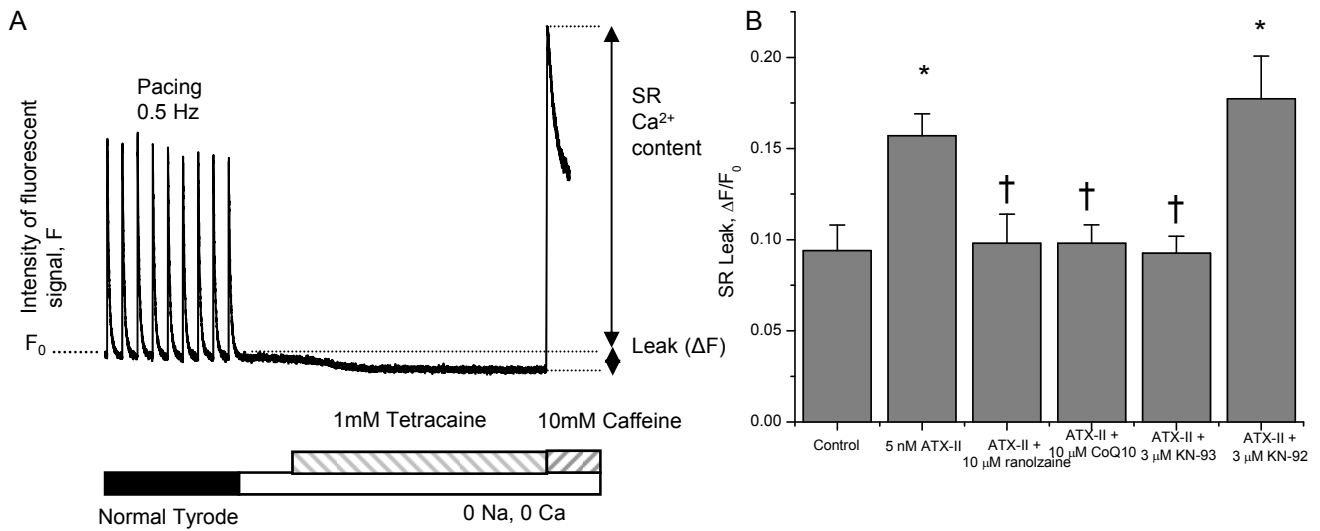
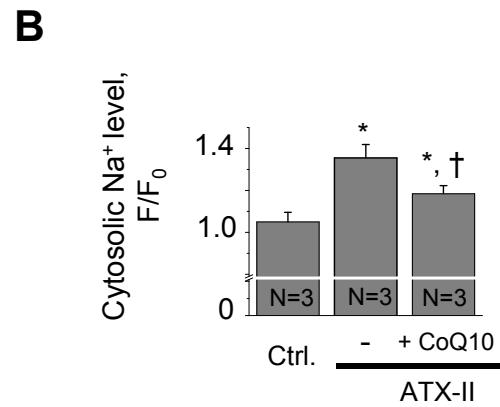
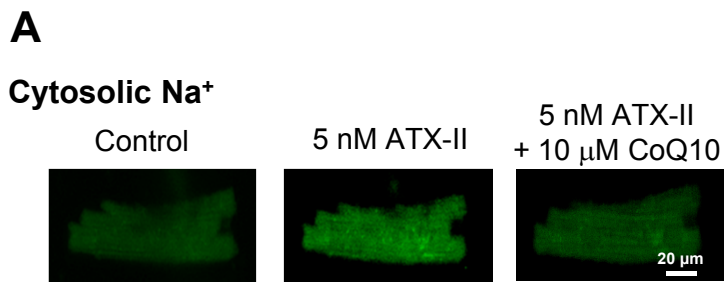


## Supplemental Figures and Table

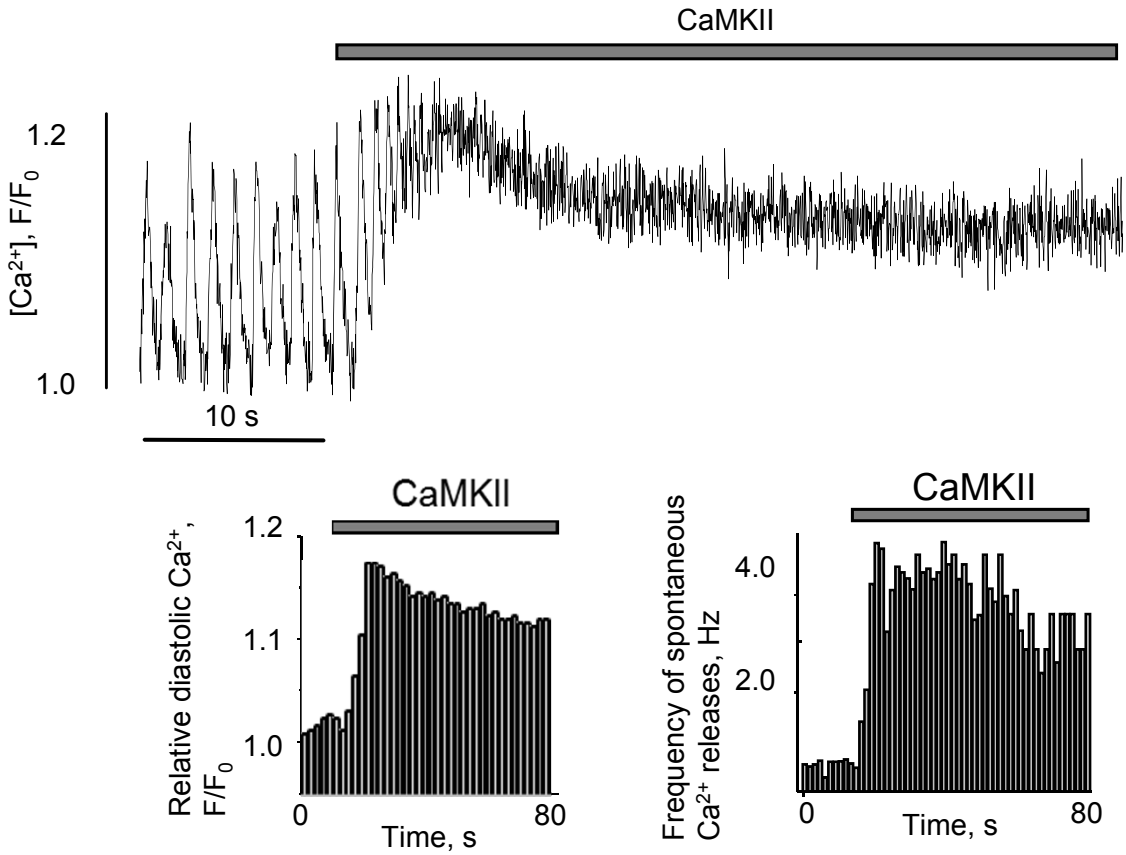


**Supplementary Figure 1.** ATX-II induced diastolic  $Ca^{2+}$  leak in rabbit ventricular myocytes. A: Scheme of protocol used to estimate diastolic SR  $Ca^{2+}$  leak. B: Pooled data on SR  $Ca^{2+}$  leak in control cells, after AXT-II application alone and after application of ATX-II + CoQ10, ATX-II + KN-93 and ATX-II + KN-92. N=5-13, \*  $P < 0.05$  vs control; † $P < 0.05$  vs ATX-II.

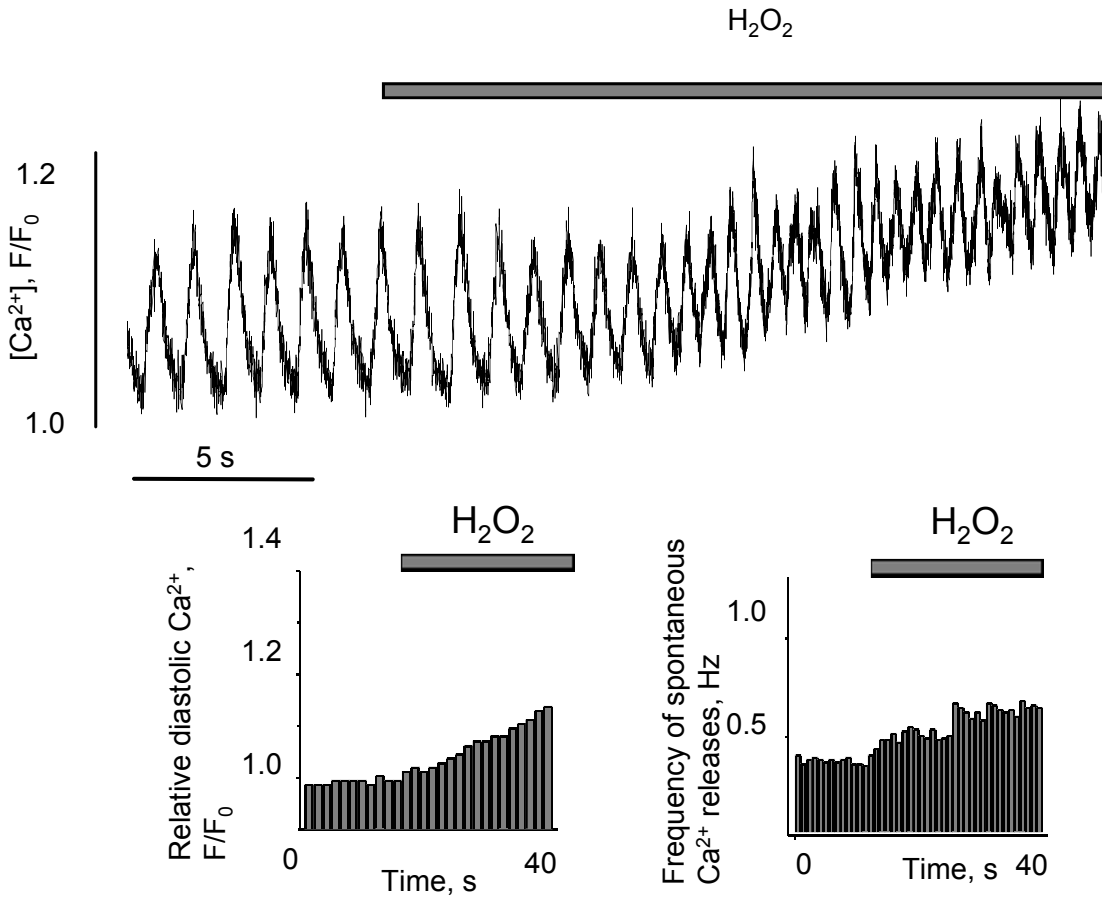


**Supplementary Figure 2.** Antioxidant CoQ10 reduced Na<sup>+</sup><sub>i</sub> overload induces by ATX-II.

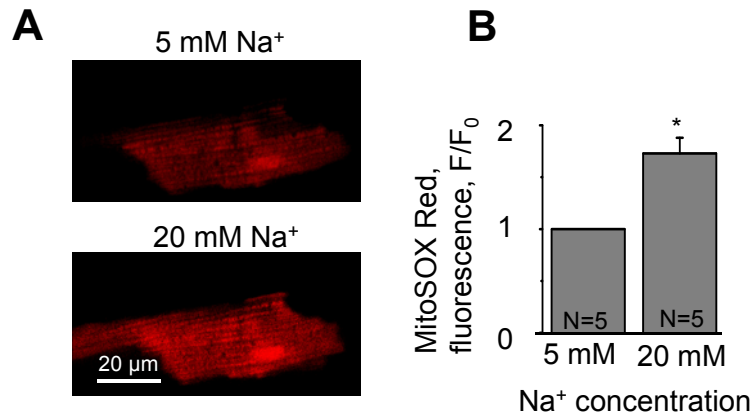
**A.** Confocal images of a rabbit ventricular myocyte paced at 1 Hz and loaded with Na<sup>+</sup> dye Asante NaTRIUM Green AM. Application of ATX-II (5 nM) for 2 min increased cytosolic Na<sup>+</sup> compared to control (Ctrl), which was reduced by CoQ10 (10 μM). **B.** Quantification of results for ATX-II-induced changes cytosolic Na<sup>+</sup>, in the absence and presence of CoQ10 (10 μM). Data are normalized to F<sub>0</sub> - the baseline fluorescence signal at the beginning of an experiment. N=3, \* p<0.05 vs control; † P<0.05 vs ATX-II.



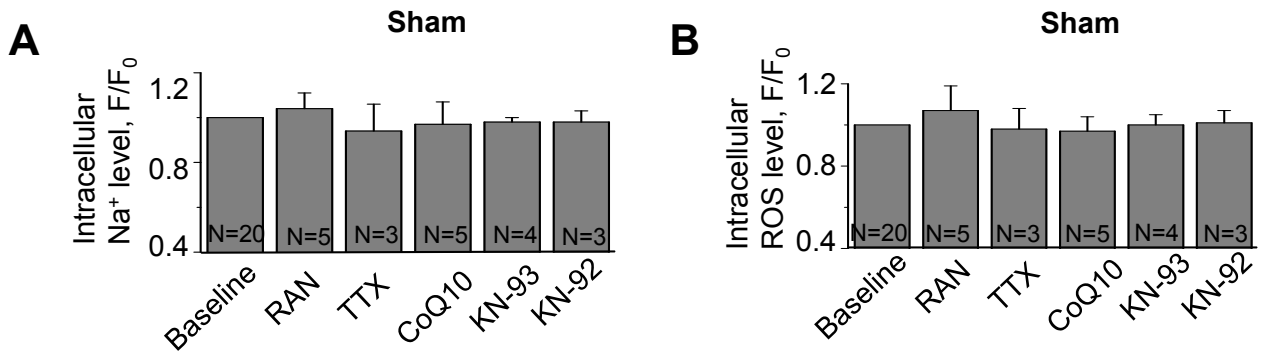
**Supplementary Figure 3.** Effect of exogenous pre-activated CaMKII (2 units activity/L) on spontaneous  $\text{Ca}^{2+}$  waves in membrane-permeabilized rabbit ventricular myocytes. Cytosolic  $\text{Na}^+$  concentration was 5 mM.



**Supplementary Figure 4.** Effect of  $\text{H}_2\text{O}_2$  (100  $\mu\text{M}$ ) on spontaneous  $\text{Ca}^{2+}$  waves in membrane-permeabilized rabbit ventricular myocytes. Cytosolic  $\text{Na}^+$  concentration was 5 mM.



**Supplementary Figure 5.** Elevated [Na<sup>+</sup>]<sub>i</sub> increases mitochondrial ROS in membrane-permeabilized cardiomyocytes. A. Mitochondrial ROS level was assessed using MitoSOX Red in permeabilized rabbit ventricular myocytes subjected to 5 mM and 20 mM Na<sup>+</sup>. B. High [Na<sup>+</sup>]<sub>i</sub> resulted in an increase in mitochondrial ROS generation. N=5, \* p<0.05 (20 mM Na<sup>+</sup> vs. 5mM Na<sup>+</sup>).



**Supplementary Figure 6.** Effects of ranolazine (10  $\mu$ M), TTX (1  $\mu$ M), CoQ10 (10  $\mu$ M), KN-93 (3  $\mu$ M), and KN-92 (3  $\mu$ M) on levels of intracellular Na<sup>+</sup> (A) and ROS (B) in myocytes isolated from sham mice and paced at a rate of 1 Hz (N = 3-20, p<0.05 compared to baseline).

	Ca <sup>2+</sup> signal amplitude, F <sub>max</sub> /F <sub>0</sub>	Rise time of Ca <sup>2+</sup> transients (ms)	τ decay Ca <sup>2+</sup> transients (ms)	Number of experiments, N
Sham	2.6 ± 0.3	28 ± 5	183 ± 11	7
TAC	2.5 ± 0.5	39 ± 4 *	201 ± 17	6
TAC + 3 μmol/L KN93	2.4 ± 0.4	27 ± 7	188 ± 11	3

**Supplementary Table 1.** Parameters of pacing-induced Ca<sup>2+</sup> transients in TAC ventricular myocytes. F<sub>0</sub> – baseline fluorescence, F<sub>max</sub> – level of fluorescence at the peak of Ca<sup>2+</sup> transient.\* p<0.05 vs sham.