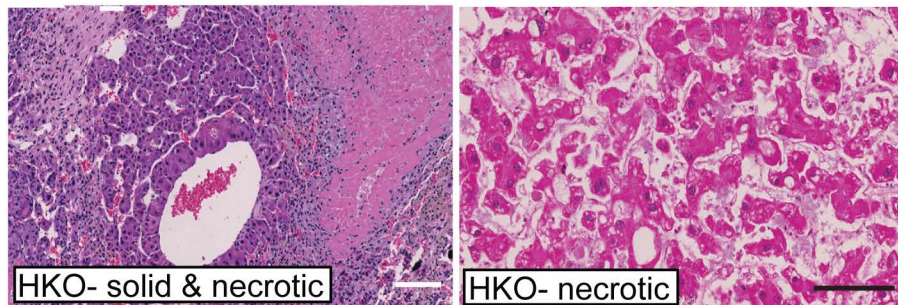
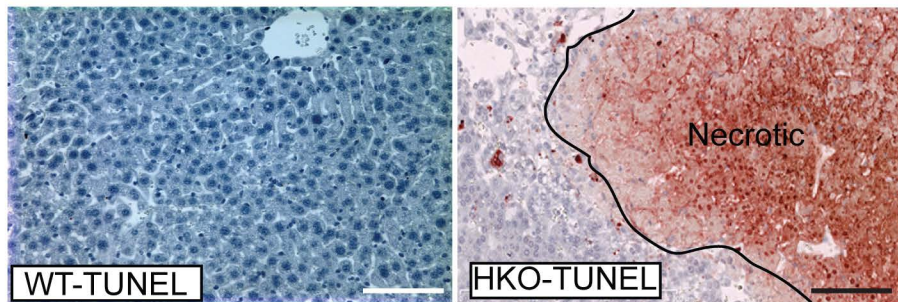


A



B



C

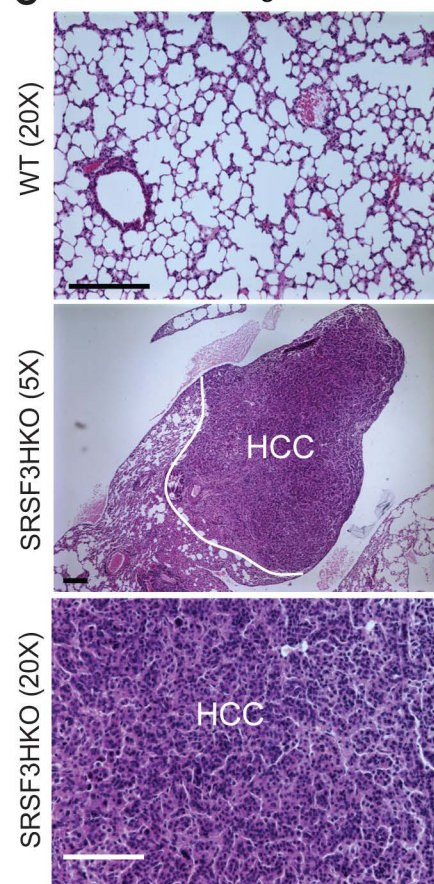


Figure S1. (A) Representative 20X images of liver sections from 24 month old SRSF3HKO mice stained with H&E. The SRSF3HKO section shows a dense cellular tumor with irregular nuclei and morphological pleomorphism and areas of necrosis. (B) TUNEL staining of necrotic tumor in SRSF3HKO liver. (C) Representative lung sections from WT and SRSF3HKO mice stained with H&E. Metastatic HCC in lung is marked by white line. For A, B, C, scale bar represents 100 μ m.

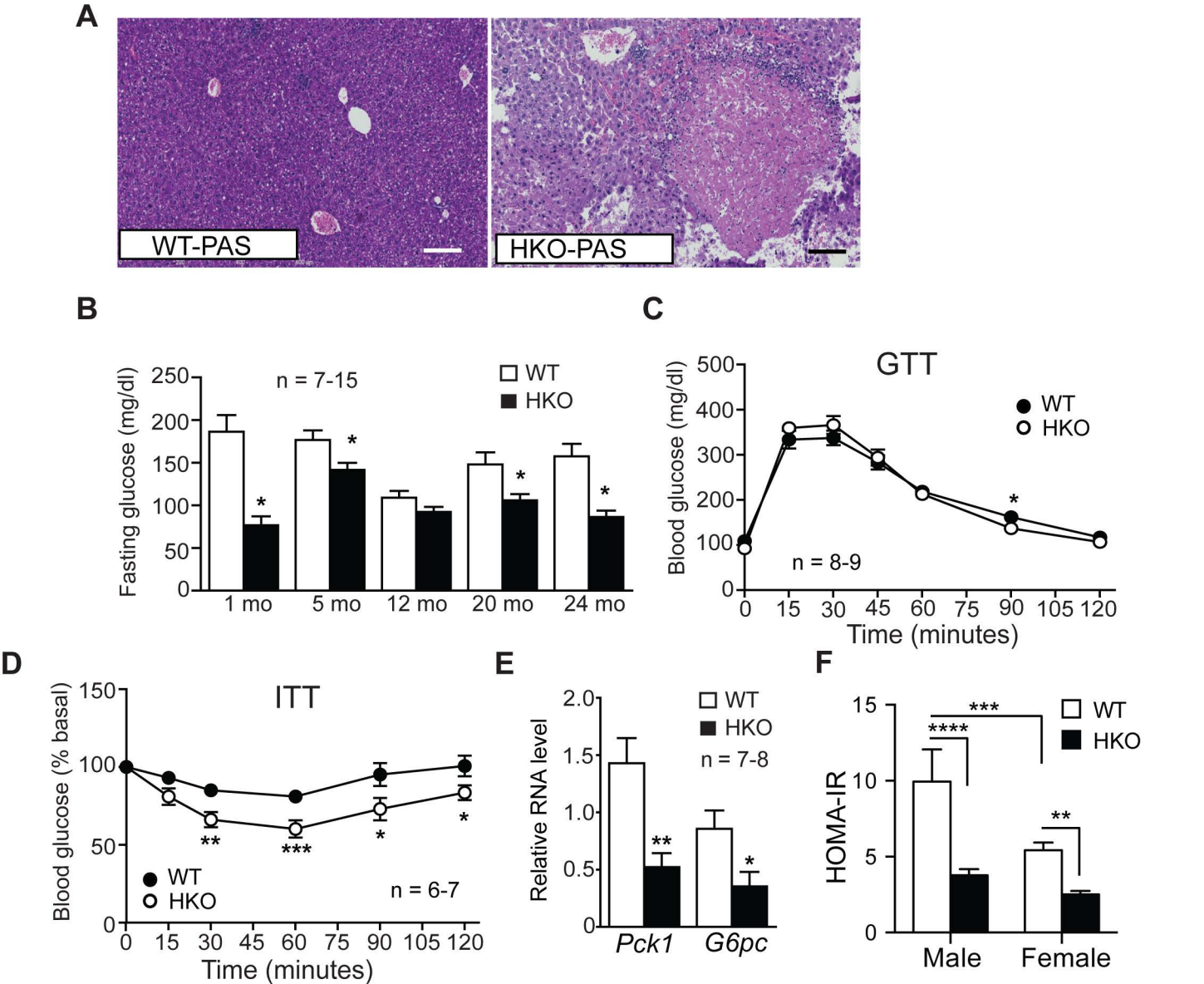


Figure S2. (A) Representative 10X images of liver sections from 24 month old WT and SRSF3-HKO mice stained for glycogen with PAS reagent. SRSF3-HKO livers have reduced glycogen in non-tumorous areas and glycogen is absent from tumors. Scale bar represents 100 μ m. (B) Fasting blood glucose values with aging showing that SRSF3-HKO mice are hypoglycemic compared to WT. (C) Glucose tolerance tests on 12 month old mice showing comparable glucose tolerance in WT and SRSF3-HKO mice. (D) Insulin tolerance tests on 12 month old mice showing greater insulin sensitivity in SRSF3-HKO mice. (E) QPCR on liver RNA from 12 month old mice showing significantly reduced PEPCK (*Pck1*) and glucose-6-phosphatase (*G6pc*) expression in SRSF3-HKO livers. Results are shown as mean \pm SEM. Asterisks indicate significance ($p < 0.05$) vs. WT. N indicates the number of animals per group. (F) Calculated HOMA insulin resistance for WT and HKO, male and female mice. WT female mice are significantly more insulin sensitive than WT males. HKO mice are significantly more sensitive than WT for either sex. Asterisks show statistical significance (** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$) as indicated. HOMA-IR shows a significant sex ($p < 0.01$) and genotype ($P < 0.0001$) effect by 2-way ANOVA.

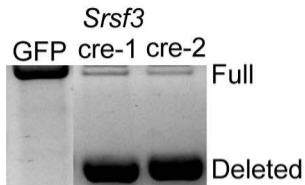
A**B**

Figure S3. A: Acute deletion of SRSF3 by adenoviral delivery of cre. Representative gel for RT-PCR showing expression of deleted form of Srsf3 in Srsf3-floxed primary hepatocytes after adeno-cre infection. GFP-cre is used as negative control. **B:** Representative western blots showing decreased expression of SRSF3 in si-Srsf3 treated HepG2 and Huh7 cells compared to si-C control. The blots were stripped and blotted for actin as loading control.

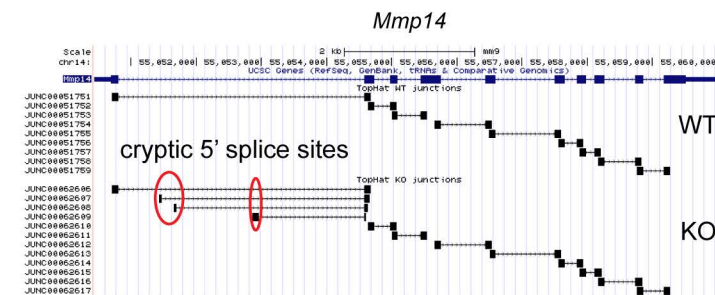
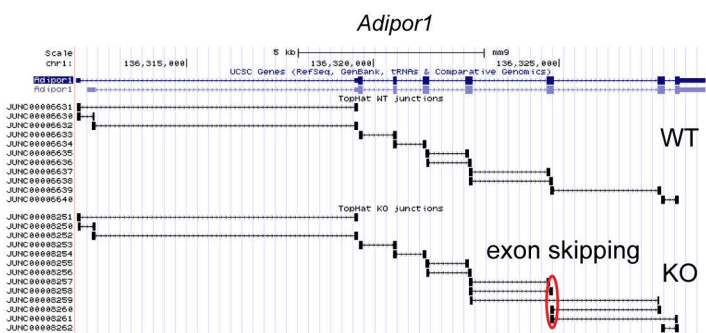
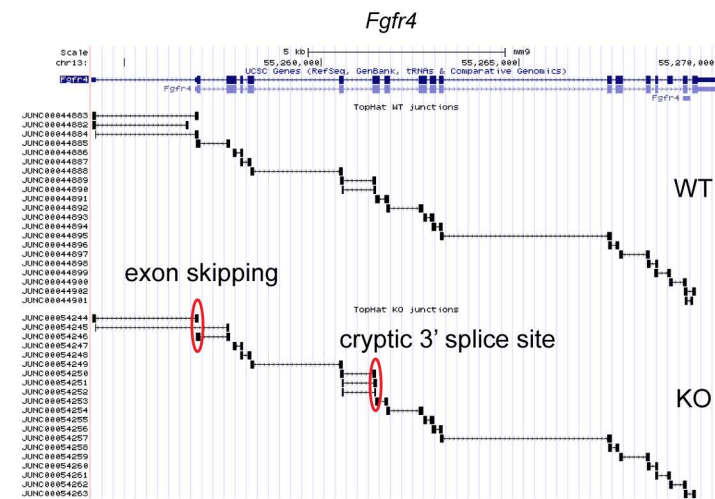
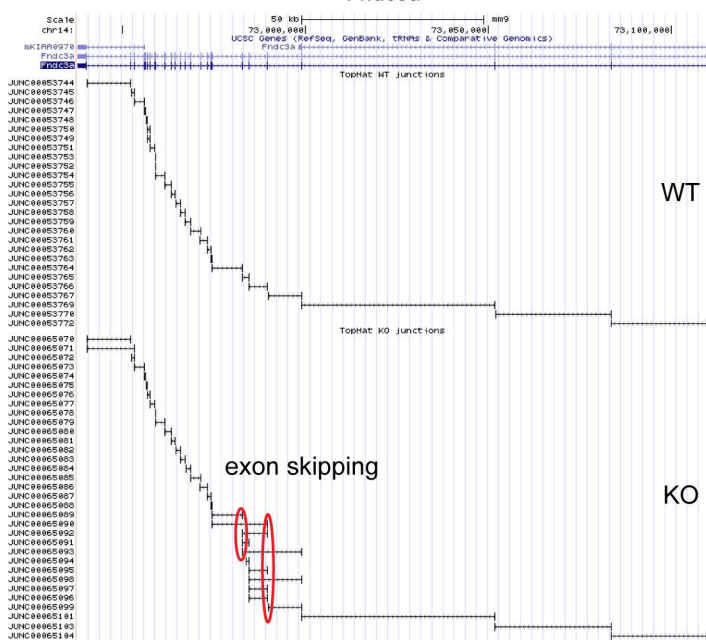
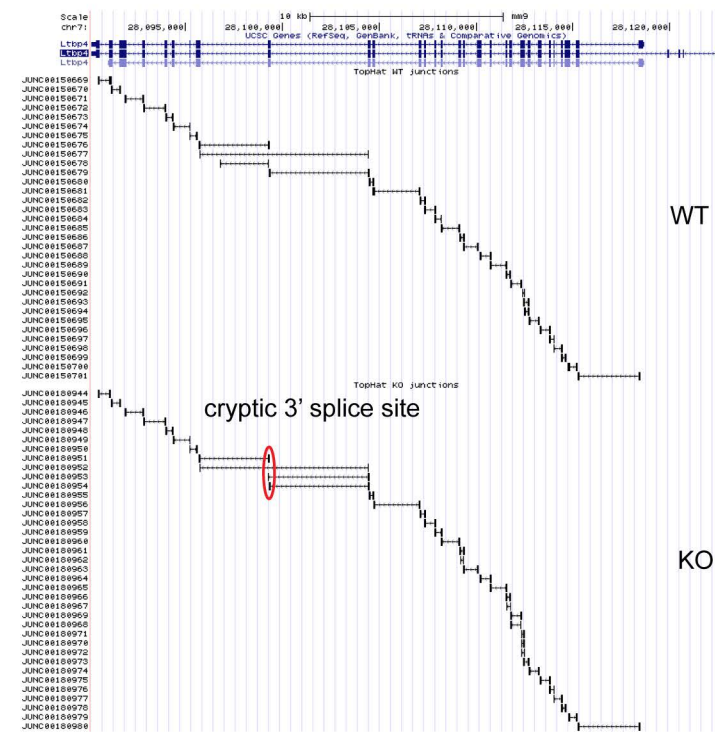
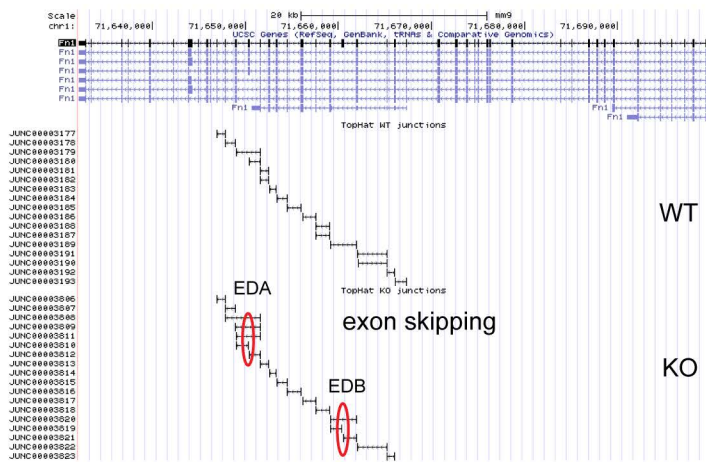


Figure S4. Altered exon-exon junction reads from RNAseq data aligned to genome using the UCSC genome browser. Altered exon usage is indicated in red. Reads from WT mice are given above reads from SRSF3HKO mice. Reads were aligned using Tophat.

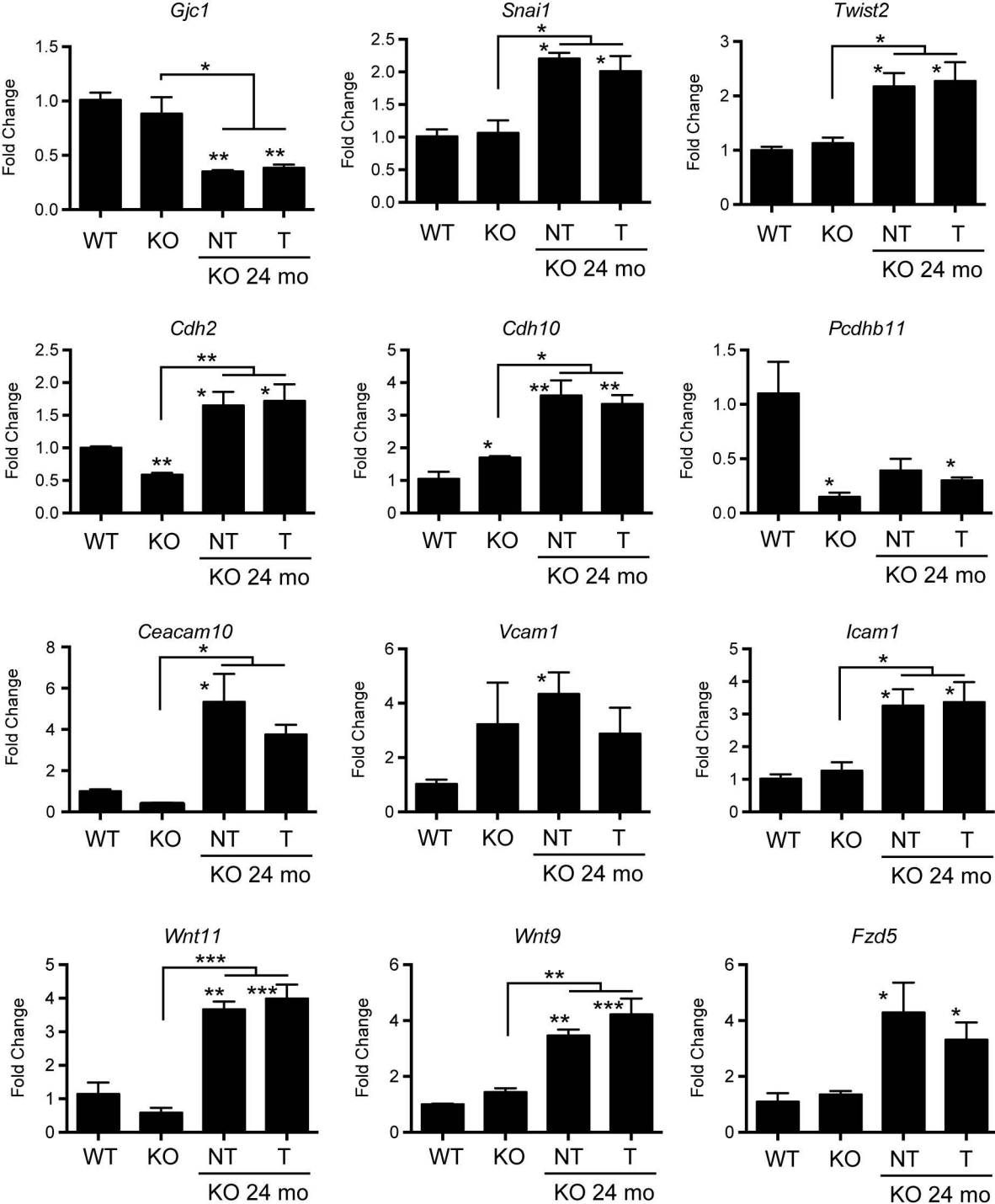


Figure S5. Alterations in EMT and Wnt signaling genes in SRSF3HKO mice. Gene expression was assessed in microarray data from wild-type mice (WT), SRSF3HKO mice at 1 month (KO), SRSF3HKO mice at 24 months (NT) and tumors from SRSF3HKO mice at 24 months (T). Asterisks indicate significance vs. WT or as indicated, *, **, *** $p < 0.05$, 0.01, or 0.001.

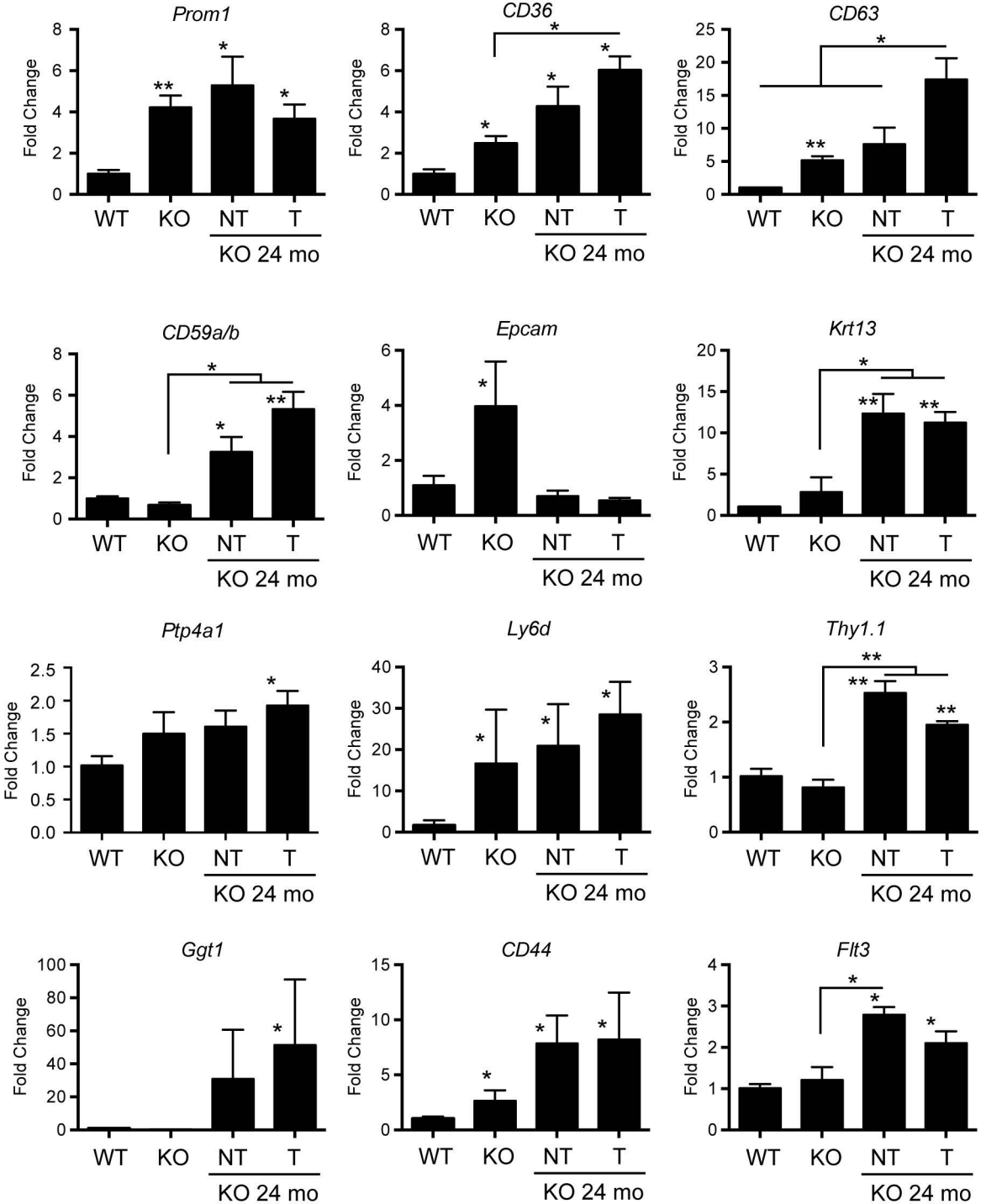


Figure S6. Alterations in progenitor and stem cell genes in SRSF3HKO mice. Gene expression was assessed in microarray data from wild-type mice (WT), SRSF3HKO mice at 1 month (KO), SRSF3HKO mice at 24 months (NT) and tumors from SRSF3HKO mice at 24 months (T). Asterisks indicate significance vs. WT or as indicated, *, ** $p < 0.05$, 0.01 .

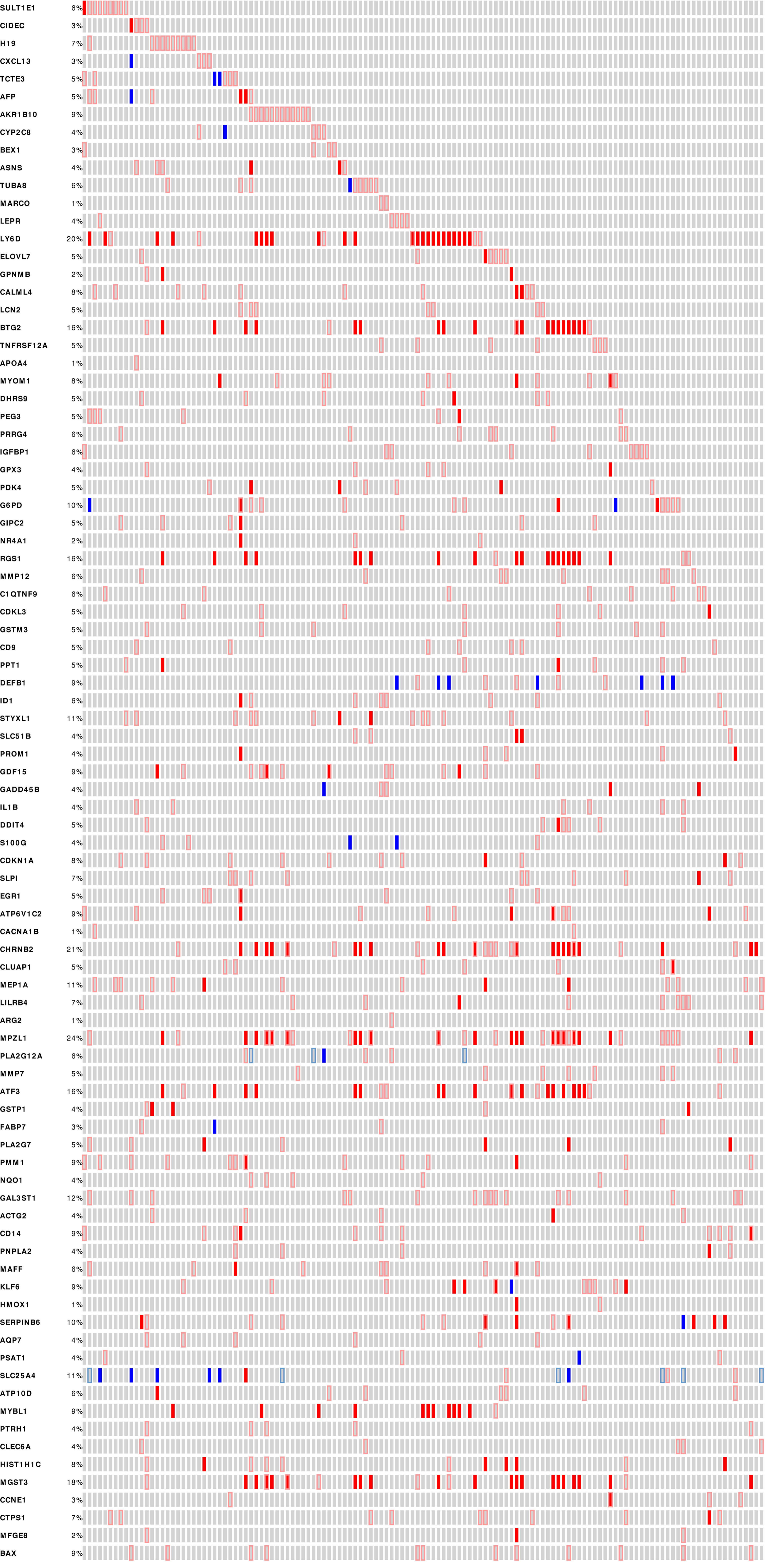


Figure S7. Alteration of the top 100 genes from the SRSF3HKO mice in the TCGA HCC database

Red indicates increased, blue indicates decreased, solid symbols represent amplification or deletion, hollow symbols represent expression changes.

Table S1: Tumor incidence in 24 month old mice

Age (month)	Genotype	# Mice Examined	# Mice w/ Tumor	Tumor Weight (g)	# Tumors per Mouse
Male Mice					
24	WT	14	1	1.2	1
24	HKO	8	8	1.59-4.16	1-10
Female Mice					
24	WT	22	0	0	0
24	HKO	15	15	1.71-3.81	1-10

Table S2: Changes of gene expression related to Fibrosis in SRSF3HKO mice at 1 mo

Locus	Description	SRSF3HKO/WT	p value
<i>Mmp12</i>	matrix metalloproteinase 12	8.013	1.36E-11
<i>Cdkl3</i>	cyclin-dependent kinase-like 3	7.207	2.87E-11
<i>Cdkn1a</i>	cyclin-dependent kinase inhibitor 1A (P21)	5.609	2.24E-10
<i>Mmp7</i>	matrix metalloproteinase 7	4.988	6.85E-10
<i>Ctgf</i>	connective tissue growth factor	3.814	1.31E-08
<i>Pdgfra</i>	platelet derived growth factor receptor, alpha polypeptide	3.582	2.85E-08
<i>Krt23</i>	keratin 23	3.425	5.05E-08
<i>Ltbp4</i>	latent transforming growth factor beta binding protein 4	0.488	1.62E-07
<i>Mmp15</i>	matrix metalloproteinase 15	0.446	5.15E-09
<i>Fgfr3</i>	fibroblast growth factor receptor 3	0.434	1.77E-09
<i>Mmp19</i>	matrix metalloproteinase 19	0.403	8.78E-11
<i>Fgfr4</i>	fibroblast growth factor receptor 4	0.332	2.37E-14
<i>Fgf1</i>	fibroblast growth factor 1	0.215	6.28E-22

Table S3: Hematology Analysis from 15-18 months old WT and SRSF3HKO mice; n=3

	WT	SRSF3HKO
WBC (K/ μ l)	2.5 \pm 0.47	1 \pm 0.1**
RBC (M/ μ l)	7.2 \pm 0.45	6.4 \pm 0.19
PLT (K/ μ l)	696 \pm 52	456 \pm 13**
Hb (g/dL)	10.4 \pm 0.67	9.9 \pm 0.38

Values are Mean \pm SE. Statistical significance was determined by Student's 2-tailed t test.

Table S5: GSEA enrichment of human HCC genesets in SRSF3HKO and WT mouse expression profiles

Geneset	Size	NES	FDR q-value
Enriched in SRSF3HKO			
CHIANG_LIVER_CANCER_SUBCLASS_UNANNOTATED_DN	141	-2.18	0.00
BOYALT_LIVER_CANCER_SUBCLASS_G3_UP	139	-2.05	0.00
CAIRO_HEPATOBLASTOMA_CLASSES_UP	406	-1.91	0.00
LEE_LIVER_CANCER_SURVIVAL_DN	76	-1.70	0.02
HOSHIDA_LIVER_CANCER_SUBCLASS_S1	184	-1.55	0.07
IIZUKA_LIVER_CANCER_PROGRESSION_G1_G2_DN	17	-1.48	0.10
BOYALT_LIVER_CANCER_SUBCLASS_G1_UP	82	-1.40	0.15
OKAMOTO_LIVER_CANCER_MULTICENTRIC_OCCURRENCE_UP	16	-1.37	0.16
HOSHIDA_LIVER_CANCER_LATE_RECURRENCE_UP	41	-1.36	0.15
YAMASHITA_LIVER_CANCER_WITH_EPCAM_UP	23	-1.31	0.17
HOSHIDA_LIVER_CANCER_SUBCLASS_S2	84	-1.31	0.16
Enriched in WT			
HSIAO_LIVER_SPECIFIC_GENES	175	2.50	0.00
CAIRO_LIVER_DEVELOPMENT_DN	189	2.37	0.00
BOYALT_LIVER_CANCER_SUBCLASS_G123_DN	40	2.30	0.00
LEE_LIVER_CANCER_SURVIVAL_UP	94	2.29	0.00
BOYALT_LIVER_CANCER_SUBCLASS_G3_DN	36	2.22	0.00
CHIANG_LIVER_CANCER_SUBCLASS_PROLIFERATION_DN	115	2.14	0.00
CHIANG_LIVER_CANCER_SUBCLASS_POLYSOMY7_UP	55	2.12	0.00
CAIRO_HEPATOBLASTOMA_DN	198	2.11	0.00
HOSHIDA_LIVER_CANCER_SUBCLASS_S3	193	1.98	0.00
CAIRO_HEPATOBLASTOMA_CLASSES_DN	145	1.95	0.00
HOSHIDA_LIVER_CANCER_SURVIVAL_DN	79	1.82	0.00
YAMASHITA_LIVER_CANCER_STEM_CELL_UP	41	1.78	0.01
CHIANG_LIVER_CANCER_SUBCLASS_CTNNB1_UP	132	1.75	0.01
ACEVEDO_LIVER_CANCER_DN	384	1.74	0.01
YAMASHITA_LIVER_CANCER_STEM_CELL_DN	49	1.74	0.01
WOO_LIVER_CANCER_RECURRENCE_DN	54	1.72	0.01
HOSHIDA_LIVER_CANCER_LATE_RECURRENCE_DN	53	1.56	0.03
ACEVEDO_LIVER_TUMOR_VS_NORMAL_ADJACENT_TISSUE_DN	192	1.53	0.04
CHIANG_LIVER_CANCER_SUBCLASS_CTNNB1_DN	127	1.42	0.07
ACEVEDO_NORMAL_TISSUE_ADJACENT_TO_LIVER_TUMOR_DN	243	1.35	0.11
YE_METASTATIC_LIVER_CANCER	19	1.31	0.14
IIZUKA_LIVER_CANCER_PROGRESSION_G2_G3_UP	20	1.30	0.14
CHIANG_LIVER_CANCER_SUBCLASS_INTERFERON_DN	38	1.27	0.16

6967969	Lrrk1	3.022E-03	0.679	0.057	0.684	0.057	135.42	11.15	91.99	7.69	7.071	0.124	6.513	0.125	leucine-rich repeat kinase 1
6815599	Pik3r1	5.359E-04	0.673	0.073	0.679	0.074	543.94	51.13	366.24	39.90	9.075	0.134	8.498	0.168	phosphatidylinositol 3-kinase, regulatory subunit, polypeptide 1 (p85 alpha)
6781689	Hs3st3a1 Hs3st3b1	2.654E-04	0.668	0.103	0.671	0.104	405.90	29.21	271.01	41.99	8.657	0.107	8.046	0.229	heparan sulfate (glucosamine) 3-O-sulfotransferase 3A1 heparan sulfate (glucosamine) 3-O-sulfotransferase 3B1
6989948	Rbpms2	8.521E-04	0.667	0.043	0.671	0.043	741.66	56.28	494.59	31.85	9.527	0.106	8.944	0.091	RNA binding protein with multiple splicing 2
6908528	Palmd	4.024E-03	0.657	0.020	0.664	0.020	1146.49	123.43	752.97	22.96	10.147	0.149	9.555	0.044	palmdelphin
6952137	Cadps2	1.893E-04	0.648	0.052	0.653	0.052	71.67	6.42	46.42	3.73	6.151	0.132	5.528	0.114	Ca2+-dependent activator protein for secretion 2
6803284	Serpina3c Serpina3	5.479E-04	0.636	0.073	0.640	0.073	236.85	20.22	150.59	17.28	7.877	0.122	7.216	0.163	serine (or cysteine) peptidase inhibitor, clade A, member 3C serine (or cysteine) peptidase inhibitor, clade A, member 3K
6899016	Arhgef11 4933430	5.968E-03	0.636	0.075	0.637	0.075	253.83	11.59	161.32	18.95	7.985	0.065	7.313	0.174	Rho guanine nucleotide exchange factor (GEF) 11 RIKEN cDNA 4933430H15 gene
6906749	Sema4a	5.809E-03	0.632	0.077	0.640	0.078	921.51	102.33	582.79	71.12	9.830	0.161	9.167	0.168	sema domain, immunoglobulin domain (Ig), transmembrane domain (TM) and short cytoplasmic domain, (semaphorin) 4A
6904979	Setd7	3.405E-03	0.619	0.057	0.625	0.057	128.50	11.99	79.58	7.30	6.993	0.135	6.302	0.135	SET domain containing (lysine methyltransferase) 7
6909278	Enpep	5.188E-03	0.605	0.063	0.608	0.063	603.82	41.02	365.34	38.00	9.231	0.102	8.498	0.144	glutamyl aminopeptidase
6945032	Smo	6.091E-04	0.597	0.040	0.604	0.041	270.24	27.64	161.32	10.84	8.062	0.154	7.327	0.100	smoothed homolog (Drosophila)
6820113	Gfra2	3.893E-03	0.593	0.078	0.594	0.078	122.75	4.45	72.76	9.62	6.938	0.052	6.160	0.191	glial cell line derived neurotrophic factor family receptor alpha 2
6778358	Sec14a	1.087E-04	0.582	0.023	0.583	0.023	1751.94	74.36	1019.95	40.78	10.772	0.063	9.992	0.058	SEC14-like 4 (S. cerevisiae)
6846197	Bbx A730021G18RI	2.411E-03	0.579	0.037	0.581	0.037	133.90	8.31	77.52	4.98	7.060	0.088	6.271	0.091	bobby sox homolog (Drosophila) RIKEN cDNA A730021G18 gene RIKEN cDNA A930011E06 gene
6749933	Adam23	2.116E-03	0.560	0.107	0.562	0.107	186.41	12.13	104.40	19.90	7.536	0.092	6.646	0.304	a disintegrin and metallopeptidase domain 23
6873133	Pik3ap1	3.010E-05	0.553	0.050	0.560	0.050	1446.64	152.58	800.10	71.75	10.481	0.162	9.633	0.126	phosphoinositide-3-kinase adaptor protein 1
6949593	Slc6a12	2.004E-03	0.551	0.027	0.563	0.027	1271.79	192.93	701.01	33.92	10.282	0.208	9.450	0.072	solute carrier family 6 (neurotransmitter transporter, betaine/GABA), member 12
7016584	Dcaf12l1	1.218E-03	0.528	0.042	0.529	0.042	105.47	5.84	55.64	4.41	6.716	0.082	5.789	0.110	DDB1 and CUL4 associated factor 12-like 1
6959254	Cyp2b13 Rnf170 C	5.10E-05	0.500	0.258	0.511	0.264	1087.66	151.87	543.41	280.38	10.055	0.220	8.728	0.691	cytochrome P450, family 2, subfamily b, polypeptide 13 polypeptide 9 ring finger protein 170 Dmx-like 1
6869545	Pde6c	4.61E-05	0.494	0.010	0.578	0.011	78.09	32.13	38.59	0.74	6.060	0.556	5.270	0.028	phosphodiesterase 6C, cGMP specific, cone, alpha prime
6943822	Casd1	7.208E-04	0.489	0.131	0.494	0.132	170.52	17.29	83.38	22.29	7.398	0.154	6.271	0.407	CAS1 domain containing 1
6874610	Bend7 Prc1	7.142E-04	0.435	0.059	0.438	0.059	183.28	17.22	79.69	10.74	7.506	0.130	6.288	0.206	BEN domain containing 7 protein regulator of cytokinesis 1
6895460	Zfhx4	7.278E-03	0.435	0.046	0.455	0.048	114.07	25.31	49.57	5.26	6.767	0.304	5.616	0.147	zinc finger homeodomain 4
6900071	Ngf	8.58E-05	0.277	0.089	0.498	0.160	472.75	347.05	130.93	41.98	8.039	1.080	6.890	0.448	nerve growth factor
6932190	Csn3	4.37E-05	0.273	0.116	0.571	0.242	178.26	140.42	48.64	20.62	6.413	1.234	5.369	0.561	casein kappa
6778972	Egfr	9.10E-05	0.272	0.039	0.316	0.045	1270.58	486.28	346.00	49.65	10.094	0.562	8.407	0.198	epidermal growth factor receptor
6915847	Lepr	1.63E-05	0.177	0.007	0.316	0.013	485.46	345.43	85.84	3.55	8.087	1.106	6.421	0.060	leptin receptor
6989195	Gldn	9.62E-06	0.156	0.022	0.242	0.034	323.04	212.68	50.30	7.06	7.698	0.932	5.623	0.209	gliomedin
6938093	Ppargc1a	1.29E-05	0.131	0.043	0.182	0.059	639.76	367.73	84.11	27.25	8.849	0.807	6.226	0.505	peroxisome proliferative activated receptor, gamma, coactivator 1 alpha
6832572	Miox	1.12E-06	0.124	0.004	0.325	0.010	340.77	291.65	42.40	1.35	7.027	1.414	5.405	0.046	myo-inositol oxygenase
6964531	Dmbt1	7.43E-07	0.101	0.003	0.179	0.006	462.16	288.94	46.70	1.47	8.031	1.208	5.544	0.045	deleted in malignant brain tumors 1
7016826	Gpc3	7.66E-07	0.085	0.002	0.180	0.005	602.14	460.57	51.35	1.40	8.160	1.289	5.681	0.039	glypican 3
6810697	Akr1c18	4.14E-07	0.042	0.005	0.110	0.013	754.10	518.98	32.02	3.84	8.188	1.733	4.981	0.167	aldo-keto reductase family 1, member C18

Supporting Materials and Methods

Blood Chemistry Analysis

Hematology analysis was performed using un-clotted blood from SRSF3HKO and WT mice. Fasting blood glucose concentrations were measured using ACCU-Check glucometer (Roche). Serum triglycerides were measured using Triglyceride ELISA Kit (LabAssay from Wako Chemicals USA) following the manufacturers' instructions.

Glucose Tolerance Test (GTT) and Insulin Tolerance Test (ITT)

To assess glucose tolerance, WT and HKO mice were fasted for 6 h. Blood glucose concentrations were measured by tail bleeding using an ACCU-Check glucometer, at time 0 and indicated times thereafter, for a total of 2 h, after an ip injection of glucose of 1 gm/kg body weight. For ITTs, mice were fasted for 4 hours and insulin used for ip injection was 0.5U/ kg body weight.

Isolation of Primary Murine Hepatocytes:

Primary hepatocytes from WT and SRSF3HKO liver were obtained by two-step perfusion with liver perfusion medium (Gibco BRL 17701-038; Gibco, Detroit, MI) followed by digestion medium, L15 (Sigma L5520; Sigma, St. Louis, MO) containing 0.15 g/ml collagenase and 10 µg/ml DNase I. Liver cells were disaggregated by passing through a 40-µm-pore nylon mesh Cell Strainer (BD Falcon). The numbers of total viable cells were determined by Trypan blue staining.

Confirmation of Altered Splicing and Gene Expression.

Total RNA was reverse transcribed using random hexamers with MultiScribe MuLV reverse transcriptase (Applied Biosystems) then subjected to PCR using GoTaq polymerase (Promega). For confirmation of exon skipping, RT-PCR was done on each sample using primers on constitutive exons flanking the altered exon. To ensure a linear PCR amplification allowing semi-quantitative assessment of the spliced products, we performed PCR analysis with 25 cycles. Representative results from each group (WT and HKO) are shown in the figures. PCR products were separated on 3% agarose or 12% non-denaturing polyacrylamide gels, stained with

ethidium bromide, visualized and quantified with Kodak Electrophoresis Documentation and Analysis System (EDAS) 290.

For mRNA expression studies, real-time quantitative RT-PCR assays were performed for each sample in triplicate in a final reaction volume of 20 μ l. The endogenous housekeeping gene *Gapdh* or the gene of interest was amplified using 20 ng of cDNA with iTaq SYBR Green supermix with ROX (Bio-Rad Laboratories) on Chromo4 Thermal Cyclers (Bio-Rad). The data were analyzed using the DDCt method and presented as the relative change in gene expression, normalized to *Gapdh*. All primers for qPCR were designed with Universal Probe Library (Roche).

Growth hormone treatment.

1-month-old mice were starved overnight and injected intraperitoneally with 5 mg of GH/kg BW in normal saline (0.9% NaCl) in a final volume of 0.2 ml. Animals injected with saline were used to evaluate basal conditions. The animals were sacrificed 7.5 min after injection, the livers were removed and snap frozen for protein isolation.

Analysis of Microarray data

Expression data from exon array were analyzed by two methods: 1) data were normalized by RMA and analyzed by ANOVA using Genespring (Agilent Technologies, Santa Clara, CA); 2) data were normalized by RMA and analyzed by ANOVA using Non-Coder (<http://noncoder.mpi-bn.mpg.de/#>); and 3) unnormalized raw data was analyzed using a Bayesian variance modeling approach in VAMPIRE. In all cases multiple testing correct was applied using a FDR q-value < 0.05. Lists of significant genes by the different approaches were imported into GeneSpring for visualization and compilation. Enrichment analyses were performed on the list of altered genes using GeneGo's Metacore software (St. Joseph, MI), and on the raw data using Gene Set Enrichment Analysis (GSEA, Broad Institute, Cambridge, MA).

IGF2 and insulin treatment:

IGF2 and insulin treatment were performed using primary hepatocytes from SRSF3HKO and age matched WT mice. After plating and attachment, primary hepatocytes were serum starved for 5 hrs and then treated with IGF2 (50 or 100 ng/ml) or insulin (30 or 100 nM) for 5 minutes.

Immunoblotting and Immunohistochemistry

For immunoblotting, livers or primary hepatocytes were lysed in extraction buffer (20mM Tris, pH 7.9; 300 mM NaCl; 1% Nonidet P-40) supplemented with protease and phosphatase inhibitors. Liver/hepatocyte lysates were separated by SDS-PAGE, transferred to PVDF membranes and immunoblotted with primary antibodies followed by HRP-conjugated secondary antibodies and developed using an ECL+ kit (GE).

Antibodies used for immunoblotting : anti-phospho STAT5 (M-186) rabbit polyclonal antibody (1:1000 dilutions, Cell Signaling Technologies), anti-insulin/insulin-like growth factor -1 receptor dual phosphospecific (pYpY1162/1163) rabbit polyclonal (Biosource International, Camarilo, CA, USA), anti-Insulin receptor beta rabbit polyclonal (BD Transduction Laboratories), non-phospho (active) beta catenin (Ser33/37/Thr41)(D13A1) rabbit monoclonal antibody (Cell Signaling Technologies), phosphor-Gsk-3 Beta rabbit polyclonal antibody (Cell Signaling Technologies), anti-cyclin D1 rabbit polyclonal antibody (abcam), anti- c-Myc rabbit polyclonal antibody (Cell Signaling Technologies), anti-total/phospho-AKT (Cell Signaling Technologies), anti-total/phospho ERK1/2 rabbit polyclonal antibody (Cell Signaling Technologies), anti-STAT5 rabbit polyclonal antibody (Cell Signaling Technologies), anti-phospho-NFkB-65 rabbit polyclonal antibody (Cell Signaling Technologies), anti-phospho-JNK/SAPK rabbit polyclonal antibody (Cell Signaling Technologies).

Immunohistochemistry was performed on formalin-fixed, paraffin-embedded mouse liver sections using the following antibodies: anti-F/480 (MCA497B, AbD Serotec, UK), anti-IGF2 (abcam, ab63984), anti-SRSF3 (abcam, ab125124), anti-Laminin (abcam, ab80580). Alpha-smooth muscle actin staining was performed using Actin, α -Smooth Muscle, Immunohistology Kit (Sigma). TUNEL staining was carried out using ApopTag Peroxidase In Situ Apoptosis Detection Kit (Millipore). H&E-stained sections were analyzed for inflammation according to the following scoring system: 0, no inflammation; 1, mild lymphocytic infiltration in the portal triad; 2, severe lymphocytic infiltration in portal triad; 3, extended infiltration of lymphocytes throughout liver. Sections were scored for steatosis according to the following scale: 0, no steatosis; 1, microsteatosis; 2, microsteatosis and mild macrosteatosis; 3, severe macrosteatosis. Sirius red stained sections were analyzed for fibrosis as follows: 0, no fibrosis; 1, mild fibrosis in the portal triad; 2, severe fibrosis in portal triad; 3: bridging fibrosis.

For the human liver cancer tissue array (LV1503, US Biomax) and the pathological samples of HCC and normal liver, immunohistochemistry was performed using anti-SRSF3 rabbit polyclonal antibody (abcam, ab125124) with Super PicTure Polymer Detection Kit (Invitrogen Corporation, Frederick, MD).