## **Contents**



## **1 Introduction**

This document supplements the paper, '**Enhancing** *in silico* **protein-based vaccine discovery for eukaryotic pathogens using predicted peptide-MHC binding and peptide conservation scores**'. It contains a compilation of *Toxoplasma gondii* proteins that are expected to make promising vaccine candidates. These proteins are used as the primary benchmark dataset to test proposed classification strategies presented in the paper.

There is currently no commercial subunit vaccine against Toxoplasmosis despite decades of research and millions of dollars spent [1]. However, the literature is full of examples of proteins observed to induce immune responses in animal models and therefore represent the type of proteins 'likely' to be worthwhile vaccine candidates. A level of caution is still warranted here because it is difficult to judge from published studies the expected efficacy of the candidates. We can only know for certain that candidates are truly worthwhile after testing in several target hosts. It is also difficult to make quantitative comparison of claimed protection levels in the studies because of differences in mouse models, ages of mice at the time-point of infection, vaccine delivery routes, adjuvants, vaccination and infection doses, parasite culture systems, challenge strains, and immunogenicity assessments [2]. For example, a candidate observed to poorly perform may give the desired protection with a different adjuvant or vaccine delivery route. Collectively, these studies indicate that the exact type and intensity of response and the immune correlates of protection are still unknown [2] but the typical type of candidate has a common theme. That is, the candidates are predominantly proteins that are naturally exposed to the immune system, such as membrane-associated proteins namely GPI-linked surface antigens (termed SAG), surface antigens known as SAG1-related sequences (SRS); and secreting proteins from specialized secretory organelles: micronemes produce MIC proteins that are involved in recognition of and adhesion to the host cell; rhoptries produce ROP proteins that drive the installation of the parasite inside the host cell; and dense granules produce GRA proteins that are involved in the maturation of the parasitophorous vacuole (PV) [3]. Three developmental stages of *T. gondii* (tachyzoite, bradyzoite and sporozoite) interact with the host immune system [1]. Therefore bradyzoite antigens (BAG) are also included in the compilation as likely candidates.

This document is in three parts: 1) Table S2-1, comprising the compiled proteins that are expected to be likely vaccine candidates with columns for Gene name, UniProt ID, Protein description, Epitope experimental evidence, Study publication reference, Machine learning (ML) classification probability for vaccine candidacy using random forest algorithm, Peptide conservation classification using predicted peptide-MHC binding and amino acid conservation scores, and Comments; 2) a brief description of some of the protein types listed in Table S2-1; and 3) Table S2-2 and S2-3, showing experimental epitopes and MHC binding information related to proteins in Table S2-1.

Some candidates in Table S2-1 have the words 'no evidence' instead of a study publication reference. This means that there is currently no publication to support the protein as a vaccine candidate. However, these proteins in question are included as likely candidates because they belong to gene families in which there are members that do have supporting immunogenic evidence. The gene families that encode *T. gondii* proteins [4] – SRS domain SAG1-like: SAG1, SAG1-related sequence 6, SAG1-related sequence 3, SAG3, SAG5D, BSR4, SAG5A, BSR4-related antigen; SRS domain SAG2-like: SAG2 (P22), SAG2B, SAG2C, SAG2D, SAG2E; Mucins and parlogs: GRA8, BAG Protein, GRA2; TRAP family: MIC2, MIC6, MIC8; EFG-like domaincontaining: MIC3, MIC6, MIC7, MIC8; TSP1 domain-containing: MIC2; Apple domain-containing: MIC4; ROP nomenclature rhoptry proteins: ROP1, ROP2, ROP4, ROP5, ROP6, ROP8, ROP10, ROP12, ROP13, ROP14, ROP15, ROP16, RON1, RON2, RON3, RON4, Rhoptry neck protein 4-like protein; and GRA nomenclature dense-granule proteins: GRA1, GRA3, GRA4, GRA5, GRA6, GRA7 (p29), GRA8, GRA9, GRA10. A universal vaccine formulation is expected to ultimately be a cocktail of immunogenic proteins that occur in multiple strains and multiple life cycle stages. Suitable adjuvants are equally important components to the formulation.

A study to be highlighted here is by Che and colleagues [5]. The study involved a comprehensive proteomic analysis of membrane proteins in *T. gondii*. In brief, three proteomics strategies were used: one-dimensional gel electrophoresis liquid chromatography-tandem mass spectrometry (1D gel LC-MS/MS), biotin labelling in conjunction with 1D gel LC-MS/MS analysis, and a novel strategy that combined three-layer 'Sandwich' Gel Electrophoresis (TLSGE) with multidimensional protein identification technology (MudPIT) [5]. The transmembrane protein clusters identified in the study were deposited in the Einstein Biodefense Proteomics Research Center (http://toro.aecom.yu.edu/cgi-bin/biodefense/main.cgi) and the data provided to ToxoDB (http://ToxodB.org), which is part of EuPathDB. Only proteins identified by all three strategies and having one or more predicted transmembrane segments were included in Table S2-1. The experimental evidence for the epitope and MHC binding information in Table S2-3 and S2-3 was extracted from the Immune Epitope Database Analysis Resource (IEDB): http://www.iedb.org/.



#### **Table S2-1. A list of proteins used in the benchmark dataset**

Abbreviation: PTM = post-translational modification

<sup>1</sup> Protein manually annotated and reviewed in UniProtKB. All other proteins are automatically annotated and not reviewed in UniProtK.

<sup>2</sup> ORF name – a name temporarily attributed to an open reading frame (ORF) by a sequencing project.  $\frac{3}{2}$ 



Abbreviation: PTM = post-translational modification

<sup>1</sup> Protein manually annotated and reviewed in UniProtKB. All other proteins are automatically annotated and not reviewed in UniProtK.

<sup>2</sup> ORF name – a name temporarily attributed to an open reading frame (ORF) by a sequencing project. 4



Abbreviation: PTM = post-translational modification

<sup>1</sup> Protein manually annotated and reviewed in UniProtKB. All other proteins are automatically annotated and not reviewed in UniProtK.

 $2^{\circ}$  ORF name – a name temporarily attributed to an open reading frame (ORF) by a sequencing project. 5

## **2 Description of protein types from Table S2-1**

Host cell invasion is the key element in the pathogenesis of Toxoplasmosis. A tachyzoite invades a host cell first by recognising host-cell surface receptors via by a family of highly abundant surface antigens (e.g. different glycoproteins, SAGs. SRSs) on its cell membrane, and then secreting proteins in a regulated and sequential progression from specialized secretory organelles in the apical complex [5]. Firstly, microneme proteins (MICs) are released to form an attachment between the apical tip of the parasite and the host cell plasma membrane (i.e. parasite-host cell adhesion). The initial attachment is randomly orientated and the tachyzoite repositions to attach its apical end against the host. Immunodominant surface antigens such SAG-related sequence 2 are involved in the attachment [84,85]. Apicomplexans are extremely polarised because secretion from the apical complex is required for invasion. Entry is through a restricted attachment called a moving junction that joins the parasite pellicle with the host. Secondly, proteins (ROPs) are secreted by rhoptries through the protruding conoid to initiate invagination of the host cell membrane. The tachyzoite penetrates by pulling the moving junction posteriorly along its pellicle. The formation of a parasitophorous vacuole (PV) membrane is created by the inversion of the host membrane as the tachyzoite penetrates in [86]. Then, proteins (GRAs) are secreted sequentially from the dense granule organelles after the parasite is fully within the PV. Mercier and colleagues provide a comprehensive review on dense granules in the context of *T. gondii* [87]. The dense granule proteins are abundantly expressed in the PV and recent data suggest that some of these proteins could be involved in building an intravacuolar membranous nanotubular network [87,88]. The exact function of this elaborate network remains unclear but is currently believed to be used to extract nutrients from the host cell. The parasite is able to shelter, grow and replicate, and utilize components from the host for its survival whilst enveloped within the PV. The release of proteins from the three secretory organelles in a sequential progression implies that their release is governed by separate signals defining distinct phases of intracellular parasitism. Upon invasion of host cells the parasite is able to partially evade the humoral and cell-mediated immune response by altering the expression and secretion of immunomodulatory cytokines or by altering the capability of immune cells. The parasite needs the survival of the host in order to develop. So in effect the parasite exists in a delicate balance of inducing and suppressing the host's immune response [89].

The following describe some of the important candidate protein types in more detail although in no particular order of importance, and are grouped into three sections: membrane-associated, secreted and miscellaneous.

#### **2.1 Membrane-associated proteins**

**SAG1, SAG2, and SAG3** are tachyzoite glycosylphosphatidylinositol (GPI)-anchored surface molecules [90] involved in host cell attachment and invasion [91] and are antigenically immunodominant [67]. SAG1/P30 was shown to elicit both humoral and cellular immune response [92] and is therefore a leading candidate for vaccine development [93] but is tachyzoite stage-specific. Using liposomes as adjuvant, purified SAG1/P30 was shown to provide protection of mice from a fatal *T. gondii* infection [62]. In another study, immune splenocytes from mice immunized with P30 appeared to lyse peritoneal macrophages infected with *T. gondii* [64]. Mice primed with recombinant influenza virus and boosted with a recombinant adenovirus encoding SAG2 elicited

both humoral and cellular immune responses specific for SAG2 [71]. A compound DNA vaccine encoding *T. gondii* antigens SAG1, SAG3 with CTXA(2)/B gene was shown to effectively enhance the humoral and cellular immune response and prolong survival time in vaccinated mice [76].

The surface of *T. gondii* is coated with developmentally expressed, GPI-linked proteins structurally related to SAG1. Collectively, these surface antigens are known as the SRS (SAG1-related sequences) superfamily of proteins [94]. **SRS2** [85] is localised on both bradyzoites and tachyzoites [95]. The SRS2 protein is involved in the host cell invasion process [96] and polyclonal and monoclonal antibodies directed against it were shown to inhibit invasion of placental ovine trophoblasts *in vitro* [97]. **SRS domain containing proteins** are present in large numbers on the parasite surface and facilitate the invasion of multiple host and cell types [94]. They are considered to be extremely immunogenic in *Toxoplasma* [23].

**RON** proteins originate from the neck of the rhoptries. All RON proteins have been demonstrated to be present at the moving junction between the apex of Apicomplexa and the host cell membrane that moves along the parasite and serves as support to propel it inside the host cell [35]. The moving junction assembly is initiated by injection of RONs into the host cell, where RON2 spans the membrane and functions as a receptor for apical membrane antigen 1 (AMA1) on the parasite [38]. The proteomics Che study included them as members of transmembrane proteins. Interestingly, the five prediction programs in the paper typically indicate RONs as both membrane-associated and secreted.

#### **2.2 Secreted proteins**

**GRA proteins** are involved in the cellular invasion process. Dense granules are secretory vesicles that play a major role in the structural modifications of the parasitophorous vacuole (PV) in which the parasite develops [98].

Both humoral and cellular immune responses against *T. gondii* was detected in sheep immunized with DNA plasmids encoding *T. gondii* GRA7 formulated in an adjuvant formulation [99]. Studies using antibodies to immunolocalize the *T. gondii* dense granule protein GRA3 have shown that this protein associates strongly with the parasitophorous vacuole membrane (PVM) i.e. GRA3 has an N-terminal secretory signal sequence and a transmembrane domain consistent with its insertion into the PVM. GRA3 possesses a dilysine 'KKXX' endoplasmic reticulum (ER) retrieval motif that interacts with PVM and the calcium modulating ligand of host cell ER in the parasitism of *T. gondii* [9,10]. The five prediction programs indicate that GRA3, and most other dense granule proteins described here, are both membrane-associated and secreted. GRA2 and GRA4 are not predicted to be membrane-associated.

**MIC proteins** are discharged by exocytosis during the attachment to the host cell surface to facilitate cell invasion [100]. Many microneme proteins also contain well-conserved functional domains associated with mainly adhesive activity (e.g. EGF-like and PAN\_1 domains) and some protease activity (e.g. Peptidase\_S8 and Rhomboid) [101].

MIC3 is expressed in all three infectious stages of *T. gondii* (tachyzoites, bradyzoites, and sporozoites). A DNA vaccine encoding the MIC3 protein has been demonstrated to elicit a strong specific immune response providing significant protection against *T. gondii* infection [47].

**ROP proteins** are involved in a variety of cellular functions related to host cell invasion, formation of the parasitophorous vacuole, and parasite-host cell interplay [102]. The protein combinations of rROP2 + rROP4 + rGRA4 and rROP2 + rROP4 + rSAG1 were shown to be very effective in the development of a high level of protection irrespective of the genetic backgrounds and innate resistance to toxoplasmosis of the laboratory mice [11]. A DNA vaccine encoding the ROP1 antigen of *T.gondii* and ovine CD154 was demonstrated to stimulate humoral and cellular immune responses in sheep. The intramuscular injection of pROP1 only induced a Th1 specific immune response [19]. ROP2/P64 expresses in all three life cycle stages. ROP2, is involved in invasion of host cells, induces humoral immune response [22,23].

#### **2.3 Miscellaneous**

The following proteins were included in the benchmark dataset because they were mentioned in published studies as possible vaccine candidates. The first notable fact about these proteins is that they are not expected to be naturally exposed to the immune system.

**Phosphoglycerate mutase** (pgam) was identified using mass spectrometry as one of three main proteins in the excretory secretory antigen (ESA) [83]. The other two proteins were MIC10 and GRA7. The study authors conclude that these three proteins demonstrated good immunogenicity and may potentially be useful in the development of vaccines against toxoplasmosis either used singly or in various combinations. The results shown in Supporting Information S3 suggest that pgam is neither secreted nor membrane-associated and there is no consensus for vaccine candidacy.

A study [60] demonstrated that protective humoral and cellular immunity against experimental toxoplasmosis in susceptible Kunming mice was induced by a DNA vaccine encoding the **perforin-like protein 1** (PLP1). Furthermore, another study [52] showed that the immune efficacy induced by DNA vaccine with MIC6 and PLP1 was better than that induced by PLP1 or MIC6 alone. The results shown in Supporting Information S3 suggest that PLP1 is neither secreted nor membrane-associated but there is consensus from the peptide-MHC binding strategies.

**Nucleoside triphosphate hydrolase** (NTPase) is released from dense granules and accumulates as a soluble protein in the vacuolar space [61]. A study [61] described that a recombinant form of NTPase co-administered with the adjuvant alum induced a strong specific Th1 immune response against toxoplasmosis in a murine model. The results shown in Supporting Information S3 suggest that NTP1 is a secreted protein but there is no consensus for vaccine candidacy.

# **3 Epitope and MHC binding evidence**









### **Table S2-3. Experimentally validated peptide-MHC I binding related to benchmark proteins in Table S2-1**











## **4 REFERENCES**

1. Kur J, Holec-Gasior L, Hiszczynska-Sawicka E (2009) Current status of toxoplasmosis vaccine development. Expert Review of Vaccines 8: 791-808.

2. Goodswen SJ, Kennedy PJ, Ellis JT (2014) Discovering a vaccine against neosporosis using computers: is it feasible? Trends in Parasitology 30: 401-411.

3. Chen Z, Harb OS, Roos DS (2008) *In Silico* Identification of Specialized Secretory-Organelle Proteins in Apicomplexan Parasites and *In Vivo* Validation in *Toxoplasma gondii*. PLoS ONE 3: e3611.

4. Louis M. Weiss KK (2014) Toxoplasma Gondii: The Model Apicomplexan - Perspectives and Methods: Academic Press.

5. Che F-Y, Madrid-Aliste C, Burd B, Zhang H, Nieves E, et al. (2010) Comprehensive proteomic analysis of membrane proteins in toxoplasma gondii. Molecular & Cellular Proteomics 10: M110 000745.

6. Jongert E, Melkebeek V, De Craeye S, Dewit J, Verhelst D, et al. (2008) An enhanced GRA1-GRA7 cocktail DNA vaccine primes anti-Toxoplasma immune responses in pigs. Vaccine 26: 1025-1031.

7. Wu X-N, Lin J, Lin X, Chen J, Chen Z-L, et al. (2012) Multicomponent DNA vaccine-encoding Toxoplasma gondii GRA1 and SAG1 primes: anti-Toxoplasma immune response in mice. Parasitology Research 111: 2001- 2009.

8. Xue M, He S, Cui Y, Yao Y, Wang H (2008) Evaluation of the immune response elicited by multi-antigenic DNA vaccine expressing SAG1, ROP2 and GRA2 against Toxoplasma gondii. Parasitology International 57: 424-429.

9. Henriquez FL, Nickdel MB, McLeod R, Lyons RE, Lyons K, et al. (2005) Toxoplasma gondii dense granule protein 3 (GRA3) is a type I transmembrane protein that possesses a cytoplasmic dilysine (KKXX) endoplasmic reticulum (ER) retrieval motif. Parasitology 131: 169-179.

10. Kim JY, Ahn H-J, Ryu KJ, Nam H-W (2008) Interaction between Parasitophorous Vacuolar Membraneassociated GRA3 and Calcium Modulating Ligand of Host Cell Endoplasmic Reticulum in the Parasitism of Toxoplasma gondii. Korean Journal of Parasitology 46: 209-216.

11. Dziadek B, Gatkowska J, Grzybowski M, Dziadek J, Dzitko K, et al. (2012) Toxoplasma gondii: The vaccine potential of three trivalent antigen-cocktails composed of recombinant ROP2, ROP4, GRA4 and SAG1 proteins against chronic toxoplasmosis in BALB/c mice. Experimental Parasitology 131: 133-138.

12. Igarashi M, Kano F, Tamekuni K, Machado RZ, Navarro IT, et al. (2008) Toxoplasma gondii: Evaluation of an intranasal vaccine using recombinant proteins against brain cyst formation in BALB/c mice. Experimental Parasitology 118: 386-392.

13. Hiszczynska-Sawicka E, Oledzka G, Holec-Gasior L, Li H, Xu JB, et al. (2011) Evaluation of immune responses in sheep induced by DNA immunization with genes encoding GRA1, GRA4, GRA6 and GRA7 antigens of Toxoplasma gondii. Veterinary Parasitology 177: 281-289.

14. Sun X-M, Zou J, Elashram SAA, Yan W-C, Liu X-Y, et al. (2011) DNA vaccination with a gene encoding Toxoplasma gondii GRA6 induces partial protection against toxoplasmosis in BALB/c mice. Parasites & Vectors 4.

15. Feliu V, Vasseur V, Grover HS, Chu HH, Brown MJ, et al. (2013) Location of the CD8 T Cell Epitope within the Antigenic Precursor Determines Immunogenicity and Protection against the Toxoplasma gondii Parasite. Plos Pathogens 9.

16. El Bissati K, Zhou Y, Dasgupta D, Cobb D, Dubey JP, et al. (2014) Effectiveness of a novel immunogenic nanoparticle platform for Toxoplasma peptide vaccine in HLA transgenic mice. Vaccine 32: 3243-3248.

17. Liu Q, Wang F, Wang G, Zhao Q, Min J, et al. (2014) Toxoplasma gondii Immune response and protective efficacy induced by ROP16/GRA7 multicomponent DNA vaccine with a genetic adjuvant B7-2. Human Vaccines & Immunotherapeutics 10: 184-191.

18. Lu B, Wu ST, Shi Y, Zhang RL, Zou LJ, et al. (2006) Toxoplasma gondii: Expression pattern and detection of infection using full-length recombinant P35 antigen. Experimental Parasitology 113: 83-90.

19. Hiszczynska-Sawicka E, Li H, Xu JB, Holec-Gasior L, Kur J, et al. (2011) Modulation of immune response to Toxoplasma gondii in sheep by immunization with a DNA vaccine encoding ROP1 antigen as a fusion protein with ovine CD154. Veterinary Parasitology 183: 72-78.

20. Ossorio PN, Schwartzman JD, Boothroyd JC (1992) A Toxoplasma-gondii rhoptry protein associated with host-cell penetration has unusual charge asymmetry. Molecular and Biochemical Parasitology 50: 1-16.

21. Debache K, Alaeddine F, Guionaud C, Monney T, Mueller J, et al. (2009) Vaccination with recombinant NcROP2 combined with recombinant NcMIC1 and NcMIC3 reduces cerebral infection and vertical transmission in mice experimentally infected with Neospora caninum tachyzoites. International Journal for Parasitology 39: 1373-1384.

22. Li W-S, Chen Q-X, Ye J-X, Xie Z-X, Chen J, et al. (2011) Comparative evaluation of immunization with recombinant protein and plasmid DNA vaccines of fusion antigen ROP2 and SAG1 from Toxoplasma gondii in mice: cellular and humoral immune responses. Parasitology Research 109: 637-644.

23. Rocchi MS, Bartley PM, Inglis NF, Collantes-Fernandez E, Entrican G, et al. (2011) Selection of Neospora caninum antigens stimulating bovine CD4(+ve) T cell responses through immuno-potency screening and proteomic approaches. Veterinary Research 42: 1-91.

24. Carey KL, Jongco AM, Kam K, Ward GE (2004) The Toxoplasma gondii rhoptry protein ROP4 is secreted into the parasitophorous vacuole and becomes phosphorylated in infected cells. Eukaryotic Cell 3: 1320-1330.

25. Zheng B, Lu S, Tong Q, Kong Q, Lou D (2013) The virulence-related rhoptry protein 5 (ROP5) of Toxoplasma Gondii is a novel vaccine candidate against toxoplasmosis in mice. Vaccine 31: 4578-4584.

26. Zhao H-G, Fan Z-G, Huang F-Y, Huang Y-H, Zhou S-L, et al. (2013) Immune response in mice induced by complex gene vaccine pcSAG1-ROP5 of Toxoplasma gondii. Zhongguo ji sheng chong xue yu ji sheng chong bing za zhi = Chinese journal of parasitology  $\&$  parasitic diseases 31: 1-5.

27. Parthasarathy S, Fong MY, Ramaswamy K, Lau YL (2013) Protective Immune Response in BALB/c Mice Induced by DNA Vaccine of the ROP8 gene of Toxoplasma gondii. American Journal of Tropical Medicine and Hygiene 88: 883-887.

28. Chen J, Zhou D-H, Li Z-Y, Petersen E, Huang S-Y, et al. (2014) Toxoplasma gondii: Protective immunity induced by rhoptry protein 9 (TgROP9) against acute toxoplasmosis. Experimental Parasitology 139: 42-48.

29. Wang P-Y, Yuan Z-G, Petersen E, Li J, Zhang X-X, et al. (2012) Protective Efficacy of a Toxoplasma gondii Rhoptry Protein 13 Plasmid DNA Vaccine in Mice. Clinical and Vaccine Immunology 19: 1916-1920.

30. Yuan Z-G, Zhang X-X, He X-H, Petersen E, Zhou D-H, et al. (2011) Protective Immunity Induced by Toxoplasma gondii Rhoptry Protein 16 against Toxoplasmosis in Mice. Clinical and Vaccine Immunology 18: 119-124.

31. Zhao Y, Yap GS (2014) Toxoplasma's Arms Race with the Host Interferon Response: A Menage a Trois of ROPs. Cell Host & Microbe 15: 517-518.

32. Cheng L, Chen Y, Chen L, Shen Y, Shen J, et al. (2012) Interactions between the ROP18 kinase and host cell proteins that aid in the parasitism of Toxoplasma gondii. Acta Tropica 122: 255-260.

33. El Hajj H, Lebrun M, Arold ST, Vial H, Labesse G, et al. (2007) ROP18 is a rhoptry kinase controlling the intracellular proliferation of Toxoplasma gondii. Plos Pathogens 3: 200-211.

34. Niedelman W, Gold DA, Rosowski EE, Sprokholt JK, Lim D, et al. (2012) The rhoptry proteins ROP18 and ROP5 mediate Toxoplasma gondii evasion of the murine, but not the human, interferon-gamma response. Plos Pathogens 8: e1002784-e1002784.

35. Besteiro S, Michelin A, Poncet J, Dubremetz J-F, Lebrun M (2009) Export of a Toxoplasma gondii Rhoptry Neck Protein Complex at the Host Cell Membrane to Form the Moving Junction during Invasion. Plos Pathogens 5.

36. Bradley PJ, Ward C, Cheng SJ, Alexander DL, Coller S, et al. (2005) Proteomic analysis of rhoptry organelles reveals many novel constituents for host-parasite interactions in Toxoplasma gondii. Journal of Biological Chemistry 280: 34245-34258.

37. Moreno-Perez DA, Montenegro M, Patarroyo ME, Patarroyo MA (2011) Identification, characterization and antigenicity of the Plasmodium vivax rhoptry neck protein 1 (PvRON1). Malaria Journal 10.

38. Tonkin ML, Roques M, Lamarque MH, Pugniere M, Douguet D, et al. (2011) Host Cell Invasion by Apicomplexan Parasites: Insights from the Co-Structure of AMA1 with a RON2 Peptide. Science 333: 463-467.

39. Poukchanski A, Fritz HM, Tonkin ML, Treeck M, Boulanger MJ, et al. (2013) Toxoplasma gondii Sporozoites Invade Host Cells Using Two Novel Paralogues of RON2 and AMA1. Plos One 8.

40. Ito D, Han E-T, Takeo S, Thongkukiatkul A, Otsuki H, et al. (2011) Plasmodial ortholog of Toxoplasma gondii rhoptry neck protein 3 is localized to the rhoptry body. Parasitology International 60: 132-138.

41. Takemae H, Sugi T, Kobayashi K, Gong H, Ishiwa A, et al. (2013) Characterization of the interaction between Toxoplasma gondii rhoptry neck protein 4 and host cellular beta-tubulin. Scientific Reports 3.

42. Curtidor H, Patino LC, Arevalo-Pinzon G, Patarroyo ME, Patarroyo MA (2011) Identification of the Plasmodium falciparum rhoptry neck protein 5 (PfRON5). Gene 474: 22-28.

43. Curtidor H, Patino LC, Arevalo-Pinzon G, Vanegas M, Patarroyo ME, et al. (2014) Plasmodium falciparum rhoptry neck protein 5 peptides bind to human red blood cells and inhibit parasite invasion. Peptides 53: 210- 217.

44. Beck JR, Fung C, Straub KW, Coppens I, Vashisht AA, et al. (2013) A Toxoplasma Palmitoyl Acyl Transferase and the Palmitoylated Armadillo Repeat Protein TgARO Govern Apical Rhoptry Tethering and Reveal a Critical Role for the Rhoptries in Host Cell Invasion but Not Egress. Plos Pathogens 9.

45. Ortega-Barria E, Boothroyd JC (1999) A Toxoplasma lectin-like activity specific for sulfated polysaccharides is involved in host cell infection. Journal of Biological Chemistry 274: 1267-1276.

46. Garcia-Reguet N, Lebrun M, Fourmaux MN, Mercereau-Puijalon O, Mann T, et al. (2000) The microneme protein MIC3 of Toxoplasma gondii is a secretory adhesin that binds to both the surface of the host cells and the surface of the parasite. Cellular Microbiology 2: 353-364.

47. Ismael AB, Sekkai D, Collin C, Bout D, Mevelec MN (2003) The MIC3 gene of Toxoplasma gondii is a novel potent vaccine candidate against toxoplasmosis. Infection and Immunity 71: 6222-6228.

48. Lourenco EV, Panunto-Castelo A, Molfetta JB, Avanci NC, Goldman MHS, et al. (2006) Immunization with MIC1 and MIC4 induces protective immunity against toxoplasma gondii. Glycobiology 16: 1112-1112.

49. Peng GH, Yuan ZG, Zhou DH, He XH, Yan C, et al. (2010) Sequence Variation in Toxoplasma gondii MIC4 Gene and Protective Effect of an MIC4 DNA Vaccine in a Murine Model Against Toxoplasmosis. Journal of Animal and Veterinary Advances 9: 1463-1468.

50. Wang H, He S, Yao Y, Cong H, Zhao H, et al. (2009) Toxoplasma gondii: Protective effect of an intranasal SAG1 and MIC4 DNA vaccine in mice. Experimental Parasitology 122: 226-232.

51. Peng G-H, Yuan Z-G, Zhou D-H, He X-H, Liu M-M, et al. (2009) Toxoplasma gondii microneme protein 6 (MIC6) is a potential vaccine candidate against toxoplasmosis in mice. Vaccine 27: 6570-6574.

52. Yan H-K, Yuan Z-G, Song H-Q, Petersen E, Zhou Y, et al. (2012) Vaccination with a DNA Vaccine Coding for Perforin-Like Protein 1 and MIC6 Induces Significant Protective Immunity against Toxoplasma gondii. Clinical and Vaccine Immunology 19: 684-689.

53. Liu MM, Yuan ZG, Peng GH, Zhou DH, He XH, et al. (2010) Toxoplasma gondii microneme protein 8 (MIC8) is a potential vaccine candidate against toxoplasmosis. Parasitology Research 106: 1079-1084.

54. Yao Y, He S-y, Wang H-x, Zhou H-y, Zhao H, et al. (2010) Protective immunity induced in mice by multiantigenic DNA vaccine with genes encoding SAG1 and MIC8 of Toxoplasma gondii. Zhongguo ji sheng chong xue yu ji sheng chong bing za zhi = Chinese journal of parasitology  $\&$  parasitic diseases 28: 81-88.

55. Zhao H-G, Huang F-Y, Guo J-L, Tan G-H (2013) Evaluation on the immune response induced by DNA vaccine encoding MIC8 co-immunized with IL-12 genetic adjuvant against Toxoplasma gondii infection. Zhongguo ji sheng chong xue yu ji sheng chong bing za zhi = Chinese journal of parasitology  $\&$  parasitic diseases 31: 284-289.

56. Li Z-Y, Chen J, Petersen E, Zhou D-H, Huang S-Y, et al. (2014) Synergy of mIL-21 and mIL-15 in enhancing DNA vaccine efficacy against acute and chronic Toxoplasma gondii infection in mice. Vaccine 32: 3058-3065.

57. Harper JM, Zhou XW, Pszenny V, Kafsack BFC, Carruthers VB (2004) The novel coccidian micronemal protein MIC11 undergoes proteolytic maturation by sequential cleavage to remove an internal propeptide. International Journal for Parasitology 34: 1047-1058.

58. Ren D, Zhou D-H, Xu M-J, Zhou Y, Yang J-F, et al. (2012) Sequence variation in Toxoplasma gondii MIC13 gene among isolates from different hosts and geographical locations. African Journal of Microbiology Research 6: 3265-3269.

59. Yuan Z-G, Ren D, Zhou D-H, Zhang X-X, Petersen E, et al. (2013) Evaluation of protective effect of pVAX-TgMIC13 plasmid against acute and chronic Toxoplasma gondii infection in a murine model. Vaccine 31: 3135-3139.

60. Yan H-K, Yuan Z-G, Petersen E, Zhang X-X, Zhou D-H, et al. (2011) Toxoplasma gondii: Protective immunity against experimental toxoplasmosis induced by a DNA vaccine encoding the perforin-like protein 1. Experimental Parasitology 128: 38-43.

61. Tan F, Hu X, Luo F-J, Pan C-W, Chen X-G (2011) Induction of protective Th1 immune responses in mice by vaccination with recombinant Toxoplasma gondii nucleoside triphosphate hydrolase-II. Vaccine 29: 2742- 2748.

62. Bulow R, Boothroyd JC (1991) Protection of mice from fatal *Toxoplasma-gondii* infection by immunization with p-30 antigen in liposomes. Journal of Immunology 147: 3496-3500.

63. Fux B, Rodrigues CV, Portela RW, Silva NM, Su CL, et al. (2003) Role of cytokines and major histocompatibility complex restriction in mouse resistance to infection with a natural recombinant strain (type I-III) of Toxoplasma gondii. Infection and Immunity 71: 6392-6401.

64. Kasper LH, Khan IA, Ely KH, Buelow R, Boothroyd JC (1992) Antigen-specific (p-30) mouse CD8+ Tcells are cytotoxic against Toxoplasma-gondii-infected peritoneal-macrophages. Journal of Immunology 148: 1493-1498.

65. Khosroshahi KH, Ghaffarifar F, D'Souza S, Sharifi Z, Dalimi A (2011) Evaluation of the immune response induced by DNA vaccine cocktail expressing complete SAG1 and ROP2 genes against toxoplasmosis. Vaccine 29: 778-783.

66. Haldorson GJ, Mathison BA, Wenberg K, Conrad PA, Dubey JP, et al. (2005) Immunization with native surface protein NcSRS2 induces a Th2 immune response and reduces congenital Neospora caninum transmission in mice. International Journal for Parasitology 35: 1407-1415.

67. Howe DK, Crawford AC, Lindsay D, Sibley LD (1998) The p29 and p35 immunodominant antigens of Neospora caninum tachyzoites are homologous to the family of surface antigens of Toxoplasma gondii. Infection and Immunity 66: 5322-5328.

68. Wasmuth JD, Pszenny V, Haile S, Jansen EM, Gast AT, et al. (2012) Integrated Bioinformatic and Targeted Deletion Analyses of the SRS Gene Superfamily Identify SRS29C as a Negative Regulator of Toxoplasma Virulence. Mbio 3.

69. Lau YL, Fong MY (2008) Toxoplasma gondii: Serological characterization and immunogenicity of recombinant surface antigen 2 (SAG2) expressed in the yeast Pichia pastoris. Experimental Parasitology 119: 373-378.

70. Li J-h, Wu S-t, Weng Y-b, Gan Y, Liu H-b, et al. (2007) Study on immuno-effect with GRA4 or SAG2 gene recombinant BCG vaccine of Toxoplasma gondii. Zhongguo ji sheng chong xue yu ji sheng chong bing za zhi = Chinese journal of parasitology & parasitic diseases 25: 217-221.

71. Machado AV, Caetano BC, Barbosa RP, Salgado APC, Rabelo RH, et al. (2010) Prime and boost immunization with influenza and adenovirus encoding the Toxoplasma gondii surface antigen 2 (SAG2) induces strong protective immunity. Vaccine 28: 3247-3256.

72. Lekutis C, Ferguson DJP, Grigg ME, Camps M, Boothroyd JC (2001) Surface antigens of Toxoplasma gondii: variations on a theme. International Journal for Parasitology 31: 1285-1292.

73. Lekutis C, Ferguson DJP, Boothroyd JC (2000) Toxoplasma gondii: Identification of a developmentally regulated family of genes related to SAG2. Experimental Parasitology 96: 89-96.

74. Zhang M, Zhao L, Song J, Li Y, Zhao Q, et al. (2013) DNA vaccine encoding the Toxoplasma gondii bradyzoite-specific surface antigens SAG2CDX protect BALB/c mice against type II parasite infection. Vaccine 31: 4536-4540.

75. Cong H, Yuan Q, Zhao Q, Zhao L, Yin H, et al. (2014) Comparative efficacy of a multi-epitope DNA vaccine via intranasal, peroral, and intramuscular delivery against lethal Toxoplasma gondii infection in mice. Parasites & Vectors 7.

76. Cong H, Zhang M, Xin Q, Wang Z, Li Y, et al. (2013) Compound DNA vaccine encoding SAG1/SAG3 with A(2)/B subunit of cholera toxin as a genetic adjuvant protects BALB/c mice against Toxoplasma gondii. Parasites & Vectors 6.

77. Spano F, Ricci I, Di Cristina M, Possenti A, Tinti M, et al. (2002) The SAG5 locus of Toxoplasma gondii encodes three novel proteins belonging to the SAG1 family of surface antigens. International Journal for Parasitology 32: 121-131.

78. Tinti M, Possenti A, Cherchi S, Barca S, Spano F (2003) Analysis of the SAG5 locus reveals a distinct genomic organisation in virulent and avirulent strains of Toxoplasma gondii. International Journal for Parasitology 33: 1605-1616.

79. McAllister MM, Parmley SF, Weiss LM, Welch YJ, McGuire AM (1996) An immunohistochemical method for detecting bradyzoite antigen (BAGS) in Toxoplasma gondii-infected tissues cross-reacts with a Neospora caninum bradyzoite antigen. Journal of Parasitology 82: 354-355.

80. Wang Q, Wu K, Chen X-g, Hao L, Cheng L (2007) Cloning and expression of a bradyzoite-specific gene of Toxoplasma gondii and immunoreactive analysis on the recombinant antigen. Zhongguo ji sheng chong xue yu ji sheng chong bing za zhi = Chinese journal of parasitology & parasitic diseases 25: 295-299.

81. Crawford J, Grujic O, Bruic E, Czjzek M, Grigg ME, et al. (2009) Structural Characterization of the Bradyzoite Surface Antigen (BSR4) from Toxoplasma gondii, a Unique Addition to the Surface Antigen Glycoprotein 1-related Superfamily. Journal of Biological Chemistry 284: 9192-9198.

82. Yin L-T, Wang F, Meng X-L, Wang H-L, Liu H-L, et al. (2012) Cloning, expression and antigenicity analysis of phosphoglycerate mutase 2 gene of Toxoplasma gondii. Zhongguo ji sheng chong xue yu ji sheng chong bing za zhi = Chinese journal of parasitology  $\&$  parasitic diseases 30: 86-89.

83. Saadatnia G, Mohamed Z, Ghaffarifar F, Osman E, Moghadam ZK, et al. (2012) Toxoplasma gondii excretory secretory antigenic proteins of diagnostic potential. Apmis 120: 47-55.

84. Hemphill A, Gottstein B, Kaufmann H (1996) Adhesion and invasion of bovine endothelial cells by Neospora caninum. Parasitology 112: 183-197.

85. Howe DK, Sibley LD (1999) Comparison of the major antigens of Neospora caninum and Toxoplasma gondii. International Journal for Parasitology 29: 1489-1496.

86. Carruthers V, Boothroyd JC (2007) Pulling together: an integrated model of Toxoplasma cell invasion. Current Opinion in Microbiology 10: 83-89.

87. Mercier C, Adjogble KDZ, Daubener W, Delauw MFC (2005) Dense granules: Are they key organelles to help understand the parasitophorous vacuole of all apicomplexa parasites? International Journal for Parasitology 35: 829-849.

88. Braun L, Travier L, Kieffer S, Musset K, Garin J, et al. (2008) Purification of Toxoplasma dense granule proteins reveals that they are in complexes throughout the secretory pathway. Molecular and Biochemical Parasitology 157: 13-21.

89. Lang C, Gross U, Lueder CGK (2007) Subversion of innate and adaptive immune responses by Toxoplasma Gondii. Parasitology Research 100: 191-203.

90. Sonda S, Fuchs N, Connolly B, Fernandez P, Gottstein B, et al. (1998) The major 36 kDa Neospora caninum tachyzoite surface protein is closely related to the major Toxoplasma gondii surface antigen. Molecular and Biochemical Parasitology 97: 97-108.

91. Hemphill A, Fuchs N, Sonda S, Gottstein B, Hentrich B (1997) Identification and partial characterization of a 36 kDa surface protein on Neospora caninum tachyzoites. Parasitology 115: 371-380.

92. Nielsen HV, Lauemoller SL, Christiansen L, Buus S, Fomsgaard A, et al. (1999) Complete protection against lethal Toxoplasma gondii infection in mice immunized with a plasmid encoding the SAG1 gene. Infection and Immunity 67: 6358-6363.

93. Liu K-Y, Zhang D-B, Wei Q-K, Li J, Li G-P, et al. (2006) Biological role of surface Toxoplasma gondii antigen in development of vaccine. World Journal of Gastroenterology 12: 2363-2368.

94. Jung C, Lee CYF, Grigg ME (2004) The SRS superfamily of Toxoplasma surface proteins. International Journal for Parasitology 34: 285-296.

95. Hemphill A (1996) Subcellular localization and functional characterization of Nc-p43, a major Neospora caninum tachyzoite surface protein. Infection and Immunity 64: 4279-4287.

96. Hemphill A, Felleisen R, Connolly B, Gottstein B, Hentrich B, et al. (1997) Characterization of a cDNAclone encoding Nc-p43, a major Neospora caninum tachyzoite surface protein. Parasitology 115: 581-590.

97. Haldorson GJ, Stanton JB, Mathison BA, Suarez CE, Baszler TV (2006) Neospora caninum: Antibodies directed against tachyzoite surface protein NcSRS2 inhibit parasite attachment and invasion of placental trophoblasts in vitro. Experimental Parasitology 112: 172-178.

98. Ellis JT, Ryce C, Atkinson R, Balu S, Jones P, et al. (2000) Isolation, characterization and expression of a GRA2 homologue from Neospora caninum. Parasitology 120: 383-390.

99. Hiszczynska-Sawicka E, Li H, Xu JB, Oledzka G, Kur J, et al. (2010) Comparison of immune response in sheep immunized with DNA vaccine encoding Toxoplasma gondii GRA7 antigen in different adjuvant formulations. Experimental Parasitology 124: 365-372.

100. Dubremetz JF, Garcia-Reguet N, Conseil V, Fourmaux MN (1998) Apical organelles and host-cell invasion by Apicomplexa. International Journal for Parasitology 28: 1007-1013.

101. Chen Z, Harb OS, Roos DS (2008) In Silico Identification of Specialized Secretory-Organelle Proteins in Apicomplexan Parasites and In Vivo Validation in Toxoplasma gondii. PLoS ONE 3.

102. Debache K, Guionaud C, Alaeddine F, Mevissen M, Hemphill A (2008) Vaccination of mice with recombinant NcROP2 antigen reduces mortality and cerebral infection in mice infected with Neospora caninum tachyzoites. International Journal for Parasitology 38: 1455-1463.