

Supporting Information

Xia and Springer 10.1073/pnas.1420645111

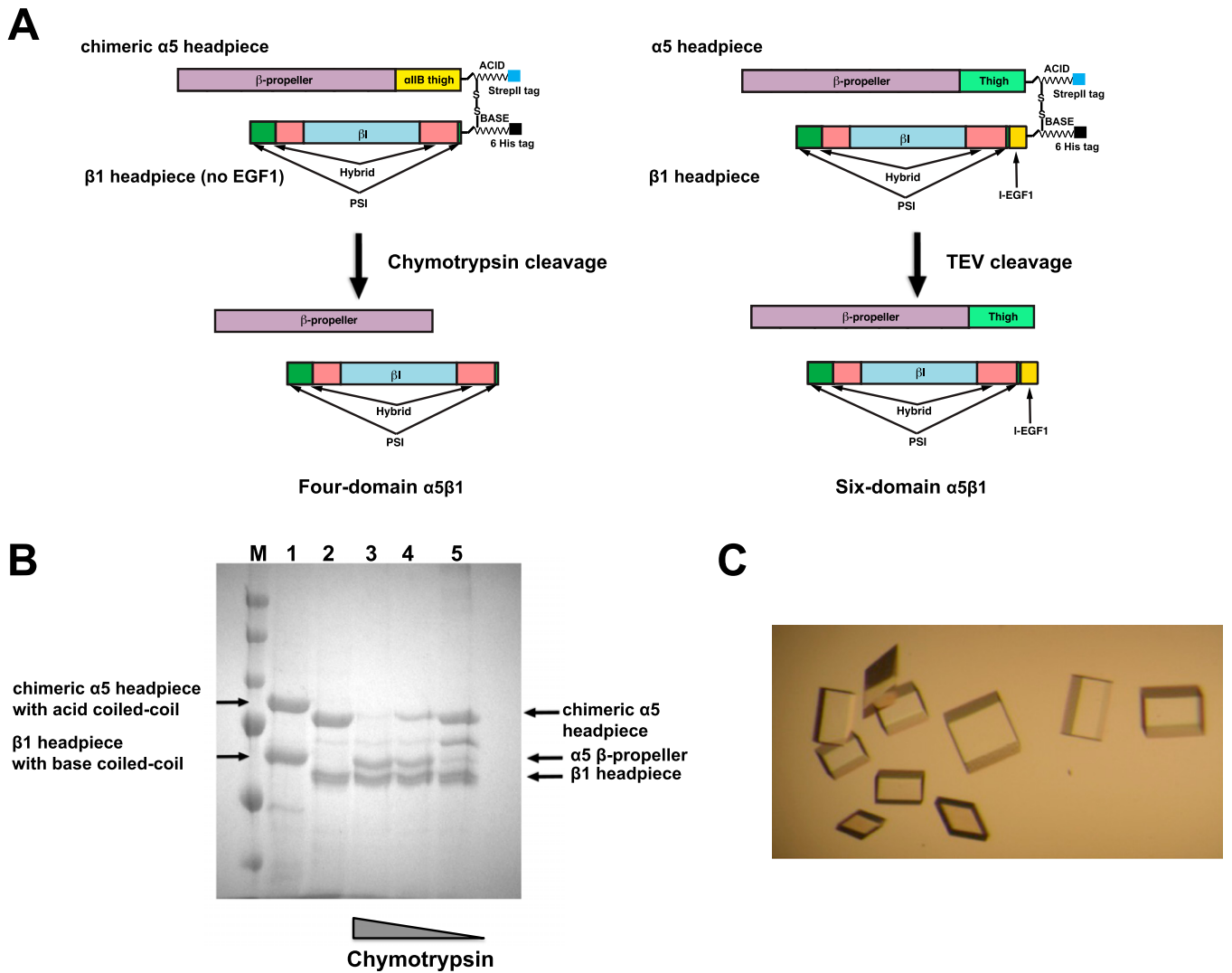


Fig. S1. $\alpha_5\beta_1$ headpiece construct design, protease digestion, and crystallization. (A) Schematic illustration of chimeric $\alpha_5\beta_1$ headpiece (Left) and wild-type $\alpha_5\beta_1$ headpiece (Right) constructs. (B) Cleavage of the thigh domain of chimeric protein analyzed by reducing SDS/PAGE. Lanes: M, markers; 1, control uncleaved integrin; 2–5, various protease:integrin ratios (wt:wt); 2, TEV:integrin 1:4; 3, chymotrypsin:integrin 1:100; 4, chymotrypsin:integrin 1:200; 5, chymotrypsin:integrin 1:500. (C) Crystals of the four-domain $\alpha_5\beta_1$ headpiece after microseeding.

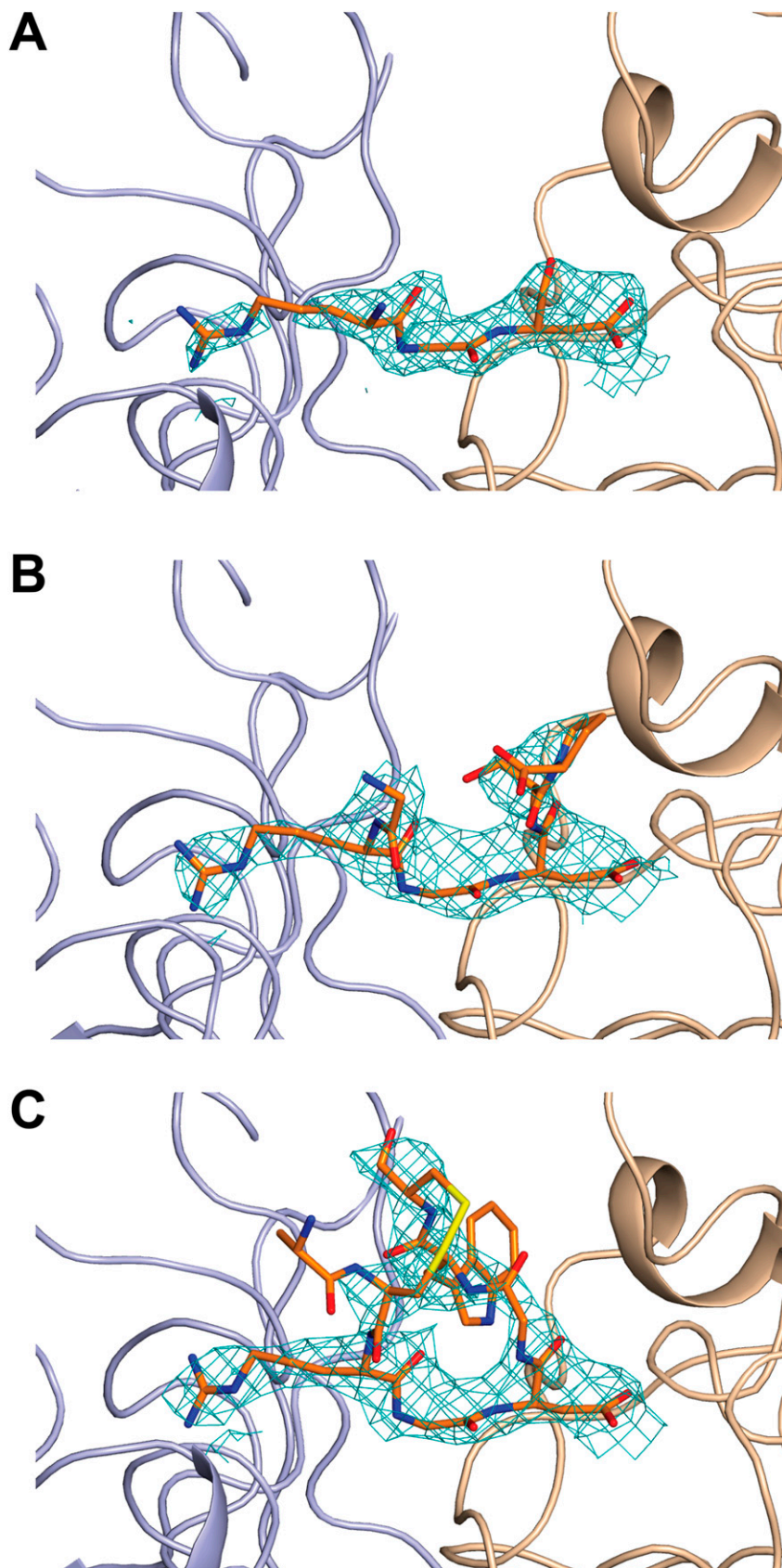


Fig. S3. Density of bound ligands. Simulated-annealing omit maps for GRGDSP peptide soaked in the presence (A) and absence (B) of Ca^{2+} and for cyclic ACRGDGWCW peptide (C). Ligands (orange) are shown in stick representation. The $2F_o - F_c$ simulated-annealing ligand omit map densities (cyan) are shown in mesh at 1σ . α_5 (light blue) and β_1 (wheat) are shown as cartoons.

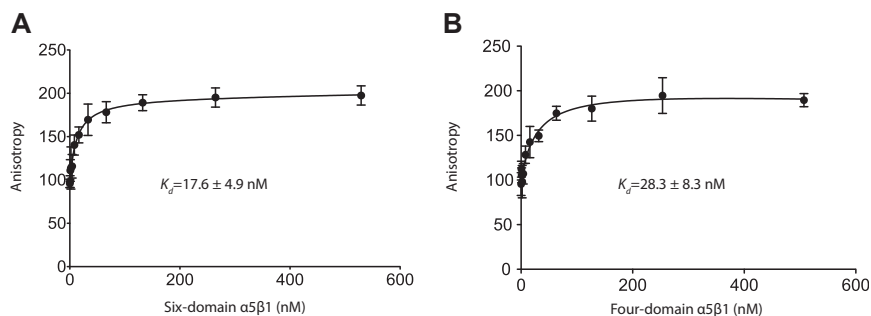


Fig. S4. Fluorescence anisotropy saturation binding. Saturation binding of six-domain $\alpha_5\beta_1$ integrin (A) and four-domain $\alpha_5\beta_1$ integrin (B) to FITC-aminohexyl-A*CRGDGWC*G cyclic peptide. Experiments were performed in 20 mM Tris-buffered saline with 1 mM Mn^{2+} and 0.1 mM Ca^{2+} (pH 7.4) with 2 nM FITC cyclic RGD peptide. Data represent mean \pm SD of triplicate samples.

Table S1. Data collection and refinement statistics

	$\alpha_5\beta_1$	$\alpha_5\beta_1$ + GRGDSP + 5 mM Mg^{2+} /1 mM Ca^{2+}	$\alpha_5\beta_1$ + GRGDSP + 5 mM Mg^{2+}	$\alpha_5\beta_1$ + ACRGDGWC + 1 mM Mg^{2+} /1 mM Ca^{2+}
Data				
Space group	P2 ₁ 2 ₁ 2 ₁	P2 ₁ 2 ₁ 2 ₁	P2 ₁ 2 ₁ 2 ₁	P2 ₁ 2 ₁ 2 ₁
Cell dimensions				
a, b, c, Å	52.1, 116.8, 159.5	61.1, 117.1, 167.1	61.0, 118.4, 170.3	57.5, 112.2, 169.6
α , β , γ , °	90, 90, 90	90, 90, 90	90, 90, 90	90, 90, 90
Resolution, Å	47.1–1.85	47.94–1.78	42.57–2.50	47.60–2.50
Reflections, total/unique	292,419/82,697	410,297/113,328	157,294/42,741	140,454/38,535
Completeness, %	98.4 (93.5)	98.2 (99.6)	98.2 (99.7)	99.3 (99.9)
$CC_{1/2}$, % [†]	99.4 (16.9)	99.4 (11.8)	99.6 (28.2)	99.6 (19.2)
CC^* , % [‡]	99.9 (53.7)	99.8 (45.9)	99.9 (60.4)	99.9 (56.8)
$I/\sigma(I)$	6.55 (0.45)	7.17 (0.34)	7.46 (0.62)	7.17 (0.66)
R_{merge} , % [§]	12.6 (210.7)	15.4 (342.2)	14.4 (276.3)	10.6 (210.0)
Redundancy	3.5 (2.7)	3.6 (3.3)	3.7 (3.7)	3.6 (3.7)
Refinement				
R_{work}	0.191 (0.397)	0.187 (0.423)	0.210 (0.403)	0.187 (0.392)
R_{free} [¶]	0.238 (0.392)	0.223 (0.468)	0.254 (0.455)	0.236 (0.388)
CC_{work} , % [#]	95.9 (39.7)	97.3 (28.8)	94.5 (53.9)	96.5 (46.8)
CC_{free} , % [#]	94.6 (36.5)	94.9 (39.8)	93.3 (48.4)	94.6 (37.8)
Ramachandran, %	96.0/3.7/0.3	96.2/3.7/0.1	95.5/4.3/0.2	93.7/5.9/0.4
Bond rmsd, Å	0.011	0.007	0.003	0.005
Angle rmsd, °	1.06	1.09	0.71	0.98
MolProbity score ^{**}	1.15 (99%)	1.24 (99%)	1.26 (100%)	1.40 (100%)
Clash score	1.38 (100%)	2.12 (99%)	1.88 (100%)	2.12 (100%)
PDB ID code	4WJK	4WK0	4WK2	4WK4

Values for highest-resolution shells are in parentheses.

[†] $CC_{1/2}$ = Pearson's correlation coefficient between average intensities of random half-datasets for each unique reflection.

[‡] $CC^* = \sqrt{2CC_{1/2}/(1+CC_{1/2})}$.

[§] $R_{merge} = \sum_i |I(i,h) - \langle I(h) \rangle| / \sum_i I(i,h)$, where $I(i,h)$ and $\langle I(h) \rangle$ are the i th and mean measurement of intensity of reflection h .

[¶] R_{free} was calculated using 5% of the data.

[#]Correlation coefficients of the observed intensities with the model-based intensities for the work (CC_{work}) and test (CC_{free}) sets.

^{||}Residues in favored, accepted, and outlier regions of the Ramachandran plot as reported by MolProbity.

^{**}MolProbity (1) score and percentile among structures of comparable resolution.

1. Chen VB, et al. (2010) MolProbity: All-atom structure validation for macromolecular crystallography. *Acta Crystallographica* D66:12–21.

