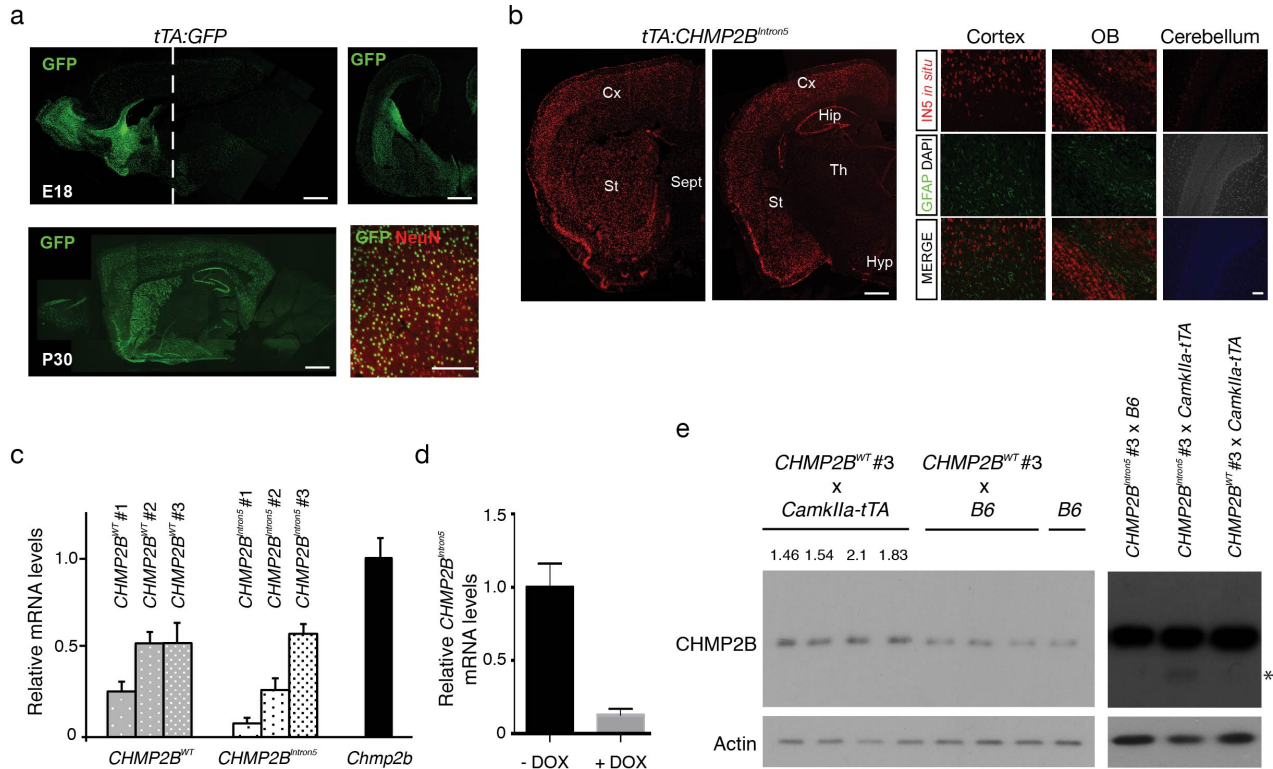


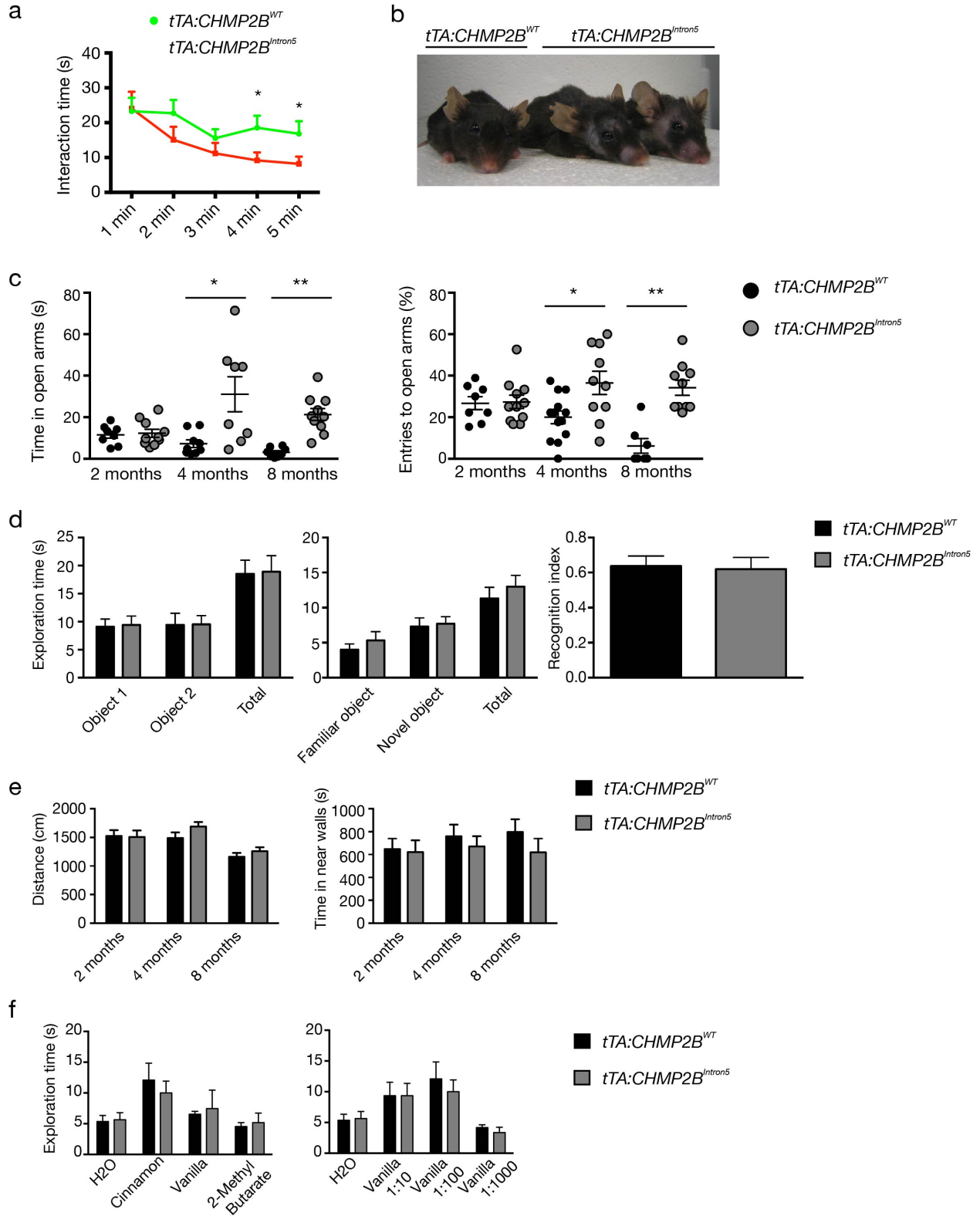
Alterations in microRNA-124 and AMPA receptors contribute to social behavioral deficits in frontotemporal dementia

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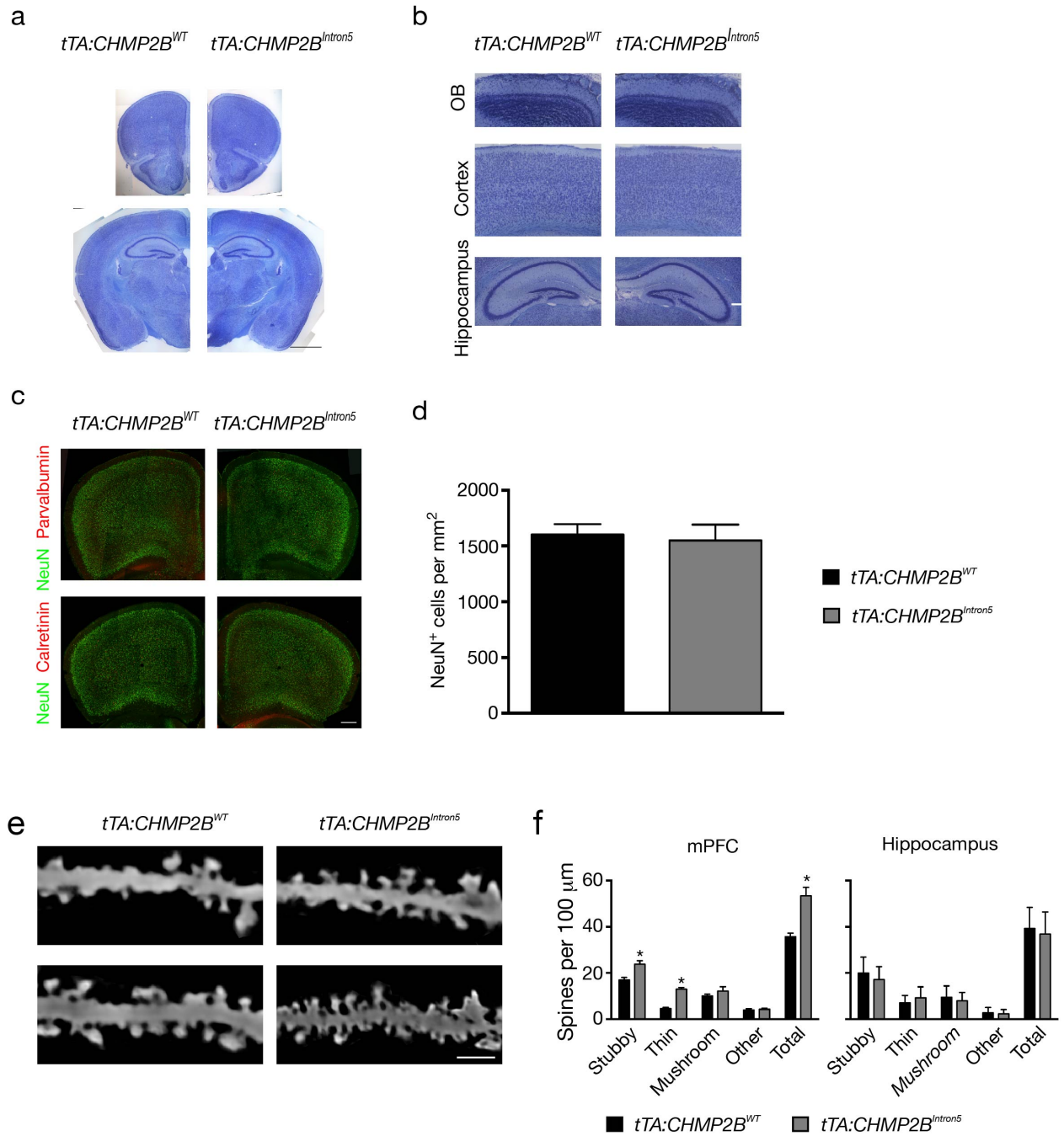


Supplementary Figure 1. Basic characterization of transgenic *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice. **(a)** GFP expression driven by *CamkIIα-tTA*. Upper panels show GFP expression in a sagittal (left) or coronal (right) brain section at E18. In the lower panel (left), a sagittal section at P30 revealed widespread expression in the cortex, striatum, olfactory bulb (OB) and hippocampus but low or no expression in other brain areas such as cerebellum or thalamus. The right lower panel shows all GFP-positive cells are neurons in the cortex of P30 mice. **(b)** Left panels show transgene expression in coronal sections of *tTA:CHMP2B^{Intron5}* mice at P30. Right panels show the absence of transgene expression in astrocytes in the cortex or olfactory bulb (OB) of *tTA:CHMP2B^{Intron5}* mice. **(c)** mRNA levels of *CHMP2B^{WT}* or *CHMP2B^{Intron5}* in the cortex of the different lines of transgenic mice (normalized to β-actin). We plotted the level of the mouse endogenous *Chmp2b* gene as reference. **(d)** *CHMP2B^{Intron5}* mRNA

levels in the cortex of mice fed a regular diet (-Dox) or a diet containing doxycycline (+Dox) for 8 months (normalized to β -actin). **(e)** Confirmation of transgene expression at the protein level by Western blot. The expression of transgenic CHMP2B^{WT} (observed as an increase in the endogenous band since mChmp2b and hCHMP2B are identical except for one amino acid) and CHMP2B^{Intron5} (observed as a lower band, asterisk). Since the antibody is directed against the C-terminal part of the protein (truncated in CHMP2B^{Intron5}), it could not be used for quantification. B6: C57BL/6. Scale bar, 200 μ m in **a**, 200 μ m (left panels) and 50 μ m (right panels) in **b**.

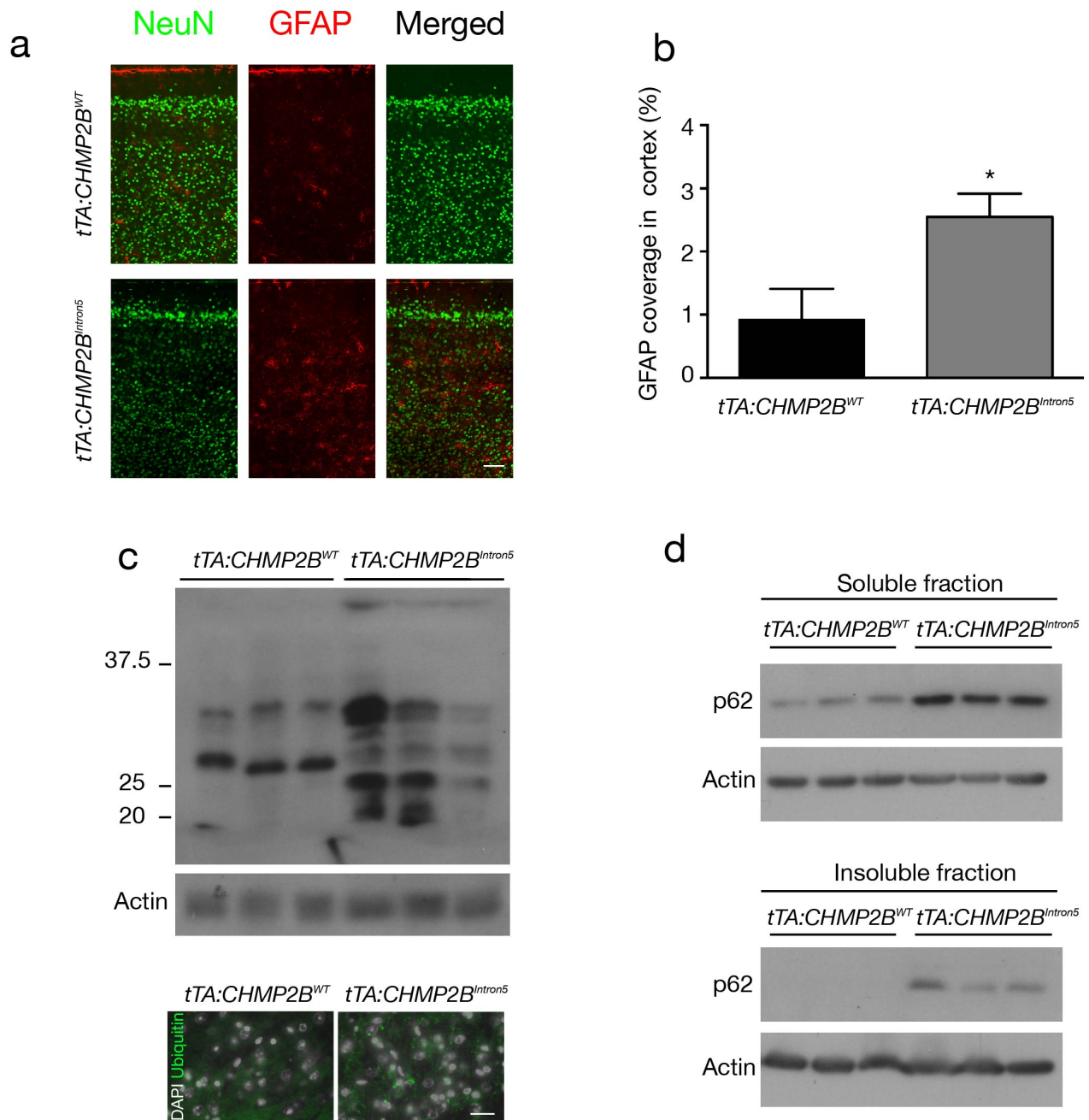


Supplementary Figure 2. Further behavioral characterization of transgenic $tTA:CHMP2B^{WT}$ and $tTA:CHMP2B^{Intron5}$ mice. **(a)** Interaction in home cage task in 8-month-old $tTA:CHMP2B^{WT}$ and $tTA:CHMP2B^{Intron5}$ mice. The difference in interaction time (defined as the time the test mouse explores the intruder mouse in 1-min intervals) is significant at the end of the task (4 min and 5 min time points. $n = 11$ mice per genotype; $P < 0.05$ at 4 min, $P < 0.05$ at 5 min by two-sided t test). **(b)** Cutaneous lesions consistent with excessive grooming are observed in a subset of individually caged $tTA:CHMP2B^{Intron5}$ (two on the right) but not in $tTA:CHMP2B^{WT}$ (first on the left) at old ages (14–20 months). **(c)** Time spent (left graph) and number of entries into the open arms (right graph) of the elevated plus maze (for $tTA:CHMP2B^{WT}$, $n = 8$ mice at all time points; for $tTA:CHMP2B^{Intron5}$, $n = 10$ at 2 and 8 months and $n = 8$ at 4 months; time spent in open arms: $P < 0.01$ at 4 months, $P < 0.001$ at 8 months by two-sided t test; entries to open arms: $P < 0.02$ at 4 months, $P < 0.001$ at 8 months by two-sided t test). **(d)** Quantification of the time $tTA:CHMP2B^{WT}$ and $tTA:CHMP2B^{Intron5}$ mice at 8 months exploring the different objects presented in the novel object recognition task during the familiarization phase (left) or the test phase (middle) ($n = 10$ for $tTA:CHMP2B^{WT}$ mice and $n = 11$ $tTA:CHMP2B^{Intron5}$ mice, $P > 0.5$ by two-way ANOVA). Proportion of time exploring the novel object in the test phase (recognition index) is showed in the right panel. **(e)** Quantification of the distance traveled (left panel) or the time spent close to the arena walls (thigmotaxis, right panel) in the open-field task ($n = 11$ mice per genotype, $P > 0.2$ by two-sided t test). **(f)** Analysis of olfactory discrimination (left) and sensitivity (right) in $tTA:CHMP2B^{WT}$ and $tTA:CHMP2B^{Intron5}$ mice at 8 months ($n = 10$ mice per genotype; $P > 0.4$ by two-sided t test). All values are mean \pm s.e.m.



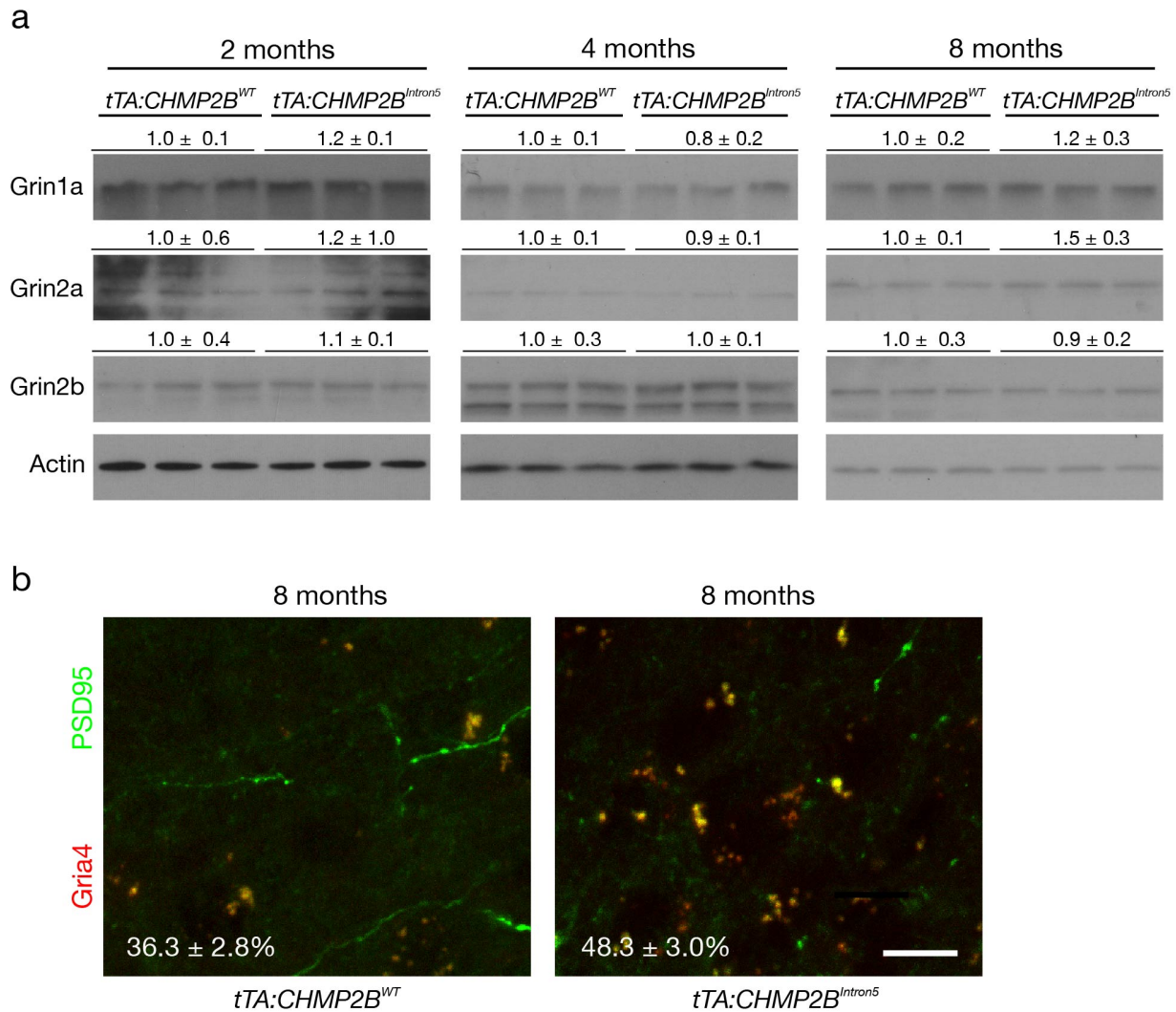
Supplementary Figure 3. Histological characterization of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice. (a) Nissl-stained sections of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}*

mice at 8 months showing that expression of $CHMP2B^{Intron5}$ leads to no overt brain defects or neuronal loss. **(b)** Higher-magnification photos of forebrain regions where the transgene was more expressed (*upper panels*, olfactory bulb; *middle panels*, cortex; *lower panels*, hippocampus) show no obvious differences between $tTA:CHMP2B^{WT}$ (*left panels*) and $tTA:CHMP2B^{Intron5}$ mice (*right panels*). **(c)** Immunofluorescence of neuronal markers in the rostral forebrain of $tTA:CHMP2B^{WT}$ and $tTA:CHMP2B^{Intron5}$ mice. In mice expressing mutant CHMP2B, we did not detect neuronal loss with a broad neuronal marker (NeuN) or an interneuron-specific marker. **(d)** NeuN density in the mPFC of $tTA:CHMP2B^{WT}$ and $tTA:CHMP2B^{Intron5}$ mice at 8 months ($n = 4$ mice per genotype; $P > 0.5$ by two-sided t test). **(e)** Representative dendritic segments of mPFC layer II-III pyramidal neurons (apical dendrite) in $tTA:CHMP2B^{WT}$ and $tTA:CHMP2B^{Intron5}$ mice at 8 months (Golgi staining). **(f)** *Left*: quantitative analysis of spine density and morphology of mPFC layer II-III pyramidal neurons (left) and on apical dendrites of CA3 hippocampal pyramidal neurons (right) ($n = 4$ mice per genotype; $P < 0.02$ for stubby spines, $P < 0.01$ for thin spines, $P < 0.001$ for total spines by two-sided two-way ANOVA corrected for multiple comparison and Bonferroni test). Scale bar, 1 mm in **a**, 300 μm in **b** and **c**, 5 μm in **e**. Values are mean \pm s.e.m. in **d** and mean \pm s.d. in **f**.

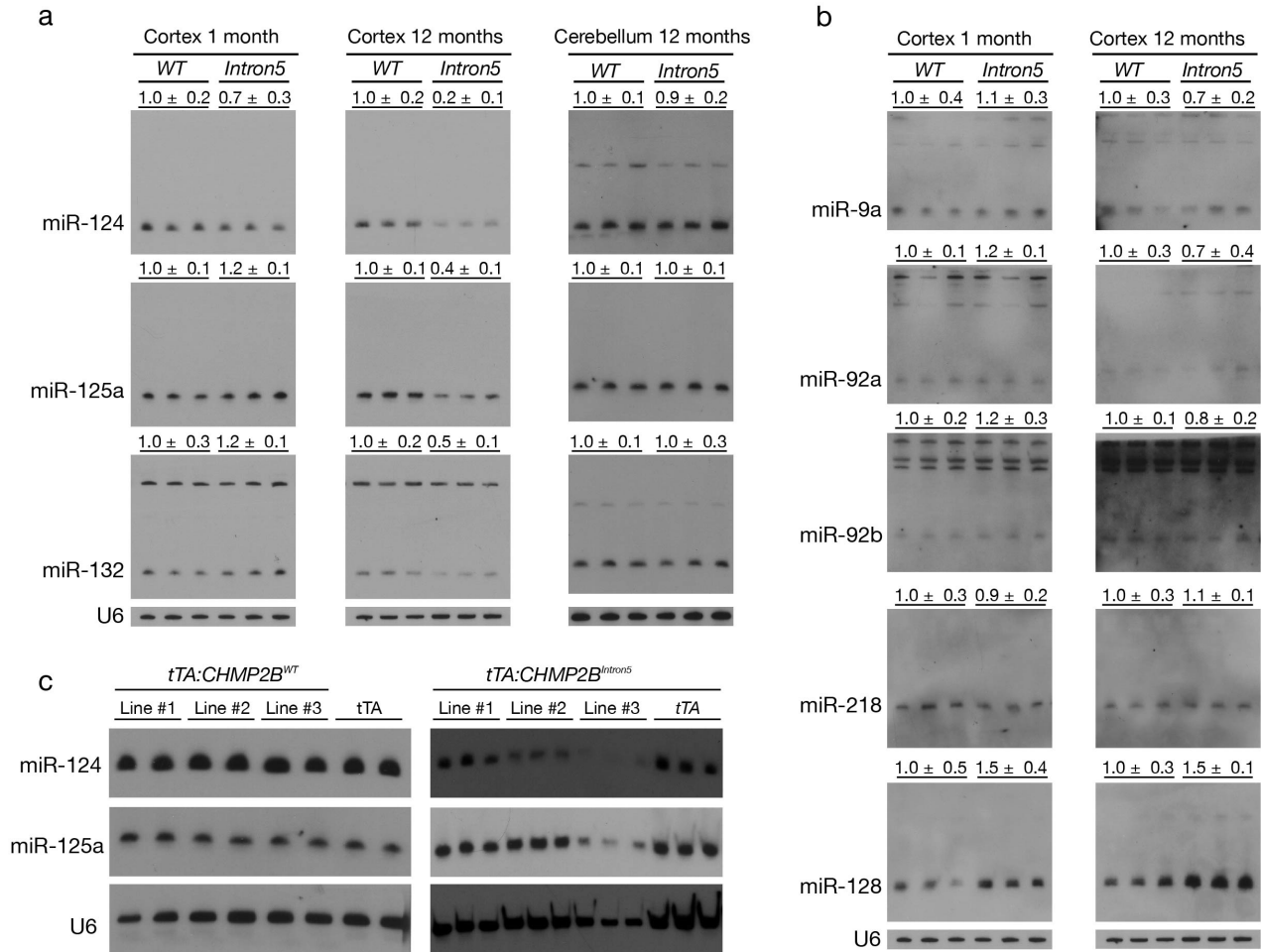


Supplementary Figure 4. Pathological phenotypes of *tTA:CHMP2B^{Intron5}* mice. **(a)** Representative photos of the mPFC of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice at 8 months stained with GFAP (red) and NeuN (green). **(b)** GFAP coverage in the cortex of *tTA:CHMP2B^{WT}*

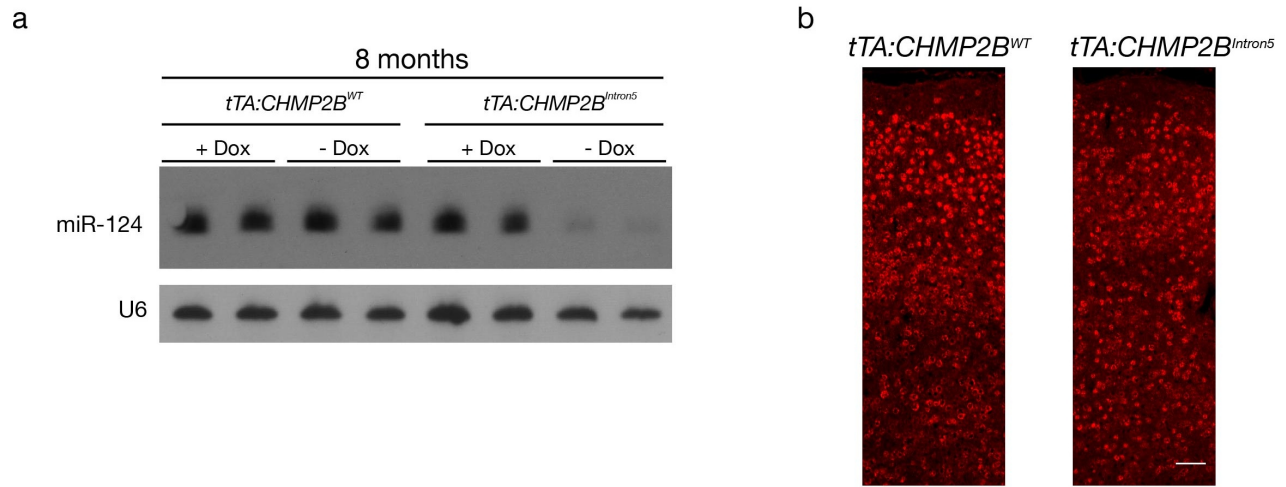
and *tTA:CHMP2B^{Intron5}* mice at 8 months ($n = 4$ mice per genotype; *: $P < 0.05$ by two-sided t test). **(c)** *Upper panel*: Western blot analysis of ubiquitinated proteins in cortical samples from *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice at 12 months. Ubiquitinated species were modestly increased in *tTA:CHMP2B^{Intron5}* mice. *Lower panels*: immunofluorescence staining with an anti-ubiquitin antibody revealed a limited amount of protein deposits in the cells. **(d)** Western blot analysis of p62 expression in cortical samples of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice at 12 months. p62 was upregulated in both soluble and insoluble fractions from mutant mice. Scale bar, 50 μm in **(a)** and 20 μm in **(c)**.



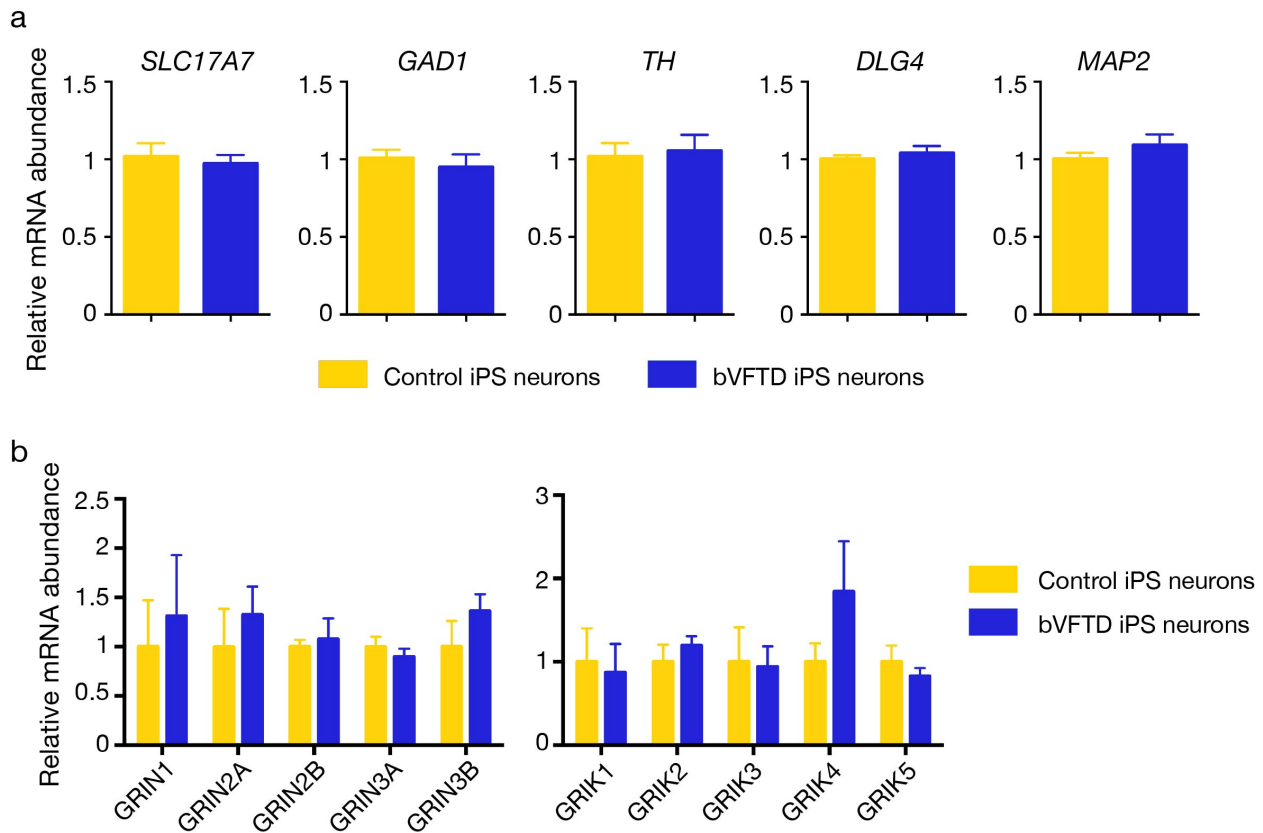
Supplementary Figure 5. Further characterization of glutamate receptors in *tTA:CHMP2B^{Intron5}* mice. **(a)** Representative western blots of PSD fractions of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice (2, 4, and 8 months) probed with anti-Grin1 (*upper panels*), anti-Grin2a (*middle panels*), and anti-Grin2b (*lower panels*). **(b)** Quantification of the percentage of Gria4-positive puncta among all PSD95-positive puncta after double immunostaining for PSD95 (green) and Gria4 (red) in the mPFC of 8-month-old *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice ($P < 0.01$). Scale bar, 200 μ m.



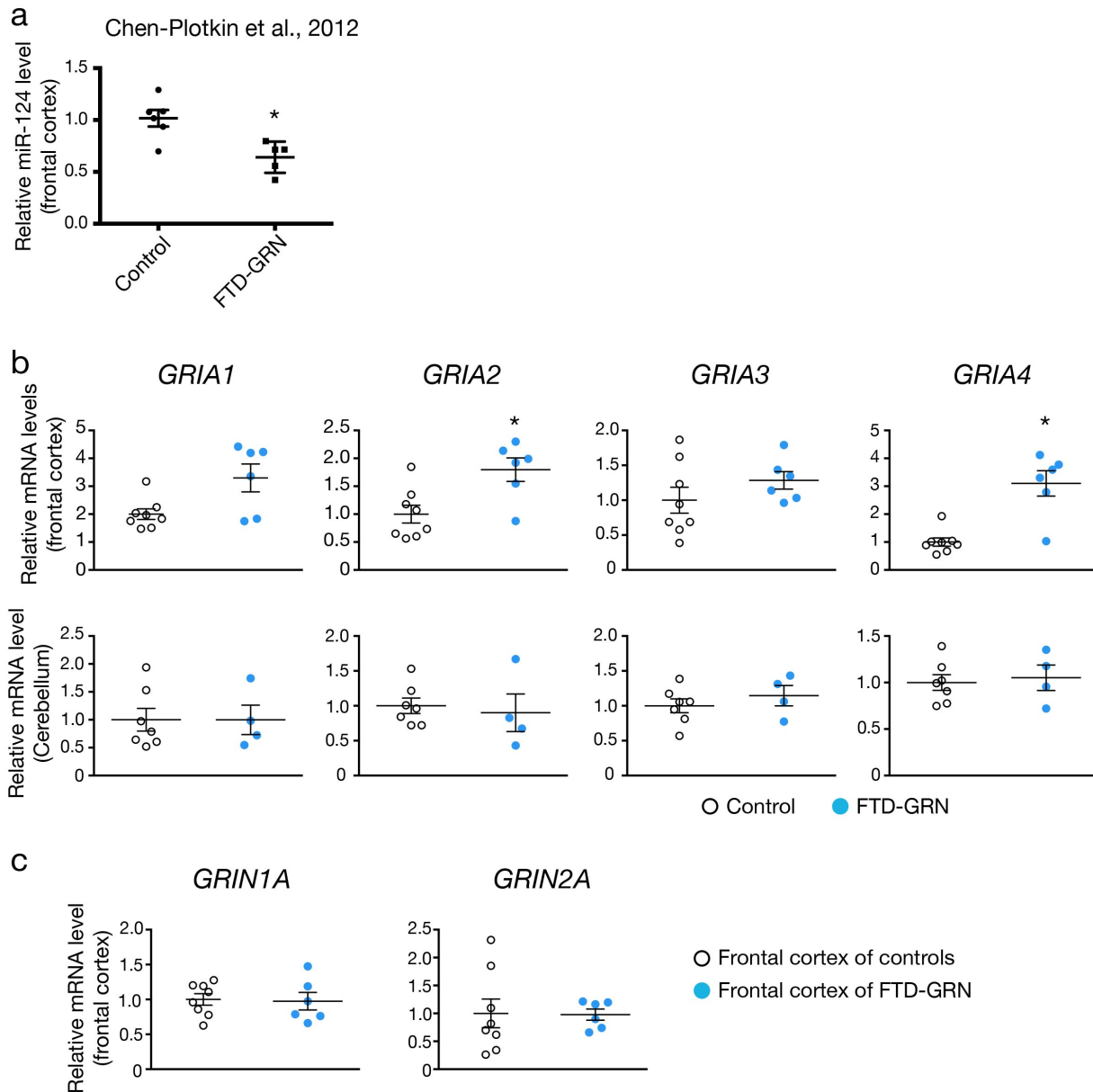
Supplementary Figure 6. Time-dependent changes in other miRNAs in *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* cortex. **(a)** Northern blot analysis of miR-124, miR-125a and miR-132, in cortical samples at 1 month and 12 months of age. As control, we extracted RNA from the cerebellum of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice (age 12 months) and also probed for those miRNAs. **(b)** Northern blot analysis of miR-9, miR-92a, miR-92b, miR-134, miR-218, and miR-128 in cortical samples of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice at 1 month and 12 months of age. **(c)** Northern blot analysis of miR-124 and miR-125a in cortical samples at 12 months of age in different lines of *tTA:CHMP2B^{WT}* (left) and *tTA:CHMP2B^{Intron5}* mice (right).



Supplementary Figure 7. miR-124 downregulation is a consequence of CHMP2B^{Intron5} expression. **(a)** Northern blot of miR-124 levels in the cortex of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice fed with regular or doxycycline-containing food from embryonic development to 8 months. RNA in each lane was isolated from one animal. **(b)** Fluorescence *in situ* analysis of miR-124 showed its similar neuronal expression in the cortex of *tTA:CHMP2B^{WT}* and *tTA:CHMP2B^{Intron5}* mice (2 months old). Scale bar, 20 μ m.

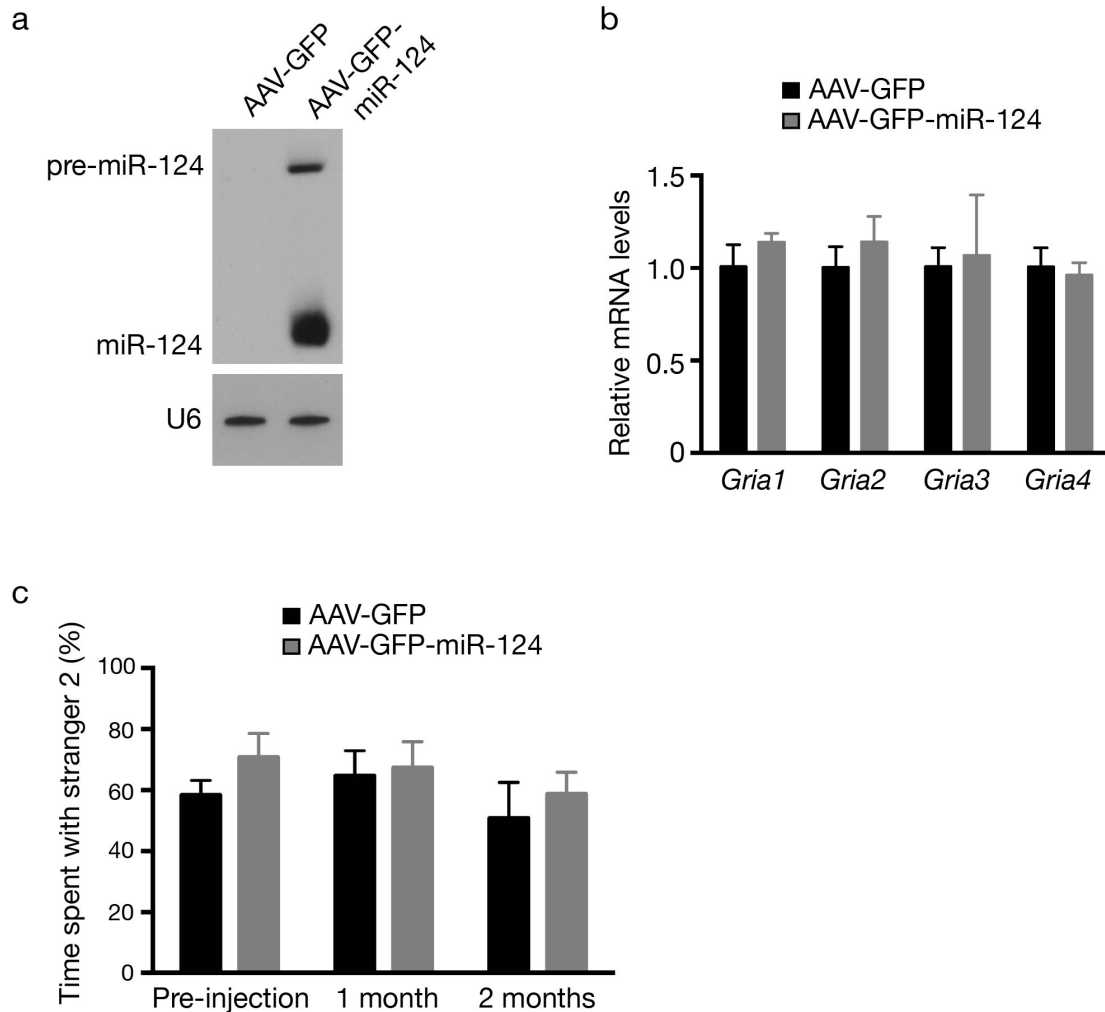


Supplementary Figure 8. Neuronal differentiation from iPSC lines. **(a)** Quantification of mRNA levels for different neuronal subtype-specific markers, such as vGLUT1 for excitatory neurons (encoded by *SLC17A7*), GAD67 for inhibitory neurons (encoded by *GAD1*), TH for dopaminergic neurons, as well as general neuron-specific markers such as PSD95 (encoded by *DLG4*) and MAP2 in 8-week-old neurons differentiated from 4 iPSC lines used in Figure 5. **(b)** Expression levels of kainate and NMDA subunits in 8-week-old neurons differentiated from 4 iPSC lines. $P > 0.5$ by two sided t test.



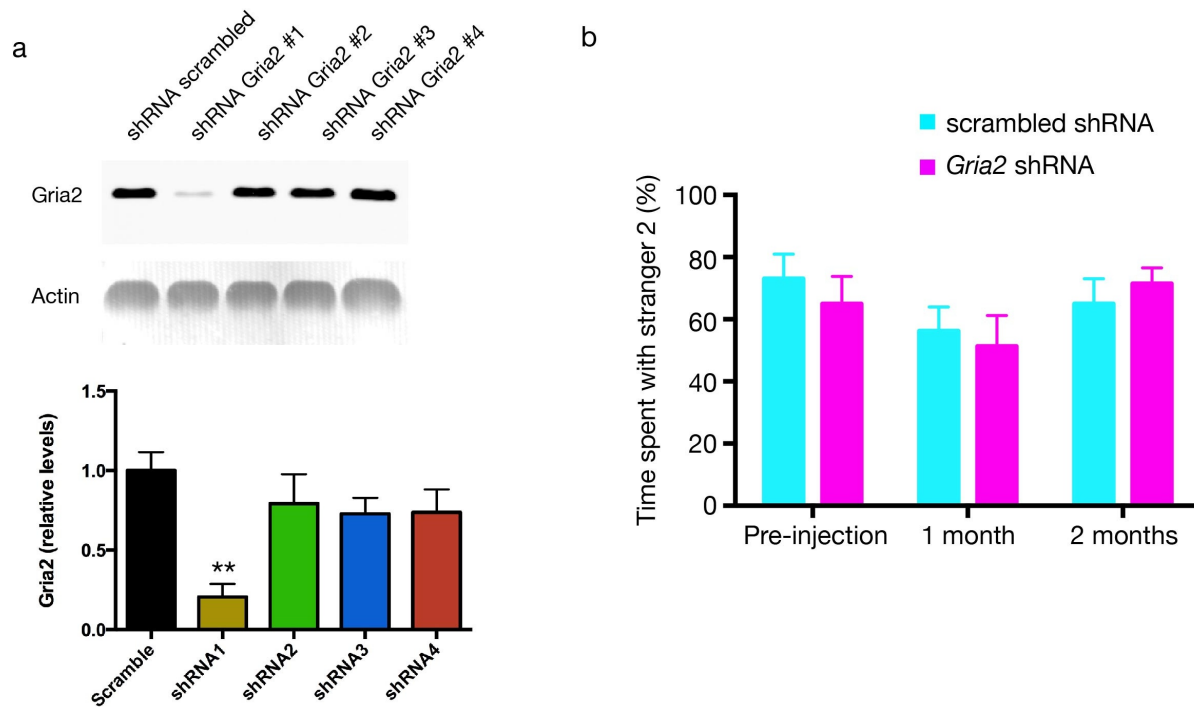
Supplementary Figure 9. Dysregulation of miR-124 and AMPARs specifically in the frontal cortex of a subset of FTD cases. **(a)** Re-analysis of miR-124 level in the frontal cortex of FTD brains from previously published profiling data³⁸. In this study, $n = 6$ for controls and $n = 5$ for FTD-GRN; *: $P < 0.05$ by Mann-Whitney test. **(b)** Re-analysis of *GRIA2* and *GRIA4* in the

frontal cortex and the cerebellum of subjects with FTD-GRN, based on published data from a microarray study³⁹. To compensate for any potential neuronal loss in FTD brains, we normalized AMPARs mRNAs contents against the geometric mean of four different neuronal-specific reference genes (MAP2, Enolase 2, GAP43 and PSD95). Cortex samples: $n = 8$ for controls, $n = 6$ for FTD-GRN; cerebellum samples: $n = 7$ for controls, $n = 4$ for FTD-GRN; *: $P < 0.05$; **: $P < 0.01$ by Mann-Whitney test. (c) Re-analysis of mRNAs encoding NMDA receptor subunits 1 and 2a using the same published dataset³⁹ (*: $P > 0.5$ by Mann-Whitney test). Values are mean \pm s.e.m.



Supplementary Figure 10. Some effects of miR-124 rescue. **(a)** Northern blot analysis of the efficiency of the AAV-GFP-miR124 construct in driving miR-124 expression in HEK cells. **(b)** qRT-PCR analysis of Gria transcripts in the occipital cortex *tTA:CHMP2B^{Intron5}* mice 2 months after AAV injection in the mPFC ($n = 5$ AAV-GFP-infected mice and $n = 4$ AAV-GFP-miR-124-infected mice; $P > 0.2$ by two-sided t test). **(c)** Analysis of Social memory and novelty after AAV-mediated delivery of miR-124. At all time points, the proportion of time spent with

Stranger 2 was similar in *tTA:CHMP2B^{Intron5}* mice injected with AAV-GFP and those injected with AAV-GFP-miR-124 ($n = 10$ per AAV vector; $P > 0.3$ by two-sided repeated-measures two-way ANOVA corrected for multiple comparison, Bonferroni test. Values are mean \pm s.d. in **b** and mean \pm s.e.m. in **c**.



Supplementary Figure 11. The effects of *Gria2* knockdown. **(a)** *In vitro* efficiency of four different shRNAs targeting *Gria2* in mouse cortical cultures 72 h after infection with lentiviral vectors expressing GFP and either a scrambled shRNA or an shRNA against mouse *Gria2*. *Lower panel* shows the quantification of 3 independent experiments (**: $P < 0.01$ one-way ANOVA followed by Bonferroni's test). **(b)** Analysis of social memory after *Gria2* knockdown in *tTA:CHMP2B^{Intron5}* mice ($n = 7$ per genotype; $P > 0.4$ by two-sided repeated-measures two-way ANOVA followed by Fisher's least-significant-difference test). Values are mean \pm s.e.m.

Supplementary Table 1. Neurotransmitter receptors whose mRNAs are targeted by miR-124 in mouse and human.

Receptor	Number of subunits	Potential miR-124 targets
GABA	16	0
Nicotinic (acetylcholine)	17	0
Muscarinic (acetylcholine)	5	0
Dopamine	5	0
Adrenergic	4	0
Serotonin	15	1
Glycine	5	1
Glutamate		
Metabotropic	8	0
Kainate	5	0
NMDA	7	1
AMPA	4	3

Supplementary Table 2. Primers used in this study.

Genes	Primer Sequences	Product Sizes (bp)
<i>CHMP2B^{Intron5}</i>	Forward TGACGGTTCTGATGACGAAGAAG Reverse GGGAGGGTGTGGGAGGTTTT	146
<i>CHMP2B</i>	Forward AAAGTGATGAATTCCCAAATGAA Reverse TCTTGTTAACTGCCTGCATTGT	76
<i>Chmp2b</i>	Forward GGACCGAGCAGCCTTAGAG Reverse CCAATCTTGGCCATCTTCTTA	69
<i>Pri-miR-124-1</i>	Forward TTCTCCCATCCTCCCTCTCT Reverse AATCAAGGTCCGCTGTGAAC	144
<i>Pri-miR-124-2</i>	Forward GTTCACAGCGGACCTTGATT Reverse TCTCCCCACCCTTCCTAACT	116
<i>Pri-miR-124-3</i>	Forward CGGTGAATGCCAAGAGAGG Reverse ATTGTTCCGCGGATTTGTC	147
<i>mGria1</i>	Forward AGGGATCGACATCCAGAGAG Reverse TGCACATTTCTGTCAAACC	63
<i>mGria2</i>	Forward CAGTTTCGCAGTCACCAATG Reverse ACCCAAAAATCGCATAGACG	65
<i>mGria3</i>	Forward CCACTTGGATTCTCCAATAGT Reverse GCATACACCCTCTGGAGAA	72
<i>mGria4</i>	Forward CTGCCAACAGTTTTGCTGTG Reverse AAATGGCAAACACCCCTCTA	66
<i>mGrik1</i>	Forward GATATTTGATTGTTTCGCACGAG Reverse GCCCATGAACAAAATCTGCT	61
<i>mGrik2</i>	Forward AGTGCCACCATAACCATCCAG Reverse GCTGGCACTTCAGAGACATTC	125
<i>mGrik3</i>	Forward AGGTCACTCATCGTCACCACT Reverse ATCAGACTTGCGGAACATGA	60
<i>mGrik4</i>	Forward GCCATTGAGTATGGCACGAT Reverse TGGTAACGGGAATTTTGGAA	65
<i>mGrik5</i>	Forward CCCCTCAGCTAGCCTCATCT Reverse GCCTCGCACCAGTTCTTCTA	67
<i>mGrin1</i>	Forward CATTTAGGGCTATCACCTCCA Reverse CACTGTGTCTTTTTGGTTTTGC	113
<i>mGrin2a</i>	Forward ATTCAACCAGAGGGGCGTA Reverse TTCAAGACAGCTGCGTCATAG	87
<i>mGrin2b</i>	Forward GGGTTACAACCGGTGCCTA Reverse CTTTGCCGATGGTGAAAGAT	65
<i>mGrin2c</i>	Forward GAAGCGGGCCATAGACCT Reverse TGGCAGATCCCTGAGAGC	90
<i>mGrin2d</i>	Forward TGCGATACAACCAGCCAAG Reverse AGATGAAGGCGTCCAGTTTC	72
<i>β-actin</i>	Forward CGTGGGCCC GCCCTAGGCACCAG Reverse TTGGCCTTAGGGTTCAGGGGGG	243

Cont.

Supplementary Table 1. (cont.)

Genes	Primer Sequences	Product Sizes (bp)
<i>hGRIA1</i>	Forward CAGCGACAGCTTTGAGATGA Reverse GATGGCATAGACTCCTTTGGA	64
<i>hGRIA2</i>	Forward TGTGGAGCCAAGGACTCTG Reverse CCCCCGACAAGGATGTAGA	89
<i>hGRIA3</i>	Forward GGAGACTGCTTAGCAAATCCTG Reverse TCATTCCTTGTACTTGCACCA	88
<i>hGRIA4</i>	Forward CCAAACAAACAGAAATTGCCTA Reverse CCACATCTTTTCATACACTGCTATTT	66
<i>hGRIK1</i>	Forward GCGGTTAGAGATGGATCAACA Reverse TCATGAAAGCCACATCTTCT	79
<i>hGRIK2</i>	Forward CACCCTTGCAACCCTGAC Reverse TTTGGGCATGAGCTCAGAA	111
<i>hGRIK3</i>	Forward GAGTATGGGGCTGTCAAGGA Reverse CGAAGGTGGAGATCTTGGAT	67
<i>hGRIK4</i>	Forward CAAGGAGGAAGATCACAGAGC Reverse TGGTAACGGGAATTTTGGAA	87
<i>hGRIK5</i>	Forward TTGGCGAGCTCATCAACC Reverse TGACCTTCTCCCGCTCAG	75
<i>hGRIN1</i>	Forward TACAAGCGGCACAAGGATG Reverse TCAGGCTCTGCTCTACCACTC	110
<i>hGRIN2A</i>	Forward CTTGGAAGAGGCAGATCGAC Reverse CTTCTCGTTGTGGCAGATCC	104
<i>hGRIN2B</i>	Forward AGCAATGGGACTGTCTCACC Reverse AACATCATCACCCATACGTCAG	68
<i>hGRIN3A</i>	Forward AAATCCAAGCTGCAATACTGG Reverse AAATGCTGCTGCTTTTCCTC	89
<i>hGRIN3B</i>	Forward ACTGCTGACCGTGGGAAA Reverse ATGAACTCGGACAGGTTGGA	93
<i>hSLC17A7</i>	Forward ACGACAGCCTTTTGTGGTTC Reverse TGAGGGGGTTCATGAGTTTC	243
<i>hGAD1</i>	Forward CCAACAGCCTGGAAGAGAAG Reverse GGTGGAGCGATCAAATGTCT	247
<i>hTH</i>	Forward ACTGGTTCACGGTGGAGTTC Reverse AGCTCCTGAGCTTGTCTTGT	230
<i>hDLG4</i>	Forward CCGACAAGTTTGGATCCTGT Reverse CCCATAGAGGTGGCTGTTGT	163
<i>hMAP2</i>	Forward CCAATGGATTCCCATACAGG Reverse CTGCTACAGCCTCAGCAGTG	179
<i>hENOLASE2</i>	Forward ACTTTGTCAGGGACTATCCTGTG Reverse TCCCTACATTGGCTGTGAACT	93
<i>hGAP43</i>	Forward GAGGATGCTGCTGCCAAG Reverse GGCACCTTTCCTTAGGTTTGGT	111
<i>hGADPH</i>	Forward CTCTGCTCCTCCTGTTCGAC Reverse GCCCAATACGACCAAATCC	119

