	Cysteine desulphurase		Scaffold		ATP-hydrolysing component		Trafficking	
	E. coli	C. <i>jejuni</i> NCTC 11168	E. coli/ H.pylori	<i>C. jejuni</i> NCTC 11168	E. coli	<i>C. jejuni</i> NCTC 11168	E. coli	<i>C. jejuni</i> NCTC 11168
ISC/NIF	lscS	Cj0240 (39%)	IscU (<i>Ec</i>) NifU (<i>Hp</i>)	Cj0239 (40%) Cj0239 (69%)	HscA HscB	DnaK (41%) -	lscA	-
SUF	SufS SufE	Cj0240 (27%) -	SufB SufD	-	SufC	LivG (26%) Cj0669 (28%) Cj1663 (27%)	SufA	-
Other	CsdA	Cj0240 (27%) Cj0791 (22%)					Mrp NfuA ErpA	Cj1606 (37%) Cj1639 (32%) -

Supplementary Table S1. Comparison between the components of the *E. coli* and *C. jejuni* NCTC 11168 Fe-S cluster biosynthesis pathways. Protein function is listed along the top row, and the specific pathways that the listed proteins belong to are in the right-hand column (other referring to those proteins not experimentally linked to any specific pathway) (Py & Barras, 2010). *C. jejuni* homologues to *E. coli* proteins are shown in bold with percentage sequence identity in brackets. Note that Cj0239 is homologous with the characterised NifU protein in *H. pylori* (Hp) rather than IscU, so this comparison is also shown. Proteins without homologues in *C. jejuni* are indicated by a – sign.

HerA	/	/	/	/	/	
11168	-MTYNEKIISMNNDLLDHQ <mark>H</mark> KELFEIS	KKLSLMNQRHVGTKELKIVLF	ELLIMINR <mark>HFSDE</mark> EAFMREI	LEYPYINH <mark>HTRIH</mark> RKII <u>L</u> EIE	EI <mark>II</mark> SEAKFVNIMTEKLNL <mark>V</mark>	VQDFIFK
81116	-MTYNEKIISMNNDLLDHQ <mark>H</mark> KELFEIS	KKLSLMNQRHVGTKELKIVLF	RELLIMINR <mark>HFSDE</mark> EAFMREI	LEYPYINH <mark>HTRIH</mark> RKII <mark>L</mark> EIE	EI <mark>II</mark> SEAKFVNIMTEKLNL <mark>V</mark>	VQDFIFK
81-176	-MTYNEKIISMNNDLLDHQ <mark>H</mark> KE <mark>L</mark> FEIS	KKLSLMNQRHVGTKELKIVLF	RELLIMINR <mark>H<u>F</u>SDE</mark> EAFMREI	IGYPYINH <mark>HTRIH</mark> RKII <mark>L</mark> EIE	EI <mark>II</mark> SEAKFVNIMTEKLNL <u>V</u>	VQDFIFK
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HerB	/	/	/	/	/	
11168	MLPKWDNSYSVHNAKIDEQ <mark>H</mark> KK <mark>L</mark> FKLA	AKVEVVSDRSVSKNEVKELLA	\EFFNYMKD <mark>H<u>F</u>NDE</mark> EKYMQLI	IGYPNLEE <mark>HRKIH</mark> KEI <mark>I</mark> QTMI	N <mark>LI</mark> K-DIKSTNDLKEKLY <mark>I</mark> V	AKKWLLE
81116	MLPKWDNSYSVHNAKIDEQ <mark>H</mark> KK <mark>L</mark> FELA	AKVEVVSDRSVSKNEVKELLA	\EFFNYMKD <mark>H<u>F</u>NDE</mark> EKYMQL]	IGYPNLEE <mark>HRKIH</mark> KEI <u>I</u> QTMI	N <mark>LI</mark> K-DIKSTNDLKEKLY <mark>I</mark> V	AKKWLLE
81-176	MLPKWDNSYSVHNAKIDEQ <mark>H</mark> KK <mark>L</mark> FKLA	AKVEVVSDRSVSKNEVKELLA	EFFNYMKD <mark>H<u>F</u>NDE</mark> EKYMQLI	IGYPNLEE <mark>HRKIH</mark> KEI <u>I</u> QTMI	N <mark>LI</mark> K-DIKSTNDLKEKLY <mark>I</mark> V	AKKWLLE
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FedA	/	/	/	/	/	
11168	MKVKWSRDFSIKNMQLDKQ <mark>H</mark> ELIFEITNLA	NDLALNIQDNNTQHKNDLKQILV	KLFQYIKI <mark>H<u>F</u>KDE</mark> EKFMESI	IDFPLIEE <mark>HKKSH</mark> QI <mark>L</mark> VEKTK	E <mark>LL</mark> E-HSNDIVKMSQELS <mark>I</mark> L	TKDWILD
81116	MKVKWSRDFSIKNMQLDKQ <mark>H</mark> ELIFEITNLA	NDLALKIQENNTQYKDDLKQILA	KLFQYIKI <mark>H<u>F</u>KDE</mark> EKFMESI	IDFPLIEE <mark>HKKSH</mark> QI <mark>L</mark> VEKTK	E <mark>LL</mark> E-HSDNIVKMSQELS <mark>I</mark> L	TKDWILD
81-176	MEVKWSRDFSIKNMQLDKQ <mark>H</mark> ELIFEITNLA	NDLALNIQDNNTQHKNDLKQILV	KLFQYIKI <mark>H<u>F</u>KDE</mark> EKFMESI	IDFPLIEE <mark>HKKSH</mark> QI <mark>L</mark> VEKTK	E <mark>LL</mark> E-HSDNIVKMSQELS <mark>I</mark> L	TKDWILD
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HerA	/					
11168	<mark>HTAKED</mark> SKIVKYYEEKFKK 133					
81116	<mark>HTAKED</mark> SKIVKYYEEKFKK 133					
81-17	<mark>HTAKED</mark> SKIVKYYEEKFKK 133					

HerB	/	/	· /	/		
11168	HILYEDMKVEKWRSSSLSTDDGGDVSF	EAAEDEDNEHPQFYLYTCN	ICPGKIHDVPYSIHQKIELQO	GRKFTCKTCKQAIKFYKKYS	199	
81116	HILYEDMKVEKWRSSSLSTDDGGDVSF	EAAEDEDNEHPQFYLYTCN	ICPGKIHDVPYSIHQKIELQO	GHKFTCKTCKQAIKFYKKHS	199	
81-176	HILYEDMKVEKWRSSSLSTDDGGDISF	EAAEDEDNEHPQFYLYTCN	CPGKIHDVPYSIHQKIELQC	GHKFT <mark>CKTC</mark> KQAIKFYKKYS	199	
	**************************************	***** **********	*****	* • * * * * * * * * * * * * * * * * * *		
FedA	/ /	/	/	/	/	/
11168	<mark>HFANED</mark> LWIANFTKKTLHLQEIHYTLE	QYIKLKSIKQDLRAEKTYDYICN	ICSLRIHAVPQTIHQELVSKE	ENTLKCEKCGQILVHLDYFDL	NQNFEKFNAIFEDALQNHHF	TTQENDMRGGG 240
81116	HFANEDLWIANFTKKTLHLQEIHYTLE	QYIKLKSIKQDLRAEKTHDYICN	ICSLRIHAVPQTIHQELVSKE	ENTLKCEKCGQILVHLDYFDL	NQNFEKFNAIFEDALQNHHF	TTQKNDMGGG- 239
81-176	<mark>HFANED</mark> LWIANFTKKPYTFK	KSIIP				145
FedA*		KSIKQDLRAEKTHDYICN	<i>ICYLRTHAVPQTIHEELVSKI</i>	ENTLK <mark>CEKC</mark> GQILVHLDYFDL	<i>NQNFEKFNAIFEDALQNHHF</i>	TTQKNDMGGG- 88
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Supplementary Figure 1: Protein sequence alignments of hemerythrins in *C. jejuni* NCTC 11168 (top line), *C. jejuni* 81116 (middle line) and *C. jejuni* 81-176 (bottom line). HerA = Cj0241, HerB = Cj1224 and FedA = Cj0045 in strain NCTC 11168. Conserved motifs characteristic of hemerythrin proteins are highlighted in yellow, hydrophobic residues potentially lining the oxygen binding pocket are in blue and underlined (French *et al.*, 2008). Potential metal binding CXC and CXXC motifs are highlighted grey. Amino-acid similarity is indicated underneath the alignment: * = fully conserved residue, : = residues sharing strongly similar properties, . = residues sharing weakly similar properties. Sequence length is indicated above each protein; / = 20 residues. *FedA** indicates the missing C-terminal sequence of the 81-176 gene is encoded in a different reading frame.



Supplementary Figure 2. *In vivo* recovery of Por and Oor activity after partial (A, B) or complete (C, D) aerobic inactivation. Assays of Por (A, C) and Oor (B, D) activity in CFE prepared from wild-type (closed circles) and *herA* mutant (open circles) cells incubated aerobically (500 ml cultures in 2.5 L baffled flasks with 250 rpm shaking in air) for 4.5 hours (A, B) or 16 hours (C, D), followed by anaerobic incubation in the presence of chloramphenicol. Dashed vertical lines indicate the point of transfer from aerobic conditions to the anaerobic recovery conditions. All activities are expressed as a percentage of the initial pre-stress activity (mean initial values in µmol min⁻¹ mg protein⁻¹ A: wt = 2.4, *herA* = 2.3; B: wt = 0.61, *herA* = 0.61; C: wt = 1.0, *herA* = 0.9; D: wt = 0.18, *herA* = 0.17). The data shown in A and B are from four independent experiments, with error bars showing standard deviation. The data shown in C and D is from a single experiment which was repeated twice with similar results.



Supplementary Figure 3. (A) *Inset*: 12% SDS-PAGE gel showing purified NCTC 11168 HerB (Cj1224) protein band of ~23 kDa (green arrow). Sizes of marker bands are also shown. *Main figure*: Absorption spectra of the purified as-isolated HerB (green spectrum) and HerB reduced with sodium dithionite (blue spectrum). (B) Absorption spectra of purified as-isolated HerB (green spectrum) and HerB plus 50 mM sodium azide (orange spectrum). (C) *Inset*: 12% SDS-PAGE gel showing purified 81-176 FedA protein band (green arrow). *Main figure*: Absorption spectra of as-isolated FedA purified from cells grown in media supplemented with ferrous ammonium sulphate (green spectrum), FedA reduced with sodium dithionite (blue spectrum) and FedA purified from cells grown with no additional ferrous ions in the medium (grey line). (D) UV absorption spectra of purified iron-loaded FedA (green spectrum) and FedA with 50 mM sodium azide (orange spectrum).