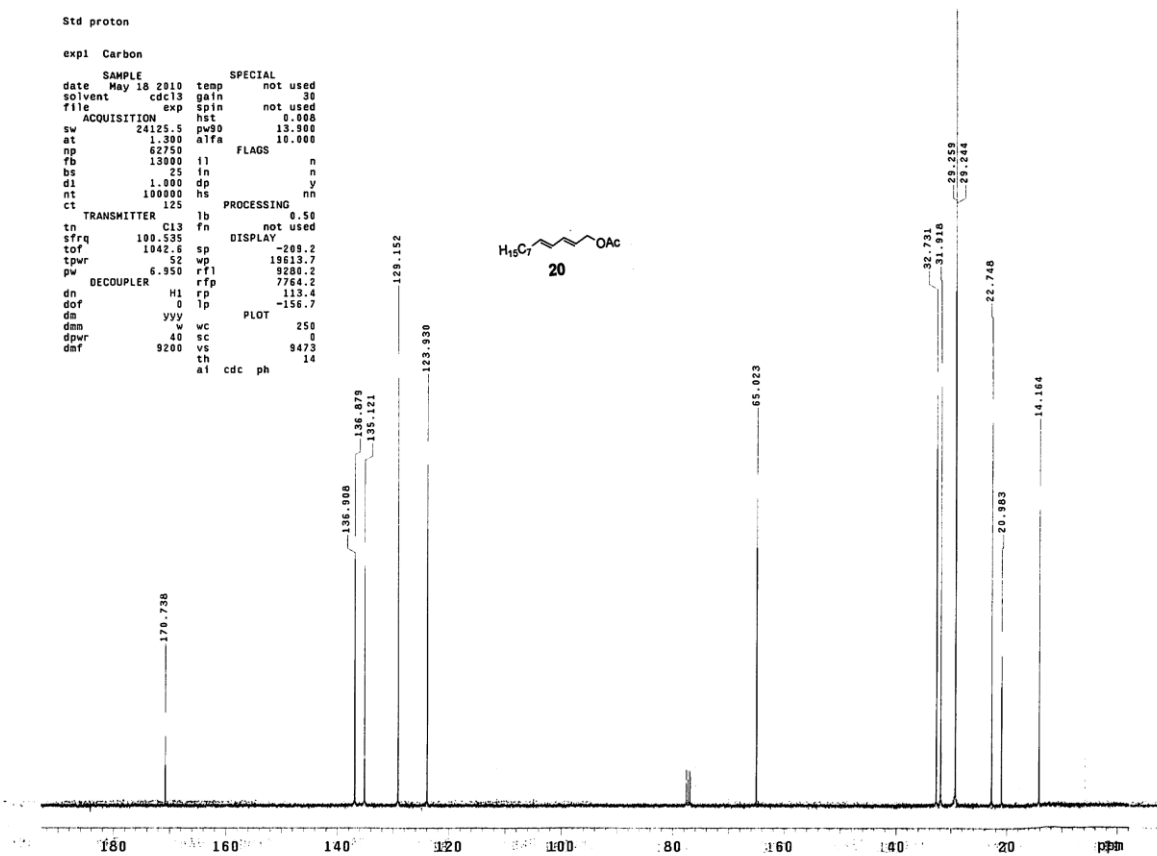
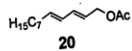


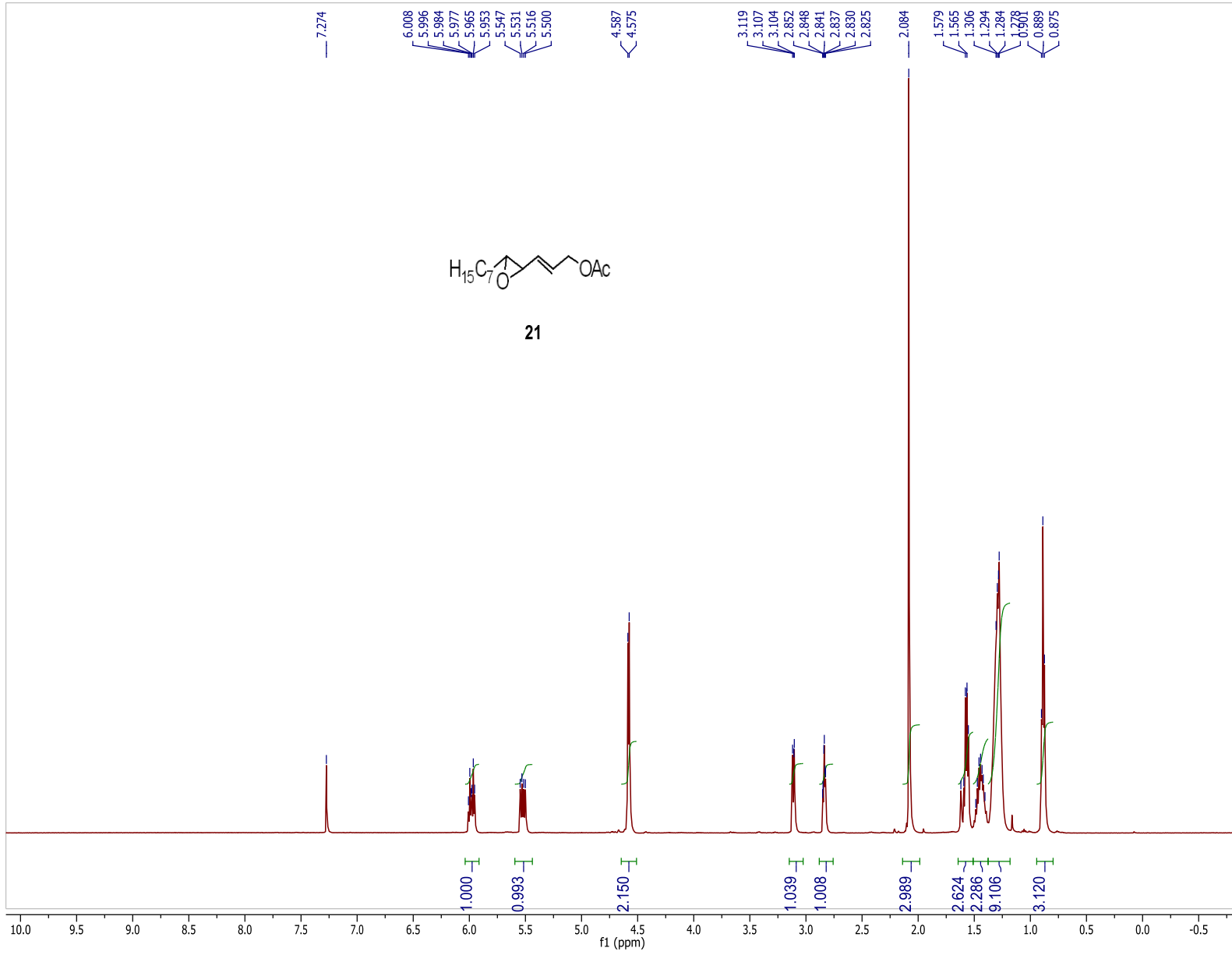


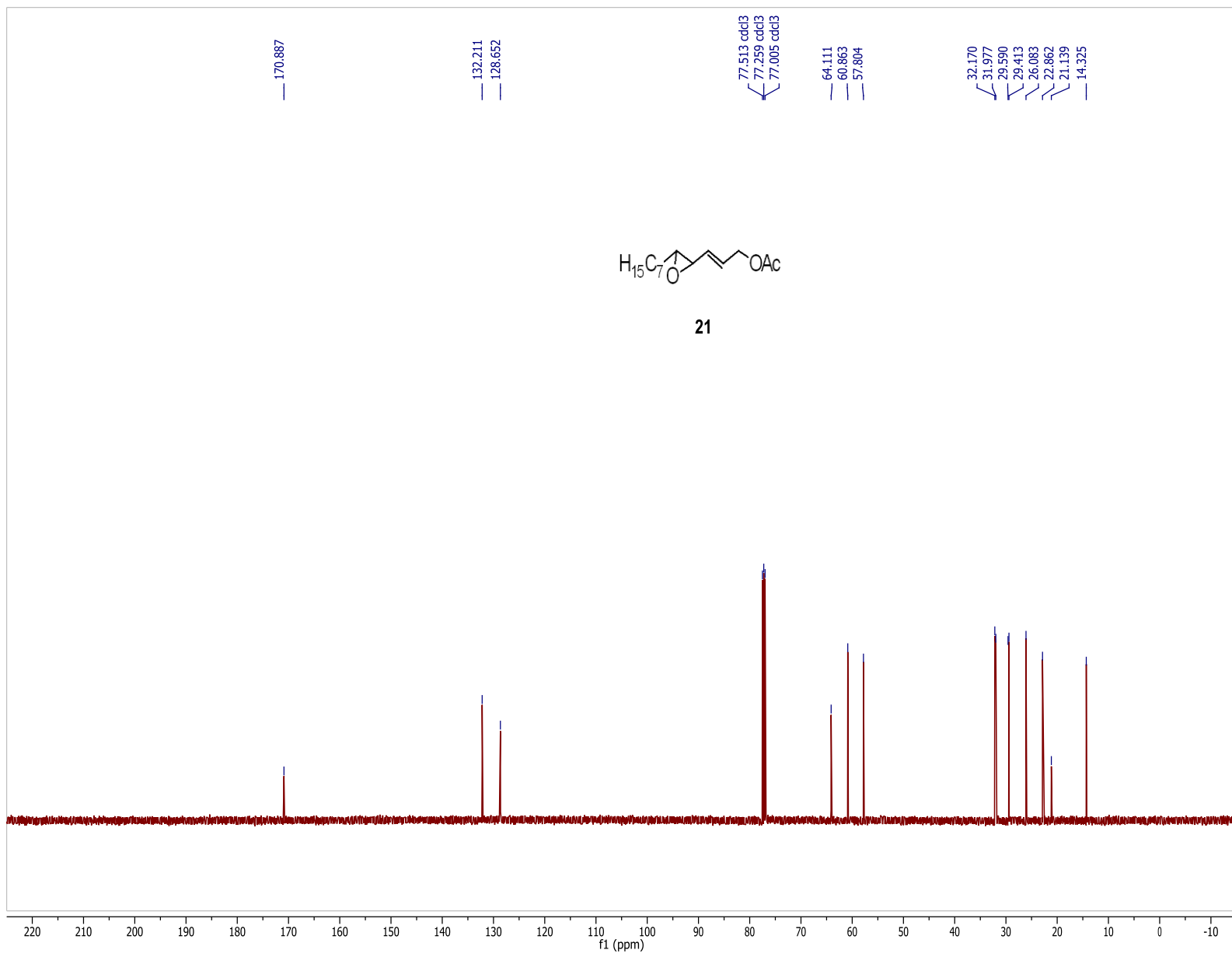
Std proton

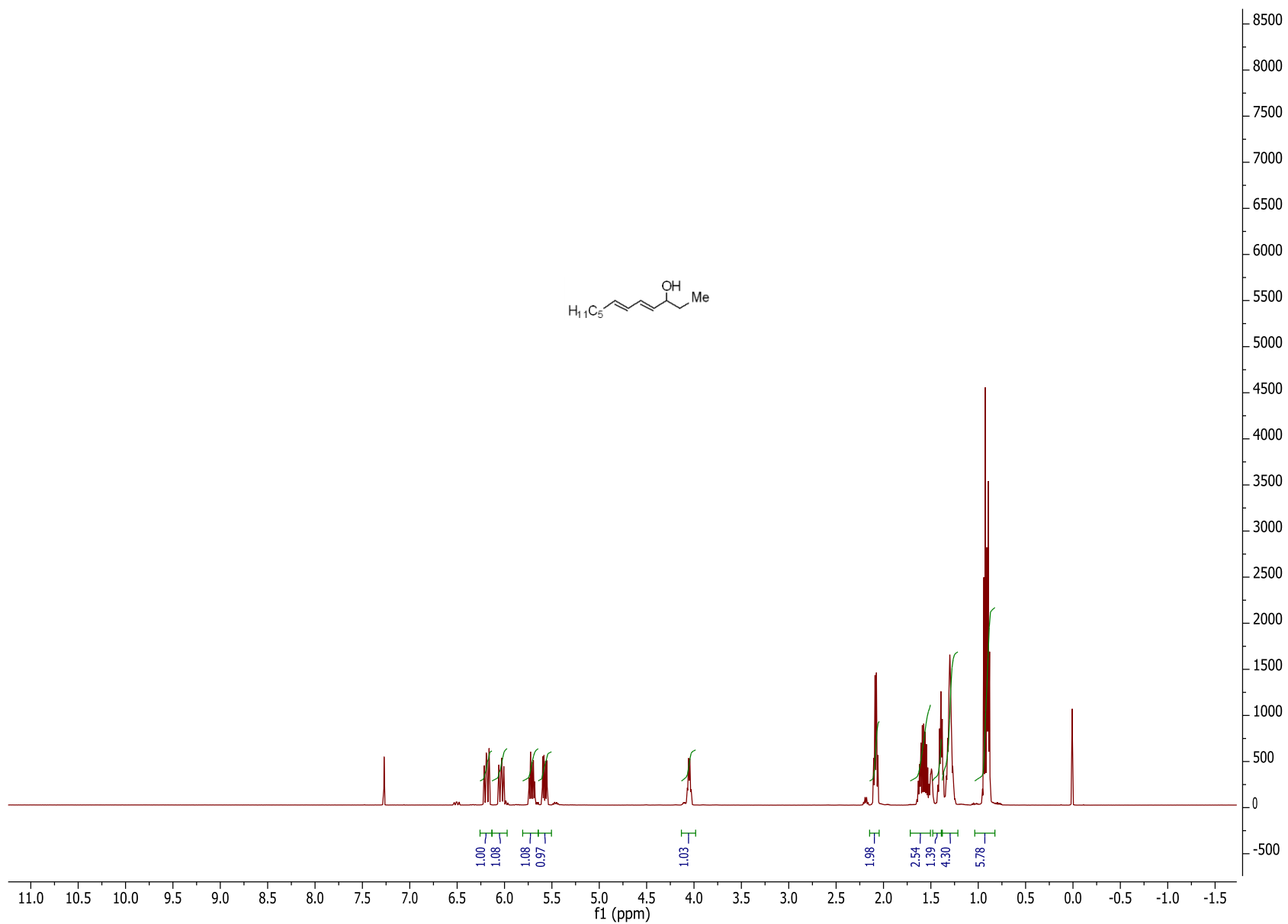
expl Carbon

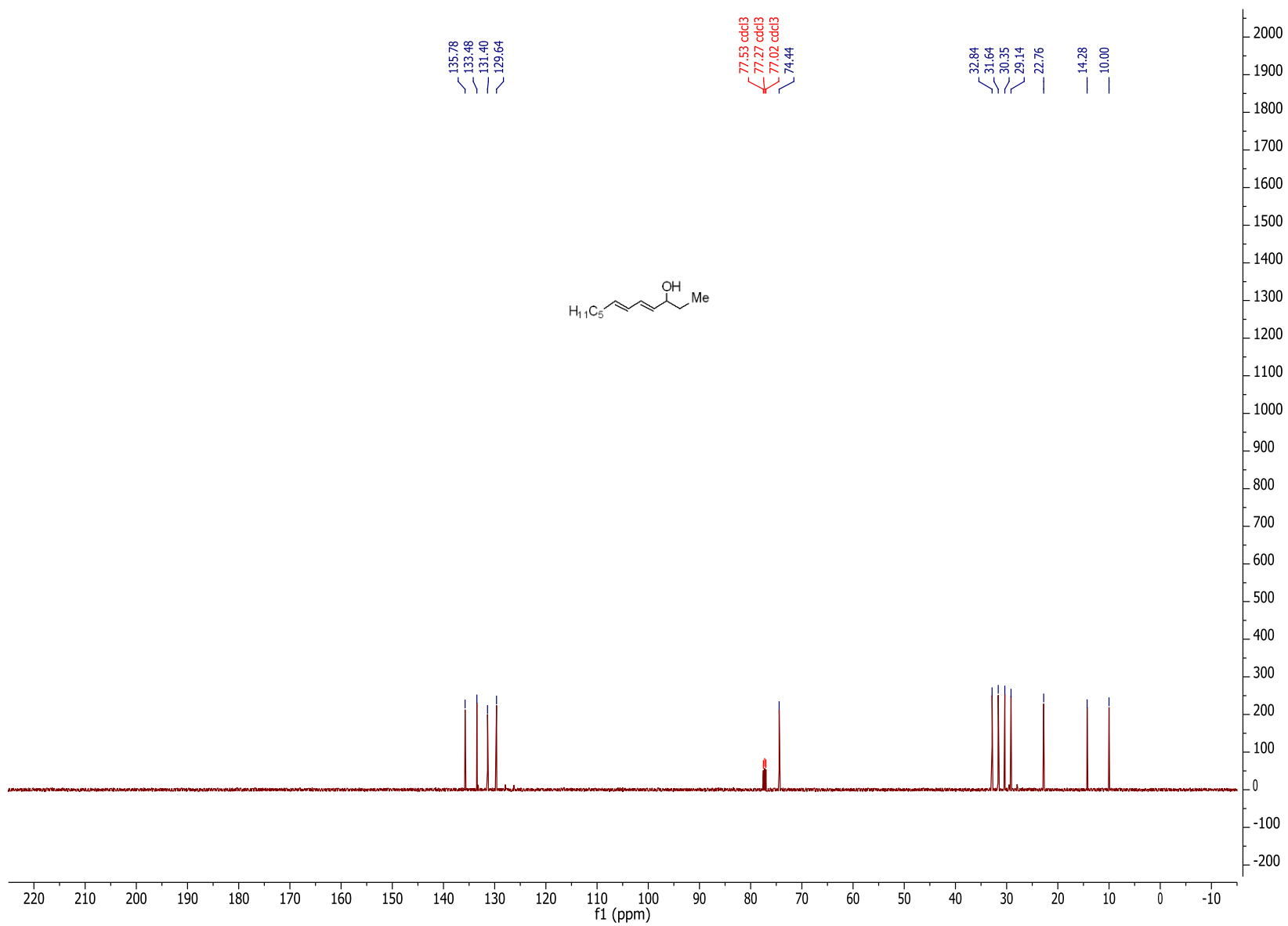
```
SAMPLE          SPECIAL
date May 18 2010 temp not used
solvent cdc13 gain 38
file exp spin not used
ACQUISITION hst 0.008
sw 24125.5 pps0 13.308
at 1.300 a1fa 10.000
np 62750
fb 13000 fl FLAGS
bs 25 fn n
dl 1.000 dp y
nt 100000 hs nn
ct 125 PROCESSING
TRANSMITTER lb 0.50
tn c13 fn not used
sfrq 100.535 DISPLAY
tof 1042.6 sp -209.2
tpwr 52 wp 19813.7
pw 6.350 rfl 3288.2
DECOUPLER rfp 7764.2
dn H1 rp 113.4
dof 0 lp -156.7
dm yyy PLOT
dmm w wc 250
dpmr 40 sc e
dmf 9200 vs 9473
a1 cdc ph 14
```



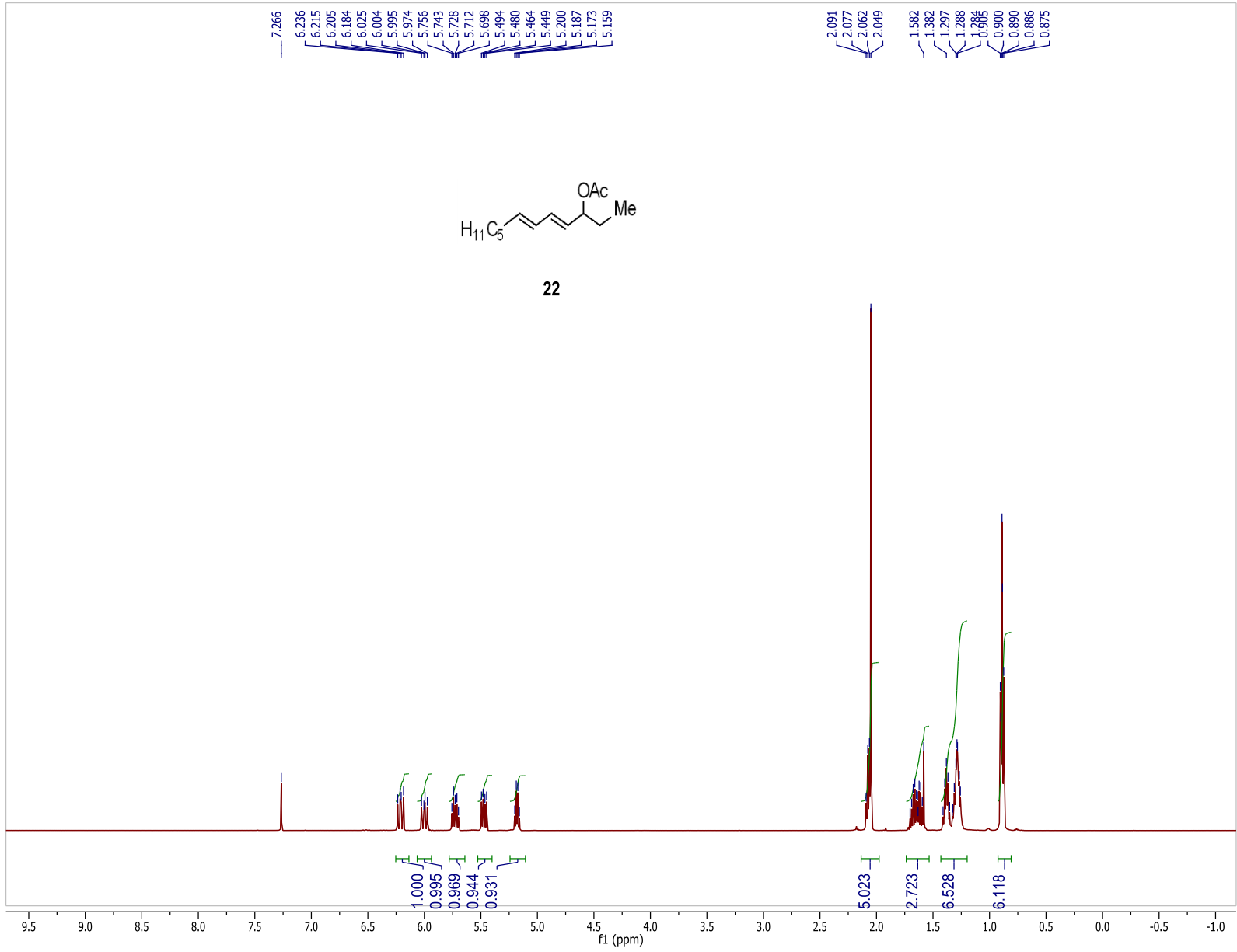


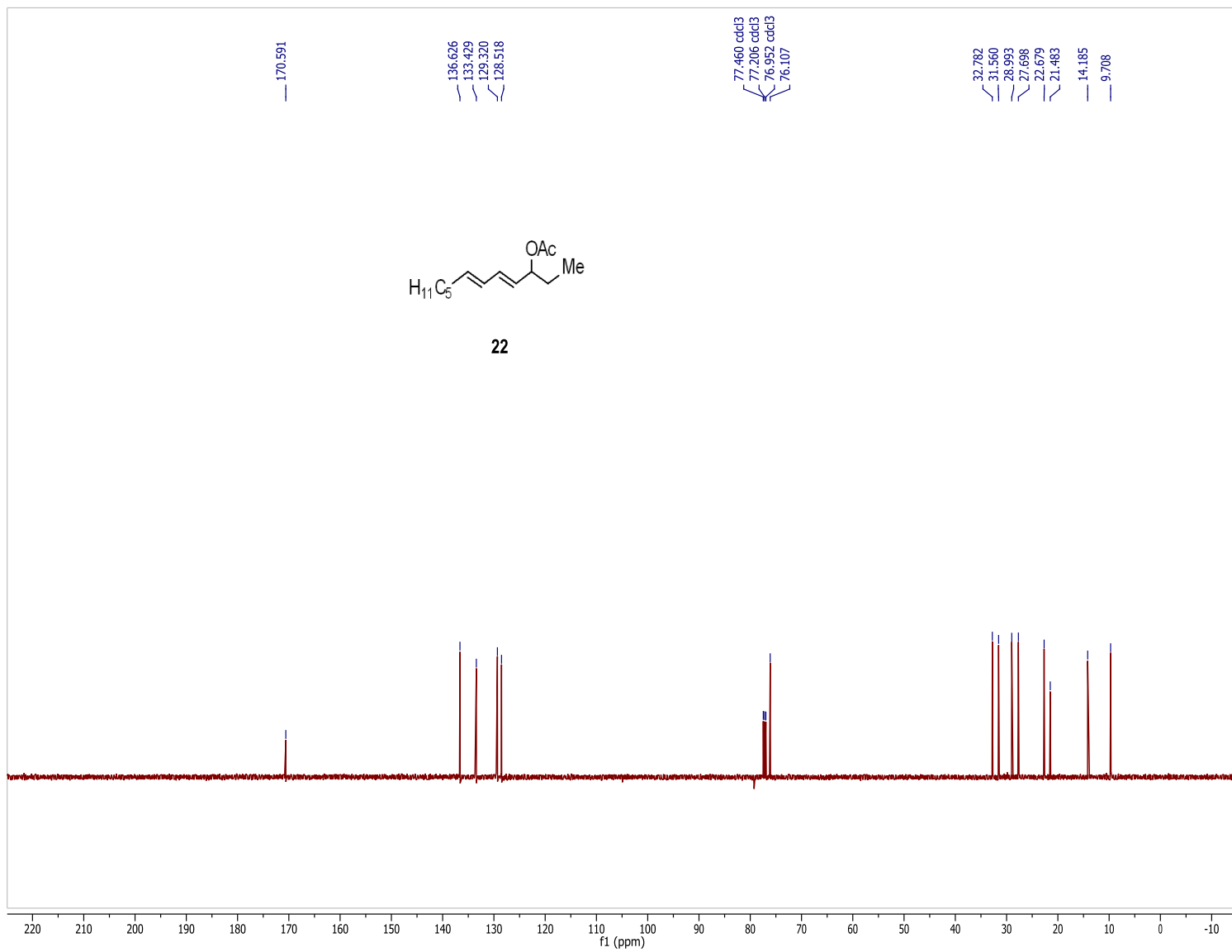


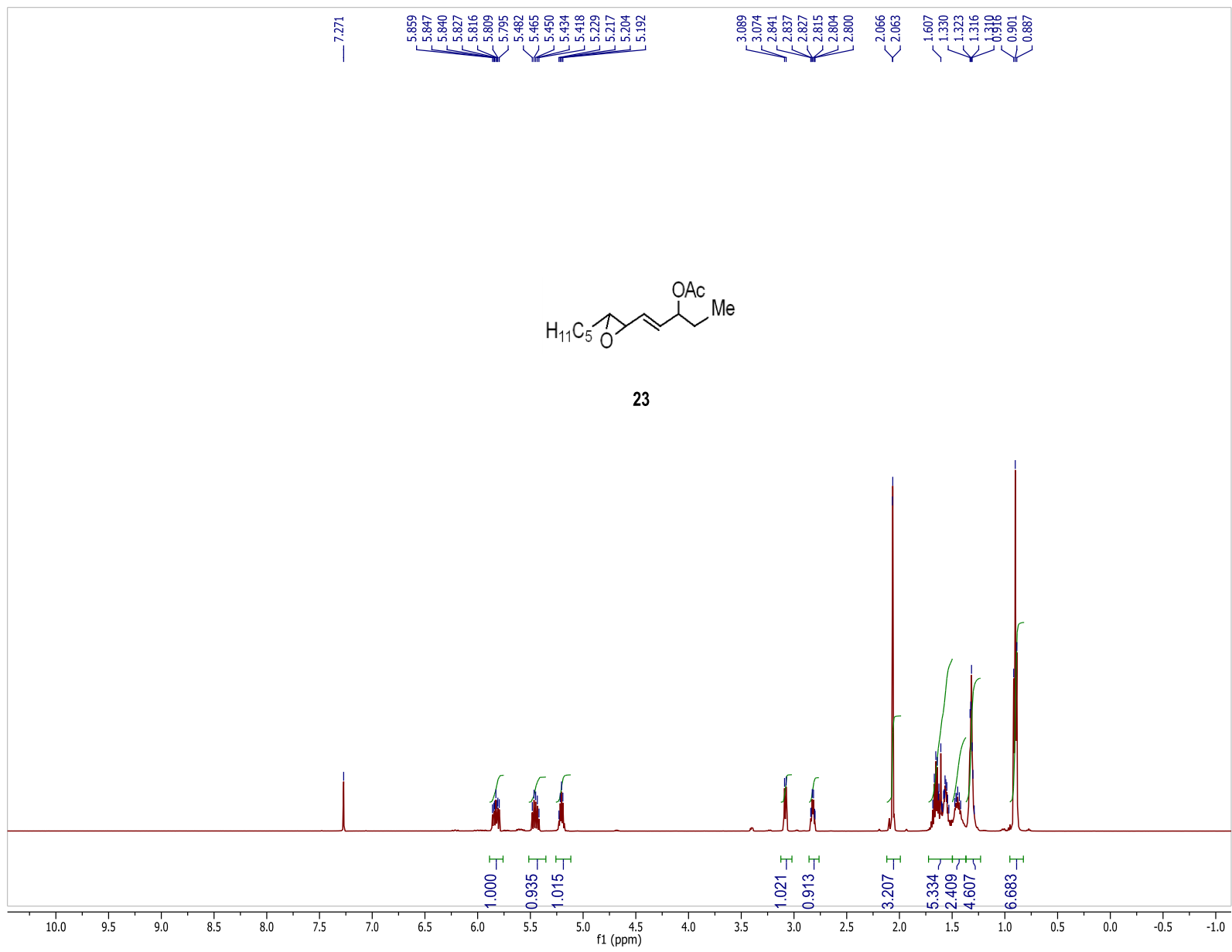


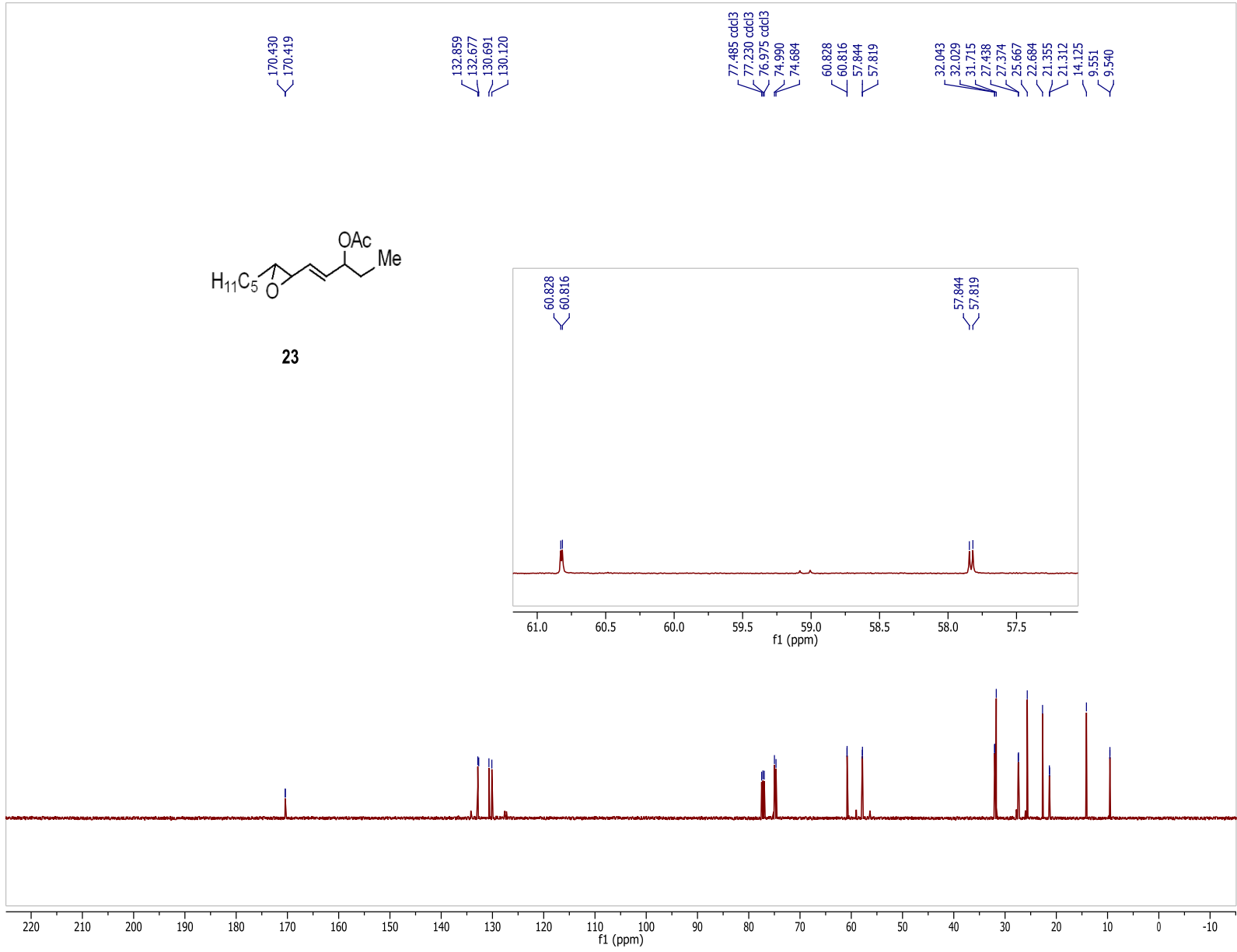


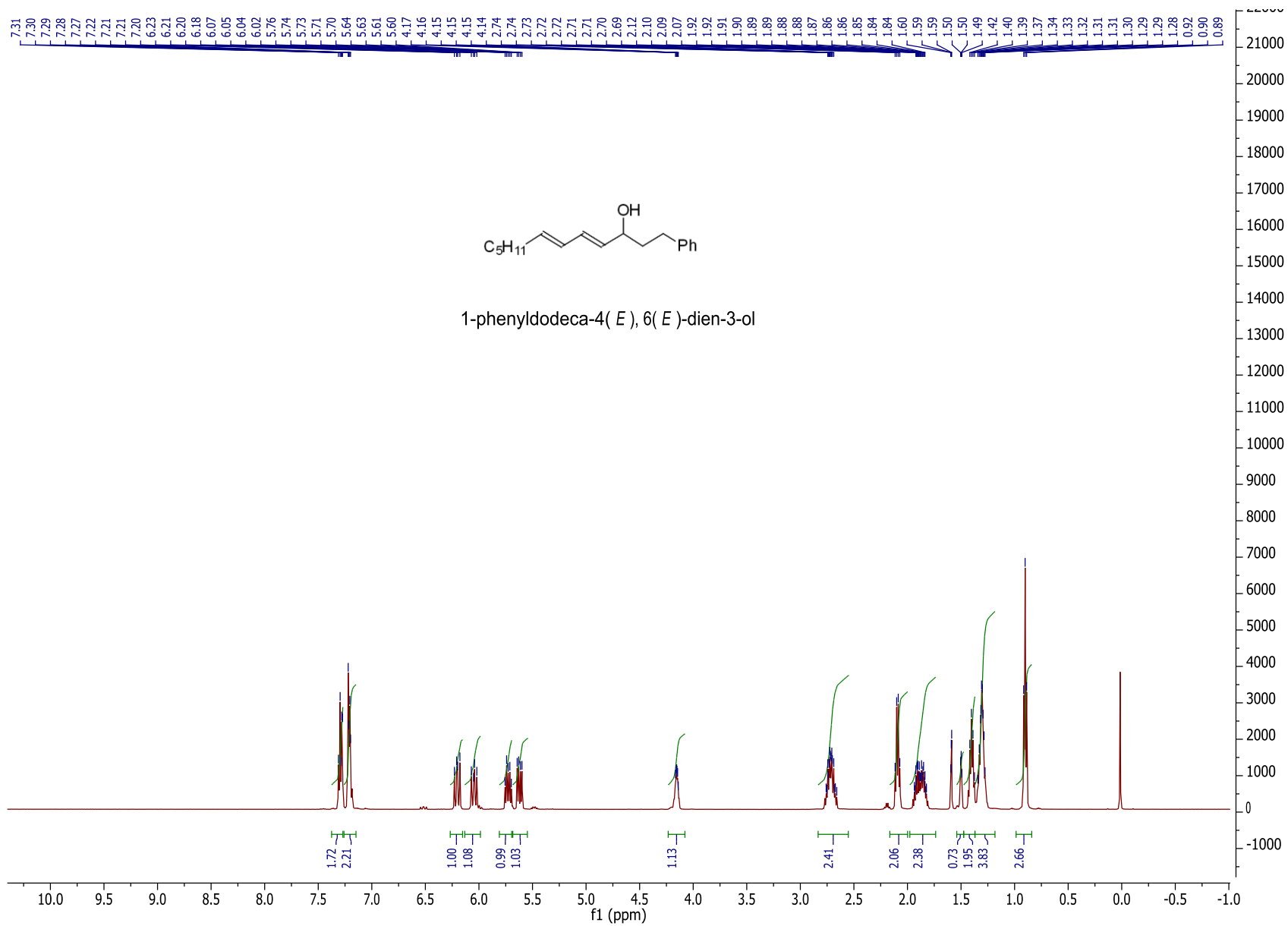


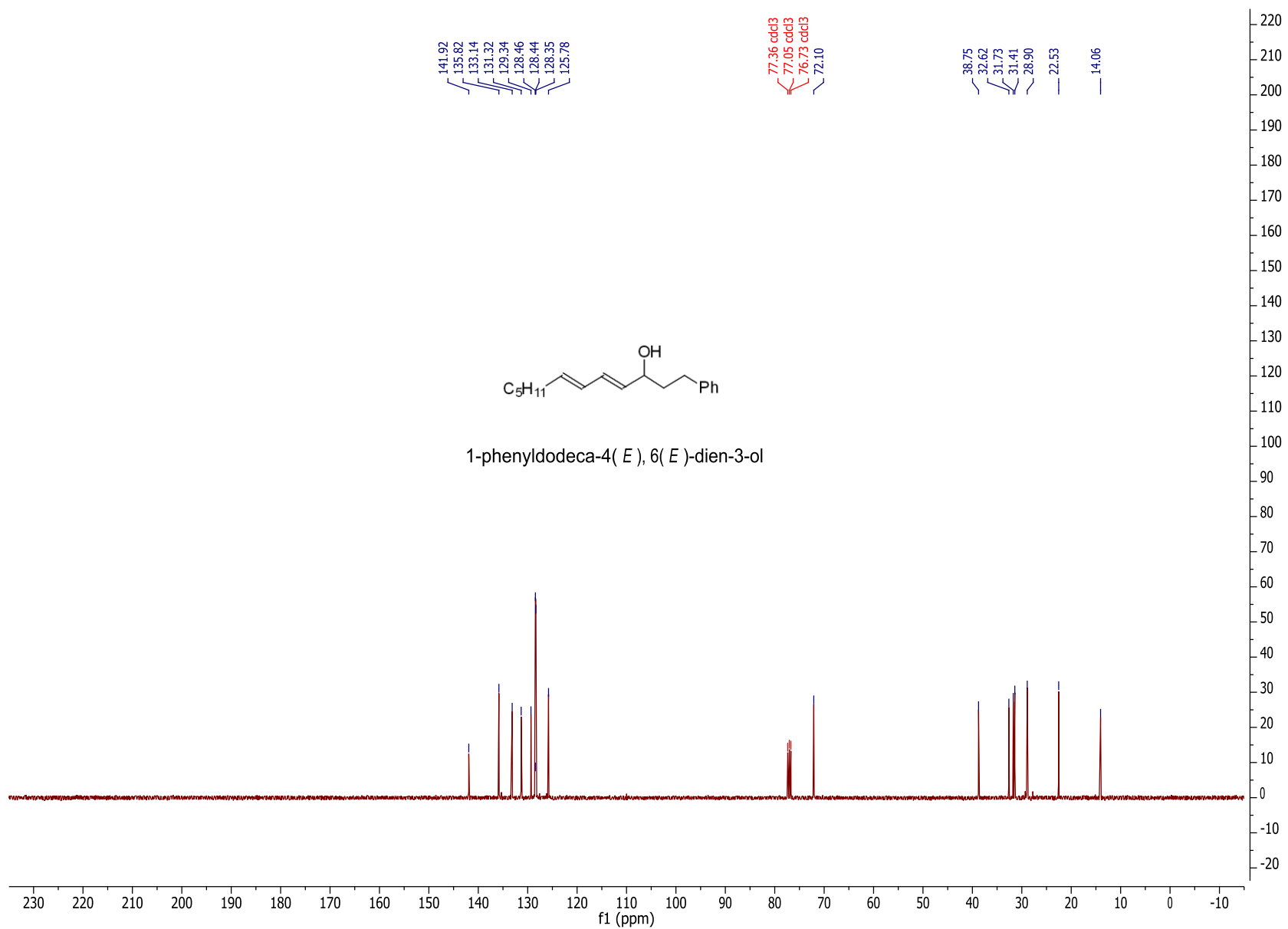


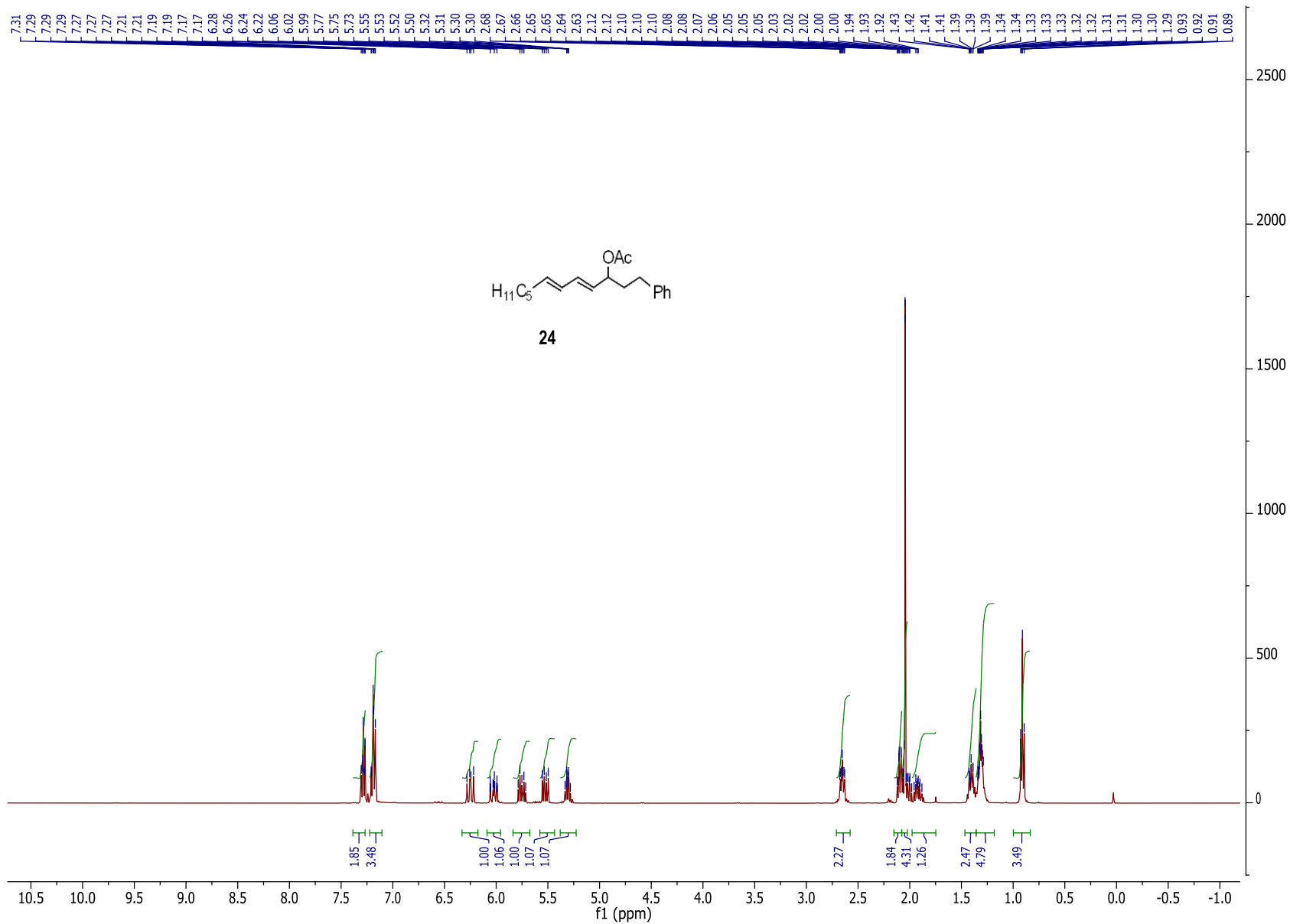


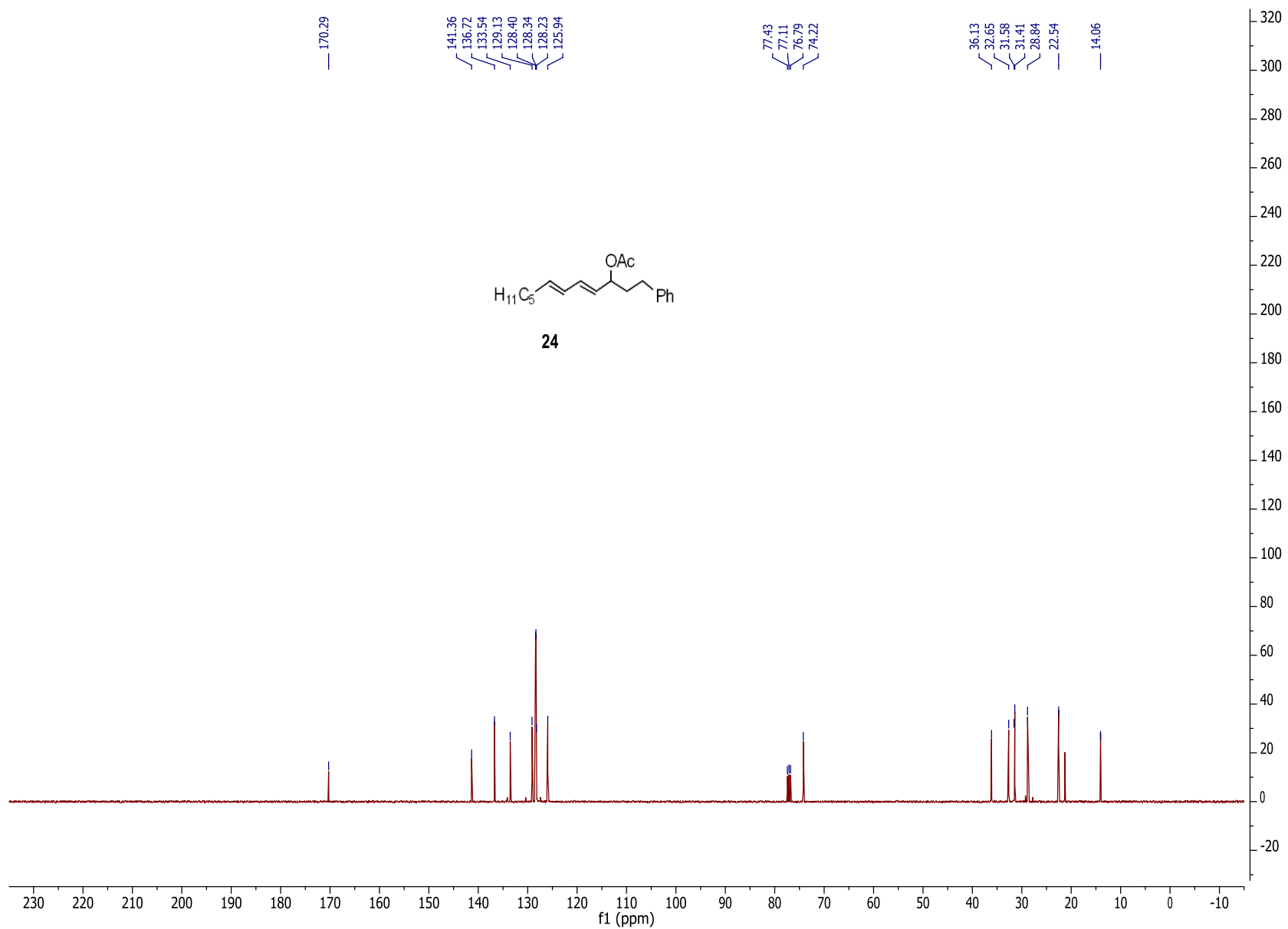




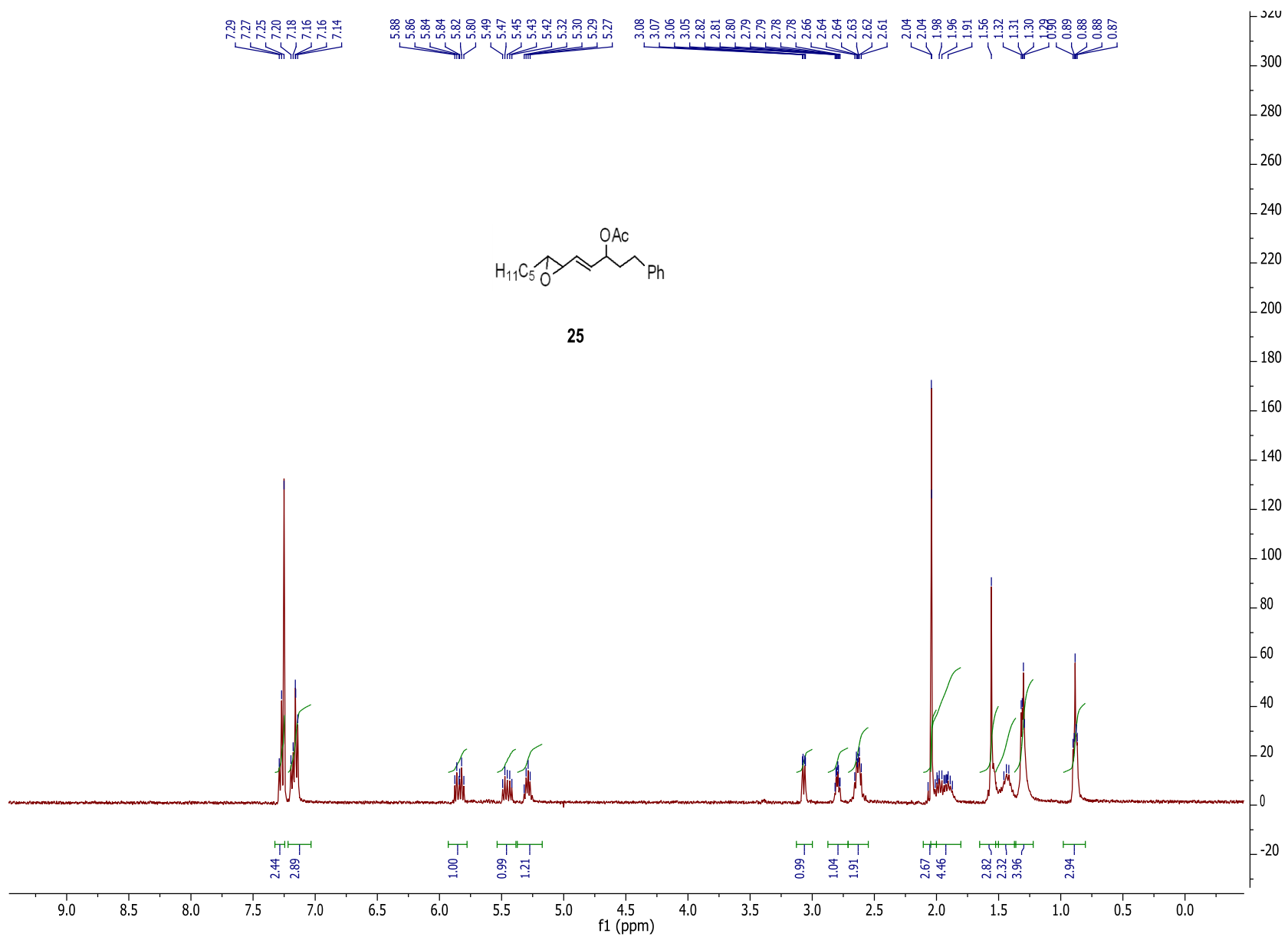


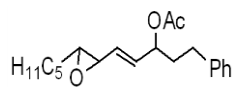




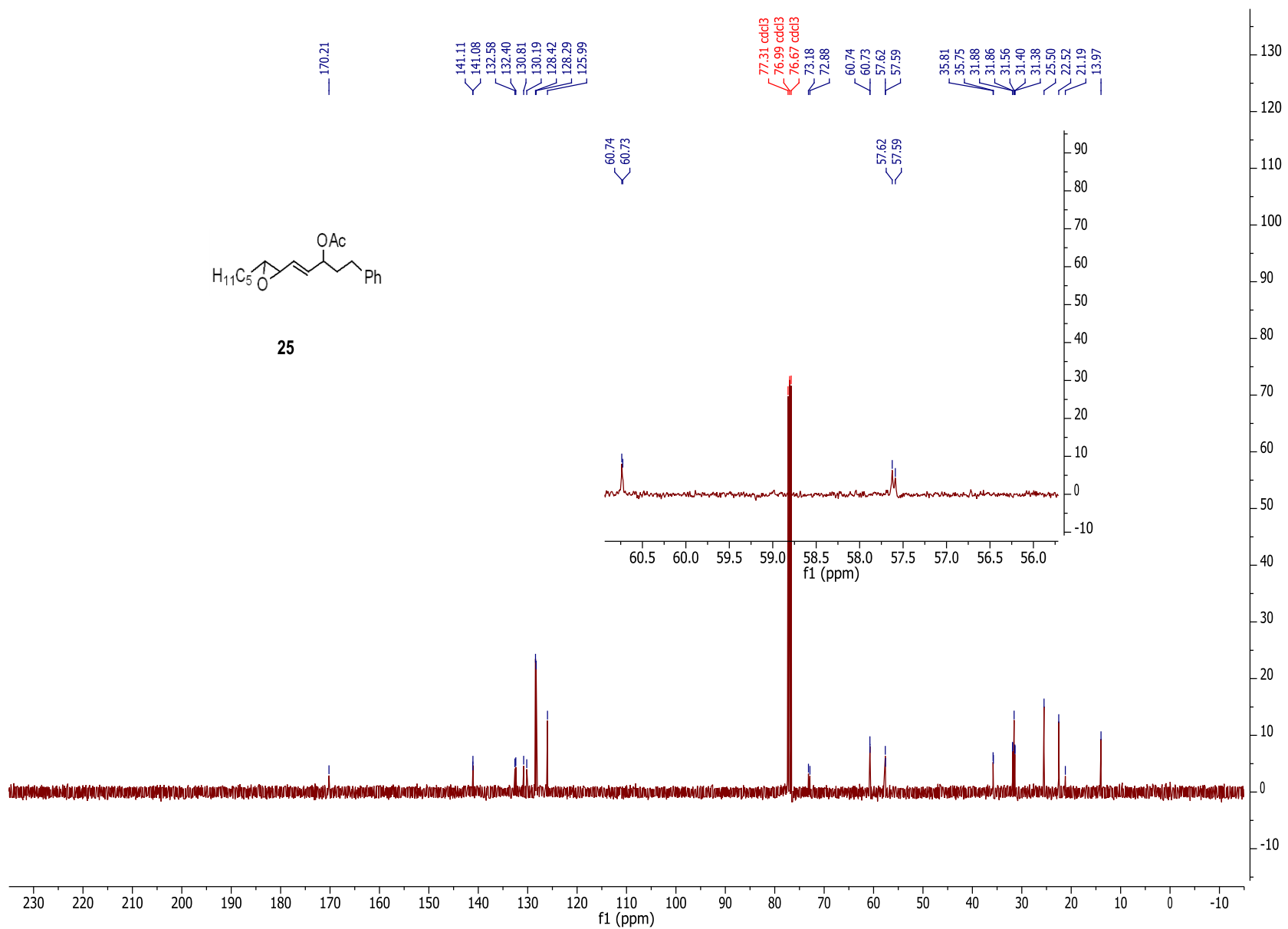




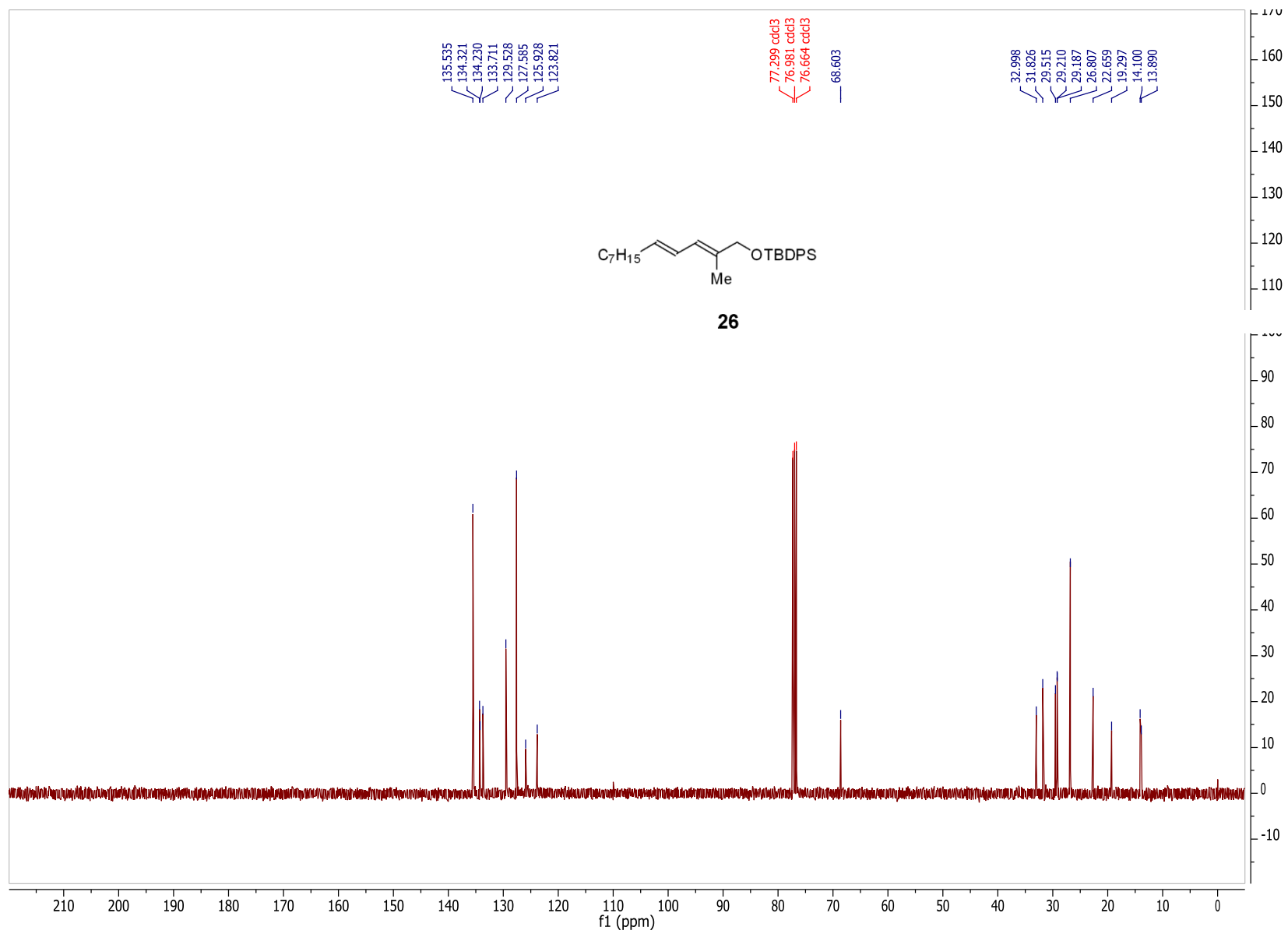




25

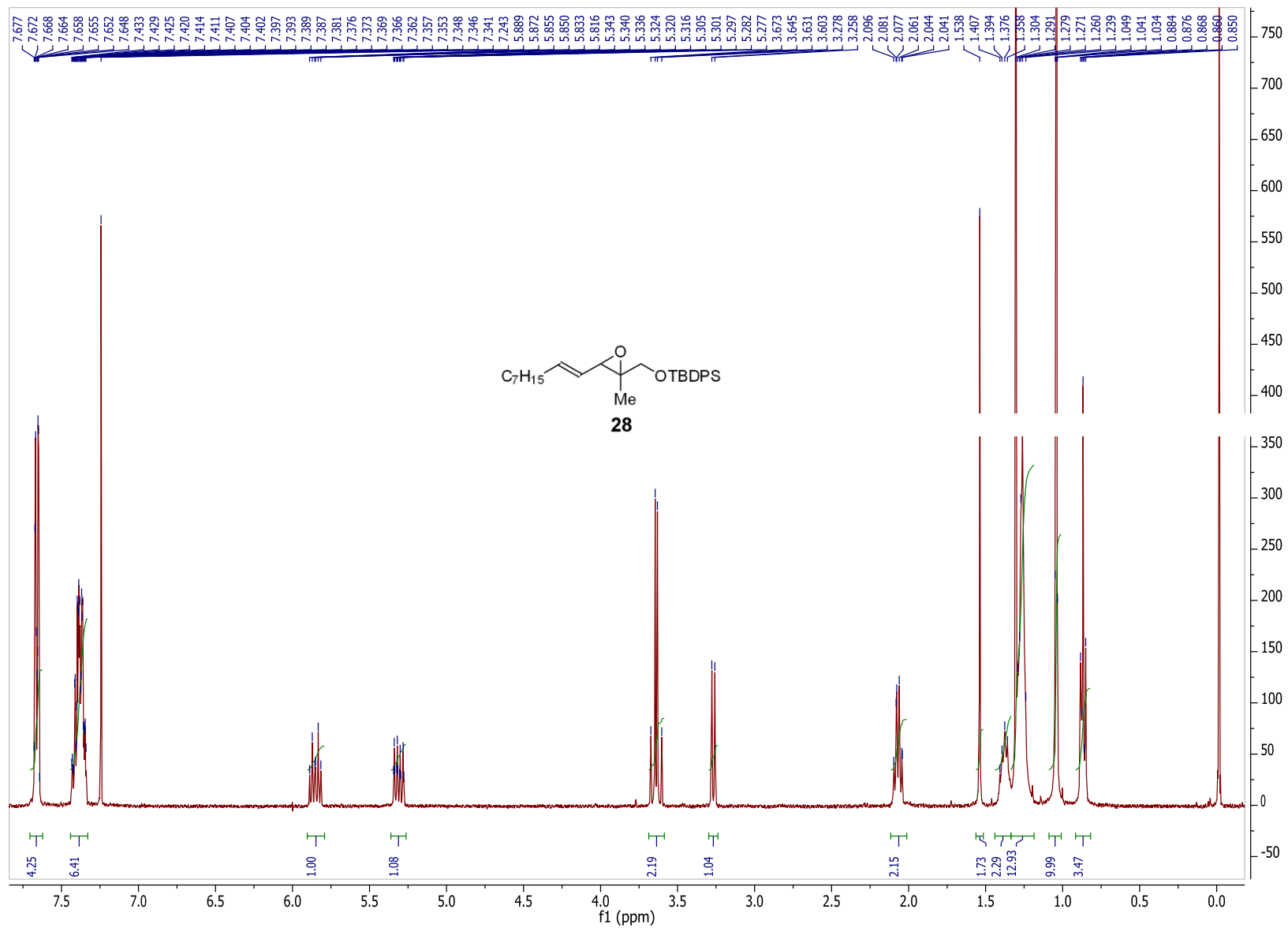


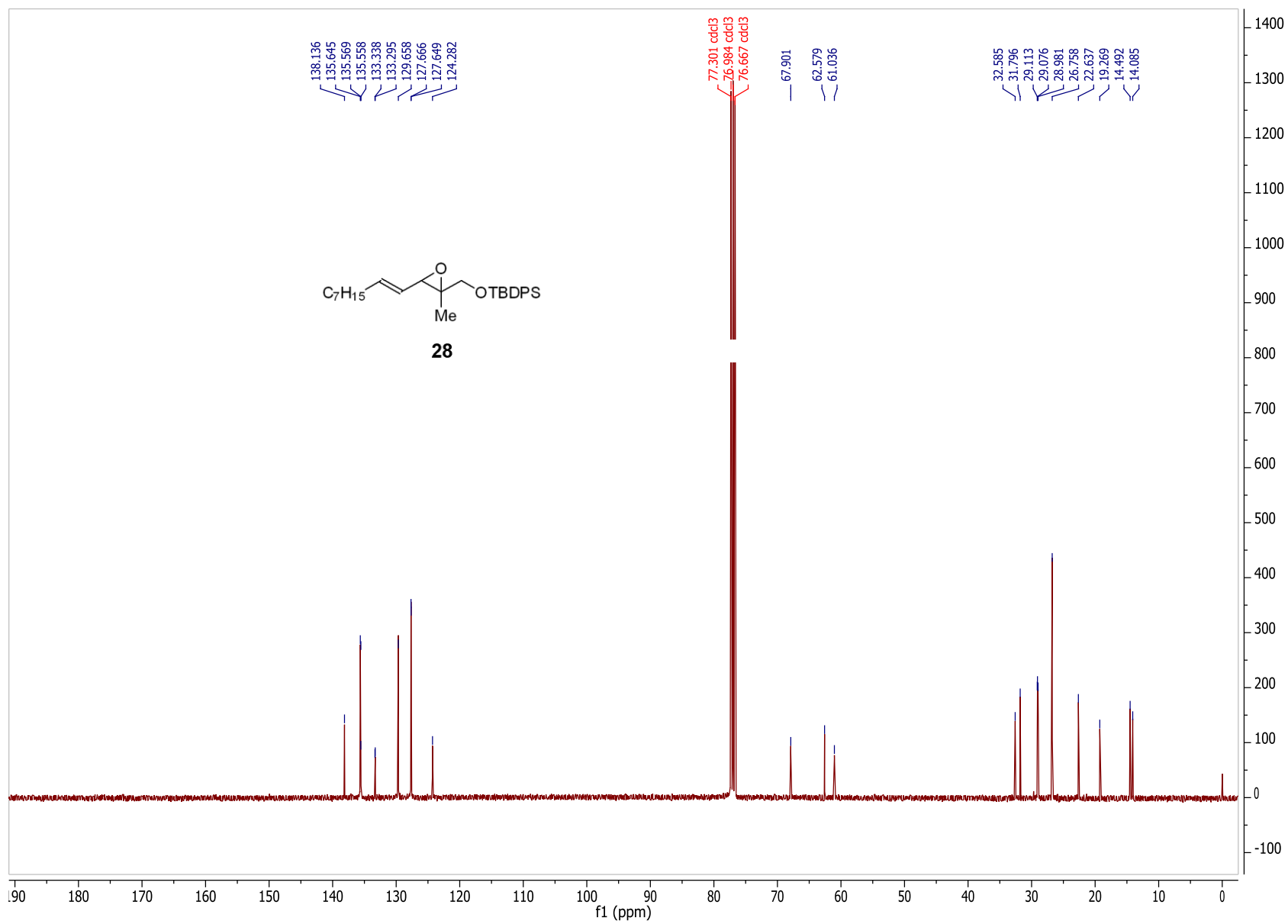




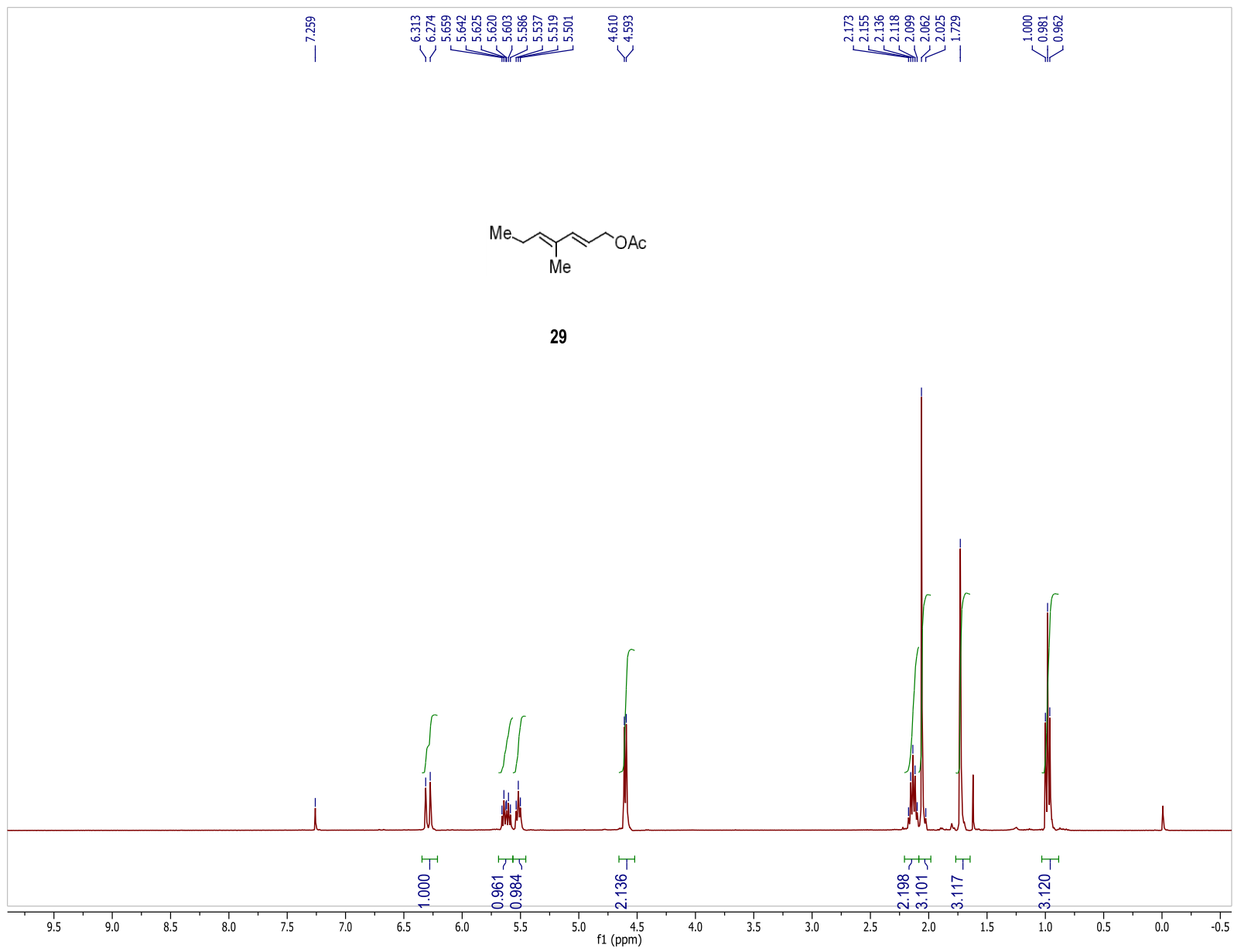


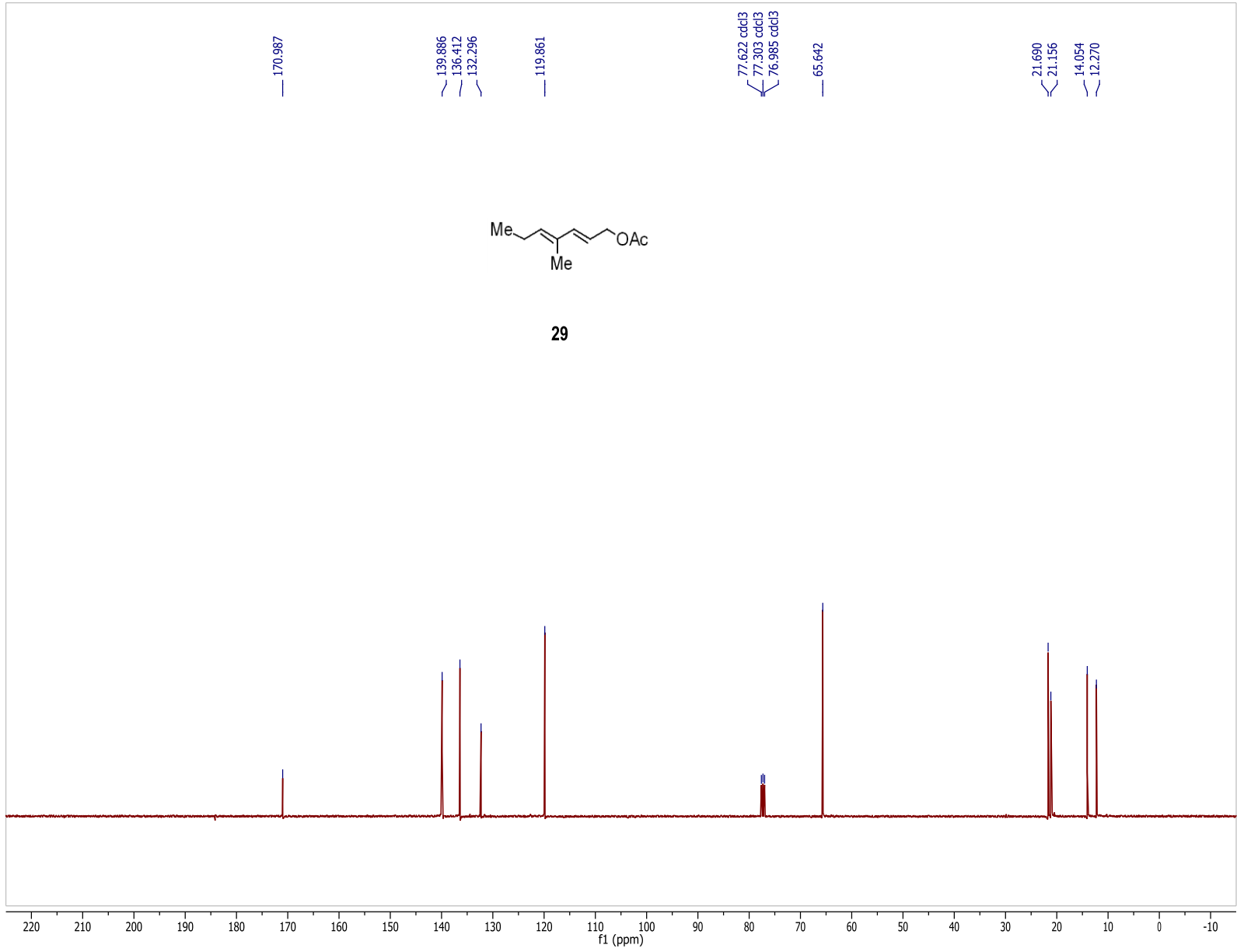


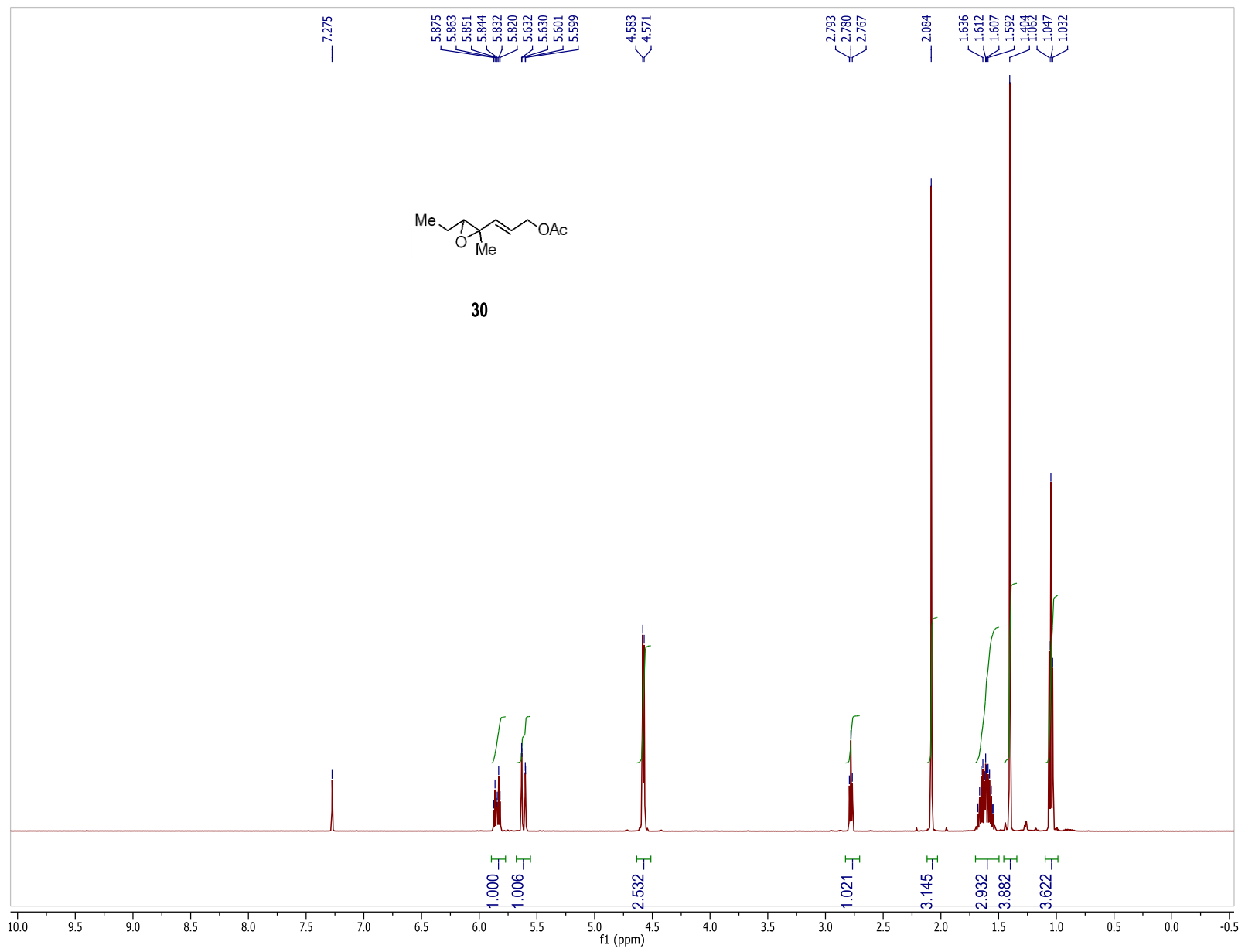


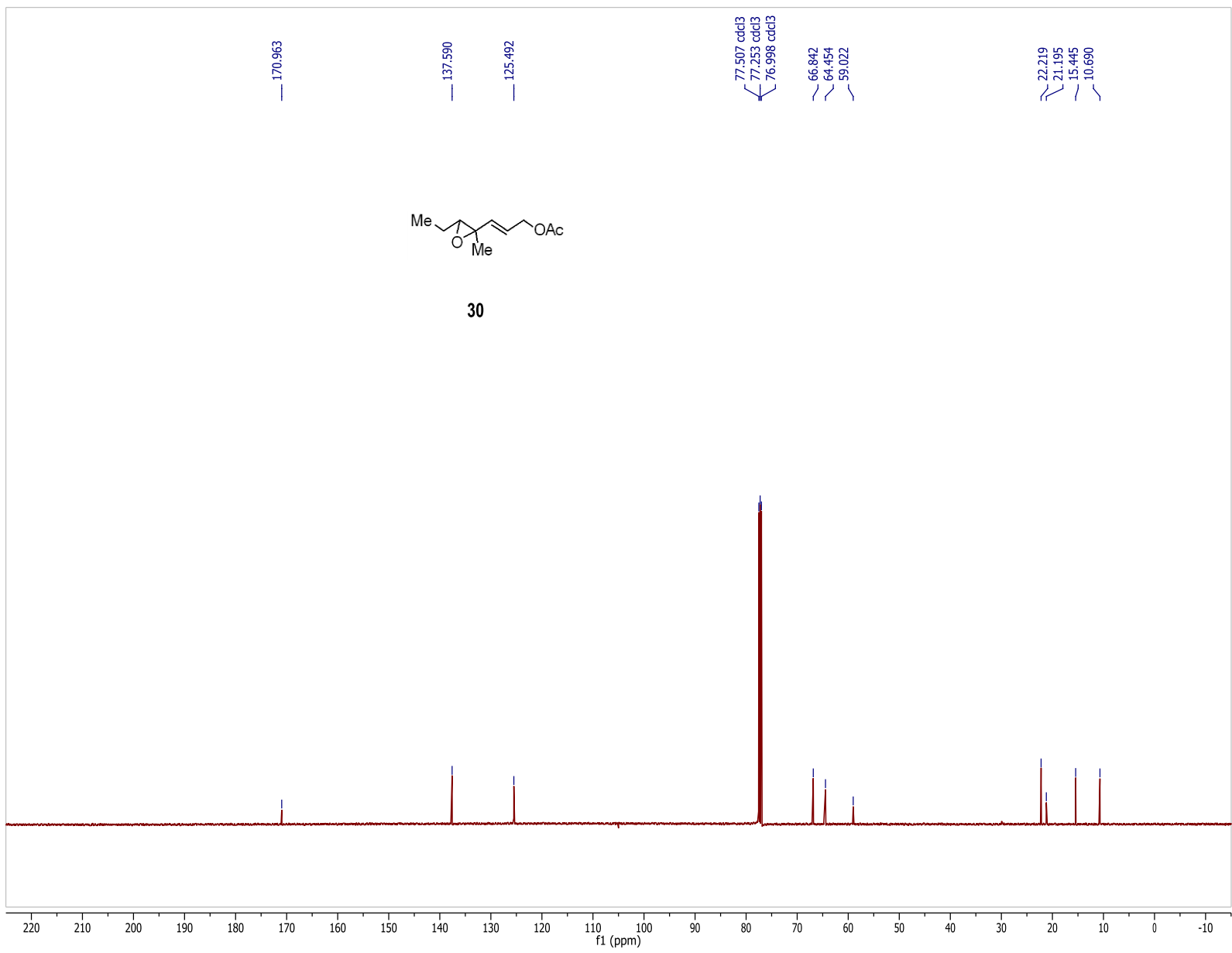


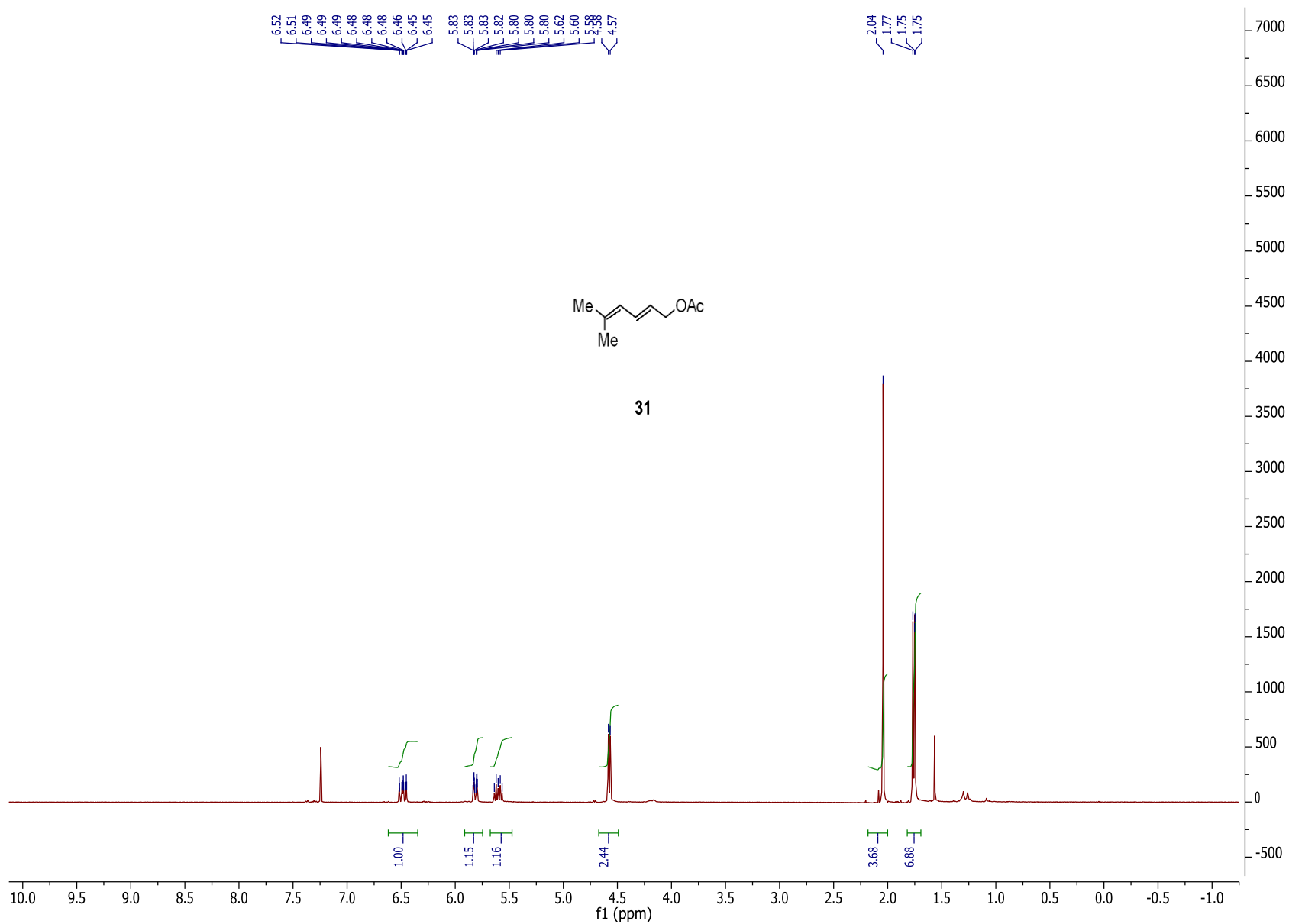


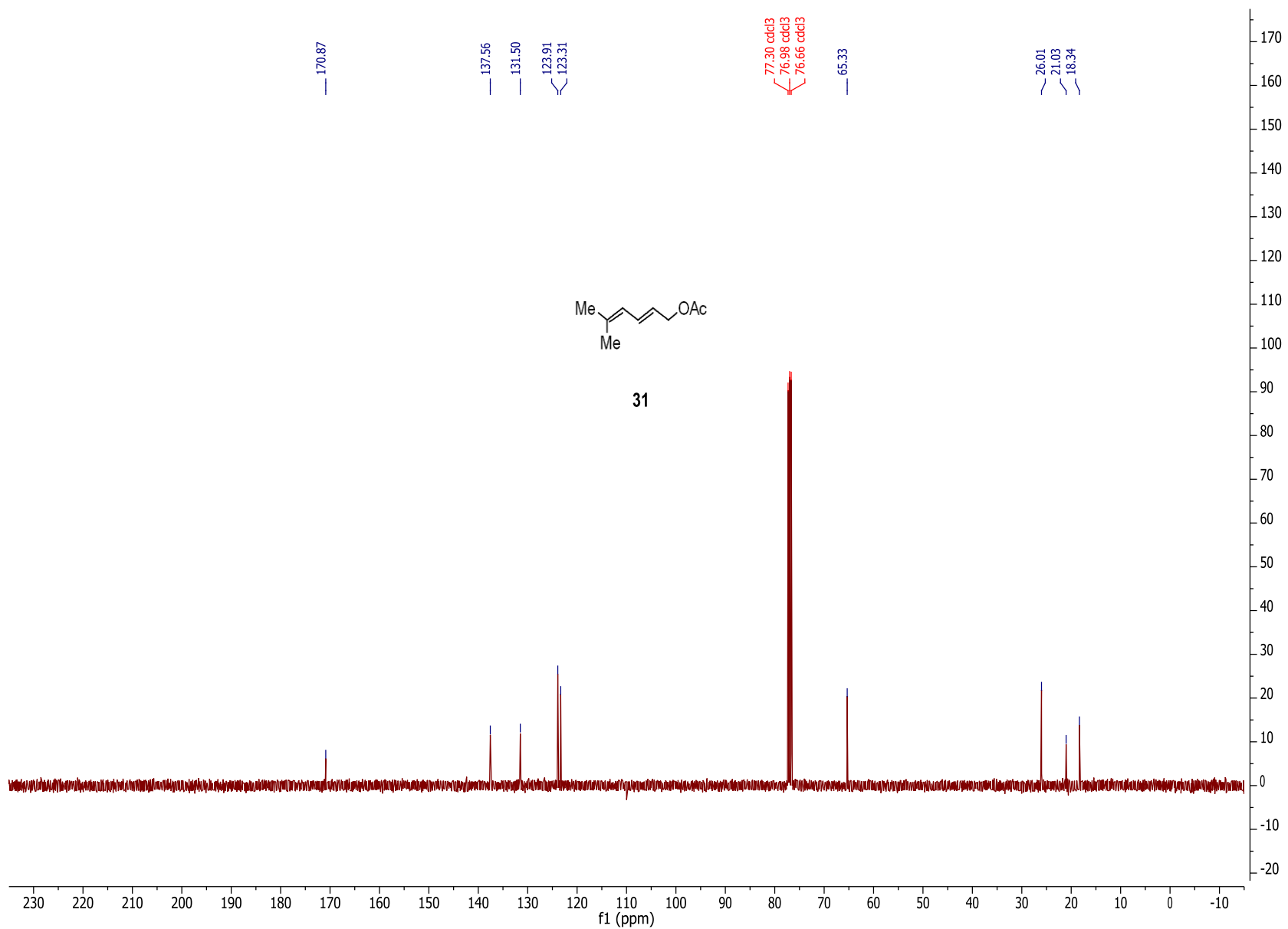


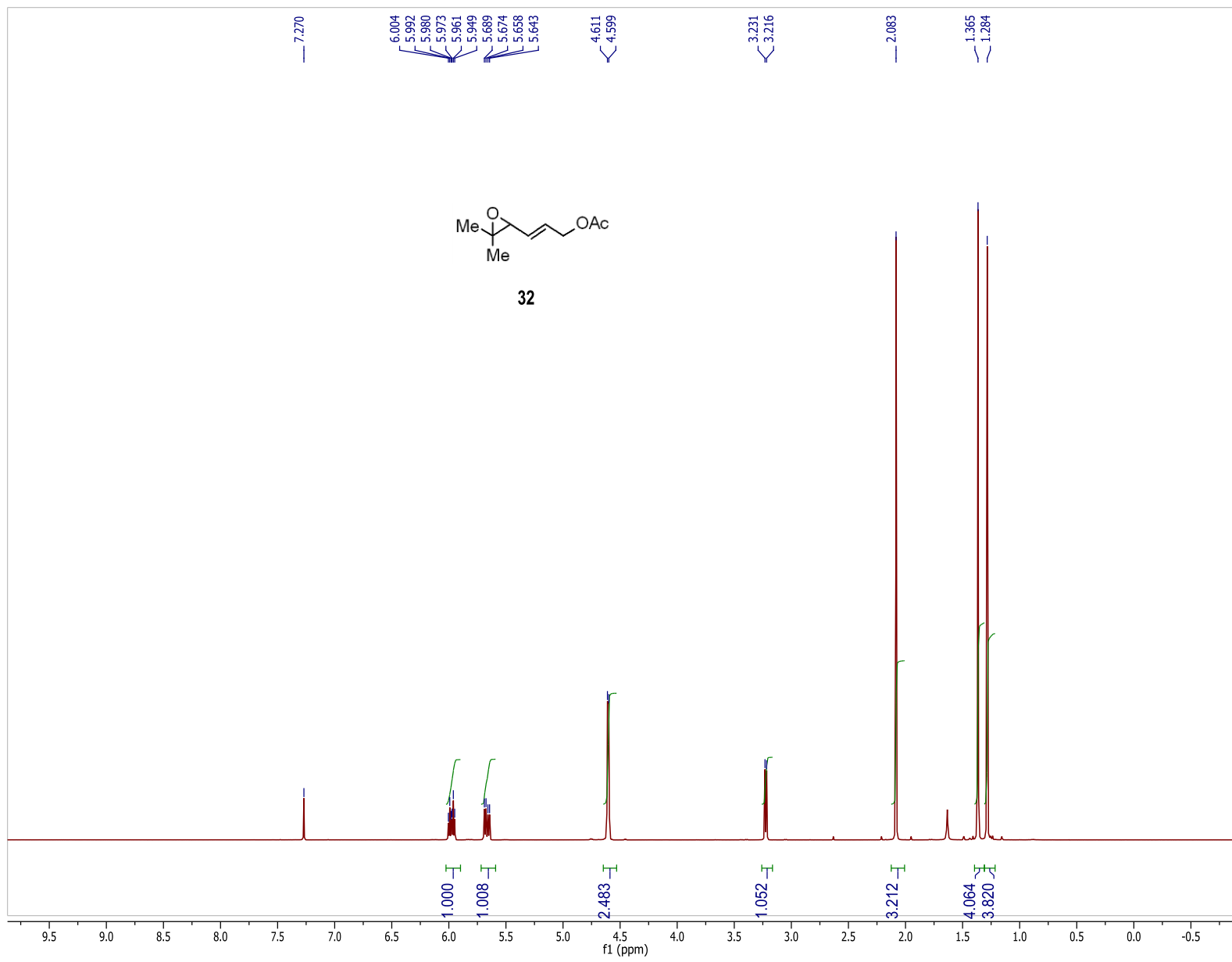


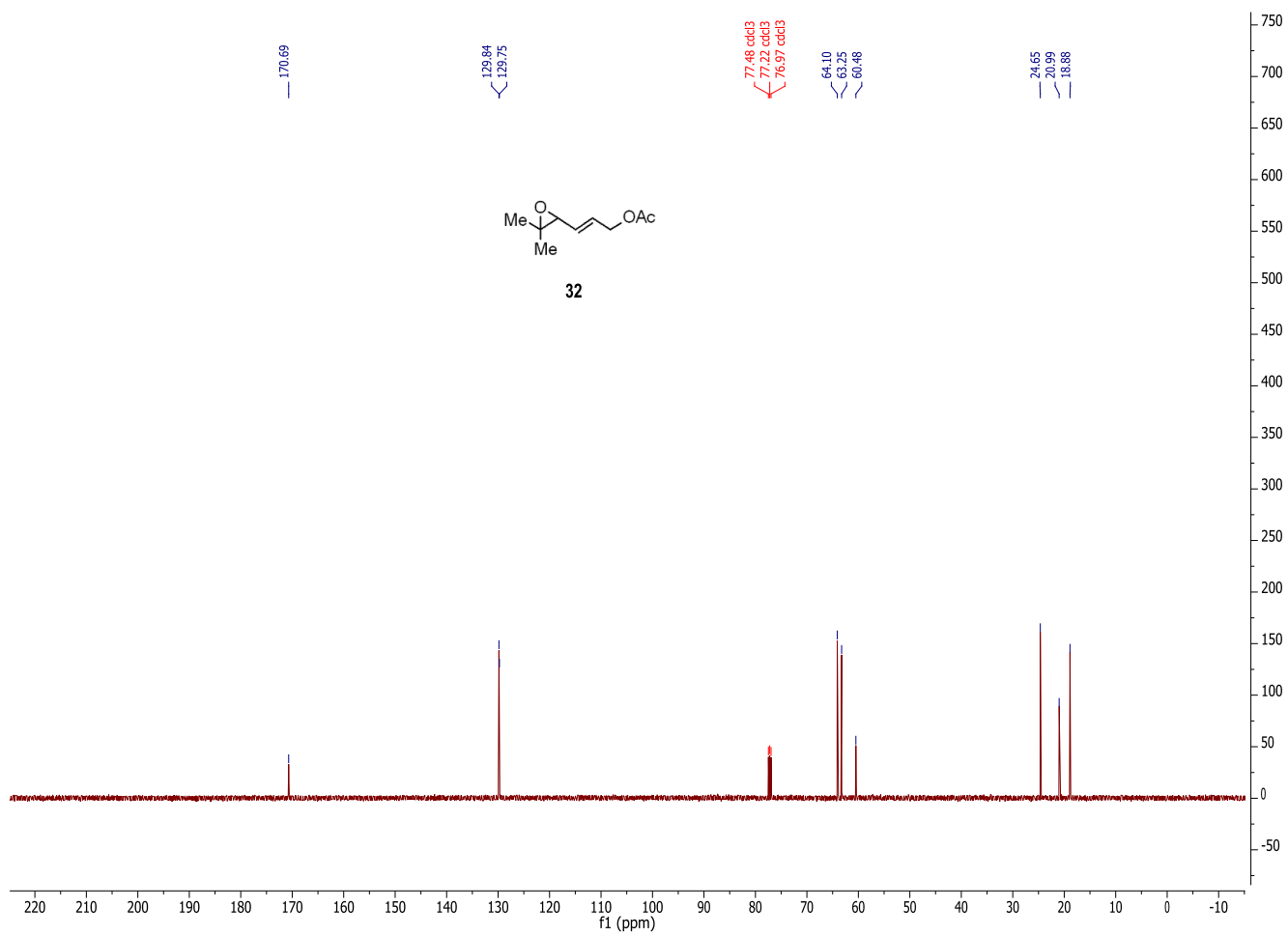




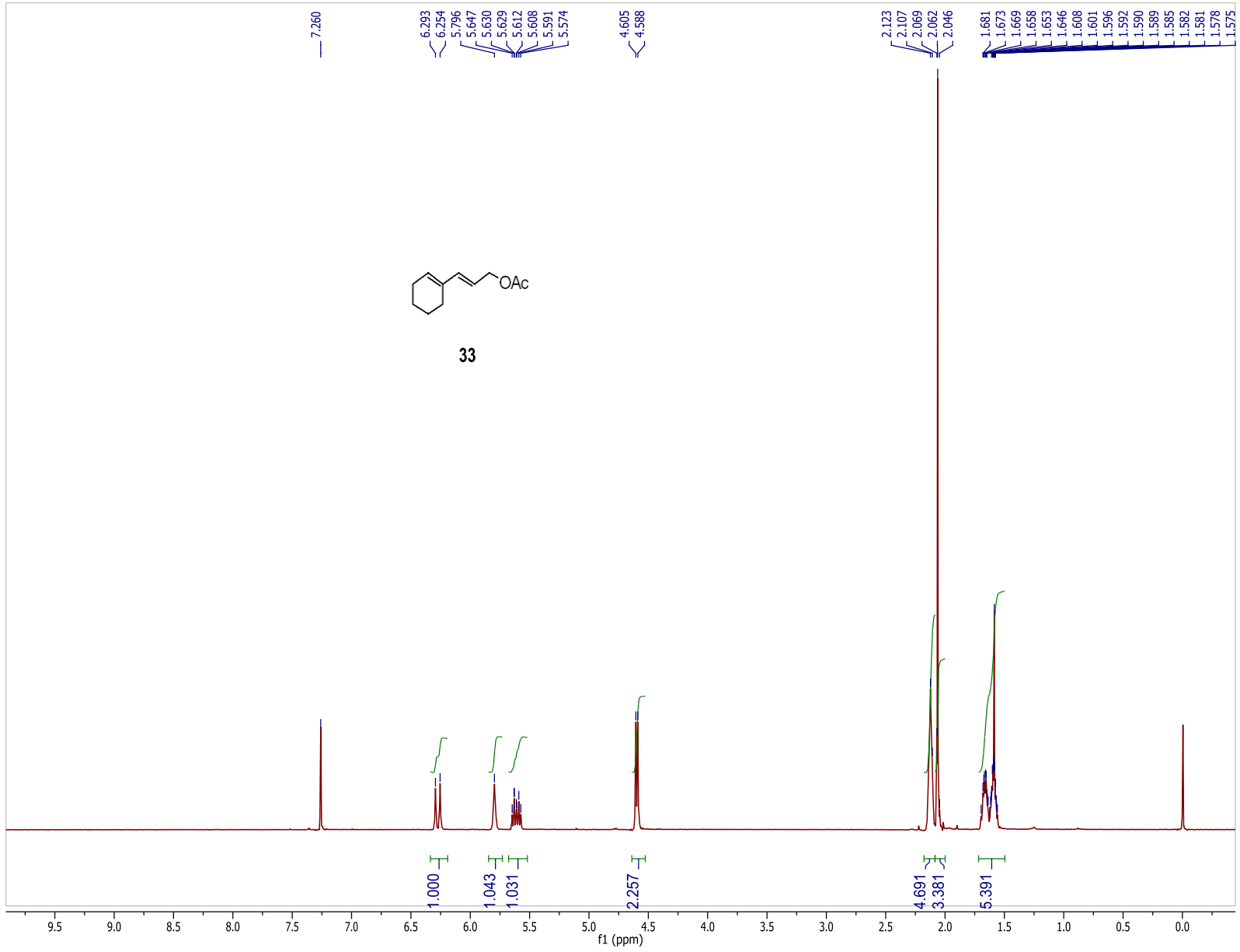


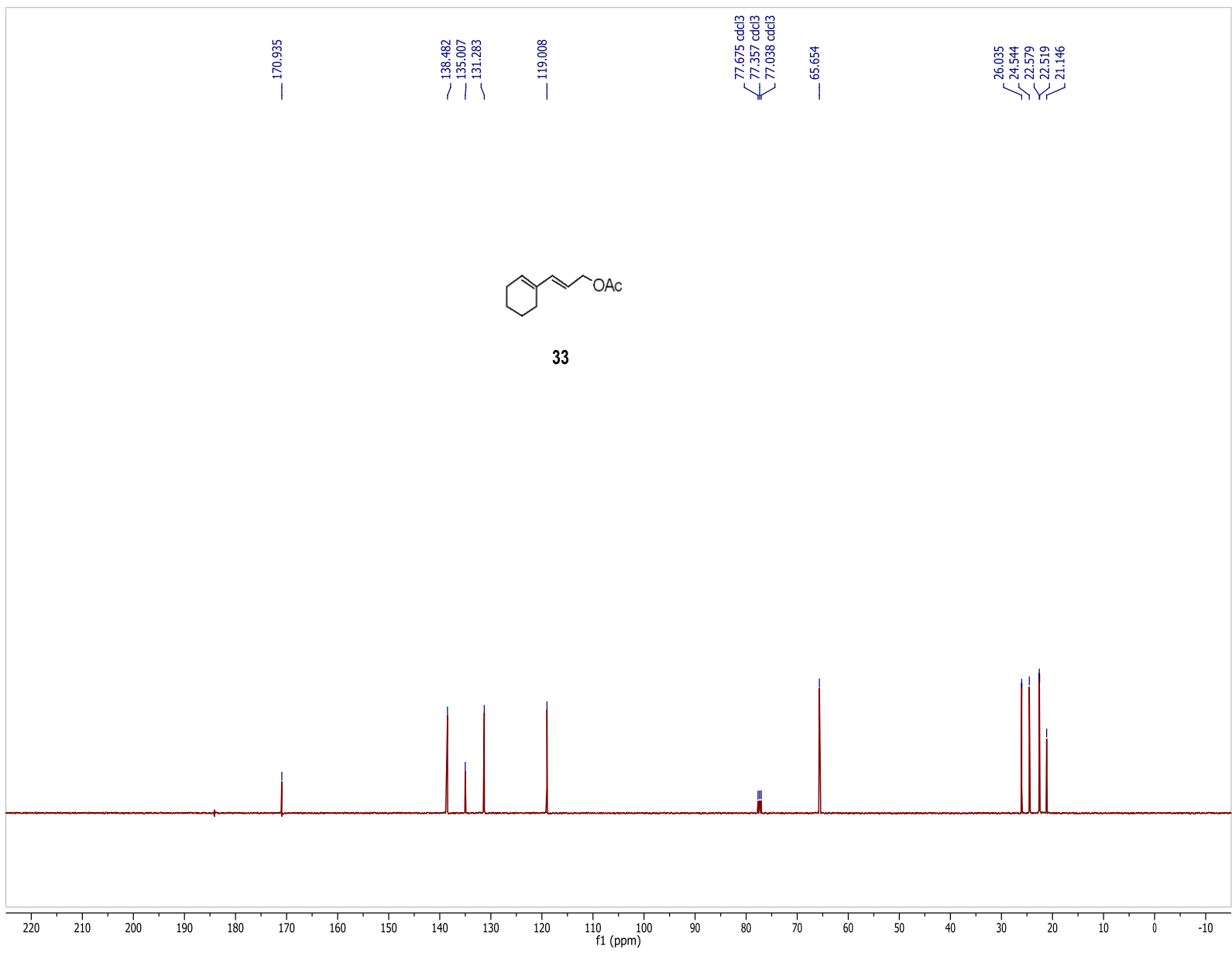


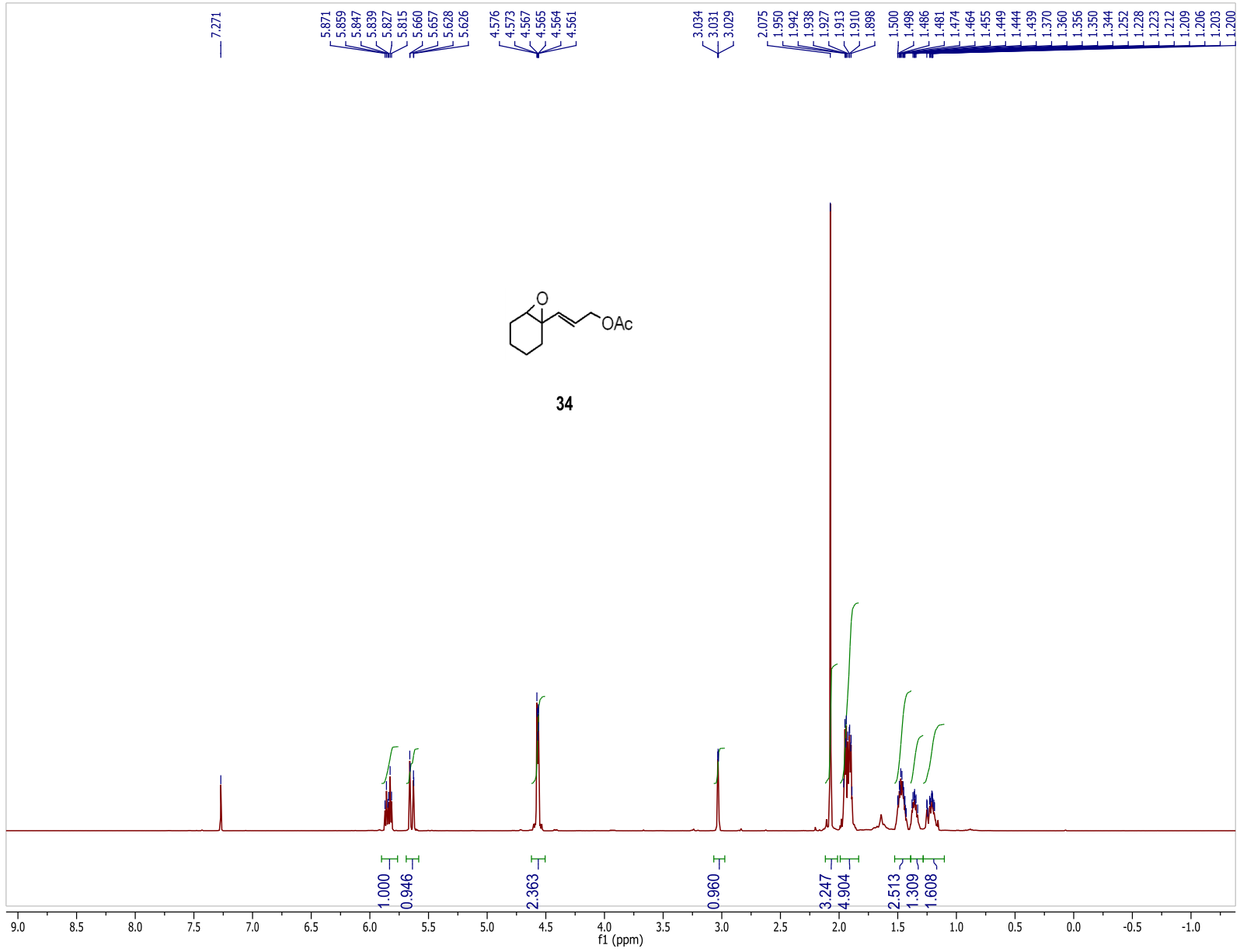


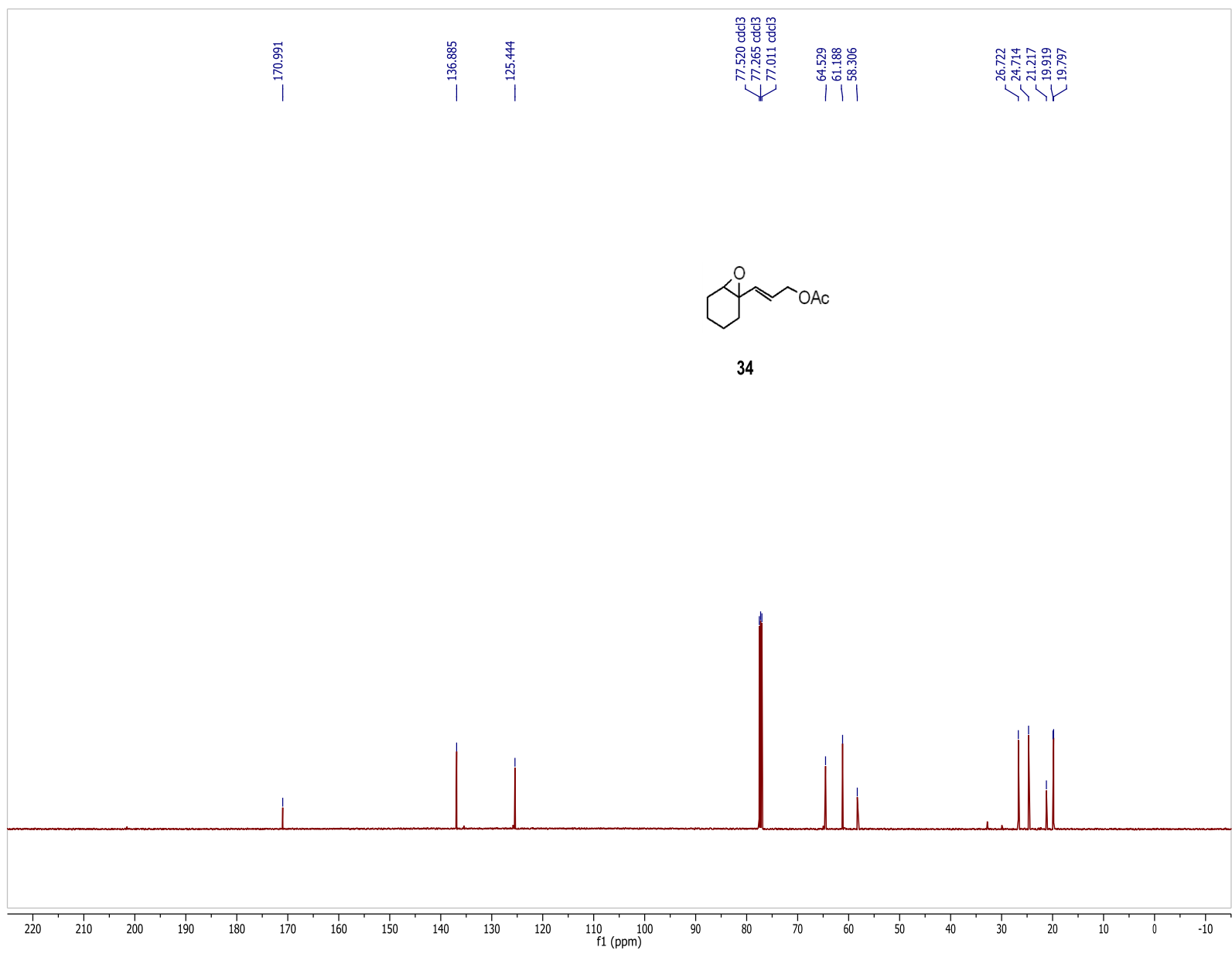


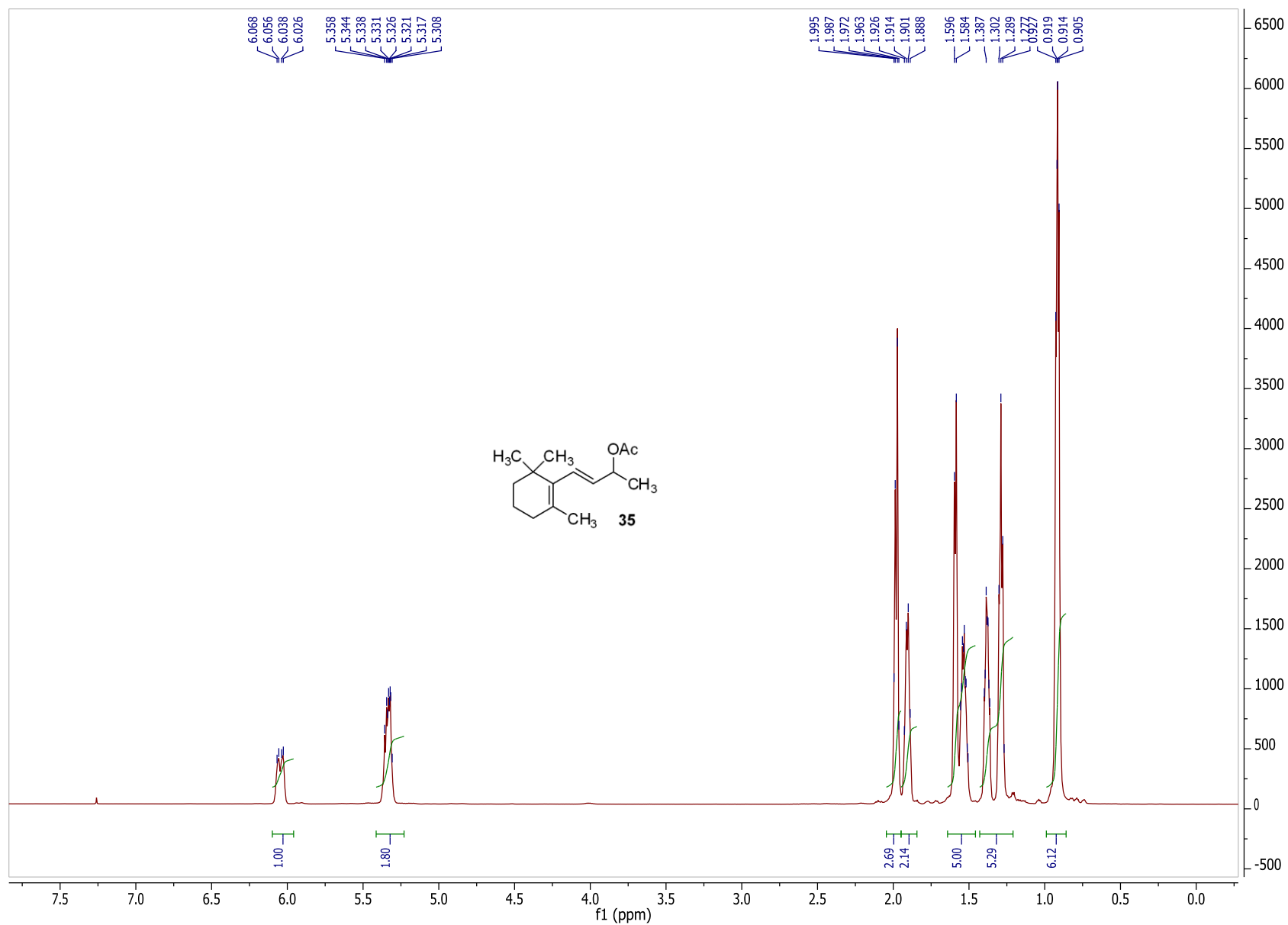


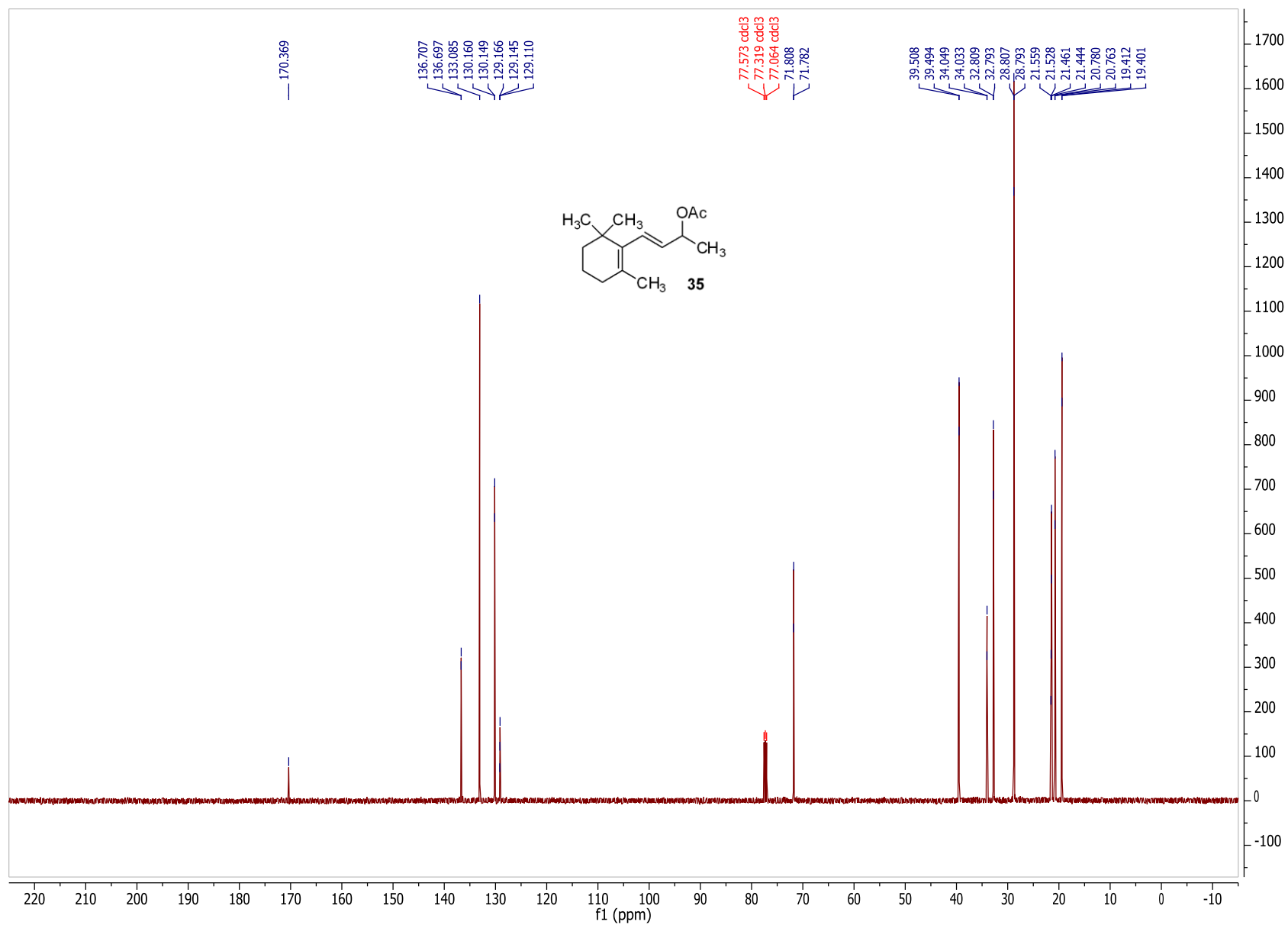


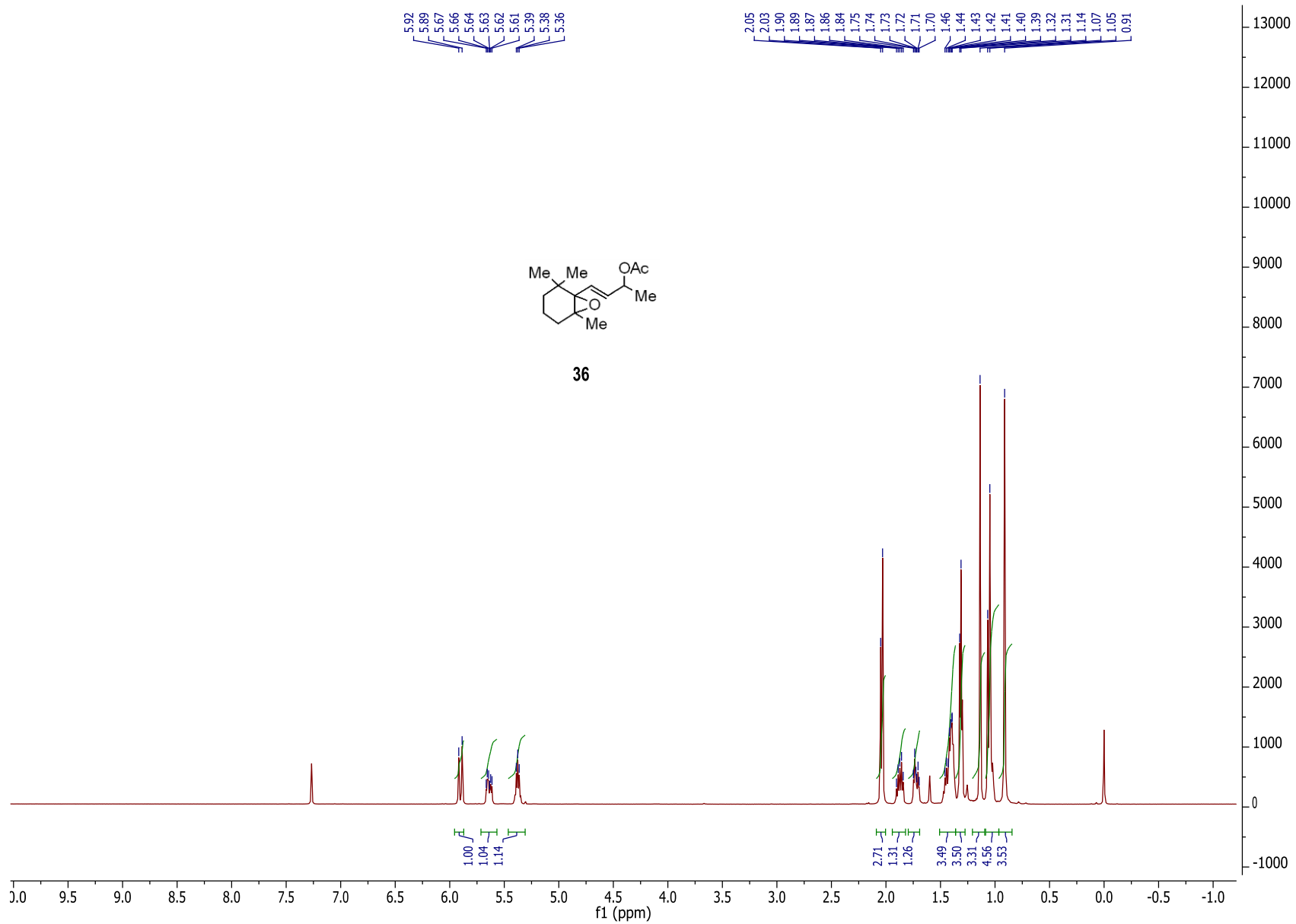


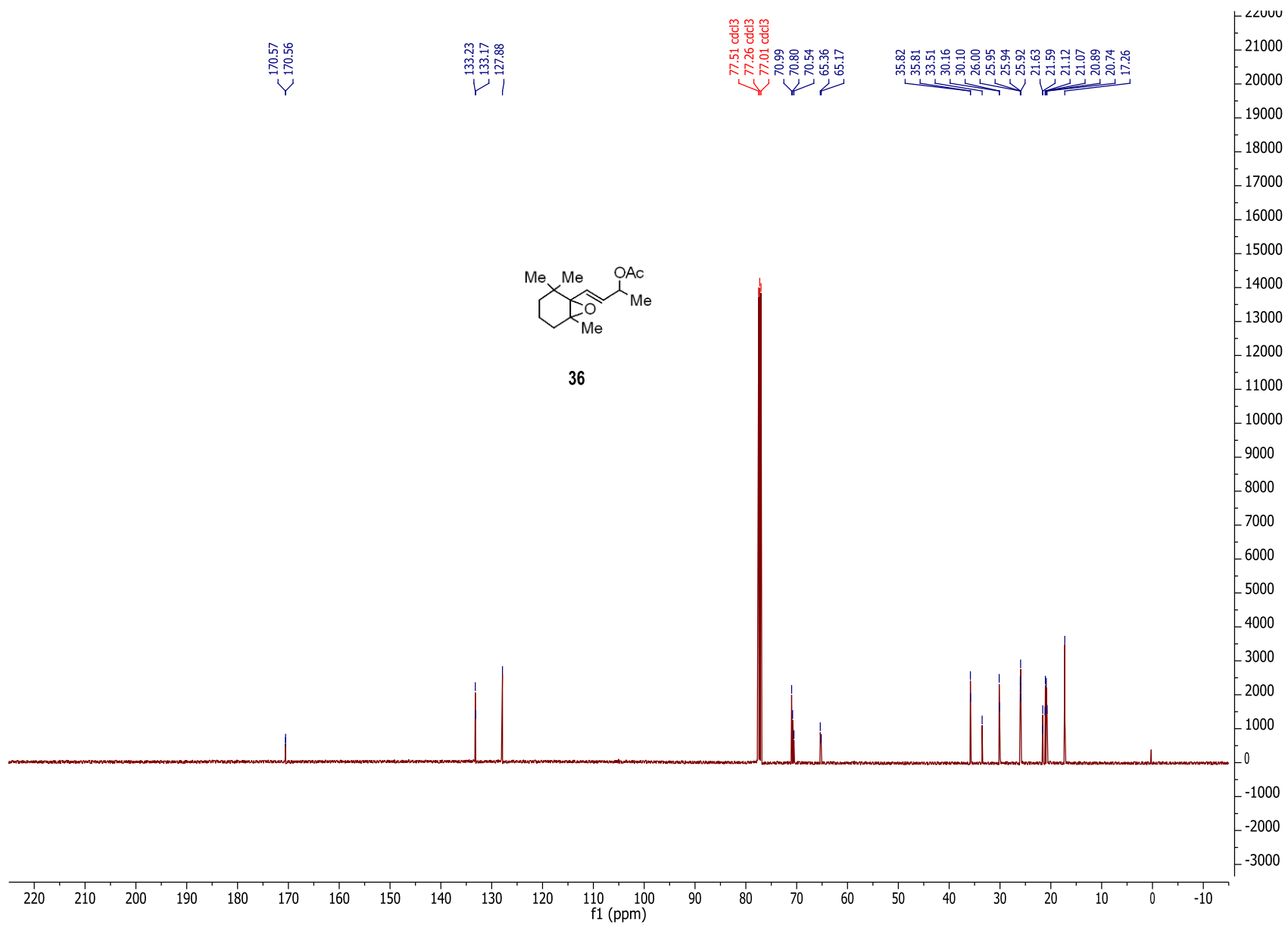




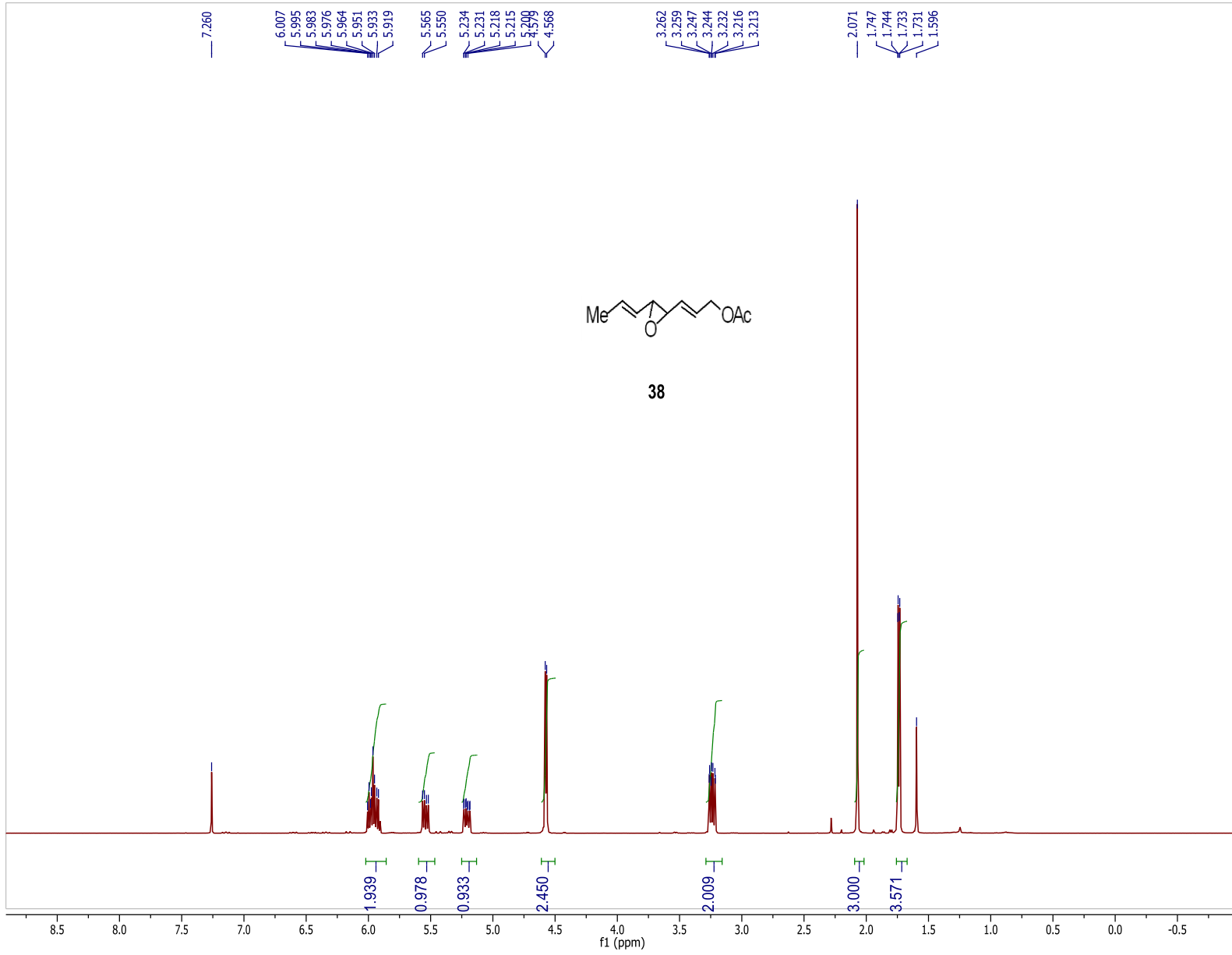


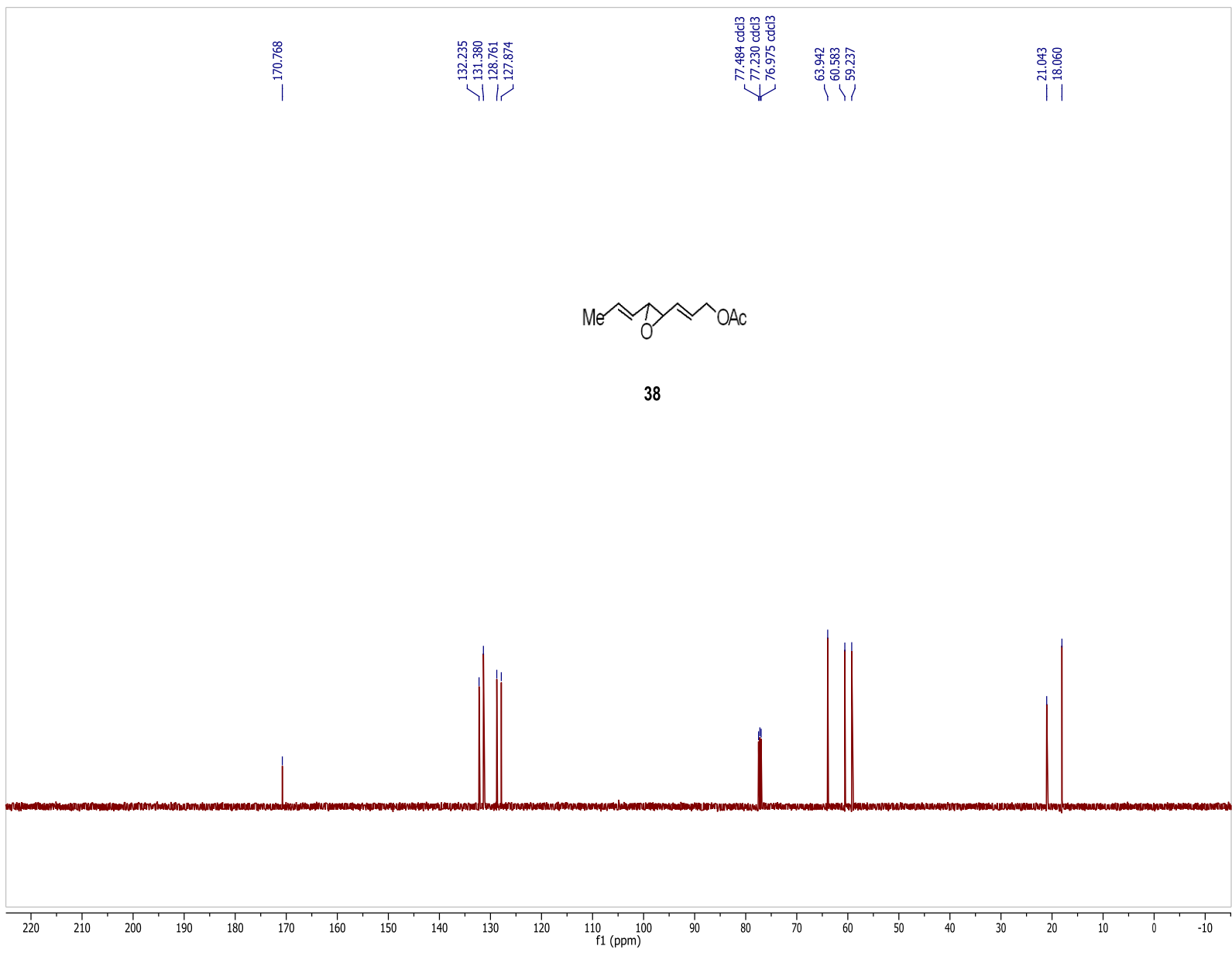


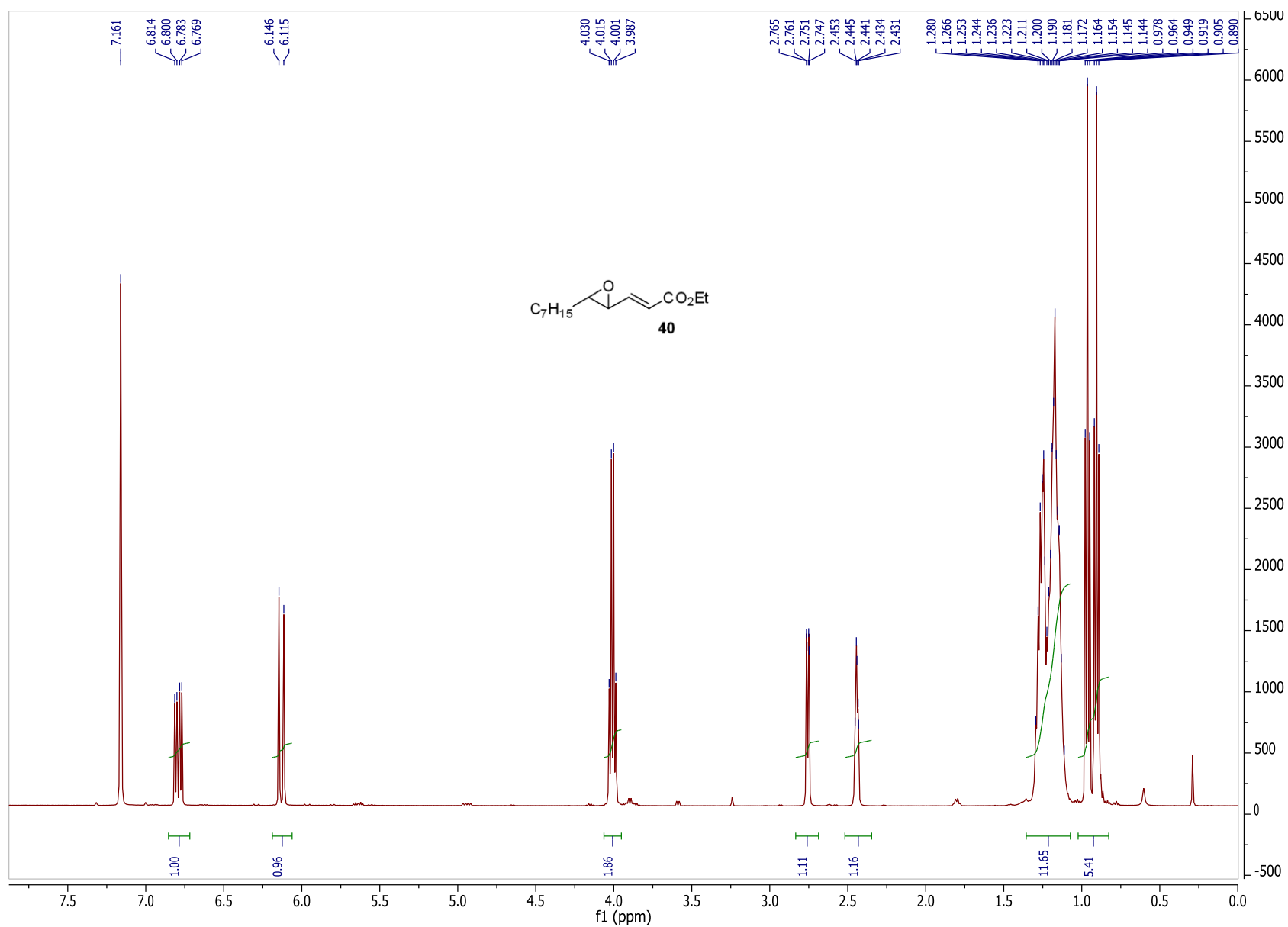


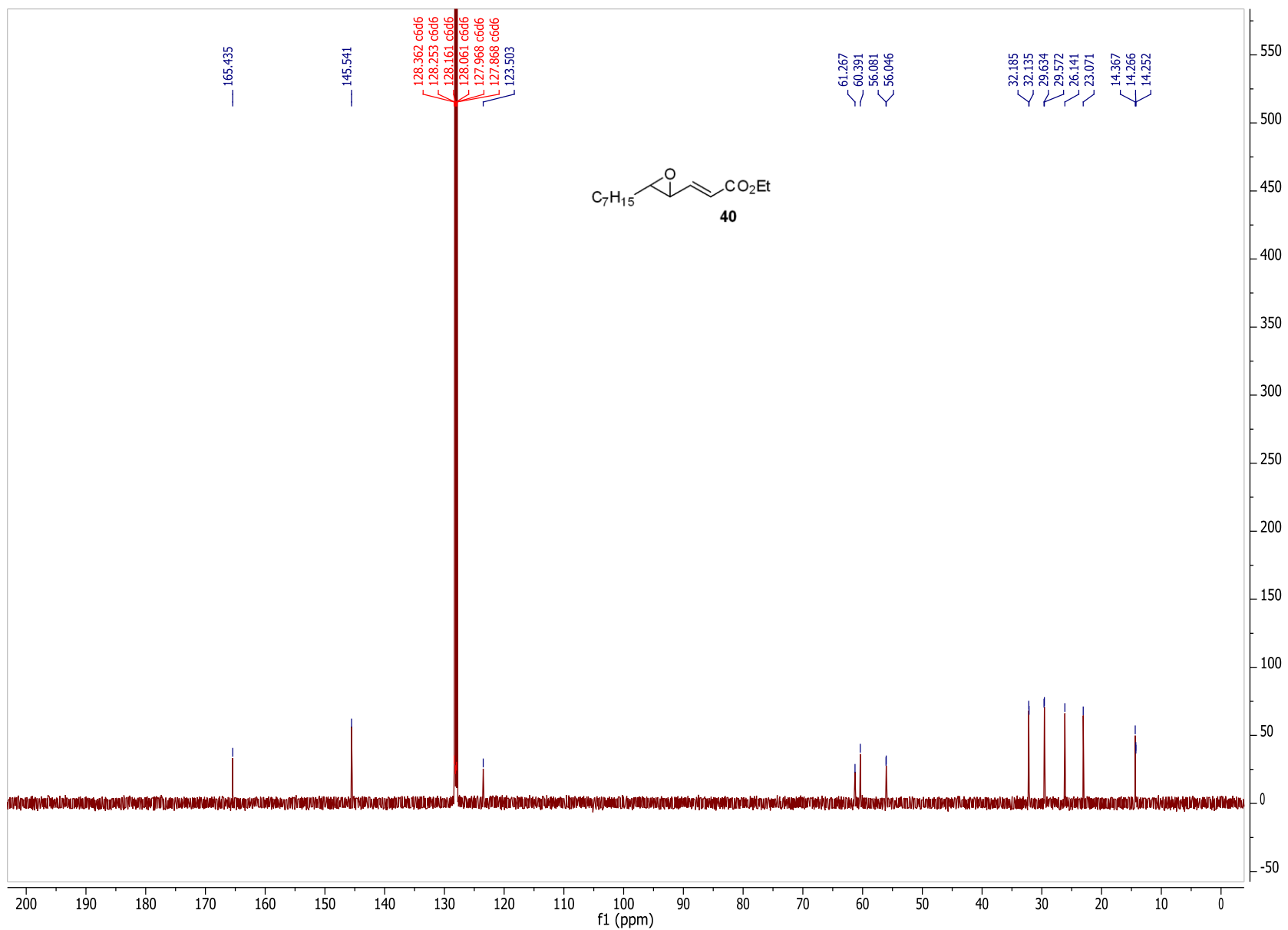


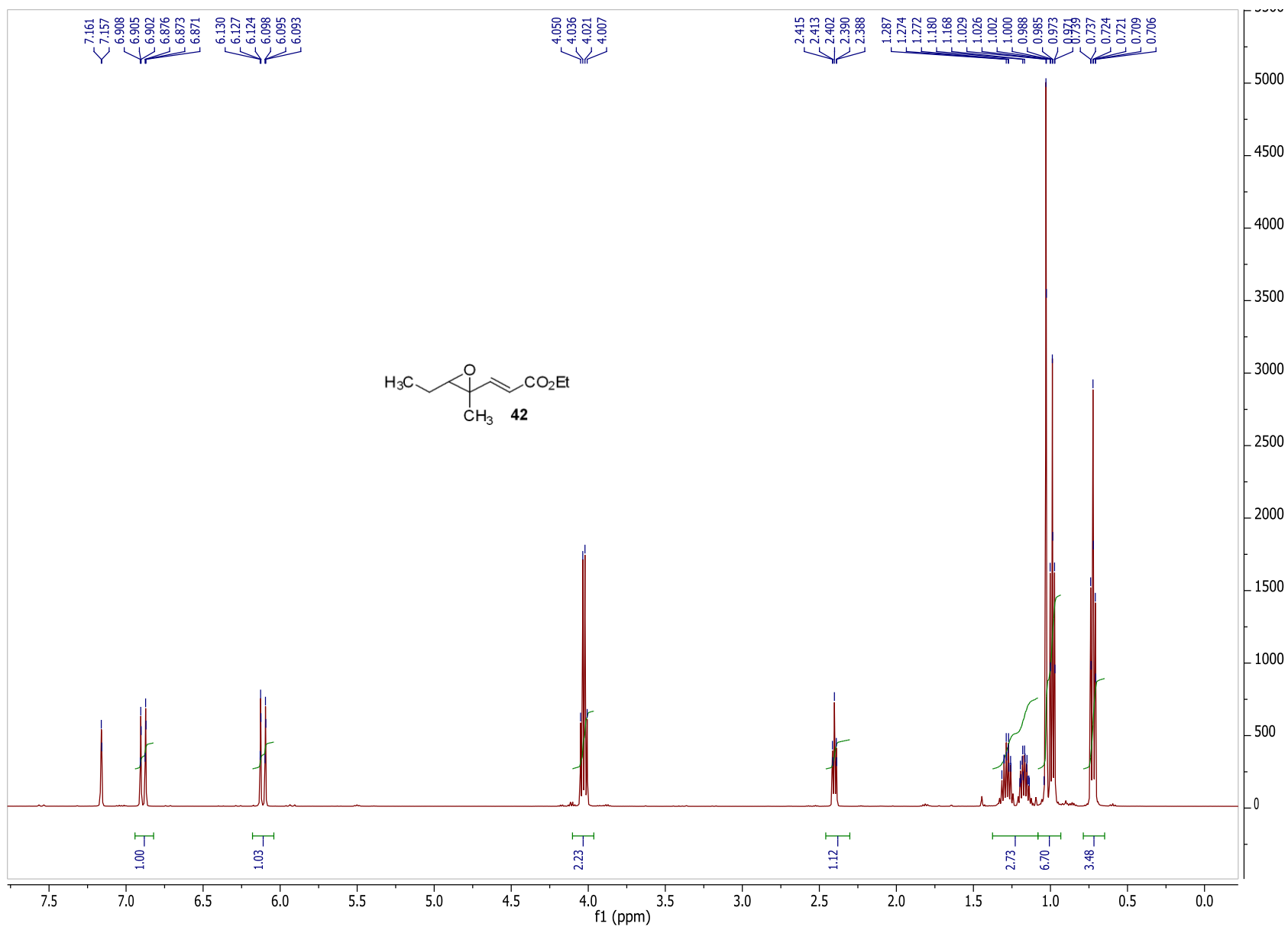


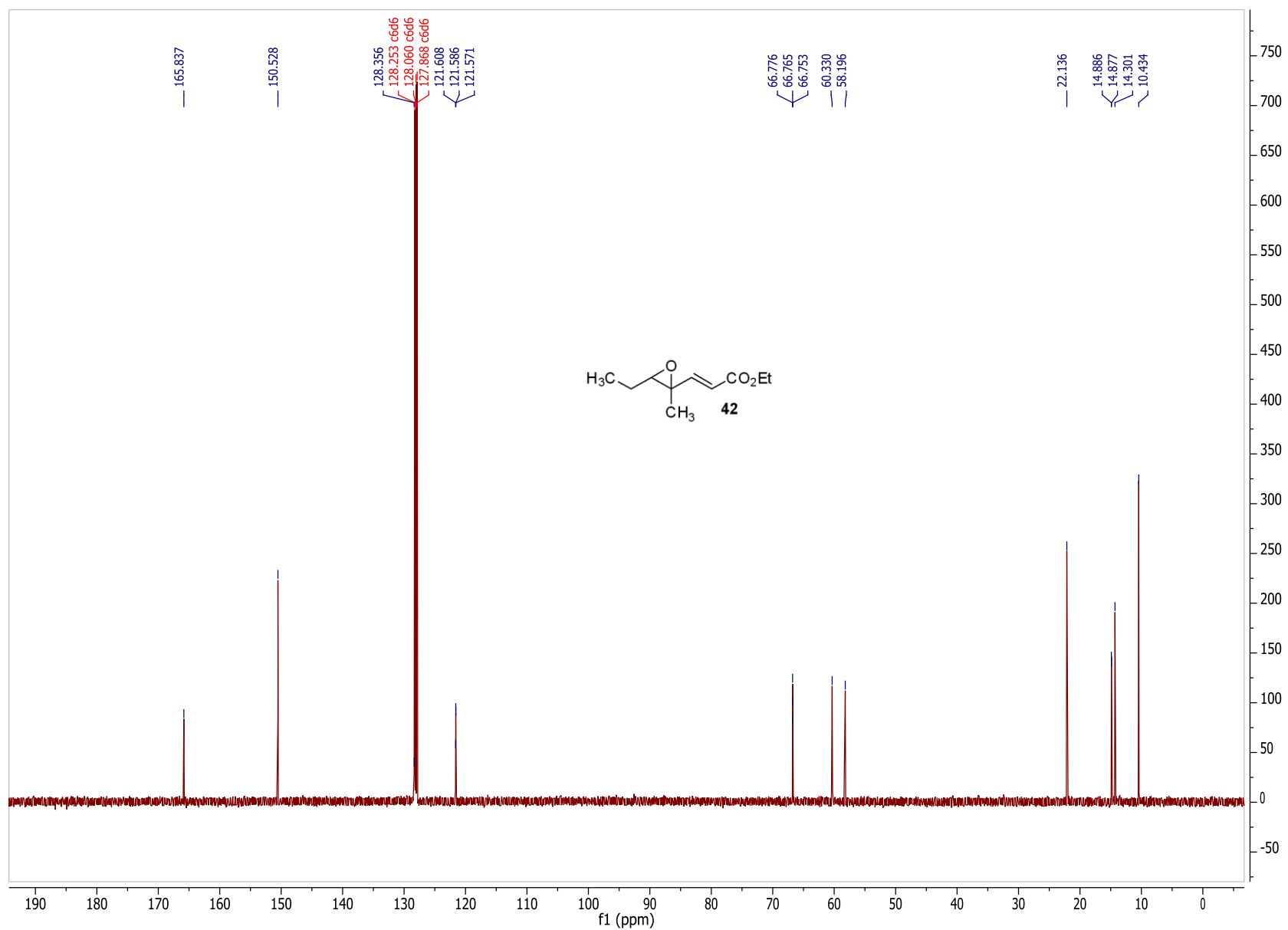






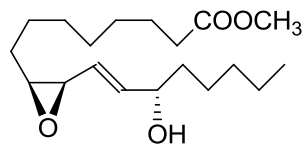




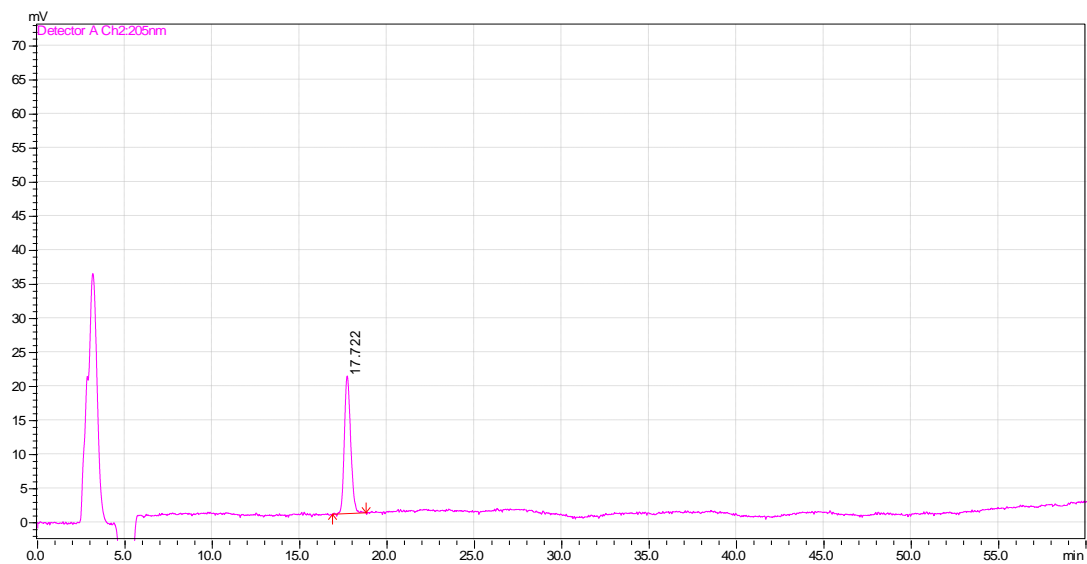


## Stereochemistry Determination:

### 1. HPLC chromatogram of standard chiral epoxide.



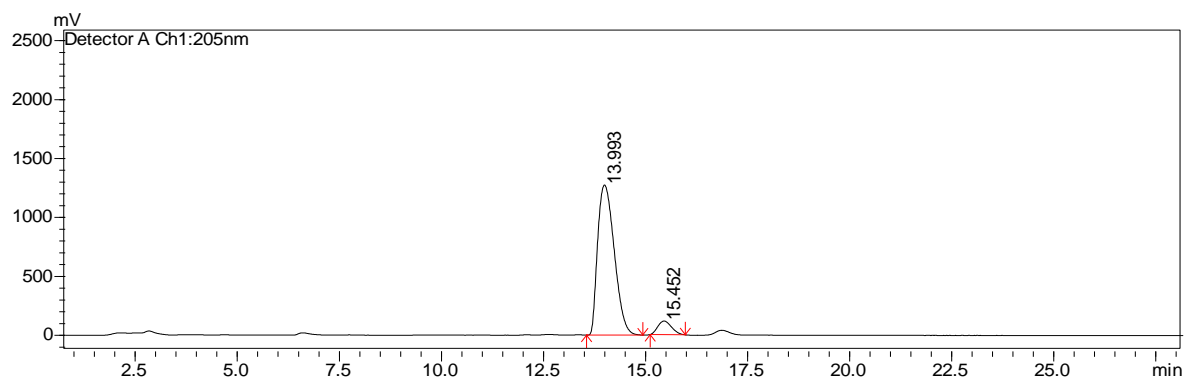
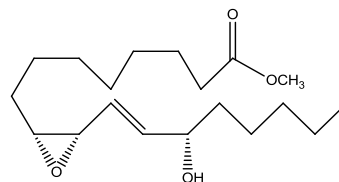
Source: Larodan Fine Chemicals,  
Cat. no.: 24-1802-12b, Lot. No.: H-074



Ascestis Express, 15cm, 4.6mm, 2.7 micron; hexane/IPA :99.9/0.1, 0.75mL/min, 205nm

**2. HPLC chromatogram of chiral epoxide prepared using chiral salan ligand and Ti(IV) using HPLC analysis condition above**

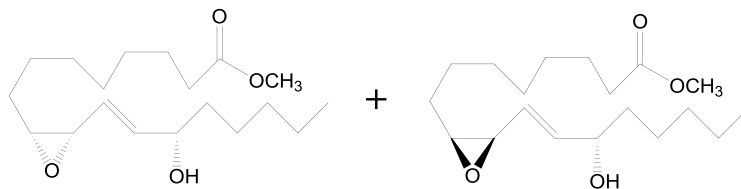
**Reference:** Jat, J. L.; De, S. R.; Kumar, G.; Adebessin, A. M.; Gandham, S. K.; Falck, J. R., submitted for publication.



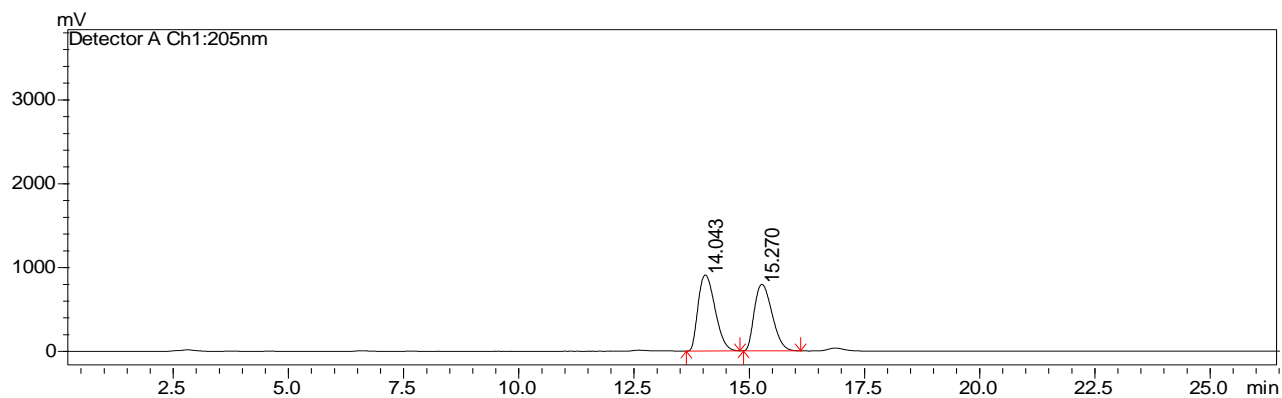
Peak	Ret. Time	Area	Height	Area %
1.	13.993	36061338	1272814	93.039
2.	15.452	2698005	115438	6.961



**3. HPLC chromatogram of 1:1 mixture of chiral Ti(salan)-generated epoxide + commercial epoxide standard using HPLC analysis condition above.**

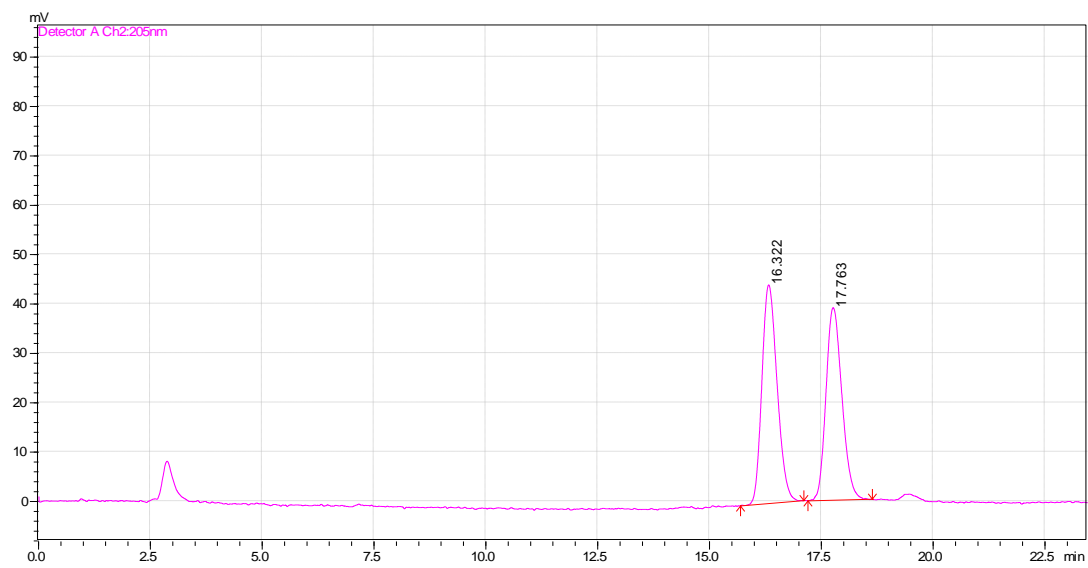
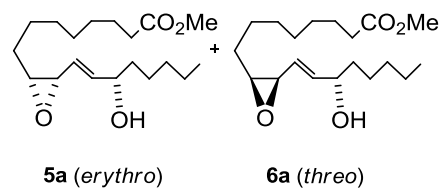


**(1:1 mixture of epoxides)**



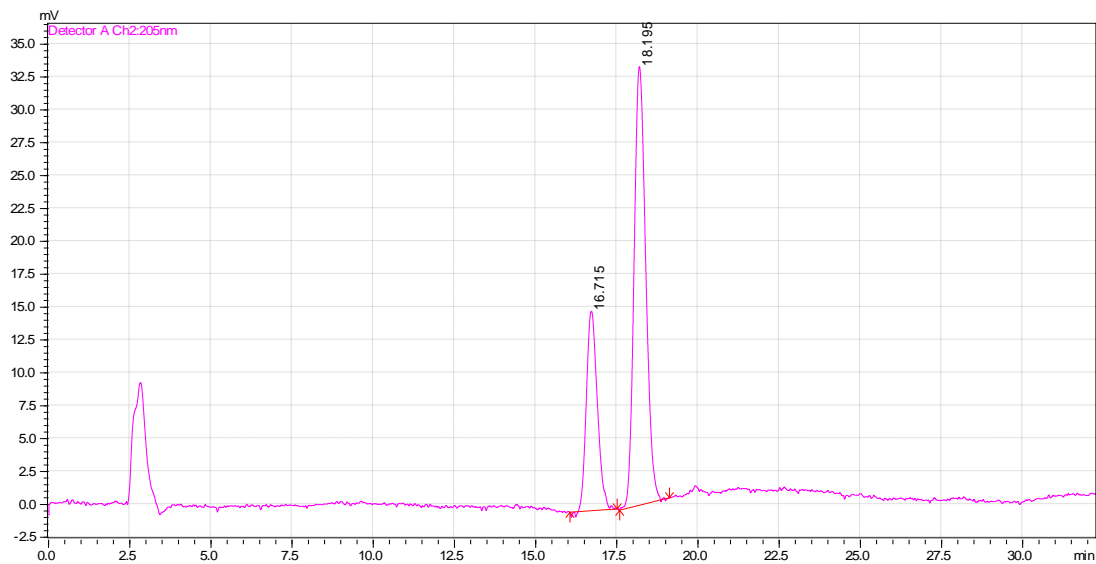
Peak	Ret. Time	Area	Height	Area %
1.	14.043	23200616	908224	52.162
2.	15.270	21276975	793732	47.838

4. HPLC chromatogram of epoxide 5a/6a from MTO-pyridine epoxidation using HPLC analysis condition above.



Peak Table				
Detector A Ch 2 205nm				
Peak #	Ret. Time	Area	Height	Area %
1	16.322	1050224	44308	52.0176
2	17.763	968752	39023	47.9824

5. HPLC chromatogram of epoxide 5a/6a from MTO-pyridine epoxidation + commercial epoxide standard using HPLC analysis condition above.



Peak Table				
Detector A Ch 2 205nm				
Peak #	Ret. Time	Area	Height	Area %
1	16.715	355843	15164	29.9270
2	18.195	833193	33361	70.0730