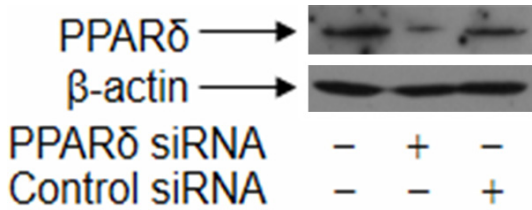
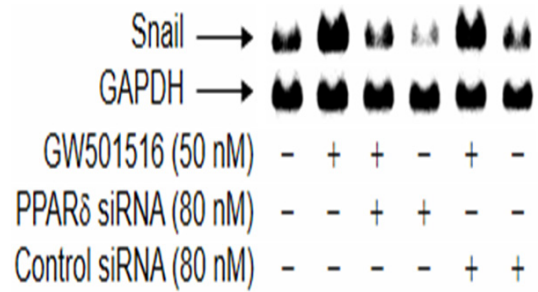


PPAR δ modulates the migration and invasion of melanoma cells



Supplementary Figure 1. Effects of small interfering (si)RNA on PPAR δ expression. A375SM cells were transfected with 80 nM PPAR δ or control siRNA for 24 h. Cells were then washed with ice-cold PBS and lysed in PRO-PREP Protein Extraction Solution. Aliquots of cell lysates were analyzed by Western blotting with anti-PPAR δ or anti- β -actin antibodies. The level of PPAR δ was markedly reduced upon transfection with the corresponding siRNAs, whereas control siRNA (consisting of a pool of nonspecific sequences) had no effect.



Supplementary Figure 3. Effects of PPAR δ siRNA on Snail expression. A375SM cells were transfected with 80 nM PPAR δ siRNA or 80 nM control siRNA. After incubation for 24 h, the cells were incubated for a further 38 h in the presence or absence of GW501516. Total RNA was extracted and fractionated by 1% agarose gel electrophoresis. Fractionated RNA was transferred onto a Hybond-N⁺ nylon membrane and hybridized with Snail or GAPDH cDNA probes. The levels of Snail were markedly reduced upon transfection with PPAR δ siRNA, whereas control siRNA had no effect.



Supplementary Figure 2. Effects of GW501516 on the expression of Snail protein. A375SM cells were incubated for 38 h with different concentrations of GW501516 (A) or exposed to 100 nM GW501516 for the indicated times (B). Cells were then washed with ice-cold PBS and lysed in PRO-PREP Protein Extraction Solution. Aliquots of cell lysates were analyzed by Western blotting with anti-PPAR δ or anti- β -actin antibodies. GW501516 induced the expression of Snail protein in a concentration- and time-dependent manner.