

## SUPPLEMENTAL DATA: MATERIAL AND METHODS

### Quantitative RT-PCR

cDNAs were synthesized from 500 ng of total DNase-treated RNA with the Maxima First Strand cDNA synthesis kit (ThermoScientific). The Platinum Sybr Green qPCR Supermix-UDG (Invitrogen) and the Lightcycler System (Roche Diagnostics) were used for analysis of *mmp9* and *il17* expression by real-time quantitative PCR. The expression of each gene was normalized to the expression of *gapdh* and the 'Fold changes' were calculated by normalization to the control sample. Experiments have been reproduced at least 5 times, error bars and statistics were calculated based on all measurements for each gene.

### Immunohistochemistry/Immunocytochemistry

Tissues were fixed in paraformaldehyde, embedded in paraffin and sectioned. Five-micrometer sections of skin were deparaffinized and rehydrated. Cytospun cells were fixed for 15 minutes in paraformaldehyde and permeabilized in cold methanol for 7 minutes. Then heat retrieval in CC1 cell conditioning solution (Ventana Medical Systems, Tucson, AZ) was performed for 25 minutes at 95°C on both tissue sections and cytospun cells. The sections were blocked with normal horse serum (0.2%; Vector Laboratories) for 30 min at room temperature. Then simultaneous staining was performed for 30 min at room temperature with goat anti-IL-17 (100 mg/ml; R&D Systems, Minneapolis, MN) and another primary antibody: either rabbit anti-human CD3 (1/50; Dako), mouse anti-human mast cell tryptase (1:150; Dako), or rabbit anti-human myeloperoxidase (1:150; Dako). This was followed by 30 min of incubation at room temperature with matched secondary antibodies: chicken anti-goat IgG Alexa Fluor 488 (chicken anti-mouse IgG Alexa Fluor 594 and chicken anti-rabbit Alexa Fluor 594 (Invitrogen). Hoescht reagent was added to the slides prior to mounting with

coverslips. Images were captured with confocal microscope (LSM 710; Zeiss) using Zen software.