

CRISPR/Cas9 mediates efficient conditional mutagenesis in *Drosophila*

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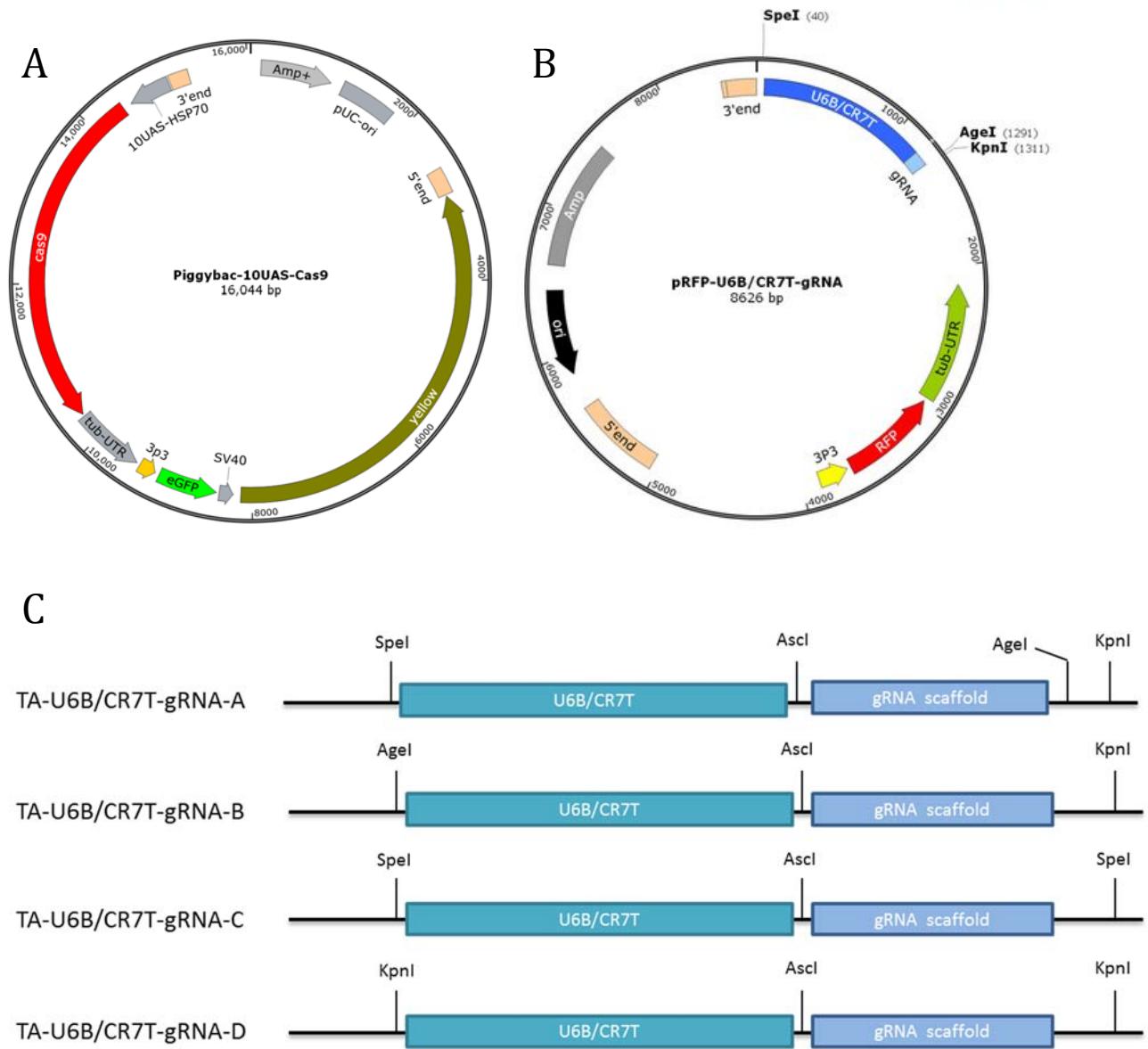
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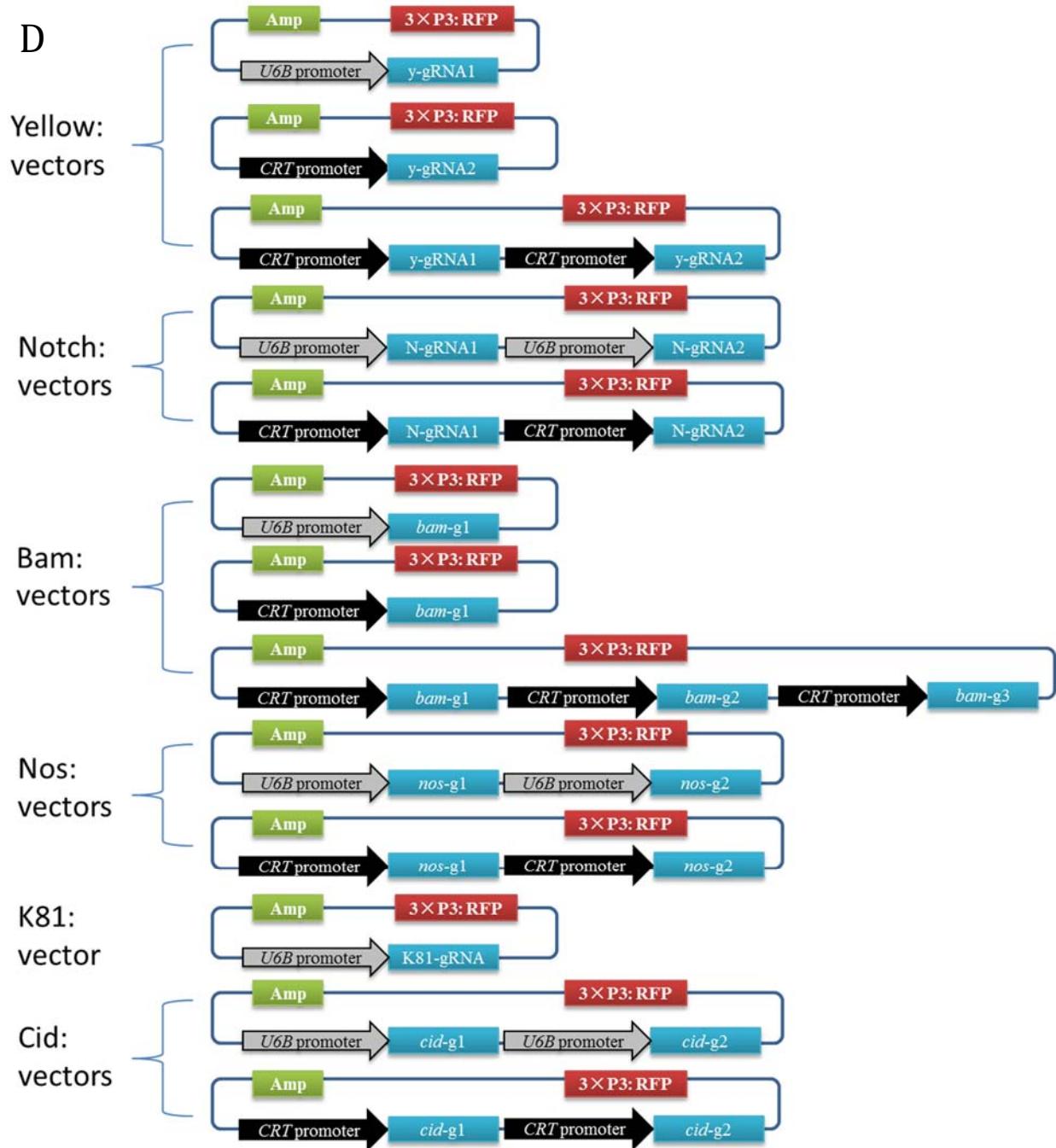


Figure S1 Maps of the plasmids. (A) Piggybac-10UAS-cas9, (B) pRFP-U6B/CR7T-gRNA, and (C) the four backbone plasmids for gRNA insertion. (D) Maps of all transgenic gRNA vectors.

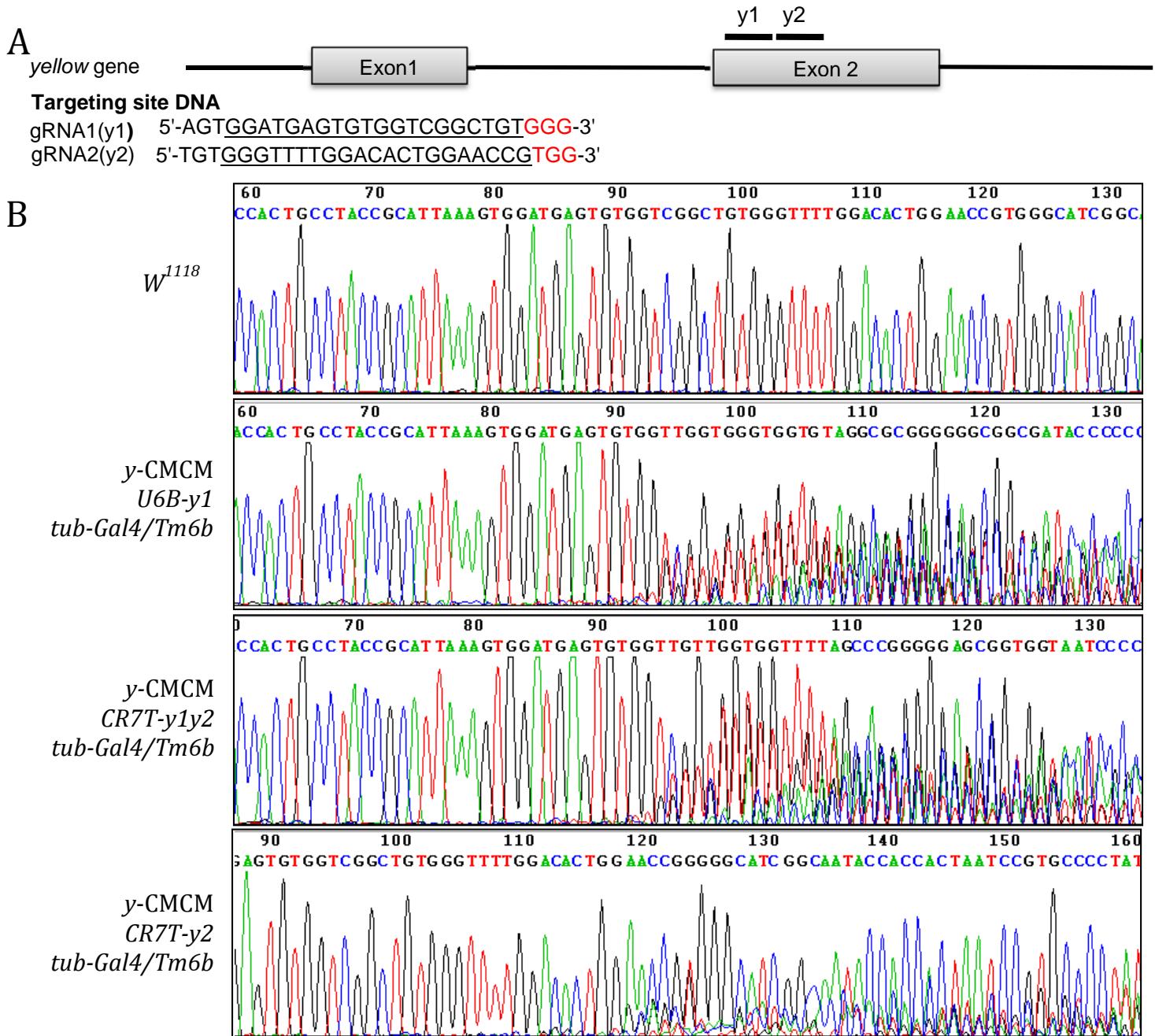
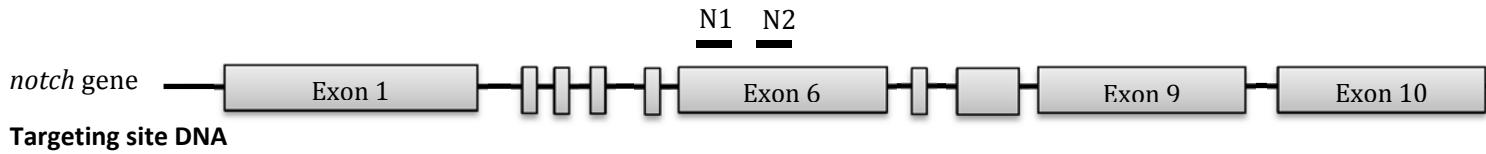


Figure S2 Sequence results for the *y* conditional mutant flies. (A) Sequences and schematic representation of the two gRNAs against the *y* gene. (B) *Tub-Gal4/Tm6b* was used to drive the expression of Cas9 in whole bodies, and three vectors, *U6B-y1*, *CR7T-y2*, and *CR7T-y1y2*, were used to drive the expression of gRNA. The mutations were induced exactly at the target locus. *w*¹¹¹⁸ was used as the control.

A



B

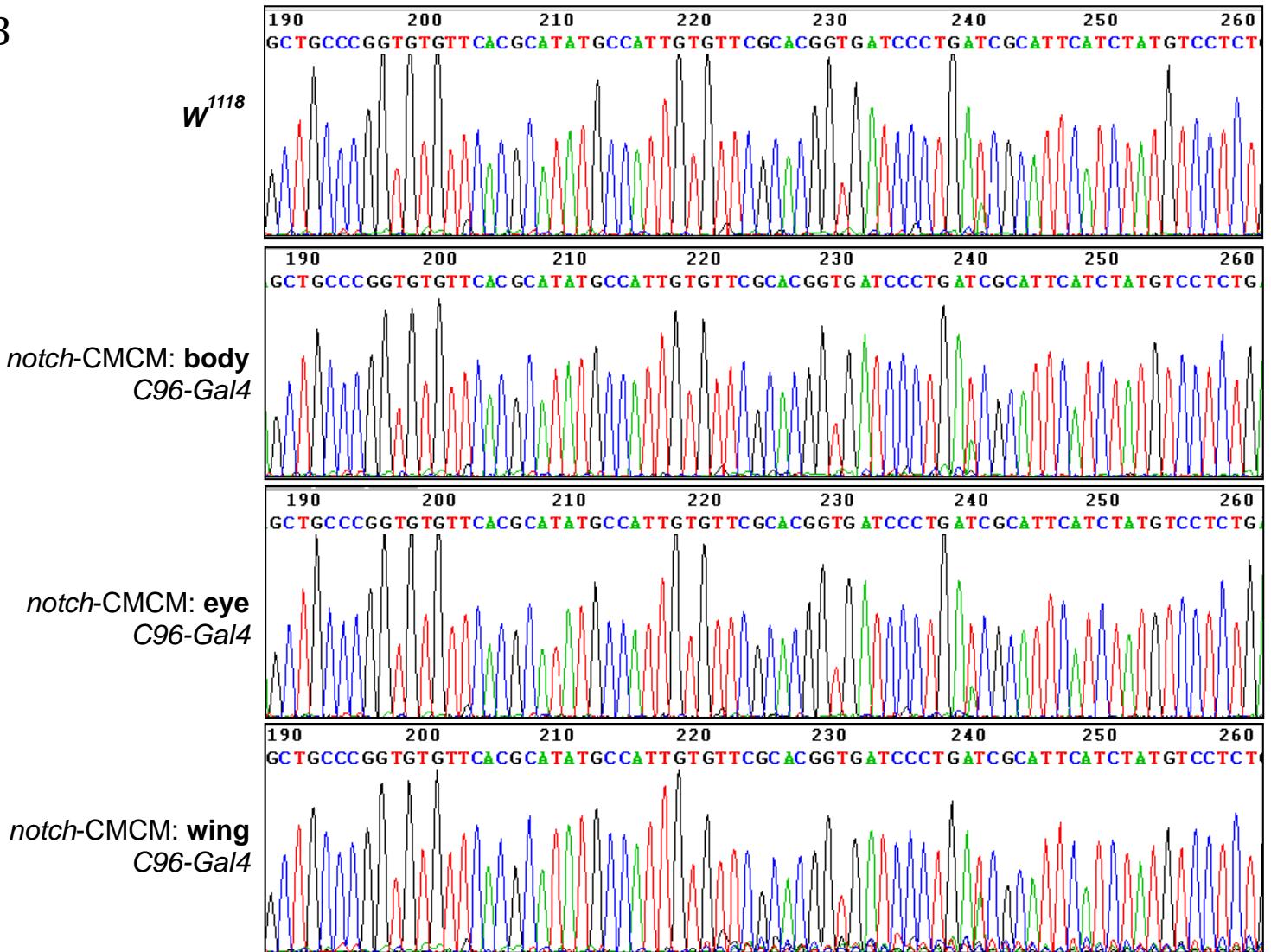
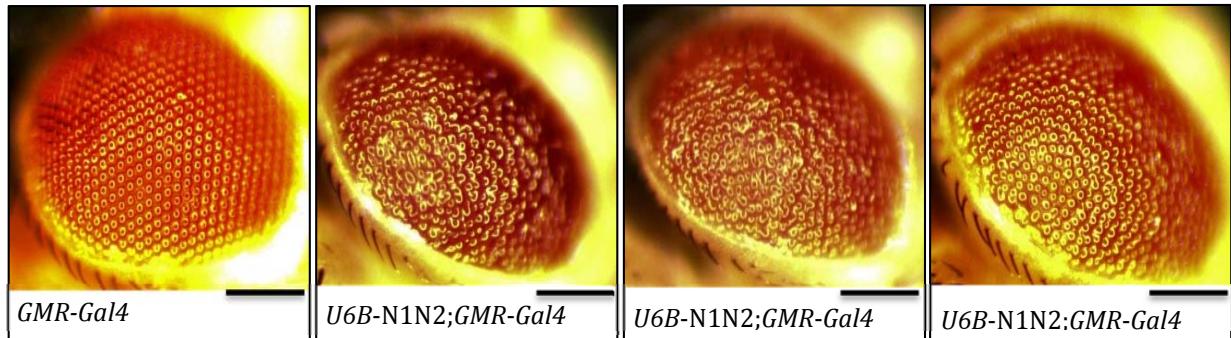


Figure S3 Sequence results for notch conditional mutant flies. (A) Sequences and schematic representation of two gRNAs against the notch gene. (B) C96-Gal4 was used to drive the expression of Cas9 specifically in the blade region of the wing imaginal disc. The body, eye, and wing from the notch conditional mutant flies were sequenced. The mutation was induced only in the wing tissue at the target locus. *w*¹¹¹⁸ was used as the control.

A



B

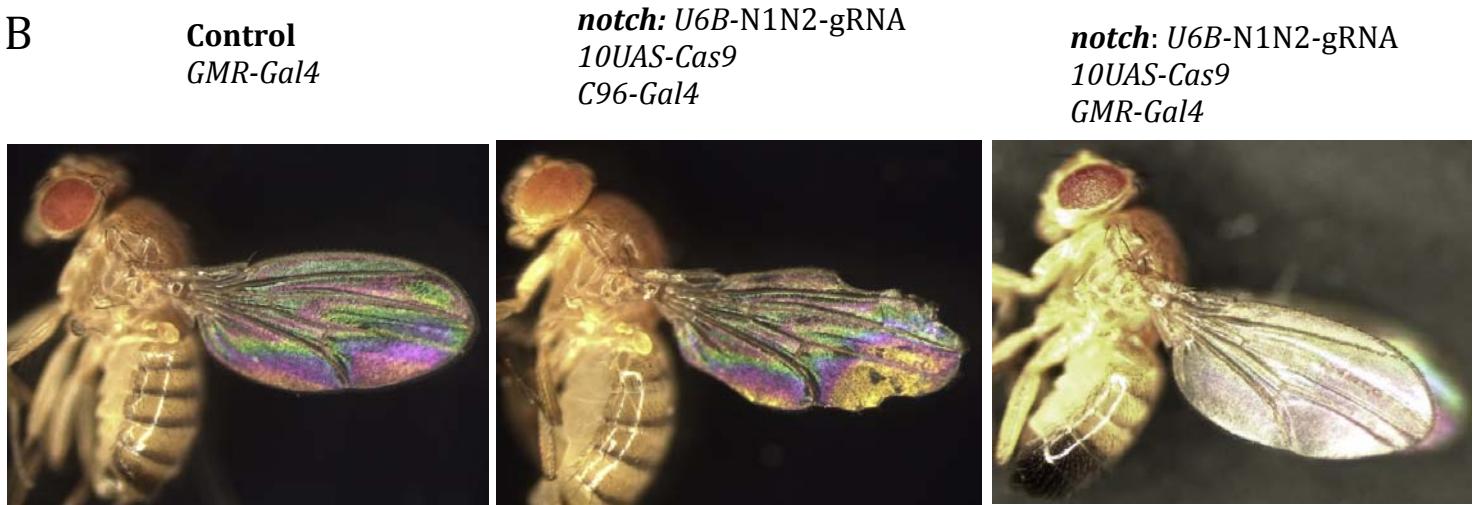


Figure S4 Phenotypes resulting from the conditional *notch* mutation in the eye and wing. (A) The conditional mutant flies showed rougher eyes than the *GMR-Gal4* control. Scale bars: 100 μm . (B) The whole-fly images for *notch* conditional mutagenesis are shown. The left-hand image used the *GMR-Gal4* fly as a control, the middle image is from the *notch* conditional mutagenesis driven by wing-specific *C96-Gal4*, and the right-hand image is from the *notch* conditional mutagenesis driven by eye-specific *GMR-Gal4*. *U6B-N1N1* was used to drive the expression of gRNA.

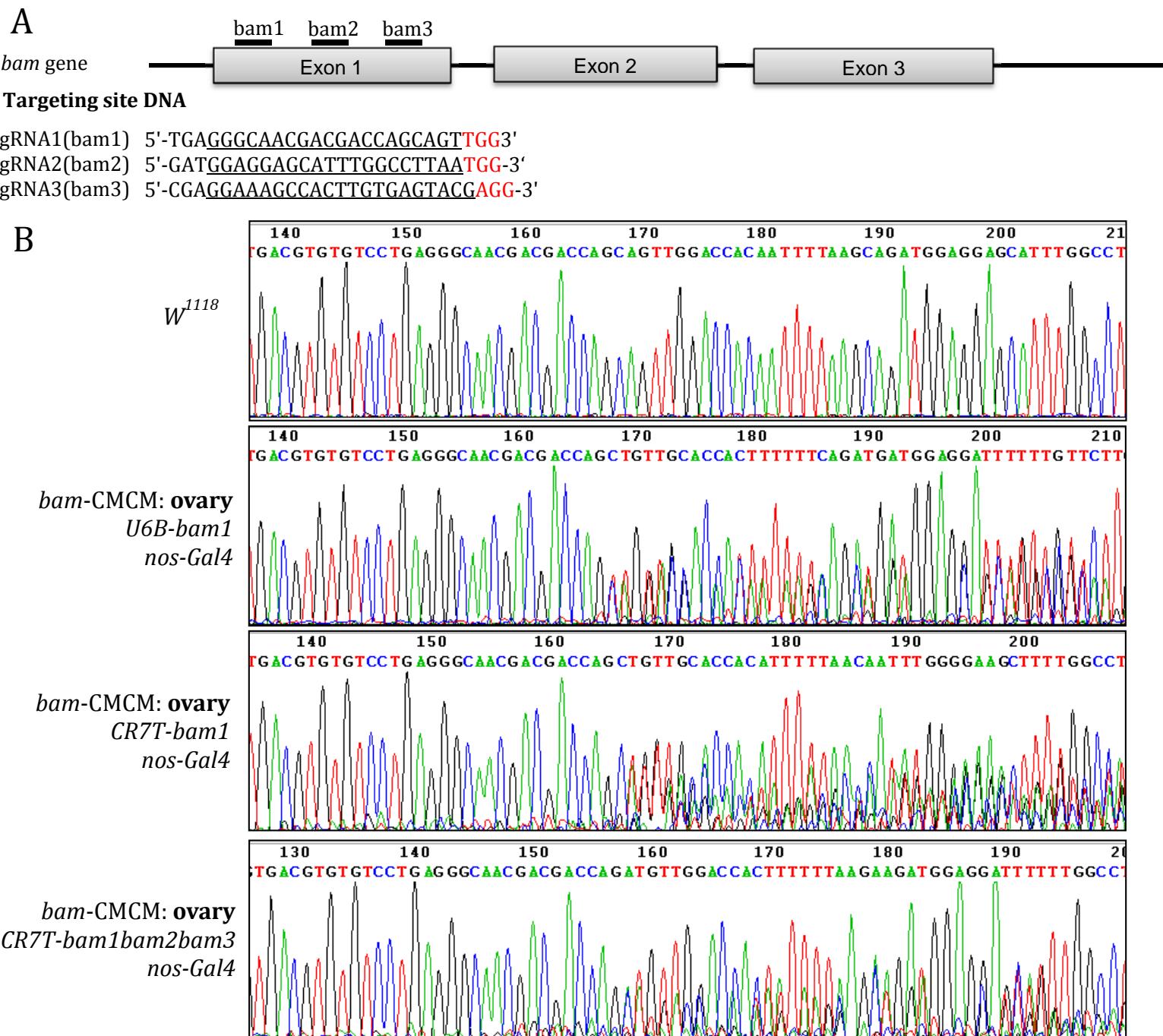
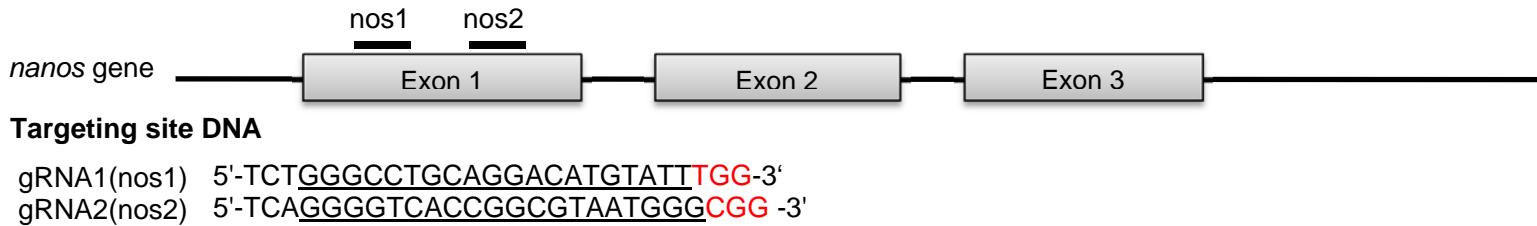


Figure S5 Sequence results for the ovaries of *bam* conditional mutant flies. (a) The sequences and a schematic representation of three gRNAs against the *bam* gene are shown. (b) *Nos-Gal4* was used to drive the expression of Cas9 specifically in the ovary, and three vectors, *U6B-bam1*, *CR7T-bam2*, and *CR7T-bam1bam2bam3*, were used to drive the expression of gRNA. The mutations were induced exactly at the target locus. A *w*¹¹¹⁸ fly was used as the control.

A



B

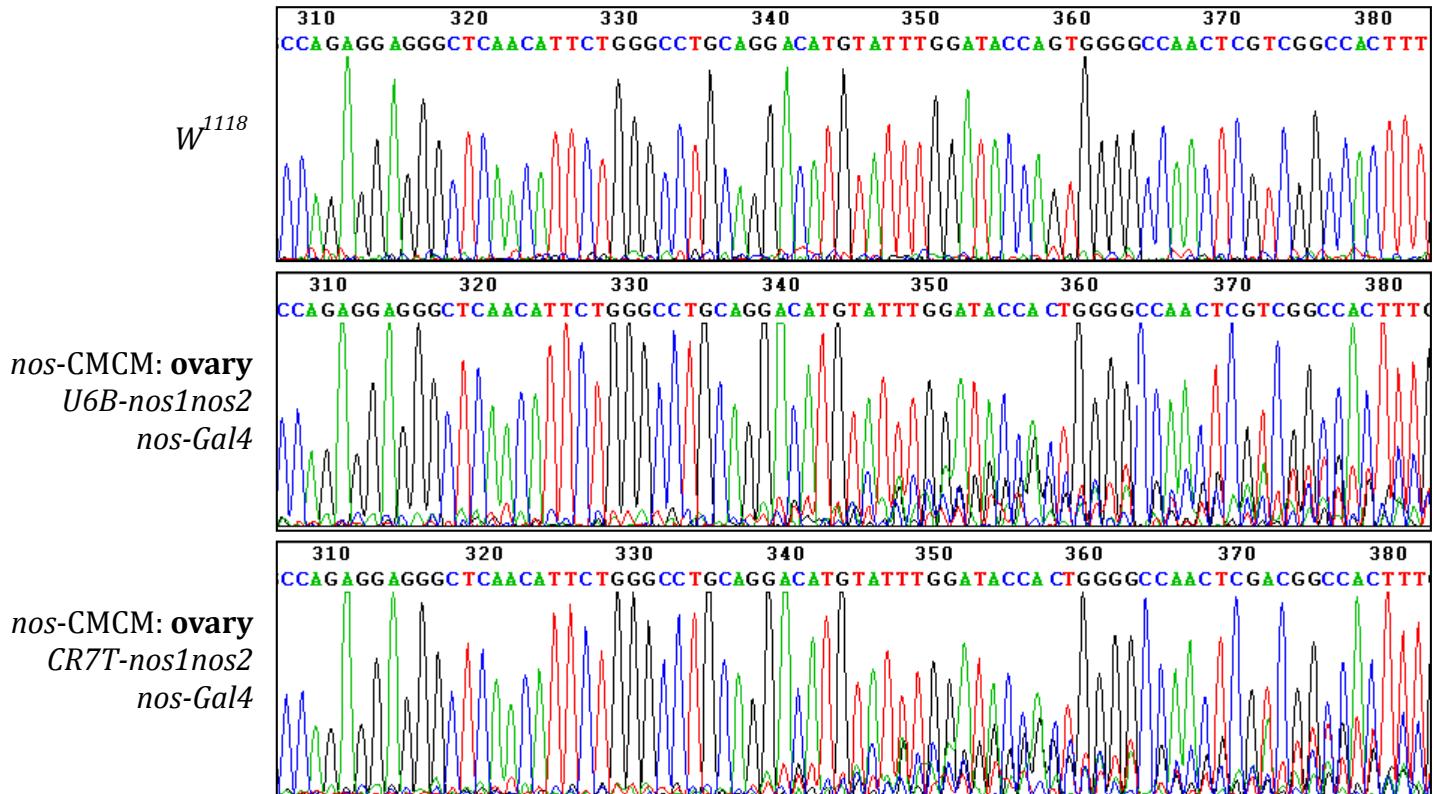


Figure S6 Sequence results for the ovaries of *nos* conditional mutant flies. (a) The sequence and a schematic representation of two gRNAs for the *nos* gene are shown. (B) *Nos-Gal4* was used to drive the expression of Cas9 specifically in the ovary, and two vectors, *U6B-nos1nos2* and *CR7T-nos1nos2*, were used to drive the expression of gRNA. The mutations were induced exactly at the target locus. A *w¹¹¹⁸* fly was used as the control.

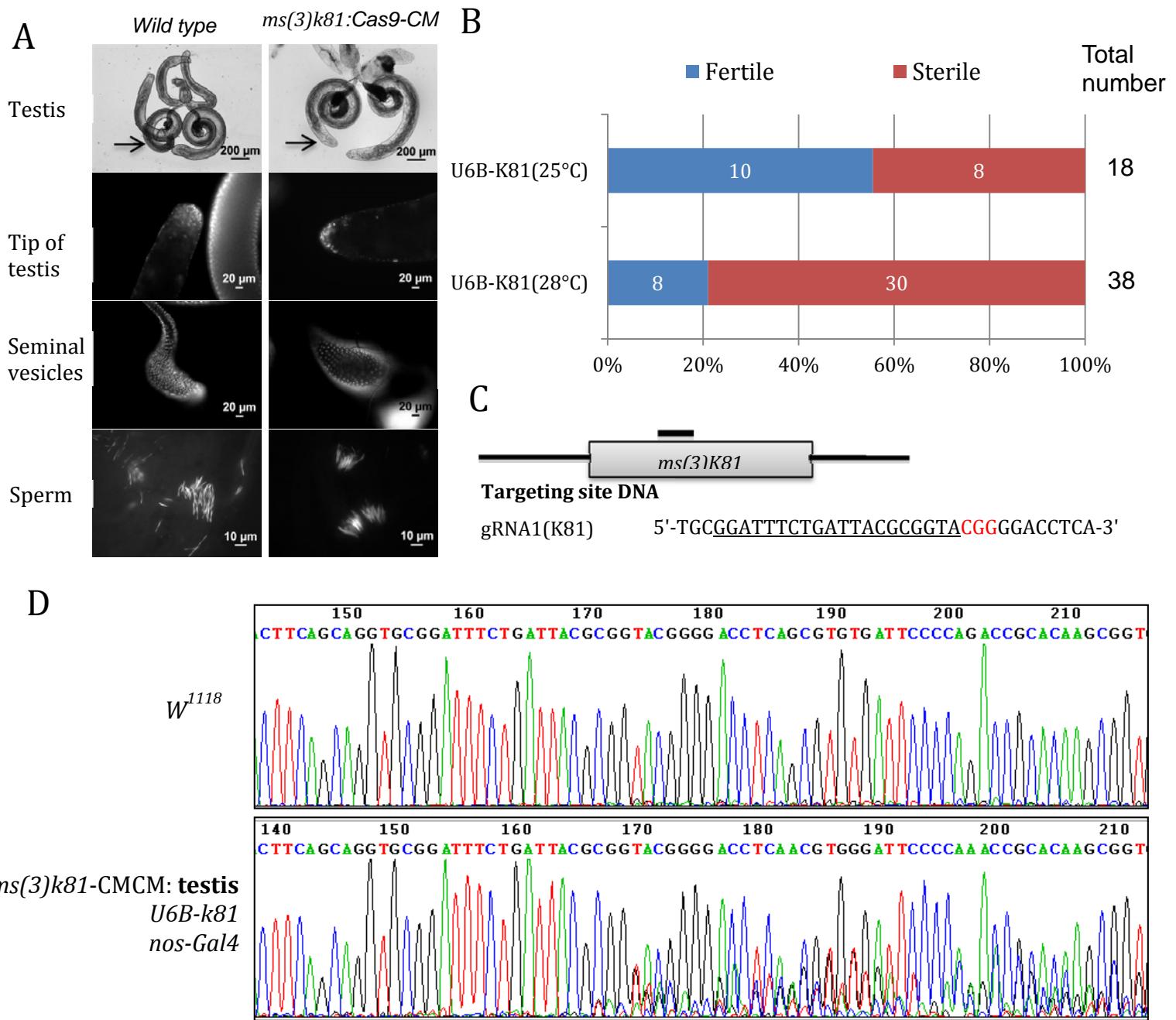


Figure S7 Conditional mutation of the *ms(3)k81* gene via the Cas9-mediated conditional mutagenesis (CMCM) system. (A) From the top to bottom of each column: whole testis (light), tip of the testis (DAPI), seminal vesicles (DAPI), and sperm detection (DAPI). *Nos-Gal4* was used to drive the expression of Cas9, and *U6B-K81* was used to drive the expression of gRNA. *ms(3)k81:Cas9-CM* was the testis from conditional mutant fly via the CMCM system. (B) Results of a fertility test are shown for flies for the CMCM system using the *ms(3)k81* gene. (C, D) The sequences and

a schematic representation of gRNAs against the *ms(3)k81* gene. Sequence results for the testes from the *ms(3)k81* conditional mutant flies. The mutations induced by CMC are located at the targeted locus.

>10UAS-HSP70 promoter

GGCTCGATCCGCTTGCATGCCTGCAGGTGGAGTACTGTCCTCCAGCGGAGTACTGTCCTCC
GAGCGGAGTACTGTCCTCCAGCGGAGTACTGTCCTCCAGCGGAGTACTGTCCTCCAGCGG
AAGCTTGATGCCTGCAGGTGGAGTACTGTCCTCCAGCGGAGTACTGTCCTCCAGCGGAG
TACTGTCCTCCAGCGGAGTACTGTCCTCCAGCGGAGTACTGTCCTCCAGCGGAGACTCTA
GCGAGCGCCGGAGTATAAATAGAGGCCTCGTCTACGGAGCACAATTCAATTAAACAG
CAAAGTGAACACCGTCTAACGAAAGCTAACGAAATAACAAGCGCAGCTGAACAAGCTA
AACAACTGCAGTAAAGTCAAGTTAAAGTGAATCAATTAAAGTAACCAGCAACCAAGTAA
ATCAACTGCAACTACTGAAATCTGCCAAGAAGTAATTATTGAATACAAGAAGAGAACTCTGA
ATAGGAATTGG

> α Tub84B 3'-UTR

CGGCCATCGAATTGAGCTCGCCCCTAACGCGTCGCCACTTCAACGCTCGATGGGAGCGTC
ATTGGTGGCGGGGTAACCGTCGAAATCAGTGTTCAGCTTCAATCGCAACAAAAATTCA
TGCAACACTGAAAAGCATACGAAAACGATGAAGATTGTACGAGAAACCATAAAGTATT
CCACAAAGACACGTATAGCAGAAAAGCCAAGTTAACCGCGATAAGTTGTACACAAGAA
TAAAATCGGCCAGATTCACTGTTGTACGAAATAAGAAAACCCACTATGTTTCTTGCCTT
TCTTCTCCAGCGATCATTCAATTCTGTTGAAAGAACGGGGTCATTGCACGGAGTTCGACT
GCGGGAAAGCAGAGCTGCCGTTCACTCGCTATAATTAGCGCTTCTATTCCCCGATTCG
GCCGCTGCTGCGCTTCCGCCTGCTGTTGGCAAGTGTAGCAGCAGGCTGTGCACGCA
GTGGCATGCACTGGCTTCCACCGCTGGTATCGATTCTCTGGGACGATGAGTCATTCTTC
GGGCCACAGCATATCGTGCAGCTACCGAAATGGTACTTCATTCTTAACTGCCGTCAA
GCATGCGATTGTACATACATATTTATATGTACATATTATGTGACTATGGTAGGTGCA
TATAATAGCAATCACGCAAGCAAATGTGTCAGTCCTGCTACAGGAACGATTCTATTAGTA
ATTTCTGTTGATAAAGTAATTATGTATGTATGAAAGCCCCATAAAATCTGAAACAATTAGGCA
AAACCATGCGAAGCTCTCA

Figure S8 Sequences of the 10UAS-HSP70 promoter and α Tub84B 3'-UTR used in this study.

Table S1 Sites targeted for each *Drosophila* gene and pRFP-gRNA constructions for the target loci.

Target gene		Target site (5' to 3') (PAM is underlined)	pRFP-gRNA	Number of gRNA
<i>yellow</i>	y1	GGATGAGTGTGGTCGGCTGT <u>GGG</u>	pRFP-U6B-y1-gRNA	1
	y2	GGGTTTGGACACTGGAACCGT <u>GG</u>	pRFP-CR7T-y2-gRNA pRFP-CR7T-y1y2-gRNA	1 2
<i>notch</i>	N1	GGACATTTGCCAGAAC <u>GGAGG</u>	pRFP-U6B-N1N2-gRNA	2
	N2	GGGATCACC <u>GTGCGAACACAATGG</u>	pRFP-CRT-N1N2-gRNA	2
<i>bag of marbles</i> (<i>bam</i>)	bam1	GGGCAACGACGACC <u>AGCAGTTGG</u>	pRFP-U6B-bam1-gRNA	1
	bam2	GGAGGAGCATTG <u>GGCCTTAATGG</u>	pRFP-CR7T-bam1-gRNA	1
	bam3	GGAAAGCC <u>ACTTGTGAGTACGAGG</u>	pRFP-CR7T-bam1bam2bam3-gRNA	3
<i>nanos</i>	nos1	GGGCCTGCAGGACAT <u>GTATTGG</u>	pRFP-U6B-nos1nos2-gRNA	2
	nos2	GGGGTCACCG <u>GGCGTAATGGCGG</u>	pRFP-CR7T-nos1nos2-gRNA	2
<i>cid</i>	cid1	GGACGCC <u>GGACGGAGGCAGCCGG</u>	pRFP-U6B-cid1cid2-gRNA	2
	cid2	GGAAAGCAA <u>ACGCGAGCAGCAGG</u>	pRFP-CR7T-cid1cid2-gRNA	2
<i>ms(3)K81</i>	K81	GGATTCTGATTAC <u>CGCGGTACGG</u>	pRFP-U6B-K81-gRNA	1

Table S2 List of primers used to construct the 10UAS-Cas9/TA-gRNA vector.

Plasmid	Primer name	Primer sequence (5' - 3') Forward and Reverse
piggyBac-10UAS-cas9	10UAS-XmaI-F	CCGGGGCTCGATCCGCTTGCATGC
	10UAS-NotI-R	GCGGCCGCCAATTCCCTATTAGAGTTCTCTTCTT
	αTub84B 3'UTR-XhoI-F	CTACTACTACTCGAGCGGCCATCGAATTGAGCTC
	αTub84B 3'UTR-SpeISalI-R	CATCATCATGTCGACACTAGTTAGAGAGCTCGATGGTTTGCC
TA-U6B/CR7T-gRNA -A	U6B-NotIspHIspEIspelFseI-F	GCGGCCGCATGCAGCTAGTGGCCGGCCGTTGACTTGCAGCCTGAAATAC
	CR7T-XhoIspelFseI-F	CTCGAGACTAGTGGCCGGCCGTTTGTATCGCTTTGTCG
	U6B/CR7T-AgeI/KpnI-R	GGTACCTGTTAAACTACCGTAAAAAAAGCACCAGACTCGGTGCCAC
TA-U6B/CR7T-gRNA -B	U6B-AgeI-F	CATACCGGTGTTGACTTGCAGCCTGAAATAC
	CR7T-AgeI-F	CATACCGGTGTTTGTATCGCTTTGTCG
	U6B/CR7T-KpnI-R	CATGGTACCAAAAAGCACCAGACTCGGTGCCAC
TA-U6B/CR7T-gRNA -C	U6B-Spel-F	CATACTAGT GTTCGACTTGCAGCCTGAAATA
	CR7T-Spel-F	CATACTAGTCGTTTGTATCGCTTTGTCG
	U6B/CR7T-Spel-R	CATACTAGTAAAAAAAGCACCAGACTCGGTGCCAC
TA-U6B/CR7T-gRNA -D	U6B-KpnI-F	CATGGTACCGTTCGACTTGCAGCCTGAAATA
	CR7T-KpnI-F	CATGGTACCCGTTTGTATCGCTTTGTCG
	U6B/CR7T-KpnI-R	CATGGTACCAAAAAGCACCAGACTCGGTGCCAC

Table S3 List of primers used to construct the transgenic gRNA vector.

Target locus	Primer name	Primer sequence (5' – 3') Forward and Reverse
<i>yellow</i>	U6B-yw-gRNA1-KOD-F	GGTCGGCTGTGTTTAGAGCTAGAAATAGCAAGTT
	U6B-yw-gRNA1-KOD-R	ACACTCATCCGAAGTATTGAGGAAAACATAACCTA
	CR7T-yw-gRNA1-KOD-F	GGTCGGCTGTGTTTAGAGCTAGAAATAGCAAGTT
	CR7T-yw-gRNA1-KOD-R	ACACTCATCCGAAGTCTTCCACTCATATACGCT
	CR7T-yw-gRNA2-KOD-F	ACTGGAACCGGTTTAGAGCTAGAAATAGCAAGTT
	CR7T-yw-gRNA2-KOD-R	GTCAAAACCCGAAAGTCTTCCACTCATATACGCT
<i>notch</i>	U6B-notch-gRNA1-KOD-R	CCGTTCTGGCAAAGATGTCCGAAGTATTGAGGAAAACATAACCTATA
	U6B-notch-gRNA2-KOD-R	TTGTGTTCGCACGGTGATCCGAAGTATTGAGGAAAACATAACCTATA
	CR7T-notch-gRNA1-KOD-R	CCGTTCTGGCAAAGATGTCCGAAGTCTTCCACTCATATACGCTA
	CR7T-notch-gRNA2-KOD-R	TTGTGTTCGCACGGTGATCCGAAGTCTTCCACTCATATACGCTA
<i>bag of marbles</i> (<i>bam</i>)	U6B-bam-gRNA1-KOD-R	ACTGCTGGTCGTCGTTGCCGAAGTATTGAGGAAAACATAACCTATA
	CR7T-bam-gRNA1-KOD-R	ACTGCTGGTCGTCGTTGCCGAAGTCTTCCACTCATATACGCTA
	CR7T-bam-gRNA2-KOD-R	TTAAGGCCAAATGCTCCTCGAAAGTCTTCCACTCATATACGCTA
	CR7T-bam-gRNA3-KOD-R	CGTACTCACAAAGTGGCTTCCGAAGTCTTCCACTCATATACGCTA
<i>nanos</i>	U6B-nos-gRNA1-KOD-R	AATACATGTCCTGCAGGCCGAAGTATTGAGGAAAACATAACCTATA
	U6B-nos-gRNA2-KOD-R	CCCATTACGCCGGTGACCCGAAGTATTGAGGAAAACATAACCTATA
	CR7T-nos-gRNA1-KOD-R	AATACATGTCCTGCAGGCCGAAGTCTTCCACTCATATACGCTA
	CR7T-nos-gRNA2-KOD-R	CCCATTACGCCGGTGACCCGAAGTCTTCCACTCATATACGCTA
<i>cid</i>	U6B-cid-gRNA1-KOD-R	GCTGCCTCCGTCGGCGTCCGAAGTATTGAGGAAAACATAACCTATA
	U6B-cid-gRNA2-KOD-R	GCTGCTCGCGTTTGCTTCCGAAGTATTGAGGAAAACATAACCTATA
	U6B-cid-gRNA3-KOD-R	AACGACGACGACACGCCCTTC GAAGTATTGAGGAAAACATAACCTATA
	U6B-cid-gRNA4-KOD-R	ACTACGGCCTCGAATTCAAC GAAGTATTGAGGAAAACATAACCTATA
<i>ms(3)k81</i>	U6B-K81-KOD-F	TTACCGGGTAGTTTAGAGCTAGAAATAGCAAGTT
	U6B-K81-KOD-R	TCAGAAATCCGAAGTATTGAGGAAAACATAACCTA
	gRNA-KOD-F	GTTTAGAGCTAGAAATAGCAAGTT

Table S4 List of primers used for PCR to verify the conditional mutations.

Target locus	Primer name	Primer sequence (5' – 3') Forward and Reverse
<i>yellow</i>	yellow-seq-F	CGGAGCTAATTCCGTATCCA
	yellow-seq-R	CGCCAGGTAGCTCGTATCTC
<i>notch</i>	Notch-seq-F	TGGAAGTGTGACCGTTTACCC
	Notch-seq-R	GTGGCAAGTCCATCGTTCAAGCA
<i>bag of marbles</i> <i>(bam)</i>	Bam-Seq-F	CAAAGAGTCTGGACGCCATCAT
	Bam-Seq-R	CGGTTCCACACATTTCTTCT
<i>nanos</i>	Nos-Seq-F	TTCGCAGTTGTTCAAGTTGTCTA
	Nos-Seq-R	ATCTCGTCCGTTGCTGGTGA
<i>ms(3)k81</i>	K81-seq-F	GAGATTCTCACTACTGCTCCTCG
	K81-seq-R	ACACGAATTGGATATGCGATAGC