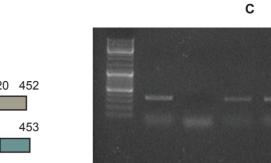
## Supplemental Materials Molecular Biology of the Cell

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Supplemental figure 1. ETV6-RUNX1 is recruited to the CDKN1A promoter. A. Schematic representation of ETV6 (TEL) and RUNX1 (AML1) proteins, and the fusion protein ETV6-RUNX1. Adapted from (Zelent et al. 2004) and UNIPROT entries P41212 and Q01196. B. ETV6-RUNX1 (E/R) knockdown in REH cells was performed by shRNA lentiviral transduction using the plasmid pLenti6/BLOCK-iT<sup>TM</sup>-DEST-shG1 and a non-targeting control vector expressing shRNA against LacZ (pLenti6/BLOCK-iT<sup>TM</sup>-DEST-shC) to generate REH-G1 and REH-C, respectively. Cells were selected for shRNA expression using blasticidin (5µg/ml) and knockdown of E/R monitored by Taqman RT-qPCR cells with at least 50% reduction were used in ChIP experiments. C. 10<sup>7</sup> REH-G1, REH-C cells were collected, DNA-protein cross-linked using formaldehyde and lysed using Diagenode lysis buffer iL1 and iL2. Chromatin DNA was sheared by sonication and ChIPs performed using anti-ETV6 (Santa Cruz N-19) or normal IgG as negative control, as indicated. 1% of input chromatin DNA was used as positive control. DNA was purified, quantified and an aliquot was used for PCR amplification using specific CDKN1A specific primers. PCR products were analyzed by agarose gel electrophoresis. **D.** Repression of CDKN1A promoter by ETV6-RUNX1. HEK293 cells were transfected with the reporter plasmid pCDKN1A-luc and indicated effector plasmids (pFlag-ETV6-RUNX1, pMX-MYC). Cells were treated with 100 µg/ml of PMA 12 hours post-transfection. Firefly luciferase data were normalized to renilla luciferase counts, and data are reported as mean and standard error of 3 independent experiments in triplicate.

**Supplemental figure 2. Involvment of** *TCF3-PBX1* **in RNA transport A.** Colocalization of NXF1 and TCF3-PBX1. HeLa cells were transfected with 1µg of pYFP-NXF1 and/or pFlag-TCF3-PBX1 plasmids using lipofectamine. Twenty-four hours post-transfection, cells were fixed in formaldehyde, permeabilized, incubated with anti-flag M2 antibody followed by Alexa 568 secondary antibody (Red). Cells were mounted with the Prolong gold Antifade reagent with DAPI (Blue) and the slides were analyzed by confocal microscopy using the Nikon A1R system and images processed with the IMARIS software. **B.** expression of *TCF3-PBX1* affects RNA localization. HeLa cells were fixed in formaldehyde, permeabilized, incubated with anti-flag M2 antibody followed by Alexa 568 secondary antibilized, incubated with anti-flag M2 antibody followed by Alexa 568 secondary antibilized, incubated with anti-flag M2 antibody followed by Alexa 568 secondary antibilized, incubated with anti-flag M2 antibody followed by Alexa 568 secondary antibody (Red). Cells were stained with the SYTO RNA marker (Green) and mounted with the Prolong gold Antifade reagent with DAPI DNA marker (Blue) and the slides were analyzed by confocal microscopy using the Nikon A1R system and images processed with the IMARIS software.

Supplemental table 1 Perturbed interactions in ETV6-RUNX1 samples Supplemental table 2 Perturbed interctions in TCF3-PBX1 samples Supplemental table 3 Perturbed interctions in BCR-ABL1 samples Supplemental table 4 Perturbed interctions confirmed by ChIP-seq Supplemental table 5 Relative expression of cell cycle genes interacting with MYC Supplemental table 6 Deregulated pathways in ETV6-RUNX1 samples Supplemental table 7 Deregulated pathways in TCF3-PBX1 samples Supplemental table 8 Perturbed interactions in TCF3-PBX1 involved in RNA processing Supplemental table 9 Similar target genes for MYC and ETV6-RUNX1

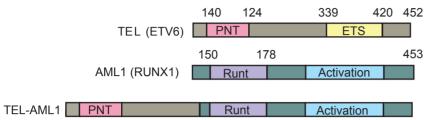


Μ

1% input

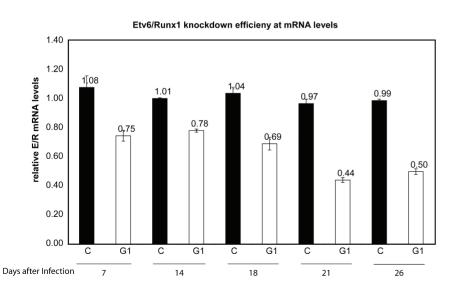
lg control

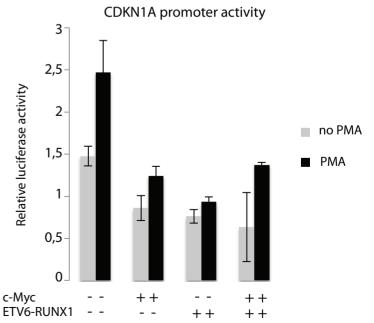
REH control



Α

В





1% input

lg control

REH G1

Anti- ETV6

Anti-ETV6

D

CDKN1A

## Figure 2

A	YFP	antiFlag+ Alexa568	DAPI	MERGE
YFP-NXF1				<u>10 µт</u>
Flag-TCF3-PBX1		9		
YFP-NFX1 + Flag-TCF3-PBX1				
В				
	Syto-RNAselect	antiFlag+ Alexa568	DAPI	MERGE
Control	10			12
Flag- TCF3-PBX1				<u>10 µт</u>