2. Fritz, I. B. (1961) *Physiol. Rev.* **41**, 52–129.
3. Tepperman, J. & Tepperman, H. M. (1970) *Fed. Proc.* **29**, 1284–1293.
4. Lowenstein, J. M. (1968) *Biochem. Soc. Symp.* **27**, 61–86.
5. Srere, P. A. (1962) in *Methods in Enzymology*, eds. Colowick, S. P. & Kaplan, N. O. (Academic Press, New York), Vol. 5, pp. 641–644.
6. Srere, P. A. (1965) *Nature* **205**, 766–770.
7. Foster, D. W. & Srere, P. A. (1968) *J. Biol. Chem.* **243**, 1926–1930.
8. Robinson, B. H. & Chappell, J. B. (1967) *Biochem. Biophys. Res. Commun.* **28**, 249–255.
9. Lowry, O. H., Rosebrough, N. J., Farr, A. L. & Randall, A. J. (1951) *J. Biol. Chem.* **193**, 265–275.
10. Robinson, B. H., Williams, G. R., Halperin, M. L. & Leznoff, C. C. (1970) *Eur. J. Biochem.* **15**, 263–272.
11. Robinson, B. H. & Williams, G. R. (1970) *Biochim. Biophys. Acta* **216**, 63–70.
12. Robinson, B. H., Williams, G. R., Halperin, M. L. & Leznoff, C. C. (1971) *Eur. J. Biochem.* **20**, 65–71.
13. Bremer, J. (1968) in *Biochemical Preparations*, ed. Lands, W. E. M. (John Wiley & Sons, Inc., New York), Vol. 12, pp. 69–73.
14. Taketa, K. & Pogell, B. M. (1966) *J. Biol. Chem.* **241**, 720–726.
15. Shug, A., Lerner, E., Elson, C. & Shrago, E. (1971) *Biochem. Biophys. Res. Commun.* **43**, 557–563.
16. Pande, S. V. & Blanchaer, M. C. (1971) *J. Biol. Chem.* **246**, 402–411.
17. McLean, P., Gumaa, K. A. & Greenbaum, A. L. (1971) *FEBS Lett.* **17**, 345–350.
18. Halperin, M. & Robinson, B. H. (1972) in *Proceedings of the Symposium on Insulin Action*, ed. Fritz, I. B. (Academic Press, New York), in press.
19. Lardy, H. A., Foster, D. O., Young, J. W., Shrago, E. & Ray, P. D. (1965) *J. Cell. Comp. Physiol.* **66** (Suppl. 1), 39–53.
20. Krebs, H. A. (1966) *Advan. Enzyme Regul.* **4**, 339–353.
21. Fritz, I. B. (1967) *Perspect. Biol. Med.* **10**, 643–677.

Correction. In the article "RNA in Human Leukemic Cells Related to the RNA of a Mouse Leukemia Virus", by Hehlmann, R., Kufe, D. & Spiegelman, S., which appeared in the February 1972 issue of *Proc. Nat. Acad. Sci. USA* **69**, 435–439, Figs. 2 and 3 (pp. 436 and 437) were transposed during printing and should be reversed (see below).

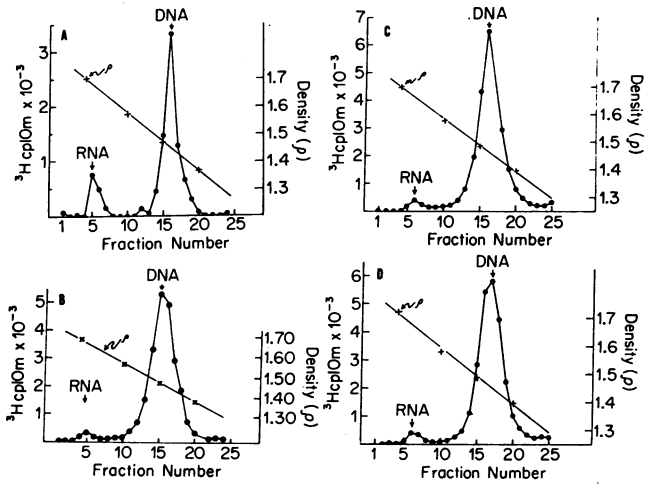

FIG. 2. (*A–D*) Cs_2SO_4 density profiles of RLV-[^{3}H]DNA hybridized to polysomal RNAs of four leukemic samples. Polysomal RNA was isolated from buffy coats of patients showing clinical manifestation of acute lymphocytic leukemia (*A*, *C*, and *D*) and of acute myelogenous leukemia (*B*). The cells were disrupted with a Dounce homogenizer, and cytoplasmic pellets were prepared as described under Fig. 1*B*. 300 μg of polysomal RNA were hybridized to RLV-[^{3}H]DNA in 60-μl volumes, and the reactions were analyzed by Cs_2SO_4 density centrifugation.

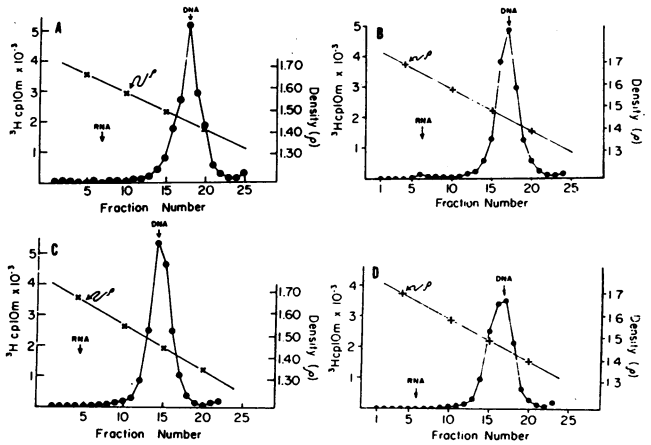

FIG. 3. (*A–D*) Cs_2SO_4 density centrifugation of RLV-[^{3}H]-DNA after annealing to polysomal RNA isolated from (*A*) normal white buffy coat, (*B*) phytohemagglutinin-stimulated lymphocytes, (*C*) buffy coat of a leukemic patient in clinical remission, and (*D*) fetal lung. The polysomal RNA input was 300 μg, except in (*C*) where 1000 μg were used per 60-μl hybridization reaction. Subsequent analysis on Cs_2SO_4 was described in Fig. 1.