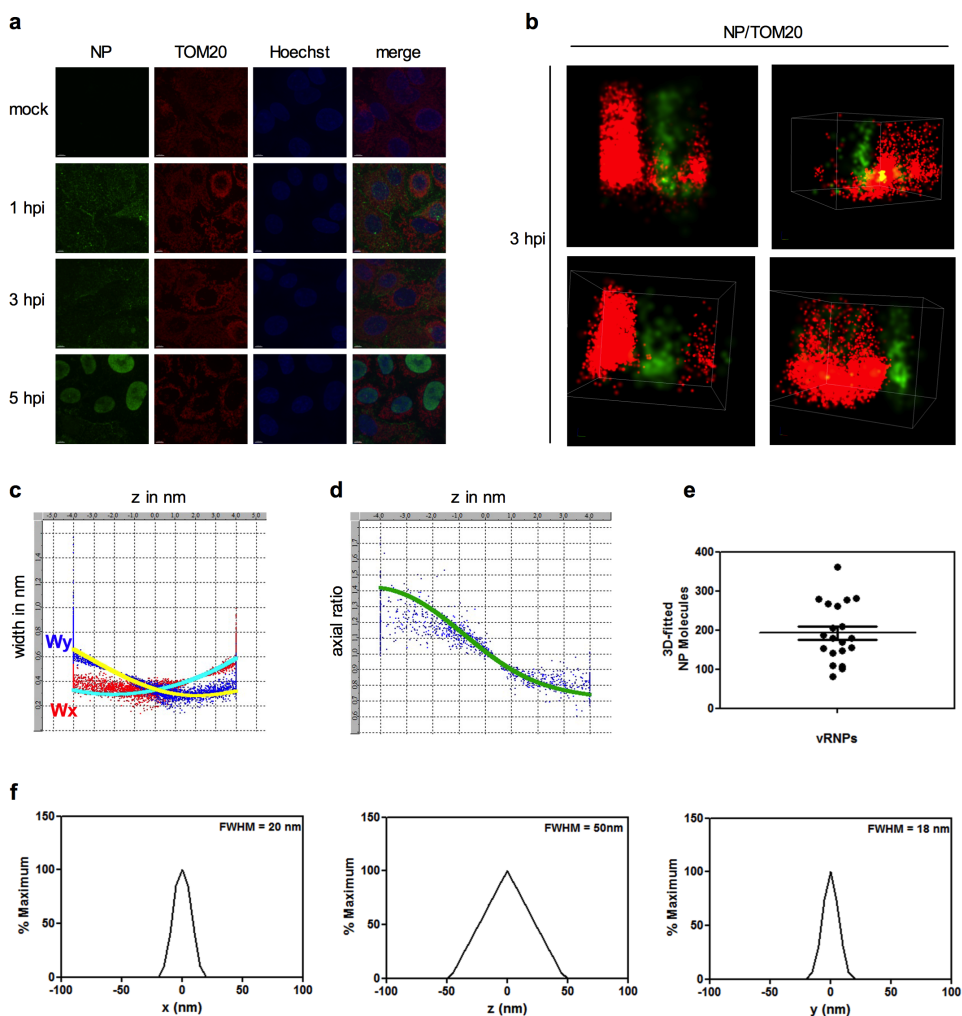


SUPPLEMENTARY INFORMATION

Viral suppressors of the RIG-I-mediated interferon response are pre-packaged in influenza virions

Authors: Swantje Liedmann¹, Eike R. Hrincius², Cliff Guy³, Darisuren Anhlan¹, Rüdiger Dierkes¹, Robert Carter⁴, Gang Wu⁴, Peter Staeheli⁵, Douglas R. Green³, Thorsten Wolff⁶, Jonathan A. McCullers^{2,7}, Stephan Ludwig^{1,8*} and Christina Ehrhardt^{1,8}

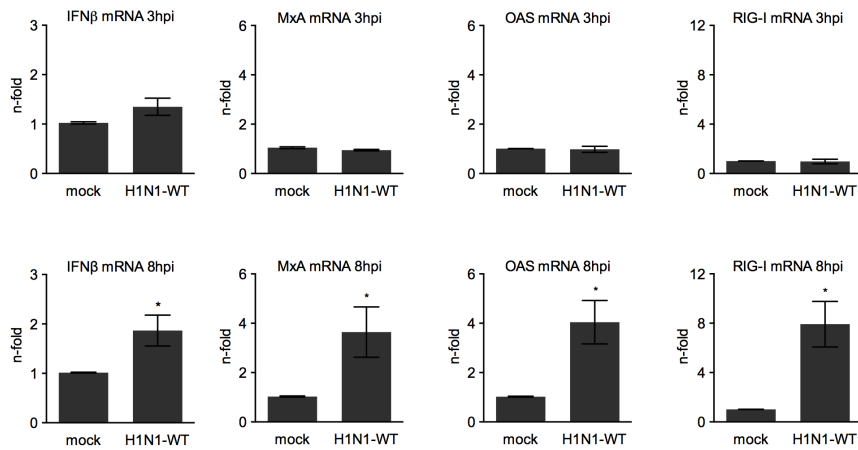
Supplementary Figure 1



Supplementary Figure 1: Determination of three-dimensional localization accuracy.

A549 cells were infected with H1N1-WT (MOI = 50) and prepared for immunofluorescence analysis. (a) Spinning disk laser scanning confocal microscopy revealed cytosolic vRNPs (green: NP-AF568) in the mitochondrial network (TOM20-AF647) previous to import into the nucleus (Hoechst) at 3 hpi. Nuclear import was completed at 5 hpi. Scale bars represent 5 μm . Shown are representative images from three independent experiments. (b) Three-dimensional stochastic optical reconstruction microscopy (STORM) revealed vRNPs (green: NP-AF568) as helical-like structure next to the outer mitochondrial membrane (red: TOM20-AF647) 3 hpi. (c, d and f) Three-dimensional localization accuracy of astigmatic STORM imaging was determined using fluorescent microbeads. (c) Width of the localized molecules (W_x or W_y) and (d) the axial ratio of the molecules are plotted against the z stepping range. (e) The number of molecules fitted to the three-dimensional calibration curves are shown for 20 randomly selected vRNPs. (f) The spatial distributions of the localizations within the bead molecules are presented for the x, y and z dimensions, with respective FWHM values as noted.

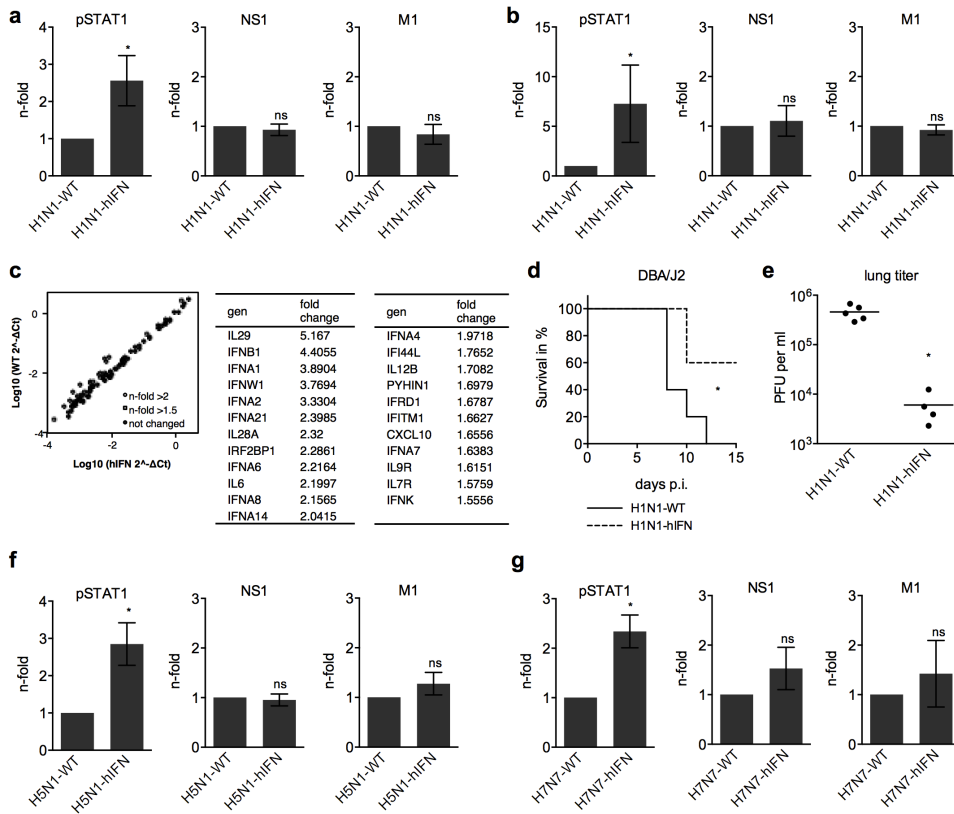
Supplementary Figure 2



Supplementary Figure 2: The type I IFN signaling pathway is not activated at 3 hours post infection.

A549 cells were infected with H1N1-WT (MOI = 5) and prepared for qRT-PCR-analysis. MessengerRNA levels of *IFNβ*, *RIG-I*, *MxA* and *OAS* were unchanged at 3 hpi. Elevated mRNA levels were measured at 8 hpi; bars show mean ± SD of three independent experiments. For statistically analysis Students t-test was performed; *P<0.05.

Supplementary Figure 3

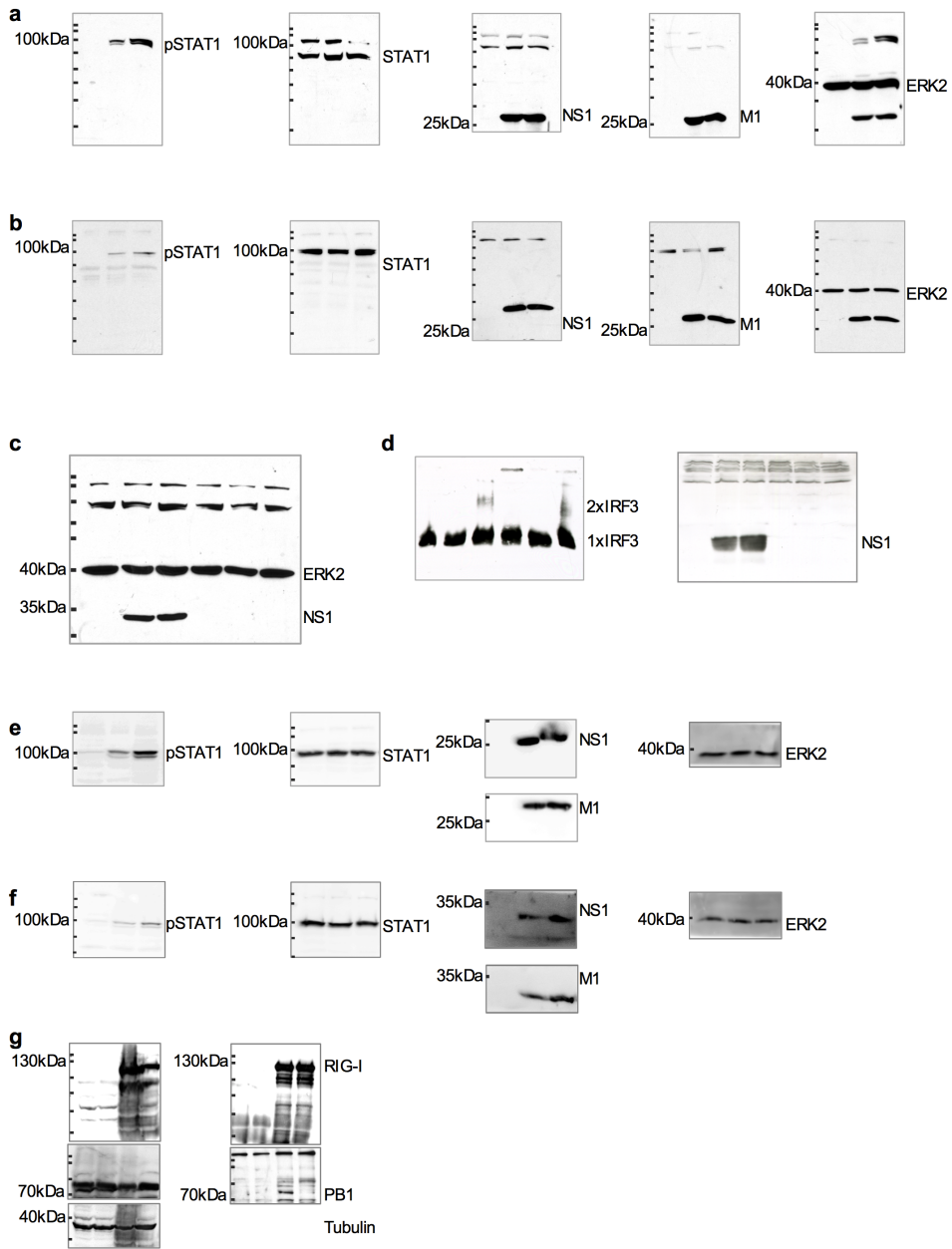


Supplementary Figure 3: Quantification of Western Blots, expression analysis of IFN-related genes and pathogenicity in DBA/J2 mice.

Western Blots were quantified using IMAGEJ 1.48v. Representative WB are presented in (a, b) Figure 2a and (f, g) Figure 2k. pSTAT1 bands were normalized to STAT1 bands. NS1 and M1 bands were normalized to ERK2 bands. Depicted are fold changes of mutant viruses compared to wild type viruses; quantified were WB of three independent experiments, bars show mean \pm SD. (c) U937 cells were infected with H1N1-WT and H1N1-hIFN (MOI = 5) and prepared for qRT-PCR-analysis. Enhanced (>1.5-fold) expression of type I IFN-related genes following disruption of the ESIE-motif was analyzed in an 88-gene-transcriptome analysis 8 hpi. (d, e) DBA/J2 mice (n = 5 per group) were infected i.n. with 10 PFU of the indicated viruses. (d) Mice were examined daily for survival for 15 days p.i. (e) Virus titers of

infected mouse lungs were determined 3 days p.i. Each dot represents an individual mouse; bars show mean. For statistical analysis (a, b, e-g) Students t-test and (d) log-rank (chi-square) test for statistical analysis of Kaplan-Meier survival data were performed; *P<0.05, ns = not statistically significant.

Supplementary Figure 4



Supplementary Figure 4: Original Western Blots.

Shown are the original Western Blots from (a-f) Figure 2 and (g) Figure 3.

Supplementary Figure 5



Supplementary Figure 5: IFN antagonism of PB1/PA impacts virulence regulation similar to RIG-I-mediated IFN antagonism of NS1.

(a, b) BALB/c mice ($n = 5$ per group) were infected i.n. with 10^4 PFU of the indicated viruses. (a) Mice were examined daily for survival for 15 days p.i. (b) Virus titers of infected mouse lungs were determined 3 days p.i. Each dot represents an individual mouse; bars show mean. For statistical analysis (a) log-rank (chi-square) test for statistical analysis of Kaplan-Meier survival data and (f) ANOVA with Tukey test for multiple comparisons were performed * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

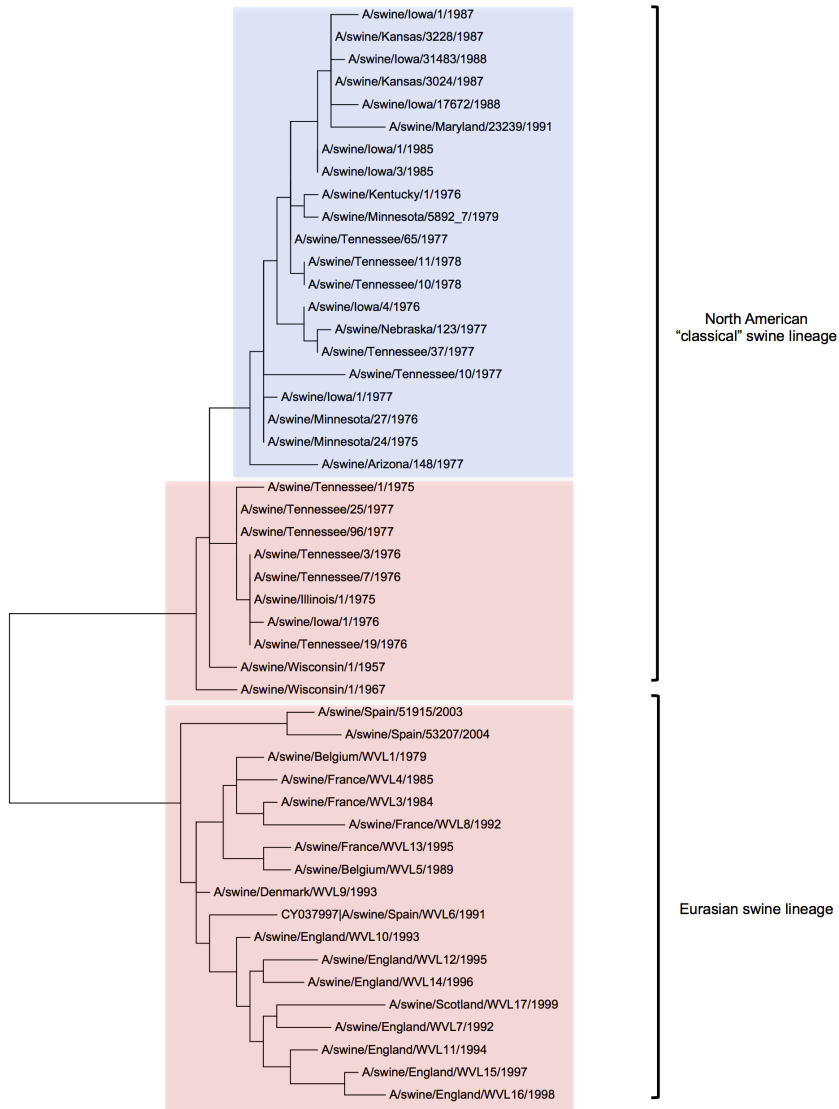
Supplementary Figure 6



Supplementary Figure 6: Evolutionary dynamics of the ESIE-motif in swine and avian IAV.

PB1 protein sequences of (a) swine and (b) avian IAV were used to construct phylogenetic tree with BEAST. Red color marks IAV with PB1 398D; blue color marks IAV with PB1 398E.

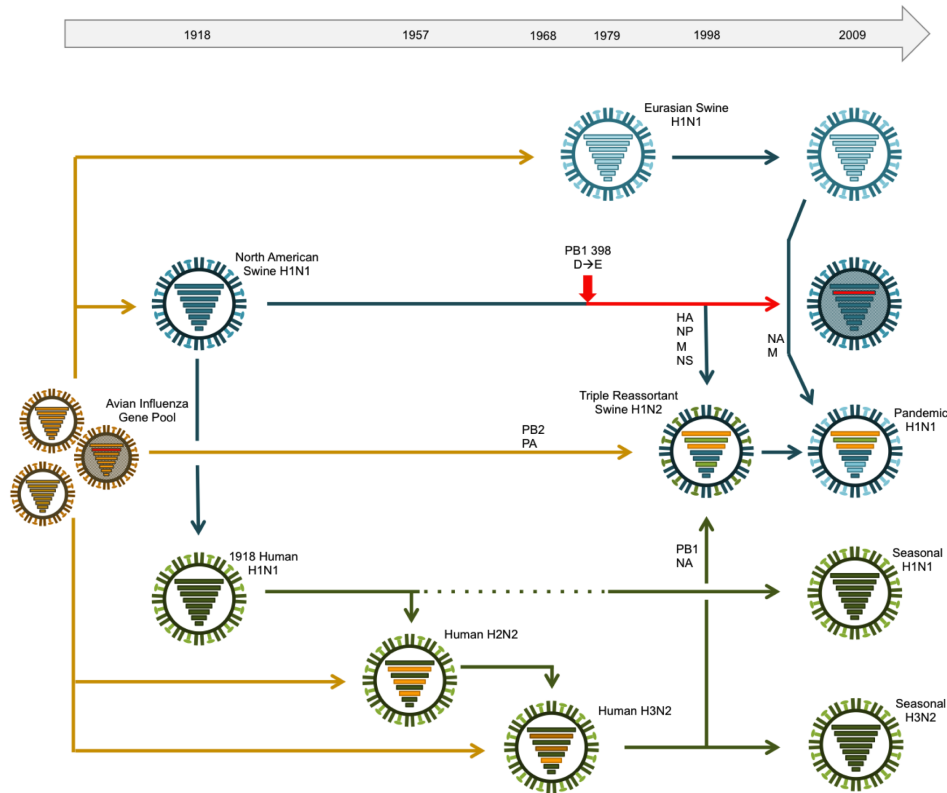
Supplementary Figure 7



Supplementary Figure 7: Completion of the ESIE-motif in the North American swine lineage and evolution of swine IAV, 1918-2009.

Amino acid sequences of the PB1 genes of representative IAV of the North American "classical" swine lineage and the Eurasian swine lineage were aligned by MUSCLE; the phylogenetic tree was constructed by using MEGA5. Red background marks IAV with PB1 398D, blue background marks IAV with PB1 398E.

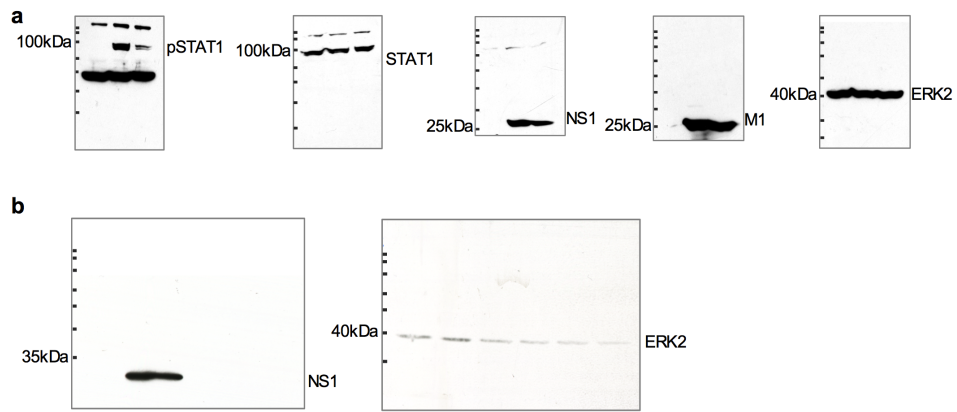
Supplementary Figure 8



Supplementary Figure 8: Evolution of swine IAV, 1918-2009.

Yellow arrows indicate derivation of gene segments from the avian IAV gene pool; evolutionary paths of swine IAV are shown with blue arrows, of human IAV with green arrows. The red arrow marks the D to E change at position 398 in the PB1 protein within the North American swine lineage. Today PB1 398E is present in an avian IAV population and persists in the North American swine lineage (shaded virus particles with red gene segment). Figure is modified from.^{1,2}

Supplementary Figure 9



Supplementary Figure 9: Original Western Blots.

Shown are the original Western Blots from Figure 4.

SUPPLEMENTARY TABLE

Supplementary Table 1: Mutagenesis primer sequences

| | |
|--------------------------------|-----------------------------------|
| H1N1-hIFN PB1 E398G for | CCGCTCTTAATAGGGGGGACTGCATC |
| H1N1-hIFN PB1 E398G rev | GATGCAGTCCCCCTATTAAGAGCGG |
| H1N1-hIFN PB1 S524G for | GAGTCAGCGGACATGGGTATTGGAGTACTG |
| H1N1-hIFN PB1 S524G rev | CAGTAACTCCAATACCCATGTCCGCTGACTC |
| H1N1-hIFN PB1 I563R for | CGTACCGATGCCATAGAGGTGACACACAAATAC |
| H1N1-hIFN PB1 I563R rev | GTATTTGTGTGTCACCTCTATGGCATCGGTACG |
| H1N1-hIFN PA E351K for | GGACATTGAGAATAAGGAGAAAATTCCAAAG |
| H1N1-hIFN PA E351K rev | CTTTGGAATTTTCTCCTTATTCTCAATGTCC |
| | |
| H5N1-hIFN PB1 D398G for | TCTATTAATAGGTGGTACAGCC |
| H5N1-hIFN PB1 D398G rev | GGCTGTACCACCTATTAATAGA |
| H5N1-hIFN PB1 S524G for | GCCGACATGGGCATTGGTGT |
| H5N1-hIFN PB1 S524G rev | AACACCAATGCCCATGTCCGGC |
| H5N1-hIFN PA E351K for | ATTGAAAATAAGGAGAAAATC |
| H5N1-hIFN PA E351K rev | GATTTTCTCCTTATTTTCAAT |
| | |
| H7N7-hIFN PB1 D398G for | CTTCTAATAGGTGGTACAGCC |
| H7N7-hIFN PB1 D398G rev | GGCTGTACCACCTATTAGAAG |
| H7N7-hIFN PB1 S524G for | GCTGACATGGGCATTGGAGTA |
| H7N7-hIFN PB1 S524G rev | TACTCCAATGCCCATGTCAGC |
| H7N7-hIFN PA E351K for | ATTGAAAATAAAGAGAAGATT |
| H7N7-hIFN PA E351K for | AATCTTCTCTTTATTTTCAAT |
| | |
| pdmH1N1-lowIFN PB1 D398E for | CTTCTAATAGAGGGCACAGCA |
| pdmH1N1-lowIFN PB1 D398E rev | TGCTGTGCCCTCTATTAGAAG |
| pdmH1N1-lowIFN PB1 R563I for A | AGGTGCCATAGAGGAGACACA |
| pdmH1N1-lowIFN PB1 R563I rev A | TGTGTCTCCTCTATGGCACCT |
| pdmH1N1-lowIFN PB1 R563I for B | AGGTGCCATATAGGAGACACA |
| pdmH1N1-lowIFN PB1 R563I rev B | TGTGTCTCCTATATGGCACCT |
| | |
| H1N1 NS1 E96A for | CATGACTCTTGCGGAAATGTCAAGG |
| H1N1 NS1 E96A rev | CCTTGACATTTCCGCAAGAGTCATG |
| H1N1 NS1 E97A for | CATGACTCTTGCGGCAATGTCAAGG |
| H1N1 NS1 E97A rev | CCTTGACATTGCCGCAAGAGTCATG |

SUPPLEMENTARY REFERENCES

1. Morens, D. M., Taubenberger, J. K., Fauci, A. S. The persistent legacy of the 1918 influenza virus. *N. Engl. J. Med.* **361**, 225-229 (2009).
2. Taubenberger, J. K., Kash, J. C. Influenza virus evolution, host adaptation, and pandemic formation. *Cell Host Microbe* **7**, 440-451 (2010).