**Supplementary Figure 1:** A metabolic map illustrating metabolic modules that are involved in methanol growth in *M. extorquens* PA1. Methanol dehydrogenase serves as the primary  $C_1$  oxidation module that oxidizes methanol to formaldehyde. Formaldehyde is further oxidized to formate by a H4MPT mediated pathway which acts as the formaldehyde oxidation module. Formate is the branch point for metabolism - a part gets oxidized to  $CO<sub>2</sub>$  by formate dehydrogenases and the rest gets reduced and assimilated via a  $H_4F$  mediated pathway and serine cycle. Genes highlighted in red, representative of a particular module, were deleted to test the role of the specific module in methylamine growth via the *N-*methylglutamate pathway.



**Supplementary Figure 2:** A metabolic map illustrating the various metabolic modules that are involved in chloromethane growth in  $M$ . *extorquens* CM4. The primary  $C_1$  oxidation module consists of CmuA and CmuB*-* that catalyze dehalogenation of chloromethane and subsequent methyl transfer to tetrahydrofolate  $(H_4F)$  to form methyl tetrahydrofolate  $(CH_3-H_4F)$  - and MetF that catalyzes the reduction of  $CH_3$ - $H_4$ F to methylene tetrahydrofolate (CH<sub>2</sub>=H<sub>4</sub>F). CH<sub>2</sub>=H<sub>4</sub>F is the branch point of metabolism –a part gets oxidized by methylene tetrahydrofolate reductase (*folD*) and formyl tetrahydrofolate hydrolase ( $purU$ ) to formate, which subsequent gets oxidized to  $CO<sub>2</sub>$  by a panel of formate dehydrogenases, and the rest gets assimilated via the serine cycle. **\****metF, folD, and purU* are absent in *M. extorquens* PA1



**Supplementary Figure 3: A)** Ratio of growth rates, of various knockout mutants versus the ∆*cel* 'wildtype' strain of PA1 and, in 3.5 mM succinate (white), 15mM methanol (light brown),nitrogen free media with 3.5 mM succinate and 7.66 mM methylamine (dark gray) and 15mM methylamine (blue) **B)** Ratio of yield (measured as the maximum OD<sup>600</sup> value during growth) of various knockout mutants versus ∆*cel* 'wildtype' strain of PA1, in 3.5 mM succinate (white), 15mM methanol (light brown), nitrogen free media with 3.5 mM succinate and 7.66 mM methylamine (dark gray) and 15mM methylamine (blue). Error bars represent the 95% C.I. of the average ratio of three biological replicates grown in each condition. A X indicates that the mutants did not grow in methanol. **NOTE: The y-axis doesn't start at 0.0 but at 0.5 (for growth rate ratios) and 0.4 (for yield ratios). This was done to highlight subtle changes in growth/yield for these mutants**





∆*fae3*

**Supplementary Figure 4:** Amino acid alignment of FAE, FAE2 and FAE3 from *M. extroquens* PA1 (using MUSCLE: <https://www.ebi.ac.uk/Tools/msa/muscle/> with standard settings and 150 iterations). The highlighted residues in FAE interact with methyl groups present in tetrahydromethanopterin  $(H<sub>a</sub>MPT)$ , absent in H<sub>4</sub>F. These interactions are responsible for the specificity of FAE to H<sub>4</sub>MPT. Two of these residues are identical and one residue (Y173) has similar properties in FAE2. Only one residue is identical in FAE3.



**Supplementary Table 1:** Growth rates of the ∆*cel* 'wildtype' strain of PA1 and the ∆*mptG* mutant (in the WT background) in 3.5 mM succinate with either 7.66 mM  $NH_4^+$  as the nitrogen source or 7.66 mM methylamine as the nitrogen source. S.E.M. represents the standard error of the mean growth rate for three biological replicates in each condition.



**Supplementary Table 2:** Growth rates and maximum OD<sub>600</sub> of the ∆*cel* 'wildtype' strain of PA1 and single-, double-, triple-knockout mutant (in the WT background) lacking ∆*fae,* ∆*fae2, and/or* ∆*fae3* in 15mM methylamine



**Supplementary Table 3:** List of primers used to construct knockout mutants in the *N-*methylglutamate pathway as well as FAE-homologs in this study

